

1 **ABSTRACT**

2 The antibiotic linezolid, the first clinically approved member of the oxazolidinone class, inhibits
3 translation of bacterial ribosomes by binding to the peptidyl transferase center. Recent work has
4 demonstrated that linezolid does not inhibit peptide bond formation at all sequences but rather acts in a
5 context-specific manner, namely when alanine occupies the penultimate position of the nascent chain. In
6 this study, we determined that the second-generation oxazolidinone radezolid also induces stalling with
7 alanine at the penultimate position. However, the molecular basis for context-specificity of these
8 inhibitors has not been elucidated. In this study, we determined high-resolution cryo-EM structures of
9 both linezolid and radezolid-stalled ribosome complexes. These structures reveal that the alanine side
10 chain fits within a small hydrophobic crevice created by oxazolidinone, resulting in improved ribosome
11 binding. Modification of the ribosome by the antibiotic resistance enzyme Cfr disrupts stalling by
12 forcing the antibiotic to adopt a conformation that narrows the hydrophobic alanine pocket. Together,
13 the structural and biochemical findings presented in this work provide molecular understanding of
14 context-specific inhibition of translation by clinically important oxazolidinone antibiotics.

15 **KEYWORDS:** linezolid, radezolid, oxazolidinone antibiotics, peptidyl transferase center, cryo-EM,
16 stalled ribosome, nascent peptide, selective inhibition of translation, Cfr, m⁸A2503

1 INTRODUCTION

2 During translation, peptide bond formation occurs within the peptidyl transferase center (PTC) of the
3 ribosome. The PTC is located within the large ribosomal subunit and catalyzes extension of the
4 polypeptide chain through proper positioning of the peptidyl-tRNA in the P-site and aminoacyl tRNA in
5 the A-site. Due to its functional importance, the PTC of the bacterial ribosome is a common target for
6 antibiotics that inhibit translation^{1,2}.

7 The PTC antibiotic linezolid (LZD, **Fig. 1a**) was the first clinically approved member of the synthetic
8 oxazolidinone class of antibiotics³. Linezolid is used to treat drug-resistant gram-positive infections
9 including those caused by methicillin-resistant *S. aureus* and vancomycin-resistant Enterococci⁴. Initial
10 cross-linking experiments identified the binding site for linezolid within the PTC A-site only when
11 performed with actively translating ribosomes^{5,6}, suggesting that other translation components may be
12 involved in linezolid binding. However, existing structures of linezolid-bound ribosomes have been
13 obtained with the ribosomes devoid of charged tRNAs^{7,8} or ribosomes containing only a P-site tRNA
14 lacking a nascent chain⁹. Recent evidence obtained through ribosome profiling and single-molecule
15 studies demonstrated that linezolid does not indiscriminately inhibit the formation of every peptide bond
16 but rather interferes with translation at certain mRNA sites. Robust inhibition of translation and
17 ribosome stalling by linezolid is strongly favored when the amino acid alanine occupies the penultimate,
18 (-1), position within the nascent chain^{10,11}. Together, these results suggest that interactions between
19 linezolid and the nascent peptide may be important for stabilizing antibiotic binding to the ribosome.
20 However, the exact nature of these interactions is yet to be elucidated.

21 The clinical success of LZD and emergence of resistance to this antibiotic through modification of the
22 ribosome by chloramphenicol-florfenicol resistance enzyme Cfr sparked the development of
23 second-generation derivatives, such as radezolid (RZD), with improved potency¹². Radezolid (**Fig. 1a**) is
24 in clinical development for bacterial acne and community-acquired pneumonia¹³. Compared to linezolid,
25 radezolid retains the aryl-oxazolidinone core (rings A and B) and C5 group but has alterations to the
26 C/D ring system. Given that key chemical elements are conserved between LZD and RZD it is plausible

1 that RZD can also act as a context-specific inhibitor of translation. The importance of both conserved
2 and distinct structural features of second generation oxazolidinones to translation inhibition and context
3 specificity, if any, is yet to be elucidated.

