1	Reduced representation sequencing to understand the evolutionary history of Torrey pine
2	(Pinus torreyana Parry) with implications for rare species conservation
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### Abstract

13 Understanding the contribution of neutral and adaptive evolutionary processes to population differences is often necessary for better informed management and conservation of 14 15 rare species. In this study, we focused on *Pinus torreyana* Parry (Torrey pine), one of the world's rarest pines, endemic to one island and one mainland population in California. Small population 16 17 size, low genetic diversity, and susceptibility to abiotic and biotic stresses suggest Torrey pine may benefit from inter-population genetic rescue to preserve the species' evolutionary potential. 18 We leveraged reduced representation sequencing to tease apart the respective contributions of 19 20 stochastic and deterministic evolutionary processes to population differentiation. We applied these data to model spatial and temporal demographic changes in effective population sizes and 21 genetic connectivity, to assess loci possibly under selection, and evaluate genetic rescue as a 22 potential conservation strategy. Overall, we observed exceedingly low standing variation 23 reflecting consistently low effective population sizes across time and limited genetic 24 25 differentiation suggesting maintenance of gene flow following divergence. However, genome scans identified more than 2000 SNPs candidates for divergent selection. Combined with 26 previous observations indicating population phenotypic differentiation, this indicates that natural 27 28 selection has likely contributed to population genetic differences. Thus, while reduced genetic diversity, small effective population size, and genetic connectivity between populations suggest 29 genetic rescue could mitigate the adverse effect of rarity, divergent selection between 30 31 populations indicates that genetic mixing could disrupt adaptation. Further work evaluating the 32 fitness consequences of inter-population admixture is necessary to empirically evaluate the tradeoffs associated with genetic rescue in Torrey pine. 33

34 **Keywords:** Rare species conservation, demographics, adaptation, genome scans, genetic rescue.

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## Introduction

36 Conservation biology aims to preserve rare species and determine the appropriate management strategies necessary for long-term persistence and maintenance of evolutionary 37 potential (Di Santo & Hamilton, 2020; Ralls et al., 2018; Swarts, Sinclair, Krauss, & Dixon, 38 2009; Young, Brown, & Zich, 1999). Rare species may have reduced effective population sizes 39 40 (Ne), impeding populations' ability to adapt to change (e.g., Ne < 1000) or increasing probability of inbreeding (e.g., Ne < 100) (Frankham, Bradshaw, & Brook, 2014), ultimately increasing the 41 risk of local extirpation. Combined, rarity and isolation are often associated with stochastic loss 42 43 of genetic variation (Aguilar, Quesada, Ashworth, Herrerias Diego, & Lobo, 2008; Hague & Routman, 2016; Young, Boyle, & Brown, 1996). Genetic rescue is one conservation strategy that 44 has been successfully used in both animals and plants to mitigate consequences of severely 45 reduced genetic diversity (Bossuyt, 2007; Hedrick, Peterson, Vucetich, Adams, & Vucetich, 46 2014; W. E. Johnson et al., 2010; Madsen, Shine, Olsson, & Wittzell, 1999; Westemeier et al., 47 1998; Willi, Van Kleunen, Dietrich, & Fischer, 2007). Genetic rescue introduces or restores gene 48 flow between populations to alleviate the fitness consequences of inbreeding through the 49 introduction of genetic variation. However, while rare species may exhibit small effective 50 51 population sizes and reduced adaptive potential, disruption of local adaptation may lead to 52 outbreeding depression, or reduced fitness of progeny following admixture between genetically differentiated lineages (Hufford & Mazer, 2003). Thus, the contribution of natural selection to 53 54 the evolution of population genetic differences is a consideration for genetic rescue, as it may 55 ultimately lead to migrants or translocated individuals being maladapted (Lowry, Rockwood, & Willis, 2008; Nosil, Vines, & Funk, 2005). For rare species conservation, an understanding of 56 57 contemporary effective population size is therefore required to assess immediate genetic threats

to population persistence. Following these threats, however, understanding the history of population connectivity, the distribution of genetic variation within and among populations, and the role of selection in shaping population differences will be critical to informed management decisions.

62 In addition to guiding conservation management strategies, understanding rare species' 63 demographic and evolutionary history may prove valuable to optimizing strategies necessary to 64 preserve neutral and nonneutral genetic diversity ex situ. Ex situ conservation collections, or the preservation of species outside their natural range of occurrence, can complement in situ 65 66 conservation strategies (Cavender et al., 2015; Potter et al., 2017; Pritchard, Fa, Oldfield, & Harrop, 2012), providing a critical resource for the preservation of genetic variation, restoration 67 or reintroduction (Guerrant Jr, Havens, & Vitt, 2014; Potter et al., 2017). Ex situ sampling 68 designs traditionally rely on neutral population genetic structure to guide sampling decisions 69 (Caujapé-Castells & Pedrola-Monfort, 2004; Gapare, Yanchuk, & Aitken, 2008; Hoban, 2019; 70 71 Hoban & Schlarbaum, 2014). However, concerns exist regarding the sole use of neutral genetic 72 variability for species conservation, as variation at neutral loci is unlikely to reflect adaptive genetic diversity (Bonin, Nicole, Pompanon, Miaud, & Taberlet, 2007; Holderegger, Kamm, & 73 74 Gugerli, 2006; McKay & Latta, 2002; Teixeira & Huber, 2021). Ex situ population sampling 75 may need to evaluate the impact different evolutionary processes have had on population genetic 76 structure to optimize neutral and adaptive variation collected. Thus, an understanding of 77 population connectivity and the impact of selection across populations can inform ex situ 78 collection design. If empirical or simulated data suggest populations are genetically connected and genetic differentiation is low, then most neutral genetic variation may be captured within one 79 80 or a few populations. However, if selection overcomes the homogenizing effects of gene flow,

ensuring adaptive genetic differences are preserved for all populations will require the inclusion of diverse population origins, separation of such populations *ex situ*, and consideration of population origin in breeding programs.

84 With the advent of next-generation sequencing (NGS) and the ever-decreasing costs associated with these technologies, genome-wide estimates of genetic diversity can be readily 85 86 assessed and used to guide conservation management strategies. Combined with statistical and simulation-based tools, these data provide a powerful and timely means to evaluate aspects of 87 population genetic variation and spatial genetic structure critical to informing genetic rescue and 88 89 ex situ conservation plans, including both populations' demographic and adaptive history (Abebe, Naz, & Léon, 2015; Liu, Zhang, Wang, & Ma, 2020; Wang, Bernhardsson, & 90 Ingvarsson, 2020; Xia et al., 2018). However, in conifers, despite extensive use in characterizing 91 genomes as well as neutral and adaptive variation (Eckert et al., 2010; Namroud, Beaulieu, Juge, 92 Laroche, & Bousquet, 2008; Nystedt et al., 2013; Stevens et al., 2016; Tyrmi et al., 2020; Wang 93 et al., 2020), these data have only rarely been used to inform conservation management 94 95 decisions.

Torrey pine (Pinus torreyana Parry) is a critically endangered pine (IUCN, 2021), 96 endemic to California. One of the rarest pine species in the world (Critchfield & Little, 1966; 97 Dusek, 1985), Torrey pine's distribution spans one island population (*Pinus torreyana* subsp. 98 insularis) of approximately 3,000 reproductive individuals (Santa Rosa Island, CA), and one 99 100 mainland population (*Pinus torreyana* subsp. torreyana) of approximately 4,000 reproductive 101 individuals (Torrey Pine State Reserve in La Jolla, CA) (J. Franklin & Santos, 2011; Hall & 102 Brinkman, 2015). In addition to low population size, and despite current in situ and ex situ 103 conservation efforts, Torrey pine suffers from exceedingly low genetic variation and faces both

104 anthropogenic and environmental disturbances (J. Franklin & Santos, 2011; Hamilton, Royauté, 105 Wright, Hodgskiss, & Ledig, 2017; Ledig & Conkle, 1983; Waters & Schaal, 1991; Whittall et al., 2010). For these reasons, the species may be at imminent risk for population-scale extirpation 106 107 events and thus a potential candidate for genetic rescue. Inter-population admixture may increase population genetic diversity, alleviating potential fitness consequences associated with Torrey 108 109 pine's low genetic diversity (Hamilton et al., 2017), and increase evolutionary potential 110 necessary to respond to current and future ecological challenges (Carlson, Cunningham, & Westley, 2014). Previous research observed heterosis following one generation of admixture 111 112 between island and mainland individuals, suggesting that genetic rescue may alleviate fitness 113 consequences associated with reduce genetic variation (Hamilton et al., 2017). However, if 114 adaptive genetic differences have evolved between island and mainland populations, fitness 115 consequences following the disruption of co-adapted gene complexes may not be observed in the first generation cross. Thus, although the combination of exceedingly low genetic diversity and 116 117 conservation status suggest Torrey pine may be a candidate for genetic rescue, evaluation of the 118 species' demographic and adaptive evolutionary history will be necessary prior to inform conservation management decisions. 119

With this study, we use genomic data to quantify and model aspects of populations' demographic and evolutionary history necessary to preserve rare species' evolutionary potential. Specifically, we delineate the contribution of stochastic and deterministic processes to genomic differentiation in *Pinus torreyana*, asking three questions: (i) what are current effective population sizes and how have they changed over time, (ii) have populations remained genetically connected following isolation, and (iii) is there evidence of adaptive divergence between populations that may indicate distinct evolutionary trajectories? We identify the most 127 probable demographic scenario for Torrey pine using Approximate Bayesian Computing (ABC) 128 and coalescent simulations. To evaluate the role of selection, we identify loci that may be important to adaptation using various  $F_{ST}$  outlier methods and assess the functional significance 129 130 of these loci by annotating candidates with the gene ontology (GO) resource. Using this knowledge, we discuss conservation strategies for Torrey pine, including ex situ sampling to 131 132 preserve neutral and nonneutral processes within species collections, and possible risks associated with genetic rescue. This study demonstrates the benefits and necessity of 133 understanding the demographic and adaptive history of rare species to guide conservation. 134

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## **Material and Methods**

### 137 **Population sampling and DNA extraction**

Between June and July 2017, needle tissue was collected from individuals spanning the 138 entire natural distribution of Pinus torreyana (Torrey pine; Fig. 1). A total of 286 individuals 139 were sampled, including 146 individuals from the mainland population at the Torrey Pine State 140 141 Reserve (TPSR) near La Jolla, CA and 140 individuals from the island population on Santa Rosa Island, CA (SRI), one of the Channel Islands. Needles were dried in silica gel following which 142 143 genomic DNA was extracted using between 25-30 mg of dry needle tissue and a modified CTAB protocol (Doyle & Doyle, 1987). To reduce DNA shearing, slow manual shaking of tubes was 144 used. Following extraction, the concentration and purity of DNA extracted was quantified for 145 146 each sample using a NanoDrop 1000 Spectrophotometer (Thermo scientific) to ensure all 147 samples had a concentration of approximately 85  $ng/\mu l$  and purity ratios of 1.4 and above (average across samples; 260/280 = 1.85, 260/230 = 1.96). 148

