

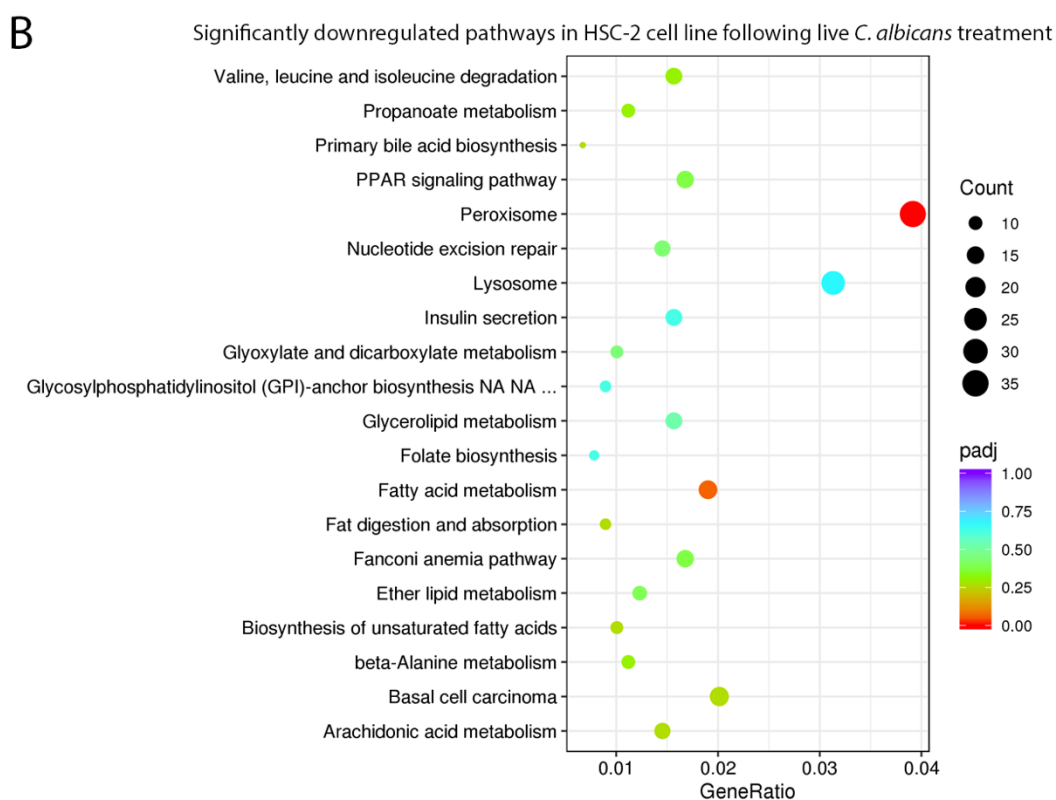
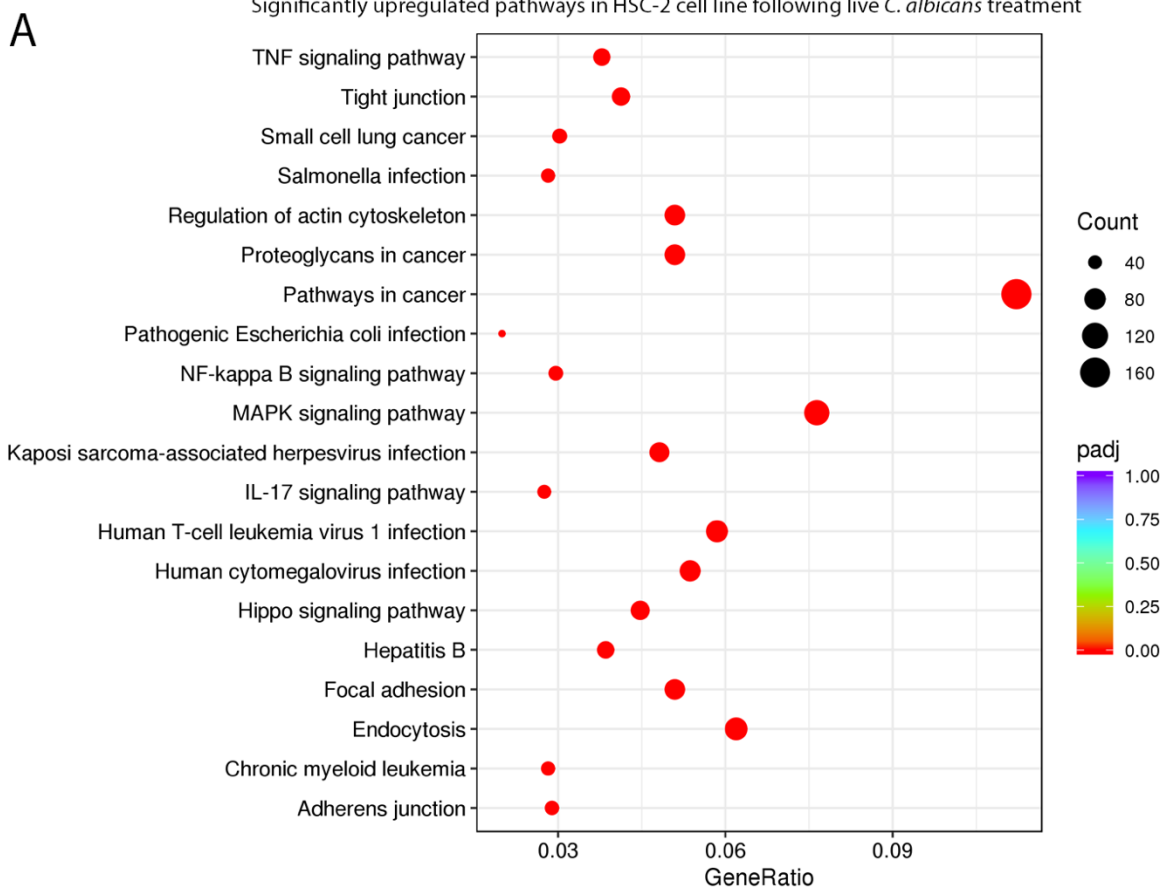
Supp Fig1

(A) Normalized proliferation activity of OSCC cells in the presence of heat- killed *C. albicans*, *C. parapsilosis* and zymosan measured by BrdU incorporation assay.

(B) Normalized proliferation activity of OSCC cells in the presence of live *C. albicans* and *C. parapsilosis* measured by BrdU incorporation assay.

(C) Normalized amount of metabolites of OSCC cells in the presence of heat- killed *C. albicans*, *C. parapsilosis* and zymosan measured by HPLC-HRMS.

(D) Normalized amount of metabolites of OSCC cells in the presence of live *C. albicans* and *C. parapsilosis* measured by HPLC-HRMS.



