Molecular diagnosis of new isolate of tomato yellow leaf curl virus in Iraq

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Abstract

Tomato yield and quality in Iraq have been threatened by a variable range of infections caused by tomato yellow leaf curl virus. In previous studies, the TYLCV isolates were partially characterized using molecular tests for small fragments not the entire length of the virus. Sample of TYLCV-infected tomato has applied in this study to diagnose complete sequence of TYLCV isolate. Three sets of primers that belong to three well-identified strains in Iraq were used in a PCR technique and interestingly the results were negative. A new Iraqi isolate has been characterized as a first novel Iraqi isolate detected ever using next generation (NGS) and bioinformatics techniques. The NGS platform has produced about 78,232,062 paired reads of the TYLCV-infected tomato var. Oula F1. The complete raw reads of the infected variety have been analyzed using RepeatExplorer pipeline and Map to reference. The full sequence of TYLCV was reconstructed and extracted to consist of 2770 nt and then deposited in Genbank under accession

number MT583814. Copy numbers and genome proportion of this sequence have been calculated that were 3523 and 0,086% respectively. Phylogenetic analyses of full nucleotide sequences confirmed close relationship to Iranian isolate (TYLCV-Kahnootj) than other published viruses. Additionally, α and β DNA satellites have not discovered in the TYLCV-infected sample.

Introduction

The family Geminiviridae is recently divided into seven genera: *Becurtovirus*, *Begomovirus*, *Curtovirus*, *Eragrovirus*, *Mastrevirus*, *Topocuvirus* and *Turcurtovirus*. Host range, insect vector, genome organization and genome-wide pairwise sequence identities have been applied to classify this family (Varsani *et al.*, 2017). Tomato yellow leaf curl virus (TYLCV) belongs to genus *Begomovirus* and has a monopartite single-stranded DNA (ssDNA) encapsidated in geminate particles. TYLCV is transmitted by the whitefly, *Bemisia tabaci* (Genn.) that effectively spread virus particles through persistent manner (Panno *et al.*, 2016). The disease caused by this virus is the most important disease on tomato in Iraq and worldwide (Mansoor *et al.*, 2003; Salati *et al.*, 2002; Al-Kuwaiti *et al.*, 2013). Infected plants become stunted with chlorotic leaves, curled-up margins, and abscised flowers. Since the first infection was recorded in Israel 1939-1940, multiple regions have been identified to have isolates from this virus over the world and classified to different viral species (Antignus and Cohen 1994; Delatte *et al.*, 2004; Pico *et al.*, 1996; Polston and Anderson 1997). Tomato yellow leaf curl Sardinia virus TYLCSV infected tomatoes in Israel and Sardinia, and was the first sequenced isolate in 1991 (Kheyr-Pour *et al.*, 1991; Navot *et al.*, 1991), and since that date many isolates have been sequenced from various locations (Moriones and Navas-Castillo 2000). In Iraq, the disease was detected based on symptoms assay and serological tests. Recently, most popular molecular works on this virus have been achieved by PCR based on amplified short fragments of TYLCV genome such as coat protein domain (Al-Ani *et al.*, 2010; Reddy *et al.*, 2005; Maruthi *et al.*, 2007). Further, many tomato genotypes have been screened against TYLCV isolates to characterize similarities and differences through these genotypes (Al Tamimi, 2020). TYLCV has been isolated not only from tomato but also from various plants such as eggplant, melon, cucumber, sesame, cowpea in addition to weeds as a primary source of

infection like *Malva parviflora*, *Chenopodium murale* and *Chenopodium quinoa* (Al-Jubouri *et al.*, 2014). Al-Kuwaiti *et al.* (2013) have reported complete sequence of TYLCV that showed 99% identity to Spain strain (Acc. AJ489258) confirming its existence in Iraq. In this study, we utilized next generation sequencing and bioinformatics techniques for the first time in Iraq to find out new isolate with complete sequence of TYLCV. Furthermore, phylogeny analyses have been performed to understand how these isolates related to each other.

