

# 1 **Assessing Confidence in Root Placement on Phylogenies:** 2 **An Empirical Study Using Non-Reversible Models for** 3 **Mammals**

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## 11 **ABSTRACT**

12 Using time-reversible Markov models is a very common practice in phylogenetic  
13 analysis, because although we expect many of their assumptions to be violated by empirical  
14 data, they provide high computational efficiency. However, these models lack the ability to  
15 infer the root placement of the estimated phylogeny. In order to compensate for the inability of  
16 these models to root the tree, many researchers use external information such as using outgroup  
17 taxa or additional assumptions such as molecular-clocks. In this study, we investigate the utility  
18 of non-reversible models to root empirical phylogenies and introduce a new bootstrap measure,  
19 the *rootstrap*, which provides information on the statistical support for any given root position.

20 Availability and implementation: rootstrap support is implemented in IQ-TREE 2 and a tutorial  
21 is available at the iqtree webpage <http://www.iqtree.org/doc/Rootstrap>. In addition, a python  
22 script is available at <https://github.com/suhanaser/Rootstrap>.

23 [phylogenetic inference, root estimation, bootstrap, non-reversible models]

24

25 MAIN TEXT

26         The most widely used method for rooting trees in phylogenetics is the outgroup method.  
27 Although the use of an outgroup to root an unrooted phylogeny usually outperforms other  
28 rooting methods (Huelsenbeck, et al. 2002), the main challenge with this method is to find an  
29 appropriate outgroup (Watrous and Wheeler 1981; Maddison, et al. 1984; Smith 1994;  
30 Swofford, et al. 1996; Lyons-Weiler, et al. 1998; Milinkovitch and Lyons-Weiler 1998).  
31 Outgroups that are too distantly-related to the ingroup may have substantially different  
32 molecular evolution than the ingroup, which can compromise accuracy. And outgroups that are  
33 too closely related to the ingroup may not be valid outgroups at all.

34         It is possible to infer the root of a tree without an outgroup using molecular clocks  
35 (Huelsenbeck, et al. 2002; Drummond, et al. 2006). A strict molecular-clock assumes that the  
36 substitution rate is constant along all lineages, a problematic assumption especially when the  
37 ingroup taxa are distantly related such that their rates of molecular evolution may vary.  
38 Relaxed molecular-clocks are more robust to deviations from the clock-like behaviour  
39 (Drummond, et al. 2006), although previous studies have shown that they can perform poorly  
40 in estimating the root of a phylogeny when those deviations are considerable (Tria, et al. 2017).

41         Other rooting methods rely on the distribution of branch lengths, including Midpoint  
42 Rooting (MPR) (Farris 1972), Minimal Ancestor Deviation (MAD) (Tria, et al. 2017), and  
43 Minimum Variance Rooting (MVR) (Mai, et al. 2017). Such methods also assume a clock-like  
44 behaviour; however, they are less dependent on this assumption as the unrooted tree is  
45 estimated without it. Similar to inferring a root directly from molecular-clock methods, the

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46 accuracy of those rooting methods decreases with higher deviations from the molecular-clock  
47 assumption (Mai, et al. 2017).

48 Other less common rooting methods that can be used in the absence of outgroup are:  
49 rooting by gene duplication (Dayhoff and Schwartz 1980; Gogarten, et al. 1989; Iwabe, et al.  
50 1989), indel-based rooting (Rivera and Lake 1992; Baldauf and Palmer 1993; Lake, et al.  
51 2007), rooting the species tree from the distribution of unrooted gene trees (Allman, et al. 2011;  
52 Yu, et al. 2011), and probabilistic co-estimation of gene trees and species tree (Boussau, et al.  
53 2013).

54 All the methods mentioned above, apart from the molecular-clock, infer the root  
55 position independently of the ML tree inference. The only existing approach to include root  
56 placement in the ML inference is the application of non-reversible models. Using non-  
57 reversible substitution models relaxes the fundamental assumption of time-reversibility that  
58 exists in the most widely used models in phylogenetic inference (Jukes and Cantor 1969;  
59 Kimura 1980; Hasegawa, et al. 1985; Tavaré 1986; Dayhoff 1987; Jones, et al. 1992; Tamura  
60 and Nei 1993; Whelan and Goldman 2001; Le and Gascuel 2008). This in itself is a potentially  
61 useful improvement in the fit between models of sequence evolution and empirical data. In  
62 addition, since non-reversible models naturally incorporate a notion of time, the position of the  
63 root on the tree is a parameter that is estimated as part of the ML tree inference. Since the  
64 incorporation of non-reversible models in efficient ML tree inference software is relatively new  
65 (Minh, et al. 2020), we still understand relatively little about the ability of non-reversible  
66 models to infer the root of a phylogenetic tree, although a recent simulation study has shown  
67 some encouraging results (Bettisworth and Stamatakis 2020).

68 Regardless of the rooting method and the underlying assumptions, it is crucial that we  
69 are able to estimate the statistical confidence we have in any particular placement of the root

70 on a phylogeny. A number of previous studies have sensibly used ratio likelihood tests such as  
71 the Shimodaira-Hasegawa (SH) test (Shimodaira and Hasegawa 1999) and the Approximately  
72 Unbiased (AU) test (Shimodaira 2002) to compare a small set of potential root placements,  
73 rejecting some alternative root placements in favour of the ML root placement e.g.(Nardi, et  
74 al. 2003; Steenkamp, et al. 2006; Jansen, et al. 2007; Moore, et al. 2007; Williams, et al. 2010;  
75 Kocot, et al. 2011; Zhou, et al. 2011; Whelan, et al. 2015; Zhang, et al. 2018), these tests are  
76 still somewhat limited in that they do not provide the level of support the data have for a certain  
77 root position.

78         There is strong empirical evidence that molecular evolutionary processes are rarely  
79 reversible (Squartini and Arndt 2008; Naser-Khdour, et al. 2019), but few studies have  
80 explored the accuracy of non-reversible substitution models to root phylogenetic trees  
81 (Huelsenbeck, et al. 2002; Yap and Speed 2005; Williams, et al. 2015; Cherlin, et al. 2018;  
82 Bettisworth and Stamatakis 2020). Most studies that have looked at this question in the past  
83 have focused on either simulated datasets (Huelsenbeck, et al. 2002; Jayaswal, et al. 2011;  
84 Cherlin, et al. 2018) or relatively small empirical datasets (Yang and Roberts 1995; Yap and  
85 Speed 2005; Jayaswal, et al. 2011; Heaps, et al. 2014; Williams, et al. 2015; Cherlin, et al.  
86 2018). In both cases, the addressed substitution models were nucleotide models, and to our  
87 knowledge, no study has yet investigated the potential of amino acid substitution models in  
88 inferring the root placement of phylogenetic trees.

89         In this paper, we focus on evaluating the utility of non-reversible amino acid and  
90 nucleotide substitution models to root the trees, and we introduce a new metric, the *rootstrap*  
91 *support value*, which estimates the extent to which the data support every possible branch as  
92 the placement of a root in a phylogenetic tree. Unlike previous studies that used Bayesian  
93 methods with non-reversible substitution models to infer rooted ML trees (Heaps, et al. 2014;  
94 Cherlin, et al. 2018), we will conduct our study in a Maximum likelihood framework using IQ-

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95 TREE (Minh, et al. 2020). A clear advantage of Maximum likelihood over the Bayesian  
96 analysis is that there is no need for a prior on the parameter distributions, which sometimes can  
97 affect tree inference (Huelsenbeck, et al. 2002; Cherlin, et al. 2018). Even though estimating  
98 the non-reversible model's parameters by maximizing the likelihood function seems more  
99 computationally intensive than calculating posterior probabilities (Huelsenbeck, et al. 2002),  
100 the IQ-TREE algorithm is sufficiently fast to allow us to estimate root placements, with  
101 *rootstrap support* for very large datasets.

