

SUPPLEMENTARY DATA - FIGURES

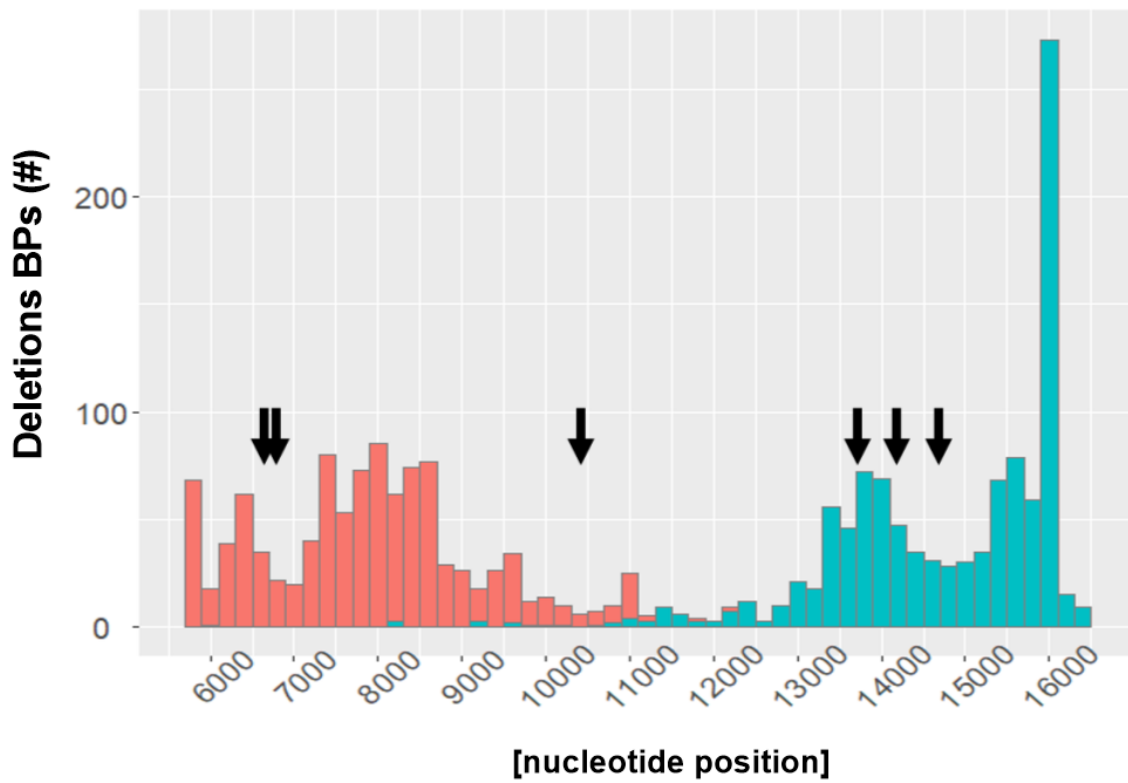


Figure S1

Distribution of breakpoints (BPs) inside the major arc with 5'-BPs shown in red, 3'-BPs shown in cyan and triplex motifs shown with arrows. Triplex motifs were detected using the triplex package in R with default scoring (min score=15). Breakpoints from the MitoBreak database.

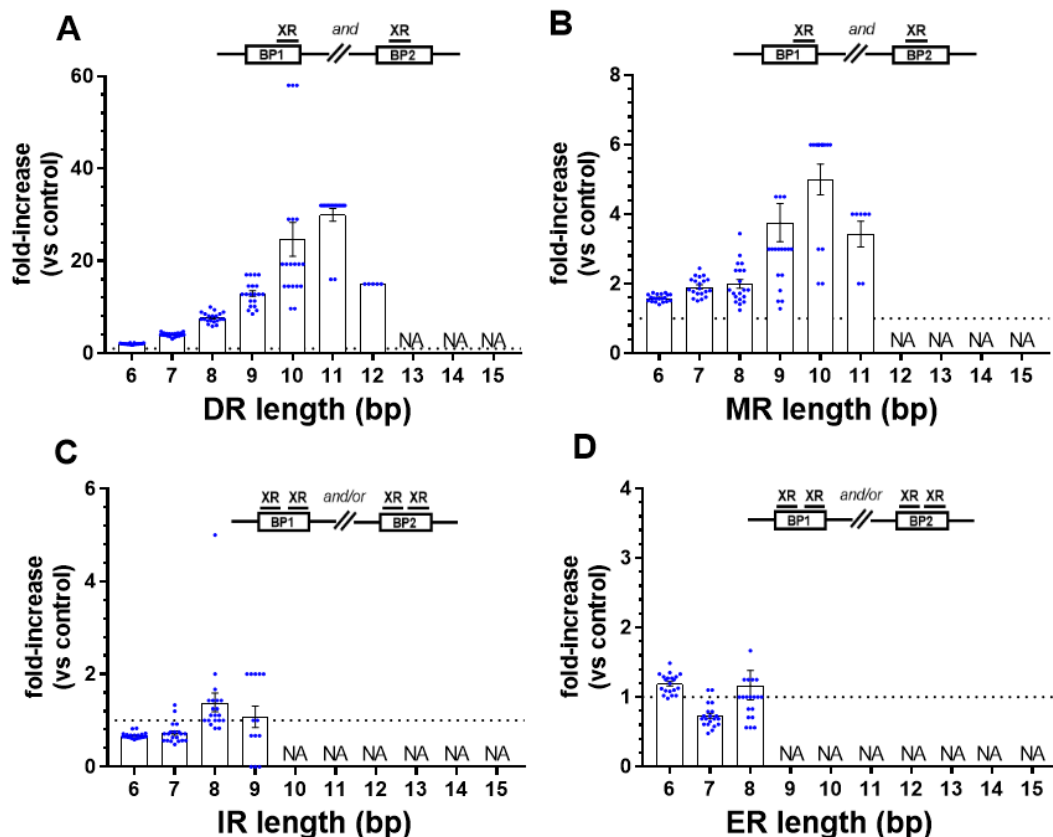


Figure S2

Direct repeat (DR; A) and mirror repeat (MR; B) motifs are enriched around actual breakpoints (BPs) compared to reshuffled breakpoints, but this is not true to the same extent for inverted repeats (IR; C) and everted repeats (ER; D). DR motifs show the strongest level of enrichment (note the y-axis). Controls were generated by reshuffling the deletion BPs while maintaining their distribution and the fold-change compared to controls was calculated ($n=20$, mean \pm SD shown). The schematic drawings above (A-C) depict the orientation of the motifs (XR) in relation to the BPs. NA, analysis not possible due to limited sample size.

(A) DR motifs between 6 and 15 bps and their level of enrichment around deletion BPs.

(B) MR motifs between 6 and 15 bps and their level of enrichment around deletion BPs.

(C) IR motifs between 6 and 15 bps and their level of enrichment around deletion BPs.

(D) ER motifs between 6 and 15 bps and their level of enrichment around deletion BPs.

Location of repeat (XR) relative to breakpoint region (BP)

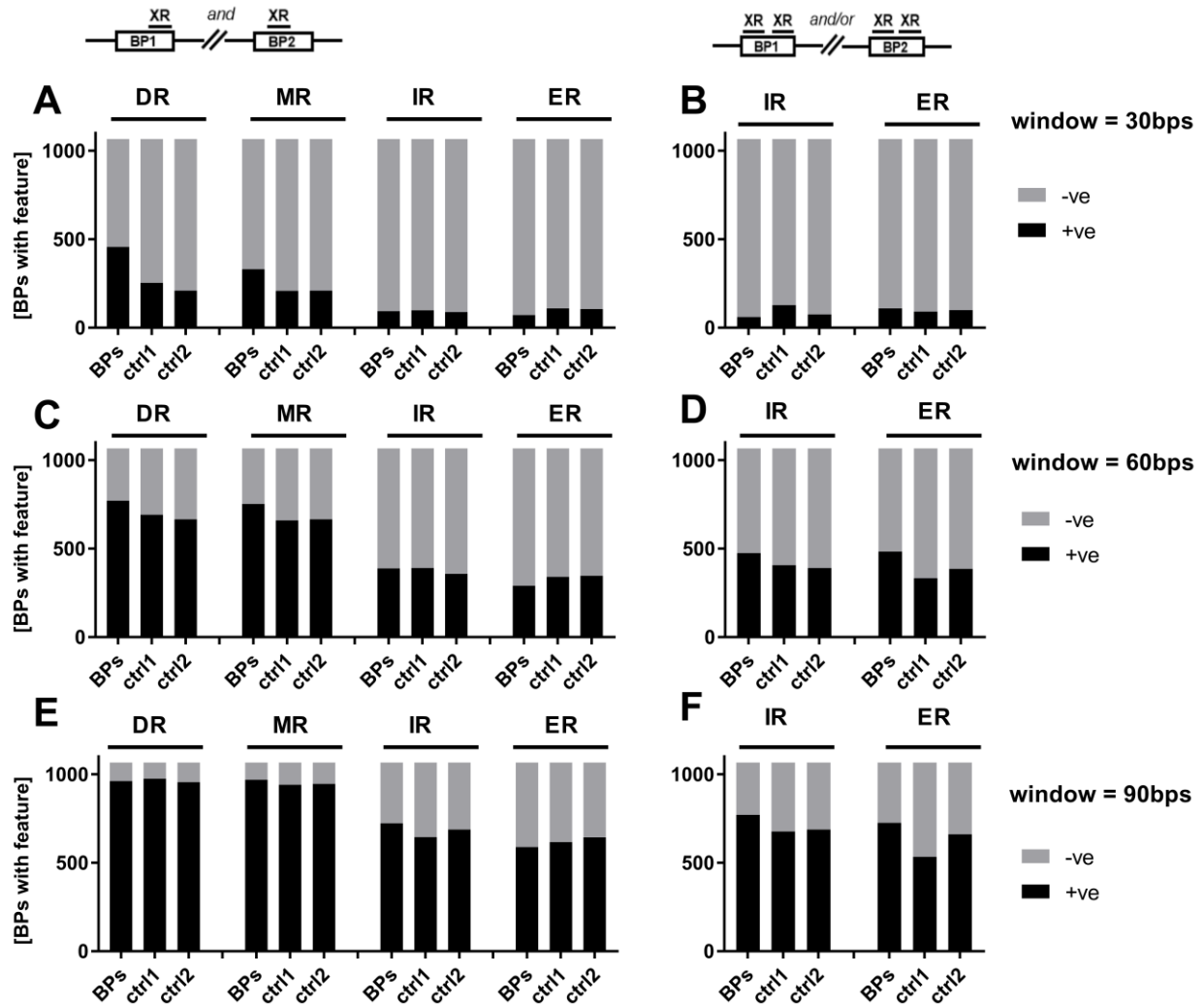


Figure S3

In this analysis we shifted each breakpoint (BP) by 200 bps to serve as its own control (ctrl1) and as a second control we generated fully random breakpoints within the major arc (ctrl2). Direct repeat (DR) and mirror repeat (MR) motifs remain somewhat enriched around actual BPs compared to reshuffled breakpoints even considering larger window sizes around BPs (A, C). In contrast, inverted repeat (IR) and everted repeat (ER) motifs are only enriched if we consider larger but not smaller window sizes (D, F) and the biological relevance of marginally increased IR motifs at BPs (+10 to 20%) is unclear.

