Title

Neither wood ash application nor phosphorus-fertilisation mitigated the negative effects of whole-tree harvesting on a temperate oligotrophic forest.

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Abstract

Background and Aims: Concerns about climate change and carbon economy have prompted the promotion of alternative energy sources, including forest-based bioenergy. An evaluation of the environmental consequences of intensive harvests (stumps and roots, and also branches and foliage) for energy wood supply, and use of wood-ash recycling as a compensatory practice, helps in the analysis of the use of forest biomass for energy production.

Methods: After 11 years, we made use of records from a split-plot experimental site crossing four different intensities of Biomass Harvesting (Stem-Only Harvest [SOH], Aboveground Additional Harvest [AAH], Belowground Additional Harvest [BAH], and Whole-Tree Harvest [WTH]) and three Compensation Methods (control [C], wood ash application [A] and phosphorus fertilisation [P]) to evaluate their effects on tree growth, soil fertility, chemical properties and soil carbon. This site is located in a maritime pine forest on a poor soil, under a warm temperate climate (SW France).

Key results: Despite their low additional biomass exports (+10% for AAH to +34% for WTH), the non-conventional harvest practices exported much higher quantities of nutrients than the conventional SOH technique (for example +145% for N and K in WTH). Consequently, these treatments had several effects on the soil nutritive status. Additional biomass harvests impacted the soil organic matter content, with negative effects on P_{organic} and soil cation exchange capacity. However, data suggested that tree growth and foliage nutrient content had not yet been significantly impacted by harvest treatments, whereas tree nutritional status was improved by P-fertiliser or wood ash. As expected, we observed a positive effect of wood ash application on soil acidity and nutrient content but, like additional harvests, wood ash application decreased the pool of soil organic carbon (~10% of the initial stock with ~7% of N_{total} losses).

Conclusions: Overall, this factorial experiment showed that exporting more forest biomass due to the additional harvesting of tree canopies, stumps and roots had negative consequences on the ecosystem. Additional harvests have aggravated the poverty of the already oligotrophic soil, which in turn may decrease tree growth and the soil organic carbon content in the future (but without any impact on either soil acidity or on trace metal contents). Importantly, applying nutrients as fertiliser or wood ash did not compensate for the negative impact of biomass exports and the method of wood ash recycling in forests could even decrease the soil organic carbon.

Introduction

Most recent reports of the Intergovernmental Panel on Climate Change have concluded that climate change is accelerating and requires rapid mitigation policies (IPCC 2014). These policies rely on two main and complementary approaches: the emission of less greenhouse gases (GHG), and the sequestration of more carbon dioxide (CO₂) from the atmosphere. Forest ecosystems can do both, depending on whether they are disturbed or not (Thom and Seidl 2016). Due to their large carbon contents (Pan et al. 2011), and because they represent a large proportion of land (Bastin et al. 2017), forests are pivotal ecosystems with respect to the Earth's carbon cycle. However, although there is a consensus that forests constitute an important leverage for climate change mitigation (IPCC 2014), there is a heated debate on how forests should be managed for mitigation purposes (Lindner and Karjalainen 2007): the *old-growth* strategy consisting of sequestering carbon in the ecosystem, conflicts with the *intensive biomass harvest* strategy, which proposes to harvest more woody biomass to replace fossil C in the production of manufactured objects and energy (Bright et al. 2012; Schulze et al. 2012; Haberl et al. 2013; Luyssaert et al. 2018). In particular, additional harvesting of forest biomass, such as foliage, branches, stumps, and roots (Nicholls et al. 2009), for energy production can impoverish forest ecosystems because these tree compartments dedicated to energy wood supply –the so-called *harvest residues* or *logging residues*– are rich in nutrients (Andre et al. 2010; Achat et al. 2015a; Augusto et al. 2015a). Review studies indicate that intensive biomass harvesting –as compared with conventional harvesting– can have negative consequences on ecosystem functioning, such as soil nutrients and organic matter pools, and consequently on the level of soil productivity for following forest establishment (Thiffault et al. 2011; Wall 2012; Achat et al. 2015a). Extracting more biomass from forests without any compensatory fertilisation seems to be an unsustainable leverage for climate change mitigation (Garcia et al. 2018). Therefore, it has been proposed that intensive harvests should be associated with compensatory practices (Nohrstedt 2001; Ranius et al. 2018; Ventura et al. 2019). Applying wood ash is often promoted as a good method to compensate for the negative effects of intensive harvests on the ecosystem nutrient budget (Hannam et al. 2018; Ranius et al. 2018; Ventura et al. 2019). Indeed, wood ash has a high nutrient content (Aronsson and Ekelund 2004), can reduce soil acidity (Reid and Watmough 2014) and contributes to the concept of *circular bioeconomy* as wood ash

returns some of the nutrients that were exported by the removal of logging residues (Pitman 2006). On the other hand, applying wood ash might contaminate ecosystems because of its relatively high content in several micronutrients or non-essential metals (Vance 1996; Nnadi et al. 2019). However, other studies conclude that, based on the metal content of wood ash, a single application per forest rotation may not result in a significant increase in soil metal content and might even improve the micronutrient status of trees (Pitman 2006). Unfortunately, while several meta-analyses have been carried out on intensive biomass harvests (Thiffault et al. 2011; Achat et al. 2015a) or wood ash application (Augusto et al. 2008a; Reid and Watmough 2014), a combination of these two practices at the time of clear-cutting and prior to reforestation have rarely been studied together to assess their interactive consequences (Hagerberg and Wallander 2002; Wang et al. 2010). Therefore, we conducted a field experiment whose main objective was to evaluate their interactions.

Located in south-western France, the *Landes de Gascogne* forest, where our study took place, has several advantages for evaluating the environmental consequences of intensive harvests and their compensatory practices. Firstly, it is a large man-made pine forest almost entirely dedicated to intensive forestry. As such, the area is already subjected to collection of harvest residues (mainly stumps and roots, and to a lesser extent also branches and needles (Mora et al. 2014; Banos and Dehez 2017)) for energy wood consumption (Augusto et al. 2010, 2015a), which in turn produces large amounts of wood ash (Alvarez-Alvarez et al. 2018). Secondly, because this forest is strongly oligotrophic (Augusto et al. 2010), it has been diagnosed to be particularly sensitive to the potential negative consequences of intensive harvest (Durante et al. 2019). Finally, it is a warm-temperate forest. Indeed, knowledge about wood ash application and removal of logging residues is based on studies carried out in boreal forests (Augusto et al. 2008a; Walmsley and Godbold 2010), whereas the consequences of these management practices are probably climate-dependent. For instance, Achat *et al.* (2015b) found that exporting tree canopies generally decreases the pool of soil organic carbon (SOC) under temperate climates, but not under cold climates. Similarly, several Nordic studies indicated that stump harvests had no effect on SOC while studies conducted in warmer latitudes concluded that the impact was highly negative (Zabowski et al. 2008; Stromgren et al. 2013; Jurevics et al. 2016).

Our initial expectations were based on the current knowledge about intensive biomass harvest or wood ash application, in comparison with the local conditions of climate, soils, and forestry. More explicitly, we expected the following effects (Table 1).

Hypothesis H1: the soil phosphorus, the most limiting nutrient in the sandy acidic soils of the study region (see Materials and Methods), would be decreased by intensive harvests (Achat et al. 2015a) and would be only partly compensated for wood ash by which has a low phosphorus content and low bioavailability (Steenari and Lindqvist 1997; Fransson et al. 1999).

H2: the sequestration rate of C into forest biomass (estimated as the growth rate of the subsequent forest stand) would be decreased by intensive harvests because the local soils are nutrient poor (Achat et al. 2009, 2015a), and would not be compensated for wood ash due to its low value as a P-fertiliser (see H1; Table 1).

