The olfactory gating of visual preferences to human skin and colors in mosquitoes

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Abstract

Mosquitoes track odors, locate hosts, and find mates visually. The color of a food resource, such as a flower or warm-blooded host, can be dominated by long wavelengths of the visible light spectrum (green to red for humans) and is likely important for object recognition and localization. However, little is known about the hues that attract mosquitoes or how odor affects mosquito visual search behaviors. We used a real-time 3D tracking system and wind tunnel that allowed careful control of the olfactory and visual environment to quantify the behavior of more than 1.3 million mosquito trajectories. We found that CO₂ induces a strong attraction to specific hues, including those that humans perceive as cyan, orange, and red. Sensitivity to orange and red correlates with mosquitoes' strong attraction to the color spectrum of human skin, which is dominated by these wavelengths. Attraction was eliminated by filtering the orange and red bands from the skin color spectrum and by introducing mutations targeting specific long-wavelength opsins or CO₂ detection. Collectively, our results show that odor is critical for mosquitoes' wavelength preferences and that the mosquito visual system is a promising target for inhibiting their attraction to human hosts.

Introduction

The behavioral preference of insects for certain bands in the visible light spectrum plays a profound role in structuring ecological communities by mediating processes such as plant-insect/predator-prey interactions and disease transmission¹⁻³. For biting insects, such as mosquitoes, tsetse flies, and kissing bugs, vision plays an essential role in various behaviors, including flight control, object tracking for host- or nectar-finding, and locating oviposition sites⁴. The visual stimuli that mediate these behaviors are integrally tied to other host-related cues, such as scent and heat. For instance, when combined with an odor lure, tsetse flies are highly attracted to what humans perceive as blue color^{5,6}, and kissing bugs prefer visual objects only when also associated with odors⁷. Visually guided mosquito behaviors are also thought to play a role in host attraction⁸⁻¹⁰. It has long been known that mosquitoes are attracted to dark, high-contrast objects^{9,11}, which has led to the development of black traps¹⁰. For high-contrast visual stimuli, recent work has shown that certain odors stimulate visual search behaviors in *Aedes aegypti* mosquitoes. This species is not attracted to black objects in the absence of CO₂, but after encountering a CO₂ plume, they become highly attracted to such objects¹¹. Other cues (heat, water vapor, skin volatiles) mediate behaviors such as landing and biting¹¹⁻¹⁴.

Despite the potential importance of color in mediating mosquito biting behaviors, surprisingly, details regarding other wavelengths that attract mosquitoes or how odors sensitize that attraction remain unclear. The visual spectra of important resources can be diverse and dominated by short and medium

wavelengths (e.g., flowers or oviposition sites) or long wavelengths (e.g., human skin) (Fig. 1a, b). Despite interest in developing traps and lures that exploit mosquito spectral preferences, only a few studies have examined these preferences, and the results of those studies have been contradictory. For instance, studies of *Ae. aegypti* have shown no difference in spectral preference in the 450–600 nm wavelength range^{15,16}. By contrast, other studies have demonstrated specific preferences but for different wavelength bands: *Ae. aegypti* mosquitoes were attracted to blue in one study¹⁷ and only to green-yellow in another¹⁸. Other studies have shown that mosquitoes sometimes prefer red^{15,19,20}, although it is thought that mosquitoes lack opsin receptors sensitive to these wavelengths. Because mosquitoes are attracted to dark visual objects, responses to red may represent achromatic responses from visual channels that are sensitive to medium-wavelengths and therefore are perceived as dark grey or black when presented against a light-colored background. Nevertheless, these prior studies did not characterize the actual flight trajectories of the mosquitoes, nor control for the change in behavioral state associated with the smell of a host. Accurate control of both a visual object's reflectance and its contrast with the background is required to determine whether mosquitoes are attracted to specific wavelengths.

Aedes aegypti provides an excellent model for studies aimed at elucidating spectral preferences and determining how these preferences are modulated by odor. *Aedes aegypti*, which are active during the dawn and dusk periods²¹, have 10 rhodopsins, 5 of which are expressed in the adult eye²². Little is known about opsin tuning, although they are orthologs of medium-wavelength sensitive opsins (green), and previous electroretinogram (ERG) studies suggested that *Ae. aegypti* is sensitive to medium-long wavelengths in the green-yellow spectrum^{23, 24}. In this study, we used a large wind tunnel and computerized vision system to close these knowledge gaps regarding mosquito visual and olfactory responses by examining *Ae. aegypti* free-flight responses to objects of different wavelengths, with and without the presence of CO₂. We show that when experiencing odor, mosquitoes become particularly attracted to hues that are dominant in human skin. We also demonstrate that knockout of either the olfactory channel that gates visual attraction or the opsins that allow detection of objects that reflect long wavelengths eliminates attraction to skin tones.

Results

Olfactory gating of spectral preferences of Ae. aegypti mosquitoes

Examining olfactory and visual search behaviors in mosquitoes often requires simulating conditions in which the statistics of the stimuli (e.g., intensity, duration) and resulting mosquito behavior are as natural as possible. We therefore examined *Ae. aegypti* behavior in a large wind tunnel spanning 450 mosquito body lengths and equipped with a 16-camera, real-time tracking system for monitoring and quantifying

mosquito behaviors^{11,25}. A checkerboard pattern was projected on the bottom (floor) of the wind tunnel, and a low-contrast grey horizon was projected on each side of the tunnel to provide optic flow (Fig. 1c). Similar to our previous assays, we placed two identically sized circles (3-cm diameter) on the floor of the tunnel in the upwind area of the working section, 18 cm apart and 33 cm from the odor source (Fig. 1c). In each experimental trial, 50 mated *Ae. aegypti* females were released into the tunnel, and their trajectories were recorded over a 3-h period (1.3 million total trajectories were recorded, with an average trajectory duration of 3 s). The tunnel was filled with filtered air for 1 h, after which a CO₂ plume (95% filtered air, 5% CO₂) located 33 cm away and separate from the visual objects was introduced into the tunnel and left for 1 h (Fig. 1d). Measurements of the plume showed an exponential decay, typical of turbulent diffusion, with a concentration of ~1500 ppm approximately 30 cm from the odor source. In the last hour of the experiment, only filtered air was released into the wind tunnel.

During exposure to filtered air, the mosquitoes exhibited random behavior to the odor source and visual objects, and they spent much of their time exploring the ceiling and walls of the tunnel and rarely investigated the visual objects (Fig. 1e). By contrast, upon exposure to the CO₂ plume, the number of flying mosquitoes more than doubled (Fig. 1f, h; Wilcoxon signed-rank test, number of trajectories in Air vs. CO₂, $P < 0.002$). During this time, the mosquitoes exhibited odor-tracking behavior, spending most of the time in the working section's central area with significantly elevated flight velocities (Figs. 1f, s1; Kruskal-Wallis test: $df = 2$, Chi-square = 597.23, $P < 0.0001$). The CO₂ also triggered an attraction to the visual objects. The mosquitoes showed no interest in the objects during the filtered air treatment (only 1–4% of mosquitoes investigated), but during CO₂ release, the percentage and number of mosquitoes investigating the visual objects increased significantly (21%; paired Student's t-test: $P = 0.002$) (Supplementary Fig. S1). After the plume was stopped, the attraction to the visual objects ceased (Fig. 1e, Supplementary Fig. S1e; Wilcoxon signed-rank test, CO₂ vs. Post-CO₂: $P < 0.001$).

To ensure that mosquito visual preference behaviors were in response to discrete wavelength bands rather than object contrast, we employed visual stimuli in the range 430 to 660 nm (violet to red to a human observer; hereafter, visual stimuli are referred to as they are perceived by the human eye without implying that mosquitoes have the same subjective experience of color), with each visual stimulus having the same approximate contrast with the background (Fig. 1g). While investigating the visual objects, the mosquitoes would fly upwind and hover immediately downwind of a visual object, at approximately 3–5 cm, while exhibiting brief excursions before returning to the objects (Fig. 1e). During CO₂ release, the same total number of mosquitoes was recruited to the visual objects of differing hues (Supplementary Fig. S1e). Relative to the white control object, however, the mosquitoes preferred certain hues, such as cyan, red, and orange,

Figure 1. Olfactory gating of mosquito color preference. (a) The *Ae. aegypti* eye. The retina is composed of ~300-400 ommatidia and ranges in spectral sensitivity from the near-ultraviolet (300 nm) to orange-red (600-700 nm) wavelengths. Image courtesy of Raúl Pardo (with permission). (b) Spectral reflectance of behaviorally important objects for *Ae. aegypti* females: human skin (brown line); flower (*P. obtusata*; green line) and small puddle filled with *Ae. aegypti* larvae (blue line). (c) Wind tunnel system with real-time tracking system, odor and visual stimulation. (d) Heat map of the CO₂ plume in the wind tunnel. (e) Example of individual trajectories (top: [x-,y-axes], bottom: side view [x-,z-axes]). The arrows represent the start of a trajectory and the circles are the visual objects. (f) Heat (occupancy) maps showing the distribution of female mosquitoes without (left panels) and with CO₂ delivery (right panels) while in presence of a white and a red objects (top), or white and green objects (bottom). The position of the plots shows the top and side views of the tunnel working section. (g) Reflectance of the visual stimuli used in the experiments. (h) Relative flight activity between the different phases of the experiments (pre-, CO₂ and post-CO₂) as the number of trajectories recorded during one phase divided by the number of trajectories recorded in the previous phase. There were no significant differences in the relative activity during the CO₂ phase when the tested visual object was black versus objects of different hues (Kruskal-Wallis test, $df = 1$, Chi-sq = 0.01, $P = 0.92$), although for both groups CO₂ significantly elevated the number of flying mosquitoes compared to the filtered Air treatment ($P < 0.002$). (i) Mean preference index for the test object (black, or different hues) vs. the control (white) object. There was a significant effect of hue on the attraction to the tested object (Kruskal-Wallis test, Preference index ~ pair of visual stimuli used: $df = 8$, Chi-sq = 597.23, $P < 0.0001$). Several hues were significantly more attractive than the control, white object (one-sample t-test: ***; $P < 0.001$). Boxplots area the mean (line) with 95% confidence interval (shaded area) ($n = 25,529; 17,729; 53,786; 23,694; 34,343; 31,037; 32,257; 24,774; 42,595; 20,929$; and 48,198 mosquito trajectories for the all-white, all-black, black-and-white, Bv-T2, Bw-, Gw-T1, Gc-, YGc-, Yw-, O- and R-Hue treatments, respectively).

as demonstrated by their focused clustering around the object (Fig. 1e). By contrast, other hues (violet, blue, green, and green-yellow) elicited no attraction responses compared to the white object (Fig. 1e, i). Across all hues, CO₂ had a strong effect on flight velocity and duration, but there were no significant differences between treatments (Supplementary Fig. S1b, d; Kruskal-Wallis test: Chi-square < 16.82, $P > 0.11$), demonstrating that the presence of CO₂ is necessary for attraction to specific hues and that visual

preference did not result from higher flight velocity increasing the probability of a mosquito randomly encountering a visual object.