4 In this work, we discovered that RZD exhibits ribosome stalling behavior similar to that of LZD,
5 arresting translating ribosomes when alanine occupies the penultimate position within the nascent
6 peptide chain. Capitalizing on the stalling preference of these two antibiotics, we generated high
7 resolution cryo-EM structures of LZD- and RZD-stalled ribosome complexes. Direct comparison of the
8 drug-stalled translation complexes with the structures of the vacant ribosomes bound to the same
9 antibiotics enabled identification of molecular contacts that improve oxazolidinone binding to the
10 ribosome. Specifically, we find that the penultimate alanine fits snugly within a shallow hydrophobic
11 pocket created by the oxazolidinone molecules, providing structural rationale for context specificity. Our
12 analysis of RZD action on ribosomes modified by the oxazolidinone resistance enzyme Cfr and the
13 structure of RZD bound to the Cfr-modified ribosome provides the first insights into how this second
14 generation oxazolidinone interacts with a LZD-resistant ribosome.

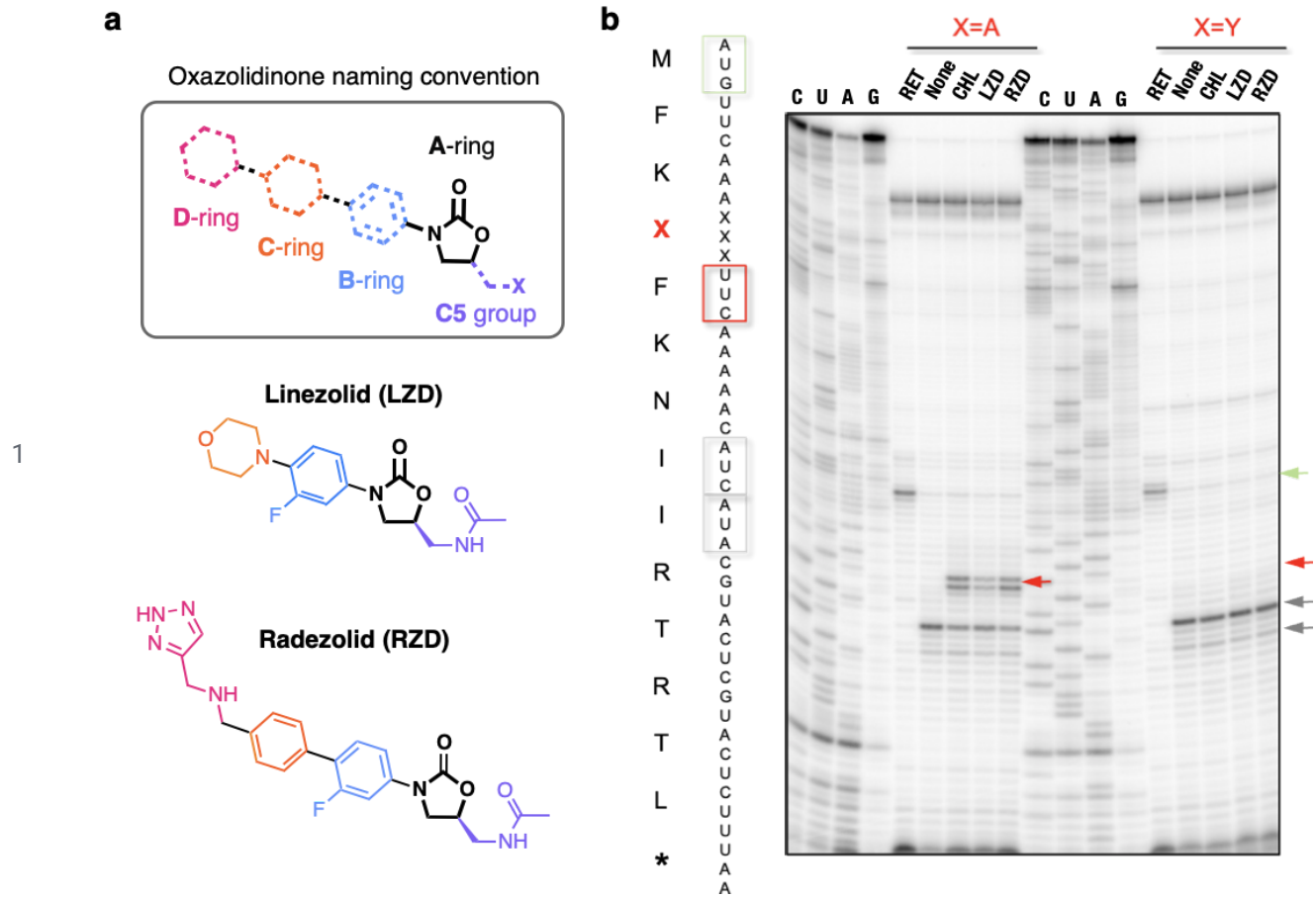
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1 RESULTS