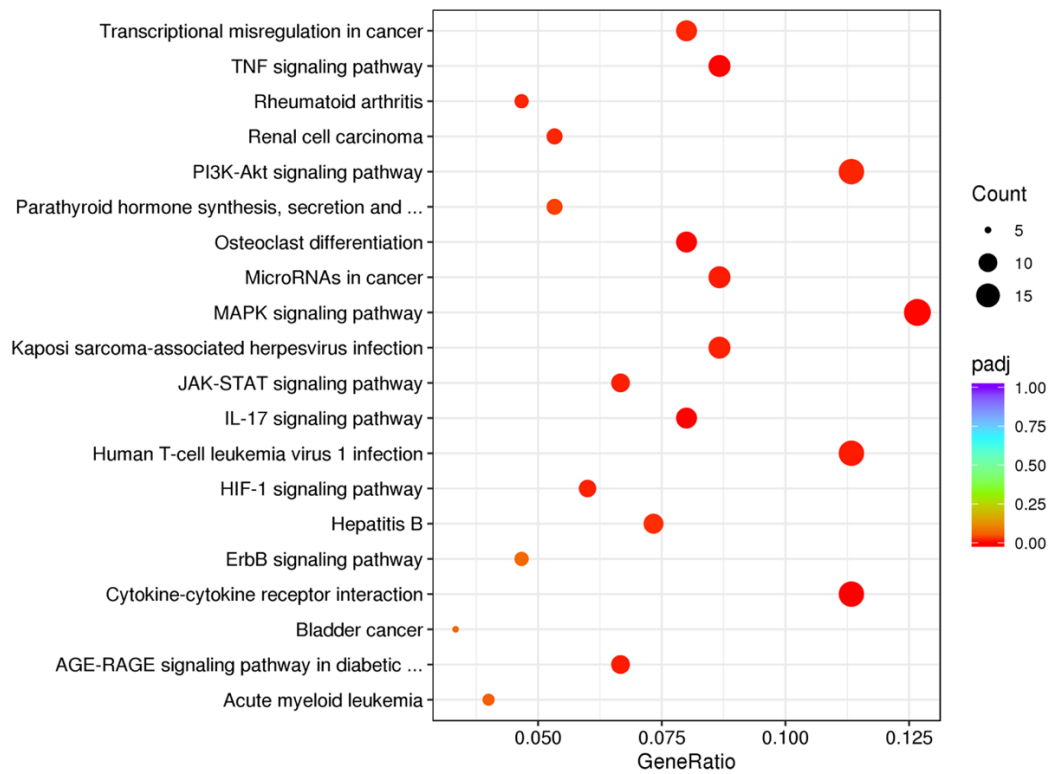
Supp Fig2

(A) Significantly upregulated pathways in HSC-2 cell line induced by *C. albicans*

(B) Significantly downregulated pathways in HSC-2 cell line following *C. albicans* treatment

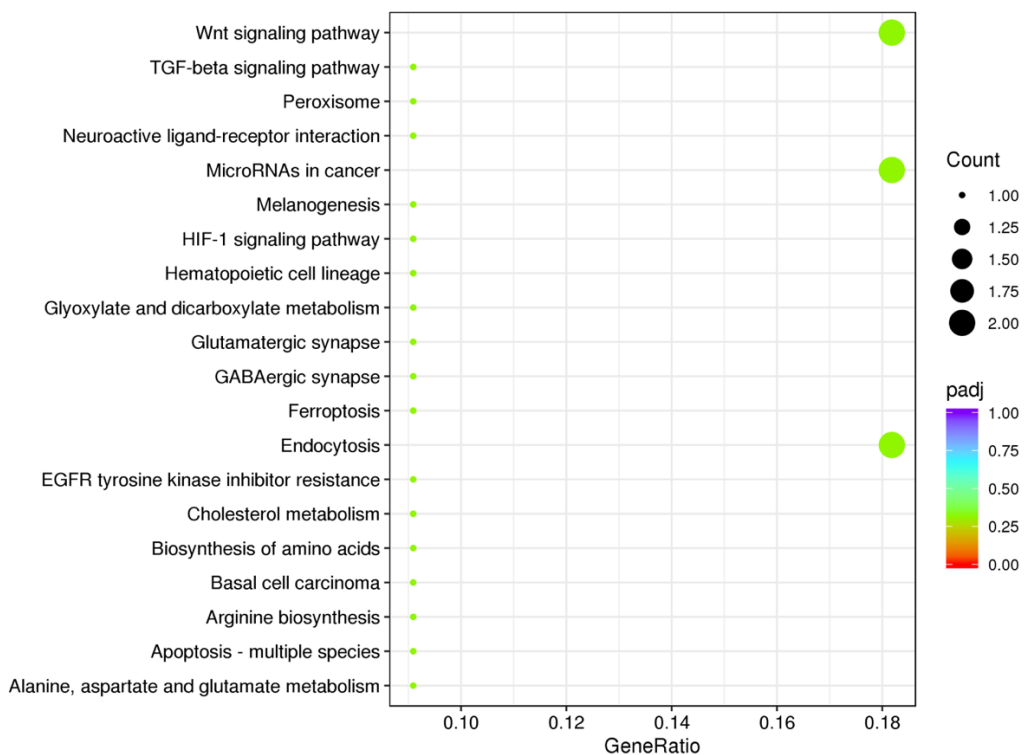
A

Significantly upregulated pathways in HO-1-N-1 cell line following live *C. albicans* treatment



B

Significantly downregulated pathways in HO-1-N-1 cell line following live *C. albicans* treatment