### 149 Genomic library preparation and ddRAD sequencing

150 Genomic libraries were prepared for all 286 individuals following the protocol of 151 Parchman & colleagues (2012). Briefly, 510 ng (6 µl at 85 ng/µl) of DNA was digested using 152 endonucleases EcoRI and MseI (New England BioLabs, Inc.) after which barcoded (EcoRI cut 153 site) and non-barcoded (MseI cut site) adapters compatible with Illumina sequencing were ligated to each end of DNA fragments using T4 ligase (New England BioLabs, Inc.). A different 154 155 barcode sequence was used for each of the 286 samples. Due to the large, highly repetitive nature 156 of pines' genome (Stevens et al., 2016), we used the methylation-sensitive enzyme EcoRI, as it effectively reduces the presence of repetitive and non-coding DNA sequences in genomic 157 158 libraries (Parchman et al., 2012). Restriction-ligation products were amplified using two 159 successive PCR reactions to produce genomic libraries with concentrations necessary for sequencing (> 2nM). All PCR-amplified genomic libraries were subsequently pooled and sent to 160 161 the Genomic Sequencing and Analysis Facility (GSAF; Austin, TX) for size selection of fragments within the range of 450-500 bp and sequenced on 5 lanes of an Illumina HiSeq 2500 162 using the 100 bp single-end sequencing protocol (1 x 100 bp). 163

### 164 **De novo assembly and SNP calling**

Demultiplexing of sequence files was performed using *ipyrad* v0.9.12 (Eaton & Overcast, 165 166 2020) allowing one mismatch in the barcode sequence. Reads were filtered, assembled *de novo*, and used to call SNPs within the *dDocent* v2.7.8 pipeline (Puritz, Hollenbeck, & Gold, 2014; 167 Puritz, Matz, et al., 2014). Reads were filtered by removing low-quality bases at the beginning 168 169 and end of reads (PHRED score < 20), Illumina adapters, and trimmed when the average 170 PHRED score fell below 10 within a 5 bp window using the program TRIMMOMATIC (Bolger, Lohse, & Usadel, 2014). As a contiguous genome assembly for *P. torreyana* is not available, we 171 172 first generated a reference of genomic regions sampled with our sequencing design using a de 173 *novo* approach. Reads were clustered based on sequence similarity and assembled into a 174 reference assembly using the program CD-HIT (Fu, Niu, Zhu, Wu, & Li, 2012; W. Li & Godzik, 175 2006). To be included in *de novo* assembly, reads had to have a minimum of 3x within-176 individual coverage and be present in at least 5 individuals. To form a cluster (locus), reads had to have a minimum of 86% sequence similarity, a cutoff previously used in pines (Menon et al., 177 178 2018). This threshold was chosen as a tradeoff to avoid the clustering of paralogous loci while 179 still accounting for the presence of missing bases, errors, or polymorphisms between true homologous sequences. Finally, reads were mapped onto *de novo* assembled loci using BWA 180 181 MEM (H. Li, 2013) and SNPs were called using the software FREEBAYES (Garrison & Marth, 182 2012). Read mapping was performed using BWA default parameters, including a match value of 1, a mismatch penalty of 4 and a gap penalty of 6. This yielded a set of 652,492 SNPs that were 183 184 subjected to downstream filtering. Variants with genotype quality (GQ) < 20 and genotype depth < 3 were first marked as missing. Then, variants with PHRED scores (QUAL)  $\leq 30$ , minor allele 185 counts < 3, minor allele frequencies < 0.01, call rate across all individuals < 0.95, mean depth 186 across samples > 57 (based on the equation:  $d + 4\sqrt{d}$ , where d is the average read depth across 187 variants, H. Li, (2014),  $F_{IS}$  estimates < -0.5 or > 0.5, and linkage score ( $r^2$ ) > 0.5 within a 95 bp 188 189 window were removed from the raw SNP dataset. Following filtering, a total of 93,085 biallelic SNPs were kept and used for analysis (hereafter referred to as the full dataset). Note that 16 190 191 individuals with > 40% missing data were also discarded, leaving 270 genotyped individuals 192 (SRI: 130 individuals, TPSR: 140 individuals) for inclusion in analyses.

### **Population structure and genetic diversity analyses**

194 To describe and quantify contemporary genetic differences between Torrey pine 195 populations, we first assessed genetic structure of populations using principal component 196 analysis (PCA) implemented in the R package ADEGENET (Jombart, 2008; Jombart & Ahmed, 197 2011). Unless otherwise stated, analyses were performed in R version 3.6.2 and 4.0.2 (R Core Team, 2019, 2020). To quantify genetic differences between island and mainland populations, 198 199 we calculated Nei's  $F_{ST}$  statistic (Nei, 1987) for each SNP using the HIERFSTAT R package 200 (Goudet & Jombart, 2020) and averaged estimates across loci. A 95% confidence interval around 201 the mean was constructed by bootstrapping the empirical  $F_{ST}$  distribution 10,000 times using R 202 packages BOOT (Canty & Ripley, 2021; Davison & Hinkley, 1997) and SIMPLEBOOT (Peng, 2019). 203

204 To determine the extent of contemporary within-population genetic structure, the most 205 likely number of genetic clusters was independently evaluated for SRI and TPSR using the function *find.clusters()* implemented in the R package ADEGENET. This function transforms 206 207 genomic data using principal component analysis and performs successive K-means clustering with an increasing number of clusters (k). For each successive value of k, the Bayesian 208 209 Information Criterion (BIC) is computed and was used to assess the optimal number of clusters 210 (k). For each population, we assessed between 1 to 10 clusters while retaining principal components necessary to explain 90% of the variation after ordination (SRI: 114, TPSR: 122). 211 For TPSR, two individuals (TPSR5107, TPSR3189) clustered distantly from the population, 212 213 which may mask subtle within-population genetic structure. Consequently, we re-ran the analysis excluding the two individuals while maintaining the same range for k (1 to 10) and retaining 121 214 215 principal components (90.38% of variation explained after ordination).

To evaluate Torrey pine evolutionary potential, contemporary standing genetic variation within the species was estimated by calculating expected heterozygosity ( $H_E$ ), inbreeding coefficients ( $F_{IS}$ ), and coancestry coefficients ( $\theta$ ) for both island and mainland populations 219 independently. Values of H<sub>E</sub> and F<sub>IS</sub> were calculated for each SNP separately using the R 220 package ADEGENET and averaged across loci to provide population-level estimates. To evaluate  $\theta$ , we used the R package RELATED (Pew, Wang, Muir, & Frasier, 2015) that 221 estimates genetic relatedness between all possible pairs of individuals within a population. 222 223 Specifically, we used the triadic likelihood (TrioML) estimate of relatedness assuming no 224 inbreeding within populations. Averaging pairwise  $\theta$  values across all individuals within TPSR 225 and SRI provided population estimates of genetic relatedness. To build 95% confidence intervals 226 around  $H_E$ ,  $F_{IS}$  and  $\theta$  averages, the empirical distribution for each parameter within each 227 population was bootstrapped 10,000 times in R using the BOOT and SIMPLEBOOT packages.

## 228 ABC demographic modeling

229 **Demographic models** – To evaluate the impact genetic drift and gene flow have had to patterns of neutral genetic variation in Torrey pine, we quantified changes in effective population 230 231 size over time, interpopulation migration rate, and time since population divergence. To do so, 232 we tested six distinct demographic models that were classified into two broad categories: (1) 233 isolation with/without migration (Fig. 2A), and (2) two-population demic expansion (Fig. 2B, C). Models of isolation with or without migration (RPM1, RPM2) were developed to test a 234 hypothesis formulated by Ledig & Conkle, (1983). This hypothesis predicts that there was once a 235 236 single ancestral population of Torrey pine that diverged to form one island and one mainland population following tectonic movement (Fig. 2A). These models assume that an ancestral 237 population with an effective size N<sub>A</sub> diverged T<sub>Div</sub> generations before present to form two 238 populations with current effective sizes N<sub>I</sub> (island, SRI) and N<sub>M</sub> (mainland, TPSR). Following 239 240 divergence, to assess whether gene flow has occurred between populations, bidirectional

migration was either prevented (RPM1,  $m_{IM}=m_{MI}=0$ ) or permitted (RPM2,  $m_{IM}=m_{MI}>0$ ). Both models assume constant island and mainland effective population size following divergence.

The remaining four models (CM1, CM2, CM3, CM4) tested two different hypotheses of 243 244 land colonization where one population was founded by the other (Fig. 2B, C). Models CM1 and CM2 specifically test the hypothesis that Santa Rosa Island was colonized by a subset of 245 246 mainland individuals (Ledig & Conkle, 1983). In this scenario, SRI was founded by N<sub>C</sub> effective migrants from TPSR T<sub>Div</sub> generations ago and grew exponentially at a rate  $r_I (r_I = log \left(\frac{N_C}{N_I}\right)/T_{Div})$ 247 to form a population with an effective size N<sub>I</sub>. TPSR effective population size (N<sub>M</sub>) was assumed 248 249 constant. As above, bidirectional migration between populations was either prevented (CM1, 250 m<sub>IM</sub>=m<sub>MI</sub>=0) or permitted (CM2, m<sub>IM</sub>=m<sub>MI</sub>>0) to evaluate whether gene flow has occurred between populations following colonization. Models CM3 and CM4 test the hypothesis that the 251 252 mainland population was founded by a subset of island individuals (Haller, 1986). This scenario 253 assumes TPSR was founded by N<sub>C</sub> effective migrants from SRI T<sub>Div</sub> generations before present. 254 SRI effective population size (N<sub>I</sub>) was assumed constant while TPSR effective population size (N<sub>M</sub>) grew exponentially at rate  $r_M (r_M = log \left(\frac{N_C}{N_M}\right)/T_{Div})$ . Once again, to test whether gene flow 255 has occurred between populations since colonization, exchange of migrants between population 256 257 was either prevented (CM3, m<sub>IM</sub>=m<sub>MI</sub>=0) or permitted (CM4, m<sub>IM</sub>=m<sub>MI</sub>>0).

For all six models, uniform priors were used except for  $T_{Div}$ ,  $N_C$ , and  $N_A$  for which we used log-uniform priors. Priors on a logarithmic scale increases the weight given to small values and is recommended when parameters' ranges span several orders of magnitude (Wegmann, Leuenberger, & Excoffier, 2009). Details on demographic parameters and their prior distributions are provided in Table 1. 263 Additional filtering and down-sampling of genetic variants - Accurate estimation of 264 demographic parameters using coalescence simulations requires the use of neutrally evolving 265 genetic markers. To ensure neutrality of SNPs, we filtered the full dataset for SNPs that did not 266 deviate significantly from Hardy-Weinberg equilibrium (HWE). Since population structure may create departures from HWE, we applied this filter to island and mainland populations 267 independently and removed SNPs that deviated significantly from HWE (P < 0.05) in both 268 populations. This was performed using a customized R function relying on R packages 269 270 ADEGENET and PEGAS (Paradis, 2010). In total, 73,928 SNPs were retained following 271 filtering for use in demographic modelling (hereafter referred to as the HWE-filtered dataset).

For computational efficiency, we down sampled the HWE-filtered dataset from 73,928 to 9,795 variants first by generating bivariate bins based on observed heterozygosity and Nei's  $F_{ST}$ (0.05-interval bins), and then by subsampling each bin proportionally to the number of SNPs they contained. In this way, each bin is subsampled to reflect its contribution to the HWE-filtered dataset (Appendix S1). Following subsampling, we conducted a principal component analysis using the down-sampled dataset to ensure patterns of genetic diversity and population structure were maintained between datasets (Appendix S2).