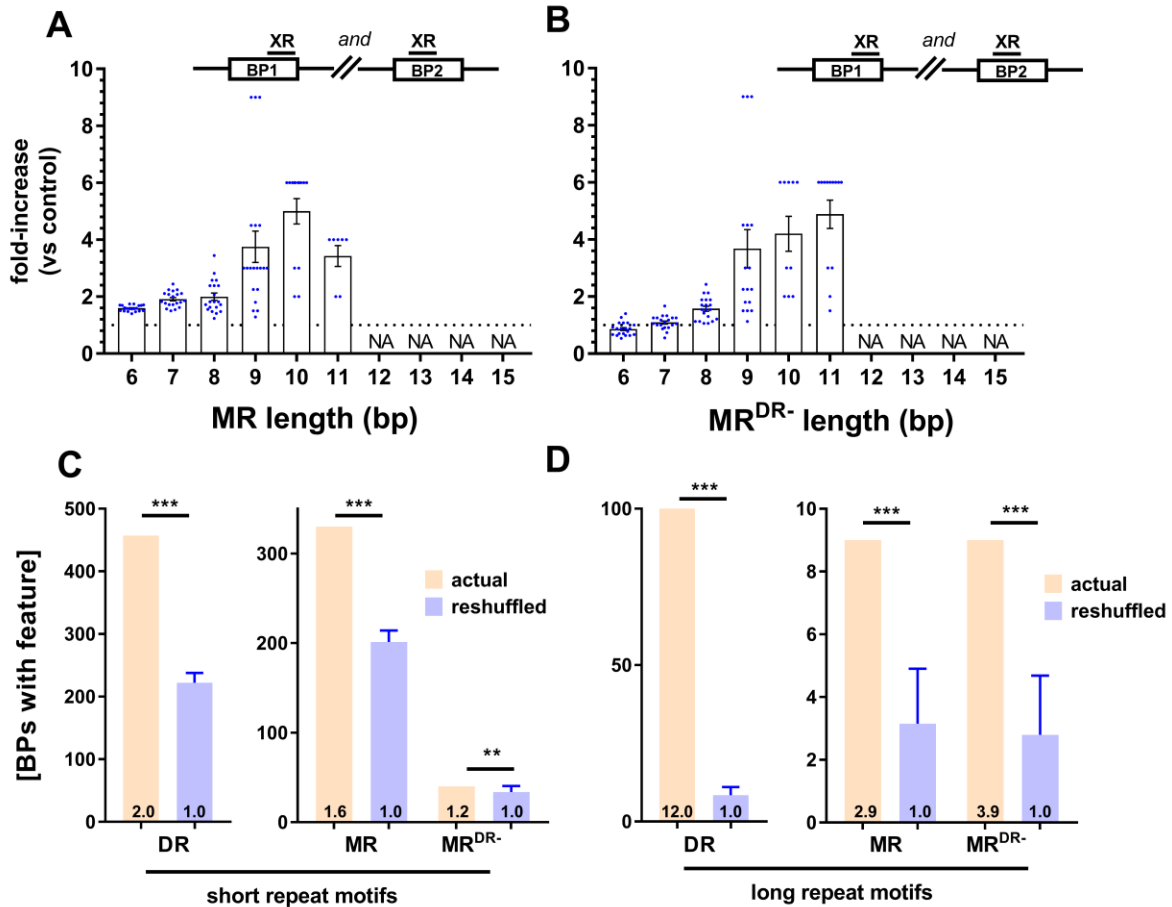


Figure S4

MR motifs of different lengths are significantly enriched around actual breakpoints (BPs) compared to reshuffled BPs (A, B). Removal of MR-DR hybrid motifs attenuates the correlation for short MR motifs but not for longer ones (MR^{DR}; B). When we separate the motifs into two groups, 6 to 8 bp (C) and 9 to 15 bp long motifs (D), longer motifs are less abundant but more specifically enriched around BPs. Controls were generated by reshuffling the deletion BPs while maintaining their distribution (n=20, mean \pm SD shown). The schematic drawings above (A, B) depict the orientation of the motifs (XR) in relation to the BPs. *** $p < 0.0001$, ** $p < 0.001$ by one sample t-test. Breakpoints from the MitoBreak database.

(A) MR motifs between 6 and 15 bps and their level of enrichment around deletion BPs.

(B) MR^{DR} motifs between 6 and 15 bps and their level of enrichment around deletion BPs.

(C) DR, MR and MR^{DR} motifs of 6 to 8 bp length and their level of enrichment around deletion BPs.

(D) DR, MR and MR^{DR} motifs of 9 to 15 bp length and their level of enrichment around deletion BPs.

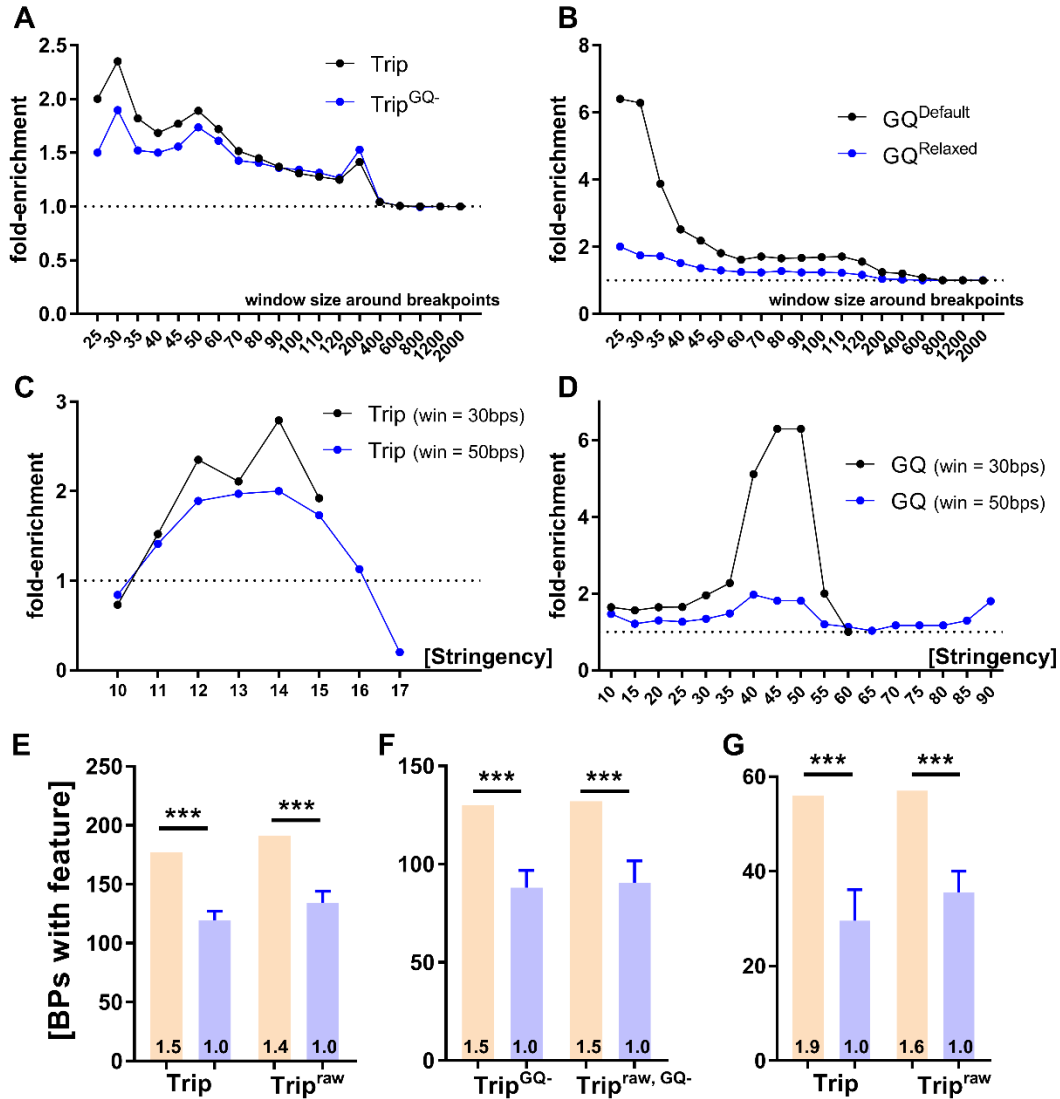


Figure S5

Triplex motifs (min score=12) are enriched around actual breakpoints (BPs) compared to shifted BPs across different window sizes (A) with similar results after removal of G-quadruplex (GQ)-triplex hybrid motifs (Trip^{GQ-}). GQ motifs are also enriched across different window sizes (B). In addition, it makes little difference if we vary the stringency (min score) of the detection algorithm. Both triplex (C) and GQ motifs (D) are enriched around actual BPs compared to shifted BPs across different min scores. Finally, in the main figure (Fig. 3) we only excluded predicted triplex motifs that were exact duplicates. Here, we show that “stringent” removal of both duplicate and partially overlapping motifs has only a marginal influence on the results (E-G), because these motifs usually overlap with the same BPs. *** p < 0.0001 by one sample t-test. Breakpoints from the MitoBreak database.

(A) Fold-enrichment of triplex motifs, and of triplex motifs excluding triplex-GQ hybrid motifs (Trip^{GQ-}), around actual BPs compared to shifted BPs (min score=12, relaxed settings). Different window sizes shown.

(B) Fold-enrichment of GQ motifs detected with default settings (min score = 47) or relaxed settings (min score = 26) around actual BPs compared to shifted BPs.

(C) Fold-enrichment of triplex motifs varying the minimum score cutoff, window sizes of 30 and 50 bps shown.

(D) Fold-enrichment of GQ motifs varying the minimum score cutoff, window sizes of 30 and 50 bps shown.

(E) Fold-enrichment after removal of all overlapping triplex motifs (n=22) compared with no removal (raw, n=33). Min score=12.

(F) Fold-enrichment after removal of all overlapping triplex motifs (n=15) compared with no removal (raw, n=33). Min score=12, excluding triplex-GQ hybrid motifs (Trip^{GQ-})

(G) Fold-enrichment after removal of all overlapping triplex motifs compared with no removal (raw). Min score=15 (default).