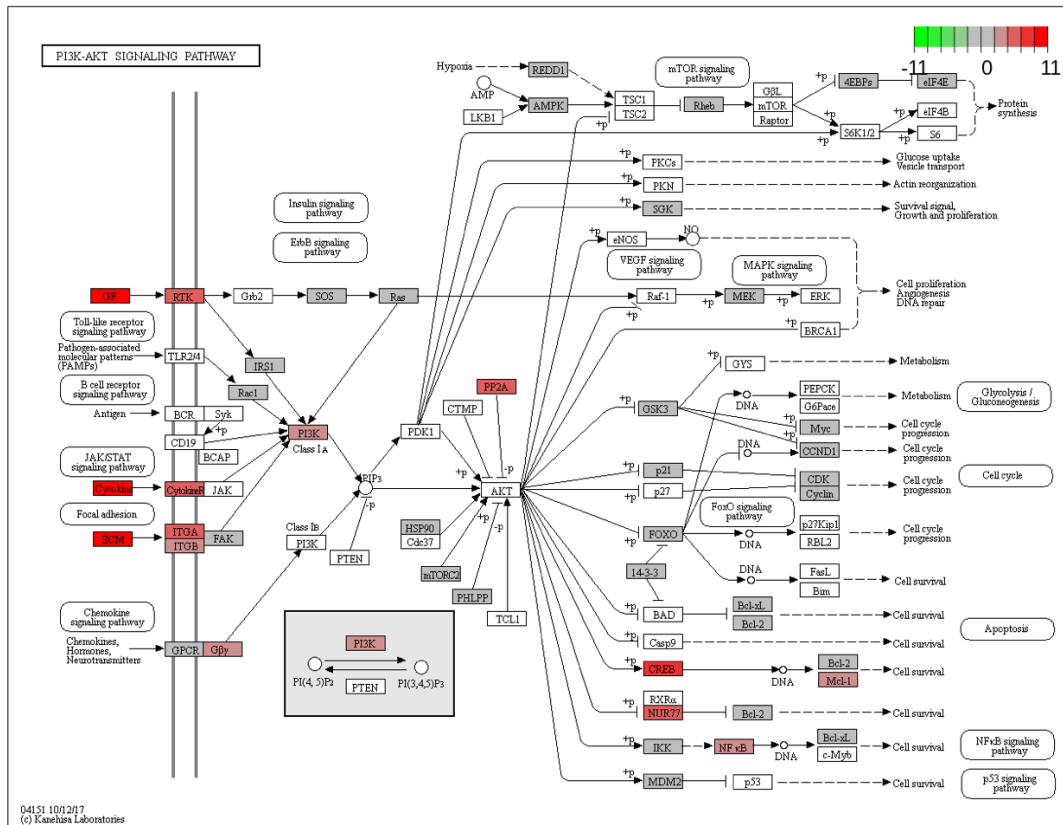
Supp Fig3

(A) Significantly upregulated pathways in HO-1-N-1 cell line induced by *C. albicans*

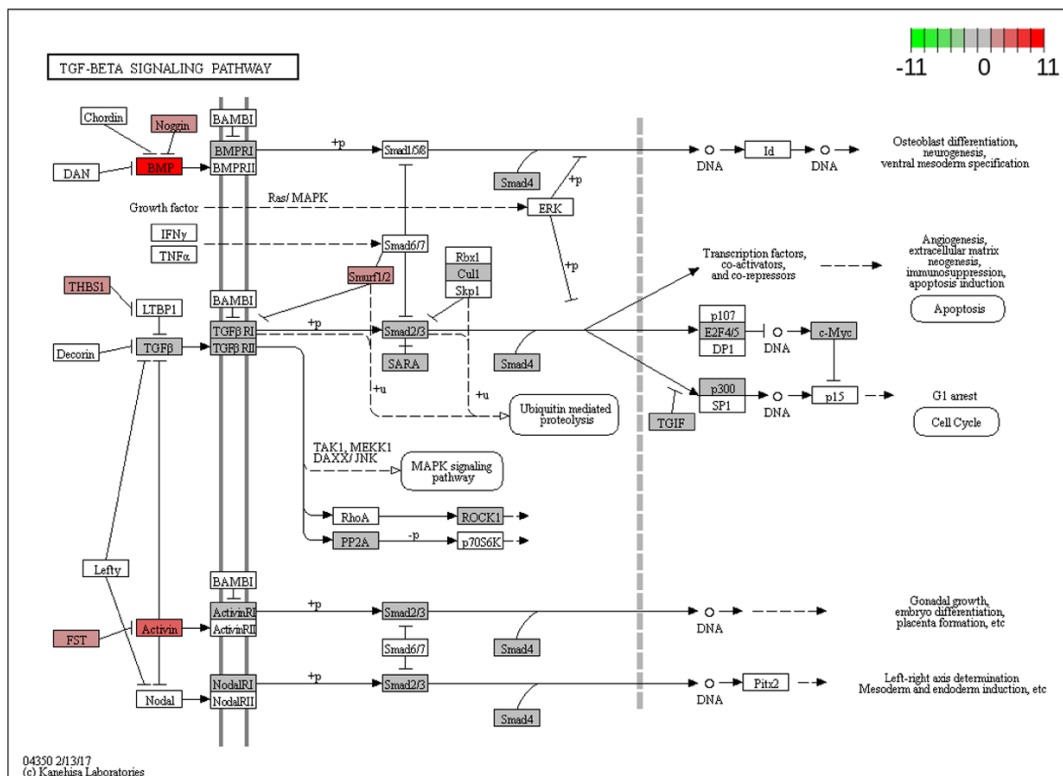
(B) Significantly downregulated pathways in HO-1-N-1 cell line following *C. albicans* treatment

Pathways activated in HSC-2 cell line after live *C. albicans* treatment

A



B



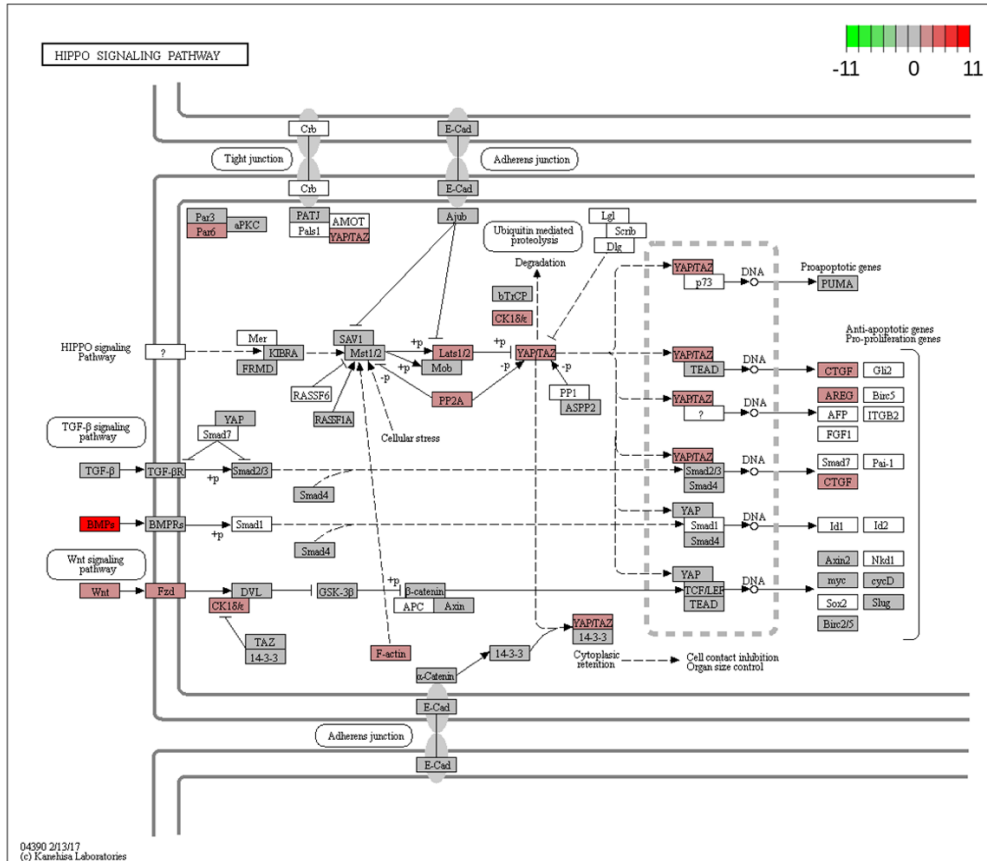
Supp Fig4

(A) PI3K-Akt signaling pathway activated in HSC-2 cell line following live *C. albicans* treatment