2 Linezolid and radezolid have similar ribosome stalling behavior

3 Previous *in vitro* toe-printing experiments have demonstrated that LZD induces ribosome stalling on a
4 model mRNA sequence when the amino acid alanine is located in the penultimate (-1) position of the
5 nascent polypeptide chain^{10,11} (**Fig. 1b**). LZD-induced ribosome stalling is abolished when the
6 penultimate alanine is replaced with tyrosine (**Fig. 1b**). To evaluate if the second generation
7 oxazolidinone RZD shows similar stalling behavior, we carried out *in vitro* toeprinting analysis to
8 monitor the position of stalled ribosomes on the previously described mRNAs encoding the following
9 peptide sequences: MFKAFKNIIRTRL and MFKYFKNIIRTRL¹¹.

10 Similarly to LZD, the presence of RZD permits formation of the first peptide bond with both mRNA
11 transcripts as templates (**Fig. 1b**). This result indicates that RZD is not a universal inhibitor of
12 translation and also does not act as initiation inhibitor as suggested for LZD by some earlier studies¹⁴.
13 While no inhibition of translation at the early mRNA codons was observed on MFKYFK-encoding
14 template, presence of RZD or LZD led to selective stalling of the ribosome during the translation of
15 MFKAFK-encoding template. The stalling occurred at the F5 codon when an alanine residue appeared
16 in the penultimate position within the nascent peptide chain. These results indicate that RZD- or LZD-
17 bound ribosomes are unable to catalyze peptide bond formation between F5 and K6 when the MFKAF
18 nascent peptide occupies the exit tunnel. Comparison of the relative intensity of the stalled ribosome
19 toeprint bands suggests that RZD is a stronger inducer of ribosome stalling than LZD. The observation
20 that RZD is unable to stall the ribosome if the critical alanine residue is replaced with a tyrosine
21 (MFKYF) is consistent with the specificity of action of LZD (**Fig. 1b**).



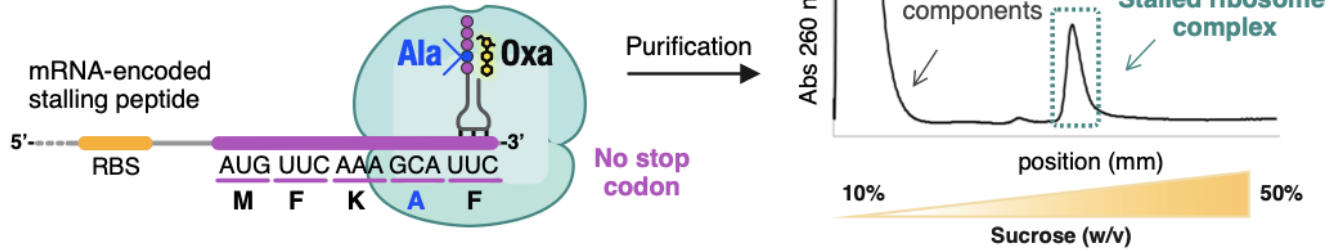
1 **Generation of stalled ribosomes complexes and cryo-EM analysis**

2 Guided by the *in vitro* stalling behavior, we designed a stalling peptide to capture LZD and RZD stalled
3 ribosome complexes (SRC) for structural analysis, herein referred to as LZD-SRC and RZD-SRC,
4 respectively. Stalled complexes were generated by conducting coupled *in vitro* transcription-translation
5 reactions with *E. coli* ribosomes in the presence of oxazolidinone antibiotics (**Fig. 2a**). To bias formation
6 of ribosomes with nascent peptide stalled at the F5 codon, we designed the DNA template where the
7 open reading frame encoding the MFKAF stalling peptide lacked a stop codon (**Fig. 2a, Supplementary**
8 **Fig. 1**). Stalled 70S ribosome complexes were purified away from other components of the translation
9 reaction by sucrose gradient fractionation and vitrified on carbon grids for cryo-EM analysis.
10 Refinement and reconstruction was performed using the cisTEM software suite¹⁶ to obtain 2.5 Å
11 resolution structure of LZD-SRC and 2.5 Å resolution of RZD-SRC (**Fig. 2b, Supplementary Fig. 2,**
12 **Supplementary Table 1**).