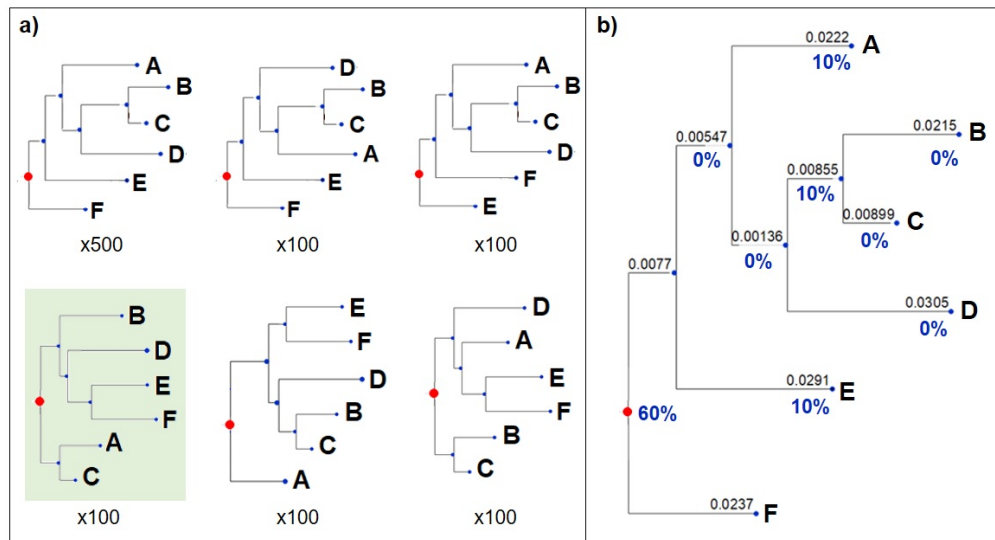
102         A recent study investigated the ability of non-reversible nucleotide models to infer the  
103 root placement of phylogenetic trees (Bettisworth and Stamatakis 2020). This study showed  
104 that IQ-TREE performs competitively with a new rooting tool, RootDigger. In most simulated  
105 datasets, IQ-TREE slightly outperformed RootDigger in terms of root placements, but no  
106 comparisons were made between RootDigger and IQ-TREE on empirical datasets. Although,  
107 RootDigger is significantly faster than IQ-TREE (Bettisworth and Stamatakis 2020), the  
108 former is limited to nucleotide substitution models. Since we are interested in both nucleotide  
109 and amino acid non-reversible models, we used IQ-TREE for tree and root inference in this  
110 study.

## 111 MATERIAL AND METHODS

### 112 *The “Rootstrap” Support, and measurements of error in root placement*

113         To compute rootstrap supports, we conduct a bootstrap analysis, i.e., resampling alignment  
114 sites with replacement, to obtain a number of bootstrap trees. We define the *rootstrap* support  
115 for each branch in the ML tree, as the proportion of bootstrap trees that have the root on that  
116 branch. Since the root can be on any branch in a rooted tree, the rootstrap support values are  
117 computed for all the branches including external branches. The sum of the rootstrap support  
118 values along the tree are always smaller than or equal to one. A sum that is smaller than one

119 can occur when one or more bootstrap replicates are rooted on a branch that does not occur in  
120 the ML tree (Fig. 1).



121

122 **FIGURE 1. Illustration of the rootstrap concept.** (a) The bootstrap replicates trees. (b)  
123 The ML tree with the rootstrap support values for each branch. Note that the sum of the  
124 rootstrap support values is less than 100% due to 100 bootstrap replicates trees (green)  
125 that have their root at a branch that does not exist in the ML tree.

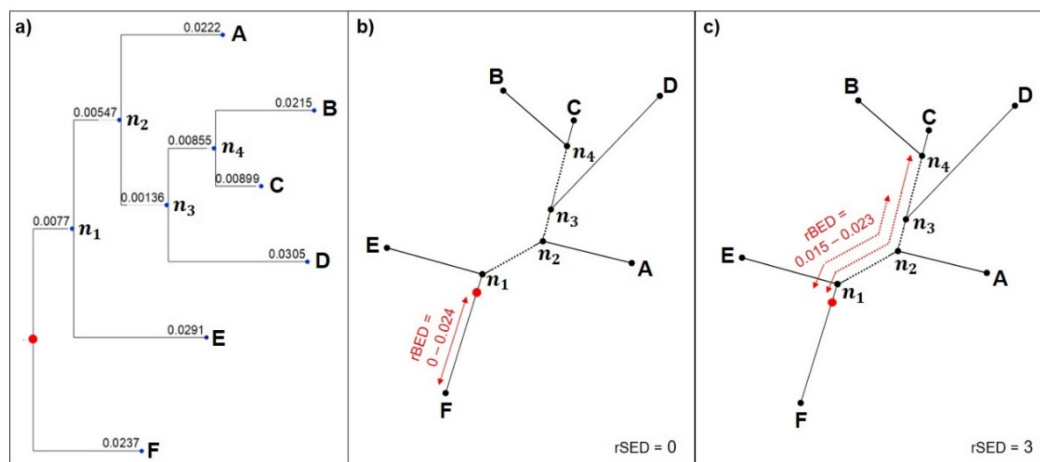
126

127 By definition, the rootstrap support values for internal branches are bounded by the  
128 bootstrap support values at those branches. On the other hand, the rootstrap support values for  
129 tips (leaf branches) are bounded by 100%, as tips always appear in all the bootstrap trees.

130 If the true position of the root is known (e.g. in simulation studies) or assumed (e.g. in  
131 the empirical cases we present below), we can calculate additional measurements of the error  
132 of the root placement. We introduce two such measurements here: *root branchlength error*  
133 *distance* (rBED) and *root split error distance* (rSED). Since the non-reversible model infers  
134 the exact position of the root on a branch, we define the *root branchlength error distance*  
135 (rBED) as the range between the minimum and maximum distance between the inferred root  
136 position and the “true root” branch. If the true root is on the same branch as the ML tree root,  
137 then rBED will be between 0 and the distance between the ML tree root and the farthest point

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138 on that branch (Fig. 2). Since rBED is based on branch lengths only, it ignores the absolute  
139 number of splits between the ML tree root and the true root; and therefore, the rBED for the  
140 true root being on the same ML root branch can be bigger than the rBED for the true root  
141 being on a different branch (e.g. Fig. 2). In order to account for the number of splits (nodes)  
142 between the ML tree root and the true root, we define *root split error distance* (rSED) as the  
143 number of splits between the ML root branch and the branch that is believed to contain the  
144 true root (Fig. 2).



145

146 FIGURE 2. An example to illustrate the root error distance. (a) the ML rooted tree, (b) the  
147 root branch-length error distance (rBED) if the true root is believed to be on the same ML  
148 root branch ( $rSED = 0$ ), (c) the rBED if the true root is believed to be on the branch between  
149 D and the clade of C + B ( $rSED = 3$ ).