Materials and methods

Plant material

From open fields of Najaf governorate, the main region of tomato production in Iraq, fresh and young leaves of TYLCV-infected tomato variety Oula F1 (Seminis Seeds Company, USA) were harvested. This variety is considered the most popular and TYLCV-sensitive commercial variety of tomato in the middle of Iraq.

DNA isolation, PCR and NGS sequencing

To extract the DNA, the young leaves (about 2 gm) of the infected tomato were applied using cetyl-trimethylammonium bromide (CTAB) method with minor modifications (Doyle and Doyle 1990). Three sets of primers that belong to strains of TYLCV-IS, TYLCV-ES and TYLCV-Mld were applied in a PCR technique to check their infection within examined sample (Table 1) (Anfoka *et al.*, 2005).

Table 1. Three primers of main strains of TYLCV in Iraq that used to amplify and characterize virus sequence in the examined sample.

Primers	Sequence (5'- 3')	Size (bp)	Strains
TYv2337 TYc138	ACGTAGGTCTTGACATCTGTTGAGCTC AAGTGGGTCCCACATATTGCAAGAC	634	TYLCV-IS Israel
TYAlmv2516 TYAlmcl15	TTTTATTTGTTGGTGTGTTGTAGTTGAAG ATATTGATGGTTTTTCAAACCTTAGAAG	433	TYLCSV-ES Spain Almeria

TYv2664	ATTGACCAAGATTTTACACTTATCCC	316	TYLCV – mld
TYc138	AAGTGGGTCCCACATATTGCAAGAC		Normal mild

80
81 PCR conditions were an initial denaturation of 10 min at 95°C, 49 cycles of 30 s at 95°C, 20 s at
82 55-63°C and 30 s at 72°C followed by a final extension of 30 s at 72°C. To obtain the whole
83 genome sequence, the extracted genomic DNA sample was sent to sequencing at DNA-link
84 Company, Republic of Korea. The genome was sequenced by using the Novaseq6000 2x150bp
85 reads technique and application WGS (PCR Free550) based on the manufacturer's procedure.

86
87 RepeatExplorer program
88 To collect virus sequences from the whole data (NGS) of infected tomato, RepeatExplorer
89 pipeline was applied to recognize these kinds of sequences in forms of clusters within repetitive
90 DNA blocks (Novák *et al.* 2013). The targeted clusters of RepeatExplorer were extracted, and
91 then Repbase dataset (Jurka *et al.* 2005), and Basic Local Alignment Search Tool (Altschul *et al.*
92 1990) were used to identify contigs of each cluster. Furthermore, the extracted sequences were
93 mapped to well-identified plant virus sequences from DPVweb (Adams and Antoniw 2005).

94 Map to reference
95 Using Geneious program V. 11 (Kearse *et al.* 2012), the raw reads were assembled against
96 TYLCV Iraq (Acc. JQ354991.1) as a reference sequence that diagnosed by Al-Kuwaiti *et al.*
97 (2013) and then consensus sequence was collected. Mapping to reference was repeated against
98 consensus sequence (TYLCV) to confirm and obtain the actual number of assembled reads.
99 Also, the raw reads were applied to assemble against α and β DNA satellites. The outcomes
100 showed in a report that has a number of assembled reads and total used reads in addition to the
101 frequent overlapped reads. This data used to calculate copy numbers (Number of assembled
102 reads x read length/reference sequence length) and genome proportions (Number of assembled
103 reads / numbers of total NGS reads x 100) (Mustafa *et al.* 2018). Finally, the two consensus
104 sequences from RepeatExplorer and Map to reference were aligned using pairwise alignment in
105 Geneious program to collect the right length of TYLCV isolate.

106

Phylogenetic analysis

The maximum likelihood (ML) method was applied to find phylogeny model using MEGA7 (Tamura *et al.* 2013). The Geneious program V. 11 (Kearse *et al.* 2012) was used for alignment and optimized manually. The entire nucleotide sequences of 13 main TYLCV isolates including our isolate of interest, Kahnootj and Iraq isolates were applied to phylogeny study. ClustalW alignment was used for extracting sequences about 2770 nt for each. The phylogenetic tree reconstructed using General Time Reversible (GTR). Bayesian phylogeny inference was used for analysis with Bayesian inference of phylogeny (MrBayes 3.2.6) (Huelsenbeck and Ronquist 2001).