To further investigate mosquito spectral preferences, a preference index value (defined as the time spent investigating a colored object minus the time investigating the white object, divided by the sum of the times spent investigating the colored and white objects) was calculated for each mosquito that investigated a visual object. The hues differed significantly in terms of mosquito preference (Fig. 1i, Kruskal-Wallis test: $df = 8$, Chi-square = 597.23, $P < 0.0001$); several hues were more attractive to mosquitoes than the white object that served as a non-attracting control (Fig. 1i, one-sample t-test: $P < 0.001$). As the hues transitioned from green to red, the attractiveness of the object also increased. For instance, orange and red visual objects were strongly preferred by female mosquitoes (multiple comparison Kruskal-Wallis test: $P < 0.05$), whereas green, blue, and violet objects were not more attractive than the white control object (multiple comparison Kruskal-Wallis test: $P > 0.05$). However, mosquitoes were not strictly attracted to the longest wavelengths, as they were also significantly attracted to cyan objects (peak reflectance at 496 nm). As a control to test for the effect of visual object attraction relative to other regions of the working section, we examined the preference between the white (control) object and a randomly selected volume in the wind tunnel. Compared with the randomly selected volume, female mosquitoes investigated the white object significantly more in presence of CO_2 (preference index = -0.53 ± 0.03 (mean $\pm$ sem), one-sample t-test: $P < 0.001$).

Behavioral preferences for orange-red wavelengths reflected from human skin

Across all skin tones, human skin has a strong signature in the long wavelength range (yellow, red) (Fig. 2a)²⁶, but it is unclear which bands in the human skin spectrum are most attractive to mosquitoes. To examine whether mosquitoes exhibit different preferences for certain spectral bands reflected by human skin, we first utilized color cards designed for cosmetics purposes to match human skin tones (Pantone SkinTone Guide) (Fig. 2a). Behavioral experiments were performed in the wind tunnel to individually test various faux skin tones (Y02, Y10, R10, and an unpleasant orange shade typical of individuals using cheap tanning lotion [which we designated “vile 45”]) using the white object as a control. Similar to the previous experiments, only during CO_2 release did mosquitoes become highly attracted to skin tones (Fig. 2c, e; Kruskal-Wallis test: $df = 8$, Chi-square = 184.37, $P < 0.001$), exhibiting no behavior characteristic of attraction before exposure to CO_2 (paired t-test: $P = 0.09$). Moreover, the mosquitoes exhibited similar levels of attraction to each skin tone (Fig. 2c, e; Kruskal-Wallis test with multiple comparisons: $P > 0.05$).

Figure 2. The contribution of orange-red wavelengths in attraction to faux human skin. (a) Spectral reflectance of human skin and faux skin used in behavioral experiments. (b) Ultra-thin optical filters (450 nm, 600 nm, and 700 nm) attenuated discrete bands in the object's reflected spectrum. (c, d) Occupancy maps of the mosquito's distribution around the visual objects during exposure to CO₂. During CO₂, mosquitoes were significantly attracted to the faux skin color compared to the white (control) object (c). However, the optical filter attenuating the 550-630 nm band reduced the numbers of mosquitoes investigating the faux skin color (d). (e, f) Mosquitoes significantly preferred the faux human skin colors (E), although optical filters in the yellow to red wavelengths significantly decreased the attractiveness of the visual object (F). Boxplots are the mean (line) with 95% confidence interval (shaded area); letters denote statistically significant differences between groups. (g) Testing a mosquito line deficient in long-wavelength opsins (*opsin1* and *opsin2*), or unable to detect CO₂ (*Gr3* mutant) allowed us to examine the contribution of olfactory and visual input into the attraction of human skin color. (h) Mean preference indices for the *Gr3* mutants (blue) and opsin mutants (orange). All mosquito lines showed similar preferences to the white and skin color visual objects during exposure to filtered air (Kruskal-Wallis test with multiple comparisons: df = 3, Chi-sq = 1.68, P = 0.64). However, during CO₂ the lines were significantly different from one another in their visual preferences (Kruskal-Wallis test with multiple comparisons: df = 3, Chi-sq = 96.01, P < 0.001): only the heterozygote (*Gr3*^{+/+}) and wild-type (LVP) lines showed significant attraction to the skin color (one-sample t-test: P < 0.001), whereas the opsin double mutant line (*op-1/op-2*) and the *Gr3*^{-/-} mutants showed no attraction (one-sample t-test: P > 0.31). Boxplots area the mean (line) with 95% confidence interval (shaded area) (n = 13,999; 15,035; 14,016; 48,624; 19,284; 20,713; 40,690; 26,135; 39,649; 16,208; 3,550; 9,277; 13,799; 10,948; and 5,679 mosquito trajectories for the Y02, Vile 45, Y10, R10, IR filter, coverslip, 450nm filter, 600nm filter, 700nm filter, wt, *Gr3*^{+/+}, *Gr3*^{+/+}, *op-1R*, *op-2G*, and *op-1R,op-2G* treatments, respectively).

To determine which region of the human skin visual spectrum is most attractive to mosquitoes, we overlaid the R10 skin color card with optical filters to attenuate discrete bands (Fig. 2b). Whereas the 450-nm optical filter had no significant effect on behavioral attraction to the skin tone compared with the

positive controls (Fig. 2f; Kruskal-Wallis test with multiple comparisons: $P > 0.58$), filters blocking longer wavelengths (550- to 700-nm) reduced the attractiveness of the visual object ($P < 0.05$). In particular, application of the 600-nm filter was associated with a 300% reduction in attraction compared with the positive controls (Fig. 2c–f). Importantly, results for controls consisting of an overlaid infrared filter or a clear nylon coverslip did not differ significantly from the unmanipulated skin tone (Kruskal-Wallis test with multiple comparisons: $P > 0.33$). To further examine the importance of visual and olfactory integration in controlling mosquito visual preferences, we examined single and double mutants of the long-wavelength photoreceptors *opsin-1* and *opsin-2* and a line with a mutation in the *Gr3* receptor¹⁴, which transduces CO₂ signals (Fig. 2g). Whereas the *Gr3*-heterozygote, *opsin-1* and *opsin-2* single mutants, and wild-type control mosquitoes were significantly attracted to the skin tone during CO₂ exposure (one-sample t-test: $P < 0.001$), neither the *opsin-1,opsin-2* double mutant nor *Gr3* mutant were attracted to the skin tones (Fig. 2h; one-sample t-test: $P > 0.31$).

The behavioral preference to the long wavelengths in the skin color cards may not reflect mosquito behaviors to the hues reflected from human skin. To examine this further, we tested mosquitoes in a smaller opaque cage (45 cm x 30 cm x 30 cm) where they were exposed to two small windows (16 cm²) (Fig. 3a,b). The cage and its environs were constructed to minimize any uncontrolled contamination from thermal or olfactory cues, and the windows being made from clear, heat absorptive glass (Supplementary Fig. S2). The back of a hand was displayed in one window, and the back of a heat-protective white glove was displayed in the other window (as a control). Similar to the assays in the wind tunnel, we found that mosquitoes were highly activated by CO₂ (Fig. 3c-e), and this increased their visual attraction to visual stimuli, including skin (Fig. 3b, c, f; Kruskal-Wallis test: $df = 2$, Chi-square = 84.04, $P < 0.001$). Mosquitoes showed no preference during control experiments with two white gloves displayed in the window, but significantly preferred skin (Fig. 3f; Kruskal-Wallis test with multiple comparisons: $P < 0.001$). However, when optical filters were placed over the window, blocking the longer wavelengths (550- to 700-nm), the attraction was significantly reduced (Kruskal-Wallis test with multiple comparisons: $P < 0.001$) and not significantly different from the negative control (Fig. 3f; Kruskal-Wallis test with multiple comparison: $P = 0.34$). Collectively, these results demonstrate that the long-wavelength band of the visual spectrum plays an important role in determining mosquito attraction to skin color. In addition, knockout of either visual or olfactory detection receptors suppresses mosquito visual attraction to long-wavelength host cues.

Figure 3. The importance of long wavelengths in attraction to human skin. (a) Cage assay with real-time tracking system, odor, and visual stimulation through two windows on the front of the cage. (b) Example of individual trajectories (top: skin and control (white glove), bottom: skin+filters (550-700nm) and control). (c) Heat maps showing the distribution of female mosquitoes during CO₂ stimulation while in presence of the skin and control (top), and the skin+filters (550-700nm) and control (bottom). (d) Relative flight activity between the different phases of the experiments (pre-, CO₂ and post-CO₂). There was no significant difference in the relative activity during the CO₂ phase between the skin and skin+600-nm filter treatments (Kruskal-Wallis test with multiple comparisons: $P > 0.57$). (e) The percentage of mosquitoes visiting the windows over the duration of the experiment. Few mosquitoes investigated the windows before the CO₂ exposure. However, exposure to CO₂ significantly increased the numbers of mosquitoes visiting the windows relative to the pre-CO₂ period (Kruskal-Wallis test with multiple comparisons: $df=5$, $\chi^2 = 277.85$, $P < 0.0001$), although during CO₂ there were no significant differences in the total number of mosquitoes investigating the windows between treatment groups (Kruskal-Wallis test with multiple comparisons: $P > 0.57$). Lines are the means and shaded areas the $\pm$ SEM. (f) Mean preference index for the different treatment groups (white glove vs. white glove, skin vs. white glove, and skin+filter (550-700nm) vs. white glove). Boxplots area the mean (line) with 95% confidence interval (shaded area). Different letters denote statistically significant differences between groups (Kruskal-Wallis test with multiple comparisons, $P < 0.01$). (n = 13,597 for the skin treatment group; n = 9,502 for the skin+filters treatment group; and n = 9,368 for the control group).