150

151 The rootstrap, rBED, and rSED assess different aspects of the root placement. While the  
152 rootstrap offers an indication of the support that the data have for a certain branch to be the  
153 root branch, rBED and rSED provide an estimation to the accuracy of the method in  
154 estimating the exact root position if the root position is known or assumed in advance. In  
155 other words, the rootstrap value is a measure for the robustness of the root placement given  
156 the model and the data and can be used on any dataset regardless of whether the true root  
157 position is known, while rBED and rSED are measures of the accuracy of the non-reversible

158 model to find the root placement given the data, and require the root position to be known or  
159 assumed in advance.

### 160 *Empirical Datasets*

161 Because non-reversible amino acid models require the estimation of a large number of  
162 parameters, and because we suspected that the information in any such analysis on the  
163 placement of the root branch of a tree might be rather limited, we searched for empirical  
164 datasets that met a number of stringent criteria:

165 (1) Existence of both DNA and amino acid multiple sequences alignments (MSA) for the  
166 same loci.

167 (2) Genome-scale MSAs to ensure that the MSAs have as much information as possible with  
168 which to estimate the non-reversible models' free parameters and the root position. Since  
169 we do not know the number of sites required to correctly infer the rooted ML tree, we  
170 define 100,000 sites as the minimum number of required sites. This also allows us to  
171 subsample the dataset to explore the ability of smaller datasets to infer root positions.

172 (3) Highly-curated alignments: since the quality of the inferred phylogeny is highly  
173 dependent on the quality of the MSA (Philippe, et al. 2011), we focussed on datasets that  
174 were highly-curated for misalignment, contamination, and paralogy.

175 (4) Existence of several clades for which there is a very strong consensus regarding their root  
176 placement. Since we are interested in evaluating the performance of non-reversible  
177 models to infer root placements in an empirical rather than a simulation context, we need  
178 to identify monophyletic sub-clades for which we can be almost certain about their root  
179 position. This enables us to divide the dataset into non-overlapping sub-clades for which  
180 we are willing to assume we know the root positions. Furthermore, we define the  
181 minimum number of taxa in each sub-dataset as five.



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182 We initially identified a number of genome-scale datasets that contained large numbers of  
183 nucleotide and amino acid MSAs. In many cases, it was difficult to determine whether these  
184 alignments had been rigorously curated, and even more challenging to find datasets for which  
185 the root position of a number of subclades could be assumed with confidence. The only dataset  
186 that met all of our criteria was a dataset of placental mammals with 78 ingroup taxa and  
187 3,050,199 amino acids (Wu, et al. 2019). This dataset was originally published as an MSA  
188 (Liu, et al. 2017) based on very high-quality sequences from Ensembl, NCBI, and GenBank  
189 databases. After receiving detailed critiques for potential alignment errors (Gatesy and Springer  
190 2017), the dataset was further processed to remove potential sources of bias and error, and an  
191 updated version of the dataset was recently published (Wu, et al. 2018). The fact that this  
192 alignment comes from one of the most well-studied clades on the planet, has been  
193 independently curated and critiqued by multiple groups of researchers and includes truly  
194 genome-scale data, makes it ideally suited for our study. The curated alignments can be found  
195 on figshare (<https://figshare.com/s/622e9e0a156e5233944b>) under the name “Wu\_2018\_aa”  
196 and “Wu\_2018\_dna” for the amino-acid and nucleotide alignments respectively.

197

### 198 *Selecting Clades with a Well-Defined Root*

199 Since our main objective in this study is to evaluate the effectiveness of non-reversible  
200 models and the rootstrap value in estimating and measuring the support for a given root  
201 placement on empirical datasets, we must identify a collection of sub-clades of the larger  
202 mammal dataset for which it is reasonable to assume a root position. We acknowledge, of  
203 course, that outside a simulation framework it is not possible to be certain of the position of the  
204 root position of a clade. Nevertheless, it is possible to identify clades for which the position of  
205 the root is well supported and non-controversial, thus minimising the chances that the  
206 assumption of a particular root position is incorrect. To achieve this, we analysed the root

207 position of each order and superorder in the dataset, and defined “*well-defined clades*” that  
208 fulfilled **all** of the following criteria:

209 (1) It contains at least five taxa. This ensures that the probability of obtaining a random ML  
210 rooted tree to be at most 0.95%. For clades with four taxa, there are 15 different rooted  
211 topologies, and therefore a 6.7% probability to get any particular root position by chance.  
212 On the other hand, for clades with at least five taxa, there are at least 105 different rooted  
213 topologies and maximum probability of 0.95% to randomly get a particular root position  
214 by chance.

215 (2) The bootstrap support for the branch leading to that clade in the phylogenetic tree  
216 calculated from the whole dataset is 100%: since the bootstrap value indicates the  
217 support the data have for a certain branch, we also require 100% support for the first  
218 direct descendants in the clade (Appendix Fig. A.1). This requirement ensures that there  
219 is strong support in the dataset for the root position of the clade when the entire dataset is  
220 rooted with an outgroup.

221 (3) The site concordance factor (sCF) for the first direct descendants in the clade is  
222 significantly greater than 33%. The site Concordance Factor (sCF) is calculated by  
223 comparing the support of each site in the alignment for the different arrangements of  
224 quartet around a certain branch. In other words, an sCF of 33% means equal support for  
225 any of the possible arrangements. Therefore, we require that the sCF of the deepest two  
226 levels of branches leading to that clade is significantly greater than 33%. Moreover, we  
227 require that the gene Concordance Factor (gCF) for the first direct descendants in the  
228 clade to be significantly greater than 33% of the sum of the gene concordance factor and  
229 the two Discordance Factors (gDF1 and gDF2). The gCF of a branch is calculated as the  
230 proportion of gene trees containing that branch, and gDFs are calculated as the

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231 proportion of gene trees containing one of the two other resolutions of that branch. Since  
232 for each branch in a bifurcating tree there are three possible arrangements of clades  
233 around that branch, we ignore all gene trees that do not contain one of these  
234 arrangements (e.g. gene trees that contribute to neither the gCF nor the gDFs). Although  
235 there is no threshold regarding the required proportion of genes concordant with a certain  
236 branch, for convenience, we define branches with gCF significantly greater than 33% of  
237 the sum  $gCF + gDF1 + gCF2$  as branches that are concordant with enough genes in the  
238 alignment (Minh, et al. 2020). To test whether the sCF and the gCF are significantly  
239 greater than 33%, we use a simple binomial test with a success probability of 0.33. The  
240 gCF, gDF1, gCF2 and sCF values are based on the tree estimated from the amino acid  
241 dataset.

242 (4) At least 95% of the studies that have been published in the last decade support this clade:  
243 we searched google scholar for all published papers since 2009 that determine the root of  
244 the addressed clade. We then checked if at least 95% of those papers agree that the root  
245 position of the clade matches that in the ML tree we estimate from the whole dataset (see  
246 supplementary material).

### 247 *Estimating unrooted Phylogenies*

248 For the whole nucleotide and amino-acid datasets with ingroup and outgroup taxa, we  
249 inferred the unrooted phylogeny using IQ-TREE2 (Minh, et al. 2020) with the best-fit fully  
250 partitioned model (Chernomor, et al. 2016) and edge-linked substitution rates (Duchene, et al.  
251 2020). We then determined the best-fit reversible model for each partition using ModelFinder  
252 (Kalyaanamoorthy, et al. 2017). See the algorithm for finding well-defined clades in  
253 Appendix Algorithm A.1.