H3: the size of the soil organic matter pool (and related organic carbon and nitrogen (N)) would be decreased by intensive harvests as for most temperate forests (Achat et al. 2015a, b), but might remain unaffected by wood ash application as observed in boreal forests (Augusto et al. 2008a).

H4: the soil would be acidified by intensive harvest, but neutralised by wood ash (Reid and Watmough 2014; Achat et al. 2015a). The local soils are already quite acidic (Augusto et al. 2010), and we expected the liming effect to prevail over the acidifying effect. The durability of neutralisation after a single ash input in this type of forest remains to be determined.

H5: the trace metal content in the ecosystem, and its bioavailability, would not be affected either by biomass harvests or by wood ash application. These expectations were based on two rationales. Firstly, the local pine trees contain moderate amounts of trace metals in their biomass (Saur et al. 1992; Trichet et al. 2018), which implies moderate exports through biomass harvest, and small returns through the dose of wood ash applied in our field trial. This assumption is in line with current knowledge, mostly based on Nordic podzols. Secondly, as the application of wood ash is expected to induce an increase in pH, the bioavailability of metals might decrease (Cappuyns and Swennen 2008).

Materials and Methods

Study region and experimental site

The *Landes de Gascogne* area is a flat region ~1.3 Mha in south-western France (between 43.5 and 45.5°N, and 1.5°W and 0.3°E). The climate is oceanic, usually with a dry season in summer and a wet season in winter and/or in spring (annual rainfall is around 900 mm per year, with a North-South gradient). Soils are sandy podzols (9–14% fine sands; 80–89% coarse sands), which are classified as more or less hydromorphic and humic (an “entic podzol” in the WRB soil classification), or with a hardpan (an “ortsteinic podzol”; (Augusto et al. 2010)). They developed in Aeolian deposits of quaternary sands (the *Sables des Landes*) and are characterized by their coarse particle size and their high quartz content (Bertran et al. 2011). Soils are acidic and poor in nutrients, particularly in phosphorus (P) and they have been identified as being among the poorest soils in P in the world (in this region = 60 µg-P_{total} g⁻¹, and a global mean value ≈ 500 µg-P_{total} g⁻¹; (Achat et al. 2009; Bredoire et al. 2016)). The vegetation was formally a mosaic of heathlands and wetlands, probably derived from human-degraded forests (Jolivet et al. 2007). During the 18th and 19th centuries, the French central government promoted the installation of big farms, most of which failed to survive due to the harsh local conditions (poor acidic soils, and extremely low soil water contents in summer alternating with periods of waterlogging). From 1857, a forest of ~1.0 Mha forest was progressively created thanks to a law inciting the sowing of a local species, the Maritime pine (*Pinus pinaster* Ait.), combined with the use of ditches for drainage. Initially grown for its resin, Maritime pine is now used for wood production and is often P-fertilised at plantation to alleviate the main nutritional limitation of the region (Trichet et al. 2009). As a result of two storms (in 1999 and 2009), with the development of stump removal practices and an incentive policy for biomass power plant installation, the practice of collecting *harvest residues* as biomass-based energy sources is currently spreading (Banos and Dehez 2017).

The site used for this study is called the *Forêt du Nezer* (44.57°N, 1.05°W; elevation = 21 m asl; slope < 1%; soil type predominantly an entic podzol). Its name came from Daniel Nezer, a Swiss banker who initiated the first attempt to develop a large agricultural complex in 1766 (the *Colonie de Nezer*), but which eventually ruined him (Sargos 1997). After this short farming period, the site returned to heathlands before becoming a pine forest during the second half of the 19th century (see above).

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164 *Experimental design and field procedures*

165 The experimental site is a split-plot factorial design, crossing four different intensities of *Biomass*
 166 *Harvesting* (BH factor in main plots) and four *Compensation Methods* by applying wood ash or
 167 phosphorus fertilisation (CM factor in subplots nested in BH plots), resulting in sixteen treatments
 168 replicated in three blocks (Figure S1). Contrary to the CM treatments, the BH treatments had to be
 169 applied to large plots (main plots) because they required the use of large machines.

170 After the clear-cut of a 45-year-old stand in April 2007, twelve large square plots, 100 × 100 m (1 ha
 171 large), were established in order to apply the three blocks × four *Biomass Harvesting* treatments,
 172 described below:

173 1- Stem-Only Harvest ('SOH'; stem harvest down to 7 cm of diameter above bark) without any residue
 174 harvest (exports = stems: this has been the local conventional practice for decades),

175 2- Aboveground Additional Harvest ('AAH'), which included stem harvest with additional harvesting
 176 of branches with part of their foliage (see below; exports = stems + canopies; canopy = branches +
 177 twigs + foliage),

178 3- Belowground Additional Harvest ('BAH'), which included stem (without branches, twigs, and
 179 foliage) harvest with additional harvesting of stumps and roots (exports = stems + stumps + roots),

180 4- Whole-Tree Harvest ('WTH') with additional harvest of all other tree compartments (exports = stems
 181 + branches + twigs + foliage + stumps + roots).

182 The conventional harvesting (clear-cut) was carried out in April 2007, with particular attention to the
 183 slash (branches and needles) which was gathered in each of the four 1 ha plots (Figure S2a). In June
 184 2007, the branches were bundled, forwarded from the six main plots (AAH-WTH
 185 plots × 3 blocks; Figure S2b-d), and finally transported to the power plant. The time-lapse between this
 186 harvest and the clear-cut has enabled a partial fall of the foliage (needles, in unquantified proportions).
 187 Then, in July-August 2007, the stumps were excavated with their main roots, fragmented into a few
 188 pieces, and left on the ground in the six main plots (BAH-WTH plots × 3 blocks; Figure S2e-g),
 189 enabling the natural removal of the soil particles by the rain. These fragments were harvested in February
 190 2008. Two years after clear-cutting, in January 2009, the area was strip ploughed to a depth of 0.3 m,

and was replanted with 1,250 per hectare (4×2 m) of one-year-old maritime pine seedlings of local origin.

Forty-eight small sub-plots, 50×50 m (0.25 ha) wide, were established within the two-year-old plantation. Four sub-plots of compensation methods (CM factor) were nested in every main plot (dedicated to a BH treatment; Figure S1). The *Compensation Methods* were applied in July 2011 for the wood ash and in December 2011 for the other fertilisers, as follows:

1- Control: no nutrient application ('C'),

2- Phosphorus fertilisation ('P'), considered as standard in this forest area (application of 60 kg-P₂O₅ per hectare in the form of superphosphate (30% P₂O₅)),

3- Phosphate-potassium fertilisation (the same P treatment as above with the addition of 60 kg-K₂O per hectare in the form of potassium sulphate (50% K₂O)). It should be noted that this CM treatment was not included in this study.

4- Wood ash application ('A') consisted of the application of 5 Mg of loose ash per hectare (provided by a local power biomass plant). Taking into account the total phosphorus content of the wood ash, it corresponded to a dose of 62 kg-P₂O₅ per hectare (regardless of P forms). The ash composition is shown in the Table S1.

The WTH-C combined treatment was included to simulate the most severe harvest condition where all forest residues were removed from the site and with no nutrient application. Although unusual, this harvest management without any compensation system could be implemented if the objective is to maximize biomass production in the case of high prices for energy wood. Conversely, the SOH-P treatment is considered as the current usual stand management of local forest owners.

Plant measurements and field sampling

Estimates of biomass and nutrient exports (*Biomass Harvesting* factor):

The quantity of biomass, and associated nutrients, exported from the plots due the Biomass Harvest treatments were estimated, based on a combination of measurements and models. In practice, before

clear-cutting the standing pine stand, all trees were identified and each diameter was measured at breast height (≈ 1.3 m).