Spectral sensitivity of the *Aedes* eye

The preference of mosquitoes for long wavelengths in the orange-red band motivated us to examine the sensitivity of the *Ae. aegypti* retina by recording ERGs that extracellularly measure the summed responses of retinal cells to visual stimuli (Fig. 4a). In the first series of experiments, a moving bar of differing hue (blue [peak 451 nm], green [537 nm], or red [>600 nm], all at the same intensity; 18° wide at 30°/s clockwise) was projected on a black background while conducting the ERG recordings (Fig. 4b). When the moving bar reached the mosquito's visual field, the ERG exhibited a negative response that quickly returned to baseline after the bar moved past the mosquito's field of view (Fig. 4b). A significant difference in hue-evoked responses was observed (Kruskal-Wallis test: $df = 3$, 40.03, $P < 0.001$), with the blue and green bars eliciting the strongest responses (Fig. 4b). Although responses to the red bar were

significantly weaker, the red hue still elicited ERG responses that were significantly higher than the baseline and those of the no-stimulus controls (Wilcoxon signed-rank test: $P < 0.001$).

To further characterize mosquito spectral sensitivity, we used a scanning monochromator to examine ERG responses across the near ultraviolet (UV) to far-red wavelength range (350–750 nm). *Ae. aegypti* exhibited the highest sensitivity to violet (410 nm, 3.2 mV) and cyan-green wavelengths (520 nm, 2.8 mV) (Fig. 4c, d). Strong ERG responses (0.47–1.27 mV) were still noted in the yellow-red wavelengths, although at >700 nm, the responses decreased to approximately 0.25 mV, which was still significantly higher than the baseline control (Wilcoxon signed-rank test, $P < 0.03$).

Figure 4. Retinal sensitivity to visual stimuli. (a) Experimental setup for the ERG experiments. A moving bar of different hue values (all at the same intensity, 18° wide at $30^\circ/\text{s}$ clockwise) was projected on a black background while conducting the ERG recordings. (b) Electroretinogram responses to a blue, green or red moving bar (mean $\pm$ sem; $n = 7$ mosquitoes). Responses to the moving object were significantly higher than the baseline for all tested hues (blue, green or red), although blue and green bars elicited higher responses (Kruskal-Wallis test, Amplitude responses $\sim$ color moving bar, $df = 2$, 40.03 , $P < 0.001$, $n = 7$ mosquitoes). (c) ERG responses to pulses of light ranging from 350 to 750 nm in 10 nm intervals. Traces are the mean responses (shaded area is the $\pm$ sem; $n = 8$ mosquitoes) to discrete wavelengths showing the elevated responses to violet (410 nm) and cyan-green (520 nm) hues. (d) Retinal sensitivity curve across the tested wavelengths from 350 to 750 nm (10 nm intervals). Two maxima occurred at the violet (420 nm) and green (~ 530 nm) wavelengths, although responses were still significantly elevated above the noise at wavelengths more than 650 nm (t-test, ERG vs. noise: $P < 0.01$).

The role of visual contrast in determining mosquito preferences

Mosquitoes are very sensitive to detecting dark objects that contrast highly with the background^{8,11}. In the above experiments, we kept the total object contrast (400–700 nm) with the background approximately the same, but the mosquitoes' preference for red objects and skin tones motivated us to evaluate whether these responses were due to contrast alone (calculated as the Weber contrast, or the difference in spectral

irradiance reflected by an object and the background, divided by the sum of the two) or whether mosquitoes can discriminate red-orange objects independently of intensity. As the first step in determining how features of a visual stimulus impact mosquito visual preference, grey objects that contrasted differently with the background were tested against a white control object (Fig. 5, Weber Contrasts: -0.28 to 0.02). Similar to the above results and across all tested stimuli, the presence of CO_2 increased mosquito flight activity and the number of visits to the visual objects (Fig. 5c, Wilcoxon signed-rank test, Air vs. CO_2 : $P < 0.001$). Female mosquitoes exhibited significantly greater attraction to the majority of the grey visual objects than the white control object (Fig. 5d, one-sample t-test: $P < 0.001$). However, when exposed to the lightest grey object, which closely approximated the background and the white object, mosquitoes showed no preference for either object (Fig. 5d; Student's t-test: $P = 0.33$). Overall, object darkness and contrast with the lighter background was significantly related to mosquito preference, with mosquitoes investigating and preferring darker objects (Fig. 5d; Kruskal-Wallis test: $df = 5$, Chi-square = 634.16 , $P < 0.001$). Although mosquitoes showed a distinct preference for darker objects, mosquito flight velocity and duration did not significantly differ across treatments (Kruskal Wallis test: Chi-square > 7.40 , $df = 5$, $P > 0.055$).

Figure 5. The effect of achromatic contrast on mosquito attraction to visual objects. (a) Occupancy map of the distribution of female mosquitoes during CO_2 delivery in presence of a white and a light grey object (Weber Contrast: -0.05). The top plot is the top view of the working section of the tunnel (x,y), and the bottom plot is the side view (x,z). (b) As in a, except the grey object has a higher contrast (-0.23). (c) Relative flight activity between the different phases of the experiments (pre-, $+\text{CO}_2$ and post- CO_2). Mosquitoes exhibited similar flight activities across all tested visual objects (Kruskal-Wallis test, $df = 1$, Chi-sq = 3.24 , $P = 0.07$). (d) Mean preference index for the test (grey, or black) vs. control object (white) with 95% confidence interval ($n = 12,764$; $27,537$; $37,085$; $28,644$; $36,050$; $25,896$; and $21,514$ mosquito trajectories for the white, grey9.5, grey6.5, grey4.5, grey4.0, grey2.5, and black treatments, respectively). Object contrast had a significant effect on the attraction to the tested object (Kruskal-Wallis test: Preference Index ~ contrast tested object, $df = 5$, Chi-sq = 634.16 , $P < 0.001$). All grey objects were significantly more attractive than the control, white object (one-sample t-test, **: $P < 0.01$, ***: $P < 0.001$) except for the lightest grey object (-0.05), which was not more attractive (one-sample t-test: $P = 0.33$).

Effects of contrast and color locus discrimination on Aedes preferences

Given the difference between high retinal sensitivity to medium-long (green) wavelengths, lack of attraction to green objects, and the relatively low ERG sensitivity to long (orange/red) wavelengths but strong behavioral attraction to those objects, we asked the following question: what role does the darkness of the object (i.e., its contrast) have on behavioral preferences *versus* the object's hue? Although we lacked the photoreceptor tunings that would allow us to normalize the inputs between spectral channels, we experimentally manipulated object contrast to equalize the perceptual contrast of the objects to the mosquitoes and then examined the mosquito's ability to discriminate between different bands of the visual spectrum.

We first performed behavioral experiments in which we altered the darkness of the grey objects to determine the contrast that matched the attraction shown to the red hue (Fig. 6a, b). Across a range of grey contrast levels, the red hue was significantly more attractive to the mosquitoes than the grey objects (Fig. 6a, c, e; Kruskal-Wallis test: Chi-square = 149.75, df = 6, $P < 0.0001$). As the grey objects became darker, however, they became more attractive to the mosquitoes, and the strong preference for the red hue decreased (Fig. 6e). The attractiveness of the red hue equaled that of the darkest grey and black objects (Fig. 6e; one-sample t-test: $P = 0.07$ and $P = 0.08$ for the darkest grey [grey 1.5] and black objects, respectively). We then determined which contrast of a green hue matched the attraction to the red object. For this purpose, we used the darkest grey object that was equally attractive as the red hue and tested it against different green objects (peak wavelength = 510 nm) that differed in terms of background contrast (Fig. 6b). The dark grey object was significantly more attractive than most green objects, but the darkest green object elicited the same level of attraction (Fig. 6f; preference index = -0.01; one-sample t-test: $P = 0.66$).

To determine whether mosquitoes can discriminate between objects of different hues but similar levels of apparent contrast, we tested attraction to the red hue *versus* the darkest green (Fig. 6g, h). During exposure to CO₂, mosquitoes were strongly attracted to the red hue but not the dark green (Fig. 6g, h; one-tailed t-test: $P < 0.0001$). We also determined whether mosquitoes could discriminate between closely related color loci. For this purpose, we matched the apparent contrast of the non-attractive blue hue to an attractive grey and then subsequently compared mosquito responses to cyan (another attractive hue) and dark blue, as well as cyan and dark green. Similar to the results observed with the red hue, mosquitoes significantly preferred the cyan objects over the dark green or dark blue objects (Fig. 6h; one-tailed t-test: $P < 0.0001$). Thus, mosquitoes easily discriminated between color hues even when the object contrasts matched.

Figure 6. The role of hue versus contrast in mosquito visual attraction. (a) Spectral reflectance of red (R-Hue), black, and grey objects used in the experiments: Grey 9.5, 6.5, 4.5, 4.0, 2.5, and 1.5, with Weber contrast values of -0.17, -0.30, -0.05, -0.10, -0.20, -0.24, -0.27, and -0.28, respectively. (b) Spectral reflectance of cyan and green objects used in the experiments: Gw-T1, Gw-T3, Gc-T1, Gc-Hue, G-S1, and G-S2, with Weber contrast values of -0.18, -0.11, -0.17, -0.20, -0.25, and -0.27, respectively. (c, d) Occupancy maps showing the distribution of female mosquitoes during CO₂ delivery in presence of the red (R-Hue) and Grey2.5 objects (c), or green (G-S2) and Grey1.5 objects (d); both the green and Grey1.5 have the same levels of contrast with the background (Weber Contrasts of -0.27). The upper and lower plots correspond to the top (x,y) and side (x,z) views of the wind tunnel. (e) Mean preference indices ($\pm$ CI) for the red versus grey objects with different levels of contrast with the background. As the grey object's contrast increased and became darker, the relative attraction to the red object decreased, until at Grey1.5 (Weber Contrast value of -0.28) the preference was not significantly different from 0 (one-sample t-test: $P = 0.07$). (f) The Grey1.5 object was subsequently used in experiments testing the attractiveness of green objects with different levels of contrast (blue arrow from E). The grey object was significantly more attractive than the majority of the tested green objects (one-sample t-test: $P < 0.001$, denoted by asterisks), although the darkest green (G-S2; Weber Contrast = -0.27) was not significantly different from 0 (one-sample t-test: $P = 0.66$) ($n = 10,098$ – $15,578$ trajectories for each tested object). (g) As in c, occupancy maps showing the distribution of trajectories around the red and dark green objects. (h) Mean preference indices ($\pm$ CI) for red versus dark green objects, and cyan versus dark blue or dark green objects. The dark green and dark blue objects, with similar apparent contrasts to the red or cyan objects, were tested as the controls. Mosquitoes significantly preferred the red and cyan objects over the dark green or dark blue objects (one-sample t-test: $P < 0.0001$, denoted by asterisks). For panels e, f and h, boxplots area the mean (line) with 95% confidence interval (shaded area), and asterisks denote $P < 0.001$ (one-sample t-test) ($n = 7,191$ – $27,717$ mosquito trajectories for each tested object).

