1 Short Communication

2 **The MAGOH paralogs - MAGOH, MAGOHB and their multiple isoforms**

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24 **ABSTRACT**

25 A central processing event in eukaryotic gene expression is splicing. Concurrent with splicing, the
 26 core-EJC proteins, eIF4A3 and RBM8A-MAGOH heterodimer are deposited 24 bases upstream of
 27 newly formed exon-exon junctions. One of the core-EJC proteins, MAGOH contains a paralog
 28 MAGOHB, and this paralog pair is conserved across vertebrates. Upon analysis of the splice variants
 29 of MAGOH-paralogs, we have found the presence of alternate protein isoforms which are also
 30 evolutionarily conserved. Further, comparison of the amino acid sequence of the principal and
 31 alternate protein isoforms has revealed absence of key amino acid residues in the alternate isoforms.
 32 The conservation of principal and alternate isoforms correlates to the importance of MAGOH and
 33 MAGOHB across vertebrates.

34 **Keywords:** MAGOH, MAGOHB, EJC, isoforms

35 **Abbreviations:** EJC – Exon Junction Complex, MAGOH-mago nashi homolog, eIF4A3 – eukaryotic
 36 initiation factor 4 A3, RBM8A- RNA binding motif 8A (also known as Y-14)

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53 1. INTRODUCTION

54 The primary step in eukaryotic gene expression involves transcription of DNA into precursor-mRNAs
 55 (pre-mRNAs). The pre-mRNAs undergo various post-transcriptional processing events before
 56 generating functional protein products. A notable processing event is splicing, where the spliceosome
 57 removes introns and ligates exons together. Concurrent with splicing, mature mRNAs are marked
 58 with the exon junction complex (EJC) at 24 nucleotides upstream of an exon-exon junction¹. The EJC
 59 forms a gene regulatory nexus composed of many proteins that are grouped under stable core- and
 60 transient peripheral-protein complexes. The core-EJC consists of three proteins, eIF4A3 and RBM8A-
 61 MAGOH heterodimer; whereas the peripheral-EJC includes ALYREF, ASAP complex or PSAP
 62 complex, and Upf proteins²⁻⁶. Additional to mRNA processing, the core-EJC proteins have essential
 63 functions in brain development and embryonic neurogenesis⁷. In this communication, we will mainly
 64 focus on one of the core-EJC proteins, MAGOH.

65 MAGOH (*mago nashi* homolog) was originally identified in *Drosophila melanogaster*, where
 66 embryos lacking functional mago protein produced sterile progeny. Thus, this protein was named
 67 “mago nashi” meaning “without grandchildren” in the Japanese language⁸. MAGOH is well
 68 conserved among all vertebrates. It functions to maintain integrity of mRNAs as a part of the EJC as
 69 well as the nonsense-mediated mRNA decay (NMD) pathway⁹. The importance of MAGOH in NMD
 70 has recently also been characterised in zebrafish¹⁰. Interestingly, vertebrates contain a paralog of
 71 MAGOH, MAGOHB which is also associated with mRNAs via EJC and NMD¹¹. In addition to their
 72 roles in mRNA processing, the MAGOH paralogs (MAGOH and MAGOHB) have been found to be
 73 associated with cancer progression. In cancers with a hemizygous deletion of MAGOH, MAGOHB
 74 was suggested as the highest dependent gene. The dependency of MAGOHB in such MAGOH
 75 deficient cancers have the potential for cancer treatment¹². MAGOH paralogs have also been found to
 76 be involved in gastric cancers. Importantly, the simultaneous knockdown of MAGOH-MAGOHB in
 77 gastric cancer cells exhibited anti-tumor effects via the BRAF/MEK/ERK signalling pathway¹³. Thus,
 78 the MAGOH paralogs are essential for proper functioning of cells, yet sufficient information on the
 79 architecture and evolutionary conservation of these genes is lacking. Concordant with most

80 eukaryotic genes, both the MAGOH-paralogs undergo alternative splicing and produce multiple splice
81 variants, which have not yet been characterised. In this communication, we will discuss the features of
82 the alternate protein isoforms of MAGOH-paralogs in humans and mice. We will also look at
83 structural features of the alternate isoforms compared to the principal MAGOH protein isoform.

