Supporting information for:

The Janus-like role of neuraminidase isoenzymes in inflammation

Md. Amran Howlader^{1†}, Ekaterina P. Demina^{2†}, Suzanne Samarani^{2,4}, Tianlin Guo¹, Ali Ahmad^{2,4}, Alexey V. Pshezhetsky*^{2,3}, Christopher W. Cairo*¹

Affiliations:

- ¹ Department of Chemistry, University of Alberta, Edmonton, Alberta, T6G 2G2, Canada;
- ² Sainte-Justine Hospital Research Center, Division of Medical Genetics, University of Montreal, Montreal, Quebec, H3T 1C5, Canada;
- ³ Department of Anatomy and Cell Biology, McGill University, Montreal, Quebec, H3A0C7, Canada
- ⁴ Department of Microbiology, Infectious Diseases & Immunology, Sainte-Justine Hospital Research Center, University of Montreal, Montreal, Quebec, H3T 1C5, Canada

[‡] Contributed equally

^{*}Correspondence

Table of Contents

Table SI1. Leukocyte subset counts in the air pouch model	3
Table SI2. Leukocyte counts in skin slices from the air pouch model	4
Table SI3. Serum cytokine levels from plasma samples	5
Table SI4. Serum cytokine levels from exudate	6
Table SI5. Transmigration of macrophages isolated from different genotypic mice	7
Table SI6. Leukocyte populations in the air pouch model after inhibitor treatment	7
Figure SI1: Raw cell counts of leukocyte subsets in the air pouch model	8
Figure SI2: Immunohistochemistry of skin slices of the air pouch model	9
Figure SI3. Cytokine levels in mice treated with LPS.	15
Figure SI4: Representative images from transmigration experiments	16
Figure SI5. Leukocyte subsets in the air pouch model after treatment with neuramin	idase
inhibitors	17
Figure SI6. Leukocyte populations in the air pouch model after inhibitor treatment	18

Table SI1. Leukocyte subset counts in the air pouch model.

MONOCYTES	Saline		LPS					
	(cells x10 ⁴)		(cells $x10^4$)					
	Mean ± SEM	N	Mean ± SEM	N				
WT C57BL6	7 ± 2	8	110 ± 24	8				
NEU3 4 DKO	20 ± 6	5	490 ± 94	5				
NEU 4 KO	36 ± 5	4	771 ± 150	4				
NEU 3 KO	34 ± 7	4	91 ± 12	3				
NEU1 KO	59 ± 12	4	35 ± 7	5				

NEUTROPHILS	Saline (cells x10 ⁴)		LPS (cells x10 ⁴)	
NEUTKOFIILS	Mean ± SEM	N	Mean ± SEM	N
WT C57BL6	5 ± 1	8	57 ± 9	8
NEU3 4 DKO	7 ± 2	5	179 ± 47	5
NEU 4 KO	10 ± 1	4	507 ± 111	4
NEU 3 KO	13 ± 5	4	20 ± 2	3
NEU1 KO	28 ± 11	4	10 ± 2	5

MACROPHAGES	Saline (cells x10 ⁴)	_	LPS (cells x10 ⁴)	
	Mean ± SEM	N	Mean ± SEM	N
WT C57BL6	2 ± 1	8	11 ± 3	8
NEU3 4 DKO	3 ± 1	5	20 ± 8	5
NEU 4 KO	3 ± 1	4	13 ± 3	4
NEU 3 KO	4 ± 1	4	8 ± 2	3
NEU1 KO	2 ± 1	4	1 ± 1	5

NK CELLS	Saline (cells x10 ⁴)		LPS (cells x10 ⁴)	104)		
	Mean ± SEM	N	Mean ± SEM	N		
WT C57BL6	9 ± 5	4	38 ± 9	4		
NEU3 4 DKO	5 ± 2	5	70 ± 22	5		
NEU 4 KO	3 ± 1	4	95 ± 6	4		
NEU 3 KO	14 ± 6	4	18 ± 6	4		
NEU1 KO	1 ± 1	2	2 ± 0.1	2		