By incorporating object contrasts, reflectance values, and peak wavelengths as independent variables into a series of linear models, the results of the behavioral tests offered a means to examine the

relative contributions of these variables toward mosquito preferences. In these models, all possible combinations were tested, and the best model was selected based on its Akaike Information Criterion (AIC) score, where the AIC estimates the value of each model and lower scores reflect the quality of the statistical model. Using combinations of the independent variables, we found the best model (and hence, lowest AIC score) relied on object contrast and wavelength, and excluded reflectance (Supplementary Fig. S3a, b). But which hues might be critical for mediating these behaviors? To further explore the relationship between the specific wavelengths of an object's hue and its attractiveness, a series of linear models were run using a multivariate analysis (PCA) of each object's visual spectrum. The PCA analysis allowed reduction of highly collinear and dimensional spectral data into reduced components, which can then be used as independent variables in the model. The best model explained approximately 17% of the variance in hue attractiveness (Supplementary Fig. S3c) and indicated that preference is negatively correlated with the content of the green band (500 to 575 nm) in the object's visual spectrum (Supplementary Fig. S3d, e).

Comparison of color preference between mosquito species

The strong and specific responses of *Ae. aegypti* to hues in the visual spectrum and the similarity in long-wavelength opsin gene expansion in other mosquito species²² motivated us to examine the color preferences in *Anopheles (An.) stephensi* and *Culex (Cx.) quinquefasciatus* mosquitoes. To examine the color preferences in *An. stephensi* and *Cx. quinquefasciatus*, we first conducted ERG recordings of 7-day-old females and examined their retinal responses to discrete wavelengths from 350 to 750 nm (Fig. 7a). Both mosquito species exhibited the strongest response to UV-visible violet wavelengths (350–420 nm), and the second strongest response was observed in the cyan-green wavelengths (500–520 nm). The strong response of *An. stephensi* and *Cx. quinquefasciatus* to the UV wavelengths contrasted with that of *Ae. aegypti* (dashed line), which exhibited the strongest response in the violet range (Figs. 3d and 7b,c). All three species exhibited similar responses to the long wavelengths in the orange to red band (620–750 nm).

Are mosquito species' behavioral preferences for visual objects correlated with their ERG responses, and are their color preferences similar? To answer these questions, we tested the responses of *An. stephensi* and *Cx. quinquefasciatus* mosquitoes to blue, green, red, and black objects in the wind tunnel using a methodology similar to that used for *Ae. aegypti*, except at lower light levels (1.28 $\mu\text{W}/\text{cm}^2$). As with the results for *Ae. aegypti*, exposure to CO₂ caused a doubling in the number of flying mosquitoes and increased the percentage of mosquitoes that investigated the visual objects 5.58- to 9.15-

fold relative to air-only treatment for *An. stephensi* and *Cx. quinquefasciatus*, respectively (Fig. 7d; Kruskal-Wallis test: $df = 2$, Chi-square > 7 , $P < 0.01$). However, in contrast to the tight clustering around attractive visual objects by *Ae. aegypti* (Fig. 1f), occupancy maps showed that the responses of *An. stephensi* and *Cx. quinquefasciatus* mosquitoes were much more diffuse (Fig. 7e, f). Nonetheless, *Cx. quinquefasciatus* formed a clustering hotspot around the outlet of the odor plume nozzle that was much stronger than the responses of the other two species (Figs. 1f; 7e,f). Although we tried to minimize the odor nozzle's visual signature by using clear acrylic and tubing, *Cx. quinquefasciatus* mosquitoes might have located the plume source based on the high CO₂ concentration or by seeing the nozzle to some degree.

Figure 7. Species-specific responses to hues. (a) ERG recordings from *Ae. aegypti*, *An. stephensi* and *Cx. quinquefasciatus* mosquitoes. Traces are responses to 1-sec. pulses of light in the 350 nm, 400 nm, and 700 nm bands. Each trace is the mean $\pm$ sem of 7-9 mosquitoes/species. (b, c) Retinal sensitivity to discrete wavelengths show that both *An. stephensi* (b) and *Cx. quinquefasciatus* (c) have the strongest responses in the UV (360 nm) and green (520 nm) bands. The dashed line is the retinal sensitivity of *Ae. aegypti*. (d) Representative flight trajectories [(x,y) and (x,z)] of *Cx. quinquefasciatus* (left) and *An. stephensi* (right) mosquitoes. *Cx. quinquefasciatus* showed a mild attraction to the blue object (top), whereas *An. stephensi* showed an attraction to the red object (bottom). (e, f) Occupancy maps (x,y) of *An. stephensi* (e) and *Cx. quinquefasciatus* (f) distribution around the blue (top) and red (bottom) objects during CO₂ exposure. (g-i) The preference indices for *Ae. aegypti* (g), *An. stephensi* (h) and *Cx. quinquefasciatus* (i) in response to the black, red (R-Hue), green (Gc-T1), and blue (Bw-T1) objects. Lines are the means and shaded bars are the confidence intervals, and letters above bars denote statistical comparisons (Kruskal-Wallis test with multiple comparisons: $P < 0.05$) (for *Ae. aegypti*, $n = 29,254$; 13,580; 18,190; and 12,086 trajectories; for *An. stephensi*, $n = 5,817$; 9,134; 2,153; and 5,627 trajectories; and for *Cx. quinquefasciatus*, $n = 3,746$; 13,553; 4,238; and 2,835 for black, red, green, and blue treatments, respectively).

We next examined the preferences of the mosquito species for different spectral objects (black, blue, green, or red) relative to the white object (the non-attractive control). Similar to *Ae. aegypti*, when both *An. stephensi* and *Cx. quinquefasciatus* mosquitoes were subjected to filtered air treatment, they showed no preference for the black object or any of the spectral objects relative to the white control object (preference indices of -0.04 and 0.08 for *An. stephensi* and *Cx. quinquefasciatus*, respectively; Kruskal-Wallis test: $df = 3$, Chi-square < 3.66 , $P > 0.30$). However, their spectral preferences changed when exposed to CO₂. After encountering the CO₂ plume, *An. stephensi* preferred the black and red objects (Fig. 7h; Kruskal-Wallis test: $df = 3$, Chi-square = 38.6, $P < 0.001$), but they were not significantly attracted to the blue or green objects (Fig. 7h; multiple comparison Kruskal-Wallis test: $P > 0.05$). By contrast, *Cx. quinquefasciatus* mosquitoes preferred the blue and red objects (Kruskal-Wallis test: $df = 3$, Chi-square = 13.6, $P < 0.01$; with multiple comparisons: $P < 0.05$) but were not significantly attracted to the green or black objects (Fig. 7i; multiple comparison Kruskal-Wallis test: $P > 0.05$). Collectively, these results show that odor strongly sensitizes attraction to visual objects across mosquito species; however, spectral preferences can be species specific.

Discussion

Free-flight behavioral experiments with mosquitoes have shown that these insects integrate olfactory, visual, skin volatiles, and thermal cues to function efficiently and robustly in complex environments^{11,14,28}. However, we know very little about mosquito visual-guided behaviors or how vision is involved in host selection. In this study, we utilized real-time tracking of mosquito behaviors in a large wind tunnel. The wind tunnel system enables control of aerodynamic conditions to structure the odor plume and allow the plume and visual objects to be decoupled in time and space. Both are important considerations when testing olfactory-visual integration. In this study, and similar to our previous work using achromatic objects^{11,28}, the presence of CO₂ increased mosquito responses to colored objects in a hue-specific manner. Both chromaticity and contrast were important components in visual object attraction and could partly explain mosquito preferences for objects that appear orange and red to human observers. These results were qualitatively similar to those reported by Smart and Brown (1957), who examined the landing responses of mosquitoes on colored cloth in a field¹⁷. The results of their study showed that mosquitoes (*Aedes* sp.) are attracted to red and black cloths, with darker colors being more attractive than lighter shades¹⁷. Our ERG results and wind tunnel assay data suggest that mosquitoes can detect and are attracted to the long wavelength bands in the orange and red portions of the human visual spectrum, although if objects contrast highly with the background and/or are darker, then they become more attractive. The demonstrated spectral preferences of mosquitoes and their lack of attraction to lighter

colors, with dominant bands in the blue and green regions, impacted the fashion industry in the early twentieth century. For instance, Nuttall and Shipley (1902)²⁹ suggested wearing khaki pants would be appropriate for ensembles in a tropical environment, and the US military changed its dress shirts from dark blue to light blue in part to mitigate mosquito biting³⁰. Nonetheless, a key question is what drives these responses in mosquitoes, particularly the attraction to darker and higher-contrast colors? An important component could be the visual OFF responses to the visual background³¹. In our wind tunnel, we projected a light grey checkerboard pattern on the bottom of the tunnel to provide optic flow and contrast with the visual objects. The lack of opsins for the red hue could cause an OFF response in downstream neurons that receive input from the photoreceptors. One complication with this hypothesis is that mosquitoes still preferred the red hue when tested against a dark green object that matched it in apparent contrast, suggesting they have an ability to detect these long wavelengths. This was further demonstrated by the ERG responses to long (red) wavelengths. Recent work in *Drosophila* has shown that photopigments provide a mechanism for flies to detect orange and red wavelengths³², and it is possible that similar processes play a role in mosquitoes.

Although the contrast with a lighter background did impact the attraction responses of *Ae. aegypti* mosquitoes, the preferences for the red and cyan hues were greater than the apparent contrasts of the competing visual cues (green, or blue). The *opsin-1* and *opsin-2* genes, which are tuned to the green to orange band of the visual spectrum, are highly expressed in the mosquito retina. The results of this study and those reported elsewhere²⁵ suggest that *opsin-1* and *opsin-2* play important roles in determining object preference and visual attraction to human skin. Additional research will be needed to identify the opsins and neural circuits involved in small-field object detection and color preference and determine how odor modulates those responses.