84 **2. RESULTS and DISCUSSION**

85 **2.1 MAGOH paralogs have multiple protein isoforms in humans**

86 MAGOH-paralogs belong to the MAGO NASHI protein family, Ensembl ID-PTHR12638. In
87 humans, *MAGOH* also known as *MAGOH1* or *MAGOHA* is located on the reverse strand of
88 chromosome 1 at 1p32.3 (*chr1:53,226,900-53,238,518, GRCh38/hg38*). Alternative splicing of
89 *MAGOH* pre-mRNA generates four transcripts, *MAGOH-201*, *MAGOH-202*, *MAGOH-203*, and
90 *MAGOH-204* (Table-1). *MAGOH-202* is the primary transcript made of five exons and encodes the
91 principal protein isoform of 146 amino acids (aa). *MAGOH-201* constitutes four exons and codes for
92 an alternate protein isoform of 109 aa. Comparison of *MAGOH-201* with *MAGOH-202* reveals
93 skipping of exon 3 (111 nucleotides) in *MAGOH-201*, generating the alternate isoform (Figure-1a).
94 The other transcripts, *MAGOH-203* and *MAGOH-204* comprise three exons and do not translate into
95 proteins due to the absence of open reading frames (ORF). Hence, *MAGOH* is transcribed to generate
96 four splice variants, of which two are protein-coding and two are non-coding. The finding of an
97 alternate isoform (109 aa) is interesting as MAGOH is primarily mentioned as a 146 aa protein, with a
98 molecular weight of 17 kDa^{14,15}. Subsequent to the finding of an alternate transcript in *MAGOH*, we
99 were interested to find out if its paralog, *MAGOHB* also produces an alternate transcript.

100 The *MAGOHB* gene in humans is located on the reverse strand of chromosome 12 at 12p13.2
101 (*chr12:10,604,193-10,613,609, GRCh38/hg38*). As per the Ensembl genome browser, *MAGOHB* pre-
102 mRNA generates eleven alternately spliced transcripts (Table-1). Of these eleven transcripts, three
103 have coding potential. Thus, similar to *MAGOH*, *MAGOHB* also produces multiple protein coding
104 transcripts. Two of the protein-coding transcripts, *MAGOHB-201* and *MAGOHB-204*, contain five
105 exons. *MAGOHB-201* is the primary transcript and codes for the 148 aa primary protein isoform,
106 while the alternate transcript, *MAGOHB-204* codes for the alternate 102 aa protein isoform. The two

transcripts differ due to the presence of an alternative 5' splice site in the first exon of *MAGOHB-204* (Figure-1a). The third protein-coding transcript, *MAGOHB-211* consists of 2 exons which code for an 81 aa protein. In the transcript, *MAGOHB-211*, its first exon is similar to the principal isoform, *MAGOHB-201*, whereas its second exon differs completely due to the presence of an alternate 3' splice site. *MAGOHB* thus produces three different protein isoforms. It will be fascinating to analyse whether the alternate protein isoforms are also generated in other species. Apart from these three protein coding transcripts, five other transcripts (*MAGOHB-205*, *-206*, *-208*, *-209*, *-210*) are also generated. Out of these five, three transcripts *MAGOHB-208*, *MAGOHB-209* and *MAGOHB-205* are made of six exons whereas *MAGOHB-206* and *MAGOHB-210* are composed of five exons. All the five transcripts are degraded via the NMD pathway due to the presence of premature termination codons (PTCs), which is one of the prerequisites of NMD. Lastly, the remaining three transcripts of *MAGOHB*, *MAGOHB-202*, *-203*, *-207* have intron retention (IR), and do not code for any protein. Here, *MAGOHB-203* and *MAGOHB-207* comprise of three exons whereas *MAGOHB-202* is made of two exons. In this article, we will focus only on the protein coding transcripts of both the *MAGOHB*-paralogs.