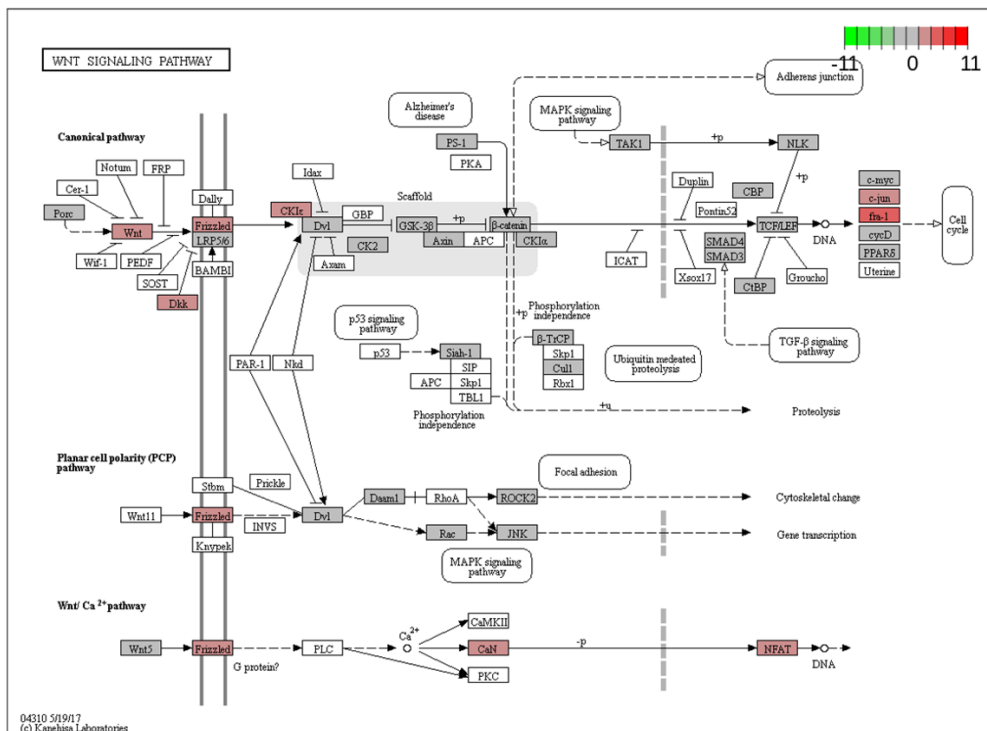
(B) TGF- β /SMAD signaling pathway activated in HSC-2 cell line following live *C. albicans* treatment

Pathways activated in HSC-2 cell line after live *C. albicans* treatment

A



B



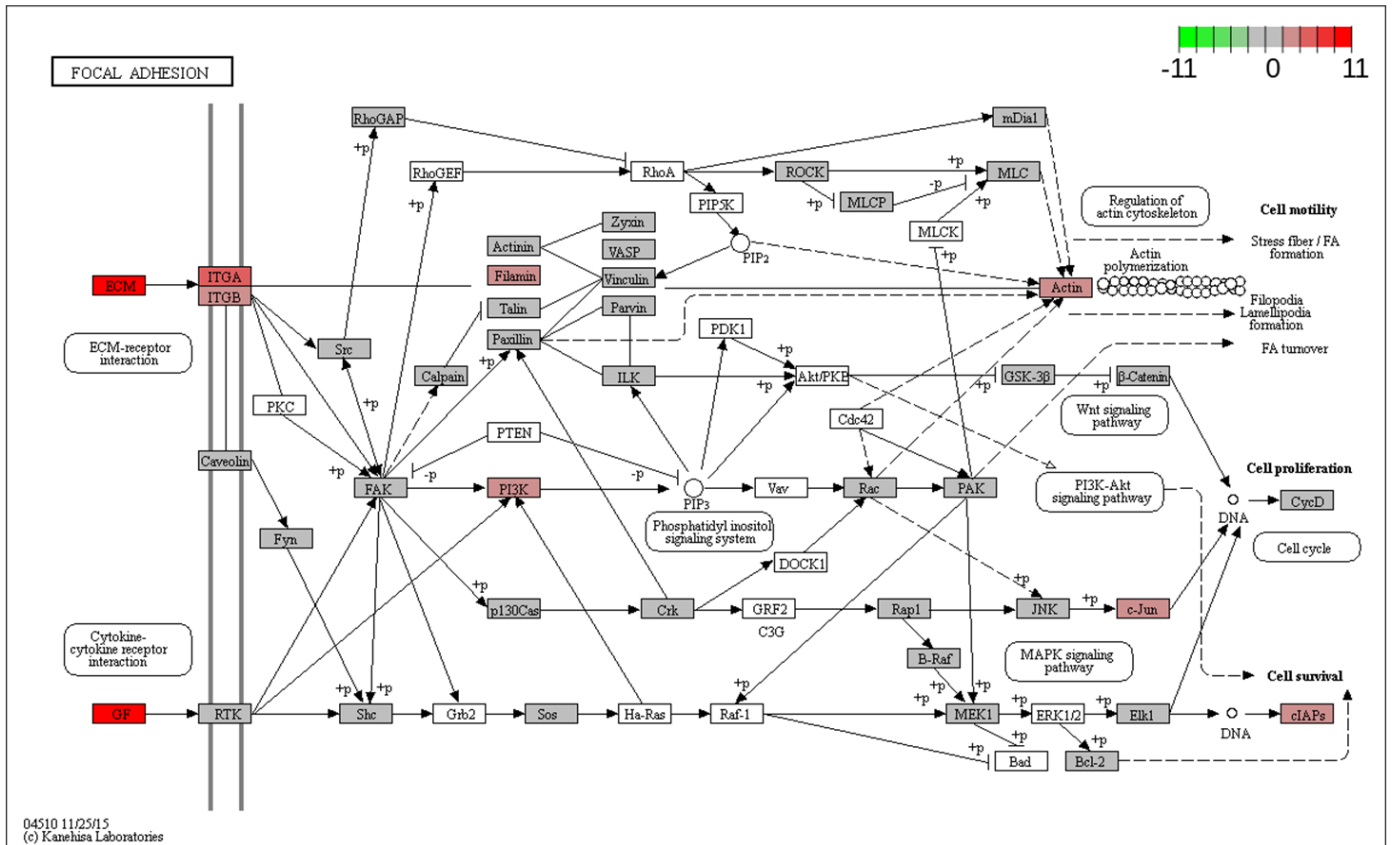
Supp Fig5

(A) Hippo signaling pathway activated in HSC-2 cell line following live *C. albicans* treatment