254 *Estimating Rooted phylogenies*

255           For each well-defined clade, we first removed all other taxa from the tree and then  
256 sought to infer the root of the well-defined clade using non-reversible models without  
257 outgroups. Using the best partitioning scheme from the reversible analysis, we inferred the  
258 rooted tree for each well-defined clade with the non-reversible models for amino acid (NR-  
259 AA) and nucleotide (NR-DNA) sequences (Minh, et al. 2020). This approach fits a 12-  
260 parameter non-reversible model for DNA sequences, and a 380-parameter non-reversible  
261 model for amino acids. Details of the command lines used are provided in the supplementary  
262 material section “Algorithm A.2”. Each analysis returns a rooted tree. We performed 1000  
263 non-parametric bootstraps of every analysis to measure the rootstrap support.

264           To assess the performance of the rootstrap and the ability of non-reversible models to  
265 estimate the root of the trees on smaller datasets, we also repeated every analysis on  
266 subsamples of the complete dataset. For each well-defined clade, we performed analysis on  
267 the complete dataset (100%) as well as datasets with 10%, 1% and 0.1% of randomly-  
268 selected loci from the original alignment.

269 *The confidence set of root branches using the Approximately Unbiased test*

270           In addition to the rootstrap support, we calculate the confidence set of all the branches  
271 that may contain the root of the ML tree using the Approximately Unbiased (AU) test  
272 (Shimodaira 2002). To do this, we re-root the ML tree with all possible placements of the root  
273 (one placement for each branch) and calculate the likelihood of each tree. Using the AU test,  
274 we then ask which root placements can be rejected in favour of the ML root, using an alpha  
275 value of 5%. We define the *root branches confidence set* as the set of root branches that are not  
276 rejected in favour of the ML root placement. An important difference between the AU test and  
277 the rootstrap support is that the AU test is conditioned on a single ML tree topology, but the  
278 rootstrap support is not. Because of this, they provide quite different information about the

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279 position of the root. The AU test assumes that the ML tree topology is true, and then seeks to  
280 determine the confidence set of root placements conditioned on that topology. The confidence  
281 set for the AU test will always therefore contain at least the ML root branch. The rootstrap does  
282 not assume any particular topology, and instead asks how many times a particular root position  
283 appears across a set of bootstrap replicates. Because of this, it is possible for every branch in  
284 the ML topology to receive 0% rootstrap support. This can occur if none of the branches in the  
285 ML topology appear as the root branch in any of the bootstrap topologies.

286 *Reducing systematic bias by removing third codon positions and loci that fail the MaxSym*  
287 *test*

288 As it is common in many phylogenetic analyses to remove third codon positions from  
289 the alignment (Swofford, et al. 1996), we wanted to assess the effect of removing third codon  
290 positions on the root inference and the rootstrap values in nucleotide datasets. For that  
291 purpose, we remove all the third codon positions from the nucleotide alignments and re-ran  
292 the analysis using the NR-DNA model.

293 Moreover, although the NR-AA and NR-DNA models relax the reversibility assumption,  
294 they still assume stationarity and homogeneity. To reduce the systematic bias produced by  
295 violating these assumptions, we used the MaxSym test (Naser-Khdour, et al. 2019) to remove  
296 loci that violate those assumptions in the nucleotide and amino acid datasets, and then re-ran  
297 all analyses as above.

298 *Applying the methods to two clades whose root position is uncertain*

299 In addition to the well-defined clades, we used the methods we propose here to infer  
300 the root of two clades of mammals whose root position is controversial; Chiroptera and the  
301 Cetartiodactyla.

302           There is a controversy around the root of the Chiroptera (bats) in literature. The two  
303 most popular hypotheses are: 1) the Microchiroptera-Megachiroptera hypothesis; where the  
304 root is placed between the Megachiroptera, which contains the family Pteropodidae, and the  
305 Microchiroptera, which contains all the remaining Chiroptera families. This hypothesis is  
306 well supported in the literature (Agnarsson, et al. 2011; Meredith, et al. 2011). However,  
307 more recent studies seem to provide less support for this hypothesis; 2) the  
308 Yinpterochiroptera-Yangochiroptera hypothesis, in which the Yangochiroptera clade includes  
309 most of Microchiroptera and the Yinpterochiroptera clade includes the rest of  
310 Microchiroptera and all of Megachiroptera. There is growing support for this hypothesis in  
311 the literature (Meganathan, et al. 2012; Tsagkogeorga, et al. 2013; Ren, et al. 2018; Reyes-  
312 Amaya and Flores 2019).

313           Similar to Chiroptera, the root of Cetartiodactyla remains contentious in the literature.  
314 The three main hypotheses regarding the root of Cetartiodactyla are: 1) Tylopoda as the sister  
315 group for all other cetartiodactylans; 2) Suina as the sister group for all other  
316 cetartiodactylans; 3) the monophyletic clade containing Tylopoda and Suina as the sister  
317 group for all other cetartiodactylans.

318           To ascertain whether certain sites or loci had very strong effects on the placement of  
319 the root we follow the approach of Shen et. al. (Shen, et al. 2017) and calculate the difference  
320 in site-wise log-likelihood scores ( $\Delta$ SLS) and gene-wise log-likelihood scores ( $\Delta$ GLS) between  
321 the supported root positions for each clade. Moreover, we analysed subsamples of each dataset  
322 to test the limits of using non-reversible models to root trees with smaller datasets.

323

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### 324 RESULTS

#### 325 *Inference of the mammal tree and selection of well-defined clades*

326 The trees inferred from the whole datasets with the nucleotide-reversible model and  
327 the amino-acid-reversible model (Appendix Fig. A.2, Appendix Fig. A.3, Appendix Table  
328 A.2) are consistent with the published tree (Liu, et al. 2017). Five clades met all the criteria of  
329 well-defined clades, namely, Afrotheria, Bovidae, Carnivora, Myomorpha, and Primates in  
330 both amino acid and nucleotide datasets (see Appendix Table A.1 and Appendix Table A.2).

331 Trees in Newick format can be found on github:

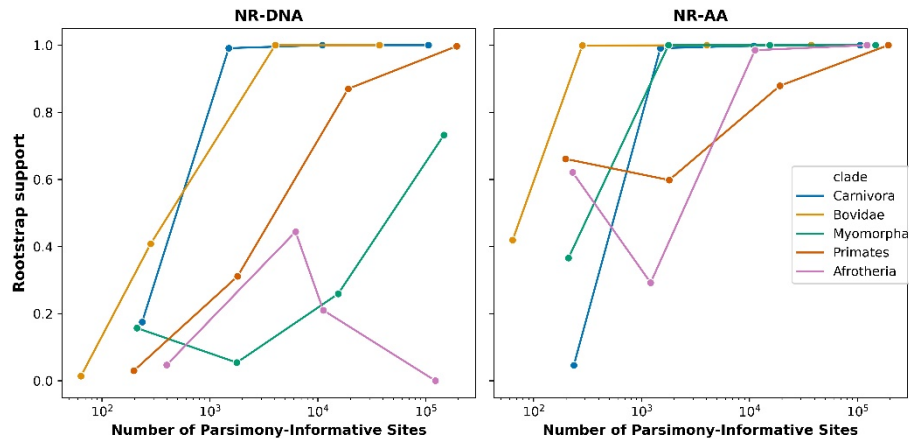
332 <https://github.com/suhanaser/Rootstrap/tree/master/trees>

#### 333 *High accuracy of the AA non-reversible model in inferring the root*

334 Using NR-AA, we inferred the correct root with very high rootstrap support for all  
335 five well-defined clades when all loci were used (Appendix Table A.3). Moreover, for all the  
336 five clades, the true root was the only root placement in the confidence set of the AU test.  
337 The average running time of the NR-AA model (model estimation + tree search + bootstrap +  
338 root inference) is 929 hrs on one core 2.6GHz CPU. However, using the optimal number of  
339 cores for each dataset reduced the average running time to 43.5 hrs per dataset.