Biomass estimates

Then, we selected 14 trees that represented the range of stem size of the forest. All these trees were cut down and studied intensively:

(i) the stem diameter was measured, and wood disks were sampled, at several heights (0 m, 0.5 m, 1.3 m, and then every 2 m); the wet mass and the dry mass of the stem disks (wood and bark, separately) were measured,

(ii) the diameter of all branches was measured at 10 cm from their insertion level,

(iii) the total dry mass of three branch classes (living branches, treetop [*i.e.* the upper part of the stem, at diameter < 7 cm], dead branches) were weighed (all branches of each class were pooled),

(iv) the dry mass of 5 living branches (wood + bark + foliage) per tree was weighed, and

(v) the dry mass values of the stem wood, and the stem bark, were measured separately.

The total dry mass by compartment of each sampled tree was then calculated. The tree size measurements (diameter at breast height) were used to estimate the biomass of the different tree compartments using allometric relationships (unpublished FCBA models, calibrated with 45 trees distributed in 5 other pine stands of a similar age). The estimated biomass values of the tree compartments were consistent with the measured values, so the models were used to calculate the biomass at the plot scale, based on the inventory of all trees (measurements of the diameter at breast height).

The effective export rate of the different tree compartments was estimated in different ways, depending on the compartment studied:

(i) stem wood was assumed to be harvested in totality (export rate = 100%),

(ii) stem bark harvests were quantified by collecting all the bark that remained on the floor of a given area, after the stems exported from five other stands of the region (FCBA unpublished data; export rate $\approx 80\%$),

(iii) the effective exports of the canopies (*i.e.* branches + twigs + foliage) and of the tree belowground (stump + roots) were estimated by direct measurements of the harvests realised. In practice, the

harvested canopies were gathered into ~2.5 m long bundles (n = 465 in total), labelled per plot, and taken to the local power biomass plant. Bundles were weighed individually (fresh weight = 225-253 kg; mean weight = 240 kg), and 20 bundles were taken at random to estimate their dry matter content (mean dry content = 67% of the initial mass). All bundles were crushed to provide pellets for the power plant. During the crushing stage, 36 samples of biomass were taken at regular time intervals (Figure S2) to estimate the dry matter content. This procedure enabled us to estimate the effective harvest rate of the canopy. Conversely, it was not logistically feasible to measure the effective rates for foliage, twigs, and large branches. Based on visual observations and on field studies (Stupak et al. 2008), we assumed that (i) the twig harvest rate was 30% higher than that of the foliage, and (ii) the branch harvest rate was 50% higher than that of the twigs. Finally, based on these ratios and on the measured harvest rate of the whole canopy, the harvest rate values were adjusted. The values of the nutrient content in the different canopy compartments (*i.e.* needles, twigs, branches, stembark, stemwood) were the mean values measured on samples from the 45 trees used to build allometric relationships (see above). These mean values were within the ranges commonly observed in the study region (Augusto et al. 2008b; Trichet et al. 2018).

A similar approach to that used for branches was used for the belowground biomass. In practice, the potentially available belowground biomass (*i.e.* stump + roots) was estimated for each tree based on allometric relationships that use the stem diameter at breast height as predictive variable (Augusto et al. 2015a). The realised belowground biomass harvest was estimated from the weight of all the fresh biomass per plot (by weighing the trucks, with and without the transported biomass). The water content of the biomass was measured on 36 samples in order to calculate the dry biomass (Figure S2g; sampling at regular intervals during the crushing flow; mean value per plot of the dry content = 65-72%). These samples were used also for nutrient analyses. The measured values of nutrient content in stumps and roots were within the ranges commonly observed (Augusto et al. 2015a), except for the potassium content, which was higher in the present study (1.50 versus 0.99 mg g⁻¹).

All these measurements and analyses enabled us to quantify the standing biomass before the Biomass Harvest treatments, and the effective exports due to harvests in terms of biomass and nutrients.

Field measurements in the subsequent forest plantation

Tree growth and needle sampling:

The growth rate of the new plantation –established after biomass harvest– was assessed simply by measuring the standing biomass of the young trees. Indeed, because the survival rate of the planted seedlings was nearly 100%, and not affected by experimental treatments, the stand density was the almost the same in all subplots. The individual size of trees was estimated by measuring at least 50 pines, chosen at random, per subplot. Tree height and stem circumference at breast height were measured in December 2015 and 2019 (*i.e.* on 7- and 11-year-old trees). The aboveground biomass was then estimated based on allometric relationships (Vidal et al. 2019).

Needles were sampled to assess the nutritive status of trees, and their degree of contamination by trace elements. For this, 8 pines were first selected at random in each subplot. Current-year needles were collected in December 2019 using a telescopic pruner. Needles were collected in the upper third of the canopy, where sunlight is always directly available, and in two opposite directions.

The needles were cleaned in a bath of demineralised water before being dried with absorbent paper and then placed in an oven for ten days at a temperature of 40°C. In order to have the most homogeneous needle composite samples, and to avoid the operator effect, about 10% of the weight of the overall sample was taken at random. For this, ten needles were taken from each of the 8 batches of needles.

Understorey survey:

The understorey composition was assessed in May 2019, using the “*phytovolume*” approach (Gonzalez et al. 2013; Vidal et al. 2019). In practice, the same pair of operators surveyed all the surface area of the studied subplot, and then estimated the soil cover percentage of the main plant functional types (*i.e.* the perennial herb *Molinia caerulea*, the bracken fern *Pteridium aquilinum*, ericaceous shrubs (*Calluna vulgaris* and *Erica scoparia*), and the common gorse *Ulex europaeus*). To help the operators’ work, we printed patterns representing surface areas with defined cover values (assessed in increasing classes: 1%, 2%, 3%, 5%, 7%, 10%, and then +5% up to 95%). The mean height of each plant functional type was estimated based on three height measurements representative of the plant height range in the surveyed area (Figure S3). The phytovolume value was computed as the product of the ground cover

(estimated in squares of 16 m²) and the plant height. Finally, the phytovolume value was converted into estimates of plant aboveground biomass using dedicated allometric relationships that were previously calibrated with destructive measurements of the standing biomass (Gonzalez et al. 2013; Vidal et al. 2019).

Soil sampling:

The topsoil was sampled in all of the 36 subplots studied. To do this, a systematic grid of 8 sampling points was used per subplot: 4 points on the tree ridges, and 4 points in the furrows, to capture the spatial distribution of the topsoil induced by the regular design of the strip ploughing and tree plantation and by subsequent operations of understorey control (*i.e.* a bladed roller passing in the furrows). At each sampling point, the forest floor layer was gently removed. Then the topsoil was sampled using a corer (length = 15 cm; diameter = 8 cm). All samples were bulked in the field, and continuously homogenised by hand until the composite sample showed no heterogeneity. The soil bulk density (kg L⁻¹) was estimated based on a pedo-transfer function specifically calibrated for the local soils (Augusto et al. 2010).

Laboratory analyses

Plant analyses:

The needle samples were dried at 40°C for 7 days to constant weight and ground (< 1µm) using a IKA M20 (mill in titanium to avoid contamination of the sample).

Aliquots of powdered samples (0.25g) were digested using a mixture of 1 ml HNO₃ (69% vol Aristar® for trace element analysis, VWR Chemicals) and 4 ml H₂O₂ (30% vol, Ultratrace®, ppttrace analysis grade, Scharlau) in DigiPREP System in dry bath blocks. Then, the solution was filtered to 0.45 µm and the volume adjusted at 50 ml with ultra-pure water. The concentrations of K, Ca, Mg, Na, Fe, Mn, Al, Cr, Cu, Ni, Zn, and Co in the extracts were analysed by ICP-AES (inductively coupled plasma atomic emission spectrometry) and Cd, Pb, Tl, and Mo by ICP-MS (inductively coupled plasma mass spectrometry) (Servicio Central de Análisis de Bizkaia, Leioa-Bizkaia). The validity and accuracy of

the procedures were checked using standard reference materials (“Standard Reference Material® 1573a” certified by the National Institute of Standards and Technology).