In line with the Ensembl annotation, we will be referring to the protein isoforms according to their transcript IDs. We observed that the two *MAGOHB* protein isoforms, principal isoform-146 aa and alternate isoform-109 aa differ in 37 residues, i.e., amino acids 50-86 of the principal isoform. In case of *MAGOHB*, three protein isoforms are generated. Coding region of the transcript *MAGOHB-204* starts from the second exon, and translates into an alternate protein isoform that varies from the principal protein isoform, *MAGOHB-201* in the N-terminus (devoid of residues 1-46). The third protein isoform, *MAGOHB-211* differs from the principal isoform in amino acids 38-81 since its second exon is formed by an alternate 3' splice site. The presence of two functionally redundant *MAGOHB* paralogs is in itself fascinating. From this characterisation we also know that both the *MAGOHB* paralogs generate multiple protein isoforms in humans. In the following sections we have analysed the presence of multiple protein isoforms of *MAGOHB* paralogs across different species,

which might hint on an evolutionarily conserved function of the alternate protein isoforms generated by the MAGOH paralogs.

2.2 MAGOHB codes for multiple protein isoforms in mice

The MAGOH paralogs are conserved across vertebrates and MAGOH has been widely studied in mice. The presence of multiple protein isoforms of the MAGOH paralogs in humans raised the next question – are such alternate protein isoforms also present in mice? Hence, we analysed the mouse genome on Ensembl for multiple isoforms of MAGOH paralogs (Table-2). Here, we have referred to the mouse orthologs of human *MAGOH* paralogs as *Magoh* and *Magohb*. In mice, *Magoh* is located on chromosome 4 (*chr 4:107,879,755-107,887,424, GRCm38*), and is transcribed into two transcripts, *Magoh-201* and *Magoh-202*. *Magoh-201* is translated into a 146 aa protein, whereas *Magoh-202* does not code for any protein because of the absence of an ORF. Therefore, unlike humans, mice do not produce any alternate protein isoform of *Magoh*. The paralog of *Magoh*, *Magohb* is located on the reverse strand of chromosome 6 (*chr 6:131,284,388-131,293,244, GRCm38*). A total of seven *Magohb* transcripts are generated in mice, where two transcripts (*Magohb-201* and *Magohb-205*) are protein-coding. *Magohb-201* translates into the 148 aa principal protein isoform, whereas *Magohb-205* codes for the alternate protein isoform of 102 aa. Thus, in humans and mice two protein isoforms of MAGOHB are generated, which are of the same length. Similar to the alternate human MAGOHB transcript, *MAGOHB-204*, the first exon of *Magohb-205* constitutes most of the 5'UTR region and ligates exon 2 via an alternate 5' splice site (Figure-1b). The start codon for *Magohb-205* is present in the second exon, thus the protein it encodes is devoid of residues 1 – 46 compared to the principal isoform. Curiously, splicing patterns of the two *Magohb* protein isoforms in mice are very similar to human MAGOHB protein isoforms. Besides the protein-coding transcripts, three transcripts, *Magohb-202*, *-203*, *-204*, are subjected to degradation via NMD. The remaining two transcripts (*Magohb-206* and *Magohb-207*) retain intronic regions and do not code for any protein. Thus, similar to its human ortholog, *Magohb* in mice also generates multiple protein-coding and non-coding splice variants.

Altogether, we find *MAGOH* transcript variants in humans and mice belong to two biotypes- protein coding transcripts and transcripts without an ORF; while *MAGOHB* transcript variants belong to three

biotypes – protein-coding, NMD-sensitive, and IR. In both the species, *MAGOH* and *MAGOHB* generate multiple splice variants. The pattern of splicing for *MAGOH*-paralogs is similar, as both the species produce protein-coding as well as non-coding transcripts. In general, multiple splice variants tend to increase the proteome diversity of the genome. Among various alternative splicing (AS) events, IR, and AS-NMD are important for post-transcriptional gene regulation^{16,17}. It is also likely that transcripts with similar exon-intron structure, referred to as iso-orthologs, have similar biological function¹⁸. The similarity in splicing pattern as well as length of the alternate protein-coding transcripts prompted us to shift our focus to the sequence conservation of the alternate protein isoforms, discussed in the next section. The reason behind generation of multiple protein-coding and non-coding splice variants of *MAGOH*-paralogs is not yet known. They may contribute either towards regulation of *MAGOH*-paralogs or may perform some unexplored function.