(B) Wnt signaling pathway activated in HSC-2 cell line following live *C. albicans* treatment

Pathways activated in HSC-2 cell line after live *C. albicans* treatment

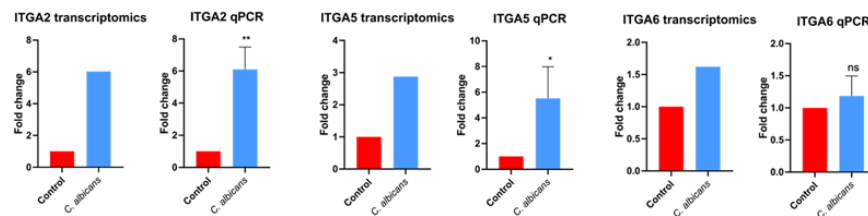
A



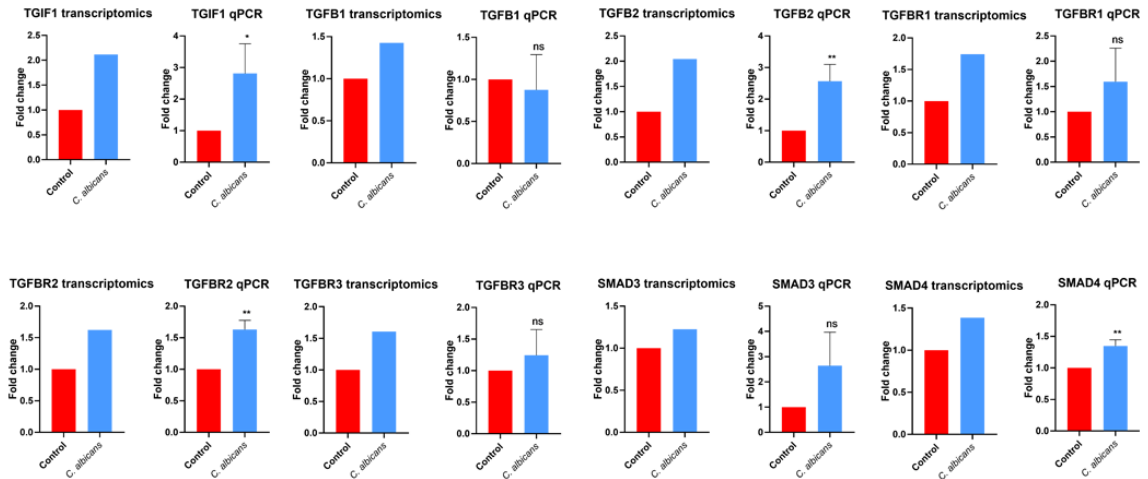
Supp Fig6

(A) Focal adhesion pathway activated in HSC-2 cell line following live *C. albicans* treatment

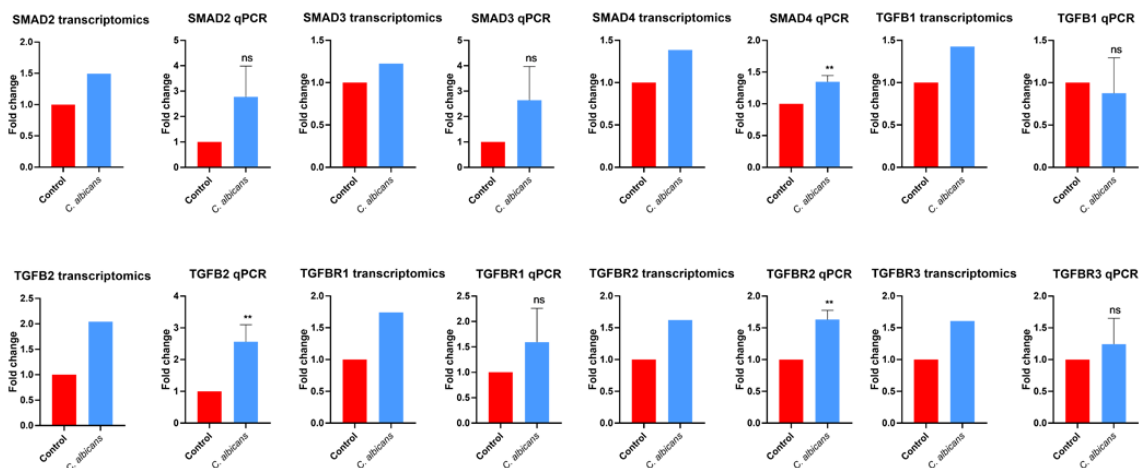
A: validation of PI3K-Akt pathway components by qPCR



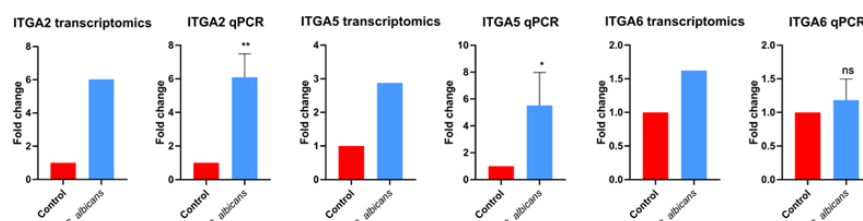
B: validation of TGFB/SMAD pathway components by qPCR



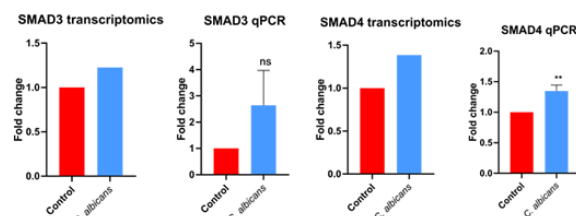
C: validation of HIPPO pathway components by qPCR