Soil characteristics:

Soils were sieved to < 2 mm and air-dried. All analyses (except pH) were carried by INRAE soil testing laboratory (INRAE-LAS, Arras, France), according to French standardised procedures or international procedures. Soil pH was determined in a distilled water (1:5 v/v soil to solution ratio) and 0.01 M CaCl₂ extract (1:10 v/v soil to solution ratio; standard NF ISO 10390:2005). The soil organic carbon content (SOC) and total nitrogen (N_{total}) were determined by dry combustion after correction for carbonate (SOC: NF ISO 10694:1995, N_{total}: NF ISO 13878). The available phosphorus (P_{Olsen}) was determined using the Olsen method (NF ISO 11263; sodium bicarbonate extraction). The cation exchange capacity (CEC) and exchangeable cations (K⁺, Ca²⁺, Mg²⁺, Na⁺, Fe²⁺, Al³⁺, and Mn²⁺) were measured at soil pH using the cobaltihexamine chloride method (NF X 31–130:1999). The soil texture (5 fractions) was determined using the Robinson pipette method (NF X 31–107:1983), but only on three soil samples (composite samples corresponding to the three blocks of the site).

Total soil mineral element contents (major and trace) were quantified after solubilisation by fluorhydric and perchloric acids (NF X 31–147:1996). After complete dissolution, the concentrations of K, Ca, Mg, Na, Fe, Mn, Al, Cr, Cu, Ni, Zn, and Co in the extracts were analysed by ICP-AES (inductively coupled plasma atomic emission spectrometry) and Cd, Pb, Tl, and Mo by ICP-MS (inductively coupled plasma mass spectrometry).

Soil solution extraction and analyses:

Two chemical extractions were realised for the soils to estimate the availability of trace metals: ultra-pure water and ammonium nitrate 1M (NH₄-NO₃). Extraction allows the quantification of the content of elements adsorbed onto the soil with variable energy depending on the chemical nature of the extractant. Water extractions make it possible to estimate the content of elements that would be found in the soil solution. Ammonium nitrate 1 M extractions quantify the elements weakly adsorbed onto the solid phase by ion exchange at pH=6.

Ultra-pure water extraction at soil pH was performed at 1:5 (M/M) soil to solution ratio, stirring on a roller-shaking table for one hour. At the end of the extraction, the solution was extracted using a Terumo® 10 ml syringe and filtered using a Sartorius Minisart® 0.22 µm filter and then part of the solution was acidified at 2 % with HNO₃ for trace metal analysis. The second part was reserved at 4°C to measure pH (using a microelectrode (pHC4000-8, Radiometer Analytical)), the concentration of dissolved organic (DOC) and inorganic (IC) carbon (by oxidative combustion (TOC-VCSH, Shimadzu)) and the anions (PO₄³⁻ (phosphate anion), NO₂⁻ (nitrogen dioxide) and NO₃⁻ (nitrate anion)) and the cation NH₄⁺ (ammonium) by colorimetric method (Technicon auto analyser II).

The NH₄-NO₃ extractions were performed at 1:2.5 (M/M) soil to solution ratio (Symeonides and McRae 1977). The suspensions were shaken on a roller-shaking table for one hour and the solution was extracted using a Terumo® 10 ml syringe and filtered using a Sartorius Minisart® 0.22 µm filter, and then acidified with 2 % HNO₃.

The concentrations of P, K, Ca, Mg, Na, Fe, Mn, and Zn in extracts from two extractions were assayed by ICP-AES and Cd and that of Pb and Cd by GF-AAS.

Data analyses

Data processing was performed with R, version 3.6.1. (R Core Team 2019). The split-plot design was analysed by a mixed effect model (*lme* function, package nlme 3.1-143). The biomass harvest factor and the compensation method factor were fixed effects (main and interaction effects). The block and biomass harvest factor nested in blocks were random effects associated with the intercept. When detected, heteroscedasticity was corrected by modelling the variance with the *varIdent()* function (nlme package) which allows different variances by level of the biomass harvest and compensation methods. Least-squares means were calculated (emmeans package 1.4.2) and linear contrasts were used to test for significant differences between treatments and their corresponding controls. When necessary, the *P*=0.05 significance probability was adjusted for multiple pairwise comparisons (Tukey's test). Mean values are shown ± one standard error. Data and scripts are available at <https://doi.org/10.15454/LCU6OZ>.

Preliminary analyses of the results showed that the interactions between the BH factor and the CM factor were almost never statistically significant, and that is why we analysed these factors separately. The lack of interactions between biomass residues removal and wood ash application has already been reported (Hagerberg and Wallander 2002).

It should be noted that, due to the split-plot design, the statistical power is greater for the CM factor nested in BH (12 replicates = 3 blocks $\times$ 4 BH treatments) compared to the BH factor (3 replicates = 3 blocks). The main consequence of this split-plot design was greater difficulty in detecting significant effects of the BH factor.

Results

Initial stand biomass and nutrient content, and effective harvests

The standing biomass of the stand prior to harvests was $179.1 \pm 3.5 \text{ Mg ha}^{-1}$ of which $40.1 \pm 0.7 \text{ Mg ha}^{-1}$ were belowground (Table S2). The standing biomass was distributed homogeneously among the experimental plots, with a coefficient of variation of 1% among the WTH plots to 8% among the SOH plots (CV=6% at the site scale). This biomass contained nutrient contents of $24.7 \pm 0.5 \text{ kg-P ha}^{-1}$ (phosphorus) and $311 \pm 6 \text{ kg-N ha}^{-1}$ (nitrogen; Table S2). The harvesting treatments did not export all this biomass and its associated nutrients: the effective rate of harvest was measured as 71% for belowground biomass (in BAH and WTH treatments) and 91% for aboveground biomass (in AAH and WTH treatments). Within the aboveground biomass pool, the measured harvest rate of canopies (foliage + twigs + branches) was 53%, with estimated rates of 34%, 44%, and 66% for foliage, twigs and branches, respectively. The realised export of biomass was $115.32 \text{ Mg ha}^{-1}$ for the conventional harvest (SOH), with moderately increased values for more intensive harvests (from +10% for AAH to +34% for WTH; Figure 1a). Despite their low additional biomass exports, due to the differences in nutrient contents between tree parts, the non-conventional harvest practices exported a lot more nutrients than the conventional SOH technique. This was particularly the case for the canopy export of nitrogen (+74% in AAH; Figure 1b) and the belowground export of potassium (+123% in BAH). In the most

impacting treatment (WTH), the increase in nutrient exports was between +64% for P (Figure 1c) and +145% for N and K (Figure 1b and 1d).

Soil properties and fertility (hypothesis H1)

The main soil characteristics were representative of the sandy podzols of the region. For instance, the mean soil particle size distribution showed the dominance of coarse sand ($85.7\pm0.2\%$, $7.5\pm0.5\%$, $0.7\pm0.3\%$, $2.4\pm0.2\%$ and $3.6\pm0.3\%$, for the coarse sand, fine sand, coarse silt, fine silt and clay, fractions, respectively), and a low initial value of the total soil P content of $44\pm17\text{ }\mu\text{g-P g}^{-1}$ (see Augusto et al. (2010) for comparisons).

The biomass harvesting treatments had many effects on the soil nutritive status, but the effects depended on the form of the nutrients and micronutrients. While the AAH treatment showed increases in values of K, Na, Fe and Mn total contents (Table 2), the pattern was quite different for exchangeable cations (Table 3) and extractable nutrients (Table 4). Indeed, the nutrients that constitute the so-called *basic cations* of the CEC (*i.e.* K^+ , Ca^{2+} , Mg^{2+} , and Na^+) were depleted to varying extents (except for K^+) by the non-conventional harvest practices (AAH, BAH, and WTH) as compared with the SOH treatment (Table 3). The same pattern applied to Ca, Mg, and to a lesser extent K, that were $\text{NH}_4\text{-NO}_3$ extracted (Table 4). The losses in these nutrients (Ca, Mg, Mn, and Zn) were generally partly compensated for by the application of wood ash (Table S1), as shown by the significant increases in their soil content as compared with the control treatment (Tables 3 and 4).