B Cells	Saline (cells x10 ⁴)		LPS (cells x10 ⁴)	
B Cens	Mean ± SEM	N	Mean ± SEM	N
WT C57BL6	2 ± 1	4	15 ± 7	4
NEU3 4 DKO	4 ± 1	5	94 ± 60	5
NEU 4 KO	4 ± 1	4	3 ± 1	4
NEU 3 KO	2 ± 1	4	6 ± 2	4
NEU1 KO	3 ± 3	2	1 ± 0.1	2

	Saline		LPS	
T CELLS	(cells x10 ⁴)		(cells x10 ⁴)	
	Mean ± SEM	N	Mean ± SEM	N
WT C57BL6	5 ± 4	4	30 ± 20	4
NEU3 4 DKO	5 ± 1	5	154 ± 46	5
NEU 4 KO	5 ± 1	4	34 ± 10	4
NEU 3 KO	12 ± 3	4	34 ± 8	4
NEU1 KO	2 ± 2	2	4 ± 1	2

Results expressed as mean \pm standard error of the mean (SEM).

Table SI2. Leukocyte counts in skin slices from the air pouch model.

Muscle layer	Saline (cells/fi	eld)	LPS (cells/field)					
	Mean ± SEM	N	Mean ± SEM	N				
WT C57BL6	20 ± 1	25	48 ± 3	24				
NEU1 KO	21 ± 1	24	26 ± 2	26				
NEU3 KO	22 ± 1	27	26 ± 2	17				
NEU4 KO	38 ± 6	15	63 ± 5	27				
NEU3/4 DKO	48 ± 3	27	30 ± 2	18				

Dermis layer	Saline (/field)		LPS (/field)	
	Mean ± SEM	N	Mean ± SEM	N
WT C57BL6	31 ± 1	8	48 ± 4	8
NEU1 KO	75 ± 11	6	25 ± 2	7
NEU3 KO	45 ± 5	9	45 ± 2	9
NEU4 KO	43 ± 2	4	74 ± 7	11
NEU3/4 DKO	52 ± 7	8	40 ± 2	5

Table SI3. Serum cytokine levels from plasma samples

			<i>y</i> 10				Plasma	Cytol	kine Pro	file ¹								
	W	Γ Sal	ine	w	T LI	PS	NEU1	KO Saline		NEU:	1 KO	LPS	NEU4 KO Saline			NEU	J4 KC	LPS
	Mean	±	SEM	Mean	±	SEM	Mean	±	SEM	Mean	±	SEM	Mean	±	SEM	Mean	±	SEM
G CSF	5	±	1	672	±	97	28	±	2	95	±	26	11	±	3	330	±	39
IL-1 alpha	13	±	3	41	±	4	49	±	7	18	±	9	9	±	3	33	±	11
IL-1 beta	1	±	0.1	7	±	1	7	±	1	3	±	0.9	2	±	0.8	7	±	2
IL-2	3	±	1	63	±	14	57	±	26	51	±	13	73	±	24	123	±	26
IL-6	19	±	7	557	\pm	92	63	±	5	181	±	80	61	±	9	407	±	23
GM CSF	8	±	2	35	±	5	34	±	4	18	±	5	14	±	4	31	±	9
IL-25 IL-17	10	±	2	181	±	27	163	±	15	65	±	24	61	±	23	152	±	50
IL-10	23	±	5	43	±	4	33	±	7	14	±	2	10	±	4	25	±	6
IL-21	3	±	0.7	21	±	5	29	±	5	12	±	4	5	±	1	33	±	12
IP-10	57	±	3	732	±	111	35	±	3	546	±	75	107	±	10	620	±	55
IL-15/ IL-15R	3	±	0.5	23	±	3	21	±	2	11	±	2	8	±	2	23	±	6
RANTES	16	±	2	924	±	144	54	±	9	414	±	114	13	±	4	362	±	21
MCP-1 CCL2	75	±	0	2473	±	306	67	±	24	1213	±	357	35	±	6	947	±	82
MIP-1 alpha	3	±	1	12	±	0.8	4	±	0.6	5	±	0.6	0.9	±	0.2	8	±	0.5
MIP-1 beta	2	±	0.3	107	±	10	7	±	1	35	±	2	3	±	0.6	51	±	7
MIP-2	4	±	0.2	17	±	2	14	±	2	10	±	2	6	±	1	14	±	2
INF-γ	2	±	0.8	5	±	0.7	4	±	0.7	2	±	0.6	0.4	±	0.2	4	±	1
TNF-α	9	±	3	244	±	20	37	±	5	421	±	60	44	±	10	297	±	37
IL-18	1092	±	166	1970	±	162	943	±	170	1456	±	160	1275	±	28	1356	±	131
IL-33	18	±	2	327	±	81	96	±	29	312	±	68	17	±	3	452	±	178
IL-12p40	37	±	7	49	±	6	42	±	6	167	±	79	24	±	3	46	±	2