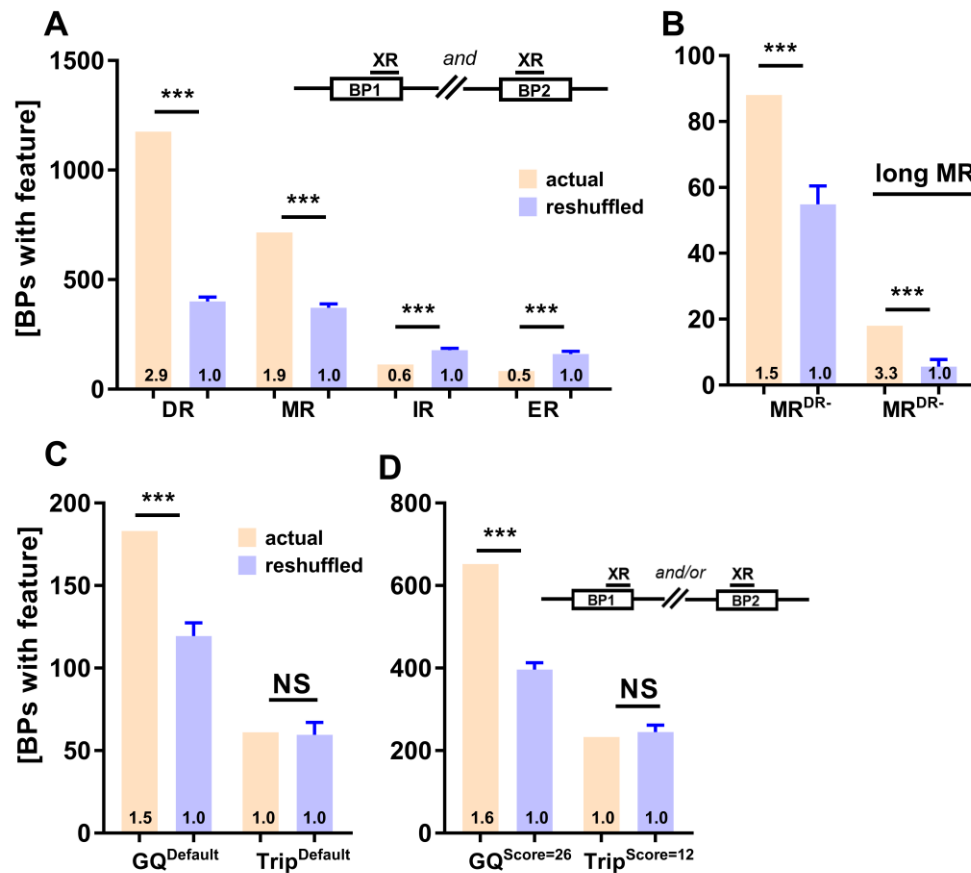


Figure S6

Our findings on the **Hjelm et al. (2019)** dataset agree with the MitoBreak data (**Fig. 2**) except for the relationship between triplex motifs and deletions (C, D). Direct repeat (DR) and mirror repeat (MR) motifs are significantly enriched around actual deletion breakpoints (BPs) compared to reshuffled BPs,

but the same is not true for inverted repeat (IR) and everted repeat (ER) motifs (A). The enrichment of MR motifs at deletion BPs is attenuated when MRs that have the same sequence as DR motifs are removed (MR^{DR-}), compared with reshuffled controls. Long MR motifs have a length of 9 to 15 bps. GQ motifs are enriched at BPs, but the data for triplex motifs is inconsistent (C, D). Controls were generated by reshuffling the deletion BPs while maintaining their distribution ($n=20$, mean \pm SD shown). *** $p<0.0001$ by one sample t-test.

(A) The number of deletion BPs associated with DR, MR, IR or ER motifs at both BPs compared with reshuffled controls.

(B) The number of BPs associated with MR motifs at both BPs after removal of hybrid MR-DR motifs (MR^{DR-}), compared with reshuffled controls. Long MR motifs have a length of 9 to 15 bps.

(C) The number of deletion BPs associated with GQ and triplex-forming motifs around BPs compared with reshuffled controls (min score = default).

(D) The number of deletion BPs associated with GQ and triplex-forming motifs around BPs compared with reshuffled controls (min score = relaxed).

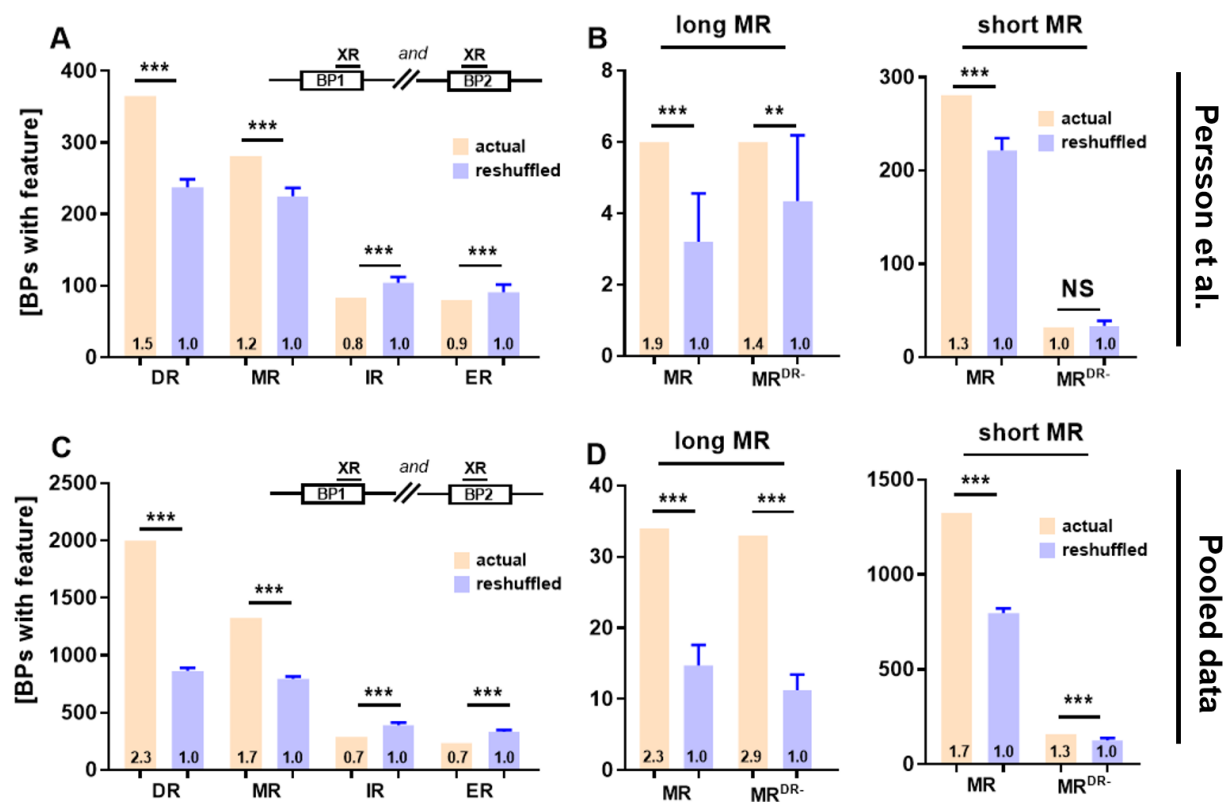


Figure S7

Our findings on the **Persson et al. (2019)** dataset and the pooled dataset (Hjelm + Persson + MitoBreak) agree with the MitoBreak data (**Fig. 2**). Direct repeat (DR) and mirror repeat (MR) motifs are significantly enriched around actual deletion breakpoints (BPs) compared to reshuffled BPs, but the same is not true for inverted repeat (IR) and everted repeat (ER) motifs (A, C). The correlation between MR motifs and deletion BPs is attenuated when adjusted for MR motifs that are equivalent to a DR motif (B, D). Controls were generated by reshuffling the deletion BPs while maintaining their distribution ($n=20$,

mean \pm SD shown). The schematic drawing above (A, C) depicts the orientation of the repeat (XR) half-sites and motifs in relation to the BPs. *** $p < 0.0001$, ** $p < 0.001$ by one sample t-test.

(A) The number of deletion BPs associated with DR, MR, IR or ER motifs at both BPs compared with reshuffled controls (based on **Persson et al. 2019**).

(B) The number of deletion BPs associated with MR motifs at both BPs, before (MR) and after removal of hybrid MR-DR motifs (MR^{DR-}), compared with reshuffled controls. Long MR motifs are defined as 9 to 15 bps and short MR motifs as 6 to 8 bps (based on **Persson et al. 2019**).

(C) Same as (A) but for the pooled dataset.

(D) Same as (B) but for the pooled dataset.

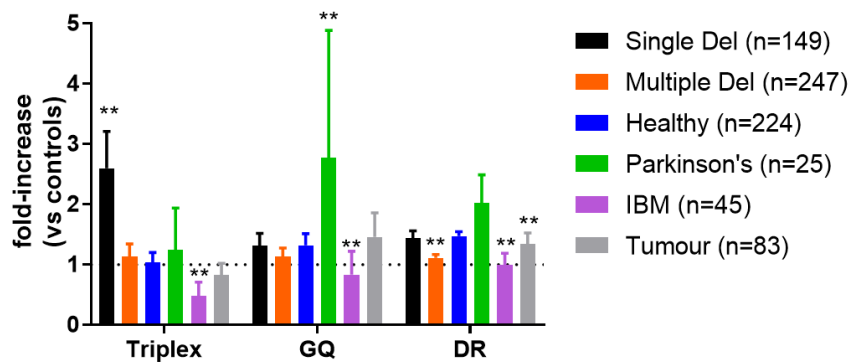


Figure S8

We split the MitoBreak data into subgroups according to disease etiology and for each subgroup we show fold-enrichment of motifs around actual breakpoints (BPs) compared to reshuffled BPs. The deletion BPs in the database comprise six groups: single mtDNA deletion syndromes, multiple mtDNA deletion syndromes, healthy tissues, Parkinson's disease, inclusion body myositis (IBM) or tumour tissues. We found that triplex motifs (min score = 12) are enriched compared to controls in several subgroups with the strongest increase in the single deletion group. This is different from the pattern seen for G-quadruplex (GQ; min score = 26) and direct repeat (DR) motifs.

Fold-enrichment calculated by comparison with $n=20$ reshuffled breakpoints (mean \pm SD), window size around breakpoints = 50 bps for GQ and triplex and 30 bps for DR. Significance based on one-way ANOVA with Dunnett's post-hoc test, ** $p < 0.05$ compared with the healthy tissues group.