The effects of the treatments on P also depended on the chemical form considered. The soil P content as measured using the sodium bicarbonate extractant (P_{Olsen}) showed no significant differences, even if the non-conventional harvest treatments resulted in a decrease in this soil content (Table 2). This not-significant trend became significant when considering the P content as measured using the $\text{NH}_4\text{-NO}_3$ extractant (Table 4). As expected, the application of wood ash did not modify the soil P status. Surprisingly the P-fertiliser treatment did not improve this P status either.

The soil N_{total} content followed the same, but insignificant, trend as for P with low values in the treatments that involved canopy harvesting (Table 2). The C/N ratio was remarkably irresponsive to all experimental treatments, but the soil N content was reduced by wood ash application.

Vegetation response (H2)

The *Biomass Harvesting* treatments and the *Compensatory Methods* treatments showed radically different effects on the tree foliage composition. While the intensity of biomass harvesting had no consequences on foliage nutrient content, applying phosphorus or wood ash influenced most nutrients. As expected, P-fertiliser improved the foliage P content, and the wood ash increased the content of many nutrients and micronutrients (N, P, K, Mg, B, Cu, and Fe; Table 5). Conversely, we did not expect the absence of a significant effect of wood ash on Ca content and, above all, the significant positive effect of the P fertilisation on most elements.

The total aboveground biomass of the spontaneous vegetation remained unaffected by the *Biomass Harvesting* treatments, but changed due to the *Compensatory Methods* treatments with an increasing standing biomass following the ranking: Control $\leq$ P fertilisation $\leq$ Ash (Figure S4). For both factors, the proportion of the prominent plant species was influenced by treatments. For the *Biomass Harvesting* factor (BH), the proportion of species adapted to oligotrophic-acidic moorlands (*i.e.* *Molinia caerulea* and ericaceous species) was increased by the intensive harvests as compared with the conventional SOH treatment (Figure S4). The opposite pattern was observed in the wood ash treatment (*Compensation Methods* factor, CM), which strongly decreased the abundance of those moorland-adapted species. The wood ash application and, to a lesser extent, the P fertilisation treatments enhanced the growth of the N-fixers (almost entirely composed of the spiny shrub *Ulex europaeus*).

As expected, the most severe treatments of the *Biomass Harvesting* factor (*i.e.* AAH and WTH) tended to depress the tree growth (field observations and Figure 2), but these trends remained statistically not-significant. Conversely we measured an unexpected significant difference between two treatments of the *Compensatory Methods* factor, with a negative effect of P fertilisation (Figure 2). In addition, the wood ash application tended to improve tree growth.

Soil organic matter and CEC (H3)

The soil Cation Exchange Capacity (CEC) was strongly and linearly correlated with the quantity of elements that are commonly found as organic matter in acidic soils, which are organic carbon (SOC) and total nitrogen (N_{total}). In the case of organic carbon, its soil content explained 76% of the variance of the exchange capacity ($CEC = 0.088 \times SOC$; $r^2 = 0.76$).

Due to the tight proportional relationship between CEC and SOC, the observed effects of the experimental treatments on these variables were quite similar. The harvest of the belowground biomass and the application of wood ash both tended to decrease the soil organic matter content and the soil CEC (Tables 2 and 3). The dissolved organic carbon (DOC) followed a similar pattern with a significant decrease in concentration after whole-tree harvesting (WTH) and wood ash application (Table 2).

Soil acidity status (H4)

Harvesting more biomass than conventionally practiced did not modify any of the soil variables used to assess the acidic status of a soil (pH-H₂O, pH-CaCl₂, Base Saturation of the CEC; Table 3). The absence of effect on the Base Saturation was the consequence of the concomitant decrease in *basic cations* (K^+ , Ca^{2+} , Mg^{2+} , and Na^+) and CEC.

Conversely to biomass harvesting, applying wood ash modified the soil acidity status. The soil pH-H₂O value increased by +0.24 unit, and the Base Saturation increased from 39% to 46% of the CEC (Table 3). The P fertilisation had no effect on soil acidity.

Ecosystem content in trace metals (H5)

Overall, the *Biomass Harvesting* treatments generally had no significant effect on the trace elements (As, Cd, Mo, Ni, Pb, Tl, and Zn) in the ecosystem (Tables 2, 4, 5, 6, S4). The exceptions were a decrease in soil Pb and Zn contents due to WTH.

The *Compensatory Methods* also had only a few significant effects on the trace element distribution in the ecosystem (Tables 2, 4, 5, 6, S4). The most noticeable exceptions were increases of soil Zn content due to wood ash application (Tables 4 and S4), an effect of P fertilisation on Cd distribution between

the soil and trees, and an improvement of Cu content of tree foliage after P or wood ash application (Table 5).

Discussion

Importance of the study design and the study region for data interpretation

Implementing a careful harvest of adult tree compartments over small areas was hardly feasible because the machines that collect stumps or canopies need space to work (Figure S2). Consequently, the *Biomass Harvesting* factor could be studied only by using large plots (*i.e.* one ha per plot). With 16 experimental treatments and three blocks, building a complete factorial experimental design (*i.e.* with 48 large plots) was not possible because the surface area of 48 ha required was incompatible with the premise of an area that was initially homogeneous in its properties and past-management. Therefore, we built our experiment based on a split-plot design. Such a design had one major consequence on data analysis. Indeed, the number of the subplots –dedicated to the *Compensation Methods* treatments– were four-fold more numerous than the experimental areas dedicated to *Biomass Harvesting* treatments (Figure S1). The power of the statistical tests was consequently much lower for the *Biomass Harvesting* factor, so isolating significant effects was sometimes difficult.

The forest region where our experiment took place is atypical from a biogeochemical perspective. As a matter of fact, the soils of the *Landes de Gascogne* region are among the poorest soils in phosphorus in the world. While in non-agricultural systems the soil P_{-total} content ranges between ~10 and 2,000 µg g⁻¹ (Achat et al. 2009; Yang and Post 2011; Bretoire et al. 2016; Augusto et al. 2017), the study region has a mean value of ~50 µg g⁻¹ and is within a range of ~10-100 µg g⁻¹ (Achat et al. 2009; Augusto et al. 2010). As such, the study region is a hotspot of P limitation and is more similar to ancient landscapes than to most European forests (Lambers et al. 2010; Vitousek et al. 2010). Being also acidic and poor in other nutrients and micronutrients (Augusto et al. 2010; Trichet et al. 2018), the soils of the study region have been identified as having the highest level of vulnerability to biomass exports according to the national system of forest evaluation (Durante et al. 2019). Poorest of the poor, the study region is also

expected to respond strongly and negatively to additional harvests of biomass, which implies that results should be extrapolated to other regions with caution.

Consequences of harvesting more biomass on the functioning of an oligotrophic forest

Exporting tree canopies generally enhances the amount of harvested biomass (~ +20% to +50%), but this is at the expense of losing huge quantities of nutrients (~ +100% to +250%; (Achat et al. 2015a)). Our results (10-34% extra biomass, and 37-145% extra nutrients) fit well with this general pattern. On the other hand, the quantities of exported nutrients of this field experiment are about half of the values used in previous modelling studies (Augusto et al. 2015b; Achat et al. 2018). This was the direct consequences of the harvesting techniques that were purposely used to reduce the losses of nutrients: letting branches shed their needles (Stupak et al. 2008), and harvesting only stumps and coarse roots (Augusto et al. 2015a). Despite these precautions, harvesting more biomass, and above all harvesting the whole-tree, reduced the most available forms of nutrients (K, Ca, Mg, and P) in the soil nutrient pools (-15% to -58%; Table 4). Additional biomass harvests likely also impacted the soil content in organic matter, with negative ripple effects on organic P and CEC, the latter being entirely dependent on organic matter in these sandy soils (Augusto et al. 2010). In the study region the organic forms of P are important for seedling and tree nutrition (Jonard et al. 2009; Achat et al. 2013), which would explain why trees of the subsequent stand have grown more slowly after intensive biomass harvests (even if a longer monitoring is needed to confirm the observed trend; see Table S3). The absence of change in the tree foliage composition might be considered as surprising as foliage nutrient content is often used to assess plant nutrition (CSIRO 1997; Mellert and Göttelein 2012). Nevertheless, maintaining the nutrient content of foliage at the expense of growth is a response commonly observed after the harvest of additional biomass (Achat et al. 2015a), and our results are consistent with this general pattern.