^{1.} Results expressed as mean \pm standard error of the mean (SEM); all values are in pg/mL.

Table SI4. Serum cytokine levels from exudate

	Exudate Cytokine Profile 1																	
	W	ΓSal	ine	W	T LI	PS	NEU1	KO	Saline	NEU:	l KO	LPS	NEU4 KO Saline		Saline	NEU4	4 KO	LPS
	Mean	±	SEM	Mean	±	SEM	Mean	±	SEM	Mean	±	SEM	Mean	±	SEM	Mean	±	SEM
G CSF	0.3	±	0.06	79	±	13	0.3	±	0.06	6	±	1	0.3	±	0.08	35	±	2
IL-1 alpha	10	±	1	16	±	1	13	±	1	12	±	1	5	±	1	14	±	1
IL-1 beta	0.6	±	0.2	5	±	0.5	0.2	±	0.06	1	±	0.2	0.2	±	0.03	7	±	0.4
IL-2	5	±	0.8	6	±	0.4	7	±	0.7	6	±	0.8	6	±	2	7	±	1
IL-6	269	±	118	1050	±	213	4	±	1	140	±	48	13	±	3	840	±	139
GM CSF	1	±	0.3	2	±	0.2	2	±	0.5	2	±	0.3	0.6	±	0.1	4	±	0.07
IL-25 IL-17	12	±	2	16	±	3	16	±	2	17	±	5	6	±	1	23	±	3
IL-10	2	±	0.4	15	±	2	3	±	0.3	2	±	0.3	1	±	0.2	11	±	0.9
IL-21	6	±	0.3	3	±	0.6	6	±	0.8	6	±	0.7	5	±	0.5	5	±	0.5
IP-10	10	±	0.4	409	±	62	14	±	2	268	±	29	23	±	5	409	±	53
IL-15/ IL-15R	0.7	±	0.1	3	±	0.3	0.9	±	0.2	1	±	0.1	0.9	±	0.1	3	±	0.3
RANTES	5	±	0.8	190	±	31	6	±	0.6	179	±	25	3	±	0.3	184	±	22
MCP-1 CCL2	196	±	36	4445	±	1054	55	±	8	3383	±	535	70	±	5	1705	±	260
MIP-1 alpha	0.6	±	0.06	16	±	2	1	±	0.2	3	±	0.6	0.7	±	0.1	28	±	2
MIP-1 beta	0.7	±	0.1	57	±	9	0.9	±	0.1	17	±	4	2	±	0.1	83	±	5
MIP-2	16	±	6	21	±	2	7	±	1	8	±	0.4	8	±	0.8	33	±	3
INF-γ	0.08	±	0.03	7	±	0.7	0.1	±	0.03	1	±	0.2	0.05	±	0.02	4	±	0.7
TNF-α	2	±	0.5	16	±	2	3	±	0.3	5	±	0.6	3	±	0.4	15	±	2
IL-18	6	±	0.8	19	±	3	15	±	2	17	±	3	4	±	0.8	25	±	2
IL-33	5	±	1	5	±	1	19	±	5	8	±	1	8	±	2	25	±	345
IL-12p40	45	±	8	208	±	38	27	±	4	154	±	31	23	±	3	167	±	17

^{1.} Results expressed as mean \pm standard error of the mean (SEM); all values are in pg/mL.