D: validation of Focal adhesion pathway components by qPCR



E: validation of Wnt pathway components by qPCR

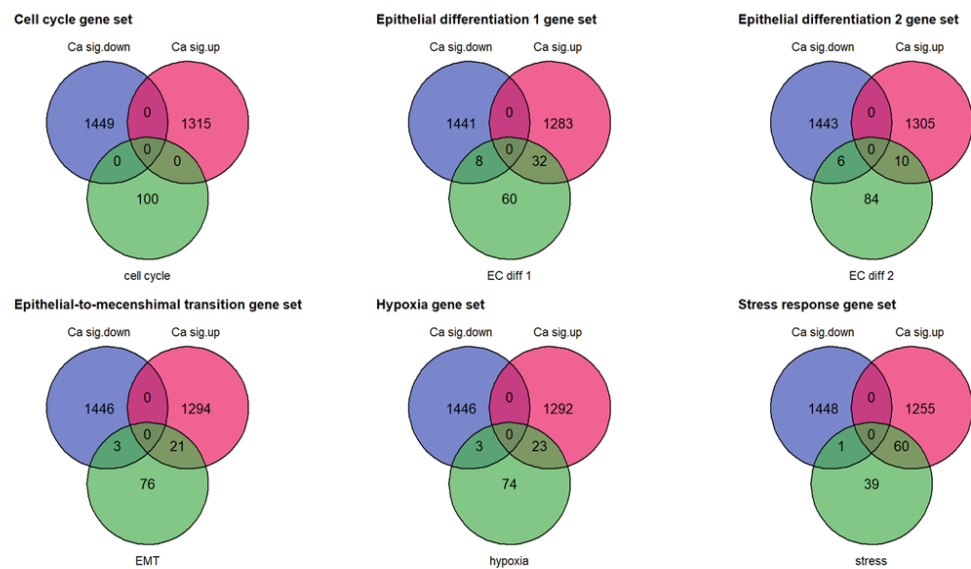


Supp Fig7

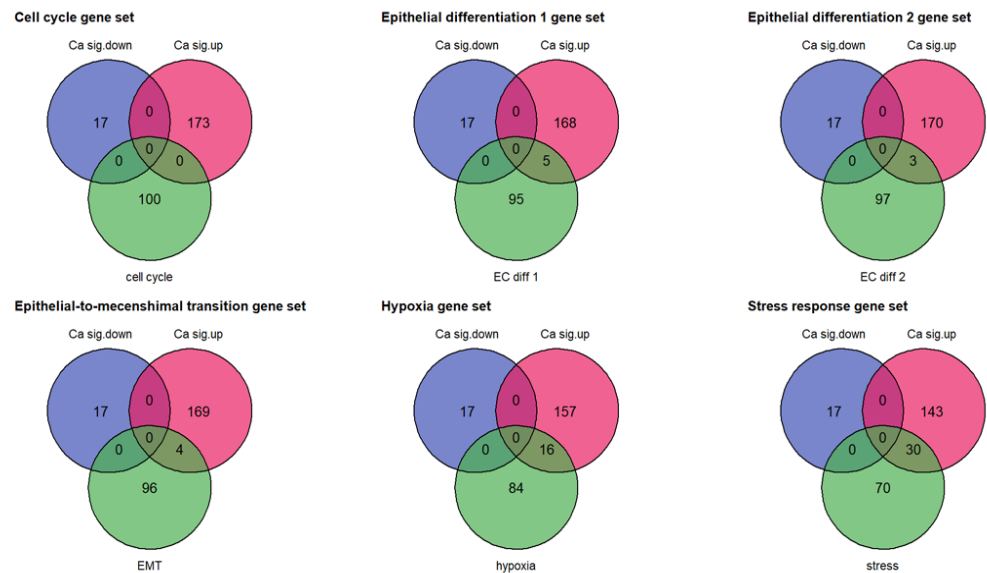
Validation of *C. albicans* activated signaling pathways. Validation was performed by qPCR analysis of pathway components.

(A) PI3K-Akt signaling pathway (B) TGF- β /SMAD (C) Hippo signaling pathway (D) Wnt signaling pathway
(E) Focal adhesion pathway

A HSC-2 cell line *C. albicans* treatment in vitro; scSec data vs. *C. albicans* induced gene set

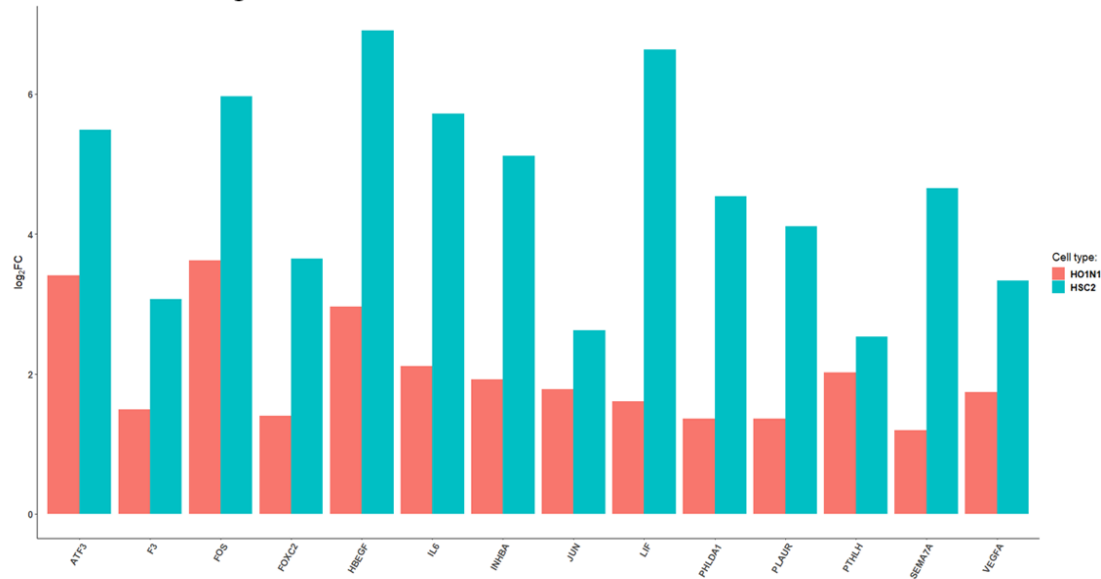


B HO-1-N-1 cell line *C. albicans* treatment in vitro; scSec data vs. *C. albicans* induced gene set



C

Live *Candida albicans* induced genes in both (HSC-2 and HO-1-N-1) cell lines, which are involved in OSCC progression