Compared with other insects, such as honeybees or the tsetse fly, we know little about mosquito visual ecology or how visual cues are integrated with other senses in these insects. Whereas shallow pools of water can be rich in green and red wavelengths and flowers dominant in blue and green, hosts' skin is dominated by long wavelengths in the green to red band of the visual spectrum (Fig. 1b). Abundant work by the cosmetics industry has shown that human skin has a lower peak in the green wavelength (530 nm, ~20%) and a dominant reflectance in the orange to red wavelengths (>600 nm, 20–60%). The diurnal *Ae. aegypti* and nocturnal and crepuscular *An. stephensi* and *Cx. quinquefasciatus* mosquitoes are all active during periods in which these longer wavelengths are dominant. For example, *Ae. aegypti* exhibit peak activity in the mornings and late afternoons, and *An. stephensi* and *Cx. quinquefasciatus* mosquitoes are especially active during moonlit nights—both environments are long-wavelength shifted^{21,33,34}. In this study, we show that mosquitoes are especially sensitive to long wavelengths (yellow to red) for host

detection; blocking these wavelengths can suppress object attraction. Moreover, we found that mosquitoes can distinguish between closely colored loci (Fig. 6h), even when their apparent contrasts match. Insect photoreceptors within an ommatidial cartridge transduce light intensity and spectral information. At the photoreceptor terminals, they also provide antagonistic inputs to downstream neuron targets, thus allowing discrimination of spectral inputs (termed ‘color opponency’). In *Drosophila melanogaster*, color opponency at the photoreceptor terminals plays an important role in their color discrimination and preference³¹, and it could be that similar processes are at play in mosquitoes. In a variety of insects, including flies, bees, and butterflies, spectrally sensitive photoreceptors form connections with transmedulla neurons that project into the lobula, where additional color and motion processing—including color opponency—occurs³⁵⁻³⁷.

It is important to note that our current experiments did not incorporate close-range cues from a host, such as heat, water vapor, or skin volatiles. These cues play critical roles in controlling landing and biting behaviors, and future work could determine how visual spectra are processed in tandem with these other stimuli. Nonetheless, previous work has shown that visual cues can promote mosquito orientation and search behaviors in combination with odor, heat, or water vapor^{11,14}. The integration of multimodal stimuli in driving behavioral responses raises questions regarding how the sensory systems are linked in the brain. Odor stimulation increases visual responses in the object-detecting neuropil in the *Ae. aegypti* lobula²⁸. Neuropil in this brain region is responsive to moving objects but not wide-field motion. Interestingly, whereas olfactory stimulation increases visual responses in the lobula, visual stimulation does not modulate glomerular responses in the antennal lobe, the primary site for processing olfactory information in the mosquito brain. Why might this occur? Mosquitoes have a relatively poor visual resolution ($\sim 10^\circ$); thus, vision may not provide fine-scale information about the identity of an object. Instead, an object’s odor may provide information about its identity, whereas vision can provide details regarding the location of the object.

Despite the potential importance, few studies have examined retinal responses to long wavelengths in mosquitoes^{23,24}, and how peripheral and downstream visual circuits, such as those in the optic lobes, process this information remains unknown. Evolutionary analyses of long-wavelength opsins in diverse mosquito species have suggested these genes are functionally important. In *Ae. aegypti*, *Anopheles coluzzii*, and *Cx. quinquefasciatus*, these genes have undergone duplication events and may be under positive selection, explaining the commonalities in ERG responses. *Aedes aegypti* has 10 putative opsins, five of which are potential medium- to long-wavelength opsins in the adult (>500 nm)²². One of these genes, *opsin-1*, is expressed in the largest group of photoreceptor cells (R1-R6) in the *Ae. aegypti* eye²³. The R1-R6 photoreceptors form a trapezoid-like structure in the ommatidial cartridge, with the R7

and R8 cells in the middle. Perhaps similar to *D. melanogaster* with rhodopsin-1 (Rh1), the *Ae. aegypti* *opsin-1* is likely involved in motion- and dim-light sensitivity³⁸. The inner R7 and R8 photoreceptors are involved in color vision, and the *Ae. aegypti opsin-2* is expressed in the R7 photoreceptors located in distinct bands on the dorsal and ventral surfaces of the eye, and possibly used for navigation and biting behaviors. For mosquitoes, opsins that differ in spectral tuning can be co-expressed in the R7 photoreceptor in the female retina (e.g., the long-wavelength *opsin-2* and short-wavelength *opsin-9*), thereby increasing their range in wavelength sensitivity³⁹. This may contrast the opsin co-expression in *D. melanogaster* R7 cells in the dorsal region of the eye, where two short (UV)-sensitive opsins are co-expressed⁴⁰, presumably to increase the spectral contrast with the green-absorbing opsins in the R8 layer. The opsin co-expression and broadening of spectral input in mosquitoes could be advantageous under low-light conditions, as photon capture would be maximized, thus allowing for detection of suitable hosts or perhaps sources of nectar^{39,41}, but this advantage would come at the cost of detection of particular wavelength bands. In our study, however, mosquitoes were able to discriminate between distinct and overlapping spectral bands (green and red; cyan and green, and cyan and blue, respectively), all at the same apparent contrasts, suggesting that other opsins, perhaps in the R8 cells, or downstream circuits, play a role in increasing the separability of those hues.

Color vision plays a critical role in a diverse array of insect vectors, and it is integrally tied to olfaction. For instance, traps that incorporate visual and olfactory cues have proven transformative as a low-cost method for controlling tsetse flies in parts of Africa⁴². Like *Ae. aegypti*, the majority of tsetse flies (e.g., *Glossina morsitans morsitans* and *G. pallidipes*) locate hosts based on smell, and once they are within close range (<10 m), visual cues cause the insects to investigate and potentially bite if the object is a host. Beginning with Vale (1974)⁴³ and continuing into the 1990s, researchers found that flies are most attracted to a specific blue (approximately 460 nm), followed by red and black, but they are not attracted to green and white^{5,6}. Researchers subsequently found that incorporating blue and black hues in traps was particularly effective at inducing flies to come into contact with insecticide-treated screens. *Triatoma infestans* (i.e., the kissing bug) is another example of an insect that integrates odor and visual cues to mediate attraction to targets. An aggregation pheromone gates the attraction of *T. infestans* to colored objects, including the red, blue, and black hues, although the bugs always reject green (approximately 525 nm) and white hues⁷. In a similar manner, our results obtained from tests of different mosquito species demonstrate the importance of olfaction in mediating mosquito color preferences. In the absence of CO₂, mosquitoes did not demonstrate any preference between white and colored objects, but they became attracted to specific hues in the presence of CO₂. However, there were species-specific differences in the attractive hues. Whereas *Ae. aegypti* was equally attracted to both red and black objects, *An. stephensi* was most attracted to black, followed by red objects. By contrast, *Cx. quinquefasciatus* was attracted to

blue, followed by red objects; surprisingly, however, this species was not attracted to small black objects. Collectively, the results of our current study and those of other studies show that the visual systems of insect disease vectors and their behaviors constitute attractive targets for the development of traps incorporating visual features that can be species-specific in terms of attraction, thus providing incentive to identify molecular targets that compromise mosquito olfactory-visual responses.

Materials and Methods

Mosquitoes, Odor Delivery, and Wind tunnel

Mosquitoes (*Aedes aegypti*: Rockefeller, Liverpool, *Gr3*[ECFP]¹⁴ and *opsin-1*, *opsin-2* mutant lines; *Anopheles stephensi* (Indian strain) and *Culex quinquefasciatus*) were raised at the University of Washington campus. Mosquito lines were provided from BEI Resources (Manassas, VA, USA) (*Ae. aegypti*: Rockefeller, Liverpool, *Gr3*[ECFP]¹⁴; *An. stephensi* and *Cx. quinquefasciatus*). In the case of the *Ae. aegypti opsin-1*, *opsin-2*, and *opsin-1,opsin-2* lines, we used existing mosquito lines that were generated as previously described²⁵. Briefly, the *op1* and *op2* alleles were generated by selecting short-guide RNAs (sgRNAs) that targeted the *GPROp1* (LOC5568060) and *GPROp2* (LOC5567680) loci²⁵. Lines were homogenized and verified by PCR before testing²⁵. Mosquitoes were raised in groups of 100 individuals and anesthetized with cold to sort males from females after cohabitating for 7 days. At this time, more than 90% of the females have been mated, as indicated by their developing embryos. For each experimental trial in the wind tunnel, we released 50 females into the wind tunnel working section 3 h prior to the mosquito's subjective sunset (time period of peak activity for *Ae. aegypti*). Each visual stimulus was tested in 4 to 12 experimental trials (on average, approximately 5,575 trajectories were quantified per trial) and 12,300 *Ae. aegypti* were flown in total across all experiments, for a total of 1,305,695 trajectories. After one hour, the 5% CO₂ plume (or filtered air in control experiments), was automatically released from a point source at the immediate upwind section of the tunnel and at a height of 20 cm and in the centerline of the tunnel. The CO₂ remained on for 1 hour, before switching off for another hour of filtered air (post CO₂). The CO₂ and filtered air were automatically delivered using 2 mass flow controllers (MC-200SCCM-D, Alicat Scientific, Tucson, AZ) that were controlled by a Python script that allowed synchronizing odor and filtered air delivery with the trajectory behaviors. The CO₂ plume was quantified using a Li-Cor LI-6262 CO₂/H₂O analyzer (Li-Cor, Lincoln, NE) for a total of 500 locations throughout the tunnel (Fig. 1). Data yielded an exponential decay similar to a model of turbulent diffusion at the air flow (40 cm/s) and turbulent intensities (5%) of this tunnel, such that 20 cm from the

source and parallel to the wind flow the plume was approximately 1700 ppm (Fig. 1c, d), which is in the range of the plume of human breath.

All behavioral experiments took place in a low-speed wind tunnel (ELD Inc., Lake City, MN), with a working section of 224 cm long, 61 cm wide, by 61 cm high with a constant laminar flow of 40 cm/sec (Fig. 1). We used 3 short-throw projectors (LG PH450U, Englewood Cliffs, NJ) and rear projection screens (SpyeDark, Spye, LLC, Minneapolis, MN) to provide a low contrast checkerboard on the floor of the tunnel and grey horizons on each side of the tunnel. The intensity of ambient light from the projectors was 96 lux across the 420-670 nm range. A 3D real-time tracking system^{11,44} was used to track the mosquitoes' trajectories. Sixteen cameras (Basler AC640gm, Exton, PA) were mounted on top of the wind tunnel and recorded mosquito trajectories at 60 frames/sec. All cameras had an opaque Infrared (IR) Optical Wratten Filter (Kodak 89B, Kodak, Rochester, NY) to mitigate the effect of light in the tracking. IR backlights (HK-F3528IR30-X, LedLightsWorld, Bellevue, WA) were installed below and the sides of the wind tunnel to provide constant illumination beyond the visual sensitivity of the mosquitoes. The temperature within the wind tunnel, measured using ibuttons and FLIR cameras, was 22.5°C and did not show any variability within the working section^{11,25}. Ambient CO₂ was constantly measured outside of the tunnel and was approximately 400 ppm.