340 Our results show that using only 10% of the sites in the amino acid alignments  
341 (around 300,000 alignment columns) still gave very high rootstrap support values (> 98%)  
342 for four of the five well-defined clades (Fig. 3) with no correlation between rSED and rBED  
343 and the size of the dataset (Table A.3). Moreover, in three of five well-defined clades, 1% of  
344 the sites (around 30,000 alignment columns) was enough to give a very high rootstrap  
345 support value for the assumed correct root placement. Using only 0.1% of the sites (around  
346 3000 alignment columns) decreased the rootstrap support value noticeably in all datasets  
347 (Appendix Table A.3). These values are shown for each dataset in Figure 3, where the X-axis

348 is plotted in terms of parsimony-informative sites to allow for a more direct comparison  
349 between datasets, and to assist those applying these methods in deciding whether to use them  
350 on their own data. Although the rootstrap support for the true root improves as the number of  
351 parsimony-informative sites increase, in some datasets (e.g. Afrotheria nucleotide dataset)  
352 this is not the case (Fig. 3).



353  
354 FIGURE 3. The rootstrap support value for each clade as a function of the number of  
355 parsimony-informative sites.

356  
357 The non-reversible amino acid models were strongly preferred to the reversible models on the  
358 complete datasets (BIC values were 93943 to 235958 units better for the non-reversible  
359 models), and for the datasets with 10% of loci subsampled (BIC values were 3577 to 15082  
360 units better for the non-reversible models), but the opposite was true for the datasets 1% and  
361 0.1% of the loci subsampled (e.g. BIC values were between 2102 and 2712 units worse for  
362 the non-reversible models for the 0.1% subsampled datasets; see Table A.7 for full results).

363  
364 *Poor performance of the DNA non-reversible model in inferring the root*

365 We correctly inferred the root for four out of the five nucleotide datasets with the NR-  
366 DNA model, when all loci were used. However, the rootstrap support was generally lower  
367 than in the amino-acid datasets (Fig. 3, Appendix Tables A.3 and A.4). Similar to amino-acid  
368 datasets, there is no correlation between rSED and rBED and the size of the dataset (Table



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369 A.4). The average running time of the NR-DNA model (model estimation + tree search +  
370 bootstrap + root inference) is 35.7 hrs on one core 2.6GHz CPU and 4 hours when the  
371 optimal number of cores for each dataset were used.

372 In contrast to the NR-AA model, there is no conclusive preference for the NR-DNA  
373 model over the reversible DNA model for the datasets we analysed (Table A.8). In fact, The  
374 BIC values of the NR-DNA models are always worse than reversible models regardless to the  
375 size of the nucleotide dataset except for three clades when all loci were included (Table A.8).  
376 In two of the datasets (Myomorpha and Primates) were the NR-DNA model was better than  
377 the reversible model the root placement was inferred correctly with high rootstrap support  
378 (>95%). In fact, the Afrotheria nucleotide dataset is the only dataset in which the non-  
379 reversible model was better than the reversible model but the root placement was inferred  
380 incorrectly.

381 Our results show that removing the third codon positions does not improve the  
382 rootstrap support value. In contrast, in some datasets removing third codon positions  
383 decreased the rootstrap support value and increased the rSED (Table 1).

384 TABLE 1. Rootstrap support and rSED values in whole nucleotide datasets and  
385 nucleotide datasets without third codon positions.

Clades	All loci		Without 3rd	
	rootstrap	rSED	rootstrap	rSED
Afrotheria	0.0%	2	0.0%	2
Primates	99.7%	0	90.1%	0
Myomorpha	73.2%	0	15.8%	1
Carnivora	100.0%	0	100.0%	0
Bovidae	100.0%	0	82.5%	0

386

387 *Removing loci that violate the stationarity and homogeneity assumptions improves the*  
388 *rootstrap support*

389 As expected, our results show that removing loci that fail the MaxSym test improves  
390 the rootstrap support values when the rootstrap support value was less than 100% and/or the  
391 root placement was inferred incorrectly, as the case in some nucleotide datasets (Table 2).

392 TABLE 2. Rootstrap support values in whole datasets and datasets with loci that passed  
393 the MaxSym test only.

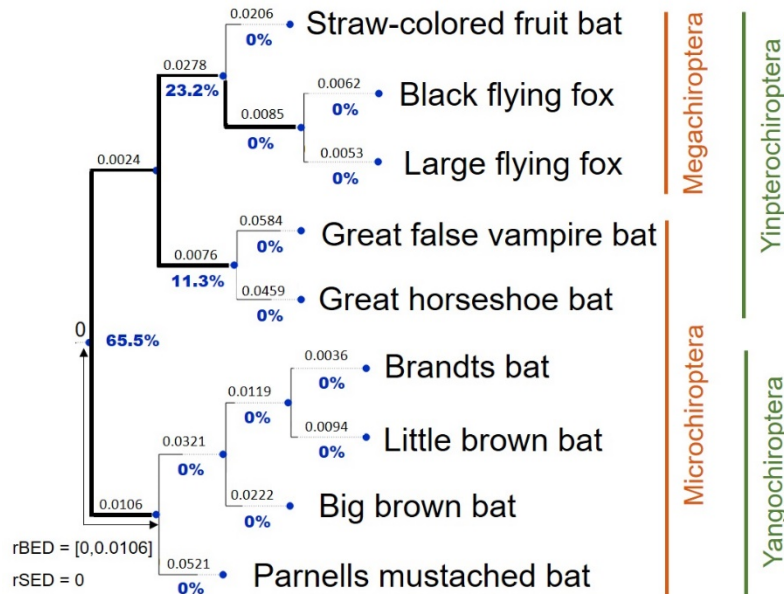
Clade	Amino Acid		Nucleotide	
	all loci	Passed MaxSym	all loci	Passed MaxSym
Afrotheria	100.0%	100.0%	0.0%	8.4%
Primates	100.0%	100.0%	99.7%	99.9%
Myomorpha	100.0%	100.0%	73.2%	88.3%
Carnivora	100.0%	100.0%	100.0%	100.0%
Bovidae	100.0%	100.0%	100.0%	100.0%

394

395 *Microchiroptera-Megachiroptera or Yinpterochiroptera-Yangochiroptera?*

396 Using the whole amino acid dataset, our results show 65.5% rootstrap support for the  
397 Yinpterochiroptera-Yangochiroptera hypothesis and 23.2% for the Microchiroptera -  
398 Megachiroptera hypothesis. The remaining 11.3% of the rootstrap support goes to supporting  
399 the branch leading to Rhinolophoidea as root branch of the bats (Fig. 4). Removing amino  
400 acid loci that fail the MaxSym test (110 loci) gives similar results, with 65.9% rootstrap  
401 support for the Yinptero-Yango hypothesis and 25.6% rootstrap support for the Micro-Mega  
402 hypothesis. In both cases, the AU test could not reject any of the three root positions that  
403 received non-zero rootstrap support (Appendix Table A.5).