2.3 Alternate isoforms of *MAGOH* paralogs are conserved across different species

We have described that *MAGOH*-paralogs produce more than one protein isoform (Figure-1). In this section, we depict the conserved nature of the alternate protein isoforms. To gain further insight into the conservation of the alternate proteins of *MAGOH* paralogs, we performed pairwise alignments of the protein sequences in humans and mice. We first compared the principal isoforms of *MAGOH*-paralogs, *MAGOH*-202 with *Magoh*-201 and *MAGOHB*-201 with *Magohb*-201, followed by comparison of the alternate isoforms of *MAGOHB*, *MAGOHB*-204 with *Magohb*-205.

The principal isoforms of *MAGOH* in humans and mice are 100 % identical (Figure-2a), whereas in the case of *MAGOHB* the identity is 98%, owing to three different residues, i.e., amino acids 2-4 (Figure-2b). Conservation of the primary isoforms is evident due to their suggested roles in EJC and NMD^{11,19}. Proceeding to the alternate protein isoforms, as discussed above, the alternate protein of *MAGOH* (*MAGOH*-201, 109 aa) does not have any ortholog in mice, meaning this isoform is either not important or might be specifically generated in higher vertebrates including humans. Interestingly, the 102 aa *MAGOHB* proteins in humans and mice are 100% identical to each other (Figure-2c). The striking identity prompted us to analyze other species for the presence of alternate isoforms of *MAGOH*-paralogs. Thus, we analyzed the alternate isoforms in different species and performed a

multiple sequence alignment of the protein sequence. Quite interestingly, the alternate isoforms of the MAGOH-paralogs are conserved across vertebrates (Table 3). Though the alternate isoform of MAGOH, 109 aa is not present in mice, it is present in 52 other species, where the protein sequence differs only at position 2 (Supplementary Figure-1 and 2). The alternate isoform of MAGOHB, 102 aa, is present in 11 different species and is 100% identical in 10 species, but differs in the alternate isoform of kangaroo rat- *Dipodomys ordii* at position 25 and 35. We have also validated the expression of the conserved alternate protein-coding transcripts, *MAGOH-201* and *MAGOHB-204* in HEK-293 cells via RT-PCR (Figure-3a). We further performed a phylogenetic analysis of cDNA sequence from four species having alternate isoforms for both MAGOH and MAGOHB (highlighted in Table 3). The tree estimated via maximum likelihood shows the isoforms clustered into two groups, specific for MAGOH and MAGOHB. This indicates that similar to the principal isoforms, the alternate isoforms are also evolutionarily conserved orthologs (Figure-3b). The conservation of the alternate protein isoforms of MAGOH paralogs across species is quite riveting. Consequently, an evolutionary pressure must have resulted in conservation of the alternate protein isoforms of both the MAGOH-paralogs. It has been previously pointed that an identity in the range of 50%-70% is the defining boundary of protein conservation, and function of conserved proteins is generally expected to be preserved above 70% identity²⁰. Thus, the presence of a conserved alternate isoform of MAGOH and MAGOHB does not seem to be a mere coincidence and might involve a functional role, yet unexplored. Due to the difference in the residues of the principal and alternate isoforms of MAGOH paralogs, there might exist structural and functional differences between them which we have elaborated below.