Visual Stimuli and Experimental Series in the Wind Tunnel

To determine the role of odor in the innate color preferences of mosquitoes, and identify the role of achromatic contrast and wavelength discrimination, a series of different experiments were conducted. In each experiment, two visual stimuli, separated by 18 cm, were presented to the mosquitoes on the floor in the upwind area of the tunnel and perpendicular to the direction of airflow. Visual stimuli consisted of paper circles that were 3 cm diameter (Color-aid Corp., Hudson Fall, NY, USA). Reflectance spectra for all the visual stimuli were characterized using an Ocean Optics USB2000 spectrophotometer with a deuterium tungsten halogen light source (DH2000) calibrated with a white Spectralon standard (Labsphere, North Sutton, NH, USA). The projector and working section light intensities (approximately 100 lux) were measured using a cosine-corrected spectrophotometer (HR-+2000, Ocean Optics, Dunedin, FL, USA) 5 cm from the projector source (63 μ W/cm²). The achromatic contrasts of the visual objects relative to the background were measured using the calibrated spectrophotometer and the Weber contrasts calculated by the intensity of the object (I_{object}) and background ($I_{background}$), where ($I_{object} - I_{background}$)/ $I_{background}$. In the first experimental series, using *wt* (ROCK) *Ae. aegypti*, we examined the innate

preference of individual colors relative to the non-attractive white object. Tested colors were violet, blue, cyan, green, green-yellow, yellow, orange, and red (Bv-T2, Bw-, Gw-T1, Gc-, YGc-, Yw-, O- and R-Hue; Color-aid Corp.). These stimuli all had similar achromatic contrasts (-0.12 to -0.18) and peak reflectance values, but had distinct peak wavelengths (Fig. 1g). The position of the respective hue and white object in each replicate trial was randomized.

In the second experimental series, experiments were performed to examine which spectral bands of the human skin might attract mosquitoes. To ensure the replicability of the experiments, and enable the control of visual object humidity, temperature, and odor, we first elected to first use faux skin mimics (Pantone SkinTone Guide; Pantone LLC, Carlstadt, NJ 07072 USA). Using *wt* (ROCK) *Ae. aegypti*, we tested four different skin tones (R10, Y10, and Y02) and a skin tone that we named “vile 45”, which matched the putrid orange color from individuals who use cheap tanning lotion (PANTONE 16-1449X, Gold Flame). Similar to our previous wind tunnel experiments, each individual skin tone was paired with a white control object. To attenuate different spectral bands reflected from the skin tone, we used ultra-thin (~200 um thick) plastic filters that were placed over the object. Filters were selected to attenuate the 450-530 nm band, the 550-630 nm band, or the 650-730 nm band (36-333 [notch filter], 35-894 [long-pass filter], and 35-896 [long-pass filter], respectively; Edmund Optics Inc., Barrington, NJ USA). Reflectance measurements of the object with the filters showed the transmission loss was <5% for bands outside of the filtered wavelengths (Fig. 2B). As a control to determine if attenuating a spectral band outside of the visual spectrum effects mosquito behavior, we used an IR filter that allows transmission of 350-750 nm band of the visual spectrum (14-547 [KG2], Edmund Optics Inc., Barrington, NJ USA). To control for the physical effect that placing the plastic filter over the object may have had on mosquito behavior, we used plastic coverslips with the same refractive index as glass (72261-22; Electron Microscopy Services, Hatfield, PA USA). Experiments were also conducted with *Gr3^{-/-}* mutants¹⁴ and the *opsin-1^{-/-}*, *opsin-2^{-/-}* double mutants²⁵ to examine how loss of olfactory or visual detection, respectively, impacted mosquito attraction to the skin color. As controls, we used heterozygote (*Gr3^{+/-}*), single mutants (*opsin-1^{-/-}*, *opsin-2^{-/-}*), and the wild-type (Liverpool) lines.

In the third series, the role of achromatic contrast, and not wavelength, on mosquito visual preferences was examined. Grey circles (3 cm diameter), differing in their Weber Contrast (-0.28 to -0.05) and ranging from near-black to very light grey (Greys 1.5, 2.5, 4.0, 4.5, 6.5, and 9.5; Color-aid Corp., Greyset), were run in combination with a white circle (Weber Contrast of 0.02). Similar to the above experiments, *wt* (ROCK) *Ae. aegypti* were used in the experiments.

In the fourth experimental series, we examined whether the spectral preferences of the mosquito (*Ae. aegypti* [ROCK]) while controlling for the apparent contrast of the different hues. We normalized the perceptual contrasts of the visual objects by testing a range of greys of different contrasts with the background (Greys 1.5, 2.5, 4.0, 4.5, 6.5, and 9.5 [Weber Contrasts of -0.28 to -0.05]; Color-aid Corp., Greyset) in combination with, and against, the red object (R-Hue; Weber Contrast = -0.17). Once we identified the grey object that was as attractive as the red, identified by the PI that not significantly from 0 (t-test: $P > 0.05$), we then tested that grey (Grey 1.5; Weber Contrast = -0.28) against a range of different green objects that had the same peak wavelength (510 nm) but different Weber Contrasts, from light to dark (-0.11 to -0.27). Based on the green object that elicited the same level of attraction as the dark grey (PI = 0), we then tested the dark green versus the red object. Similar to experiments with the green objects, we tested the dark grey (Grey 1.5; Weber Contrast = -0.28) against a range of different blue objects that had the same peak wavelength (470 nm) but different Weber Contrasts, from light to dark (-0.13 to -0.28), followed by testing the red object versus the dark blue, and the attractive cyan (495 nm) object versus the dark blue or dark green objects with the same apparent contrast.

Visual Attraction to Skin in a Cage-Assay

To assay the visual attraction to the skin, acrylic cages (45 x 30 x 30 cm; McMaster-Carr; cat. # 8560K171) were constructed to allow for video recording and tracking from above. Thermal insulation and white sheeting were wrapped around the cage's exterior to prevent any heat cues from outside of the cage while providing a uniform visual environment to the interior. One side of the cage was partially made from a white mesh allowing 5% CO₂ input to the cage. To conduct the visual preference assays, two 4 x 4 cm windows, spaced 18 cm apart, were cut into the acrylic. Windows were sealed with heat absorptive glass (Schott KG2, Edmund Optics). Similar to the visual stimuli used in the wind tunnel experiments, mosquitoes were tested with a white "control" in one window (white glove), and the other window displaying either human skin, or human skin through long-wavelength optical filters (550-730 nm band; 36-333 and 35-894 filters; Edmund Optics Inc., Barrington, NJ USA). Positions of the visual stimuli were randomized between experimental replicates. To test for any side preference or contamination in the cage, control experiments were also conducted using two white gloves as the visual stimuli in the windows. Experiments were performed in a chamber held at approximately 20 to 22.5°C, and the cage was situated underneath a hood ventilation system allowing air exchange. A CO₂ Flypad (Genesee Scientific; cat. # 59-119) was placed immediately adjacent to the side of the cage, and similar to the wind tunnel experiments, CO₂ was controlled by two mass flow controllers (MC-200SCCM-D, Alicat

Scientific, Tucson, AZ) via a Python script that allowed synchronizing odor and filtered air delivery with the trajectory behaviors. Filtered air was released for the first 8 minutes of each experiment, followed by the release of 5% CO₂ for 8 minutes, before switching off for another 8 minutes (post-CO₂). Two cameras (Basler AC640gm, Exton, PA) were mounted above the cage and recorded mosquito trajectories at 100 frames/sec. IR backlights (HK-F3528IR30-X, LedLightsWorld, Bellevue, WA) were installed above the cage. Human skin reflectance measurements and assays were from 3 males and 3 female wild type individuals on the University of Washington (Seattle) campus (ages 25-46 years old). Volunteers were from various backgrounds: Hispanic (one male), white (one male and one female), and Asian (one male and two females). Protocols were reviewed and approved by the University of Washington Institutional Review Board, and all human volunteers gave their informed consent to participate in the research. As the wind tunnel experiments, we used 6—8 day-old, non-blood-fed, mated females who were sucrose deprived for 24 hours but had access to water. We released 50 *Ae. aegypti* (ROCK) females into the cage for each experiment, and the assays were initiated 3 hr before lights off (ZT12). Ambient CO₂ was constantly measured both inside and outside of the cage.

Visual preferences in *Cx. quinquefasciatus* and *An. stephensi*:

Anopheles stephensi (Indian strain) and *Culex quinquefasciatus* mosquitoes were separately raised in groups of 100 individuals and anesthetized with CO₂ to sort males from females after cohabitating for 7 days. At the time of their subjective sunset, groups of 50 female mosquitoes were released into the working section of the wind tunnel. After 1 h of filtered air, 5% CO₂ was released from a point source at the upwind section of the tunnel (height of 20 cm, and in the center of the tunnel) for 1 h, after which filtered air was released from the point source. The intensity of ambient light from the projectors was approximately 1.3 $\mu\text{W}/\text{cm}^2$. The low light intensity, relative to that used with *Ae. aegypti*, was necessary to recruit females to the visual objects. At higher light intensities *An. stephensi* and *Cx. quinquefasciatus* mosquitoes responded to the CO₂ plume but they did not respond to the visual objects. We found that these two species began to investigate the visual objects only at light intensities <5 lux. Like our experiments with *Ae. aegypti*, the temperature within the wind tunnel was approximately 22.5°C.

Trajectories analysis:

Our tracking system is unable to maintain mosquito identities for extended periods of time, but we considered individual trajectories as independent for the sake of statistical analysis. Analyses were restricted to trajectories that were at least 90 frames (1.5 seconds) long. Only trajectories that lasted for more than 1.5 seconds were analyzed (average length trajectory: 3.1 sec., longest trajectory: 96.4 sec., total number of 1,305,695). To examine the mosquito behaviors and preferences to the two visual stimuli in the tunnel, a fictive volume was created around the visual cues (area: 14x14 cm, height: 4 cm). The volume was centered over the object in the crosswind direction, and shifted slightly downwind in the wind line direction. This volume was chosen as it captures the area of primary activity of the mosquitoes. A sensitivity analysis was performed by adjusting the volume size and demonstrated that this volume best captured the mosquitoes investigating the visual objects while excluding mosquitoes transiting to other areas of the working section.

