Research Article:

Sex differences and estradiol effects in MAPK and Akt cell signalling across subregions of the hippocampus

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Abstract

Introduction: Rapid effects of estrogens within the hippocampus of rodents are dependent upon cell signaling cascades, and activation of these cascades by estrogens varies by sex. Whether these pathways are rapidly activated within the dentate gyrus (DG) and CA1 by estrogens and across the anatomical longitudinal axis has been overlooked.

Methods: Gonadally-intact female and male rats were given either vehicle or physiological systemic low (1.1µg/kg) or high (37.3µg/kg) doses of 17β-estradiol thirty minutes prior to tissue collection. To control for the effects of circulating estrogens, an additional group of female rats was ovariectomized (OVX) and administered 17β-estradiol. Brains were extracted and tissue punches of the CA1 and DG were taken along the longitudinal hippocampal axis (dorsal and ventral) and analyzed for key MAPK and Akt cascade phosphoproteins.

Results: Intact females had higher Akt pathway phosphoproteins (pAkt, pGSK-3β, pp70S6K) than males in the DG (dorsal, ventral) and lower pERK1/2 in the dorsal DG. Most effects of 17β-estradiol on cell signalling occurred in OVX animals. In OVX animals, 17β-estradiol increased cell signalling of MAPK and Akt phosphoproteins (pERK1/2, pJNK, pAkt, pGSK-3β) in the CA1 and pERK1/2 and pJNK DG.

Discussion/Conclusions: Systemic 17β-estradiol treatment rapidly alters phosphoprotein levels in the hippocampus dependent on reproductive status and intact females have greater expression of Akt phosphoproteins than intact males across the hippocampus. These findings shed light on underlying mechanisms of sex differences in hippocampal function and response to interventions that affect MAPK or Akt signaling.

Introduction

Estrogens rapidly affect object and spatial memory consolidation (1–3), short-term social memory (2,4), response learning (5), neurogenesis (6), and spine and synapse formation (7–9). Most of these effects of estrogens occur rapidly and likely via cell signaling pathways after binding with the estrogen receptors (ERs). Estrogens exert both rapid (non-classical) and delayed (classical) actions. The classical actions of estrogens occur when they bind with ERs that dimerize, translocate to the nucleus, and act directly to affect gene expression (10). In contrast, the rapid effects of estrogens initiate a myriad of intracellular processes, including the activation of cell signaling cascades (reviewed in (2–4)). Rapid (within minutes) or delayed (within hours) effects of estrogens on molecular, cellular, and behavior can vary by brain region (reviewed in (2,3,11)). Rapid effects of estrogens on social, object, and spatial location recognition rely on signaling through mitogen-activated protein kinase (MAPK) and Akt (protein kinase B) pathways, whereas the delayed effects are driven predominantly by ER-estrogen response element interactions and effects on gene expression (2,3). There is, however, overlap in these effects, with the less well understood rapid effects occurring concurrently with the more established delayed effects. Thus, understanding the mechanisms, such as cell signaling cascades, through which the rapid effects act is imperative to a comprehensive view of actions by estrogens.

Two of the most studied cell signaling cascades are the MAPK and Akt pathways. These pathways are involved in a number of processes including cell proliferation, migration, and survival, apoptosis, metabolism, differentiation, immune responses, and development (12–15). There is much crosstalk between MAPK and Akt pathways (16,17), and these pathways often act in parallel to elicit similar downstream mechanisms and outputs in complementary or synergistic manners (16,18,19). However, little work has investigated both pathways in the same animals and those that do typically only measure specific phosphoproteins of interest, despite the intricacy and interconnectedness of these signaling cascades.

Sex differences have been noted in the influence of 17β -estradiol on the hippocampus. In older rats, acute 17β -estradiol increases spine density in older females but decreases spine density in males (20). Furthermore, whereas gonadectomy reduces spine density in both males and females in the CA1 (21–23), estradiol increases spine density in females (24,25) but not in males (22). Few studies have investigated sex differences in the effects of 17β -estradiol on cell signalling. Female rats have higher baseline levels of phosphorylated extracellular signal-regulate protein kinase 1/2 (pERK1/2; both p44 [ERK1] and p42 [ERK2] isoforms), pAkt, and phosphorylated glycogen synthase kinase 3 β (pGSK-3 β in the prefrontal cortex (prelimbic and infralimbic cortices), nucleus accumbens, and rostral caudate putamen of rats than male rats (26). However, to our knowledge, no studies have examined sex differences in expression of both MAPK and Akt phosphoproteins in different subregions of the hippocampus thus far. Few studies have explored sex differences in these cell signalling proteins after exposure to 17β -estradiol. Intrahippocampal 17β -estradiol increases dorsal hippocampal pAkt and pERK2 in OVX females, but not in gonadally-intact or castrated males (27). Similarly, 17β -estradiol increases pERK1/2 in anteroventral periventricular (AVPV) nucleus and the medial preoptic area in females but not in males (28). Thus, there is a dearth of evidence exploring influence of 17β -estradiol on cell signalling across the sexes and within subregions across the hippocampus.

The hippocampus is a heterogeneous structure with distinct subfields across the longitudinal axis (dorsal, ventral), as well as dentate gyrus (DG), CA3 and CA1 subregions. Based on gene expression, receptor levels, activity, and connectivity, the dorsal hippocampus is thought to be important for learning and memory, whereas the ventral region is more important for regulation of stress and anxiety (29). Estradiol rapidly increases cell proliferation in both the dorsal and ventral DG (6,30). Furthermore, 17 β -estradiol increases apical dendritic spines in the CA1 region but not in the DG (21,31). Given that 17 β -estradiol influences both dorsal and ventral hippocampal plasticity, it is important to examine possible subregional (i.e. DG and CA1) effects across the longitudinal axis that show rapid 17 β -estradiol action in both sexes.

Most previous studies have examined the effects of estrogens on cell signaling in either the whole hippocampus (32), dorsal hippocampus (27,33–35), or dorsal CA1 (36) in OVX female rodents. Estrogens rapidly increase phosphorylated (activated) proteins in the MAPK and Akt cascades including the MAPKs pERK and pJNK and Akt pathway proteins pAkt, pGSK-3 β , and p70S6K. Another MAPK protein, p38, has divergent effects from other MAPKs and is minimally affected in the hippocampus by the delayed effects of 17 β -estradiol treatment in OVX female rats (37). However, even in studies investigating the rapid effects of 17 β -estradiol on cell signaling cascades, methodologies (including, for example, timing and/or dose of 17 β -estradiol treatment) vary and make comparison between pathways and proteins difficult.

The current study was designed to examine sex differences in cell signaling cascades, with and without 17 β -estradiol, in the dorsal and ventral fields of DG and CA1 of the hippocampus. We hypothesized that sex and 17 β -estradiol would differentially affect MAPK and Akt cell signaling cascades and that these differences would vary by region across the longitudinal axis. Furthermore, we hypothesized that in OVX rats 17 β -estradiol would increase activation of cell signaling cascades in both the CA1 and DG, but that these effects may be more prominent in the dorsal versus ventral axis.