Table SI5. Transmigration of macrophages isolated from different genotypic mice.

	(Cells/field)	
	Mean ± SEM	N
WT C57BL6	8 ± 1	32
NEU1 KO	2 ± 1	19
NEU3 KO	17 ± 1	24
NEU4 KO	6 ± 1	32

Table SI6. Leukocyte populations in the air pouch model after inhibitor treatment.

MONOCYTES	Saline		LPS	
	(cells x10 ⁴)		(cells x10 ⁴)	
	Mean ± SEM	N	Mean ± SEM	N
WT C57BL6	7 ± 2	8	75 ± 8	8
IN1	17 ± 4	4	157 ± 48	5
IN3	37 ± 13	4	78 ± 22	4
IN4	25 ± 1	3	182 ± 46	3

NEUTROPHILS	Saline		LPS	
	(cells x10 ⁴)		(cells x10 ⁴)	
	Mean ± SEM	N	Mean ± SEM	N
WT C57BL6	5 ± 1	8	57 ± 9	8
IN1	7 ± 1	4	121 ± 56	4
IN3	26 ± 7	4	42 ± 9	4
IN4	22 ± 7	4	146 ± 41	3

MACROPHAGES	Saline		LPS	
	(cells x10 ⁴)		(cells x10 ⁴)	
	Mean ± SEM	N	Mean ± SEM	N
WT C57BL6	2 ± 1	8	11 ± 3	8
IN1	2 ± 1	5	2 ± 1	5
IN3	2 ± 1	4	1 ± .2	4
IN4	2 ± 1	4	5 ± 2	3

NK CELLS	Saline		LPS	
	(cells x10 ⁴)		(cells x10 ⁴)	
	Mean ± SEM	(N)	Mean ± SEM	(N)
WT C57BL6	4 ± 1	(3)	38 ± 9	(4)
IN1	13 ± 2	(4)	24 ± 8	(4)
IN3	3 ± 1	(4)	7 ± 1	(4)
IN4	30 ± 10	(4)	65 ± 31	(3)

Number of cells found in the air pouch was counted by FACS.