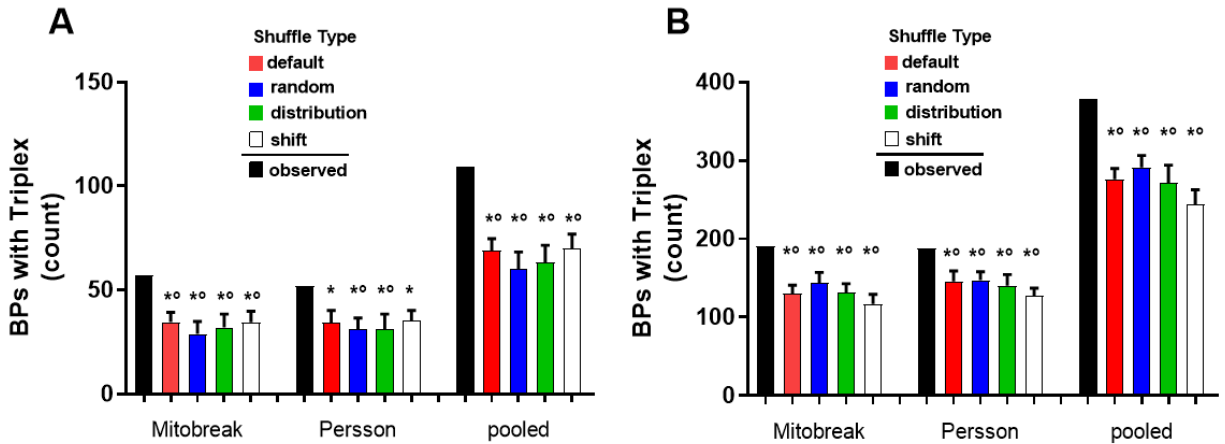


Figure. S9

The enrichment of triplex motifs around mtDNA deletion breakpoints (BPs) in the MitoBreak and Persson dataset is consistent when different BP shuffling methods are used and robust to statistical assumptions (one sample t-test vs. Fisher's exact test). "Default" shuffle, each mtDNA deletion as a whole is randomly redistributed within the major arc. "Random", individual mtDNA BPs are distributed randomly within the major arc. "Distribution", individual mtDNA BPs are redistributed in a way that approximates their original distribution. "Shift", each BP is randomly shifted by 100 to 300 bps, thereby maintaining the original distribution. Controls were generated by reshuffling the deletion BPs as described above (n=20, mean \pm SD shown). * $p < 0.05$ by one sample t-test and ° $p < 0.05$ by Fisher's exact test.

(A) The number of observed BPs associated with triplex motifs compared to the number of reshuffled control BPs associated with triplex motifs (min score=15; default).

(B) Same as in (A) but with relaxed criteria for the detection of triplex motifs (min score=12).

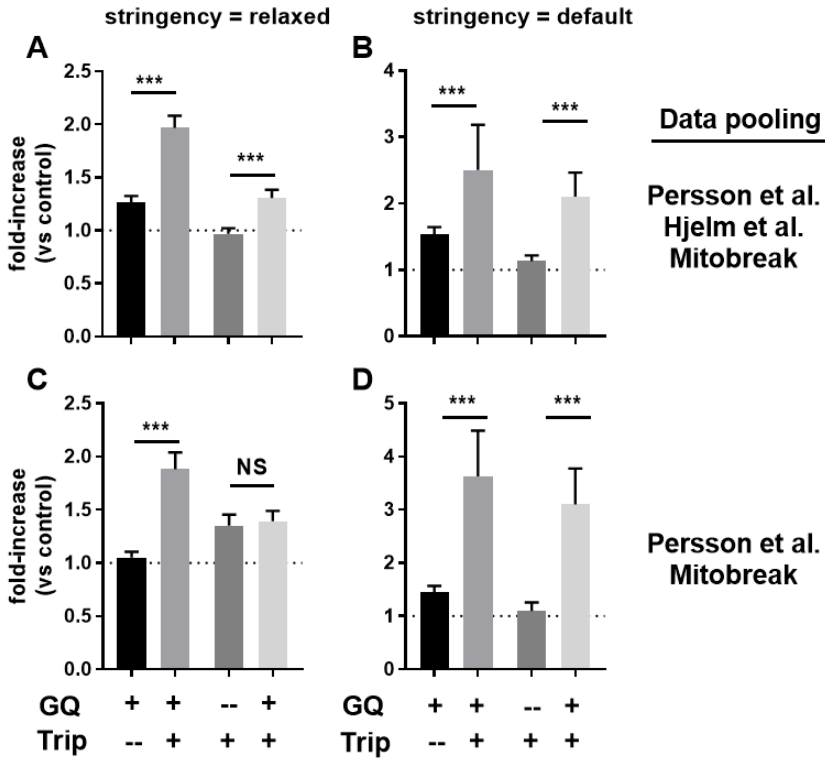


Figure S10

Fold-enrichment of motifs around actual breakpoints (BPs) compared to reshuffled breakpoints is shown. We define partially overlapping G-quadruplex (GQ)-triplex hybrid motifs if the sequence midpoints are within 50 bps of each other and we show that such hybrid motifs (GQ⁺, Trip⁺) are more strongly enriched around BPs compared to GQ (GQ⁺, Trip⁻) and triplex (Trip⁺, GQ⁻) motifs in isolation. Fold-enrichment calculated by comparison with n=20 reshuffled breakpoints (mean \pm SD). *** p < 0.0001 by student's t-test.

(A) Fold-enrichment of the above motifs around mtDNA deletion BPs compared to reshuffled BPs. Data pooled from three studies, GQ min score = 26, Triplex min score = 12.

(B) Fold-enrichment of the above motifs around mtDNA deletion BPs compared to reshuffled BPs. Data pooled from three studies, GQ min score = 47, Triplex min score = 15.

(C) Fold-enrichment of the above motifs around mtDNA deletion BPs compared to reshuffled BPs. Data pooled from two studies, GQ min score = 26, Triplex min score = 15.

(D) Fold-enrichment of the above motifs around mtDNA deletion BPs compared to reshuffled BPs. Data pooled from two studies, GQ min score = 47, Triplex min score = 15.

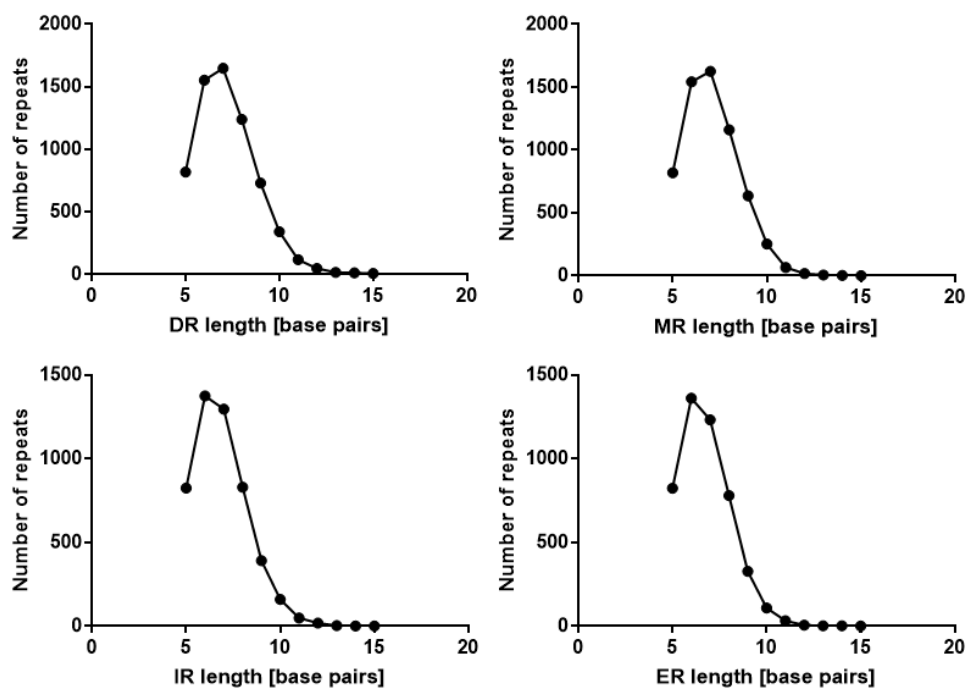


Figure S11

Long repeat motifs of any kind are rare in the human mitochondrial genome and their number decreases approximately in a log-linear fashion (not shown here). Since we plot unique repeats here, not counting multiple occurrences, the numbers for very short repeats decrease again due to their limited sequence diversity.

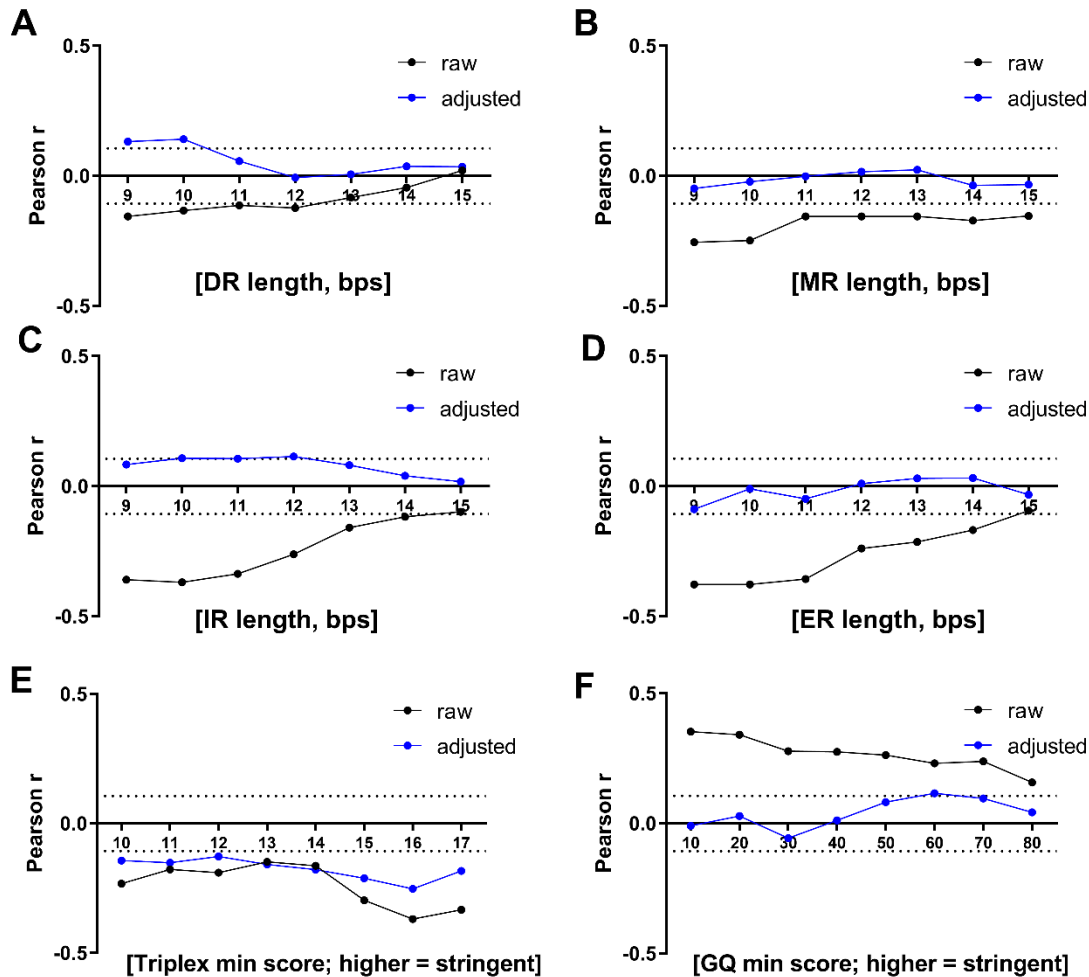


Figure S12

Secondary structures that are more thermodynamically stable would pose a larger threat to genome stability and should show a stronger inverse correlation with MLS. Although longer repeats will form more stable structures, they do not show a stronger correlation with MLS (A-D). Triplex (E) and G-quadruplex (GQ) motifs (F) with a higher minimum score should also be more stable, however, stable GQ motifs do not show an inverse correlation with MLS (F). Only for triplex motifs do we see the expected trend, a stronger inverse correlation with MLS in the case of more stable motifs (E). The critical R value at which $p < 0.01$ is indicated.