The structure of MAGOH is composed of three α helices and six β strands that form an extended sheet¹⁵. The helices form a hydrophobic core to interact with the RNA-binding domain of Y-14, which is primarily involved in protein-protein interactions. The extended β sheet contains conserved residues, forming the binding platform for EJC factors or other associated proteins^{15,19}. On aligning the principal and alternate protein isoforms in humans, we found the alternate isoform of MAGOH to have 74.7% sequence identity to the principal MAGOH isoform, whereas the alternate MAGOHB

isoform has 68.9% sequence identity to the principal MAGOHB isoform (Figure-2d, 2e). Comparison of structural features is shown in Figure-4. As shown, the alternate isoform of MAGOH lacks residues 50-86 corresponding to the principal MAGOH isoform, MAGOH-202. These residues constitute a portion of the α_1 helix of MAGOH. Helix α_1 with helix α_3 form a part of the hydrophobic core which binds the RNA-binding domain (RBD) of Y-14. Consequently, the alternate isoform, MAGOH-201, might lack the ability to bind Y-14 or may bind to it without masking Y-14's RBD. If the alternate isoform of MAGOH binds Y-14 it will be interesting to find out the stability and possible function of this alternate heterodimer complex.

The N-terminus of the principal MAGOH isoform contains conserved residues, Tyr6, Tyr10, and Phe21 of the β -sheet¹⁵. Since the alternate isoform of MAGOHB, MAGOHB-204, lacks residues 1-46, it also lacks these conserved residues present in the β sheet. Consequently, mutations in residues 16/17, 20, 39/40, 41/42 of the principal MAGOH protein result in loss of interaction with pre-mRNA, spliced mRNA as well as EJC components, BTZ, UPF3b and eIF4A3, whereas interaction with Y-14 and PYM remains intact²¹. Thus, MAGOHB-204 might have functional Y-14 binding but may lack binding with other EJC factors.

To summarize, we find that the generation of alternate protein isoforms by MAGOH-paralogs is not only limited to humans, but also extends to other species. The high degree of conservation of the alternate protein isoforms, MAGOH-201 (109 aa) and MAGOHB-204 (102 aa), may point out to the presence of an evolutionary pressure maintaining multiple isoforms of MAGOH and MAGOHB. These alternate protein isoforms might be involved in regulating the levels of principal MAGOH proteins or may have functions entirely different from the principal isoforms.

Delineating the function of these isoforms is challenging because of the sequence similarity between the primary and alternate protein isoforms. Hence, the function of the alternate isoforms might need to be analysed in conditions where the principal isoform is absent, and is the subject of further experimentations.

3. MATERIALS AND METHODS

3.1 Sequence information for isoforms

All the sequence information for *Homo sapiens* and *Mus musculus* were taken from Ensembl (Release 101) (<https://asia.ensembl.org/index.html>)²². The Ensembl IDs are - ENSG00000162385(MAGOH), ENSG00000111196(MAGOHb) for human MAGOH paralogs, and ENSMUSG00000028609(Magoh), ENSMUSG00000030188(Magohb) for Magoh paralogs in mice. Genome assembly versions referred were Human-GRCh38.p13 and Mouse-GRCm38.p6.

3.2 Sequence alignment

Alignments were generated using the CLC Main Workbench's alignment tool and the identity among aligned proteins was analysed using the EMBOSS Needle pairwise alignment tool²³. The proteins were aligned with the BLOSUM62 matrix. Gap opening penalty was kept at 10 and gap extension penalty at 0.5. Multiple sequence alignments of the alternate isoforms (109 aa and 102 aa) were performed using ClustalW in the MEGA-X²⁴ tool with the following parameters- gap opening penalty 10.00, gap extension penalty 0.20. Jalview was used for visualisation of the multiple sequence alignments²⁵. Phylogenetic trees were generated using MEGA-X as previously described²⁶, briefly the cDNA sequence was downloaded from Ensembl, codons aligned using MUSCLE followed by model generation and using the highest model to estimate the phylogenetic tree via maximum likelihood.

3.3 Isoform expression analysis

RNA for isoform analysis was extracted from HEK-293 cells, cultured in DMEM at 37°C in a humidified CO₂ incubator. The RNA was converted into cDNA using the Super Reverse Transcriptase MuLV cDNA kit (Biobharati). The primers sequence for the principal isoform were used as described previously¹¹. Primer sequences for the alternate isoforms are mentioned in the supplementary file.

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CONFLICTS OF INTEREST

The authors declare no competing conflicts.