Methods

Animals

Ten- to eleven-week-old, gonadally intact female (n=65) and male (n=19) Sprague-Dawley rats (Charles River, St. Constant, Quebec, Canada) were received at the University of British Columbia and pair-housed (with the exception of one triple-housed cage of males) upon arrival. Rats were housed in transparent polyurethane bins (48×27×20cm) with aspen chip bedding and were given Purina rat chow and tap water *ad libitum*. Rats were maintained under a 12/12h light/dark cycle (lights on 07:00h) in standard housing conditions (21 ± 1°C; 50 ± 10% humidity). Animals were handled for 5 minutes each, every other day prior to the experiment for a minimum of five handling sessions. Animals were 12-13 weeks old when administered 17 β -estradiol and brains were collected. The means and standard errors of the means for body mass were as follows: females 295±3.55g and males 477±7.51g. To determine estrous phase during testing, intact females underwent vaginal lavage immediately following tissue collection. Lavage samples were stained with cresyl violet and estrous phase was evaluated (38). One week following arrival, 36 female rats were bilaterally ovariectomized using aseptic techniques under isoflurane anesthesia, with ketamine (30 mg/kg, Bimeda-MTC, Cambridge, ON), xylazine (2 mg/kg, Bayer HealthCare, Toronto, ON), and bupivacaine (applied locally; 4 mg/kg, Hospira Healthcare

Corporation, Montreal, QC). Following surgery, rats were single-housed and allowed to recover for at least one week before testing. All experiments were conducted in accordance with the ethical guidelines set by the Canada Council for Animal Care and were approved by the University of British Columbia Animal Care Committee. All efforts were made to reduce the number and the suffering of animals.

Treatment

Rats were subcutaneously administered 1.1µg/kg 17β-estradiol in sesame oil (low 17β-estradiol; Females n=10, Males n=6, OVX n=13), 37.3µg/kg 17β-estradiol in sesame oil (high 17β-estradiol; Females n=10, Males n=7, OVX n=13), or sesame oil vehicle (Females n=9, Males n=6, OVX n=10) 30 minutes prior to tissue extraction. These sample sizes were selected as they have previously produced large effect sizes (Cohen's $d > 0.8$) in similar analyses (6,39). Estradiol doses were chosen because they result in circulating levels of 17β-estradiol observed on the morning of proestrus (high 17β-estradiol) or during diestrus (low 17β-estradiol)(40), and both doses enhance cell proliferation 30 minutes after injection in OVX females (6,30). There were no significant differences in average body mass per treatment group (all $p > 0.12$). All rats were injected between 09:00h and 11:15h. Rats were sacrificed by decapitation, and brains were excised, hemisected, and flash-frozen on dry ice.

Brain tissue processing

Brains were sliced into 300µm sections at -10°C using a Leica CM3050 S cryostat (Nußloch, Germany). Punches were taken from the DG and CA1 of both the dorsal and ventral horns of the hippocampus using modified 18G needles with an internal punch diameter of 0.838mm (see Figure 1). Tissue was collected and homogenized using an Omni Bead Ruptor (Omni international, Kennesaw, GA) with 40µl of cold lysis buffer. Homogenates were centrifuged at 4°C and 1000×g for 15min then stored at -20°C. Total protein concentrations in homogenates were quantified using the Pierce Micro BCA Protein Assay Kit (ThermoFisher Scientific) and used according to manufacturer instructions, with samples run in triplicates.

Phosphoprotein quantification

Multiplex electrochemiluminescence immunoassay kits (MAPK Kinase Phosphoprotein and Akt Signaling Panel Assay Whole Cell Lysate Kits) were purchased from Meso Scale Discovery (Rockville, MD) and used according to manufacturer instructions – with the exception of incubating the samples at 4°C overnight – to measure phosphoprotein levels. Electrochemiluminescent assays are highly selective and quantitative, produce low background, have a broad dynamic range of detection, and require relatively small volume of sample compared to ELISAs and Western blots (41,42). The antibody precoated plates allowed for the simultaneous quantification of the following phosphoproteins: pERK1/2 (combined p42 and p44 isoforms), pJNK, and pp38 (MAPK kit) and pAkt, pGSK-3β, and pp70S6K (Akt kit). Samples were run in technical replicates and plates were read with a Sector Imager 2400 (Meso Scale Discovery) using the Discovery Workbench 4.0 (Meso Scale Discovery). Phosphoprotein values were averaged across technical replicates and normalized to total protein concentrations for statistical analyses (as in (39,43–45)). Inter- and intra-assay coefficients of variation (CV) were below 10%. Phosphoprotein signal data were excluded when the CV between duplicates exceeded this threshold.

Statistical analyses

Statistical analyses were run using Statistica v.8.0 (StatSoft Inc, Tulsa, OK) software. Bartlett's homogeneity of variance tests were performed and, in analyses where homogeneity of variance was violated, data were subjected to a Box-Cox transformation ($\lambda=-0.5$). Two-way ANOVAs were conducted for each analyte and region with treatment (oil, low 17 β -estradiol, high 17 β -estradiol) and sex (female, male) as between groups factors. To determine if there were baseline differences between males and females, vehicle treated rats were used as reference for calculation of percent differences. Phosphoprotein levels in intact vehicle treated female rats were compared across the estrous cycle (proestrus vs non-proestrus, corresponding to high and low 17 β -estradiol phases, respectively) using Welch's t-tests. One-way ANOVAs were conducted for each analyte and region in the OVX animals, with treatment (vehicle, low 17 β -estradiol, high 17 β -estradiol) as the between-subjects factor. *Post hoc* comparisons used Newman-Keuls. When one-way ANOVAs violated homogeneity of variance, Brown-Forsythe ANOVAs were performed with Dunnett's T3 *post hoc*s. *A priori* analyses were subjected to Bonferroni correction. Any statistical outliers (± 2 standard deviations from the group mean) were removed. Statistical significance was set at $p<0.05$. Eta squared (η^2) and Cohen's d effect sizes were calculated where appropriate.

Results

First, we analysed the effects of estrous cycle on phosphoproteins across the dorsal/ventral axis and in all regions. There were no significant effects (all $ps>0.245$; Supplementary Figures 1 and 2) and, as such, these were not considered further.

Intact males and females

Akt pathway: Females have higher levels of Akt pathway phosphoproteins in the dorsal and ventral dentate gyrus but not in the CA1 region.

Female rats had up to 400% higher levels of pAkt, pGSK-3 β , and pp70S6K than males in the dorsal and ventral DG, but not in the CA1 region. Females had greater phosphoprotein signal than males for each analyte in the Akt pathway in both the dorsal (109-400%) and ventral DG (123-262%; main effects of sex: dorsal DG: pAkt – $F_{(1,39)}=58.51$, $p<0.0001$, $\eta^2=0.262$; pGSK-3 β – $F_{(1,36)}=37.21$, $p<0.0001$, $\eta^2=0.488$; pp70S6K – $F_{(1,39)}=22.33$, $p<0.0001$, $\eta^2=0.358$; Figure 2A; ventral DG: pAkt – $F_{(1,36)}=51.22$, $p<0.0001$, $\eta^2=0.581$; pGSK-3 β – $F_{(1,35)}=15.67$, $p=0.0004$, $\eta^2=0.292$; pp70S6K – $F_{(1,37)}=11.96$, $p=0.0014$, $\eta^2=0.205$; Figure 2B). There were no other main or interaction effects in this pathway (all $ps>0.094$).