Supp Fig8

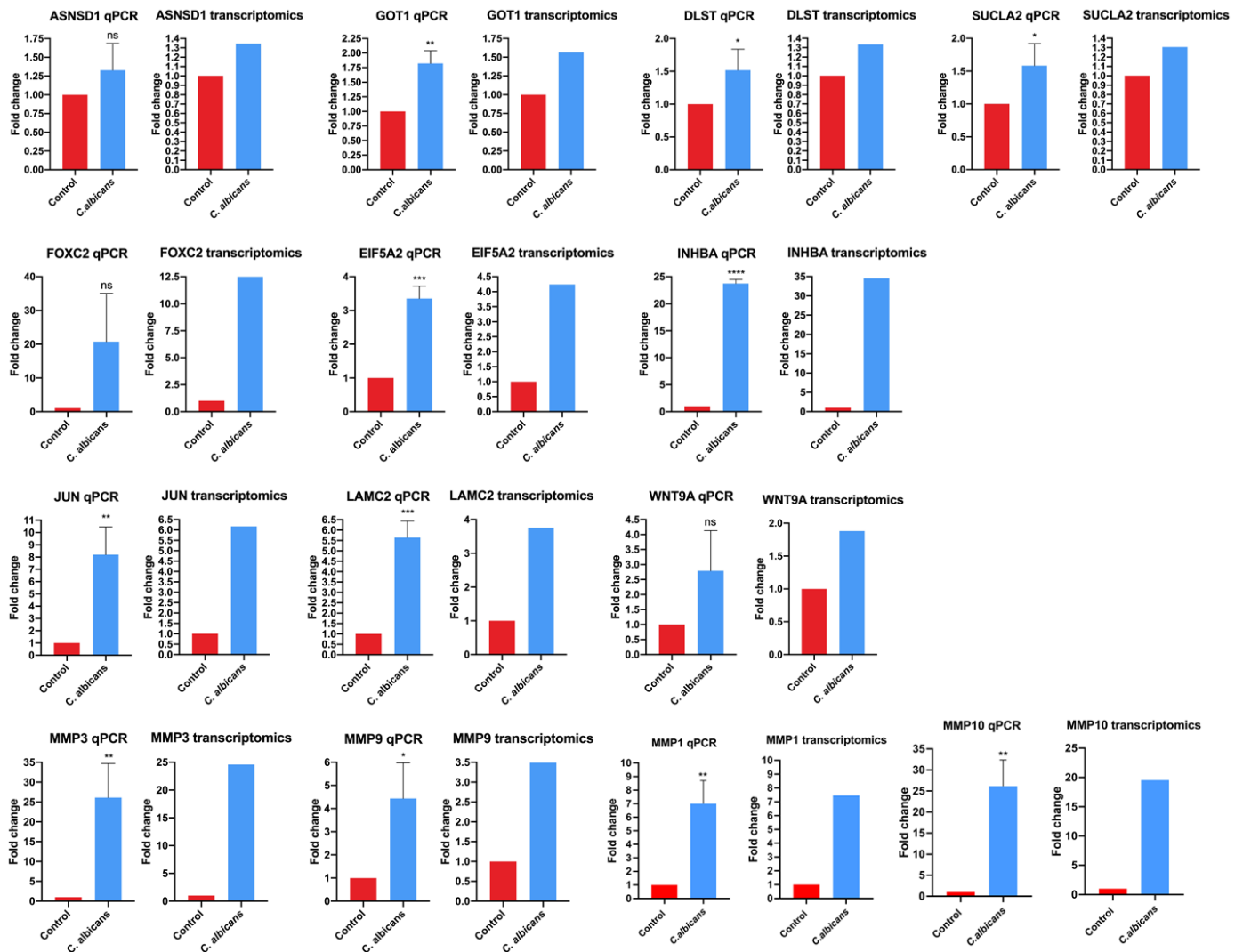
(A) Comparison of *Candida induced* genes in HSC-2 cell line to genes involved in different tumor progression processes. Differentially expressed gene list derived from an OSCC single cell sequencing study.

(B) Comparison of *Candida induced* genes in HO-1-N-1 cell line to genes involved in different tumor progression processes. Differentially expressed gene list derived from an OSCC single cell sequencing study.

(C) Live *C. albicans* induced genes in both (HSC-2 and HO-1-N-1) cell lines, which are involved in OSCC progression. OSCC progression marker gene list derived from literature and OSCC single cell sequencing study.

Validation of transcriptomics data by qPCR (HSC-2 cell line)

A



Supp Fig9

(A) Histopathological samples of *Candida*-colonized and *Candida*-free tumors, analysed and scored manually by a pathologist after H&E staining.

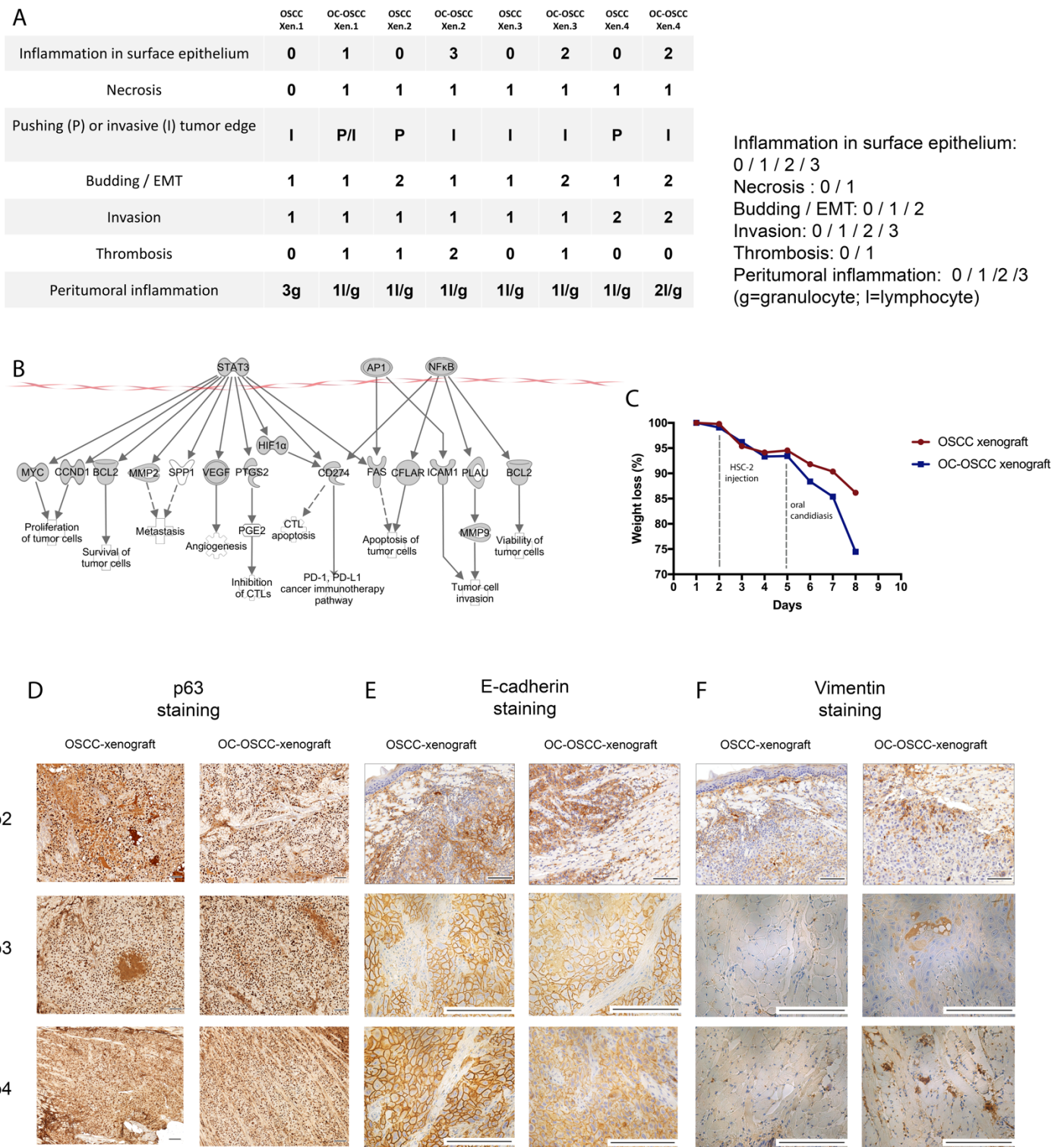
(B) Causal analyses of the genes which expression changed in OC-OSCC xenograft samples.