Results

The extracted DNA concentration and quality were 418 ng/ml and 1.92 which was in a high quality level for sequencing approach (Fig. 1. A). PCR has surprisingly failed to get amplified sequences for the three TYLCV isolates indicating that our sample has probably a new virus sequence. High quality raw reads have been obtained from NGS Illumina platform with about 78,232,062 paired reads; 150 bp read length and 36.9% GC content (Fig. 1. B).

RepeatExplorer has been clustered the whole uploaded raw reads that showed tandemly repeated sequences, TYLCV sequences involved in cluster number 76 as a putative satellite with a genome proportion 0,084% and the consensus sequence was 2763 nt (Fig. 1. C). On the other side, the report of Map to reference revealed that 65,303 reads have been assembled to count genome proportion and copy numbers of the isolate that were 0,086% and 3523 respectively whereas the consensus sequence was 2770 nt. In order to obtain complete sequence, the aligned consensus sequences from both RepeatExplorer and Map to reference showed 100% similarity and the longest sequence was extracted. Lastly, we determined 2770 nt of TYLCV and named as Najaf isolate and then deposited in Genbank under accession number MT583814.1.

Phylogenetic analyses showed what we have proven earlier by pairwise alignment between Najaf isolate and each of Kahnootj and Iraq isolates. Kahnootj was most likely similar to Najaf isolate than Iraq. TYLCV nucleotide and amino acid sequence alignments showed that Najaf isolate is

closely related to Kahnootj isolate the Iranian TYLCV in 93.8% pairwise identity while it was 90.3% with Iraq isolate (Fig. 2). However, polymorphic sequences have shown higher rate in intergenic region (IR) (75%), V2 (91.9%), C1 (Rep) (93.1%), and C4 (88.3%) than other coding regions V1 (97.3%), C3 (95.9%) and C2 (96.5%).

Our research has been extended trying to find DNA-a and β satellites using Map to reference tool but we did not get any positive hits.

Discussion

The deep sequencing of whole genome sequencing technique has revolutionized genomic research collecting a massive data of the host genome with complete sequences of cellular components. Additionally, this method is considered as unbiased approach to detect plant viruses with no previous knowledge about the suspected elements. The NGS technique does not use specific primers, antibodies, and virus-specific reagents to diagnose pathogenic viruses (Adams *et al.*, 2009). In this method, full image of plant viruses and complex infection could be investigated in the host genome unlikely to biased methods that focus on almost one expected virus (Jones *et al.*, 2017). In Iraq, most of molecular tests concentrated on partial sequences of TYLCV such as coat protein domain to detect the virus and apply these amplified parts in multiple alignments and phylogeny analyses. The only full-length sequenced of this virus from Iraq was Iraq isolate (JQ354991.1) that achieved by Al-Kuwaiti *et al* (2013) and named as TYLCV Iraq but it shared 99% similarity to Spain strain (Acc. AJ489258), Reunion Islands (Acc. AM409201) and Mauritius (Acc. HM448447). Additionally, Sadeq Al-Kuwaiti (2013) has applied rolling circle amplification (RCA) and non-degenerate primers to catch the full length of virus. Therefore, our new isolate is considered the first full sequenced Iraqi isolate of TYLCV with totally new data yet. We found Kahnootj isolate from Iran is closer than other isolates even from Iraq with 93.8 % similarity. It is highly expected that Najaf and Kahnootj isolates are sharing the same ancestor and most likely the virus was imported into Iraq from Iran by international trading (Mali *et al.*, 2003; Gibbs *et al.*, 2008; Al-Kuwaiti 2013).

Mapping reads to the reference has revealed the estimated copy numbers of this virus was 3523 within infected cell. Copy number value has been always considered as a good sign for variation

and variability of endogenous components within host genome especially transposable elements including viruses (Mustafa *et al.* 2018; Alisawi 2019). Interestingly, this is one of NGS and bioinformatics benefits to calculate copy numbers of the components of interest and this approach has been applied in many studies to predict viral copies (Yang *et al.*, 2017; Alisawi 2019). So far, the number of copies indicates virus particles indeed infected the host genome. In fact, this little amount of copies strongly explains the early stage of symptoms emerging in the infected sample. However, further works need to be achieved regarding isolate distribution and diversity over tomato varieties and different regions in Iraq.