High 17 β -estradiol increased dorsal CA1 pAkt by 36% over vehicle treatment in females ($p=0.05$, Cohen's $d=0.929$), but there were no other significant pairwise comparisons (interaction: $F_{(2,37)}=3.731$, $p=0.0334$, $\eta^2=0.143$; Figure 2C)). Although there were main effects of sex in the dorsal CA1 in the Akt pathway (pAkt – $F_{(1,37)}=7.725$, $p=0.0085$, $\eta^2=0.148$; pGSK-3 β – $F_{(1,38)}=7.608$, $p=0.0089$, $\eta^2=0.143$; pp70S6K – $F_{(1,39)}=5.796$, $p=0.0209$, $\eta^2=0.117$; Figure 2C), *a priori* comparisons failed to show a sex differences in the vehicle conditions (all $ps>0.05$; Figure 2C). There were no other significant effects in the ventral CA1 (all $ps>0.079$; Figure 2D).

MAPK pathway: Males had greater pERK1/2 in the dorsal dentate gyrus than females. 17 β -estradiol reduced pp38 in the ventral DG in intact females but not in intact males.

Male rats had significantly greater pERK1/2 (292%) than females in dorsal DG ($p=0.0004$, Cohen's $d=1.389$; main effect of sex: $F_{(1,37)}=23.87$, $p<0.0001$, $\eta^2=0.372$; Figure 3A). Despite the significant main effects for sex for the other analytes (pJNK – $F_{(1,41)}=5.832$, $p=0.0203$, $\eta^2=0.117$; pp38 – $F_{(1,42)}=38.25$, $p<0.0001$, $\eta^2=0.427$), there were no significant differences in pJNK or pp38 between vehicle treated males and females ($ps>0.05$) or other significant main or interaction effects in the dorsal DG (all $ps>0.257$).

Both doses of 17 β -estradiol decreased pp38 in the ventral DG of females (low 17 β -estradiol: $p<0.0001$, Cohen's $d=2.260$; high 17 β -estradiol: $p<0.0001$, Cohen's $d=3.876$; Figure 3B; interaction: $F_{(2,34)}=8.955$, $p=0.0008$, $\eta^2=0.128$). There were also main effects of sex on pp38 ($F_{(1,34)}=45.13$, $p<0.0001$, $\eta^2=0.323$) and 17 β -estradiol ($F_{(2,34)}=18.54$, $p<0.0001$, $\eta^2=0.266$). Although there was main effect of sex for pERK1/2 ($F_{(1,39)}=6.067$, $p=0.0183$, $\eta^2=0.133$; Figure 3B), the vehicle treated males and females did not significantly differ ($p>0.05$). There were no significant effects on pJNK in the ventral DG (all $ps>0.415$).

There were no significant sex (all $ps>0.145$) or 17 β -estradiol (all $ps>0.134$) effects observed for any MAPK analyte in either the dorsal or ventral CA1 in intact males or females (Figures 3C-D).

Ovariectomized females

Akt pathway: 17 β -Estradiol increased Akt phosphoproteins in the CA1 region of OVX rats.

In ovariectomized rats, 17 β -estradiol had more significant effects on cell signalling phosphoproteins in the hippocampus than in intact rats but these effects were largely restricted to the CA1 region. There were no significant effects of 17 β -estradiol on Akt pathway phosphoproteins in the DG of OVX rats (all $ps>0.270$, Figures 4A-B). In the dorsal CA1, 17 β -estradiol increased pGSK-3 β by ~34-43%, regardless of dose (43% increase with low 17 β -estradiol: $p=0.024$, Cohen's $d=2.37$; 34% increase with high 17 β -estradiol: $p=0.032$, Cohen's $d=1.32$; main effect of 17 β -estradiol: $F_{(2,13)}=4.889$, $p=0.026$, $\eta^2=0.4293$; Figure 4C). However, in the ventral CA1 only the low dose of 17 β -estradiol increased pAkt by 89% ($p=0.042$, Cohen's $d=1.397$; Figure 4D) and pGSK-3 β by 79% ($p=0.008$, Cohen's $d=1.859$; Figure 4D). There were no other 17 β -estradiol effects in the dorsal or ventral CA1 (all $ps>0.205$).

MAPK pathway: 17 β -Estradiol increases MAPK phosphoproteins in the CA1 and DG of OVX rats

17 β -estradiol increased pERK1/2 and pJNK activation in the dorsal DG, (pERK1/2: low 17 β -estradiol – 147% increase, $p=0.031$, Cohen's $d=1.356$; pJNK: low 17 β -estradiol – 57% increase, $p=0.028$, Cohen's $d=1.401$, main effect of 17 β -estradiol: $F^*_{(2, 6.337)}=5.56$, $p=0.0403$; Figure 5A). There were no other significant effects of 17 β -estradiol in the dorsal or ventral DG (all $p>0.303$, Figure 5B).

In the dorsal CA1, both 17 β -estradiol doses increased pJNK by 27% (low 17 β -estradiol: $p=0.017$, Cohen's $d=1.56$; high 17 β -estradiol: $p=0.009$, $d=1.89$; main effect of 17 β -estradiol: $F_{(2,15)}=6.288$, $p=0.0104$, $\eta^2=0.456$; Figure 5C). There were no other significant effects on MAPK phosphoproteins in the dorsal CA1.

In the ventral CA1, low 17 β -estradiol, but not high 17 β -estradiol, increased pERK1/2 by 114% and pJNK by 58% in OVX rats (pERK1/2 – p=0.003, Cohen’s d=2.434, main effect of 17 β -estradiol: $F_{(2,15)}=8.731$, p=0.0031, $\eta^2=0.538$; pJNK – p=0.027, Cohen’s d=1.537, main effect of 17 β -estradiol: $F_{(2,14)}=5.64$, p=0.0160, $\eta^2=0.446$; Figure 5D). There were no other significant effects of 17 β -estradiol of any MAPK phosphoproteins in the ventral CA1 (all ps>0.22).

Discussion

Past characterization of the rapid effects of estrogens on cell signaling cascades has predominantly focused on the CA1 of the dorsal hippocampus or the dorsal hippocampus as a whole and has rarely investigated sex differences. Here, we found sex differences in phosphoprotein levels in the dorsal and ventral DG and dorsal CA1 in MAPK and Akt cascades. 17 β -estradiol had more dramatic effects in OVX females compared to intact females and males on MAPK and Akt pathways. In intact rats, 17 β -estradiol significantly decreased MAPK phosphoproteins in females but a low dose of 17 β -estradiol increased pp38 in males in the ventral DG. Dose-dependent effects of 17 β -estradiol were observed mainly in the CA1 in OVX female mice in both Akt and MAPK pathways. Collectively, these findings are the first characterization of sex differences in MAPK and Akt phosphoprotein levels across the anatomical longitudinal axis of the hippocampus in the DG and CA1. These findings illustrate that both sex and 17 β -estradiol differentially influence phosphoprotein levels in region-specific ways in the hippocampus.

Sex differences in MAPK and Akt cell signalling pathways in the DG.