(C) Weight loss of the animal groups of OSCC xenograft and OC-OSCC xenograft.

(D) p63 staining of OSCC xenograft and OC-OSCC xenograft sections.

(E) E-cadherin staining of OSCC xenograft and OC-OSCC xenograft sections.

(F) Vimentin staining of OSCC xenograft and OC-OSCC xenograft sections.



Supp Fig10

(A) Scoring OSCC and OC-OSCC xenograft samples manually by a pathologist.

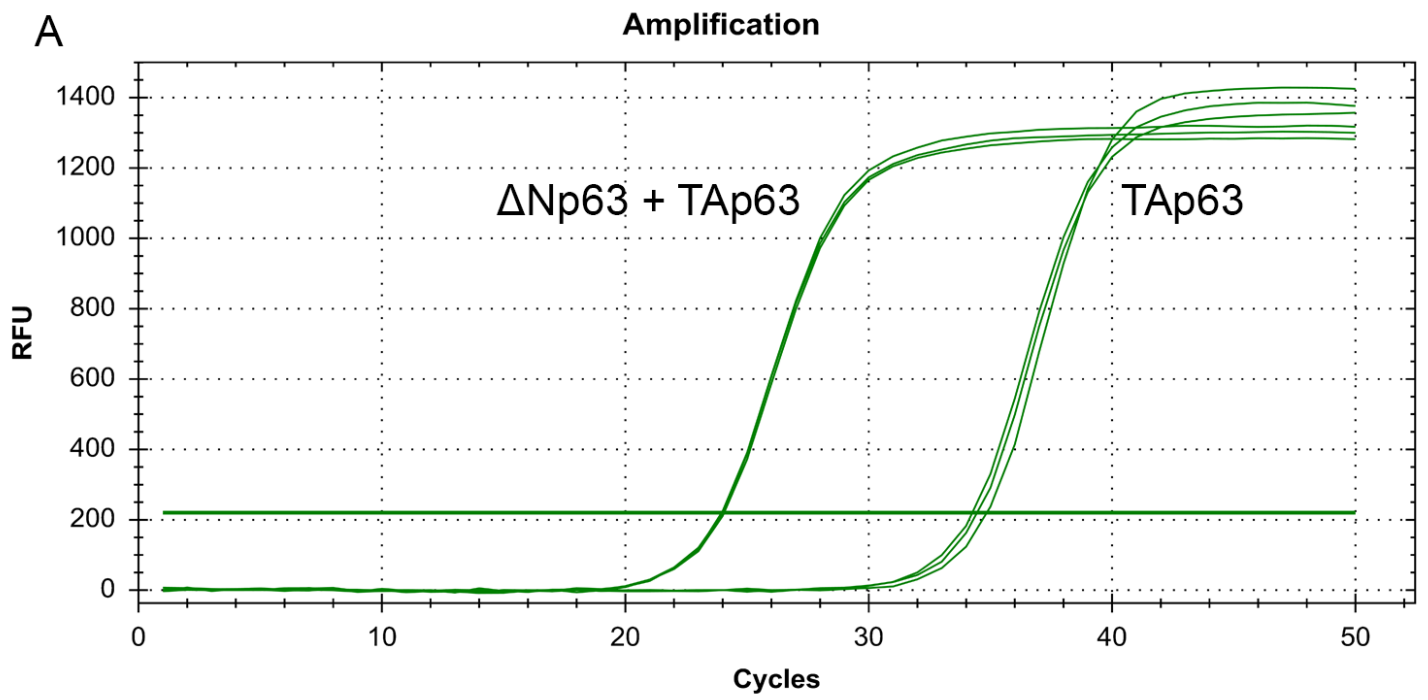
(B) Causal analyses of the genes which expression changed in OC-OSCC samples after oral candidiasis.

(C) Weight loss of the animals after HSC-2 tumor cell injection (OSCC xenograft) and HSC-2 injection combined with oral candidiasis (OC-OSCC xenograft).

(D) p63 staining of the histopathological samples (animal 2, 3, 4)

(E) E-cadherin staining of the histopathological samples (animal 2, 3, 4)

(F) vimentin staining of the histopathological samples (animal 2, 3, 4)



B

	qC
$\Delta Np63 + TAp63$	24.02
$\Delta Np63 + TAp63$	23.96
$\Delta Np63 + TAp63$	24.06
TAp63	34.45
TAp63	34.26
TAp63	34.85

Supp Fig11

(A) qPCR curve of TAp63 and $\Delta Np63 + TAp63$ splice variants. First primer pair was designed close to C-terminal region, second primer pair in the N-terminal region. First primer pair amplifies both splice variant group ($\Delta Np63 + TAp63$), second primer pair amplifies only splice variant possessing N-terminal region (TAp63).

(B) qC value of the transcript variants.

Supp Table 1

The precursor mass, fragment ion mass, polarity, retention time and fragmentation energy of the metabolites

Supp Table 2

Differentially expressed genes. *In vitro* and *in vivo* sequencing data.

Supp Table 3

qPCR primer list of the genes which were validated

Supp Table 4

120 genes derived from literature data (25 papers) which are involved in OSCC progression.