We could not reveal the existence of DNA satellites in our infected sample probably due to moderate level of infection in the examined sample. A and β DNA satellites are often associated with begomoviruses in their replication, movement within infected plant and horizontal transmission between plants, as well as involved in induction of virus symptoms and suppression of transcriptional gene silencing. This result is consistent with most published outcomes that reported the importance of DNA satellites for disease severity and inducing symptoms. However, we do expect to find these satellites in severely TYLCV-infected plants as no more researches have been covered DNA satellites in Iraq (Laufs *et al.*, 1995; Saunders *et al.*, 2004; Cui *et al.*, 2005; Yang *et al.*, 2011; Amin *et al.*, 2011; Zhou, 2013).

Conclusion

This work concluded that whole genome sequencing technique alongside bioinformatics is highly recommended to detect complete sequence of TYLCV isolate within infected tomato genome raw reads. Consequently, a clear insight of virus pathway and identity has been revealed by phylogenetic analysis. More investigations need to be done for detecting full sequences of highly pathogenic plant viruses in Iraq.

Conflict of Interest Statement

The authors have no conflicts of interest to declare.

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193 Statement of Ethics

194 The authors have no ethical conflicts to disclose.

195 Author Contributions

196 A.A. carried out the experiment. O.A. analyzed the NGS data and wrote the manuscript with
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198 **References**

199 Adams, I. P., Glover, R. H., Monger, W. A., Mumford, R., Jackeviciene, E., Navalinskiene, M.,
200 ... & Boonham, N., 2009. Next-generation sequencing and metagenomic analysis: a universal
201 diagnostic tool in plant virology. *Molecular plant pathology*, 10(4), 537-545.

202 Adams M.J. & Antoniw J.F, 2005. DPVweb: An open access internet resource on plant viruses
203 and virus diseases. *Outlooks on Pest Management* 16, 268-270.

204 Al-Ani, R. A., Diwan, S. N. H., & Adhab, M. A, 2010. Efficiency of *Thuja orientalis* and
205 *Artimisia campestris* extracts to control of *Potato leaf roll virus* (PLRV) in potato
206 plants. *Agriculture and Biology Journal of North America*, 1(4), 579-583.

207 Alisawi, O. N, 2019. Virus integration and tandem repeats in the genomes of *Petunia* (Doctoral
208 dissertation, University of Leicester).

209 Al-Jubouri, A. A., Al-Fadhal, F. A., & Samaka, H. M, 2014. Detection of *Tomato yellow leaf*
210 *curl virus* (TYLCV) and its strains on tomato plants and host rang determine by Polymerase
211 Chain Reaction. *Kufa Journal for Agricultural Sciences*, 6(4), 99-117.

212 Al-Kuwaiti, N.; B. Otto; C. Collins; S. Seal and Maruthi, M, 2013. Molecular characterization
213 and first complete genome sequence of *Tomato yellow leaf curl virus* (TYLCV) infecting
214 tomato in Iraq. *New Disease Reports*, 27: 17.

215 Altschul S.F., Gish W., Miller W., Myers E.W. & Lipman D.J, 1990. Basic local alignment
216 search tool. *Journal of Molecular Biology* 215, 403-410.

217 Al Tamimi, S. Q, 2020. Molecular identification of some isolates of *Tomato yellow leaf curl*

218 virus and test response some tomato genotypes and their control(Doctoral dissertation,
219 University of Kufa)(In Arabic).

220 Amin, I., Hussain, K., Akbergenov, R., Yadav, J. S., Qazi, J., Mansoor, S., ... & Briddon, R. W,
221 2011. Suppressors of RNA silencing encoded by the components of the cotton leaf curl
222 begomovirus-betasatellite complex. *Molecular plant-microbe interactions*, 24(8), 973-983.