Consistent with investigations of other brain regions, we observed sex differences in cell signaling pathways in the dorsal and ventral DG, mostly in the Akt pathway (see Table 1 for summary). Females had higher Akt pathway phosphoproteins across the anatomical longitudinal axis and between subregions of the hippocampus than males. On the other hand, males only had higher pERK1/2 than females in the dorsal DG. These findings are partially consistent to past work in other areas of the brain, as female rats had higher baseline levels of pERK1/2, pAkt, and pGSK-3 β in the frontal cortex (prelimbic and infralimbic cortices), nucleus accumbens, and rostral caudate putamen than males (26). Those findings, at least the Akt pathway proteins, mirrors our findings in the hippocampus. In male mice, pERK1/2 expression in the AVPV was higher, irrespective of gonad status, compared to females (28), similar to our findings in the dorsal DG. Regrettably, while much has been made of sex differences in MAPK and Akt pathway proteins in skeletal muscle, heart, and other peripheral tissues (e.g. (46,47)), these are few investigations into sex differences in MAPK or Akt signaling in the brain. Our findings suggest sex differences, favouring females, in the Akt signalling pathways across the dorsal and ventral axis of the DG of the hippocampus and with males showing greater activation of ERK1/2 in the dorsal DG.

It is intriguing that females had higher levels of Akt phosphoproteins than males in the DG, which adds to the literature showing greater Akt phosphoproteins in the brain (26,48) and periphery (46) of females compared to males. The Akt pathway provides a mechanism through which extrinsic factors such as neurotrophic factors can activate transcription factors to affect the regulation of neural stem cells and neurogenesis in the DG (49). Recent evidence suggests that Akt pathway activation can enhance

ischemia-induced neurogenesis and cell migration in males (50) and inactivation can attenuate the exercise-induced neurogenesis in the DG in male mice (51). The fact that the DG was the region that showed sex differences suggests that this may be involved in sex differences in neurogenesis that exist in maturation timelines (52) but also in neurogenic response to androgens (53) and stress exposure (54). Voluntary running increases neurogenesis in the DG of females more so than in males (55,56), suggesting another possible mechanism for this sex difference in Akt phosphoproteins. Collectively these findings suggest that one function of the increased Akt phosphoproteins may be differences in DG neurogenesis in response to certain neurotropic factors in females compared to males. Clearly more work needs to be done to determine the sex specific roles of Akt in the hippocampus.

Understanding the functional significance of these differences is perhaps more important than understanding from whence they arise. Convergent sex differences – those where the underlying cellular mechanisms differ but the behavioural outcome is similar between the sexes – have been observed now in a number of brain regions and behaviours, including spatial tasks, social learning, eye-blink learning, fear behaviors, and synaptic potentiation (57–60). Finally, these data may have implications with regards to latent sex differences; that is, sex differences that only become apparent following environmental or pharmacological interventions and/or aging/disease. For example, females are resistant to the behavioural effects of the neonatal ventral hippocampal lesion model of schizophrenia, potentially resulting from greater MEK-ERK pathway activity in the prefrontal cortex and striatum (26). Perhaps greater Akt pathway phosphoproteins present in the DG of females would make them more resilient to disruptions of this pathway. We suggest that when assessing behaviour and its underlying mechanisms that researchers pay close attention to the potential sex differences in those underlying mechanisms, especially in the case of cell signalling cascades, even when no change in behaviour is present between the sexes.

17 β -estradiol on hippocampal cell signalling pathways in OVX rats

We found more dramatic effects of 17 β -estradiol on cell signalling activation in the hippocampus of OVX rats compared to intact rats (see Table 2 for summary). Estradiol rapidly increased MAPK and Akt pathway phosphoproteins dependent on dose and longitudinal axis within the CA1 region of OVX rats. It is perhaps not surprising that the CA1 region was particularly sensitive to activation of cell signalling proteins in response to 17 β -estradiol as others have found that the dorsal CA1 region shows rapid upregulation of spine density and memory consolidation (27,33,34,36,61–64). Our results are consistent with Koss et al. (2018) who found that whole dorsal hippocampi of OVX female mice, but not gonadally-intact males had increased pAkt and pERK2 (p42 isoform) following acute 17 β -estradiol treatment. Although we did not see statistically significant effects in the dorsal CA1 or ventral DG, the direction of the means favours an increase in pERK1/2 with 17 β -estradiol. It is important to note here that Koss and colleagues (27) saw differences in pERK only in pERK2 and not pERK1 whereas our analysis did not discriminate between isoforms. In addition to the hippocampus, estradiol rapidly increases pERK1/2 in the medial preoptic area and AVPV (28). Further, effects of 17 β -estradiol on pAkt were present only in the ventral CA1 and not in the dorsal CA1, which is interesting given the functional implications of these regions. Future research should consider the anatomical longitudinal axis, as findings can differ based on this and may be important for therapeutic outcomes.

As alluded to above, 17 β -estradiol treatment increased pERK1/2 in the DG and ventral CA1 of OVX rats but not in gonadally intact male and female rats. Consistent with our findings, Koss et al (27) found that 17 β -estradiol increased pERK2 in OVX female mice, and not in intact males. Much of the previous work on the rapid effects of 17 β -estradiol has focused on comparing to an OVX group. However, except for pAkt in the dorsal CA1 and pp38 in the ventral DG, the effects of 17 β -estradiol treatment in this study were only present in OVX rats and not intact rats. It is possible that more effects would be seen at a different (earlier) timepoint than 30 minutes (discussed below). Another possible reason is that the intracellular response to estrogens is already at ceiling in gonadally-intact female rats. While not directly compared, phosphoprotein levels (e.g. Akt analytes in the dorsal DG) in OVX rats were lower than in gonadally-intact females. Further supporting this is the inverted U-shaped dose response in OVX rats in which low 17 β -estradiol increased phosphoproteins whereas high 17 β -estradiol largely did not. This is frequently seen in sex steroid research (9,65) and likely results from higher doses activating off target effects. As such, treatment of gonadally-intact female mice with 17 β -estradiol may have resulted in fewer effects on MAPK and Akt analytes due to the ongoing presence of gonad-derived hormones. This has substantial implications for the conclusions and translatability of preclinical 17 β -estradiol research. For example, if 17 β -estradiol treatment leads to an improvement in a cognitive task in OVX rodents in a MAPK- or Akt-dependent manner, it should not be assumed that the same improvements would be seen in gonadally-intact rodents.

Differences across hippocampal subregions and along the longitudinal axis

Estrogens, in particular 17 β -estradiol, affect neuroplasticity in a subregion-dependant manner in the hippocampus. For example, 17 β -estradiol increases apical dendritic spine density in the dorsal CA1 (21,25,31,61–64) but not in the DG (21,31). However, 17 β -estradiol rapidly increases cell proliferation in the dorsal and ventral DG of OVX rats (6,30), with the effects on males or gonadally-intact females to be determined. Inhibition of ERK signaling blocks the increased dendritic spine densities in dorsal CA1 and medial prefrontal cortex following intrahippocampal 17 β -estradiol treatment (64). Given our findings that low 17 β -estradiol increases pERK, pJNK, and pGSK-3 β in dorsal DG, perhaps the rapid effects of 17 β -estradiol on cell proliferation in the DG (6) involves activation of these phosphoproteins. As such, investigations into the intracellular mechanisms driving rapid increases in cell proliferation in the DG are required.

It remains possible that the other subregions of the hippocampus drive effects seen in previous studies that were absent in ours. Here, we focused on the CA1 and DG as many of the established rapid effects of estrogens occur within these regions (e.g. (6,30,61–64); for recent reviews, see (3,9)). As many previous studies had less regional specificity, perhaps the effects, such as increases in pERK in the dorsal hippocampus response to 17 β -estradiol, are driven by hippocampal regions other than the CA1 and DG, such as the CA3 region.