*Generating coalescent simulations and estimating summary statistics* – To evaluate and compare demographic scenarios, a set of 200,000 simulations was generated using ABCSAMPLER for each model, a wrapper program included in the package ABCTOOLBOX (Wegmann, Leuenberger, Neuenschwander, & Excoffier, 2010). For each simulation, ABCSAMPLER samples prior ranges of demographic parameters and uses these values as inputs for coalescence simulations within a user-defined simulation program. We used FASTSIMCOAL version 2.6.0.3 (Excoffier, Dupanloup, Huerta-Sánchez, Sousa, & Foll, 2013; 286 Excoffier & Foll, 2011) to simulate 9,795 unlinked SNPs with a minor allele frequency of 0.01, 287 reflecting the composition of the down-sampled genetic dataset. For each model, simulated data were output as genotypes and fed to a user-defined program by ABCSAMPLER to estimate 288 289 population genetic summary statistics. ARLEQUIN version 3.5.2.2 (Excoffier & Lischer, 2010) 290 was used to calculate ten distinct population genetic summary statistics, specifically aiming at 291 quantifying genetic variation and divergence within and between Torrey pine populations. These 292 included genetic diversity (i.e., population-specific heterozygosity and number of alleles, average heterozygosity and number of alleles over loci and populations, and mean total heterozygosity), 293 294 genetic differentiation (i.e., pairwise  $F_{ST}$ ), and variance (i.e., standard deviation over populations of the average heterozygosity and number of alleles) statistics. Finally, to obtain observed 295 296 population genetic summary statistics, ARLEQUIN version 3.5.2.2 was rerun using the down-297 sampled dataset. Characterizing and summarizing the amount and distribution of genetic variation within each dataset, these statistics were used to calculate posterior probabilities and 298 299 identify the demographic model with greatest support, as well as estimate demographic 300 parameters associated with that model (see below).

ABC parameter estimation – Demographic parameters were estimated in R using the 301 302 ABC package (Csilléry, François, & Blum, 2012). Cross-validation simulations were conducted first to evaluate the ability of summary statistics to distinguish between demographic models 303 (Appendix S3). We performed leave-one-out cross-validations, consisting of selecting one 304 305 simulation of a demographic scenario and then assigning it to one of the six models using 306 posterior probabilities estimated from all remaining simulations. This was repeated one hundred times for each demographic model, generating a confusion matrix. If misclassification rates 307 308 (proportions of simulations incorrectly assigned to a model) are low, then computed summary 309 statistics can distinguish between our different demographic scenarios. The posterior probability 310 of each demographic model was approximated as the proportion of accepted simulations and 311 used to select the best model following cross-validation. To ensure the best model provided a 312 good fit to the data, we performed a goodness-of-fit test as implemented in the function gfit(). Finally, demographic parameters associated with the best model were estimated as the weighted 313 314 medians of posterior distributions using the *weighted.median()* function implemented in the R package SPATSTAT (Baddeley, Rubak, & Turner, 2015). Posterior distributions were created 315 from the set of accepted simulations using a non-linear postsampling regression adjustment 316 317 conducted on log-transformed data (Blum & François, 2010). Ninety-five percent confidence 318 intervals around weighted medians were estimated using 10,000 bootstrap replicates of posterior distributions in R, using BOOT, SIMPLEBOOT and SPATSTAT packages. The validity and 319 320 accuracy of each estimated parameter were tested using additional, yet distinct, 100-fold leaveone-out cross-validation simulations (Appendix S4). The cross-validation process begins with 321 322 the random selection of one simulation generated by the best demographic model. Summary 323 statistics associated with that simulation are considered as pseudo-observed data and its demographic parameters are estimated using remaining simulations for the model. If pseudo-324 325 observed parameters can accurately be predicted, then inferred demographic parameters from true observed data can be considered valid and accurate. Cross-validation simulations, model 326 selection, model validation, and parameters estimation were conducted using a tolerance 327 328 threshold of 0.01, a tradeoff between retaining a reasonable number of simulations to estimate 329 posterior distributions and keeping the tolerance value as low as possible (S. Li & Jakobsson, 2012). Finally, as census sizes for Torrey pine populations are available (J. Franklin & Santos, 330 331 2011; Hall & Brinkman, 2015), we calculated the proportion of the census size (N) to effective

population size (Ne) for each population separately. Of all reproductive trees present within a
 population (census size), this ratio estimated the proportion contributing to the next generation
 (effective size).

335 Simulating the null F<sub>ST</sub> distribution

To evaluate the influence natural selection may have had on the genomic structure of 336 337 Torrey pine populations, we compared the distribution of Nei's  $F_{ST}$  estimated from the full SNP dataset with a simulated distribution based on 93,085 independent SNPs from 270 individuals 338 generated using SIMCOAL2 version 12.09.07 (Laval & Excoffier, 2004). We used weighted 339 340 medians estimated from posterior distributions borrowed from the best demographic model (see Results) as input parameters for neutral simulations. For each simulated SNP, the minimum 341 allele frequency was set to 0.01 to reflect filters applied to the full dataset. Locus-specific Nei's 342 F<sub>ST</sub> values were estimated for both full and simulated datasets in R using the HIERFSTAT 343 package (Appendix S5). 344

#### 345 **Outlier detection analyses**

To estimate the potential contribution of local adaptation to genomic differentiation 346 between Torrey pine populations, we used the full dataset of 93,085 SNPs to identify loci 347 348 putatively under selection using three distinct methods: BAYESCAN version 2.1 (Foll & Gaggiotti, 2008), OUTFLANK version 0.2 (Whitlock & Lotterhos, 2014) and PCADAPT 349 version 4.3.3 (Privé, Luu, Vilhjálmsson, & Blum, 2020). These three approaches were selected 350 351 for their ability to account for neutral population structure (BAYESCAN, OUTFLANK) and to 352 handle genetic admixture between individuals (PCADAPT). For both BAYESCAN and OUTFLANK, we grouped all 270 individuals by populations (mainland: 140 individuals, island: 353

130 individuals). Below is a brief description of all three methods used to identify candidateSNPs.

The first approach we used was BAYESCAN. For each locus, BAYESCAN uses a 356 357 Bayesian approach to decompose  $F_{ST}$  coefficients into population- and locus-specific components using a logistic regression. Loci are identified as putatively under selection if the 358 359 locus-specific component is needed to explain the observed distribution of genetic diversity. Our analysis was conducted using BAYESCAN default parameters. Results were visualized and 360 analyzed in R. Only SNPs with a false discovery rate of 1% or below were retained and 361 362 considered as candidate loci. The second approach we used was OUTFLANK, an R package which identifies outliers by inferring a null  $F_{ST}$  distribution approximated from empirical  $F_{ST}$ 363 values. This distribution was produced by removing loci with an expected heterozygosity below 364 365 0.1 (Hmin = 0.1), trimming 5% of lowest and highest empirical  $F_{ST}$  values (RightTrimFraction = LeftTrimFraction = 0.05), and retaining only loci passing a built-in neutrality test with a false 366 367 discovery rate of 0.1% (qthreshold = 0.001). Using the latter threshold provided a conservative 368 estimate for neutral F<sub>ST</sub>. Outlier loci were identified by comparing the empirical F<sub>ST</sub> distribution 369 to the inferred null  $F_{ST}$  distribution using a built-in chi-squared test with a false discovery rate of 1% (qthreshold = 0.01). Note that loci with  $H_E < 0.1$  were excluded while conducting the chi-370 371 squared test and therefore could not be identified as potential outliers (Hmin = 0.1). The third and last approach we used was PCADAPT. Also implemented in R, PCADAPT is a package that 372 373 assesses population structure using PCA. Consequently, this approach does not require 374 individuals to be grouped into populations. Following PCA, candidate loci are identified as those substantially correlating with population structure. We ran PCADAPT retaining the first axis of 375 376 differentiation (K = 1) and SNPs with a minor allele frequency of 0.01 (min.maf = 0.01). We

only considered the first principal component to calculate the test statistic, as additional axes did not ascertain population structure (Appendix S6). Candidate loci were identified as the set of SNPs with a false discovery rate of 1% or below. To minimize the presence of false positives within our dataset, only outlier SNPs common to all three approaches were considered as candidate loci and included in subsequent analyses. Finally, to both visualize and quantify genetic structure at putatively adaptive loci, we conducted a principal component analysis based only on candidate SNPs shared by all three methods using the R package ADEGENET.

## 384 Functional categorization of candidate loci

385 To identify biological processes or molecular functions that may play a role in adaptation 386 to mainland or island environments, de novo assembled sequences containing candidate SNPs common to all genome scans were first extracted and then annotated using BLASTx version 387 388 2.6.0+. Sequences were blasted against the UniProt protein database filtered for sequences from species within the *Pinaceae* family (Taxon identifier 3318). Sequence similarity was assessed 389 using BLASTx default parameters, an e-value hit filter of  $10^{-3}$  (-evalue 0.001), and a number of 390 391 database hits to retain of 1 (-max\_target\_seqs 1). Gene ontology terms were mapped onto annotated sequences in R using the UNIPROTR package (Soudy et al., 2020). 392

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### Results

# **395 Population genetic structure and variation**

Using all 93,085 SNP markers, the principal component analysis revealed little genomewide differentiation in Torrey pine with the first two principal components (PC1, PC2) explaining only 1.9 and 0.6% of observed genetic differences, respectively (Fig. 3). Nonetheless, PC1 unambiguously separated island from mainland individuals, suggesting some level of genetic differentiation exists between populations. These results were further supported by the 401 low average coefficient of genetic differentiation found across loci (Nei's F<sub>ST</sub>), estimated at 402 0.013 (95% CI: 0.012 - 0.013). In addition to population-scale genetic differentiation, withinpopulation genetic differentiation was estimated to evaluate whether local inbreeding, possibly 403 404 increased in small populations, could have contributed to fine-scale genetic structure in Torrey pine. For both island and mainland populations, we found no evidence of within-population 405 406 genetic structure. Population-specific principal component analyses identified no clear genetic clusters and revealed that, combined, the first two axes of differentiation (PC1, PC2) only 407 explained approximately 1.9% and 2.2% of mainland and island within-population genetic 408 409 differences, respectively (Fig. 4). In addition, evaluating the likelihood between 1 to 10 genetic 410 clusters (k) within each population using BIC indicated that populations appear largely homogeneous (most likely k = 1) (Appendix S7). Interestingly, average estimates of inbreeding 411 and coancestry coefficients across loci were low for both the island and the mainland population 412 (Table 2). While low inbreeding coefficients support the lack of observable within-population 413 genetic structure, this also indicates that reproduction among relatives or unequal reproductive 414 415 success among individuals is unlikely to have contributed to low expected heterozygosity observed within populations (Table 2). Combined, our results indicate that Torrey pine exhibits 416 exceedingly low genetic diversity, with the majority of variation distributed within genetically 417 unstructured populations. 418

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## **Demographic history of Torrey pine**

Of the six demographic models evaluated (Fig. 2), the isolation with migration model (RPM2) received the most support with a posterior probability of 92.18%. The remaining five models exhibited lower posterior probabilities ranging from 0% to 3.98%. With low misclassification rates, cross-validations indicated that simulated summary statistics were able to 424 confidently distinguish between different demographic scenarios (Appendix S3). The goodness-425 of-fit test revealed that simulated summary statistics did not significantly differ from observed 426 ones (P = 0.76), providing a good fit to the data.

427 Based on RPM2, an ancestral Torrey pine population with an effective size of approximately 1,124 individuals (95% CI: 939 - 1,213) diverged during the early Pleistocene 428 approximately 1.2 million YBP (95% CI: 1,195,367 - 1,296,186, assuming a generation time of 429 ten years) to form one island and one mainland population with effective sizes of approximately 430 2,305 (95% CI: 2,166 - 2,338) and 1,715 (95% CI: 1,616 - 1,759) individuals, respectively (Fig. 431 432 5). This resulted in a 0.75 ( $N_I/N = 2,305/3,063$ ) proportion of the census to effective population size on the island, and a 0.45 ( $N_M/N = 1,715/3,806$ ) proportion of the census to effective 433 population size on the mainland. Following divergence, some gene flow was maintained between 434 populations with an estimated migration rate of 8.34 x  $10^{-3}$  (95% CI: 8.15 x  $10^{-3}$  - 8.76 x  $10^{-3}$ ) per 435 generation. In general, cross-validation simulations indicated low prediction errors (Appendix 436 S4), suggesting high accuracy of inferred parameters. Nonetheless, note that for  $T_{Div}$ , the 437 438 associated prediction error was higher than for other parameters.