223 Anfoka, G.H., Abhary, M. and Nakhla, M.K., 2005. Molecular identification of species of the
224 Tomato yellow leaf curl virus complex in Jordan. *Journal of Plant Pathology*, pp.65-70.

225 Antignus, Y., & Cohen, S, 1994. Complete nucleotide sequence of an infectious clone of a mild
226 isolate of *Tomato yellow leaf curl virus* (TYLCV). *Phytopathology*, 84(7), 707-712.

227 Cui, X., Li, G., Wang, D., Hu, D., & Zhou, X., 2005. A begomovirus DNA β -encoded protein
228 binds DNA, functions as a suppressor of RNA silencing, and targets the cell nucleus. *Journal of*
229 *Virology*, 79(16), 10764-10775.

230 Delatte H, Naze F, Granier M, Reynaud B, Peterschmitt M, Lett M., 2004. Tomato-infecting
231 begomoviruses in the South Western islands of the Indian Ocean. *Presented at the 4th*
232 *International Geminivirus Symposium*, W3-4.

233 Doyle J. & Doyle J., 1990. Isolation of DNA from small amounts of plant tissues. *BRL Focus*
234 12, V15.

235 Gibbs, A. J., Ohshima, K., Phillips, M. J., & Gibbs, M. J., 2008. The prehistory of potyviruses:
236 their initial radiation was during the dawn of agriculture. *PloS one*, 3(6), e2523.

237 Huelsenbeck J.P. & Ronquist F., 2001. MRBAYES: Bayesian inference of phylogenetic trees.
238 *Bioinformatics* 17, 754-755.

239 Jones, S., Baizan-Edge, A., MacFarlane, S., & Torrance, L., 2017. Viral diagnostics in plants
240 using next generation sequencing: computational analysis in practice. *Frontiers in Plant*
241 *Science*, 8, 1770.

242 Jurka J., Kapitonov V.V., Pavlicek A., Klonowski P., Kohany O. & Walichiewicz J., 2005.
243 Repbase Update, a database of eukaryotic repetitive elements. *Cytogenetic and Genome*
244 *Research* 110, 462-467.

245 Kheyr-Pour A, Bendahmane M, Matzeit V, Accotto GP, Crespi S, Gronenborn B., 1991. *Tomato*
246 *yellow leaf curl virus* from Sardinia is a whitefly-transmitted monopartite geminivirus. *Nucleic*
247 *Acids Res* 19:6763–6769.

248 Kears M., Moir R., Wilson A., Stones-Havas S., Cheung M., Sturrock S., Buxton S., Cooper A.,
249 Markowitz S. & Duran C., 2012. Geneious Basic: an integrated and extendable desktop software
250 platform for the organization and analysis of sequence data. *Bioinformatics* 28, 1647-1649.

251 Laufs J., Traut W., Heyraud F., Matzeit V., Rogers S. G., Schell J., Gronenborn B., 1995. In
252 vitro cleavage and joining at the viral origin of replication by the replication initiator protein of
253 *Tomato yellow leaf curl virus*. *Proc Natl Acad Sci U S A* 92:3879–3883.

254 Mali, V. R., Subr, Z., & Kudela, O., 2003. Seed transmission of como and potyviruses in faba
255 bean and vetch cultivars introduced into Slovakia. *Acta phytopathologica et entomologica*
256 *hungarica*, 38(1-2), 87-97.

257 Mansoor, S., Briddon, R. W., Zafar, Y., & Stanley, J., 2003. Geminivirus disease complexes: an
258 emerging threat. *Trends in plant science*, 8(3), 128-134.

259 Maruthi M, Rekha A, Mirza S, Alam S, Colvin J., 2007. PCR-based detection and partial
260 genome sequencing indicate high genetic diversity in Bangladeshi begomoviruses and their
261 whitefly vector, *Bemisia tabaci*. *Virus Genes* 34, 373–385.

262 Moriones E, Navas-Castillo J., 2000. *Tomato yellow leaf curl virus*, an emerging virus complex
263 causing epidemics worldwide. *Virus Res* 71:123–134.