Estrous cycle

In the present study, we did not observe any statistical effects of estrous cycle, although our study was not designed to specifically examine estrous cycle changes. There is a paucity of research making comparisons across the estrous cycle with regards to cell signaling cascades in the brain. One study found

that transcription of MAPK and, potentially, Akt cascade phosphoprotein mRNA in the medial prefrontal cortex of rats was upregulated in proestrus (66). In the ventral hippocampus, chromatin dynamics and resulting MAPK pathway protein gene expression vary across the estrous cycle and between male and female mice (67). In addition to changes in dendritic spine densities in the rat hippocampus across the estrous cycle (68,69), changes in total hippocampal volume across the mouse estrous cycle have been observed, with high estrogen phases associated with greater hippocampal volume (70). Similar changes have been found across the human menstrual cycle (71,72). With so many neuroplastic changes occurring with natural fluctuations of estrogens, further investigations into the effects of the estrous cycle on estrogen-responsive kinase pathways are needed.

Limitations and Future Directions

One limitation is that we only examined a 30 min window after 17 β -estradiol exposure. We selected a 30 min delay in the present study as others have noted changes to hippocampal physiology and behaviour have been observed at this timepoint (6,30,64,73) following 17 β -estradiol or shortly thereafter (e.g 40 min (61–63)). However, other studies (using intrahippocampal infusion) have found increases in many MAPK and Akt phosphoproteins in the hippocampus as early as 5–10 min following 17 β -estradiol (33–36). Due to the transient nature of phosphorylation in cell signalling cascades, it remains possible that the effects of treatment occurred at an earlier timepoint than in our study. Furthermore, it is possible that both sex and/or gonadal status may affect the rate at which these rapid effects of 17 β -estradiol occur and should be investigated further.

The present data were normalized to total protein in sample (as is done in regularly in the literature: (39,43–45), for example). One possible limitation is that phosphorylated proteins were not normalised to the specific protein under investigation (e.g. pERK1/2 normalized to total ERK1/2). It remains possible that there are sex differences in and estradiol effects on total protein levels that could confound the interpretation of the present results. Genomic synthesis of novel MAPK, Akt, or other proteins triggered by 17 β -estradiol is unlikely due to the time of tissue collection (30 min). It remains possible, however, that 17 β -estradiol could rapidly increase the total protein levels via non-genomic mechanisms, such as the translation of mRNA transcripts already in the cell. Additionally, it is through the phosphorylation of these proteins that functional outcomes occur (e.g. DNA transcription, protein translation, actin polymerization, etc.), thus obtaining the functional unit of these pathways.

The electrochemiluminescent assay is highly sensitive and selective, provides quantitative data for analyses, and requires relatively little tissue for analysis compared to similar assays (41,42). Other cell signaling cascades or proteins, such as the eukaryotic translation initiation factor 4E-binding protein 1, a translation repressor protein downstream of many Akt and MAPK phosphoproteins including Akt, ERK1/2, and p38 (74) would be important proteins to examine in the future. Estradiol only had significant effects on phosphoprotein level in the ventral DG where pp38 was decreased in female rats only. This reduction could result from inactivation by upstream kinases or as a compensatory mechanism following transient activation (75), which would be important to examine in the future. In addition, in the present study we focussed on the CA1 and DG as they are rapidly affected by estrogens (9). However, the CA3 is an additional region of interest, given sex and estradiol differences in morphology and response to stress in this region (20,68,76).

Conclusions

Sex differences in MAPK and Akt cell signaling phosphoproteins exist in both the dorsal and ventral DG in rats. Estradiol treatment increased MAPK and Akt signaling in the CA1 and dorsal DG in OVX rats with far fewer effects in intact females or males. These results shed light on the possible underlying mechanisms of sex differences in hippocampal function in response to 17 β -estradiol that affect MAPK and Akt kinase pathways. They further show the heterogeneity of estrogen and sex effects between hippocampal subregions and along the dorsal/ventral axis. With these subregions serving often distinct purposes, and the functional dissociation between the dorsal and ventral hippocampus, the present results provide possible mechanisms through which these differences may arise. Furthermore, with many neurological disorders having profound effects on the hippocampus, these sex, hormonal status, and regional differences provide valuable insights not only into the potential underlying mechanisms, but also into therapeutic possibilities.

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Statement of Ethics

All experiments were conducted in accordance with the ethical guidelines set by the Canada Council for Animal Care and were approved by the University of British Columbia Animal Care Committee. All efforts were made to reduce the number and the suffering of animals.

Conflicts of Interest

The authors have no conflicts of interest to declare.

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Author Contributions

PASS – conception and design of the work; data acquisition, analysis, and interpretation; writing and revising

TAP - conception and design of the work; data acquisition, analysis, and interpretation; writing and revising

LAMG - conception and design of the work; data acquisition, analysis, and interpretation; writing and revising

Data Availability

All data is available upon request to corresponding author.

- 473 1. Frick KM. Molecular mechanisms underlying the memory-enhancing effects of estradiol. *Horm*
474 *Behav.* 2015 Aug;74:4–18.
- 475 2. Sheppard PAS, Koss WA, Frick KM, Choleris E. Rapid actions of oestrogens and their receptors on
476 memory acquisition and consolidation in females. *J Neuroendocrinol.* 2018 Feb;30(2):e12485.
- 477 3. Taxier LR, Gross KS, Frick KM. Oestradiol as a neuromodulator of learning and memory. *Nat Rev*
478 *Neurosci.* 2020 Oct;21(10):535–50.
- 479 4. Paletta P, Sheppard PAS, Matta R, Ervin KSJ, Choleris E. Rapid effects of estrogens on short-term
480 memory: Possible mechanisms. *Horm Behav.* 2018 Aug;104:88–99.
- 481 5. Zurkovsky L, Serio SJ, Korol DL. Intra-striatal estradiol in female rats impairs response learning
482 within two hours of treatment. *Horm Behav.* 2011 Nov;60(5):470–7.
- 483 6. Barha CK, Lieblich SE, Galea L a. M. Different Forms of Oestrogen Rapidly Upregulate Cell
484 Proliferation in the Dentate Gyrus of Adult Female Rats. *J Neuroendocrinol.* 2009;21(3):155–66.
- 485 7. Sellers K, Raval P, Srivastava DP. Molecular signature of rapid estrogen regulation of synaptic
486 connectivity and cognition. *Front Neuroendocrinol.* 2015 Jan 1;36:72–89.
- 487 8. Sellers KJ, Erli F, Raval P, Watson IA, Chen D, Srivastava DP. Rapid modulation of synaptogenesis
488 and spinogenesis by 17 β -estradiol in primary cortical neurons. *Front Cell Neurosci.* 2015;9:137.
- 489 9. Sheppard PAS, Choleris E, Galea LAM. Structural plasticity of the hippocampus in response to
490 estrogens in female rodents. *Mol Brain.* 2019 Mar 18;12(1):22.
- 491 10. Nilsson S, Mäkelä S, Treuter E, Tujague M, Thomsen J, Andersson G, et al. Mechanisms of
492 estrogen action. *Physiol Rev.* 2001 Oct;81(4):1535–65.
- 493 11. Lai Y-J, Yu D, Zhang JH, Chen G-J. Cooperation of genomic and rapid nongenomic actions of
494 estrogens in synaptic plasticity. *Mol Neurobiol.* 2017;54(6):4113–26.
- 495 12. Marshall CJ. MAP kinase kinase kinase, MAP kinase kinase and MAP kinase. *Curr Opin Genet Dev.*
496 1994 Feb;4(1):82–9.
- 497 13. Seger R, Krebs EG. The MAPK signaling cascade. *FASEB J.* 1995 Jun;9(9):726–35.
- 498 14. Manning BD, Cantley LC. AKT/PKB signaling: Navigating downstream. *Cell.* 2007 Jun
499 29;129(7):1261–74.
- 500 15. Dorard C, Vucak G, Baccarini M. Deciphering the RAS/ERK pathway in vivo. *Biochem Soc Trans.*
501 2017 Feb 8;45(1):27–36.
- 502 16. Cao Z, Liao Q, Su M, Huang K, Jin J, Cao D. AKT and ERK dual inhibitors: The way forward? *Cancer*
503 *Lett.* 2019 Sep 10;459:30–40.