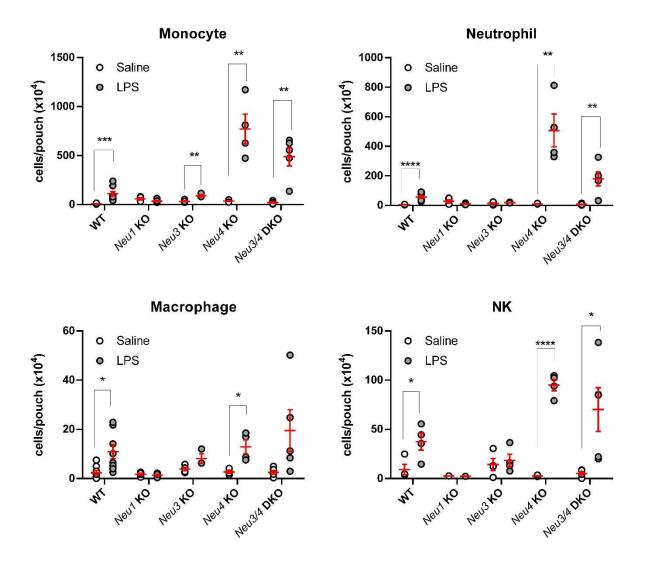


Figure SI1: Raw cell counts of leukocyte subsets in the air pouch model. Leukocytes collected from animals after saline or LPS treatment were counted by FACS and identified based on antibody markers. Cell types shown are Monocytes, Neutrophils, Macrophages, and NK cells. WT, NEU1 KO, NEU3 KO, NEU4 KO and NEU3/4 DKO mice were used in this study. The air pouch exudate was collected after 9 h. The data is presented as mean \pm SEM, and conditions were compared to control using two-way ANOVA followed by Bonferroni multiple comparison test (*, $p \le 0.05$; **, $p \le 0.01$; ***, $p \le 0.005$; ***, $p \le 0.005$; ***, $p \le 0.005$; ****, $p \le 0.0001$).