- (A) Longer direct repeat (DR) motifs do not consistently correlate with MLS after adjustment.
- (B) Longer mirror repeat (MR) motifs do not consistently correlate with MLS after adjustment.
- (C) Longer inverted repeat (IR) motifs do not consistently correlate with MLS after adjustment.
- (E) Longer everted repeat (ER) motifs do not consistently correlate with MLS after adjustment.
- (F) Triplex motifs show a consistent inverse correlation with MLS that is stronger for high scoring triplex motifs (i.e. motifs more likely to be stable).
- (G) G-quadruplex motifs show a consistent positive correlation with MLS that is attenuated after adjustment. In contrast to triplex motifs, higher scoring GQ motifs (i.e. motifs more likely to be stable) do not show a stronger correlation with MLS.

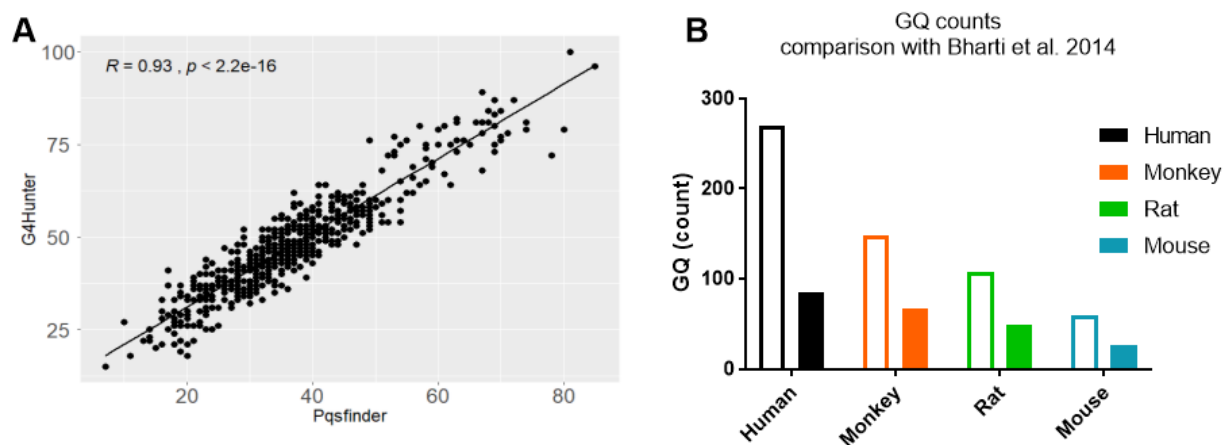


Figure S13

Two different G-quadruplex (GQ) prediction tools, pqsfinder and G4Hunter (both ran with default settings), perform similarly across 600 mammalian species (A). The data we generated using pqsfinder (filled bars) is comparable to Bharti et al. 2014 (open bars), although, our thresholds are more conservative yielding lower GQ counts (B). Our data is based on the mt genomes of NC_005943.1 (*Macaca mulatta*), AC_000022.2 (*Rattus norvegicus*), NC_005089.1 (*Mus musculus*) and NC_012920.1 (*Homo sapiens*).

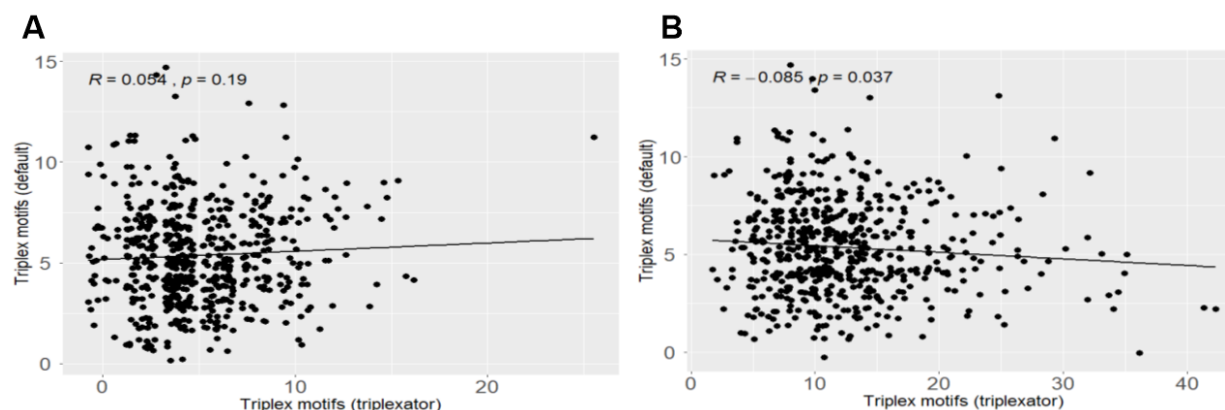


Figure S14

In contrast to the GQ prediction tools (Fig. S12), two different triplex motif prediction tools, the triplex package in R and triplexator (both ran with default settings), do not predict similar numbers of triplex motifs across 600 mammalian species. We use triplexator in -ds mode to detect potential triplex target sites (A) and in -ss mode to detect potential triplex forming oligonucleotides (B). Data shown as jitterplot for better visualization.

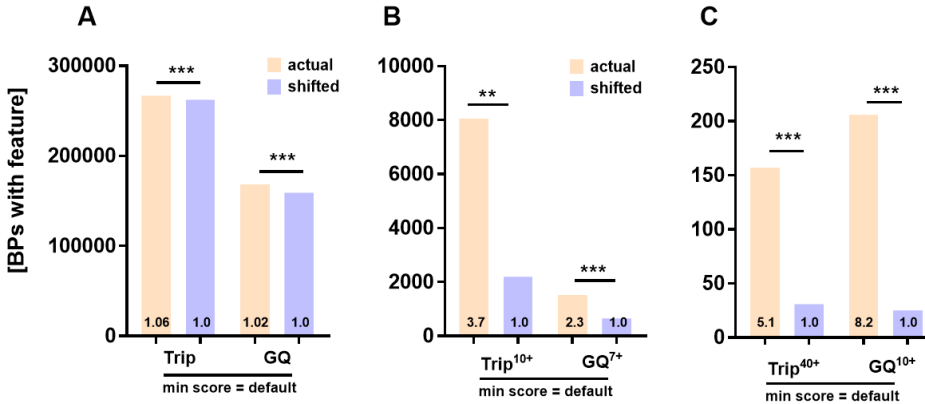


Figure S15

More actual, cancer-related breakpoints (BPs) compared to shifted control BPs are associated with triplex and G-quadruplex (GQ) motifs (A). The difference, however, is more pronounced when we consider highly unstable regions. More actual BPs compared to shifted control BPs are associated with multiple mutagenic motifs and thus lie in a highly unstable region (B, C; the number of mutagenic motifs is indicated by superscript). Each BP has a paired control and significance is determined by comparing the number of motifs per BP via Wilcoxon signed-rank test. ** $p < 0.01$; *** $p < 0.001$.

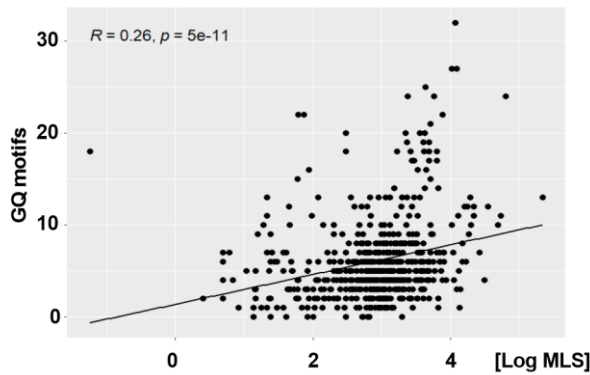


Figure S16

Across mammals, mitochondrial G-quadruplex (GQ) motifs show a positive correlation with species maximum lifespan (MLS) in an unadjusted analysis. GQ motifs were detected using default settings (min score = 47).

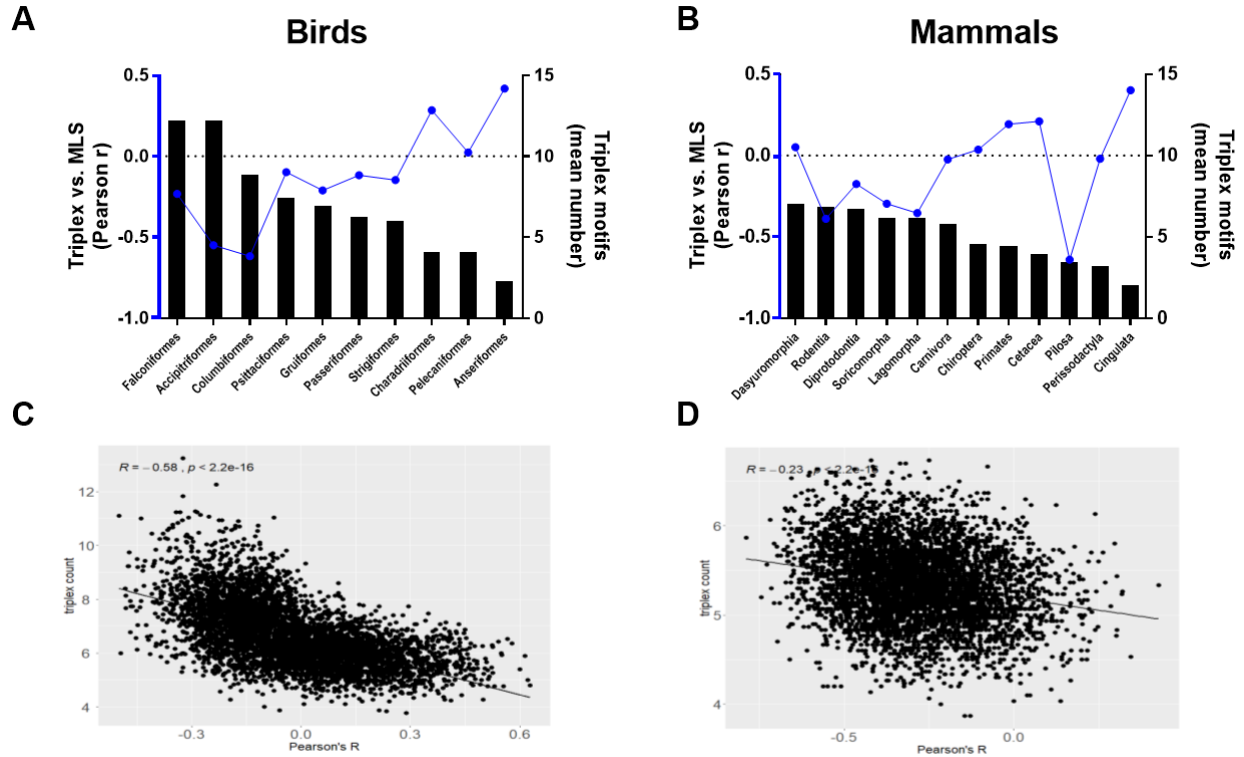


Figure S17

Phylogenetic orders (A, B) and randomly selected subgroups (C, D) of birds and mammals with a high number of triplex motifs in their mtDNA show a more robust inverse correlation between maximum lifespan (MLS) and triplex motifs, suggesting that levels of triplex motifs in mtDNA above a certain threshold could be detrimental to lifespan. Triplex motifs were detected using default settings (min score = 15).