- 504 17. Titolo D, Mayer CM, Dhillon SS, Cai F, Belsham DD. Estrogen facilitates both phosphatidylinositol
505 3-kinase/Akt and ERK1/2 mitogen-activated protein kinase membrane signaling required for
506 long-term neuropeptide Y transcriptional regulation in clonal, immortalized neurons. *J Neurosci*.
507 2008 Jun 18;28(25):6473–82.
- 508 18. Winter JN, Jefferson LS, Kimball SR. ERK and Akt signaling pathways function through parallel
509 mechanisms to promote mTORC1 signaling. *Am J Physiol - Cell Physiol*. 2011 May;300(5):C1172–
510 80.
- 511 19. Zhou J, Du T, Li B, Rong Y, Verkhatsky A, Peng L. Crosstalk between MAPK/ERK and PI3K/AKT
512 signal pathways during brain ischemia/reperfusion. *ASN NEURO*. 2015 Oct 6;7(5):
513 1759091415602463.
- 514 20. Miranda P, Williams CL, Einstein G. Granule cells in aging rats are sexually dimorphic in their
515 response to estradiol. *J Neurosci*. 1999 May 1;19(9):3316–25.
- 516 21. Gould E, Woolley CS, Frankfurt M, McEwen BS. Gonadal steroids regulate dendritic spine density
517 in hippocampal pyramidal cells in adulthood. *J Neurosci*. 1990 Apr;10(4):1286–91.
- 518 22. Leranthy C, Petnehazy O, MacLusky NJ. Gonadal hormones affect spine synaptic density in the
519 CA1 hippocampal subfield of male rats. *J Neurosci*. 2003 Mar 1;23(5):1588–92.
- 520 23. Wallace M, Luine V, Arellanos A, Frankfurt M. Ovariectomized rats show decreased recognition
521 memory and spine density in the hippocampus and prefrontal cortex. *Brain Res*. 2006 Dec
522 18;1126(1):176–82.
- 523 24. MacLusky NJ, Luine VN, Hajszan T, Leranthy C. The 17alpha and 17beta isomers of estradiol both
524 induce rapid spine synapse formation in the CA1 hippocampal subfield of ovariectomized female
525 rats. *Endocrinology*. 2005 Jan;146(1):287–93.
- 526 25. Woolley CS, McEwen BS. Roles of estradiol and progesterone in regulation of hippocampal
527 dendritic spine density during the estrous cycle in the rat. *J Comp Neurol*. 1993 Oct
528 8;336(2):293–306.
- 529 26. Bychkov E, Ahmed MR, Gurevich EV. Sex differences in the activity of signalling pathways and
530 expression of G-protein-coupled receptor kinases in the neonatal ventral hippocampal lesion
531 model of schizophrenia. *Int J Neuropsychopharmacol*. 2011 Feb;14(1):1–15.
- 532 27. Koss WA, Haertel JM, Philippi SM, Frick KM. Sex differences in the rapid cell signaling
533 mechanisms underlying the memory-enhancing effects of 17β-estradiol. *eNeuro*. 2018 Oct
534 16;5(5):e0267-18.2018.
- 535 28. Barabás K, Szegő ÉM, Kaszás A, Nagy GM, Juhász GD, Ábrahám IM. Sex Differences in oestrogen-
536 induced p44/42 MAPK phosphorylation in the mouse brain in vivo. *J Neuroendocrinol*. 2006 Aug
537 1;18(8):621–8.
- 538 29. Fanselow MS, Dong H-W. Are the dorsal and ventral hippocampus functionally distinct
539 structures? *Neuron*. 2010 Jan 14;65(1):7–19.

- 540 30. Duarte-Guterman P, Lieblich SE, Chow C, Galea LAM. Estradiol and GPER activation differentially
541 affect cell proliferation but not GPER expression in the hippocampus of adult female rats. PLoS
542 One. 2015;10(6):e0129880.
- 543 31. Jacome LF, Barateli K, Buitrago D, Lema F, Frankfurt M, Luine VN. Gonadal hormones rapidly
544 enhance spatial memory and increase hippocampal spine density in male rats. Endocrinology.
545 2016 Apr 1;157(4):1357–62.
- 546 32. Cardona-Gomez P, Perez M, Avila J, Garcia-Segura LM, Wandosell F. Estradiol inhibits GSK3 and
547 regulates interaction of estrogen receptors, GSK3, and beta-catenin in the hippocampus. Mol
548 Cell Neurosci. 2004 Mar 1;25(3):363–73.
- 549 33. Fan L, Zhao Z, Orr PT, Chambers CH, Lewis MC, Frick KM. Estradiol-induced object memory
550 consolidation in middle-aged female mice requires dorsal hippocampal extracellular signal-
551 regulated kinase and phosphatidylinositol 3-kinase activation. J Neurosci. 2010 Mar
552 24;30(12):4390–400.
- 553 34. Fortress AM, Fan L, Orr PT, Zhao Z, Frick KM. Estradiol-induced object recognition memory
554 consolidation is dependent on activation of mTOR signaling in the dorsal hippocampus. Learn
555 Mem. 2013 Feb 19;20(3):147–55.
- 556 35. Kim J, Szinte JS, Boulware MI, Frick KM. 17 β -estradiol and agonism of G-protein-coupled
557 estrogen receptor enhance hippocampal memory via different cell-signaling mechanisms. J
558 Neurosci. 2016 Mar 16;36(11):3309–21.
- 559 36. Fernandez SM, Lewis MC, Pechenino AS, Harburger LL, Orr PT, Gresack JE, et al. Estradiol-
560 induced enhancement of object memory consolidation involves hippocampal extracellular
561 signal-regulated kinase activation and membrane-bound estrogen receptors. J Neurosci. 2008
562 Aug 27;28(35):8660–7.
- 563 37. Pinceti E, Shults CL, Rao YS, Pak TR. Differential effects of E2 on MAPK activity in the brain and
564 heart of aged female rats. PLoS ONE. 2016 Aug 3;11(8):e0160276.
- 565 38. Cora MC, Kooistra L, Travlos G. Vaginal cytology of the laboratory rat and mouse: Review and
566 criteria for the staging of the estrous cycle using stained vaginal smears. Toxicol Pathol. 2015 Aug
567 1;43(6):776–93.
- 568 39. Eid RS, Lieblich SE, Wong SJ, Galea LAM. Ovarian status dictates the neuroinflammatory and
569 behavioral consequences of sub-chronic stress exposure in middle-aged female mice. Neurobiol
570 Stress. 2019 Nov 20;12:100199.
- 571 40. Viau V, Meaney MJ. Variations in the hypothalamic-pituitary-adrenal response to stress during
572 the estrous cycle in the rat. Endocrinology. 1991 Nov;129(5):2503–11.
- 573 41. Chen M, Ning Z, Chen K, Zhang Y, Shen Y. Recent advances of electrochemiluminescent system in
574 bioassay. J Anal Test. 2020 Apr 1;4(2):57–75.