### 439 Divergent selection between island and mainland populations

Despite limited genetic variation within populations, we found some evidence for the evolution of genetic differences among populations at a subset of loci. We compared the neutral  $F_{ST}$  distribution generated from the simulated RPM2 demographic model with the empirical distribution based on our full dataset of SNP variants (Appendix S5). For select SNPs, moderate to high empirical  $F_{ST}$  values (from approximately 0.2 to 0.65) could not be generated through neutral simulations, suggesting they may be candidate for selection. PCADAPT, BAYESCAN, and OUTFLANK identified 3,138 (3.37%), 2,163 (2.32%), and 3,942 (4.23%) outlier SNPs, 447 respectively (Fig. 6A). Of these outlier loci, 2,053 (2.21%) were common to all three methods 448 and contribute to genomic structure between Torrey pine populations (Fig. 6B). Indeed, the principal component analysis revealed that the first axis of differentiation (PC1) unambiguously 449 450 differentiated island from mainland populations based on common outlier SNPs, explaining over 20% of observed genetic differences. Consistent with these results, F<sub>ST</sub> values for putatively 451 452 adaptive loci ranged between 0.1 and 0.63 and either could not be generated or could only be 453 generated at low frequency through neutral simulations, representing only 0.078% of all simulated F<sub>ST</sub> values. 454

455 Functional categorization of common outlier loci was performed by blasting *de novo* 456 assembled contigs carrying outlier SNPs against the Pinaceae UniProt protein database and 457 retrieving each hit's Gene Ontology terms. Overall, 110 (7.51%) contigs were annotated. After 458 accounting for redundancy in the data (i.e., different contigs aligning to the same locus), we identified a total of 80 putative adaptive genes with homologous sequences in Larix, Picea, or 459 *Pinus* species (Appendix S8) that may be targets of selection. Functionally, these genes are 460 461 primarily encoded in mitochondria, involved in the process of DNA integration, or with processes associated with molecular functions such as RNA-directed DNA polymerase activity 462 463 or nucleic acid binding (Table 3).

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### Discussion

An understanding of demographic and adaptive evolutionary history is invaluable for rare species of conservation concern, particularly when management decisions impact populations at risk of extinction. Teasing apart the contribution of both stochastic and deterministic evolutionary processes to population genomic differentiation over time and space can be used to 470 inform species conservation decisions, including the potential consequences of genetic rescue 471 (Frankham et al., 2011; Hufford & Mazer, 2003; Ralls et al., 2018). Here, we evaluated the genomics of Torrey pine, a critically endangered endemic isolated to two populations in 472 473 California. We modeled demographic change and connectivity over time and tested the influence 474 of neutral and selective processes have had on contemporary population genomic structure. We 475 observed that Torrey pine populations exhibited exceedingly low genetic variation, particularly 476 for a conifer (see below), with little within- or among-population structure. Although some 477 connectivity has been maintained between island and mainland populations, demographic 478 modeling indicates that Torrey pine has consistently suffered from low effective population size. Genome scans revealed over 2000 loci that were candidates for selection. Consistent with 479 previous observations indicating phenotypic differences between island and mainland 480 populations, these data suggest adaptive genetic differences have evolved among populations (Di 481 Santo et al., in review; Haller, 1986; Hamilton et al., 2017). From a conservation standpoint these 482 results lead to contrasting recommendations with respect to genetic rescue. A history of reduced 483 484 effective population size and low genome-wide differentiation at neutral loci indicate little genetic differentiation among populations that may impact a genetic rescue program. However, 485 previous observations of phenotypic differences paired with loci associated with divergent 486 selection point towards the importance of adaptive evolution among Torrey pine populations. 487 These results suggest increased genetic variation via inter-population crosses may be needed in 488 489 this species, but admixture should be evaluated first to quantify its fitness impact.

490 Standing genetic variation

491 Torrey pine populations exhibited extremely low genetic variation in comparison with 492 other conifers which often exhibit an expected heterozygosity around 0.3 within populations 493 (e.g., Namroud et al., 2008; Tsumura, Uchiyama, Moriguchi, Ueno, & Ihara-Ujino, 2012). 494 Average expected heterozygosity  $(H_{\rm E})$  and contemporary effective population size (Ne) were 495 estimated at 0.185 and 2,305 individuals for the island population, and 0.184 and 1,715 496 individuals for the mainland population, respectively (Table 2; Fig. 5). Although low, this study is the first to find genetic variability within island and mainland populations of Torrey pine. 497 498 While previous genetic analyses using allozymes and chloroplast DNA markers identified fixed genetic differences between populations, they failed to observe genetic variation within 499 populations (Ledig & Conkle, 1983; Whittall et al., 2010). Ledig & Conkle, (1983) hypothesized 500 501 reduced genetic diversity was attributable to drastically reduced mainland and island populations 502 in the recent geological past. They suggested that Torrey pine populations declined to fewer than 50 individuals, following which a recovery led to approximately 3,000 to 4,000 reproductively 503 504 mature trees (J. Franklin & Santos, 2011; Hall & Brinkman, 2015). Demographic models herein, 505 however, suggest that effective population size has always been low for Torrey pine (Fig. 5). The 506 best fit demographic model predicted an ancestral effective population size  $(N_A)$  of 1,124 and 507 only limited change following population divergence (Fig. 5). Given these observations, longterm reduced effective population size has likely exacerbated the consequences of genetic drift 508 509 leading to an extreme lack of genetic variation within Torrey pine populations.

Despite extremely low genetic variation and small effective population sizes, there was no evidence for inbreeding ( $F_{IS}$ ) or excessive relatedness ( $\theta$ ) within populations (Table 2). These findings support high ratios between effective and census population size (Ne/N) found in both Torrey pine populations (island = 0.75, mainland = 0.45) and may, at least partly, explain why these estimates were higher than ratio averages of 0.1 to 0.2 typically recommended for conservation management (Frankham et al., 2014). Indeed, it is not uncommon for plants to 516 exhibit Ne/N ratios greater than 0.2 (Hoban et al., 2020; Waples, Luikart, Faulkner, & Tallmon, 517 2013). Combined with a lack of within population genetic structure (Fig. 4, Appendix S7), these results also indicate that neither reproduction among relatives nor unequal reproductive success 518 519 has likely contributed to reduced genetic variation within populations. Wind pollination and zoochorous seed dispersal have likely contributed to homogenizing the gene pool within 520 521 populations (M. Johnson, Vander Wall, & Borchert, 2003; Loveless & Hamrick, 1984). Pines also possess mechanisms that can reduce the probability of self-fertilization, including the 522 embryo lethal system (Williams, 2009; Williams, Zhou, & Hall, 2001). This self-incompatibility 523 524 system selectively induces death in embryos resulting from self-fertilization (Bramlett & Popham, 1971; Williams, Barnes, & Nyoka, 1999). Consequently, increased dispersal potential 525 paired with post-zygotic barriers limiting the probability of mating among relatives have likely 526 527 reduced within-population genetic structure in Torrey pine.

## 528 Neutral genetic differences across populations over time

529 Demographic modelling using neutral genomic variation supports the maintenance of 530 some genetic connectivity following population divergence approximately 1.2 MYA (Fig. 5), estimating the probability of gene exchange at  $8.34 \times 10^{-3}$  per generation. Despite geographic 531 532 isolation among populations and reduced potential for inter-population gene flow, contemporary estimates of F<sub>ST</sub>=0.013 indicate only subtle genome-wide differentiation between island and 533 mainland populations. Reduced genetic differentiation is typical of many conifers (Eckert et al., 534 535 2010; Namroud et al., 2008; Tyrmi et al., 2020), as pollen may maintain connectivity over very 536 long distances (Campbell, McDonald, Flannigan, & Kringayark, 1999; Varis, Pakkanen, Galofré, & Pulkkinen, 2009; Williams, 2010). Gene flow between populations may also have been 537 538 maintained via seed dispersal. Birds represent potential seed dispersers for Torrey pine and may

play a prominent role in long-distance seed dispersal (M. Johnson et al., 2003; Pesendorfer, 539 540 Sillett, Koenig, & Morrison, 2016; Viana, Gangoso, Bouten, & Figuerola, 2016). Interestingly, higher estimates of contemporary effective population size (N<sub>I</sub>, N<sub>M</sub>) relative to the ancestral 541 542 population size  $(N_A)$  suggest that island and mainland populations have experienced genetic bottlenecks following one or multiple moderate population expansion events (Fig. 5). Overall, 543 our findings indicate that despite the increased probability of genetic drift due to genetic 544 bottlenecks and low population sizes, gene flow maintained between island and mainland 545 populations may have been sufficient to prevent extensive genomic differentiation at neutral loci 546 547 following population isolation. Note, however, that coalescent simulations assume nonoverlapping generations, which may limit their ability to accurately estimate demographic 548 parameters in long-lived species, including conifers. Consequently, gene flow estimated between 549 island and mainland populations may have been overestimated or may possibly be an artefact 550 resulting from an attempt of the demographic model to account for shared ancestral genetic 551 552 variation among populations.

### 553 **Evidence for local adaptation**

Phenotypic monitoring using common garden experiments or in situ morphological 554 555 observations for cone, seed, and needle morphology have previously suggested genetically-based 556 phenotypic divergence among Torrey pine populations (Di Santo et al., *in review*; Haller, 1986; Hamilton et al., 2017). Thus, subtle genetic differentiation observed among populations likely 557 558 reflects the fact that most genome divergence among populations is the result of neutral rather 559 than adaptive differentiation among populations. To test for the role of selection across loci, we simulated a null F<sub>ST</sub> distribution to compare with our empirical F<sub>ST</sub> distribution, which indicated 560 561 that a few thousand loci may be under divergent selection (Appendix S5). Genome scans further

562 supported this observation, identifying 2,053 (2.21%) candidate SNPs with accentuated 563 divergence between island and mainland populations (Fig. 6). Annotation of sequences 564 containing these outliers SNPs suggested that adaptive evolution in Torrey pine may not only 565 result from genetic differentiation at the nuclear level, but also at the mitochondrial level (Table 3). This could be consistent with previous observations of the importance of cytoplasmic genetic 566 567 differences as a factor contributing to local adaptation in plants (Hamilton & Aitken, 2013; Leinonen, Remington, Leppälä, & Savolainen, 2013; Leinonen, Remington, & Savolainen, 2011; 568 569 for a review see Bock, Andrew, & Rieseberg, 2014). For example, Leinonen et al., (2011) found 570 using a reciprocal transplant experiment that individuals of Arabidopsis lyrata harboring the local cytoplasmic genome had higher fitness than individuals harboring the non-local 571 cytoplasmic genome, suggesting that cytoplasmic genetic variation may contribute to local 572 573 adaptation.