Fertilisation and wood ash application as possible mitigation practices

Because phosphorus is the first-order factor that limits tree growth in the study region (Trichet et al. 2009), we initially expected that the application of P-fertiliser to mitigate the negative effects of

additional harvests, but this was not the case in our experiment. Because the soil was also impoverished in other major nutrients, and because they are considered as second-order nutritional limitations in this context (Trichet et al. 2008), tree growth may have been constrained in our experiment by a multi-element scarcity. Multi-element fertilisation is often put forward as a major advantage of applying wood ash in forests (Ranius et al. 2018). Both wood ash application and P fertilisation were tested in the present study.

The major effect of applying wood ash was expected to be a change in the acidity status of soils (Reid and Watmough 2014). A simple approximation states that two tons of ash have the same neutralising effect as one ton of calcium carbonate (Vance 1996). Our experiment was not an exception and the soil pH, base saturation and concomitant available calcium and magnesium, were improved by wood ash application (for results in a similar context to ours, see Gomez-Rey et al. (2013)). The available soil micronutrient contents of Mn and Zn also logically increased because wood ash contains them in substantial quantities (Table S1). Whereas the effects of wood ash were expected and straight forward to interpret, the P fertilisation had surprising outcomes: the soil P status was not improved, and the soil contents of available Ca and Mg actually decreased by P fertilisation. These results may be explained by the tree foliage composition that was modified by both P fertilisation and wood ash application, with increases in concentration values for most nutrients and micronutrients (and even some non-essential trace metals). Hence, it seemed that the two compensatory methods improved tree nutrition but, for P fertilisation, it was at the expense of the soil reserves in Ca and Mg.

In turn, these changes in the soil properties modified the dynamics of the spontaneous vegetation and of the tree plantation. The vegetation experienced a shift of species prominence after the wood ash application, with a decrease in the species typical of acidic moorlands and heathlands (*i.e. Molinia caerulea* and ericaceous species) to the benefit of the local main N-fixer (the shrub *Ulex europaeus*). The decrease of the abundance of the acidophilus species is logical since wood ash decreased the soil acidity. The increase of the N-fixer shrub was expected as we are used to observing a positive response of this leguminous species to P fertilisation (*e.g.* Delerue et al. (2015)), but although the N-fixers abundance was increased by P fertilisation, it was relatively low compared with previous experiments (Augusto et al. 2005; Vidal et al. 2019), and lower than results after the wood ash application. This effect

has already been reported in the literature (Campillo et al. 2005) and may be the positive consequence of reducing the ambient acidity on the symbionts of the leguminous species (Slattery and Coventry 1995).

Trees that were planted responded differently to the *Compensatory Methods*. Firstly, trees that received wood ash tended to grow the fastest. This result is consistent with our interpretation that plant growth in this site was limited by several elements at the same time. Since wood ash application improved the availability of many nutrients and micronutrients, it was logical that plant growth was enhanced. On the other hand, the depressive effect of P fertilisation was surprising considering that this practice has proven its efficiency in the study region for more than half a century (Trichet et al. 2009). As explained above, it seems that P fertilisation improved the tree foliage composition at the expense of soil fertility, and we speculate that this soil impoverishment might be at the origin of subsequent degraded tree growth. However, this atypical phenomenon requires further investigation.

Considering the positive effects of wood ash application on soil acidity and nutrient content, and its positive repercussion on tree growth, one might conclude that this compensatory method should be promoted in oligotrophic forests submitted to intensive biomass harvestings. Nevertheless, this conclusion does not take into account one major drawback of wood ash application in our experiment, which was a strong decrease of the soil organic matter content, with negative consequences on organic carbon and total nitrogen contents and soil exchange capacity. Even after correcting for the change in soil bulk density, the losses of organic carbon were ~10-11% of the initial stock of the topsoil (N_{total} losses were ~7%). We interpret this decrease as the result of the acidity alleviation by wood ash, which probably enhanced the soil microbial activity and respiration (Bååth and Arnebrant 1994; Jokinen et al. 2006; Omil et al. 2013).

In a meta-analysis, we previously concluded that wood ash had no influence on soil organic carbon (Augusto et al. 2008a), but this study was almost entirely based on Nordic, or cold temperate experiments (mean annual temperature < 8.5°C; e.g. (Feldkirchner et al. 2003)). Under a warm temperate climate, wood ash application seems to increase SOC stocks when applied in recent afforestation of former croplands (Sartori et al. 2007; Ventura et al. 2019), which can be explained by the initial soil conditions (i.e. low SOC values and high pH values) and improved necromass production

by plants (*i.e.* litterfall and fine root turnover). In areas with a long history of forest occupation (*i.e.* high SOC values and often low pH values), to our knowledge, our experiment is only the second study under a warm temperate climate that has quantified the effect of wood ash on SOC: both studies observed a possible decrease in organic carbon soil contents (Solla-Gullon et al. 2006).

Initial expectations and future anticipation

Most, but not all, of our initial expectations were confirmed by the field experiment. Firstly, as anticipated, harvesting more biomass aggravated the P scarcity of the local soils, and the application of wood ash did not modify the soil P content (Hypothesis H1; Table S5). Still in line with our expectations, the impoverishment of the soil caused by additional harvests of biomass could result in reduced growth of the subsequent forest stand; wood ash application had positive effects on early tree growth (H2). The fact that, unexpectedly, P application did not enable trees to overcome P limitation suggested that other nutritional limitations –such as Ca and Mg– prevented the experimental fertilisation from having an effect. If our two first hypotheses were fairly well supported by results, the third and fourth hypotheses received mixed support. Indeed, both high rates of biomass harvesting and wood ash application decreased the pool of soil organic carbon, but only the former treatment was supposed to do so (H3). The SOC destocking caused by wood ash application opened perspectives on a large scale (see below). Similarly, while wood ash application logically improved the acidity status of the soil to a moderate extent, intensive biomass exports did not deteriorate it, as in many other studies (H4; see above for possible explanations). Finally, none of the experimental factors had the anticipated major effects on the trace metal distribution within the ecosystem (H5).

Overall, this factorial experiment showed that exporting more forest biomass through the additional harvests of tree canopies, stumps, and roots had negative consequences on the ecosystem. Additional harvests have aggravated the poverty of the already oligotrophic soil, which in turn may decrease tree growth and the soil content of organic carbon in the future (but without any impact either on soil acidity or on trace metals content). Importantly, applying nutrients as fertiliser or wood ash did not fully compensate for the negative impact of biomass exports.

Beyond the management consequences for the *Les Landes de Gascogne* region, this study may have an interesting repercussion on research about climate change mitigation. Indeed, the *intensive biomass harvest* strategy (which proposes to harvest more woody biomass to replace fossil C in the production of energy), in conjunction with the application of wood ash to return part of the exported nutrients, can be seen as a winning approach in boreal forests because neither biomass harvests (Achat et al. 2015b) nor wood ash (Augusto et al. 2008a) destock soil carbon in this biome. In temperate forests, intensive biomass harvests are already known to decrease soil organic carbon (Achat et al. 2015b), but wood ash was not suspected to do so, probably because current knowledge is based on research carried out in Nordic countries (Augusto et al. 2008a). This study is among the first field experiments to have tested the effect of wood ash application on soil organic carbon under a warm temperate climate. Although we acknowledge that our study is based on only one field experiment and should be followed by others, we anticipate that, if the negative impact of wood ash on soil carbon sequestration was confirmed in several other temperate forests, the *intensive biomass harvest* strategy would be inefficient in such ecosystems and should be replaced by the *old-growth* strategy –consisting of sequestering carbon in the ecosystem– to mitigate climate change.