A) The higher the mean number of triplex motifs in a bird order (bar graphs) the stronger the inverse correlation between triplex motifs and MLS in the same order (blue line). All bird orders with more than 5 species in our dataset are included in this graph.

(B) The higher the mean number of triplex motifs in a mammalian order (bar graphs) the stronger the inverse correlation between triplex motifs and MLS in the same order (blue line). All mammalian orders with more than 5 species in our dataset are included in this graph.

(C, D) We resampled 30 species from the bird (C) and mammal (D) dataset 5000 times. Then we calculated the correlation coefficient between triplex count and species MLS for each resampled subgroup. After this we plot the mean triplex count of each subgroup against the correlation coefficient of that subgroup.

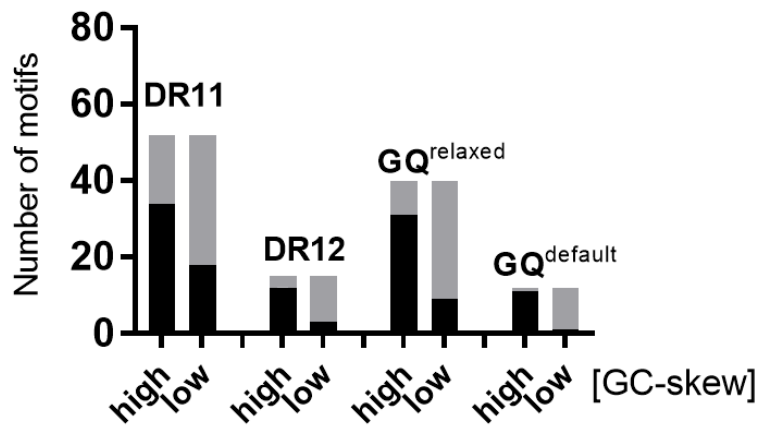


Figure S18

The human mtDNA was split into 100 bp windows and the number of direct repeat (DR) and G-quadruplex (GQ) motifs within highly and moderately GC-skewed windows is shown. Windows with a more negative skew than the median are defined as highly skewed and windows with a more positive skew are defined as low skew. We find that both DR and G-quadruplex motifs localize preferably in highly GC-skewed regions. DR11, DR12 ... 11 and 12 bp long DR motifs. The dark shaded area shows the number of motifs associated with a given window and the grey shaded area the number of motifs not associated with a given window.

SUPPLEMENTARY DATA - TABLES

Triplex package (default, min score = 15)

	start	width	score	pvalue	ins	type	s	
[1]	6676	26	17	7.3e-01	0	7	+	[TAACTTACTACTCCGGAAAAAAGAA]
[2]	6689	31	15	9.9e-01	0	6	+	[CGGAAAAAAGAACCATTGGATACATAGGT]
[3]	10925	31	16	3.4e-01	0	1	+	[TTTTCTCCGACCCCTAACAACCCCTCC]
[4]	13751	20	15	6.2e-01	0	1	+	[TTTCCCCCGCATCCCCCTTC]
[5]	14097	19	15	6.2e-01	0	0	+	[CTCCTCTCTTCTTCTTC]
[6]	14599	21	15	6.2e-01	0	2	-	[TTTTCTTCTAAGCCTTCTCT]

Triplex package (relaxed, min score = 12)

	start	width	score	pvalue	ins	type	s	
[1]	5830	25	13	1e+00	0	6	+	[AAAAAGAGGCCTAACCCCTGTCTTT]
[2]	6676	26	17	7.3e-01	0	7	+	[TAACTTACTACTCCGGAAAAAAGAA]
[3]	6689	31	15	9.9e-01	0	6	+	[CGGAAAAAAGAACCATTGGATACATAGGT]
[4]	7434	19	14	1e+00	0	7	+	[ATAAAATCTAGACAAAAAA]
[5]	7465	19	12	1e+00	0	3	-	[GAAACCAGCTTTGGGGGGT]
[6]	7515	18	13	9.9e-01	0	2	-	[TTTTCTAATACCTTTTT]
[7]	7515	18	12	1e+00	0	3	-	[TTTTCTAATACCTTTTT]
[8]	8414	18	12	1e+00	0	0	+	[CTCCTTACACTATTCCTC]
[9]	8414	18	12	1e+00	0	1	+	[CTCCTTACACTATTCCTC]
[10]	9460	25	14	1e+00	0	5	-	[AAAAAACTTCTGAGGTAATAAATA]
[11]	9477	27	14	8.9e-01	0	1	+	[GTTTTTTTCTTCGAGGATTTTTCTGA]
[12]	9478	22	13	1e+00	0	4	-	[AAAAATCCTGCGAAGAAAAAA]
[13]	10048	16	12	1e+00	0	6	+	[AAAAAAGAGTAATAAA]
[14]	10192	23	14	8.9e-01	0	0	+	[CCCCCGCCCGCTCCCTTTCTCC]
[15]	10814	26	12	1e+00	0	6	+	[AAAAAACACATAATTGAATCAACAC]

[16]	10925	26	14	8.9e-01	0	2 -	[GGGGTTGTTAGGGGGTCGGAGGAAAA]
[17]	10925	31	16	3.4e-01	0	1 +	[TTTTCTCCGACCCCCCTAACACCCCCCTCC]
[18]	10936	16	12	1e+00	0	4 -	[GGGGTTGTTAGGGGG]
[19]	11032	25	13	1e+00	0	6 +	[AAAAAACTCTACCTCTCTATACTA]
[20]	12106	29	12	1e+00	0	3 -	[AGGAAAAACCCGGTAATGATGTCGGGGTTG]
[21]	12374	17	12	1e+00	0	1 +	[CTTCCCTAATTCCCCC]
[22]	12383	26	14	8.9e-01	0	0 +	[TTCCCCCATCCTTACCACCCTCGTT]
[23]	12396	30	13	1e+00	0	7 +	[TACCACCCTCGTTAACCCCTAACAAAAAAA]
[24]	12418	24	13	1e+00	0	6 +	[AAAAAACTCATACCCCCATTAT]
[25]	13218	20	13	1e+00	0	7 +	[ACAAAATGACATCAAAAAA]
[26]	13751	20	15	6.2e-01	0	1 +	[TTCCCCCGCATCCCCCTTC]
[27]	13752	18	14	8.9e-01	0	0 +	[TTCCCCCGCATCCCCCTTC]
[28]	14097	19	15	6.2e-01	0	0 +	[CTTCTCTCTTTCTTCTTC]
[29]	14492	19	12	1e+00	0	7 +	[CCTAAATAAATTAAAAAA]
[30]	14504	27	13	1e+00	0	6 +	[AAAAAACTATTAAACCCATATAACCT]
[31]	14599	21	15	6.2e-01	0	2 -	[TTTTCTTCTAAGCCTTCTCT]
[32]	15443	18	13	9.9e-01	0	1 +	[CTTCTCTTCTTCTCTCC]
[33]	16006	24	13	1e+00	0	5 -	[AAAGAACAGAGAATAGTTTAAATT]

Non-B DNA motif search tool (nBMST)

type	start	end	width	sequence
Mirror_Repeat	10077	10115	39	ttaataatcaacaccctcctagccttactactaataatt
Mirror_Repeat	11050	11111	62	tatactaattctccctacaaatctccttaattataac(...)

Triplexator (default, max error rate 5%)

	Start	End	Score	Motif	Guanine.rate	TFO
01	6213	6232	19	Y	0.55	CTCTTACCTCCCTCTCTCCT
02	6213	6232	19	R	0.55	CTCTTACCTCCCTCTCTCCT
03	6219	6238	19	Y	0.60	CCTCCCTCTCTCCTACTCCT
04	6219	6238	19	R	0.60	CCTCCCTCTCTCCTACTCCT
05	8803	8818	16	M	0.56	ACACCAACCAACCAAC
06	10831	10850	19	M	0.45	<u>AATCAACACAACCACCCACA</u>
07	10834	10853	19	M	0.55	<u>CAACACAACCACCCACAGCC</u>
08	12540	12560	20	M	0.48	AGCCACAACCCAAACAACCCA
09	12542	12562	20	M	0.52	CCACAACCCAAACAACCCAGC
10	14092	14117	25	Y	0.42	<u>CTTTACTTCCTCTCTTTCTTCTTCCC</u>
11	14092	14117	25	R	0.42	<u>CTTTACTTCCTCTCTTTCTTCTTCCC</u>
12	14097	14121	24	Y	0.48	<u>CTTCCTCTCTTTCTTCTTCCCACTC</u>
13	14097	14121	24	R	0.48	<u>CTTCCTCTCTTTCTTCTTCCCACTC</u>
14	14327	14347	20	M	0.57	ACCACAACCACCACCCCATCA
15	14613	14633	20	M	0.43	<u>AAGAAAACCCCAACACCCCA</u>
16	15439	15462	23	Y	0.42	CTTACTTCTCTTCTTCTCTCTT
17	15439	15462	23	R	0.42	CTTACTTCTCTTCTTCTCTCTT
18	16277	16296	19	M	0.55	ACCAACAAACCTACCCACCC

Triplexator (relaxed, max error rate 7.5%)

	Start	End	Score	Motif	Guanine.rate	TFO
01	5931	5946	15	M	0.31	ACAAACCACAAAGACA
02	6213	6232	19	Y	0.55	CTCTTACCTCCCTCTCTCCT
03	6213	6232	19	R	0.55	CTCTTACCTCCCTCTCTCCT
04	6219	6238	19	Y	0.60	CCTCCCTCTCTCCTACTCCT
05	6219	6238	19	R	0.60	CCTCCCTCTCTCCTACTCCT
06	6486	6502	16	Y	0.53	CTTCTCCTATCTCTCCC
07	6486	6502	16	R	0.53	CTTCTCCTATCTCTCCC
08	6542	6557	15	M	0.56	CAACCTCAACACCACC
09	7397	7415	18	M	0.68	CCCCCACCCTACCACACA
10	7443	7460	17	R	0.28	<u>AGACAAAAAAGGAAGGAA</u>
11	7443	7460	17	Y	0.28	<u>AGACAAAAAAGGAAGGAA</u>
12	7717	7733	16	M	0.35	AACACTCACAAACAAAC