Overall, GO annotation of both nuclear and mitochondrial encoded candidate genes 574 575 indicated that genes important for mechanisms such as DNA integration, methylation, gene 576 silencing, carbohydrate transport and metabolic processes, and defense against pathogens (bacteria and fungi) were candidates for selection (Table 3). This suggests that between the 577 island and mainland environments, modification of genetic composition and architecture 578 579 following DNA integration, changes in gene expression or protein function following methylation and gene silencing, and direct or indirect selection against pathogens may have 580 581 played an important role in population divergence following isolation. For example, a candidate 582 gene associated with defense against bacteria and fungi (UniProt accession: B8LLJ5, GO terms: GO:0042742, GO:0050832, GO:0031640) suggests that phenotypic differentiation may have 583 584 evolved in response to pests or pathogens. Indeed, the mainland population of Torrey pine may 585 have faced substantial selection associated with the recent outbreak of the California five-spined 586 ips (Ips paraconfusus Lanier) (J. Franklin & Santos, 2011; Shea & Neustein, 1995), whereas the 587 island population may not have been exposed to that selective pressure. Noteworthy with these 588 results is the fact that pine genomes are enormous (Grotkopp, Rejmánek, Sanderson, & Rost, 2004; Stevens et al., 2016), and our sequencing approach (reduced representation sequencing; 589 590 see Material and Methods) represents only a fraction of the Torrey pine genome. This suggests, despite the differences observed, most likely some variation has been overlooked that plays a 591 critical role in local adaptation for this species. 592

## 593 Applying neutral and adaptive evolutionary processes to rare species conservation

While identification of the appropriate effective population size necessary to protect 594 adaptive evolutionary potential for rare species is still debated, recommendations generally range 595 596 between 500 to 5000 individuals (Frankham et al., 2014; I. R. Franklin & Frankham, 1998; Lynch & Lande, 1998). Torrey pine, critically endangered and endemic to just two native 597 598 populations, suffers from extremely low effective population size ( $N_I = 2,305$ ,  $N_M = 1,715$ ) 599 relative to other pines (Menon et al., 2018; Xia et al., 2018). In addition, there is clear evidence of the impact of adaptive evolutionary processes alongside neutral processes structuring genomic 600 601 variation within and among populations. Given historical and contemporary estimates of 602 effective population size as well as contemporary estimates of expected heterozygosity, our results indicate that Torrey pine may not retain the genetic variation within populations needed to 603 604 adapt to change. Current monitoring within the mainland population suggests that a lack of 605 recruitment (personal observation), infestation by *Ips* beetles (personal observation), and climate warming (Diffenbaugh, Swain, & Touma, 2015) may increase extinction risk. Thus, for Torrey 606 607 pine, increased Ne and greater genetic diversity may be required for long-term persistence.

608 As genetic variation is extremely low within populations, one conservation strategy that 609 may facilitate the maintenance of genetic variation within populations at risk is a genetic rescue 610 program. A genetic rescue program would facilitate inter-population breeding as a means to 611 increase heterozygosity, increasing rates of inter-population gene flow. Indeed, demographic modeling indicates that following population isolation some gene flow has been maintained and 612 there are low levels of genome-wide differentiation among populations (Nei's  $F_{ST} = 0.013$ ). 613 However, the combination of observed phenotypic differences and large number of genes that 614 appear targets of selection suggest island and mainland populations of Torrey pine have 615 616 undergone distinct evolutionary trajectories necessary for adaptation following isolation. Thus, a 617 genetic rescue program should be considered with caution as gene flow between populations may disrupt local adaptation and further reduce population performance (Goto, Iijima, Ogawa, & 618 619 Ohya, 2011; Hufford & Mazer, 2003; Montalvo & Ellstrand, 2001). Despite this word of caution, preliminary data comparing mainland, island, and F1 individuals from a common garden 620 experiment planted outside the species natural distribution indicate that F1s exhibit increased 621 622 fitness relative to mainland and island populations (Hamilton et al., 2017). Consequently, future monitoring is needed to empirically quantify fitness consequences of advanced-generation 623 624 admixture (F2, Backcross-Island (BC-I), Backcross-Mainland (BC-M)) following earlygeneration heterosis. 625

Given the challenge to conserve and manage rare species in a rapidly changing environment, the use of genomic data to model evolutionary history, assess demographic change, and tease apart the contributions of neutral and adaptive processes will be critical. For Torrey pine, the fact that there is low genome-wide differentiation among populations, a consistent history of low effective population size, and indications that some gene flow is maintained among populations may suggest that one population (island or mainland) could be targeted to effectively preserve neutral genetic variation. However, the combination of outlier loci and previously observed phenotypic differences suggest if the goal is to preserve adaptive genetic variation, a strategy that favors conservation efforts across both mainland and island populations will be needed. If conservation strategies such as genetic rescue are considered, assessment of multiple admixed generations within a common environment will provide the necessary empirical test to evaluate the consequences of enhancing genetic exchange among populations.

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652	corretakers of the Terrey pine acceptations sampled for this study. Any use of product names is for
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656	References
657	Abebe, T. D., Naz, A. A., & Léon, J. (2015). Landscape genomics reveal signatures of local
658	adaptation in barley (Hordeum vulgare L.). Frontiers in Plant Science, 6, 813. doi:
659	10.3389/fpls.2015.00813
660	Aguilar, R., Quesada, M., Ashworth, L., Herrerias Diego, Y., & Lobo, J. (2008). Genetic
661	consequences of habitat fragmentation in plant populations: susceptible signals in plant
662	traits and methodological approaches. Molecular Ecology, 17(24), 5177-5188. doi:
663	10.1111/j.1365-294X.2008.03971.x
664	Baddeley, A., Rubak, E., & Turner, R. (2015). Spatial point patterns: methodology and
665	applications with R. New York: CRC press.
666	Blum, M. G. B., & François, O. (2010). Non-linear regression models for Approximate Bayesian
667	Computation. Statistics and Computing, 20(1), 63-73. doi: 10.1007/s11222-009-9116-0
668	Bock, D. G., Andrew, R. L., & Rieseberg, L. H. (2014). On the adaptive value of cytoplasmic
669	genomes in plants. <i>Molecular Ecology</i> , 23(20), 4899–4911. doi: 10.1111/mec.12920
670	Bolger, A. M., Lohse, M., & Usadel, B. (2014). Trimmomatic: a flexible trimmer for Illumina
671	sequence data. <i>Bioinformatics</i> , 30(15), 2114–2120. doi: 10.1093/bioinformatics/btu170
672	Bonin, A., Nicole, F., Pompanon, F., Miaud, C., & Taberlet, P. (2007). Population adaptive
673	index: a new method to help measure intraspecific genetic diversity and prioritize
674	populations for conservation. Conservation Biology, 21(3), 697-708. doi: 10.1111/j.1523-

## 675 1739.2007.00685.x

Bossuyt, B. (2007). Genetic rescue in an isolated metapopulation of a naturally fragmented plant
species, Parnassia palustris. *Conservation Biology*, *21*(3), 832–841. doi: 10.1111/j.1523-

678 1739.2006.00645.x

- Bramlett, D. L., & Popham, T. W. (1971). Model relating unsound seed and embryonic lethal
  alleles in self-pollinated pines. *Silvae Genet*, *20*, 192–193.
- 681 Campbell, I. D., McDonald, K., Flannigan, M. D., & Kringayark, J. (1999). Long-distance
- transport of pollen into the Arctic. *Nature*, *399*(6731), 29–30. doi: 10.1038/19891
- 683 Canty, A., & Ripley, B. D. (2021). *boot: Bootstrap R (S-Plus) Functions*.
- Carlson, S. M., Cunningham, C. J., & Westley, P. A. H. (2014). Evolutionary rescue in a
  changing world. *Trends in Ecology and Evolution*, 29(9), 521–530. doi:
  10.1016/j.tree.2014.06.005
- Caujapé-Castells, J., & Pedrola-Monfort, J. (2004). Designing ex-situ conservation strategies
  through the assessment of neutral genetic markers: application to the endangered
  Androcymbium gramineum. *Conservation Genetics*, 5(2), 131–144. doi:
  10.1023/B:COGE.0000029997.59502.88
- 691 Cavender, N., Westwood, M., Bechtoldt, C., Donnelly, G., Oldfield, S., Gardner, M., ...
- McNamara, W. (2015). Strengthening the conservation value of ex situ tree collections.
- 693 *Oryx*, 49(3), 416–424. doi: 10.1017/S0030605314000866
- 694 Critchfield, W. B., & Little, E. L. (1966). *Geographic distribution of the pines of the world*.
  695 Washington, DC: US Department of Agriculture, Forest Service.

696	Csilléry, K., François, O., & Blum, M. G. B. (2012). abc: an R package for approximate
697	Bayesian computation (ABC). Methods in Ecology and Evolution, 3(3), 475-479. doi:
698	10.1111/j.2041-210X.2011.00179.x

- Davison, A. C., & Hinkley, D. V. (1997). *Bootstrap methods and their application*. Cambridge:
  Cambridge University Press.
- voo Camonage Oniversity Press.
- Di Santo, L. N., & Hamilton, J. A. (2020). Using environmental and geographic data to optimize
- ex situ collections and preserve evolutionary potential. *Conservation Biology*, 35(2), 733–

703 744. doi: 10.1111/cobi.13568

- Di Santo, L. N., Polgar, M., Nies, S., Hodgskiss, P., Canning, C. A., Wright, J. W., & Hamilton,
- J. A. (n.d.). Morphological traits as a tool to quantify variation maintained in ex situ
  collections: a case study in Pinus torreyana (Parry). *AoB Plants (In Review)*.
- 707 Diffenbaugh, N. S., Swain, D. L., & Touma, D. (2015). Anthropogenic warming has increased
- drought risk in California. Proceedings of the National Academy of Sciences, 112(13),
- 709 3931–3936. doi: 10.1073/pnas.1422385112
- Doyle, J. J., & Doyle, J. L. (1987). A rapid DNA isolation procedure for small quantities of fresh
  leaf tissue. *Phytochemical Bulletin*, *19*(1), 11–15.
- 712 Dusek, K. H. (1985). Update on our rarest pine. *American Forests*, *91*, 26–29, 61, 63.
- Eaton, D. A. R., & Overcast, I. (2020). ipyrad: Interactive assembly and analysis of RADseq
- datasets. *Bioinformatics*, *36*(8), 2592–2594. doi: 10.1093/bioinformatics/btz966
- 715 Eckert, A. J., van Heerwaarden, J., Wegrzyn, J. L., Nelson, D. C., Ross-Ibarra, J., González-
- 716 Martínez, S. C., & Neale, D. B. (2010). Patterns of population structure and environmental

- associations to aridity across the range of loblolly pine (Pinus taeda L., Pinaceae). *Genetics*,
- 718 *185*(3), 969–982. doi: 10.1534/genetics.110.115543
- 719 Excoffier, L., Dupanloup, I., Huerta-Sánchez, E., Sousa, V. C., & Foll, M. (2013). Robust
- demographic inference from genomic and SNP data. *PLoS Genetics*, *9*(10), e1003905. doi:
- 721 10.1371/journal.pgen.1003905
- 722 Excoffier, L., & Foll, M. (2011). Fastsimcoal: a continuous-time coalescent simulator of
- genomic diversity under arbitrarily complex evolutionary scenarios. *Bioinformatics*, 27(9),
- 724 1332–1334. doi: 10.1093/bioinformatics/btr124
- Excoffier, L., & Lischer, H. E. L. (2010). Arlequin suite ver 3.5: a new series of programs to
   perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources*, 10(3), 564–567. doi: 10.1111/j.1755-0998.2010.02847.x
- Foll, M., & Gaggiotti, O. (2008). A genome-scan method to identify selected loci appropriate for
  both dominant and codominant markers: a Bayesian perspective. *Genetics*, 180(2), 977–
  993. doi: 10.1534/genetics.108.092221
- Frankham, R., Ballou, J. D., Eldridge, M. D. B., Lacy, R. C., Ralls, K., Dudash, M. R., &
  Fenster, C. B. (2011). Predicting the probability of outbreeding depression. *Conservation Biology*, 25(3), 465–475. doi: 10.1111/j.1523-1739.2011.01662.x
- Frankham, R., Bradshaw, C. J. A., & Brook, B. W. (2014). Genetics in conservation
  management: Revised recommendations for the 50/500 rules, Red List criteria and
  population viability analyses. *Biological Conservation*, *170*, 56–63. doi:
  10.1016/j.biocon.2013.12.036