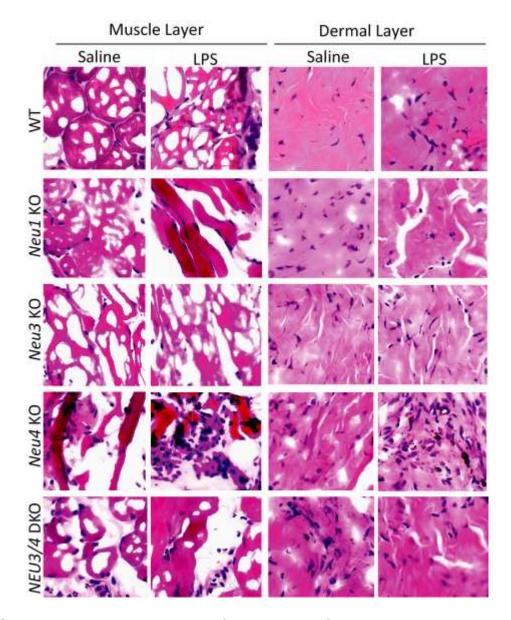
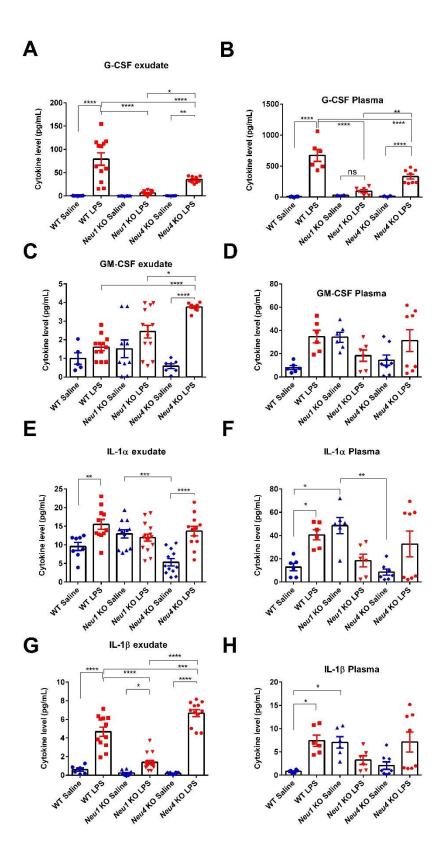
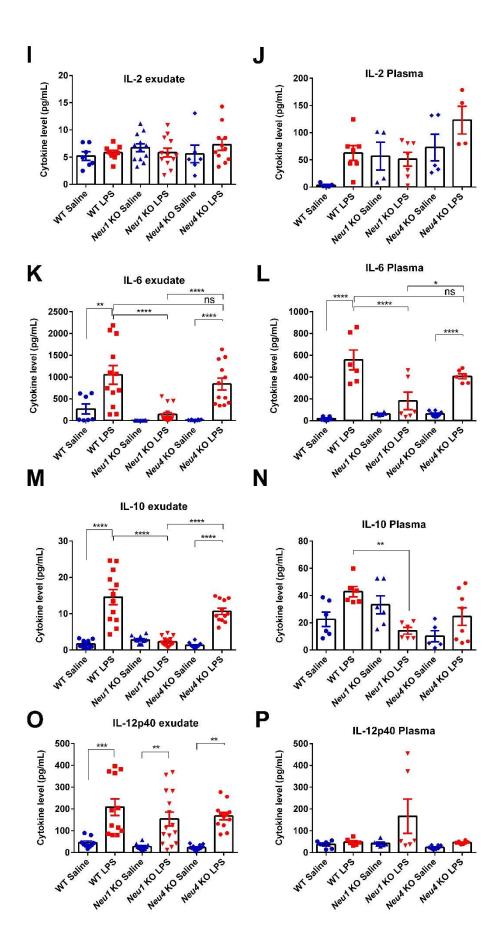
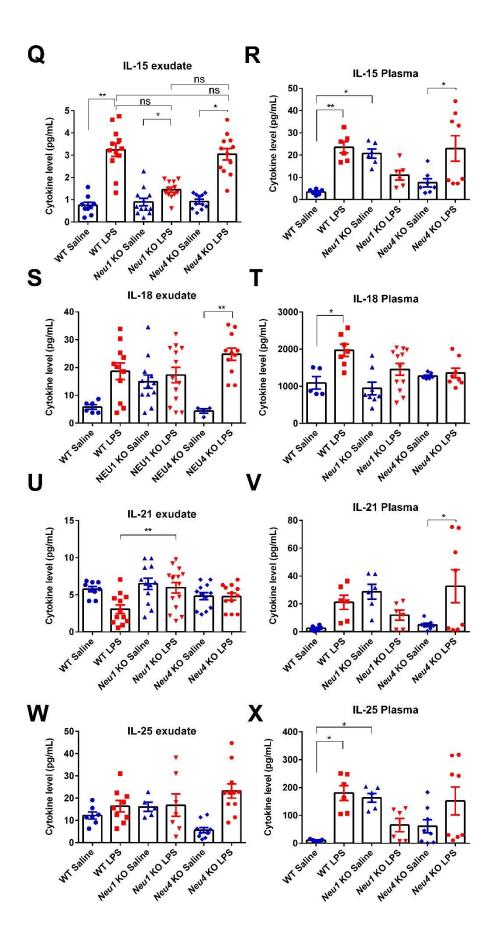
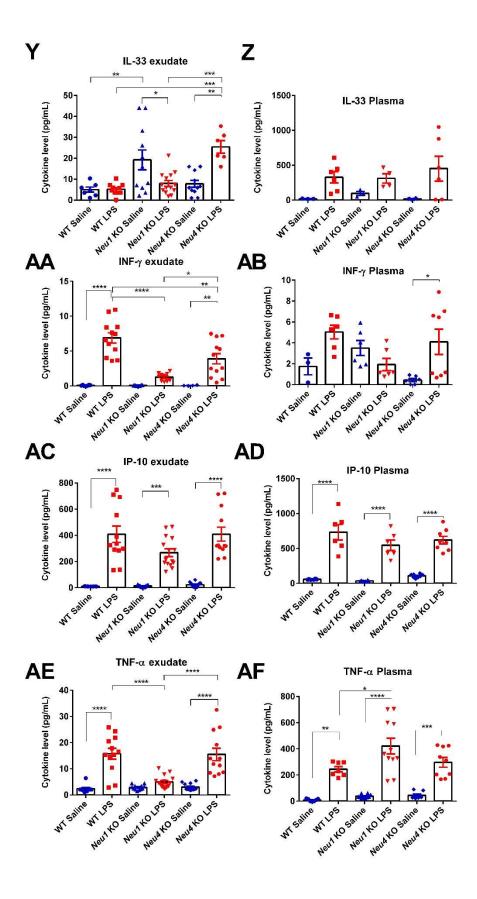