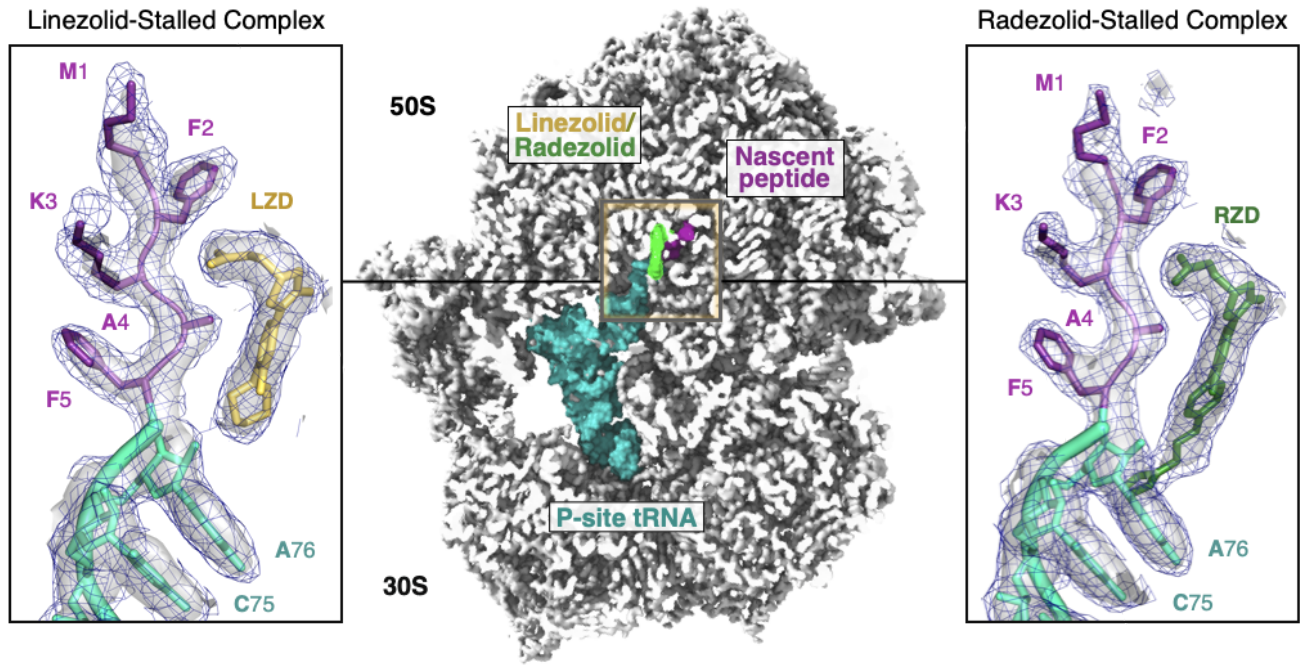
13 The generated cryo-EM maps of LZD-SRC and RZD-SRC have well-defined densities for the
14 oxazolidinone antibiotic and rRNA nucleotides, especially those within the PTC. Both cryo-EM maps
15 also have well defined densities for the peptidyl-tRNA located in the P-site. No A-site or E-site tRNAs
16 are present. Modeling of the density maps unambiguously assigns the penultimate residue in the nascent
17 peptide as alanine, as expected based on the DNA template used in the experiment (**Fig. 2b,**
18 **Supplementary Fig. 2**). This assignment is further supported by cryo-EM densities corresponding to
19 mRNA:tRNA interaction. The codon-anticodon densities are best modeled by the interaction between
20 UUC:GAA-tRNA^{Phe} rather than GCA:UGC-tRNA^{Ala} (**Supplementary Fig. 3**).

21 To directly compare how presence of the nascent peptide may influence positioning of the antibiotic
22 and/or conformation of rRNA nucleotides, we also generated structures of vacant *E. coli* 50S ribosomal
23 subunits in complex with LZD or RZD alone, herein referred to as LZD-50S and RZD-50S, at 2.4 Å
24 resolution and 2.5 Å resolution, respectively (**Supplementary Fig. 2, Supplementary Table 1**).
25 Similarly to the stalled complexes, antibiotic-only bound structures have unambiguous densities for the
26 oxazolidinone antibiotic.

a *in vitro* translation reaction:



b



2 **Figure 2. Cryo-EM structures of linezolid and radezolid-stalled ribosome complexes.** (a) Stalled
3 complexes were generated by performing coupled *in vitro* transcription-translation reactions in the
4 presence of the oxazolidinone (Oxa) antibiotic linezolid or radezolid. Complexes were further purified
5 by sucrose gradient fractionation. (b) Cross-section of the cryo-EM density map of the 70S ribosome in
6 complex with peptidyl-tRNA and oxazolidinone. Inserts are close-up views of the linezolid (LZD) or
7 radezolid (RZD) in complex with the MFKAF nascent peptide. Coulomb potential density is contoured
8 at 4.0σ in surface representation and 1.0σ in mesh representation. Figure was made using unsharpened
9 maps.