- 575 42. Zanut A, Fiorani A, Canola S, Saito T, Ziebart N, Rapino S, et al. Insights into the mechanism of
576 coreactant electrochemiluminescence facilitating enhanced bioanalytical performance. *Nat*
577 *Commun.* 2020 May 29;11(1):2668.
- 578 43. Zaworski P, Herrmann KM von, Taylor S, Sunshine SS, McCarthy K, Risher N, et al. SMN protein
579 can be reliably measured in whole blood with an electrochemiluminescence (ECL) immunoassay:
580 Implications for clinical trials. *PLoS ONE.* 2016 Mar 8;11(3):e0150640.
- 581 44. Bass V, Gordon CJ, Jarema KA, MacPhail RC, Cascio WE, Phillips PM, et al. Ozone induces glucose
582 intolerance and systemic metabolic effects in young and aged brown Norway rats. *Toxicol Appl*
583 *Pharmacol.* 2013 Dec 15;273(3):551–60.
- 584 45. Aldridge GM, Podrebarac DM, Greenough WT, Weiler IJ. The use of total protein stains as
585 loading controls: an alternative to high-abundance single protein controls in semi-quantitative
586 immunoblotting. *J Neurosci Methods.* 2008 Jul 30;172(2):250–4.
- 587 46. Camper-Kirby Dreama, Welch Sara, Walker Angela, Shiraishi Isao, Setchell Kenneth D. R.,
588 Schaefer Erik, et al. Myocardial Akt activation and gender. *Circ Res.* 2001 May 25;88(10):1020–7.
- 589 47. Nicoll JX, Fry AC, Mosier EM. Sex-based differences in resting MAPK, androgen, and
590 glucocorticoid receptor phosphorylation in human skeletal muscle. *Steroids.* 2019 Jan;141:23–9.
- 591 48. Wong H, Levenga J, LaPlante L, Keller B, Cooper-Sansone A, Borski C, et al. Isoform-specific roles
592 for AKT in affective behavior, spatial memory, and extinction related to psychiatric disorders.
593 *eLife.* 2020 Dec 16;9:e56630.
- 594 49. Otaegi G, Yusta-Boyo MJ, Vergaño-Vera E, Méndez-Gómez HR, Carrera AC, Abad JL, et al.
595 Modulation of the PI 3-kinase–Akt signalling pathway by IGF-I and PTEN regulates the
596 differentiation of neural stem/precursor cells. *J Cell Sci.* 2006 Jul 1;119(13):2739–48.
- 597 50. Shioda N, Han F, Morioka M, Fukunaga K. Bis(1-oxy-2-pyridinethiolato)oxovanadium(IV)
598 enhances neurogenesis via phosphatidylinositol 3-kinase/Akt and extracellular signal regulated
599 kinase activation in the hippocampal subgranular zone after mouse focal cerebral ischemia.
600 *Neuroscience.* 2008 Aug 26;155(3):876–87.
- 601 51. Sun L, Cui K, Xing F, Liu X. Akt dependent adult hippocampal neurogenesis regulates the
602 behavioral improvement of treadmill running to mice model of post-traumatic stress disorder.
603 *Behav Brain Res.* 2020 Feb 3;379:112375.
- 604 52. Yagi S, Splinter JEJ, Tai D, Wong S, Wen Y, Galea LAM. Sex differences in maturation and attrition
605 of adult neurogenesis in the hippocampus. *eNeuro.* 2020 Jul-Aug;7(4):e0468-19.2020
- 606 53. Duarte-Guterman P, Lieblich SE, Wainwright SR, Chow C, Chaiton JA, Watson NV, et al.
607 Androgens enhance adult hippocampal neurogenesis in males but not females in an age-
608 dependent manner. *Endocrinology.* 2019 Sep 1;160(9):2128–36.
- 609 54. Falconer EM, Galea LAM. Sex differences in cell proliferation, cell death and defensive behavior
610 following acute predator odor stress in adult rats. *Brain Res.* 2003 Jun 13;975(1–2):22–36.

- 611 55. Cahill SP, Cole JD, Yu RQ, Clemans-Gibbon J, Snyder JS. Differential effects of extended exercise
612 and memantine treatment on adult neurogenesis in male and female rats. *Neuroscience*. 2018
613 Oct 15;390:241–55.
- 614 56. Ma X, Hamadeh MJ, Christie BR, Foster JA, Tarnopolsky MA. Impact of treadmill running and sex
615 on hippocampal neurogenesis in the mouse model of amyotrophic lateral sclerosis. *PLoS ONE*.
616 2012 Apr 25;7(4):e36048.
- 617 57. Shors TJ, Chua C, Falduto J. Sex differences and opposite effects of stress on dendritic spine
618 density in the male versus female hippocampus. *J Neurosci*. 2001 Aug 15;21(16):6292–7.
- 619 58. Choleris E, Galea LAM, Sohrabji F, Frick KM. Sex differences in the brain: Implications for
620 behavioral and biomedical research. *Neurosci Biobehav Rev*. 2018;85:126–45.
- 621 59. Jain A, Huang GZ, Woolley CS. Latent sex differences in molecular signaling that underlies
622 excitatory synaptic potentiation in the hippocampus. *J Neurosci*. 2019 Feb 27;39(9):1552–65.
- 623 60. Gruene TM, Roberts E, Thomas V, Ronzio A, Shansky RM. Sex-specific neuroanatomical
624 correlates of fear expression in prefrontal-amygdala circuits. *Biol Psychiatry*. 2015 Aug
625 1;78(3):186–93.
- 626 61. Phan A, Suschkov S, Molinaro L, Reynolds K, Lymer JM, Bailey CDC, et al. Rapid increases in
627 immature synapses parallel estrogen-induced hippocampal learning enhancements. *Proc Natl*
628 *Acad Sci U S A*. 2015 Dec 29;112(52):16018–23.
- 629 62. Phan A, Gabor CS, Favaro KJ, Kaschack S, Armstrong JN, MacLusky NJ, et al. Low doses of 17 β -
630 estradiol rapidly improve learning and increase hippocampal dendritic spines.
631 *Neuropsychopharmacol*. 2012 Sep;37(10):2299–309.
- 632 63. Sheppard PAS, Asling HA, Walczyk-Mooradally A, Armstrong SE, Elad VM, Lalonde J, et al. Protein
633 synthesis and actin polymerization in the rapid effects of 17 β -estradiol on short-term social
634 memory and dendritic spine dynamics in female mice. *Psychoneuroendocrinology*. 2021 Apr
635 15;105232.
- 636 64. Tuscher JJ, Luine V, Frankfurt M, Frick KM. Estradiol-mediated spine changes in the dorsal
637 hippocampus and medial prefrontal cortex of ovariectomized female mice depend on ERK and
638 mTOR activation in the dorsal hippocampus. *J Neurosci*. 2016 Feb 3;36(5):1483–9.
- 639 65. Luine VN. Estradiol and cognitive function: Past, present and future. *Horm Behav*. 2014 Sep
640 1;66(4):602–18.
- 641 66. Duclot F, Kabbaj M. The estrous cycle surpasses sex differences in regulating the transcriptome
642 in the rat medial prefrontal cortex and reveals an underlying role of early growth response 1.
643 *Genome Biol*. 2015 Dec 2;16(1):256.
- 644 67. Jaric I, Rocks D, Greally JM, Suzuki M, Kundakovic M. Chromatin organization in the female
645 mouse brain fluctuates across the oestrous cycle. *Nat Commun*. 2019 Jun 28;10(1):2851.