- Franklin, I. R., & Frankham, R. (1998). How large must populations be to retain evolutionary
  potential? *Animal Conservation*, 1(1), 69–70. doi: 10.1111/j.1469-1795.1998.tb00228.x
- 740 Franklin, J., & Santos, E. V. (2011). A spatially explicit census reveals population structure and
- recruitment patterns for narrowly endemic pine, Pinus torreyana. *Plant Ecology*, 212(2),
- 742 293–306. doi: 10.1007/s11258-010-9822-x
- Fu, L., Niu, B., Zhu, Z., Wu, S., & Li, W. (2012). CD-HIT: accelerated for clustering the nextgeneration sequencing data. *Bioinformatics*, 28(23), 3150–3152. doi:
  10.1093/bioinformatics/bts565
- Gapare, W. J., Yanchuk, A. D., & Aitken, S. N. (2008). Optimal sampling strategies for capture
- of genetic diversity differ between core and peripheral populations of Picea sitchensis
  (Bong.) Carr. *Conservation Genetics*, 9(2), 411–418. doi: 10.1007/s10592-007-9353-8
- Garrison, E., & Marth, G. (2012). Haplotype-based variant detection from short-read sequencing.
   *ArXiv Preprint ArXiv:1207.3907.*
- 751 Goto, S., Iijima, H., Ogawa, H., & Ohya, K. (2011). Outbreeding depression caused by
- intraspecific hybridization between local and nonlocal genotypes in Abies sachalinensis.

753 *Restoration Ecology*, *19*(2), 243–250. doi: 10.1111/j.1526-100X.2009.00568.x

- Goudet, J., & Jombart, T. (2020). hierfstat: Estimation and Tests of Hierarchical F-Statistics.
- 755 Retrieved from https://cran.r-project.org/package=hierfstat
- 756 Grotkopp, E., Rejmánek, M., Sanderson, M. J., & Rost, T. L. (2004). Evolution of genome size
- in pines (Pinus) and its life-history correlates: supertree analyses. *Evolution*, 58(8), 1705–
- 758 1729. doi: 10.1111/j.0014-3820.2004.tb00456.x

- Guerrant Jr, E. O., Havens, K., & Vitt, P. (2014). Sampling for effective ex situ plant
  conservation. *International Journal of Plant Sciences*, *175*(1), 11–20. doi: 10.1086/674131
- Hague, M. T. J., & Routman, E. J. (2016). Does population size affect genetic diversity? A test
- with sympatric lizard species. *Heredity*, *116*(1), 92–98. doi: 10.1038/hdy.2015.76
- Hall, T., & Brinkman, A. (2015). Population Dynamics of the Island Torrey Pine (Pinus torreyana ssp. insularis) on Santa Rosa Island, CA. California State University Channel Islands.
- Haller, J. R. (1986). Taxonomy and Relationships of the Mainland and Island Populations of
  Pinus torreyana (Pinaceae). *Systematic Botany*, *11*(1), 39–50. doi: 10.2307/2418944
- Hamilton, J. A., & Aitken, S. N. (2013). Genetic and morphological structure of a spruce hybrid
  (Picea sitchensis× P. glauca) zone along a climatic gradient. *American Journal of Botany*, *100*(8), 1651–1662. doi: 10.3732/ajb.1200654
- Hamilton, J. A., Royauté, R., Wright, J. W., Hodgskiss, P., & Ledig, T. F. (2017). Genetic
  conservation and management of the California endemic, Torrey pine (Pinus torreyana
  Parry): Implications of genetic rescue in a genetically depauperate species. *Ecology and Evolution*, 7(18), 7370–7381. doi: 10.1002/ece3.3306
- Hedrick, P. W., Peterson, R. O., Vucetich, L. M., Adams, J. R., & Vucetich, J. A. (2014).
- Genetic rescue in Isle Royale wolves: genetic analysis and the collapse of the population.

777 *Conservation Genetics*, *15*(5), 1111–1121. doi: 10.1007/s10592-014-0604-1

Hoban, S. (2019). New guidance for ex situ gene conservation: Sampling realistic population
systems and accounting for collection attrition. *Biological Conservation*, 235, 199–208. doi:

## 780 10.1016/j.biocon.2019.04.013

- Hoban, S., Bruford, M., D'Urban Jackson, J., Lopes-Fernandes, M., Heuertz, M., Hohenlohe, P.
- A., ... Laikre, L. (2020). Genetic diversity targets and indicators in the CBD post-2020
- Global Biodiversity Framework must be improved. *Biological Conservation*, 248, 108654.
- 784 doi: 10.1016/j.biocon.2020.108654
- Hoban, S., & Schlarbaum, S. (2014). Optimal sampling of seeds from plant populations for ex situ conservation of genetic biodiversity, considering realistic population structure.
   *Biological Conservation*, 177, 90–99. doi: 10.1016/j.biocon.2014.06.014
- Holderegger, R., Kamm, U., & Gugerli, F. (2006). Adaptive vs. neutral genetic diversity:
  implications for landscape genetics. *Landscape Ecology*, 21(6), 797–807. doi:
  10.1007/s10980-005-5245-9
- Hufford, K. M., & Mazer, S. J. (2003). Plant ecotypes: genetic differentiation in the age of
  ecological restoration. *Trends in Ecology & Evolution*, 18(3), 147–155. doi:
  10.1016/S0169-5347(03)00002-8
- 794 IUCN. (2021). The IUCN Red List of Threatened Species. Version 2021-1. Retrieved April 19,
  795 2021, from https://www.iucnredlist.org
- Johnson, M., Vander Wall, S. B., & Borchert, M. (2003). A comparative analysis of seed and
- cone characteristics and seed-dispersal strategies of three pines in the subsection
  Sabinianae. *Plant Ecology*, *168*(1), 69–84. doi: 10.1023/A:1024470224134
- Johnson, W. E., Onorato, D. P., Roelke, M. E., Land, D. E., Cunningham, M., Belden, R. C., ...
- 800 Shindle, D. (2010). Genetic restoration of the Florida panther. Science, 329(5999), 1641–

- 801 1645. doi: 10.1126/science.1192891
- Jombart, T. (2008). adegenet: a R package for the multivariate analysis of genetic markers.
   *Bioinformatics*, 24(11), 1403–1405. doi: 10.1093/bioinformatics/btn129
- Jombart, T., & Ahmed, I. (2011). adegenet 1.3-1: new tools for the analysis of genome-wide

805 SNP data. *Bioinformatics*, 27(21), 3070–3071. doi: 10.1093/bioinformatics/btr521

- Laval, G., & Excoffier, L. (2004). SIMCOAL 2.0: a program to simulate genomic diversity over
  large recombining regions in a subdivided population with a complex history. *Bioinformatics*, 20(15), 2485–2487. doi: 10.1093/bioinformatics/bth264
- Ledig, T. F., & Conkle, T. M. (1983). Gene diversity and genetic structure in a narrow endemic,
- 810 Torrey Pine (Pinus torreyana Parry ex Carr.). *Evolution*, *37*(1), 79–85. doi:
  811 10.2307/2408176
- Leinonen, P. H., Remington, D. L., Leppälä, J., & Savolainen, O. (2013). Genetic basis of local
  adaptation and flowering time variation in Arabidopsis lyrata. *Molecular Ecology*, 22(3),
  709–723. doi: 10.1111/j.1365-294X.2012.05678.x
- Leinonen, P. H., Remington, D. L., & Savolainen, O. (2011). Local adaptation, phenotypic
  differentiation, and hybrid fitness in diverged natural populations of Arabidopsis lyrata.
- 817 Evolution: International Journal of Organic Evolution, 65(1), 90–107. doi: 10.1111/j.1558-
- 818 5646.2010.01119.x
- Li, H. (2013). Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *ArXiv Preprint ArXiv:1303.3997.*
- Li, H. (2014). Toward better understanding of artifacts in variant calling from high-coverage

822	samples. <i>Bioinformatics</i>	, 30(20)	, 2843–2851. doi:	10.1093/bioinformatics/btu356
-----	--------------------------------	----------	-------------------	-------------------------------

- Li, S., & Jakobsson, M. (2012). Estimating demographic parameters from large-scale population
  genomic data using Approximate Bayesian Computation. *BMC Genetics*, *13*(1), 1–15. doi:
  10.1186/1471-2156-13-22
- Li, W., & Godzik, A. (2006). Cd-hit: a fast program for clustering and comparing large sets of protein or nucleotide sequences. *Bioinformatics*, 22(13), 1658–1659. doi: 10.1093/bioinformatics/btl158
- Liu, D., Zhang, L., Wang, J., & Ma, Y. (2020). Conservation genomics of a threatened
  Rhododendron: contrasting patterns of population structure revealed from neutral and
  selected SNPs. *Frontiers in Genetics*, *11*, 757. doi: 10.3389/fgene.2020.00757
- Loveless, M. D., & Hamrick, J. L. (1984). Ecological determinants of genetic structure in plant populations. *Annual Review of Ecology and Systematics*, *15*(1), 65–95. doi: 10.1146/annurev.es.15.110184.000433
- Lowry, D. B., Rockwood, C. R., & Willis, J. H. (2008). Ecological reproductive isolation of
- coast and inland races of Mimulus guttatus. *Evolution: International Journal of Organic*

837 *Evolution*, 62(9), 2196–2214. doi: 10.1111/j.1558-5646.2008.00457.x

- Lynch, M., & Lande, R. (1998). The critical effective size for a genetically secure population.
- Animal Conservation, 1(1), 70–72. doi: 10.1111/j.1469-1795.1998.tb00229.x
- Madsen, T., Shine, R., Olsson, M., & Wittzell, H. (1999). Restoration of an inbred adder
  population. *Nature*, 402(6757), 34–35. doi: 10.1038/46941
- McKay, J. K., & Latta, R. G. (2002). Adaptive population divergence: markers, QTL and traits.