13	8272	8288	16	Y	0.71	CCCCCTCTACCCCCCTCT
14	8272	8288	16	R	0.71	CCCCCTCTACCCCCCTCT
15	8452	8468	16	M	0.41	AAACACAAACTACCACC
16	8597	8612	15	Y	0.44	TTCTATTTCCCCCTCT
17	8597	8612	15	R	0.44	TTCTATTTCCCCCTCT
18	8652	8667	15	M	0.44	AATCACCACCCAACAA
19	8803	8820	17	M	0.50	ACACCAACCACCCAACCTA
20	9350	9366	16	M	0.41	AACCAACACACTAACCA
21	9664	9681	17	M	0.28	AAAACAACCGAAACCAAA
22	10267	10282	15	Y	0.56	CCCTCCTTTTACCCCT
23	10267	10282	15	R	0.56	CCCTCCTTTTACCCCT
24	10610	10625	15	M	0.56	AACCTCAACACCCAC
25	10831	10850	19	M	0.45	<u>AATCAACACAACCACCCACA</u>
26	10834	10853	19	M	0.55	<u>CAACACAACCACCCACAGCC</u>
27	10886	10904	18	M	0.37	AACCAAATCAACAACAACC
28	10935	10952	17	M	0.67	<u>ACCCCTAACAACCCCCC</u>
29	11227	11242	15	Y	0.62	CTCCCTTCCCCTACTC
30	11227	11242	15	R	0.62	CTCCCTTCCCCTACTC
31	12083	12099	16	Y	0.59	TCCCCCATTCTCCTCCT
32	12083	12099	16	R	0.59	TCCCCCATTCTCCTCCT
33	12090	12106	16	Y	0.53	<u>TTCTCCTCCTATCCCTC</u>
34	12090	12106	16	R	0.53	<u>TTCTCCTCCTATCCCTC</u>
35	12409	12426	17	M	0.28	<u>AACCCTAACAACAAAAAC</u>
36	12540	12560	20	M	0.48	AGCCACAACCCAAACAACCCA
37	12542	12562	20	M	0.52	CCACAACCCAAACAACCCAGC
38	13290	13306	16	M	0.53	CATCAACCAACCACACC
39	13293	13308	15	M	0.50	CAACCAACCACACCTA
40	13770	13786	16	M	0.53	<u>CAAACAACAATCCCCC</u>
41	14064	14079	15	M	0.44	CACCTCAACCCCAAAAA
42	14092	14121	28	Y	0.43	<u>CTTTACTTCCTCTCTTTCTTCTTCCCACTC</u>
43	14092	14121	28	R	0.43	<u>CTTTACTTCCTCTCTTTCTTCTTCCCACTC</u>
44	14097	14126	28	Y	0.47	<u>CTTCCTCTCTTTCTTCTTCCCACTCATCCT</u>
45	14097	14126	28	R	0.47	<u>CTTCCTCTCTTTCTTCTTCCCACTCATCCT</u>
46	14181	14196	15	M	0.31	ATACACCAACAACAA
47	14327	14347	20	M	0.57	ACCACAACCACCCCATCA
48	14387	14402	15	M	0.44	AACCCCACTAAAACAC
49	14551	14568	17	M	0.56	AACACACCCGACCACACC
50	14613	14633	20	M	0.43	<u>AAGAAAACCCCAACAAACCCA</u>
51	14639	14654	15	M	0.44	AAACCCCACTCAACA
52	14809	14825	16	M	0.65	CCCCACCCCATCAAAA
53	15439	15462	23	Y	0.42	<u>CTTACTTCTCTTCTTCTCTCTCTT</u>
54	15439	15462	23	R	0.42	CTTACTTCTCTTCTTCTCTCTCTT
55	15532	15548	16	M	0.59	AAACACCCCTCCCCACA
56	16162	16188	25	M	0.44	AAAAACCCAATCCACATCAAAACCCCC
57	16179	16194	15	M	0.62	CAAAACCCCTCCCCA
58	16277	16296	19	M	0.55	ACCAACAAACCTACCCACCC

Table S1. Triplex motifs detected by different motif prediction tools

List of triplex motifs within the mtDNA major arc detected by the triplex package, the non-B DNA motif search tool (nBMST) and Triplexator. Before exclusion of redundant and overlapping motifs. Motifs found by Triplexator that overlap with motifs from the triplex package are underlined. The strand (s) is indicated for motifs found by the triplex package. Plus strand corresponds to the light-strand of mtDNA and minus strand to the heavy-strand.

Motif	%error rate	window size (bps)				
		20	30	40	50	60

triplexator	5	1.50	1.86	1.31	1.33	1.24
triplexator	7.5	1.53	1.45	1.21	1.08	1.09
triplexator	10	1.10	1.33	1.27	1.13	1.10
triplexator	15	1.23	1.17	1.10	1.08	1.07
rmGQ	5	3.00	4.25	2.26	2.00	1.90

Motif	Stringency	20	30	40	50	60
triplex	default	6.00	1.92	1.62	1.73	1.65
triplex	relaxed	1.75	2.35	1.68	1.89	1.72
rmGQ	default	1.00	1.12	1.28	1.63	1.45
rmGQ	relaxed	2.00	3.14	2.05	2.21	1.84

Motif	Stringency	20	30	40	50	60
GQ	default	6.33	6.29	2.52	1.80	1.61
GQ	relaxed	2.52	1.80	1.51	1.29	1.25

Table S2. Comparison of Triplexator, triplex package and pqsfinder data (fold-enrichment vs. control)

This table shows the fold-enrichment of triplex motifs detected using two different methods (Triplexator and triplex package), and of G-quadruplex (GQ) motifs detected by the pqsfinder package, around actual deletion breakpoints (BPs) compared to a shifted control, where each BP was shifted by 200 bp towards the midpoint of the major arc. The acceptable maximal error rate (%) in Triplexator dictates the stringency of detection (higher = relaxed stringency). Analysis based on MitoBreak BPs. rmGQ, overlapping GQ motifs removed.

Pqsfinder (default, min score = 47)

	start	width	score	sequence
[1]	6290	30	52	[CCCTCCCTTAGCAGGGAACACTCCCACCC]
[2]	7397	16	53	[CCCCCACCCTACCAC]
[3]	7807	20	51	[CCTCATCGCCCTCCCATCCC]
[4]	8262	34	67	[CCCTATAGCACCCCTCTACCCCTCTAGAGCCC]
[5]	9243	28	55	[CCCAGCCCATGACCCCTAACAGGGGCCC]
[6]	9526	49	85	[CCCCACCCCTTACTAACATTAACGAAAATAACCCCAACAGGCATCA...]
[7]	10184	24	60	[CCCTATATCCCCCGCCCGCTCCC]
[8]	10918	34	92	[CCCAACCTTTTCTCCGACCCCTAACAACCCCC]
[9]	12084	32	67	[CCCCATTCTCCTCCTATCCCTCAACCCCGAC]
[10]	12359	45	78	[CCACCCTAACCCCTGACTTCCCTAATTCCCCCATCCTTACCACCC]
[11]	13026	36	90	[CCCCTGACTCCCCTCAGCCATAGAAGGCCCCACCCC]
[12]	13647	49	62	[CCCCACCCTTACTAACATTAACGAAAATAACCCCAACCCCTACTAAAC...]
[13]	13755	31	61	[CCCCGCATCCCCCTTCCAAACAACAATCCCC]
[14]	14245	40	54	[CCCCGCACCAATAGGATCCTCCCGAATCAACCCTGACCCC]
[15]	14389	40	69	[CCCCACTAAAACACTACCAAGACCTCAACCCCTGACCCC]
[16]	14771	47	49	[CCCCTAATAAAATTAATTAACCACTCATTCATCGACCTCCCCACCCC]

[17]	15526	31	83	[CCCCCTTAAACACCCCTCCCCACATCAAGCCC]
[18]	16159	35	51	[CATAAAAACCCAATCCACATCAAAACCCCCCTCCCC]
[19]	16353	28	55	[CCCTTCTCGTCCCCATGGATGACCCCCC]

Pqsfinder (relaxed, min score = 26)

	start	width	score	sequence
[1]	6154	35	34	[CCCTAATAATCGGTGCCCCGATATGGCGTTTCCC]
[2]	6257	15	27	[GGAGGCCGGAGCAGG]
[3]	6290	30	52	[CCCTCCCTTAGCAGGGAACCTACTCCACCC]
[4]	6421	48	43	[CCCCTGCCATAACCCAATACCAAACGCCCCCTCTTCGTCTGATCCG...]
[5]	6539	49	40	[CCGCAACCTCAACACCACCTTCTTCGACCCCGCCGGAGGAGGAGAC...]
[6]	7095	46	29	[CCCCTATTCTCAGGCTACACCCTAGACCAAACCTACGCCAAATCC]
[7]	7199	49	29	[CCTATCCGGAATGCCCCGACGTTACTCGGACTACCCCCGATGCATAC...]
[8]	7397	16	53	[CCCCCACCCCTACCAC]
[9]	7466	28	38	[CCCCCAAAGCTGTTTTCAAGCCAACCC]
[10]	7807	20	51	[CCTCATCGCCCTCCCATCCC]
[11]	8262	34	67	[CCCTATAGCACCCCTCTACCCCTCTAGAGCCC]
[12]	8369	41	42	[CCCCAACTAAATACTACCGTATGGCCCACCATAATTACCCC]
[13]	8464	12	32	[CCACCTACCTCC]
[14]	8559	50	41	[CCCCACAATCCTAGGCCTACCCGCCGAGTACTGATCATTCTATTT...]
[15]	8619	10	35	[CCCCACCTCC]
[16]	8806	42	35	[CCAACCAACCAACTATCTATAAACCTAGCCATGGCCATCCCC]
[17]	8914	30	41	[CCACAAGGCACACCTACACCCCTTATCCCC]
[18]	9243	28	55	[CCCAGCCCATGACCCCTAACAGGGGGCCC]
[19]	9289	14	29	[CCTCCGGCCTAGCC]
[20]	9413	15	27	[CCACCACACACCACC]
[21]	9526	49	85	[CCCCTACCCCCAATTAGGAGGGCACTGGCCCCCAACAGGCATCAC...]
[22]	10184	24	60	[CCCTATATCCCCCGCCCCGCTCCC]
[23]	10267	27	42	[CCCTCCTTTTACCCCTACCATGAGCCC]
[24]	10612	26	42	[CCCTCAACACCCACTCCCTCTTAGCC]
[25]	10918	34	92	[CCCAACCTTTTCTCCGACCCCTAACAACCCCC]
[26]	10968	15	27	[CCTGACTCCTACCCC]
[27]	11130	33	26	[CCACACTTATCCCCACCTTGGCTATCATCACCC]
[28]	11206	31	39	[CCTATTCTACACCTAGTAGGCTCCCTTCCC]
[29]	11407	25	43	[CCCTAAAGCCCATGTGAAGCCCCC]
[30]	11532	12	32	[CCTACCCCTTCC]
[31]	11848	27	35	[CCTCGCTAACCTCGCCTTACCCCCCAC]
[32]	12084	32	67	[CCCCATTCTCCTCCTATCCCTCAACCCCGAC]
[33]	12359	45	78	[CCACCCTAACCTGACTTCCCTAATCCCCCATCCTTACCACCC]
[34]	12542	27	37	[CCACAACCAACCAACCCAGCTCTCCC]
[35]	12927	45	33	[CCCACAACAAATAGCCCTTCTAAACGCTAATCCAAGCCTCACCCC]
[36]	13026	36	90	[CCCCTGACTCCCTCAGCCATAGAAGGCCCCACCCC]
[37]	13119	13	30	[CCGCTTCCACCCC]
[38]	13647	49	62	[CCCCACCCTTACTAACATTAACGAAAATAACCCACCCCTACTAAAC...]
[39]	13755	31	61	[CCCCGCATCCCCCTTCCAAACAACAATCCCC]
[40]	13947	47	42	[CCCCTATCTAGGCCTTCTTACGAGCCAAAACCTGCCCTACTCCTCC]
[41]	14043	16	26	[CCAAATCTCCACCTCC]
[42]	14115	23	28	[CCCACTCATCCTAACCTACTCC]
[43]	14245	40	54	[CCCCGCACCAATAGGATCCTCCCGAATCAACCCCTGACCCC]
[44]	14334	40	37	[CCACCACCCCATCATACTCTTTCACCCACAGCACCAATCC]
[45]	14389	40	69	[CCCCATAAAACACTACCAAGACCTCAACCCCTGACCCC]
[46]	14490	46	37	[CCCCTAAATAAATTAAAAAACTATTAAACCCATATAACCTCCCCC]
[47]	14620	25	31	[CCCCACAAACCCCATTAATAAACCC]
[48]	14771	47	49	[CCCCTAATAAAATTAATTAACCACTCATTATCGACCTCCCCACCCC]
[49]	14862	15	27	[CCTGCCTGATCCTCC]
[50]	15263	42	26	[CCCACCCTCACACGATTCTTTACCTTTCACTTCATCTTGCCC]
[51]	15368	48	28	[CCCCTAGGAATCACCTCCATTCCGATAAAATCACCTTCCACCCT...]
[52]	15526	31	83	[CCCCCTTAAACACCCCTCCCCACATCAAGCCC]
[53]	15658	11	33	[CCCCATCCTCC]
[54]	16159	35	51	[CATAAAAACCCAATCCACATCAAAACCCCCCTCCCC]
[55]	16260	37	44	[CCCTCACCCACTAGGATACCAACAAACCTACCCACCC]
[56]	16353	28	55	[CCCTTCTCGTCCCCATGGATGACCCCCC]