- 646 68. Mendell AL, Atwi S, Bailey CDC, McCloskey D, Scharfman HE, MacLusky NJ. Expansion of mossy
647 fibers and CA3 apical dendritic length accompanies the fall in dendritic spine density after
648 gonadectomy in male, but not female, rats. *Brain Struct Funct*. 2017 Jan;222(1):587–601.
- 649 69. Woolley CS, Gould E, Frankfurt M, McEwen BS. Naturally occurring fluctuation in dendritic spine
650 density on adult hippocampal pyramidal neurons. *J Neurosci*. 1990 Dec;10(12):4035–9.
- 651 70. Qiu LR, Germann J, Spring S, Alm C, Vousden DA, Palmert MR, et al. Hippocampal volumes differ
652 across the mouse estrous cycle, can change within 24hours, and associate with cognitive
653 strategies. *NeuroImage*. 2013 Dec 1;83:593–8.
- 654 71. Lisofsky N, Mårtensson J, Eckert A, Lindenberger U, Gallinat J, Kühn S. Hippocampal volume and
655 functional connectivity changes during the female menstrual cycle. *NeuroImage*. 2015 Sep
656 1;118:154–62.
- 657 72. Pletzer B, Harris T, Hidalgo-Lopez E. Subcortical structural changes along the menstrual cycle:
658 Beyond the hippocampus. *Sci Rep*. 2018 Oct 30;8: 16042.
- 659 73. Barha CK, Galea LAM. Influence of different estrogens on neuroplasticity and cognition in the
660 hippocampus. *Biochim Biophys Acta BBA - Gen Subj*. 2010 Oct 1;1800(10):1056–67.
- 661 74. Qin X, Jiang B, Zhang Y. 4E-BP1, a multifactor regulated multifunctional protein. *Cell Cycle*. 2016
662 Feb 22;15(6):781–6.
- 663 75. Cuenda A, Rousseau S. p38 MAP-Kinases pathway regulation, function and role in human
664 diseases. *Biochim Biophys Acta BBA - Mol Cell Res*. 2007 Aug 1;1773(8):1358–75.
- 665 76. Galea LA, McEwen BS, Tanapat P, Deak T, Spencer RL, Dhabhar FS. Sex differences in dendritic
666 atrophy of CA3 pyramidal neurons in response to chronic restraint stress. *Neuroscience*. 1997
667 Dec;81(3):689–97.

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Figure 1: Example of CA1 and dentate gyrus punch location

Blue circles indicate approximate CA1 tissue punch location. Red circles indicate approximate dentate gyrus tissue punch location. Punches were 0.838mm in diameter.

Figure 2: Akt phosphoprotein levels in the dentate gyrus (DG) and CA1 of female and male rats

Phosphoprotein signal normalized by amount of total protein in sample. A) Phosphoprotein levels were higher in females in all analytes in the dorsal DG (pAkt, pGSK-3 β , and pp70S6K: $p < 0.0001$). B) Phosphoprotein levels were higher in females in all analytes in the ventral DG (pAkt: $p < 0.0001$; pGSK-3 β : $p = 0.0004$; pp70S6K: $p = 0.0014$). C) High 17 β -estradiol increased dorsal CA1 pAkt in females ($p = 0.05$). There were no effects of sex between vehicle-treated rats. D) There were no effects of sex or treatment on Akt analytes in the ventral CA1. Error bars are $\pm$ standard error of the mean. Main effects of sex: ## $p < 0.01$, ### $p < 0.001$, #### $p < 0.0001$; effects of 17 β -estradiol: * $p = 0.05$.

Figure 3: MAPK phosphoprotein levels the dentate gyrus (DG) and CA1 of in female and male rats

Phosphoprotein signal normalized by amount of total protein in sample. A) Males had higher pERK1/2 ($p < 0.0001$) in the dorsal DG than females. There were no significant effects of sex among vehicle-treated rats in the dorsal DG for pJNK or pp38. B) In the ventral DG, there were no significant effects of sex among vehicle-treated rats. Low 17 β -estradiol reduced ventral DG pp38 in females ($p < 0.0001$), with high 17 β -estradiol reducing pp38 still further (compared to vehicle, $p < 0.0001$; compared to low 17 β -estradiol $p < 0.01$). C-D) There were no significant effects of sex or treatment on MAPK analytes in the dorsal CA1 (C) or the ventral CA1 (D). Error bars are $\pm$ standard error of the mean. Main effects of sex: #### $p < 0.0001$; effects of 17 β -estradiol: ** $p < 0.01$, **** $p < 0.0001$.

Figure 4: Akt phosphoprotein levels in the dentate gyrus (DG) and CA1 of ovariectomized female rats

Phosphoprotein signal normalized by amount of total protein in sample. A-B) 17 β -estradiol did not affect Akt pathway phosphoprotein levels in the dorsal DG (A) or ventral DG (B) of OVX rats ($ps > 0.27$). C) Low ($p = 0.0113$) and high ($p = 0.0306$) 17 β -estradiol increased pGSK-3 β in the dorsal CA1. D) In the ventral CA1, low 17 β -estradiol increased pAkt (relative to vehicle [$p = 0.0142$] and high 17 β -estradiol [$p = 0.0471$]) and pGSK-3 β ($p = 0.0025$). Error bars are $\pm$ standard error of the mean SEM. Effects of 17 β -estradiol: * $p < 0.05$, ** $p < 0.01$.

Figure 5: MAPK phosphoprotein levels in the dentate gyrus (DG) and CA1 of ovariectomized female rats

Phosphoprotein signal normalized by amount of total protein in sample. A) Low 17 β -estradiol increased pERK ($p = 0.031$) and pJNK ($p = 0.028$) in the dorsal DG of OVX rats. B) 17 β -estradiol did not affect MAPK pathway phosphoprotein levels in the ventral DG of OVX rats ($ps > 0.303$). C) Low ($p = 0.017$) and high ($p = 0.009$) 17 β -estradiol increased pJNK in the dorsal CA1 of OVX rats. D) In the ventral CA1, low 17 β -estradiol increased pERK1/2 (relative to vehicle [$p < 0.01$] and high 17 β -estradiol [$p < 0.05$]) and pJNK (relative to vehicle and high 17 β -estradiol, $ps < 0.05$). Error bars are $\pm$ standard error of the mean. Effects of 17 β -estradiol: * $p < 0.05$, ** $p < 0.01$.

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Akt Pathway Analytes