843 Trends in Ecology & Evolution, 17(6), 285–291. doi: 10.1016/S0169-5347(02)02478-3

- Menon, M., Bagley, J. C., Friedline, C. J., Whipple, A. V, Schoettle, A. W., Leal Sàenz, A., ...
- 645 Gonzalez Elizondo, S. M. (2018). The role of hybridization during ecological divergence
- of southwestern white pine (Pinus strobiformis) and limber pine (P. flexilis). *Molecular*
- *Ecology*, 27(5), 1245–1260. doi: 10.1111/mec.14505
- Montalvo, A. M., & Ellstrand, N. C. (2001). Nonlocal transplantation and outbreeding
  depression in the subshrub Lotus scoparius (Fabaceae). *American Journal of Botany*, 88(2),
  258–269. doi: 10.2307/2657017
- Namroud, M.-C., Beaulieu, J., Juge, N., Laroche, J., & Bousquet, J. (2008). Scanning the
  genome for gene single nucleotide polymorphisms involved in adaptive population
  differentiation in white spruce. *Molecular Ecology*, *17*(16), 3599–3613. doi:
  10.1111/j.1365-294X.2008.03840.x
- Nei, M. (1987). *Molecular evolutionary genetics*. New York: Columbia University Press.
- Nosil, P., Vines, T. H., & Funk, D. J. (2005). Reproductive isolation caused by natural selection
  against immigrants from divergent habitats. *Evolution*, *59*(4), 705–719. doi: 10.1111/j.00143820.2005.tb01747.x
- 859 Nystedt, B., Street, N. R., Wetterbom, A., Zuccolo, A., Lin, Y.-C., Scofield, D. G., ...
- Alexeyenko, A. (2013). The Norway spruce genome sequence and conifer genome
  evolution. *Nature*, 497(7451), 579–584. doi: 10.1038/nature12211
- Paradis, E. (2010). pegas: an R package for population genetics with an integrated–modular
  approach. *Bioinformatics*, 26(3), 419–420. doi: 10.1093/bioinformatics/btp696

- Parchman, T. L., Gompert, Z., Mudge, J., Schilkey, F. D., Benkman, C. W., & Buerkle, A. C.
- 865 (2012). Genome  $\Box$  wide association genetics of an adaptive trait in lodgepole pine.
- 866 *Molecular Ecology*, 21(12), 2991–3005. doi: 10.1111/j.1365-294X.2012.05513.x
- Peng, R. D. (2019). *simpleboot: Simple Bootstrap Routines*. Retrieved from https://cran.rproject.org/package=simpleboot
- 869 Pesendorfer, M. B., Sillett, S. T., Koenig, W. D., & Morrison, S. A. (2016). Scatter-hoarding
- 870 corvids as seed dispersers for oaks and pines: a review of a widely distributed mutualism
- and its utility to habitat restoration. *The Condor: Ornithological Applications*, 118(2), 215–
- 872 237. doi: 10.1650/CONDOR-15-125.1
- Pew, J., Wang, J., Muir, P., & Frasier, T. (2015). related: an R package for analyzing pairwise
  relatedness data based on codominant molecular markers.
- Potter, K. M., Jetton, R. M., Bower, A., Jacobs, D. F., Man, G., Hipkins, V. D., & Westwood, M.
- (2017). Banking on the future: progress, challenges and opportunities for the genetic
  conservation of forest trees. *New Forests*, *48*, 153–180. doi: 10.1007/s11056-017-9582-8
- Pritchard, D. J., Fa, J. E., Oldfield, S., & Harrop, S. R. (2012). Bring the captive closer to the
  wild: redefining the role of ex situ conservation. *Oryx*, 46(1), 18–23. doi:
  10.1017/S0030605310001766
- Privé, F., Luu, K., Vilhjálmsson, B. J., & Blum, M. G. B. (2020). Performing highly efficient
- genome scans for local adaptation with R package pcadapt version 4. *Molecular Biology*
- *and Evolution*, *37*(7), 2153–2154. doi: 10.1093/molbev/msaa053
- Puritz, J. B., Hollenbeck, C. M., & Gold, J. R. (2014). dDocent: a RADseq, variant-calling

- pipeline designed for population genomics of non-model organisms. *PeerJ*, 2, e431. doi:
  10.7717/peerj.431
- 887 Puritz, J. B., Matz, M. V, Toonen, R. J., Weber, J. N., Bolnick, D. I., & Bird, C. E. (2014).
- 888 Demystifying the RAD fad. *Molecular Ecology*, *23*(24), 5937–5942. doi: 889 10.1111/mec.12965
- R Core Team. (2019). *R: A language and environment for statistical computing*. Vienna: R
  Foundation for Statistical Computing, Austria. Retrieved from https://www.r-project.org/
- 892 R Core Team. (2020). R: A language and environment for statistical computing. Vienna: R

Foundation for Statistical Computing, Austria. Retrieved from https://www.r-project.org/

- Ralls, K., Ballou, J. D., Dudash, M. R., Eldridge, M. D. B., Fenster, C. B., Lacy, R. C., ...
  Frankham, R. (2018). Call for a paradigm shift in the genetic management of fragmented
  populations. *Conservation Letters*, *11*(2), e12412. doi: 10.1111/conl.12412
- Shea, P. J., & Neustein, M. (1995). Protection of a rare stand of Torrey pine from Ips
  paraconfusus. *General Technical Report Intermountain Research Station, USDA Forest Service*, (INT-318), 39–43.
- 900 Soudy, M., Anwar, A. M., Ahmed, E. A., Osama, A., Ezzeldin, S., Mahgoub, S., & Magdeldin,
- 901 S. (2020). UniprotR: retrieving and visualizing protein sequence and functional information
- from Universal Protein Resource (UniProt knowledgebase). Journal of Proteomics, 213,
- 903 103613. doi: 10.1016/j.jprot.2019.103613
- 904 Stevens, K. A., Wegrzyn, J. L., Zimin, A., Puiu, D., Crepeau, M., Cardeno, C., ... Holtz-Morris,
- A. (2016). Sequence of the sugar pine megagenome. *Genetics*, 204(4), 1613–1626. doi:

## 906 10.1093/genetics/204.4.NP

- Swarts, N. D., Sinclair, E. A., Krauss, S. L., & Dixon, K. W. (2009). Genetic diversity in
  fragmented populations of the critically endangered spider orchid Caladenia huegelii:
  implications for conservation. *Conservation Genetics*, 10(5), 1199–1208. doi:
  10.1007/s10592-008-9651-9
- 911 Teixeira, J. C., & Huber, C. D. (2021). The inflated significance of neutral genetic diversity in
  912 conservation genetics. *Proceedings of the National Academy of Sciences*, 118(10),
  913 e2015096118. doi: 10.1073/pnas.2015096118
- Tsumura, Y., Uchiyama, K., Moriguchi, Y., Ueno, S., & Ihara-Ujino, T. (2012). Genome
  scanning for detecting adaptive genes along environmental gradients in the Japanese
  conifer, Cryptomeria japonica. *Heredity*, *109*(6), 349–360. doi: 10.1038/hdy.2012.50
- 917 Tyrmi, J. S., Vuosku, J., Acosta, J. J., Li, Z., Sterck, L., Cervera, M. T., ... Pyhäjärvi, T. (2020).
- Genomics of clinal local adaptation in Pinus sylvestris under continuous environmental and
  spatial genetic setting. *G3: Genes, Genomes, Genetics, 10*(8), 2683–2696. doi:
  10.1534/g3.120.401285
- Varis, S., Pakkanen, A., Galofré, A., & Pulkkinen, P. (2009). The extent of south-north pollen
  transfer in Finnish Scots pine. *Silva Fennica*, 43(5), 717–726. doi: 10.14214/sf.168
- 923 Viana, D. S., Gangoso, L., Bouten, W., & Figuerola, J. (2016). Overseas seed dispersal by
- 924 migratory birds. Proceedings of the Royal Society B: Biological Sciences, 283(1822),
- 925 20152406. doi: 10.1098/rspb.2015.2406
- 926 Wang, X., Bernhardsson, C., & Ingvarsson, P. K. (2020). Demography and natural selection

927 ha	ive shaped	genetic	variation	in the	widely	distributed	conifer	Norwav s	pruce (	Picea	abies)
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- 928 *Genome Biology and Evolution*, *12*(2), 3803–3817. doi: 10.1093/gbe/evaa005
- 929 Waples, R. S., Luikart, G., Faulkner, J. R., & Tallmon, D. A. (2013). Simple life-history traits
- 930 explain key effective population size ratios across diverse taxa. *Proceedings of the Royal*
- 931 Society B: Biological Sciences, 280(1768), 20131339. doi: 10.1098/rspb.2013.1339
- Waters, E. R., & Schaal, B. A. (1991). No variation is detected in the chloroplast genome of
  Pinus torreyana. *Canadian Journal of Forest Research*, 21(12), 1832–1835. doi:
- 934 10.1139/x91-253
- Wegmann, D., Leuenberger, C., & Excoffier, L. (2009). Using ABCtoolbox. Retrieved from
  http://cmpg.unibe.ch/software/ABCtoolbox/
- Wegmann, D., Leuenberger, C., Neuenschwander, S., & Excoffier, L. (2010). ABCtoolbox: a
  versatile toolkit for approximate Bayesian computations. *BMC Bioinformatics*, *11*(1), 1–7.
  doi: 10.1186/1471-2105-11-116
- 940 Westemeier, R. L., Brawn, J. D., Simpson, S. A., Esker, T. L., Jansen, R. W., Walk, J. W., ...
- Paige, K. N. (1998). Tracking the long-term decline and recovery of an isolated population. *Science*, 282(5394), 1695–1698. doi: 10.1126/science.282.5394.1695
- 943 Whitlock, M. C., & Lotterhos, K. (2014). *OutFLANK: Fst outliers with trimming*.
- Whittall, J. B., Syring, J., Parks, M., Buenrostro, J., Dick, C., Liston, A., & Cronn, R. (2010).
- Finding a (pine) needle in a haystack: chloroplast genome sequence divergence in rare
  widespread pines. *Molecular Ecology*, 19(s1), 100–114. doi: 10.1111/j.1365294X.2009.04474.x

- Willi, Y., Van Kleunen, M., Dietrich, S., & Fischer, M. (2007). Genetic rescue persists beyond 948 949 first-generation outbreeding in small populations of a rare plant. Proceedings of the Royal 950 London *B*: **Biological** 2357-2364. Society of Sciences. 274(1623), doi: 951 10.1098/rspb.2007.0768
- 952 Williams, C. G. (2009). *Conifer reproductive biology*. Dordrecht: Springer.
- Williams, C. G. (2010). Long □ distance pine pollen still germinates after meso □ scale dispersal. *American Journal of Botany*, *97*(5), 846–855. doi: 10.3732/ajb.0900255
- Williams, C. G., Barnes, R. D., & Nyoka, I. (1999). Embryonic genetic load for a neotropical
- 956 conifer, Pinus patula Schiede et Deppe. *Journal of Heredity*, 90(3), 394–398. doi:
  957 10.1093/jhered/90.3.394
- Williams, C. G., Zhou, Y., & Hall, S. E. (2001). A chromosomal region promoting outcrossing in
  a conifer. *Genetics*, 159(3), 1283–1289. doi: 10.1093/genetics/159.3.1283
- Xia, H., Wang, B., Zhao, W., Pan, J., Mao, J. F., & Wang, X. R. (2018). Combining
  mitochondrial and nuclear genome analyses to dissect the effects of colonization,
  environment, and geography on population structure in Pinus tabuliformis. *Evolutionary Applications*, 11(10), 1931–1945. doi: 10.1111/eva.12697
- Young, A. G., Boyle, T., & Brown, T. (1996). The population genetic consequences of habitat
  fragmentation for plants. *Trends in Ecology & Evolution*, 11(10), 413–418. doi:
  10.1016/0169-5347(96)10045-8
- Young, A. G., Brown, A. H. D., & Zich, F. A. (1999). Genetic structure of fragmented
  populations of the endangered daisy Rutidosis leptorrhynchoides. *Conservation Biology*,

969 *13*(2), 256–265. doi: 10.1046/j.1523-1739.1999.013002256.x

## 970

## Tables

- 971 **Table 1.** Demographic parameters with their prior distributions and occurrence in each of the six
- 972 models simulated.

Present in models	Parameter	Symbol	Prior distribution	Unit
RPM1, RPM2	Ancestral effective population size	$N_A^{\dagger}$	logU(100:100,000)	ind.
All	Island effective population size	$N_I^{\ddagger}$	U(100:6,000)	ind.
All	Mainland effective population size	$N_M^{\ddagger}$	U(100:6,000)	ind.
CM1, CM2, CM3, CM4	Initial effective population size following colonization	$N_C$ <sup>§</sup>	logU(2:300)	ind.
All	Time of population divergence	$T_{Div}$ ¶	logU(400:50,000)	gen.
RPM2, CM2, CM4	Migration from island to mainland and vice-versa.	$m_{IM}, m_{MI}$	U(0.001:0.01)	-
All	Minor allele frequency	maf <sup>i</sup>	0.01	-

<sup>†</sup> A wide prior range for Torrey pine ancestral effective population size was used to enable the simulation
 of complex demographic histories, including population size expansion, population size reduction or a
 combination of both.