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370 **Figure 1- Isoforms of MAGOH-paralogs** in (a) humans and (b) mice. The pre-mRNA generated
371 from each gene is shown on top, where boxes represent exons, and introns are represented by
372 horizontal lines. Constitutive splicing is denoted with solid lines, while alternative splicing is denoted
373 with dashed lines. MAGOH is represented in blue and MAGOHB is represented in pink. The pre-
374 mRNA shown here includes exons of the protein-coding transcripts. Principal and alternate protein
375 isoforms are shown below each pre-mRNA, where each box represents coding exons from the pre-
376 mRNA.

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Figure 2- Pairwise alignment of MAGOH protein isoforms. The protein isoform alignments were generated using the CLC Main Workbench. **a)** MAGOH principal isoform alignment in humans (MAGOH-202) and mice (Magoh-201) **b)** MAGOHB principal isoform alignment in humans (MAGOHB-201) and mice (Magohb-201) **c)** MAGOHB alternate isoform alignment in humans (MAGOHB-204) and mice (Magohb-205). The principal and alternate isoforms of MAGOH/MAGOHB in humans were also aligned to each other to analyse similarity - **d)** MAGOH principal isoform (MAGOH-202) aligned with alternate isoform (MAGOH-201) **e)** MAGOHB principal isoform (MAGOHB-201) aligned with its alternate isoform (MAGOHB-204). The different residues are coloured in red, gaps are coloured in pink boxes and represented as dashes.

407 **Table 1- MAGOH paralog isoforms in *Homo sapiens***

Transcript ID	Ensembl ID	Length (bases)	Protein (amino acids)	Biotype	5'UTR (bases)	Exon 1 (bases)	Exon 2 (bases)	Exon3 (bases)	Exon4 (bases)	Exon5 (bases)	Exon6 (bases)	3' UTR (bases)
MAGOH-201	ENST00000371466.4	500	109	Protein-coding	51	51* + 88	59	83	100 + 119	-	-	119
MAGOH-202	ENST00000371470.8	656	146	Protein-coding	70	70 + 88	59	111	83	100 + 145	-	145
MAGOH-203	ENST00000462941.1	738	-	Processed transcript (no ORF)	-	148	59	531	-	-	-	-
MAGOH-204	ENST00000495868.1	695	-	Processed transcript	-	398	83	214	-	-	-	-
MAGOHB-201	ENST00000320756.7	2606	148	Protein-coding	77	77 + 94	59	111	83	100 + 2082	-	2082
MAGOHB-202	ENST00000398874.8	549	-	IR*	-	187	362	-	-	-	-	-
MAGOHB-203	ENST00000537852.5	2671	-	IR	-	162	59	2088	-	-	-	-
MAGOHB-204	ENST00000539554.5	666	102	Protein-coding	95	51	44 + 15	111	83	100 + 262		262
MAGOHB-205	ENST00000540074.5	705	44	NMD*	95	51	45 + 15	111	9 + 277	83	115	475
MAGOHB-206	ENST00000543929.5	841	81	NMD	68	68 + 94	152 + 187	59	111	170	-	527
MAGOHB-207	ENST00000544176.1	559	-	IR	-	162	59	338	-	-	-	-
MAGOHB-208	ENST00000544850.5	1057	90	NMD	62	62 + 94	59	111	9 + 277	83	362	722
MAGOHB-209	ENST00000545236.5	868	81	NMD	77	77 + 94	152 + 187	59	111	83	105	545
MAGOHB-210	ENST00000546173.5	576	81	NMD	41	41 + 94	152 + 12	59	111	107	-	289
MAGOHB-211	ENST00000625272.1	319	81	Protein-coding	73	73 + 94	152	-	-	-	-	-

408 *NMD = Nonsense-mediated mRNA decay, IR= Intron retention, * Number of bases written in red correspond to untranslated regions (UTR)