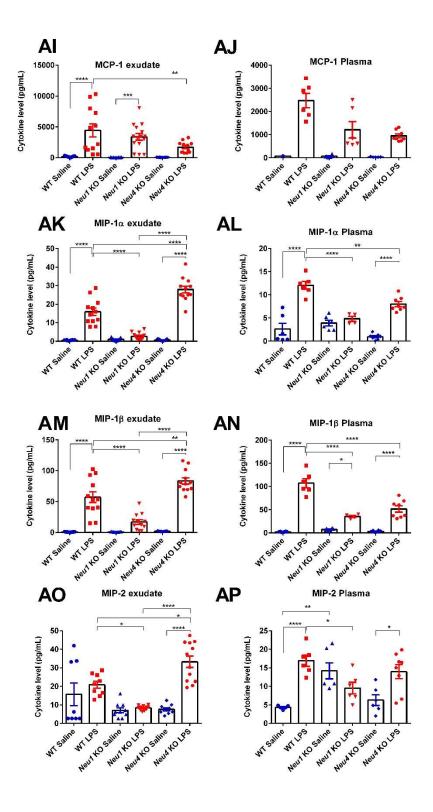
Figure SI2: Immunohistochemistry of skin slices of the air pouch model. Tissue from the air pouch model was collected, sectioned, and stained (H&E). Regions of tissue were identified as dermis or muscle. Representative images of the dermis and muscle regions of different mice groups. Random fields from each region were used to determine leukocyte counts shown in **Figure 3.**











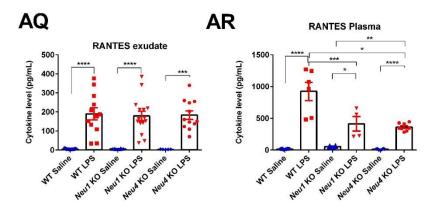


Figure Sl3. Cytokine levels in mice treated with LPS. Cytokines were analyzed using the Autoplex Analyser CS1000 system (Perkin Elmer, Waltham, MA) with a commercial ProcartaPlex Mouse Cytokine Panel Assay kit (Thermo Fisher Scientific Inc., Rockford, USA) in accordance with the manufacturer's instruction. Levels for cytokines from exudate and plasma samples are shown for: (A-B) G-CSF, (C-D) GM-CSF, (E-F) IL-1 α , (G-H) IL-1 β , (I-J) Il-2, (K-L) IL-6, (M-N) IL-10, (O-P) IL-12p40, (Q-R) IL-15, (S-T) IL-18, (U-V) IL-21, (W-X) IL-25, (Y-Z) IL-33, (AA-AB) INF- γ , (AC-AD) IP-10, (AE-AF) TNF- α , (AI-AJ) MCP-1, (AK-AL) MIP-1 α , (AM-AN) MIP-1 β , (AO-AP) MIP-2, and (AQ-AR) RANTES. Bars represent the mean values \pm standard error of the mean (SEM) in units of pg/mL. Samples were compared to controls using a one-way ANOVA followed by Tukey's multiple comparison test (*, p < 0.05; **, p < 0.01; ***, p < 0.001; ****, p < 0.001).

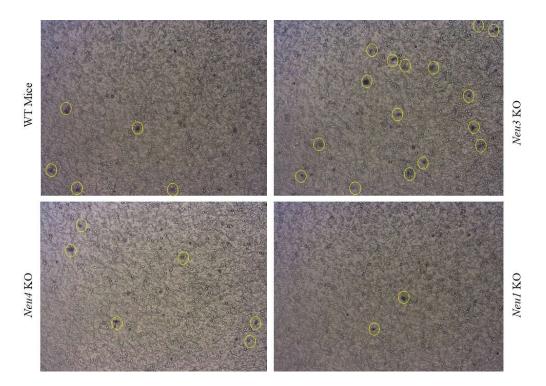


Figure Sl4: Representative images from transmigration experiments. Macrophages were isolated and differentiated from bone marrow of WT, NEU1 KO, NEU3 KO, and NEU4 KO mice. Cells (2.5×10^4) were placed in the upper chamber of the transmigration plate and the experiment was carried out for 5 h. The number of cells that infiltrated the FN-coated membrane was determined by counting of stained cells. The number of cells that infiltrated the FN-coated membrane was determined by counting of stained cells. A representative set of brightfield images is shown in the upper panel using a 10 X objective. Cell counts are shown in **Figure 6.**