<sup>‡</sup>Census size of reproductively mature trees is of approximately 3,000 to 4,000 individuals in island and

977 mainland population respectively (J. Franklin & Santos, 2011; Hall & Brinkman, 2015). We selected a

978 wide prior range around those estimates to account for the fact that effective and not census population 979 sizes were simulated.

<sup>§</sup> Number of potential effective migrants were chosen based on seed capacity of a Torrey pine cone,
which can hold up to several hundred seeds (unpublished data).

<sup>¶</sup> Time since populations' isolation was estimated between 4,300 and 430,000 years ago (Ledig & Conkle,

1983 1983). Generation time for Torrey pine being approximately 10 years, this translates into a time since
divergence between 430 and 43,000 generations.

985 <sup>\$</sup> Low genome-wide differentiation observed between populations in this study ( $F_{ST} = 0.013$ ) suggest that 986 gene flow may have occurred between populations of Torrey pine. We believe a probability of migration

between 0.001 and 0.01 should appropriately reflect low to moderate gene exchange expected between

geographically distant populations (approximately 280 km) of this wind-pollinated, long-lived, and

primarily outcrossing pine species ( $F_{IS} = -0.127$  and -0.124 for island and mainland population respectively).

<sup>i</sup> Minor allele frequency of 0.01 was chosen to reflect empirical genetic data, filtered for variants with
 frequencies of or above 0.01 (see Material and Methods).

993	<b>Table 2.</b> Average genetic summary statistics across loci, including expected heterozygosity (H <sub>E</sub> ),
994	inbreeding coefficient (F <sub>IS</sub> ), and coancestry (relatedness) coefficient ( $\theta$ ) for island (SRI) and
995	mainland (TPSR) Torrey pine populations. Ninety-five percent confidence interval (95% CI)
996	around mean estimates were obtained by bootstrapping.

Population	H <sub>E</sub> (95% CI)	F <sub>IS</sub> (95% CI)	θ (95% CI)
T 1 1	0.185	-0.127	0.023
Island	(0.184, 0.186)	(-0.128, -0.126)	(0.023, 0.024)
M · 1 1	0.184	-0.124	0.022
Mainland	(0.183, 0.184)	(-0.125, -0.124)	(0.022, 0.023)

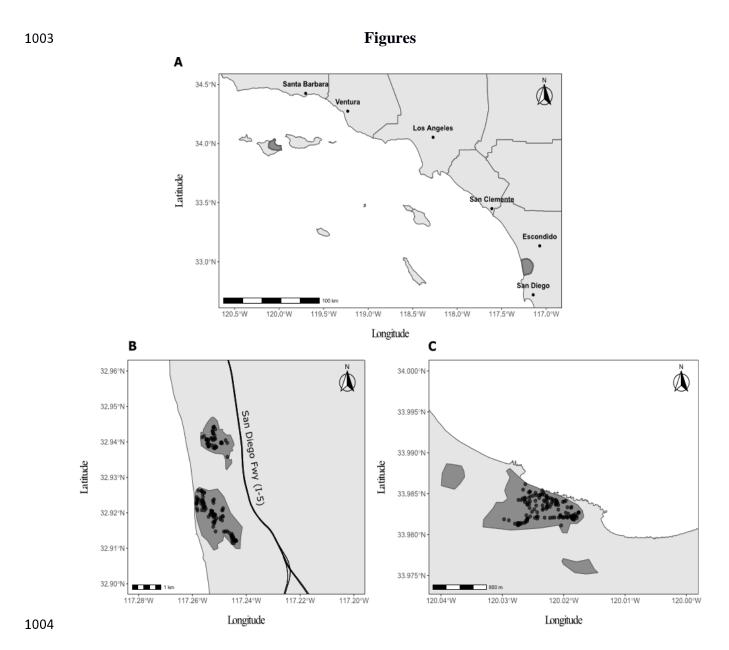
## 998 **Table 3.** Functional categorization of 80 putatively adaptive genes between Torrey pine

999 populations. Listed are the ten most frequent GO terms within each of the three GO classes

1000 (Biological Process, Molecular Function, Cellular Component).

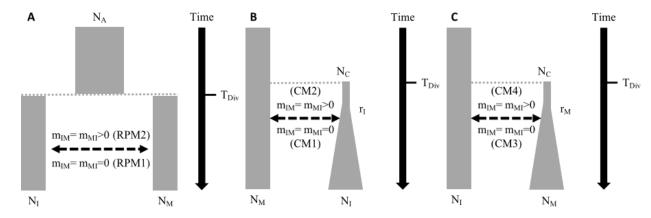
GO class	GO term	Frequency (%)
	DNA integration	6.2
	Methylation	2.5
	Actin filament polymerization	1.2
	Arp2/3 complex-mediated actin nucleation	1.2
Biological Process	Carbohydrate metabolic process	1.2
	Carbohydrate transport	1.2
	Cell differentiation	1.2
	Defense response to bacterium	1.2
	Defense response to fungus	1.2
	Gene silencing by RNA	1.2
	RNA-directed DNA polymerase activity	11.2
	Nucleic acid binding	7.5
	ATP binding	3.8
	ADP binding	2.5
Mala sala a Francisca	Magnesium ion binding	2.5
Molecular Function	O-methyltransferase activity	2.5
	Oxidoreductase activity	2.5
	Protein kinase activity	2.5
	4-lactone oxidase activity	1.2
	Actin binding	1.2
	Mitochondrion	38.8
	Integral component of membrane	7.5
	Arp2/3 protein complex	1.2
	Cell wall	1.2
	Chloroplast	1.2
Cellular Component	Chloroplast thylakoid membrane	1.2
	Cytoplasm	1.2
	Extracellular region	1.2
	Membrane	1.2
	Nucleus	1.2

1001

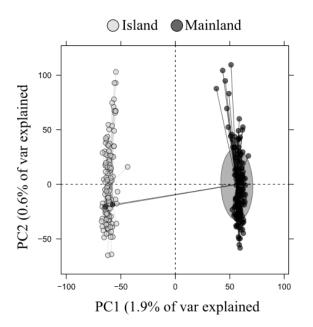


**Figure 1**. (A) Distribution of the last two remanent populations of Torrey pine (dark grey shades). Top left: Santa Rosa Island, Channel Islands, CA. Bottom right: Torrey Pine State Reserve, La Jolla, CA. (B-C) Population-specific distribution of Torrey pine (dark grey shades) and trees sampled for needle tissue (circles) at the Torrey Pine State Reserve (TPSR, B) and on Santa Rosa Island (SRI, C).

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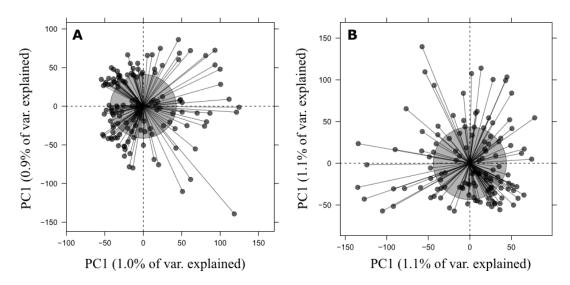


1012 Figure 2. Schematics of demographic scenarios simulated. Rectangles represent current or ancestral populations, dashed arrows represent migration between population, and solid arrows 1013 1014 represent time. (A) Scenarios of isolation without (RPM1) or with (RPM2) migration between 1015 populations. (B) Island colonization scenarios without (CM1) or with (CM2) subsequent 1016 migration between populations. (C) Mainland colonization scenarios without (CM3) or with 1017 (CM4) subsequent migration between populations. N<sub>A</sub>, ancestral effective population size; N<sub>I</sub>, 1018 island effective population size; N<sub>M</sub>, mainland effective population size; N<sub>C</sub> initial effective 1019 population size following migration (number of migrants); m<sub>IM</sub>, migration probability from island to mainland; m<sub>MI</sub>, migration probability from mainland to island; T<sub>Div</sub>, time of population 1020 1021 divergence; r<sub>I</sub>, island (exponential) growth rate; r<sub>M</sub>, mainland (exponential) growth rate 1022



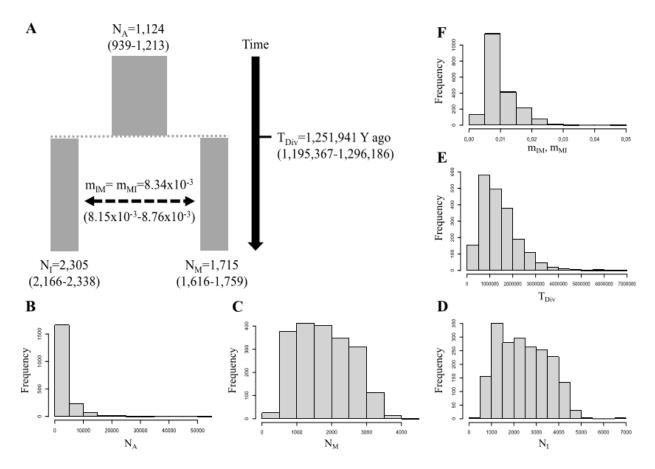
1023

Figure 3. Principal component analysis using 93,085 SNPs for 270 Torrey pine individuals,
including individuals from both mainland (black) and island (grey) populations. Variation
explained by the first two principal components is provided in parentheses.



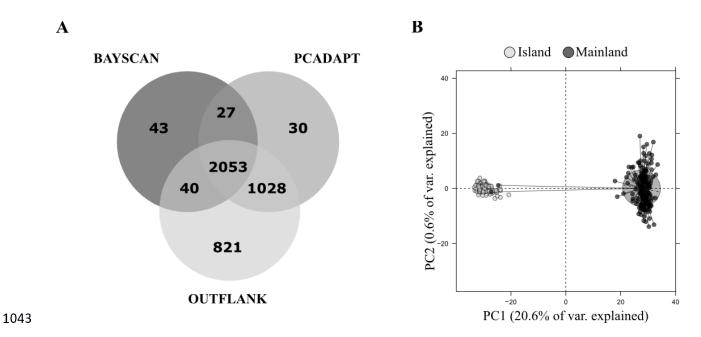
1029Figure 4. Population-specific principal component analysis based on 93,085 SNP variants. (A)1030Mainland population (TPSR, n = 138). Note that two individuals were removed from analysis to1031visualize within-population structure on a finer scale (see Material and Methods). (B) Island1032population (SRI, n = 130).

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Figure 5. (A) Demographic parameters (weighted medians) and 95% confidence intervals (in
parentheses) estimated using RPM2. Each rectangle represents a population, either contemporary
or ancestral. The solid arrow represents time, while the dashed arrow indicates gene flow
between populations. (B-F) Posterior distribution of each demographic parameter inferred using
a tolerance rate of 0.01. N<sub>A</sub>, ancestral effective population size; N<sub>I</sub>, island effective population
size; N<sub>M</sub>, mainland effective population size; m<sub>IM</sub>, migration probability from island to mainland;
m<sub>MI</sub>, migration probability from mainland to island; T<sub>Div</sub>, time of population divergence.



**Figure 6.** (A)Venn diagram showing the number of putatively unique and shared adaptive SNPs detected by BAYESCAN, PCADAPT, and OUTFLANK. (B) Principal component analysis based on all 2,053 shared candidate SNPs for Torrey pine individuals, including individuals from both mainland (black, n = 140) and island (grey, n = 130) populations. Variation explained by the first two principal components is provided in parentheses.