Occupancy maps were calculated by dividing the wind tunnel into 0.3-cm² squares. For each replicate experiment, the number of mosquito occurrences within each square was summed and divided by the total number of occurrences in all squares to yield a percentage of residency. We did not quantify landings on the spots due to limitations of the camera angles needed to identify landings. During the filtered “air” treatment, mosquitoes often investigated certain areas of the working section, such as the top or corners of the working section, causing hot spots in the occupancy maps. This is typical for mosquito activity without a stimulus. By contrast, when CO₂ is released, these hot spots are no longer apparent, and instead, the mosquitoes investigate the visual objects or navigate to the odor source, as demonstrated by a hot spot in the central area of the working section or near the odor source. For each mosquito line, the replicate trials were pooled to create a occupancy heatmap for the tested visual stimulus.

To calculate the fractions of trajectories that approached either visual object, for each trajectory we calculated a preference index by determining the amount of time a trajectory spent in each volume divided by the total time it spent in both volumes. If the trajectory spent all of its time in only one volume then it was assigned a preference index of 1 (test object) or -1 (white neutral object). Approximately 25 to 50% of the trajectories approached either object. From these preference indices, we calculated the global mean, and bootstrapped the 95% confidence interval of the mean through random resampling of the individual trajectories 500 times. To determine whether the mosquitoes preferred the visual objects compared to elsewhere in the tunnel, we calculated the preference index for each trajectory at each time point as the amount of time the mosquito spent in a particular 4x14x14 cm volume that was randomly selected in the tunnel and compared them to the volumes containing the visual objects. Mean flight velocities were calculated from the 3D tracks of each individual trajectory. To further examine whether mosquito responses to the visual objects changed throughout the experiment, the percent of time (per each

minute interval) the mosquitoes investigated the visual objects was calculated. Statistically significant groups were estimated using a Kruskal-Wallis, Mann-Whitney *U*-test with Bonferroni correction at a $P = 0.01$ level, or the one-sample *t*-test. All recorded data were analyzed using Matlab (Mathworks, 2019a release).

Electroretinogram (ERG) recordings

ERG recordings were performed by fixing 6 day-old, non-blood-fed female mosquitoes to a coverslip using Bondic glue. Mosquitoes were dark-adapted for 1 h prior to stimulation. The recording glass electrode (thin-wall glass capillaries; OD, 1.0 mm; length, 76 mm; World Precision Instruments, cat. # TW100F-3) was pulled using a micropipette puller (Sutter Instrument, p-2000), and filled with Ringer's solution (3 mM CaCl_2 , 182 mM KCl, 46 mM NaCl, 10 mM Tris pH 7.2). The reference electrode, a sharpened tungsten wire, was placed into one compound eye in a small drop of electrode gel (Parker, cat. # 17-05), and the recording electrode was placed immediately on the surface of the contralateral eye. Two different types of visual stimuli were presented to the mosquitoes. In the first series, the mosquitoes were placed at the center of a semi-cylindrical visual arena (frosted mylar, 10 cm diameter, 10 cm high); a video projector (Acer K132 WXGA DLP LED Projector, 600 Lumens) positioned in front of the arena projected the visual stimuli. To test the response to moving objects, similar to what the mosquito might encounter in flight, we tested responses to a 19° wide bar moving from left to right (Clockwise) (Fig. 4). The mosquito was randomly tested with blue, green, and red bars (distinct peaks at 455, 547 nm, and 633 nm, 18 lux), and each colored bar was tested 10-30 times per mosquito ($n = 7$ mosquitoes).

The second stimulation method used a digital monochromator to examine responses to different wavelengths across the mosquito visual spectra (350-750 nm). Mosquitoes were exposed to a 1-sec pulses of light (10 lux) from a light source (35-watt Halogen; ThorLabs) and a fiber optic scanning monochromator (MonoScan 2000, Mikropak GmbH, Ostfildern, Germany) that provided control of the transmitted wavelengths (± 2 nm). Light was transmitted via optical fibers (QP600-1-SR-B X, Ocean Optics, FL 32792, USA) and through a neutral density filter (fused silica, Thorlabs Inc., 0-1 OD). Each mosquito preparation was tested to wavelengths of 350-750 nm in 10 nm increments ($n=8$ mosquitoes/species). The visual stimuli were calibrated using a cosine-corrected spectrophotometer (HR-+2000, Ocean Optics, Dunedin, FL, USA) that was placed immediate to the recording preparation, allowing us to scale the irradiance of the tested stimuli. The light-induced responses were amplified by using an A-M Systems amplifier (10-100x; A-M Systems, 1800) and digitized using a Digidata data

acquisition system (Digidata 1550B, Molecular Devices, San Jose, CA 95134). Data were visualized and analyzed using Matlab software (Mathworks).

Linear Models

Linear models were created using R 4.0.3 and the lm function with the default option. Comparison between models were performed using the AIC function that calculates the Akaike's Information Criterion (AIC) for each model. For the first series of models, the dataset consisted of the mean preference index per experiment, the contrast value, peak wavelength and brightness value for the tested object. For the second series of models, the dataset contained the mean preference index per experiment and the area under the curve (AUC) of the reflectance measurement for the tested object calculated with bins of 25 nm from 350 to 675 nm. A Principal Component Analysis (PCA) was applied to the AUC vector to remove collinearity in the object's spectrum.

Data Availability

Data on the behavioral and electroretinogram experiments can be found on Mendeley Data. Software is available on <https://github.com/riffelllab>.

References

1. Briscoe, A.D. and L. Chittka, *The evolution of color vision in insects*. Annual review of entomology, 2001. **46**(1): p. 471-510.
2. Hannah, L., A.G. Dyer, J.E. Garcia, A. Dorin, and M. Burd, *Psychophysics of the hoverfly: categorical or continuous color discrimination?* Current zoology, 2019. **65**(4): p. 483-492.
3. van der Kooi, C.J., D.G. Stavenga, K. Arikawa, G. Belušič, and A. Kelber, *Evolution of insect color vision: From spectral sensitivity to visual ecology*. Annual Review of Entomology, 2021. **66**.
4. Clements, A.N., *The biology of mosquitoes. Volume 2: sensory reception and behaviour*. 1999: CABI publishing.
5. Green, C., *Effects of colours and synthetic odours on the attraction of Glossina pallidipes and G. morsitans morsitans to traps and screens*. Physiological Entomology, 1986. **11**(4): p. 411-421.
6. Torr, S. and G. Vale, *Know your foe: lessons from the analysis of tsetse fly behaviour*. Trends in Parasitology, 2015. **31**(3): p. 95-99.
7. Reisenman, C.E., A.N. Lorenzo Figueiras, M. Giurfa, and C.R. Lazzari, *Interaction of visual and olfactory cues in the aggregation behaviour of the haematophagous bug Triatoma infestans*. Journal of Comparative Physiology A, 2000. **186**(10): p. 961-968.

- 777 8. Bidlingmayer, W.L. and D.G. Hem, *Mosquito (Dipteral Culicidae) flight behaviour near*
778 *conspicuous objects*. Bulletin of Entomological Research, 2009. **69**(4): p. 691-700.
- 779 9. Kennedy, J.S., *The visual responses of flying mosquitoes*. Proceedings of the Zoological Society
780 of London, A, 1939. **109**: p. 221-242.
- 781 10. WL, B., *How mosquitoes see traps: role of visual responses*. Journal of the American Mosquito
782 Control Association, 1994. **10**: p. 272.
- 783 11. van Breugel, F., J.A. Riffell, A. Fairhall, and M.H. Dickinson, *Mosquitoes use vision to associate*
784 *odor plumes with thermal targets*. Current Biology, 2015. **25**(16): p. 2123-2129.
- 785 12. Kline, D.L. and G.F. Lemire, *Field evaluation of heat as an added attractant to traps baited with*
786 *carbon dioxide and octenol for Aedes taeniorhynchus*. Journal of the American Mosquito Control
787 Association-Mosquito News, 1995. **11**(4): p. 454-456.
- 788 13. Lacey, E. and R. Cardé, *Activation, orientation and landing of female Culex quinquefasciatus in*
789 *response to carbon dioxide and odour from human feet: 3-D flight analysis in a wind tunnel*.
790 Medical and Veterinary Entomology, 2011. **25**(1): p. 94-103.
- 791 14. McMeniman, C.J., R.A. Corfas, B.J. Matthews, S.A. Ritchie, and L.B. Vosshall, *Multimodal*
792 *integration of carbon dioxide and other sensory cues drives mosquito attraction to humans*. Cell,
793 2014. **156**(5): p. 1060-1071.
- 794 15. Fay, R. and W. Prince, *A trap based on visual responses of adult mosquitoes*. . Journal of the
795 American Mosquito Control Association-Mosquito News, 1968. **28**(1): p. 1-7.
- 796 16. Muir, L.E., B.H. Kay, and M.J. Thorne, *Aedes aegypti (Diptera: Culicidae) vision: response to*
797 *stimuli from the optical environment*. Journal of Medical Entomology, 1992. **29**(3): p. 445-450.
- 798 17. MR, S. and B. AWA, *Studies on the responses of the female Aedes mosquito. Part VII—the effect*
799 *of skin temperature, hue and moisture on the attractiveness of the human hand*. Bulletin of
800 Entomological Research, 1957. **47**: p. 89.
- 801 18. Snow, W., *The spectral sensitivity of Aedes aegypti (L.) at oviposition*. Bulletin of Entomological
802 Research, 1971. **60**(4): p. 683-696.
- 803 19. Brett, G., *On the relative attractiveness to Aedes aegypti of certain coloured cloths*. Transactions
804 of the Royal Society of Tropical Medicine and Hygiene, 1938. **32**(1): p. 113-124.
- 805 20. Muir, L.E., M.J. Thorne, and B.H. Kay, *Aedes aegypti (Diptera: Culicidae) vision: spectral*
806 *sensitivity and other perceptual parameters of the female eye*. Journal of Medical Entomology,
807 1992. **29**(2): p. 278-281.
- 808 21. Trpis, M., McClelland, G. A., Gillett, J. D., Teesdale, C. and T.R. Rao. *Diel periodicity in the*
809 *landing of Aedes aegypti on man*. Bulletin of the World Health Organization, 1973. **48**, 623-629.
- 810 22. Giraldo-Calderón, G.I., M.J. Zanis, and C.A. Hill, *Retention of duplicated long-wavelength*
811 *opsins in mosquito lineages by positive selection and differential expression*. BMC Evolutionary
812 Biology, 2017. **17**(1): p. 84.
- 813 23. Hu, X., J.H. England, A.C. Lani, J.J. Tung, N.J. Ward, S.M. Adams, K.A. Barber, M.A. Whaley,
814 and J.E. O'Tousa, *Patterned rhodopsin expression in R7 photoreceptors of mosquito retina:*
815 *Implications for species-specific behavior*. Journal of Comparative Neurology, 2009. **516**(4): p.
816 334-342.
- 817 24. Hu, X., M.A. Whaley, M.M. Stein, B.E. Mitchell, and J.E. O'Tousa, *Coexpression of spectrally*
818 *distinct rhodopsins in Aedes aegypti R7 photoreceptors*. PloS one, 2011. **6**(8): p. e23121.
- 819 25. Zhan, Y., D. Alonso San Alberto, C. Rusch, J. Riffell, and C. Montell, *Aedes aegypti vision-*
820 *guided target recognition behavior requires two redundant rhodopsins*. BioRxiv, 2020: p. 1-30.