[57] 16400 12 32 [CCACCATCCTCC]

Table S3. G-quadruplex motifs in the major arc of mtDNA

List of G-quadruplex (GQ) motifs detected by pqsfinder. All GQ motifs but one were found on the heavy-strand (motif #2).

Motif	DR ⁺	DR ⁻	ratio (DR ⁺ vs DR ⁻)	p-value
GQ ⁺ (default)	82	42	2.0	p<0.05
Triplex ⁺ (default)	28	30	0.9	
GQ ⁺ (relaxed)	189	114	1.7	p<0.0001
Triplex ⁺ (relaxed)	86	119	0.7	

Table S4. Colocalization of G-quadruplex and triplex motifs with DR motifs

G-quadruplex (GQ) motifs preferably colocalize with deletions that are associated with direct repeat motifs (DR), whereas triplex motifs do not. Significance based on Fisher's exact test. Deletion data from the MitoBreak database.

	5'-Breakpoints		3'-Breakpoints		deletion		
	median	mean	median	mean	median size (bp)	major arc deletions (N)	subjects (N)
MitoBreak	7962	8077	15149	14741	7122	1066	NA
Persson	8396	8708	15063	14571	6125	1114	5
Hjelm	8106	8446	14341	14075	5926	1894	93

Table S5. Comparison of mtDNA deletion breakpoint datasets

Breakpoint positions and other characteristics of the three datasets we used (MitoBreak, Persson et al. 2019 and Hjelm et al 2019).

Motif	Type	Mammals		Birds		Ray-finned fishes	
		Raw	Adjusted	Raw	Adjusted	Raw	Adjusted
DR11	11bp	<u>-0.113</u>	0.055	0.090	<u>0.140</u>	0.117	-0.026
MR11	11bp	<u>-0.155</u>	-0.002	0.037	0.115	0.065	-0.003
IR11	11bp	<u>-0.336</u>	<u>0.105</u>	-0.125	-0.016	0.086	<u>0.250</u>
ER11	11bp	<u>-0.356</u>	-0.047	-0.073	0.002	-0.020	0.087

DR	9 to 15bp	<u>-0.149</u>	<u>0.127</u>	0.059	0.107	<u>0.231</u>	<u>0.212</u>
MR	9 to 15bp	<u>-0.255</u>	-0.025^	0.068	<u>0.167</u>	0.140	0.053
IR	9 to 15bp	<u>-0.369</u>	-0.058	-0.012	0.140	-0.032	0.113
ER	9 to 15bp	<u>-0.384</u>	<u>0.134</u>	-0.074	0.052	-0.093	<u>0.212</u>
triplex	default	<u>-0.296</u>	<u>-0.211**</u>	-0.073	0.003	-0.083	0.040
triplex	relaxed	<u>-0.190</u>	<u>-0.127^</u>	-0.012	0.077	0.091	<u>0.354**</u>
GQ	default	<u>0.264</u>	0.068	0.108	<u>0.174</u>	0.017	<u>0.292^</u>
GQ	relaxed	<u>0.283</u>	-0.097**	0.042	0.119	-0.020	<u>0.269</u>

Table S6. Correlation between motifs and MLS in birds and ray-finned fishes (actinopterygii)

The adjusted model takes into account GC content, GC skew, AT skew and number of effective codons. Significant correlations in the raw or adjusted model are bolded/underlined ($p < 0.05$). The PGLS model additionally considers phylogeny. ^denotes p-values of $0.05 < p < 0.10$ in the PGLS model and ** p-values of $p < 0.05$. The table shows Pearson's R.

		non-D loop mtDNA		major arc mtDNA	
Motif	Type	Raw	Adjusted	Raw	Adjusted
DR11	11bp	<u>-0.113</u>	0.055	<u>-0.162</u>	0.052
MR11	11bp	<u>-0.155</u>	-0.002	<u>-0.126</u>	-0.006
IR11	11bp	<u>-0.336</u>	<u>0.105</u>	<u>-0.314</u>	0.043
ER11	11bp	<u>-0.356</u>	-0.047	<u>-0.313</u>	-0.046
triplex	default	<u>-0.296</u>	<u>-0.211**</u>	<u>-0.240</u>	<u>-0.127^</u>
triplex	relaxed	<u>-0.190</u>	<u>-0.127^</u>	<u>-0.135</u>	-0.059
GQ	default	<u>0.264</u>	0.068	<u>0.263</u>	0.057
GQ	relaxed	<u>0.283</u>	-0.097**	<u>0.278</u>	<u>-0.107**</u>

Table S7. Correlation between potentially mutagenic motifs in the mtDNA and species lifespan

The adjusted model takes into account body mass, GC content, GC skew, AT skew and number of effective codons. Significant correlations in the raw and adjusted model are bolded/underlined ($p < 0.05$). The PGLS model additionally considers phylogeny. ^denotes p -values of $0.05 < p < 0.10$ in the PGLS model and ** p -values of $p < 0.05$. The table shows Pearson's R .

	Triplex motifs			GQ motifs		
	actual	control	fold-enrichment	actual	control	fold-enrichment
mean (motifs/BP)	0.948	0.674	1.41	0.822	0.785	1.05
pseudo-median	2	1.999942	NA	1.500065	1.500044	NA
CI max	2.000036	1.999936	NA	1.500029	1.500063	NA
CI min	2.000002	1.999948	NA	1.500006	1.50004	NA
Total BPs	577994	577994	NA	577994	577994	NA
BP's with X motifs:						
0	408279	418334	0.98	311129	315643	0.99
1	77810	79537	0.98	143692	144915	0.99
2	33749	34258	0.99	72928	70344	1.04
3	15129	14471	1.05	30653	29684	1.03
4	11192	10262	1.09	11909	11294	1.05

5	9137	7736	1.18	4351	4032	1.08
6	6108	4847	1.26	1656	1367	1.21
7	3706	2889	1.28	635	385	1.65
8	2153	1775	1.21	434	144	3.01
9	1564	1044	1.50	253	103	2.46
10+	8058	2197	3.67	206	25	8.24

Table S8. Triplex and G-quadruplex motifs are enriched at actual breakpoints

Although both triplex and G-quadruplex (GQ) motifs are enriched around actual, cancer-associated breakpoints (BPs) compared to control BPs, the difference is more pronounced for triplex motifs on average. Most BPs were not associated with any motif and for both GQ and triplex motifs the results are strongest in the subset of BPs associated with multiple motifs. The pseudo-median and confidence interval (CI) was calculated by Wilcoxon rank sum test in R.

triplex min score=12				
Species	Subgroup	Triplex (count)	Correlation vs MLS	Sample (N)
mammals	low	4.2	<u>0.217</u>	230
mammals	high	6.1	<u>-0.444</u>	172
birds	low	5.2	-0.136	123
birds	high	9.1	<u>-0.239</u>	68
fish	low	6.3	<u>0.459</u>	59
fish	high	9.3	-0.124	78

triplex min score=15				
Species	Subgroup	Triplex (count)	Correlation vs MLS	Sample (N)
mammals	low	4.2	0.046	230
mammals	high	6.1	<u>-0.280</u>	172
birds	low	5.2	-0.142	123
birds	high	9.1	<u>-0.384</u>	68
fish	low	6.3	-0.183	59
fish	high	9.3	-0.064	78

Table S9. Correlation between triplex motifs and MLS is strongest in species with many triplex motifs in their mtDNA

Triplex (count) refers to the mean number of mtDNA triplex motifs in the corresponding subgroup. Each “low” subgroup is composed of orders with triplex counts below the median for the whole phylogenetic class, and the “high” group of orders with triplex counts above the median. When we pool data from mammalian orders with fewer triplex motifs than the median for all mammals (n=172 species) we find an inverse relationship with maximum lifespan (MLS) and the same is true for birds. Significant correlations are bolded and underlined ($p < 0.05$).

Motif	IR ⁺	IR ⁻	ratio (IR ⁺ vs IR ⁻)	p-value
ER ⁺ (6-15bp)	18	53	0.33	
GQ ⁺ (default)	8	106	0.07	$p < 0.001$
DR ⁺ (6-15bp)	39	418	0.09	$p < 0.001$

Table S10. Colocalization of ER, GQ and DR motifs

Everted repeat (ER) motifs preferably colocalize with inverted repeat (IR) motifs. Neither G-quadruplex (GQ) nor direct repeat (DR) motifs show such an enrichment around IR motifs. Significance based on Fisher’s exact test. Deletion data from the MitoBreak database.