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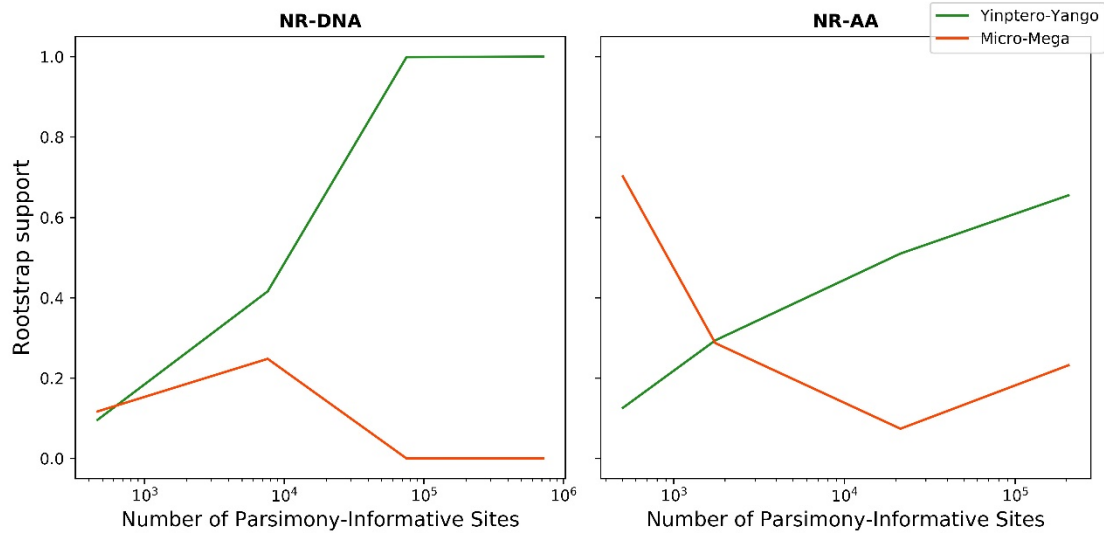


404

405 FIGURE 4. The ML rooted tree as inferred from the whole Chiroptera amino acid dataset.  
 406 Bold branches are branches in the AU confidence set. Blue values under each branch are the  
 407 rootstrap support values.

408 Using the NR-DNA model gives 100% rootstrap support for the Yinptero-Yango  
 409 hypothesis, and we can confidently reject the Micro-Mega hypothesis in favour of the  
 410 Yinptero-Yango hypothesis using the AU test (Appendix Fig. A.4). Yet, removing nucleotide  
 411 loci that fail the MaxSym test (~25% of the loci) decreases the support for the Yinptero-  
 412 Yango hypothesis to 90.1%, although we can still confidently reject the Micro-Mega  
 413 hypothesis using the AU test (Appendix Table A.5).

414 Interestingly, when we randomly subsample 10%, 1%, and 0.1% of the loci in the  
 415 nucleotide dataset, we consistently get the Yinptero-Yango hypothesis as the ML tree and the  
 416 solely rooted topology in the AU confidence set (Appendix Table A.5). Moreover, the  
 417 rootstrap support value for the Yinptero-Yango hypothesis increases and the rootstrap support  
 418 value for the Micro-Mega hypothesis decreases as more parsimony-informative sites are  
 419 added to the alignment, for both nucleotide and amino acid datasets (Fig. 5, Appendix Table  
 420 A.5). These results are consistent with previous studies that used smaller datasets (Appendix  
 421 Figure A.10)



422

423 FIGURE 5. Rootstrap support value as a function of the number of parsimony-informative  
424 characters in the Chiroptera nucleotide and amino acid datasets using the NR-DNA model (to  
425 the left) and the NR-AA model (to the right).

426 The  $\Delta$ GLS and  $\Delta$ SLS values (Shen, et al. 2017) reveal that approximately half of the  
427 nucleotide and amino acid loci prefer the Yinptero-Yango hypothesis while the other half  
428 prefers Micro-Mega hypothesis. Furthermore, slightly less than half of the nucleotide sites  
429 prefer the Yinptero-Yango hypothesis. However, more than two-thirds of the amino acid sites  
430 prefer the Yinptero-Yango hypothesis (Appendix Fig. A.5). The distributions of  $\Delta$ GLS and  
431  $\Delta$ SLS (Appendix Fig. A.6) show that a small proportion of the amino acid loci (~1%) have  
432 very strong support for the Micro-Mega hypothesis, and removing those loci from the  
433 alignment increased the rootstrap support for the Yinptero-Yango hypothesis to 76.6%.  
434 Nonetheless, both root placements are still in the confidence set of the AU test (Appendix  
435 Table A.5) with the amino acid dataset. On the other hand, removing nucleotide loci with the  
436 highest absolute  $\Delta$ GLS value still gives the Yinptero-Yango hypothesis as the ML tree and  
437 the sole topology in the AU confidence set. Although the nucleotide data show a clear  
438 preference to the Yinptero-Yango hypothesis, in terms of BIC scores, the NR-DNA model  
439 performs worse than reversible models in all datasets except for the dataset where we  
440 removed loci that failed the MaxSym test (Table A.5). On the other hand, the NR-AA

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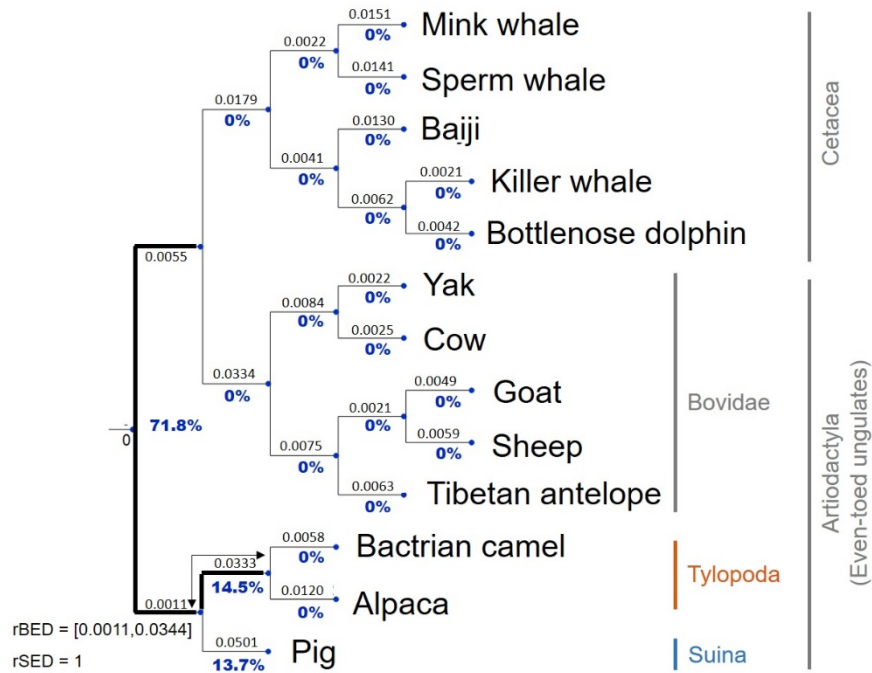
441 performs better than reversible models in big datasets (Table A.5). Yet, the amino acid data  
442 do not allow us to distinguish between the two leading hypotheses for the placement of the  
443 root of the Chiroptera based on rooting with non-reversible models (Table A.5).