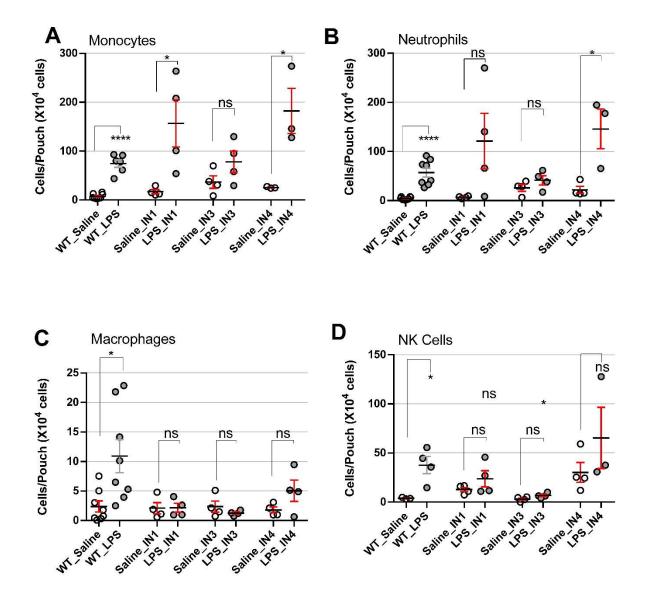


Figure SI5. Leukocyte subsets in the air pouch model after treatment with neuraminidase inhibitors. Leukocytes collected from animals were counted by FACS and identified based on antibody markers (see Methods). Inhibitors that target NEU1 (IN1, CG14600), NEU3 (IN3, CG22600), and NEU4 (IN4, CY16600) were used. Cell types were A. Monocytes, B. Neutrophils, C. Macrophages, and D. NK cells. Animals were WT only and inhibitor treatments were at 1 mg/kg. The air pouch exudate was collected after 9 h. The data is presented as mean \pm SEM, and conditions were compared to control using Student's t-test (*, p \leq 0.05; **, p \leq 0.01; ***, p \leq 0.005; ****, p \leq 0.001).

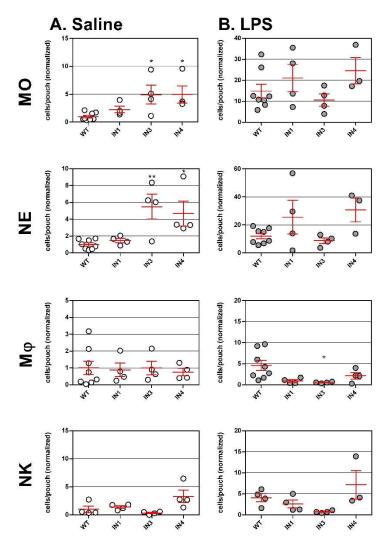


Figure SI6. Leukocyte populations in the air pouch model after inhibitor treatment. Leukocytes from mice treated with either treated with saline or neuramindase inhibitors (IN1, IN3, and IN4). After which animals were either injected with saline (O), or LPS (\bullet) treatment. Collected leukocytes were processed with marker antibodies and were counted by flow cytometry. The cells were identified after staining with marker-specific fluorochrome-conjugated antibodies as monocyte (MO), neutrophil (NE), macrophage (M Θ), or natural killer (NK) cells. The air pouch exudate was collected after 9 h. The data is presented as cell counts (Mean \pm SEM) compared to the respective WT saline controls. Means were compared to control using one-way ANOVA following Dunnet's t-test (*, p \leq 0.05; **, p \leq 0.01; ***, p \leq 0.005; ****, p \leq 0.0001). Raw cell counts are presented in Fig SI5.