Acknowledgements

We are grateful to the ADEME, GIS G-PMF and the Council of the Nouvelle-Aquitaine region for their financial support (D.V. PhD grant; SYLVOGENE and PINASTER projects). Similarly, we thank the ISPA lab for providing the F.B. Master grant. Finally, we are grateful to the Forest Experimental Facility (UEFP-<https://doi.org/10.15454/1.5483264699193726E12>) for their assistance during the successive field campaigns, and to Sylvie Bussière, Cécile Coriou, and Catherine Lambrot for their help during the sample analyses.

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Figure 1 – Realised exports of biomass, nitrogen, and phosphorus

SOH = stem-only harvest; AAH = aboveground additional harvest; BAH = belowground additional harvest; WTH = whole-tree harvest.

All data (including data about calcium, and magnesium) are presented in Table S2.

The methods used to quantify the biomass and nutrient exports are detailed in the manuscript in the section “Plant measurements and field sampling”, in particular in the subsection “Estimates of biomass and nutrient exports”.

Figure 2 – Tree size as affected by biomass harvest and compensation methods

SOH = stem-only harvest; AAH = aboveground additional harvest; BAH = belowground additional harvest; WTH = whole-tree harvest. Control = no nutrient application. *P* values for differences between treatments of reference (*i.e.* SOH and Control) on the one hand, and other treatments on the other hand, are indicated in the graph. Trees were measured at 11 years old. Other tree metrics are shown in Table S3.

Table 1 – Initial expected consequences of intensive biomass harvesting and wood ash application on a temperate oligotrophic forest
(➔: means “has no effect on...”; ⬇: means “decreases...”; ⬆: means “increases...”; ?: means “unknown effect on...”)

Ecosystem trait		Intensive harvests	Wood ash
Soil fertility (phosphorus) {hypothesis H1}	<i>General knowledge</i>	⬇ soil P	➔ soil P
	<i>Local conditions</i>	P-limited soils	
	<i>Expected results</i>	⬇ (aggravated P-limitation)	➔ (or ⬆) soil P
Tree growth (carbon sequestration) {H2}	<i>General knowledge</i>	⬇ tree growth	➔ (or ⬆) tree growth
	<i>Local conditions</i>	oligotrophic P-limited ecosystem	
	<i>Expected results</i>	⬇ tree growth	➔ tree growth
Soil Organic Matter SOM (carbon and nitrogen) {H3}	<i>General knowledge</i>	➔ boreal forests ⬇ temperate forests	➔ boreal forests ? temperate forests
	<i>Local conditions</i>	warm temperate	
	<i>Expected results</i>	⬇ SOM	➔ SOM
Soil acidity (pH, base saturation) {H4}	<i>General knowledge</i>	⬇ acido-basic status	⬆ acido-basic status
	<i>Local conditions</i>	acidic soils	
	<i>Expected results</i>	⬇ acido-basic status	⬆ acido-basic status
Trace metals in ecosystem (micronutrients: [B, Cu, Fe, Mn, Mo, Zn] and toxic trace metals [As, Pb, Cd]) {H5}	<i>General knowledge</i>	➔ or ⬇ trace metals (due to exports)	➔ for podzol soils (boreal soils in Nordic areas)
	<i>Local conditions</i>	low content in tree biomass growing on podzol soils	
	<i>Expected results</i>	➔ trace metals	➔ or ⬇ bioavailability at moderate ash dose + increase pH

Table 2 – Soil properties related to site fertility as affected by biomass harvest and compensation methods

	SOC (mg g ⁻¹)	DOC (mg L ⁻¹)	N (mg g ⁻¹)	C/N unit less	P _{Olsen} (µg g ⁻¹)	K (mg g ⁻¹)	Ca (mg g ⁻¹)	Mg (mg g ⁻¹)	Na (mg g ⁻¹)	Fe (mg g ⁻¹)	Mn (µg g ⁻¹)	Mo (µg g ⁻¹)	Zn (µg g ⁻¹)
<i>Biomass Harvesting</i>													
SOH	38.6±3.9	44±2	1.41±0.15	27.7±0.8	29.3±2.2	4.07±0.2	0.82±0.06	0.17±0.09	1.17±0.09	1.8±0.4	41.5±3.5	0.16±0.01	6.60±0.29
AAH	35.4±3.9	42±4	1.46±0.15	24.5±0.8	25.3±2.5	5.40±0.2 *	0.98±0.06	0.25±0.09	1.65±0.08 *	3.1±0.3 *	62.0±3.5 *	0.21±0.01	7.37±0.48
BAH	30.2±3.9	39±2	1.14±0.15	26.3±0.8	25.3±2.9	4.31±0.2	0.71±0.04	0.15±0.08	1.22±0.08	2.2±0.3	43.6±3.5	0.15±0.01	5.77±0.36
WTH	31.1±3.9	31±2 *	1.10±0.15	28.2±0.8	21.0±3.5	4.09±0.2	0.72±0.03	0.11±0.04	1.11±0.05	1.5±0.3	35.4±3.5	0.13±0.01	3.85±0.54 *
<i>Compensation Methods</i>													
Control	37.1±2.9	39±1	1.36±0.11	27.2±0.7	27.1±2.4	4.47±0.13	0.85±0.03	0.18±0.10	1.31±0.05	2.1±0.3	46.6±3.2	0.16±0.01	5.69±0.31
Phosphorus	33.6±2.9	42±2	1.28±0.11	26.3±0.7	24.5±2.4	4.45±0.26	0.74±0.03 *	0.16±0.08	1.26±0.09	2.0±0.3	45.0±4.9	0.17±0.01	5.57±0.32
Ash	30.8±2.9 *	36±1 *	1.18±0.11 *	26.5±0.7	24.0±2.4	4.50±0.15	0.85±0.03	0.17±0.09	1.31±0.06	2.3±0.3	45.2±2.0	0.16±0.01	6.42±0.36

All values are for the total content of the elements in the soil (except for P). SOC = soil organic carbon; DOC = dissolved organic carbon. For phosphorus (P), values are extractable P using the Olsen method.
SOH = stem-only harvest; AAH = aboveground additional harvest; BAH = belowground additional harvest; WTH = whole-tree harvest. Control = no nutrient application.
Treatments with an asterisk differ at $P<0.05$ from their reference treatment (SOH or Control, respectively; see Methods).

892 **Table 3** – Soil acidity status as affected by biomass harvest and compensation methods

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	pH-H ₂ O unit less	pH-CaCl ₂ unit less	CEC _m (cmol. _c kg ⁻¹)	BS (%)	K (cmol. _c kg ⁻¹)	Ca (cmol. _c kg ⁻¹)	Mg (cmol. _c kg ⁻¹)	Na (cmol. _c kg ⁻¹)
<i>Biomass Harvesting</i>								
SOH	4.43±0.07	3.43±0.08	3.51±0.23	43±2	0.06±0.00	0.92±0.04	0.36±0.02	0.06±0.00
AAH	4.68±0.07	3.69±0.08	2.90±0.24	38±2	0.06±0.00	0.67±0.03 *	0.21±0.01 *	0.05±0.00
BAH	4.49±0.07	3.57±0.08	2.51±0.23 *	42±2	0.05±0.00	0.65±0.04 *	0.23±0.02 *	0.04±0.00 *
WTH	4.51±0.07	3.46±0.08	2.90±0.24	39±2	0.06±0.01	0.69±0.06	0.28±0.02 *	0.04±0.00 *
<i>Compensation Methods</i>								
Control	4.41±0.06	3.47±0.07	3.45±0.21	39±2	0.06±0.00	0.74±0.05	0.29±0.01	0.05±0.00
Phosphorus	4.53±0.05	3.51±0.07	2.58±0.19 *	38±2	0.06±0.00	0.57±0.04 *	0.22±0.02 *	0.04±0.00 *
Ash	4.65±0.05 *	3.64±0.07	2.84±0.19 *	46±2 *	0.06±0.00	0.89±0.04 *	0.30±0.02	0.05±0.00

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895 SOH = stem-only harvest; AAH = aboveground additional harvest; BAH = belowground additional harvest; WTH = whole-tree harvest. Control = no nutrient application.