### 444 *The ambiguous root of Cetartiodactyla*

445 The ML tree inferred with the whole amino acid dataset places the clade containing  
446 Tylopoda (represented by its only extant family; Camelidae) and Suina as the sister group to  
447 all other cetartiodactylans with 71.8% rootstrap support (Fig. 6). Yet, The AU test did not  
448 reject Tylopoda alone as the sister group to all other cetartiodactylans. On the other hand, the  
449 ML tree inferred with the whole nucleotide dataset places Tylopoda as the only sister group  
450 to all other cetartiodactylans with 71.0% rootstrap support, and we can confidently reject the  
451 Tylopoda + Suina hypothesis using the AU test (Appendix Fig. A.7).

452 Removing the amino acid loci that failed the MaxSym test (~1%) still places Tylopoda +  
453 Suina as the sister group to all other cetartiodactylans, yet, it decreases the rootstrap support  
454 for the Tylopoda + Suina hypothesis to 63.3% and increases the rootstrap support for the  
455 Tylopoda hypothesis to 28.5%. However, we still cannot reject either of the hypotheses using  
456 the AU test (Appendix Table A.6).

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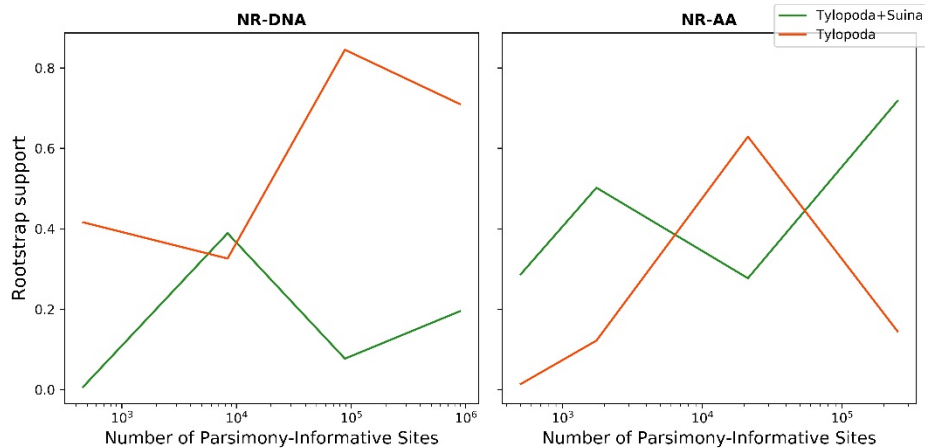
457

458 FIGURE 6. The ML rooted tree of as inferred from the whole Cetartiodactyla amino acid  
 459 dataset. Bold branches are branches in the AU confidence set. Blue values under each branch  
 460 are the rootstrap support values.

461 Removing the nucleotide loci that failed the MaxSym test (~1%) still places Tylopoda  
 462 as the only sister group to all other cetartiodactylans and the only rooted topology in the AU  
 463 confidence set. However, it decreases the rootstrap support for the Tylopoda hypothesis to  
 464 68.7% and increases the rootstrap support for the Tylopoda + Suina hypothesis to 20.1%  
 465 (Appendix Table A.6).

466 The results from the subsample datasets are mixed (Fig. 7). Analyses on smaller datasets  
 467 show no clear pattern in the placement of the root (Appendix Table A.6), leading us to  
 468 conclude only that the analyses of the whole dataset is likely to provide the most accurate  
 469 result, but that it is plausible that adding more data may lead to different conclusions in the  
 470 future.

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471

472 FIGURE 7. rootstrap support value as a function of the number of parsimony-informative  
473 characters in the Cetartiodactyla nucleotide and amino acid datasets using the NR-DNA  
474 model (to the left) and the NR-AA model (to the right).

475  $\Delta$ GLS analyses reveal that approximately, half of the amino acid and nucleotide loci  
476 favour the Tylopoda+Suina hypothesis, while the other half of loci favour the Tylopoda  
477 hypothesis (Appendix Figs. A.8-9). On the other hand, two-thirds of the amino acid sites and  
478 more than 80% of the nucleotide sites favour the Tylopoda+Suina hypothesis. Removing 1%  
479 of the amino acid loci with the highest absolute  $\Delta$ GLS values still places Tylopoda + Suina as  
480 the sister group to all other cetartiodactylans. However, the rootstrap support of the Tylopoda  
481 + Suina decreased to 63.2% and the rootstrap support for the Tylopoda hypothesis remains  
482 approximately the same (~14.5%), while the rootstrap support for the Suina hypothesis  
483 increases from 13.7% to 22.4%. Yet, both the Tylopoda + Suina hypothesis and the Tylopoda  
484 hypothesis are in the confidence set of the AU test, while the Suina hypothesis is rejected by  
485 the AU test (Appendix Table A.6).

486 Removing 1% of the nucleotide loci with the highest absolute  $\Delta$ GLS values gives the  
487 Tylopoda+Suina as the sister group to all other cetartiodactylans with 39.7% rootstrap  
488 support. However, the solely rooted topology in the AU confidence set is the topology in  
489 which the root is placed on the branch leading to Suina (Appendix Table A.6). Similar to  
490 Chiroptera and the well-defined clades, NR-AA model preforms, in terms of the BIC score,

491 better than reversible models in big amino-acid datasets, while the NR-DNA performs worse  
492 than reversible models in all datasets (Table A.6). We conclude that neither the nucleotide  
493 nor the amino acid data are adequate to infer the root placement of Cetartiodactyla with non-  
494 reversible models.

## 495 DISCUSSION

496 In this paper, we introduced a new measure of support for the placement of the root in  
497 a phylogenetic tree, the rootstrap support value, and applied it to empirical amino acid and  
498 nucleotide datasets inferred using non-reversible models implemented in IQ-TREE (Minh, et  
499 al. 2020). The rootstrap is a useful measure because it can be used to assess the statistical  
500 support for the placement of the root in any rooted tree, regardless of the rooting method. In a  
501 Maximum Likelihood setting, interpretation of the rootstrap support is similar to the  
502 interpretation of the classic nonparametric bootstrap. In a Bayesian setting, the same  
503 procedure could be used to calculate the posterior probability of the root placement given a  
504 posterior distribution of trees. It is noteworthy that the rootstrap support value is not a  
505 measure of the accuracy of the root placement and therefore should not be interpreted as  
506 such. However, it provides information about the robustness of the root inference with regard  
507 to resampling the data. This interpretation is consistent with the interpretation of the  
508 nonparametric bootstrap (Holmes 2003) but with regard to the root placement instead of the  
509 whole tree topology.

510 In addition to the rootstrap support value, we introduced another two metrics; the root  
511 branch-length error distance (rBED), and the root split error distance rSED. Similar to the  
512 rootstrap metric, these additional metrics can be used in with any approach that generates  
513 rooted phylogenetic trees. We note that both metrics require the true position of the root to be



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514 known (or assumed) and that the rBED requires the rooting method to be able to accurately  
515 place the root in a specific position of the root branch.

516 In this study, we used these and other methods to assess the utility of non-reversible  
517 models to root phylogenetic trees in a Maximum Likelihood framework. We focussed on  
518 applying these methods to a large and very well curated phylogenomic dataset of mammals,  
519 as the mammal phylogeny provides perhaps the best opportunity to find clades for which the  
520 root position is known with some confidence. As expected, our results show an exponential  
521 increase in the rootstrap support for the true root as we add more information to the MSA.  
522 Our results suggest that non-reversible amino-acid models are more useful for inferring root  
523 positions than non-reversible DNA models. One explanation for this difference between the  
524 NR-DNA and the NR-AA models is the bigger character-state space of the NR-AA models.  
525 These models have 400 parameters (380 rate parameters and 20 amino acid frequencies)  
526 whereas NR-DNA models have only 16 parameters (12 rate parameters and 4 nucleotide  
527 frequencies). This could allow the NR-AA model to capture the evolutionary process better  
528 than the NR-DNA model, potentially providing more information on the root position of the  
529 phylogeny. This hypothesis requires some further exploration though, and we note that the  
530 actual character-space of amino acids is much smaller than accommodated in NR-DNA  
531 models due to functional constraints on protein structure (Dayhoff, et al. 1978).