264 Mustafa S.I., Schwarzacher T. & Heslop-Harrison J., 2018. Complete mitogenomes from
265 Kurdistan sheep: abundant centromeric nuclear copies representing diverse ancestors.
266 *Mitochondrial DNA Part A*, 1-14.

267 Navot N, Pichersky E, Zeidan M, Zamir D, Czosnek H., 1991. *Tomato yellow leaf curl virus*: a
268 whitefly-transmitted geminivirus with a single genomic component. *Virology* 185:151–161.

269 Novák P., Neumann P., Pech J., Steinhaisl J. & Macas J., 2013. RepeatExplorer: a Galaxy based
270 web server for genome-wide characterization of eukaryotic repetitive elements from next-
271 generation sequence reads. *Bioinformatics* 29, 792-793.

272 Panno, S., Iacono, G., Davino, M., Marchione, S., Zappardo, V., Bella, P., ... & Davino, S., 2016.
273 First report of *Tomato leaf curl New Delhi virus* affecting zucchini squash in an important
274 horticultural area of southern Italy. *New Dis. Rep*, 33(6), 2044-0588.

275 Pico B, Diez MJ, Nuez F., 1996. Viral diseases causing the greatest economic losses to the
276 tomato crop. II. The *Tomato yellow leaf curl virus*. *Sci Hort* 67:151–196.

277 Polston JE, Anderson PK., 1997. The emergence of white fly transmitted geminiviruses in
278 tomato in the Western Hemisphere. *Plant Dis* 81:1358–1369.

- 279 Reddy, R. C., Colvin, J., Muniyappa, V., & Seal, S., 2005. Diversity and distribution of
280 begomoviruses infecting tomato in India. *Archives of virology*, 150(5), 845-867.
- 281 Sadeq Al-Kuwaiti, N. A., 2013. Molecular characterization of viruses infecting potato and
282 vegetables in Iraq (Doctoral dissertation, University of Greenwich).
- 283 Salati, R., Nahkla, M. K., Rojas, M. R., Guzman, P., Jaquez, J., Maxwell, D. P., & Gilbertson, R.
284 L., 2002. *Tomato yellow leaf curl virus* in the Dominican Republic: characterization of an
285 infectious clone, virus monitoring in whiteflies, and identification of reservoir
286 hosts. *Phytopathology*, 92(5), 487-496.
- 287 Saunders, K., Norman, A., Gucciardo, S., & Stanley, J., 2004. The DNA β satellite component
288 associated with ageratum yellow vein disease encodes an essential pathogenicity protein
289 (β C1). *Virology*, 324(1), 37-47.
- 290 Tamura K., Stecher G., Peterson D., Filipski A. & Kumar S., 2013. MEGA6: molecular
291 evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution* 30, 2725- 2729.
- 292 Varsani, A., Roumagnac, P., Fuchs, M., Navas-Castillo, J., Moriones, E., Idris, A., ... & Martin,
293 D. P., 2017. *Capulavirus* and *Grablovirus*: two new genera in the family
294 Geminiviridae. *Archives of Virology*, 162(6), 1819-1831.
- 295 Yang, Z., Mammel, M., Papafragkou, E., Hida, K., Elkins, C. A., & Kulka, M., 2017.
296 Application of next generation sequencing toward sensitive detection of enteric viruses isolated
297 from celery samples as an example of produce. *International journal of food microbiology*, 261,
298 73-81.
- 299 Yang, X., Xie, Y., Raja, P., Li, S., Wolf, J. N., Shen, Q., *et al.*, 2011. Suppression of
300 methylation-mediated transcriptional gene silencing by β C1-SAHH protein interaction during
301 geminivirus-betasatellite infection. *PLoS Pathog.* 7:e1002329. doi:
302 10.1371/journal.ppat.1002329.
- 303 Zhou, X., 2013. Advances in understanding begomovirus satellites. *Annual review of*
304 *phytopathology*, 51.

305 **Figures**

Fig. 2. Phylogenetic tree of 13 full lengths TYLCV isolates showing close relationship between Najaf and Kahnooj isolates. The tree was generated using Geneious tree builder with bootstrape analyses and 50% support, the out group member was *Tomato leaf curl Sudan virus*.

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