- 821 26. Angelopoulou, E., 1999. *The reflectance spectrum of human skin*. Technical Reports (CIS),
822 p.584.
- 823 27. Lacey, E.S., A. Ray, and R.T. Carde, *Close encounters: contributions of carbon dioxide and*
824 *human skin odour to finding and landing on a host in Aedes aegypti*. Physiological Entomology,
825 2014. **39**(1): p. 60-68.
- 826 28. Vinauger, C., F. Van Breugel, L.T. Locke, K.K. Tobin, M.H. Dickinson, A.L. Fairhall, O.S.
827 Akbari, and J.A. Riffell, *Visual-olfactory integration in the human disease vector mosquito Aedes*
828 *aegypti*. Current Biology, 2019. **29**(15): p. 2509-2516. e5.
- 829 29. Nuttall, G.H. and A.E. Shipley, *Studies in relation to malaria. II.(Cont.) The structure and*
830 *biology of Anopheles (Anopheles maculipennis)*. Epidemiology & Infection, 1902. **2**(1): p. 58-84.
- 831 30. Packard, A., *Color-preference in insects*. Journal of the New York Entomological Society, 1903.
832 **11**(3): p. 132-137.
- 833 31. Schnaitmann, C., M. Pagni, and D.F. Reiff, *Color vision in insects: insights from Drosophila*.
834 Journal of Comparative Physiology A, 2020. **206**(2), p.183-198.
- 835 32. Sharkey, C.R., J. Blanco, M.M. Leibowitz, D. Pinto-Benito, and T.J. Wardill, *The spectral*
836 *sensitivity of Drosophila photoreceptors*. Scientific Reports, 2020. **10**(1), p.1-13.
- 837 33. Day, J.F., *Host-seeking strategies of mosquito disease vectors*. Journal of the American Mosquito
838 Control Association, 2005. **21**(sp1): p. 17-22.
- 839 34. Silver, J.B., *Designing a Mosquito Sampling Programme*. Mosquito Ecology: Field Sampling
840 Methods, 2008: p. 1-23.
- 841 35. Paulk, A.C., J. Phillips-Portillo, A.M. Dacks, J.-M. Fellous, and W. Gronenberg, *The processing*
842 *of color, motion, and stimulus timing are anatomically segregated in the bumblebee brain*.
843 Journal of Neuroscience, 2008. **28**(25): p. 6319-6332.
- 844 36. Swihart, S., *The neural basis of colour vision in the butterfly, Heliconius erato*. Journal of Insect
845 Physiology, 1972. **18**(5): p. 1015-1025.
- 846 37. Yonekura, T., J. Yamauchi, T. Morimoto, and Y. Seki, *Spectral response properties of higher*
847 *visual neurons in Drosophila melanogaster*. Journal of Comparative Physiology A, 2020. **206**(2):
848 p. 217-232.
- 849 38. Leming, M.T., *Light-dark and circadian effects on the visual response of Aedes aegypti*. 2015:
850 University of Notre Dame.
- 851 39. Hu, X., M.T. Leming, M.A. Whaley, and J.E. O'Tousa, *Rhodopsin coexpression in UV*
852 *photoreceptors of Aedes aegypti and Anopheles gambiae mosquitoes*. Journal of Experimental
853 Biology, 2014. **217**(6): p. 1003-1008.
- 854 40. Mazzoni, E.O., Celik, A., Wernet, M.F., Vasilaiuskas, D., Johnston, R.J., Cook, T.A., Pichaud,
855 F., and C. Desplan. Iroquois complex genes induce co-expression of rhodopsins in Drosophila.
856 PLoS biology, 2008 **6**(4): e97.
- 857 41. Peach, D.A., E. Ko, A.J. Blake, and G. Gries, *Ultraviolet inflorescence cues enhance*
858 *attractiveness of inflorescence odour to Culex pipiens mosquitoes*. PloS one, 2019. **14**(6): p.
859 e0217484.
- 860 42. Shaw, A.P., I. Tirados, C.T. Mangwiro, J. Esterhuizen, M.J. Lehane, S.J. Torr, and V. Kovacic,
861 *Costs of using "tiny targets" to control Glossina fuscipes fuscipes, a vector of gambiense*
862 *sleeping sickness in Arua District of Uganda*. PLoS Neglected Tropical Disease, 2015. **9**(3): p.
863 e0003624.

43. Vale, G., *The responses of tsetse flies (Diptera, Glossinidae) to mobile and stationary baits*. Bulletin of Entomological research, 1974. **64**(4): p. 545-588.
44. Stowers, J.R., M. Hofbauer, R. Bastien, J. Griessner, P. Higgins, S. Farooqui, R.M. Fischer, K. Nowikovsky, W. Haubensak, and I.D. Couzin, *Virtual reality for freely moving animals*. Nature methods, 2017. **14**(10): p. 995-1002.

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Author contributions

D.A.S.A., C.R., and J.A.R. designed research; D.A.S.A., C.R., and J.A.R. performed wind tunnel experiments; C.R. and J.A.R. conducted electroretinogram experiments; A.D.S. assisted in visual stimulus, wind tunnel software, and experimental designs; Y.Z. and C.M. generated the opsin mutant lines. All authors wrote and edited the paper.

Competing Interests

The authors declare no competing interests.

Supplementary Information Figures and Legends

Figure S1. Flight behaviors to different color stimuli, and characterization of stimuli in ERG experiments. (a) Flight velocities of mosquitoes in different CO₂ exposure treatments (AIR-, +CO₂, and post-CO₂), across all color stimuli. Exposure to CO₂ significantly elevated the flight velocities of mosquitoes (Kruskal-Wallis test with multiple comparisons: df=2, Chi-sq. = 597.23, $P < 0.001$), although there was no significant difference between AIR and post-CO₂ treatments (Kruskal-Wallis test with multiple comparisons: $P > 0.05$). Bars are the mean $\pm$ sem. (b) The flight velocities for each tested hue. There was no significant difference between hue treatment groups (Kruskal-Wallis test: df=11, Chi-sq. = 10.17, $P = 0.42$). Bars are the mean $\pm$ sem. (c) As in a, except for the mosquito flight durations. Exposure to CO₂ significantly the duration of the flight trajectories (Kruskal-Wallis test with multiple comparisons: df=2, Chi-sq. = 87.83, $P < 0.001$), although there was no significant difference between AIR and post-CO₂ treatments (Kruskal-Wallis test with multiple comparisons: $P > 0.05$). (d) As in b, except for the flight durations of each color stimulus. There was no significant difference between hue treatment groups (Kruskal-Wallis test: df=11, Chi-sq. = 16.82, $P = 0.16$). (e) The number of mosquitoes visiting the visual

objects over the duration of the experiment. Few mosquitoes investigated the visual objects before or after the CO₂ exposure (Air and post-CO₂, respectively), and there was no significant difference between those two time periods (Kruskal-Wallis test with multiple comparisons between AIR and post-CO₂: $P > 0.98$). However, exposure to CO₂ significantly increased the numbers of mosquitoes visiting the visual objects (Kruskal-Wallis test with multiple comparisons: $df=11$, Chi-sq. = 258.72, $P < 0.001$; $P < 0.001$). During CO₂, there were no significant differences in the number of mosquitoes investigating the different hues (Kruskal-Wallis test with multiple comparisons: $P > 0.99$). Lines are the means and shaded areas the $\pm$ sem. **(f)** The lux measurements of stimuli used in the ERG experiments. Experiments used either a short-throw projector (left) or a digital monochromator (right).

Figure S2. CO₂ and temperature measurements in the cage assay. (a) CO₂ measurements were taken outside (black trace) and inside (blue trace) the cage during experiments to measure if background contamination was occurring. CO₂ concentrations only increased in the cage during pre-programmed release from the mass flow controllers (grey shaded area). (b) Image taken from a FLIR camera (FLIR One Pro, FLIR Systems Inc., Goleta, CA USA) showed a constant temperature range, and did not show any variability including the region of the cage where the volunteer's skin was displayed through the window. The thermal shielding and IR absorptive windows in the cage prevented a radiant heat signature that is attractive mosquitoes.

Figure S3. Models on the contributions of contrast and hue wavelength on mosquito preferences.

(a) Linear model results based on contrast and peak wavelength values. (b) AIC of all the models significantly different from a null model and based on combinations of the contrast (C), peak wavelength (P) and brightness (R) values. The model in (a) was selected as the best having the lowest AIC values. (c) Linear regression parameters, standard errors, t-values and P-values for the model based on the PC2 and PC3. The PCs were obtained from the area-under-the-curve (AUC) of the reflectance curves calculated every 25 nm from 325 to 675 nm. (d) AIC of all the models significantly different from a null model and based on combinations of the first 4 PCs (representing more than 99% of the variance of the original dataset). The model in (c) was selected as the best having the lowest AIC values. (e) Correlation matrix between the AUC values from 325 to 675 nm and the 14 resulting PCs. The first three PCs correlated with different parts of the spectrum: PC1 was strongly positively correlated with the 575-675 nm range whereas PC2 and PC2 strongly negatively correlated with the 475-575 nm and 375-475 nm range respectively.