532 Another explanation for the difference in performance between the NR-AA and NR-  
533 DNA models is that higher compositional heterogeneity in nucleotide datasets may bias tree  
534 inference. The fact that each amino acid can be specified by more than one codon, and that  
535 synonymous substitutions are more frequent than non-synonymous substitutions, makes  
536 amino acid datasets less compositionally heterogeneous than nucleotide datasets. In principle,  
537 this bias can be alleviated by removing loci that violate the stationarity and homogeneity  
538 assumptions (Naser-Khdour, et al. 2019). Our results suggest that this may be the case for the

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539 datasets we analysed: we show that removing loci that violate the stationarity and  
540 homogeneity assumptions improves the accuracy and statistical support for the placement of  
541 the root. This is not surprising since the robustness of the rootstrap, similar to the bootstrap,  
542 relies on the consistency of the inference method, so removing systematic bias should  
543 improve its performance.

544         We used the non-reversible approach to rooting trees along with the rootstrap support  
545 to assess the evidence for different root placements in the Chiroptera and Cetartiodactyla.  
546 Using the amino acid datasets we found that in both cases, although there tended to be higher  
547 rootstrap support for one hypothesis, neither of the current hypotheses for either dataset could  
548 be rejected. These results emphasize the importance of the rootstrap support value as a  
549 measure of the robustness of the root estimate given the data. In both the Chiroptera and  
550 Cetartiodactyla datasets the root placement varied among subsamples of the dataset, and the  
551 rootstrap support reflects this uncertainty. However, checking the stability of root placement  
552 estimate by randomly subsampling from the whole Chiroptera dataset show an obvious trend  
553 towards the Yinpterochiroptera-Yangochiroptera hypothesis as the dataset increases in size.  
554 This trend is consistent with a small number of influential sites or loci having their signal  
555 progressively drowned out in favour of the Yinpterochiroptera-Yangochiroptera hypothesis  
556 as more data are added to the alignment. In both the Chiroptera and Cetartiodactyla cases, the  
557 amino acid data is inadequate to distinguish between certain root placements. On the other  
558 hand, in both the Chiroptera and Cetartiodactyla, the nucleotide datasets appear to show  
559 stronger support for a single root placement.

560         Comparing BIC scores of reversible and non-reversible models show that in most of  
561 the nucleotide datasets the reversible model was a much better fit to the data than the NR-  
562 DNA model. This is likely due to the limitations of the method we used to infer the NR-DNA  
563 model. Specifically, when inferring the trees with reversible DNA models, we used a

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564 partitioned model such that each partition was able to have an independent DNA substitution  
565 model. On the other hand, when we inferred the NR-DNA model we estimated a single  
566 model for the entire alignment. Thus, the NR-DNA model we inferred was unable to account  
567 for heterogeneity in the evolutionary process among partitions, possibly leading to its worse  
568 fit to the data when assessed using BIC scores. This suggests that using either mixture models  
569 or partitioned models may improve the fit of non-reversible DNA models to the data. The  
570 DNA results are consistent with results from previous study using the NR-DNA model and  
571 RootDigger (Bettisworth and Stamatakis 2020), although that study did not compare the  
572 performance of IQ-TREE and RootDigger on empirical datasets. Its results indicate that the  
573 NR-DNA model in IQ-TREE could not infer the correct root placement for any of the three  
574 tested datasets.

575         Our results demonstrate that the amino-acid non-reversible model can often be  
576 surprisingly accurate for inferring the root placement of phylogenies in the absence of  
577 additional information (such as outgroups) or assumptions (such as molecular clocks). In all  
578 of the well-defined clades that we examined, the non-reversible amino-acid model  
579 successfully identified the root that we identified a-priori as correct, and with very high  
580 rootstrap support. Importantly, the non-reversible amino-acid models also tended to fit the  
581 data far better than their reversible counterparts did. Indeed, we show that root placements  
582 appear to be accurate even with datasets as small as 50 well-curated loci between fairly  
583 closely-related taxa such as orders of mammals. Nevertheless, the application of the non-  
584 reversible amino acid models to two clades where the root position has previously been  
585 contentious failed to shed much additional light on the true root placement. Thus, while we  
586 show that the use of non-reversible models certainly has promise, we also show that it is no  
587 silver bullet.

588           Where a reliable outgroup taxon can be found, without the issues that can confound  
589 the inference of root placements using outgroups (Dalevi, et al. 2001; Braun and Kimball  
590 2002; Graham, et al. 2002; Brady, et al. 2006), we suggest relying on the use of outgroups.  
591 Nevertheless, where no reliable outgroups can be found, or where there is some reason to  
592 question the position of a root inferred using an outgroup (e.g. references about questionable  
593 outgroup rooting), our study suggests that using non-reversible models can provide a useful  
594 additional line of evidence for the position of the root of a phylogeny. We note also that the  
595 rootstrap value and the AU test could be used to provide estimates of the uncertainty of root  
596 placement using an outgroup taxon

597           Our work suggests a practical approach to inferring the root of a phylogenetic tree  
598 using non-reversible models. First, estimate an unrooted tree topology using the best  
599 reversible models available, excluding outgroup sequences. Next, fix the tree topology and  
600 use the best non-reversible models available to infer the Maximum Likelihood (ML) root  
601 position of that tree. Finally, determine to what extent the ML root position should be trusted.  
602 The degree of trust that researchers should put in an inferred ML root position should be  
603 influenced by three factors (noting of course that all phylogenetic inferences are susceptible  
604 to be misled by model misspecification). First, the fit of the non-reversible model to the data  
605 should be better than the fit of the reversible model. This can be assessed using common  
606 criteria like AICc or BIC scores. A better fit of the non-reversible model provides some  
607 assurance that the data contain sufficient signal that using a non-reversible model is advisable  
608 in the first place. Our results show that the root placement was inferred correctly with high  
609 rootstrap support in 12 out of the 13 datasets in which the non-reversible model was  
610 preferable.

611           In the absence of a better fit for a non-reversible model, we do not think any inferred  
612 ML root position should be trusted. Second, root positions with higher rootstrap support

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613 should be trusted more, because a higher rootstrap support indicates less variance among sites  
614 in the signal for the placement of the root. Third, ML root positions should be trusted more  
615 when the number of root placements included in the confidence set of an AU test is small,  
616 because a smaller confidence set indicates that there is less uncertainty in the root placement  
617 when the analysis is conditioned on the full alignment and the unrooted ML tree topology. A  
618 conservative approach to inferring root placements with non-reversible models would be to  
619 consider any root placement that has a substantial fraction of the rootstrap support and/or is  
620 included in the set of possible root placements identified by the AU test as a possible root  
621 placement given the assumptions of the model.

622 We hope that the combination of non-reversible, rootstrap support, and AU tests will  
623 add another tool to the phylogeneticist's arsenal when it comes to inferring rooted  
624 phylogenies.

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