896 CEC_m = cationic exchange capacity (measured value); BS = base saturation of the CEC, which is the proportion of charges occupied by non-acidic exchangeable cations (K,
897 Ca, Mg, Na).

898 For consistency between the different terms of its equation, BS was calculated using the effective CEC (CEC_e: calculated as the sum all the charges of all the analysed cations),
899 whose values differ slightly from CEC_m values.

900 Treatments with an asterisk differ at $P < 0.05$ from their reference treatment (SOH or Control, respectively; see Methods).

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907 **Table 4** – Soil composition in NH₄-NO₃ extractable elements as affected by biomass harvest and compensation methods
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	P (µg g ⁻¹)	K (µg g ⁻¹)	Ca (µg g ⁻¹)	Mg (µg g ⁻¹)	Fe (µg g ⁻¹)	Mn (µg g ⁻¹)	Zn (µg g ⁻¹)
<i>Biomass Harvesting</i>							
SOH	1.2±0.3	27±2	178±8	47±2	7.0±1.2	1.5±0.1	0.60±0.05
AAH	0.5±0.2 *	21±2 (*)	117±10 *	25±2 *	9.7±1.2	1.3±0.1	0.48±0.04
BAH	0.8±0.2	21±2 (*)	128±8 *	30±2 *	8.2±1.2	1.2±0.1	0.47±0.04
WTH	0.9±0.2	21±2 (*)	151±12	38±2	6.1±1.2	1.2±0.1	0.45±0.04
<i>Compensation Methods</i>							
Control	0.9±0.2	23±1	131±9	35±2	7.2±1.3	1.1±0.1	0.44±0.02
Phosphorus	0.7±0.2	22±1	117±9	30±2 *	8.5±0.9	1.1±0.1	0.48±0.03
Ash	0.9±0.1	22±1	183±9 *	40±1 *	7.5±0.5	1.8±0.1 *	0.58±0.06 *

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910 SOH = stem-only harvest; AAH = aboveground additional harvest; BAH = belowground additional harvest; WTH = whole-tree harvest. Control = no nutrient application.
911 Treatments with an asterisk differ at $P < 0.05$ (except for K: P value < 0.10 ; symbol in brackets) from their reference treatment (SOH or Control, respectively; see Methods).
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917 **Table 5** – Tree nutritional status (needle composition) as affected by biomass harvest and compensation methods
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	N (mg g ⁻¹)	P (mg g ⁻¹)	K (mg g ⁻¹)	Ca (mg g ⁻¹)	Mg (mg g ⁻¹)	B (μg g ⁻¹)	Cu (μg g ⁻¹)	Fe (μg g ⁻¹)	Mn (μg g ⁻¹)	Mo (μg g ⁻¹)	Zn (μg g ⁻¹)
<i>Biomass Harvesting</i>											
SOH	12.8±0.7	0.64±0.04	4.6±0.1	2.3±0.1	1.4±0.1	16.3±1.0	1.8±0.2	27.8±2.0	96.3±11.0	0.011±0.003	29.8±2.5
AAH	13.9±0.7	0.69±0.04	4.4±0.1	2.2±0.1	1.5±0.1	16.7±1.0	1.8±0.2	32.1±2.1 *	104.2±10.9	0.014±0.002	32.1±2.5
BAH	12.1±0.7	0.65±0.04	4.5±0.1	2.2±0.1	1.4±0.1	17.2±1.0	2.0±0.2	27.1±2.0	92.9±11.7	0.009±0.003	30.5±2.5
WTH	12.6±0.7	0.65±0.05	4.6±0.1	2.3±0.1	1.4±0.1	17.2±1.0	1.8±0.3	28.9±2.9	81.0±11.7	0.012±0.002	32.1±2.5
<i>Compensation Methods</i>											
Control	12.3±0.7	0.64±0.04	4.2±0.1	2.2±0.1	1.4±0.1	16.0±0.9	1.7±0.1	27.0±2.1	92.3±9.8	0.011±0.002	29.1±2.2
Phosphorus	13.6±0.7 *	0.66±0.04 *	4.5±0.1 *	2.3±0.1	1.5±0.1 *	18.0±0.9 *	1.9±0.1	30.4±2.1 *	100.1±9.4 *	0.012±0.002	32.7±2.2 *
Ash	12.7±0.7 *	0.67±0.04 *	4.8±0.1 *	2.3±0.1	1.4±0.1	16.6±0.9 *	2.0±0.1	29.6±2.0 *	88.4±9.8	0.012±0.002	31.7±2.2 *

919 SOH = stem-only harvest; AAH = aboveground additional harvest; BAH = belowground additional harvest; WTH = whole-tree harvest. Control = no nutrient application.
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921 Treatments with an asterisk differ at $P<0.05$ from their reference treatment (SOH or Control, respectively; see Methods).
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926 **Table 6** – Ecosystem content in non-essential trace metals

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	Cadmium			Lead			Arsenic
	<i>Soil</i> (µg-Cd kg ⁻¹)	<i>Extractable</i> (µg-Cd kg ⁻¹)	<i>Foliage</i> (µg-Cd g ⁻¹)	<i>Soil</i> (µg-Pb g ⁻¹)	<i>Extractable</i> (µg-Pb g ⁻¹)	<i>Foliage</i> (µg-Pb g ⁻¹)	<i>Foliage</i> (µg-As kg ⁻¹)
<i>Biomass Harvesting</i>							
SOH	33±2	9.4±0.6	0.17±0.01	9.45±0.79	0.24±0.01	0.17±0.01	19±3
AAH	33±2	8.9±0.6	0.18±0.01	9.53±0.58	0.17±0.01	0.18±0.01	21±3
BAH	30±2	8.4±0.6	0.16±0.01	7.59±0.45	0.17±0.01	0.16±0.01	15±3
WTH	26±2	7.6±0.6	0.17±0.01	6.76±0.24 *	0.16±0.01	0.07±0.01	21±3
<i>Compensation Methods</i>							
Control	31±1	8.2±0.5	0.16±0.01	8.20±0.39	0.20±0.01	0.16±0.01	16±2
Phosphorus	28±1 *	8.2±0.4	0.18±0.01 (*)	8.24±0.39	0.18±0.01	0.18±0.01 (*)	20±4
Ash	32±1	9.4±0.7	0.17±0.01	8.57±0.39	0.17±0.01	0.17±0.01	20±4

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929 “Soil” = soil substrate (total content of Cd and Pb); “Extractable” = soil extractable with 1M NH₄-NO₃ extractant; “Foliage” = tree foliage (*i.e.* pine needles) content. It should
930 be noted that the units used differ among Cd variables.

931 SOH = stem-only harvest; AAH = aboveground additional harvest; BAH = belowground additional harvest; WTH = whole-tree harvest. Control = no nutrient application.

932 Treatments with an asterisk differ at $P < 0.05$ (for P value < 0.10 , the symbol is in brackets) from their reference treatment (SOH or Control, respectively; see Methods).

933 Total soil content of nickel (Ni) and thallium (Tl) are not presented. No significant differences were observed for these variables (Ni=1.49 µg g⁻¹; Tl=0.11 µg g⁻¹).

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