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Transcript ID	Ensembl ID	Length (bases)	Protein (amino acids)	Biotype	5' UTR (bases)	Exon 1 (bases)	Exon 2 (bases)	Exon3 (bases)	Exon4 (bases)	Exon5 (bases)	Exon6 (bases)	3' UTR (bases)
Magoh-201	ENSMUST00000030348.5	692	146	Protein coding	103	103* + 88	59	111	83	100 + 148	-	148
Magoh-202	ENSMUST000000141376.1	483	-	Processed transcript	-	189	83	211	-	-	-	-
Magohb-201	ENSMUST00000032307.11	767	148	Protein coding	79	79 + 94	59	111	83	100 + 241	-	241
Magohb-202	ENSMUST000000172883.7	460	35	NMD*	-	60	47 + 64	83	206	-	-	353
Magohb-203	ENSMUST000000173198.7	561	44	NMD	-	80	53 + 147	59	111	83	28	428
Magohb-204	ENSMUST000000173332.1	395	44	NMD	-	80	54 + 147	59	56	-	-	262
Magohb-205	ENSMUST000000173837.5	897	102	Protein coding	564	520	44 + 15	111	83	100 + 24	-	24
Magohb-206	ENSMUST000000174488.1	447	-	IR*	-	231	216	-	-	-	-	-
Magohb-207	ENSMUST000000174781.3	546	-	IR	-	288	111	83	64	-	-	-

410 **Table 2- Magoh paralogs in *Mus musculus***

411 *NMD = Nonsense-mediated mRNA decay, IR= Intron retention, * Number of bases written in red correspond to untranslated regions (UTR)

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413 **Table 3- Species with conserved alternate isoforms of MAGOH paralogs**

Gene	Size of protein (amino acids)	Species with conserved isoform
MAGOH	109	<i>Anser brachyrhynchus</i> , <i>Anser cygnoides</i> , <i>Aotus nancymae</i> , <i>Bison bison bison</i> , <i>Bos grunniens</i> , Hybrid - <i>Bos Indicus</i> (<i>Bos indicus</i> x <i>Bos taurus</i>), <i>Bos mutus</i> , Hybrid - <i>Bos Taurus</i> (<i>Bos indicus</i> x <i>Bos taurus</i>), <i>Callithrix jacchus</i> , <i>Camelus dromedarius</i> , <i>Canis lupus familiaris</i> , <i>Canis lupus familiaris</i> , <i>Carlito syrichta</i> , <i>Catagonus wagneri</i> , <i>Cebus capucinus imitator</i> , <i>Cercocebus atys</i> , <i>Colobus angolensis palliatus</i> , <i>Equus caballus</i> , <i>Felis catus</i> , <i>Gorilla gorilla gorilla</i> , <i>Homo sapiens</i> , <i>Lynx canadensis</i> , <i>Macaca fascicularis</i> , <i>Macaca nemestrina</i> , <i>Mandrillus leucophaeus</i> , <i>Microcebus murinus</i> , <i>Monodelphis domestica</i> , <i>Moschus moschiferus</i> , <i>Neovison vison</i> , <i>Nomascus leucogenys</i> , <i>Ornithorhynchus anatinus</i> , <i>Oryctolagus cuniculus</i> , <i>Bonobo</i> , <i>Pan paniscus</i> , <i>Pan troglodytes</i> , <i>Panthera leo</i> , <i>Panthera pardus</i> , <i>Panthera tigris altaica</i> , <i>Papio anubis</i> , <i>Physeter catodon</i> , <i>Ptilocolobus tephrosceles</i> , <i>Podarcis muralis</i> , <i>Prolemur simus</i> , <i>Propithecus coquereli</i> , <i>Rhinolophus ferrumequinum</i> , <i>Rhinopithecus bieti</i> , <i>Rhinopithecus roxellana</i> , <i>Saimiri boliviensis boliviensis</i> , <i>Suricata suricatta</i> , <i>Sus scrofa</i> , <i>Theropithecus gelada</i> , <i>Ursus thibetanus thibetanus</i> , <i>Vombatus ursinus</i>

MAGOHB	102	<i>Cavia porcellus</i> , <i>Cricetulus griseus</i> , <i>Dipodomys ordii</i> , <i>Gorilla gorilla gorilla</i> , <i>Homo sapiens</i> , <i>Mus musculus</i> , <i>Mus pahari</i> , <i>Mus spretus</i> , <i>Nannospalax galili</i> , <i>Pan troglodytes</i> , <i>Rhinopithecus bieti</i>
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