1 **The penultimate alanine facilitates oxazolidinone antibiotic binding to the ribosome**

2 The overall binding modes of LZD and RZD within the PTC in antibiotic-only bound and stalled
3 ribosome complexes are similar to those described previously^{7-9,17}. The fluorophenyl moiety (B-ring) sits
4 in the A-site cleft, a hydrophobic pocket formed by splayed out nitrogen bases of nucleotides C2452 and
5 A2451 (**Supplementary Fig. 4a,b**). The oxazolidinone ring (A-ring) of LZD and RZD is positioned in
6 an offset π - π stacking interaction with Ψ 2504. Interestingly, we find that the carbonyl of the
7 oxazolidinone ring does not interact with rRNA but is rather chelated to what is likely a Mg^{2+} ion based
8 on the coordination geometry (**Supplementary Fig. 4c,d**), corroborating previous SAR studies that
9 demonstrated the importance of an electron-pair donor for activity¹⁸.

10 The binding poses for both LZD and RZD are near-identical between the antibiotic-only and
11 stalled-ribosome structures (**Supplementary Fig. 5**). We do, however, observe improved density for
12 both antibiotics, suggesting that presence of the stalling nascent peptide stabilizes the placement of LZD
13 and RZD in the ribosome (**Fig. 3a,b**). Specifically, we observe improved density for the C5 acetamide
14 group for both antibiotics, as well as enhanced density for the D-ring of RZD in the stalled structures. Of
15 note, in previously published density maps^{7-9,17}, the acetamide is less well resolved and has been
16 modeled in a variety of positions, likely due to its ability to sample multiple conformations. Our
17 structures suggest that the nascent peptide present in the stalled ribosome complex stabilizes the C5
18 group thereby providing a more biologically relevant view on the drug placement.

19 The nascent protein chain, and more specifically, the penultimate Ala residue, contributes directly to
20 formation of the drug binding site (**Fig. 2b, Fig. 3c,d**). In contrast to the orientation of the flanking
21 residues within the nascent peptide, the side chain of the penultimate alanine faces towards the
22 oxazolidinone binding pocket. In this orientation, the methyl group of alanine fits snugly within the
23 hydrophobic crevice formed by the C5 group and A/B-ring system of the antibiotic. The crevice is deep
24 enough to accommodate the small side chain of alanine, but is too shallow to fit bulkier side chains,
25 including that of tyrosine, providing rationale for differential stalling preferences observed in our
26 toeprinting experiments (**Fig. 1b**). In the stalled complex, the alanine's side chain methyl group engages

1 in a CH- π interaction with the aryl B-ring of the oxazolidinone (3.6 Å and 3.9 Å between C atom and
2 plane of the B-ring for LZD-SRC and RZD-SRC, respectively) (**Fig. 3c,d**). This interaction likely
3 facilitates antibiotic binding to the A-site of the PTC, resulting in enhanced occlusion of incoming
4 aminoacyl tRNAs. Glycine, the smallest amino acid residue and which lacks a side chain, does not clash
5 with the ribosome-bound antibiotic when present in the penultimate position of the nascent peptide, yet
6 it is not favored for drug-induced ribosome stalling^{10,11}. The inability of glycine to form the
7 alanine-specific CH- π interaction with the drug molecule likely explains this observation.

8 A similar CH- π interaction has been recently observed between the penultimate alanine and aryl ring of
9 chloramphenicol (CHL), another PTC-targeting antibiotic that also exhibits context-specific inhibition
10 of translation¹⁹. In contrast to CHL, which also induces robust ribosome stalling when serine or
11 threonine occupy the penultimate position, LZD exhibits a strong preference for alanine¹⁰. While *in*
12 *silico* modeling of serine or threonine at the penultimate position in LZD-SRC reveals that serine can be
13 accommodated, the methyl group of threonine generates a steric clash for all favored rotamers
14 (**Supplementary Fig. 6**). Recent work hypothesized that CHL-induced stalling with serine in the
15 penultimate position of the nascent peptide is stabilized by a H-bond between the serine hydroxyl and
16 chlorine atom of CHL¹⁹. Since the C5 group of LZD does not have an analogous electron-pair donor, it
17 is likely that the unsatisfied H-bond acceptor, rather than sterics, accounts for LZD's strong preference
18 for alanine over serine.

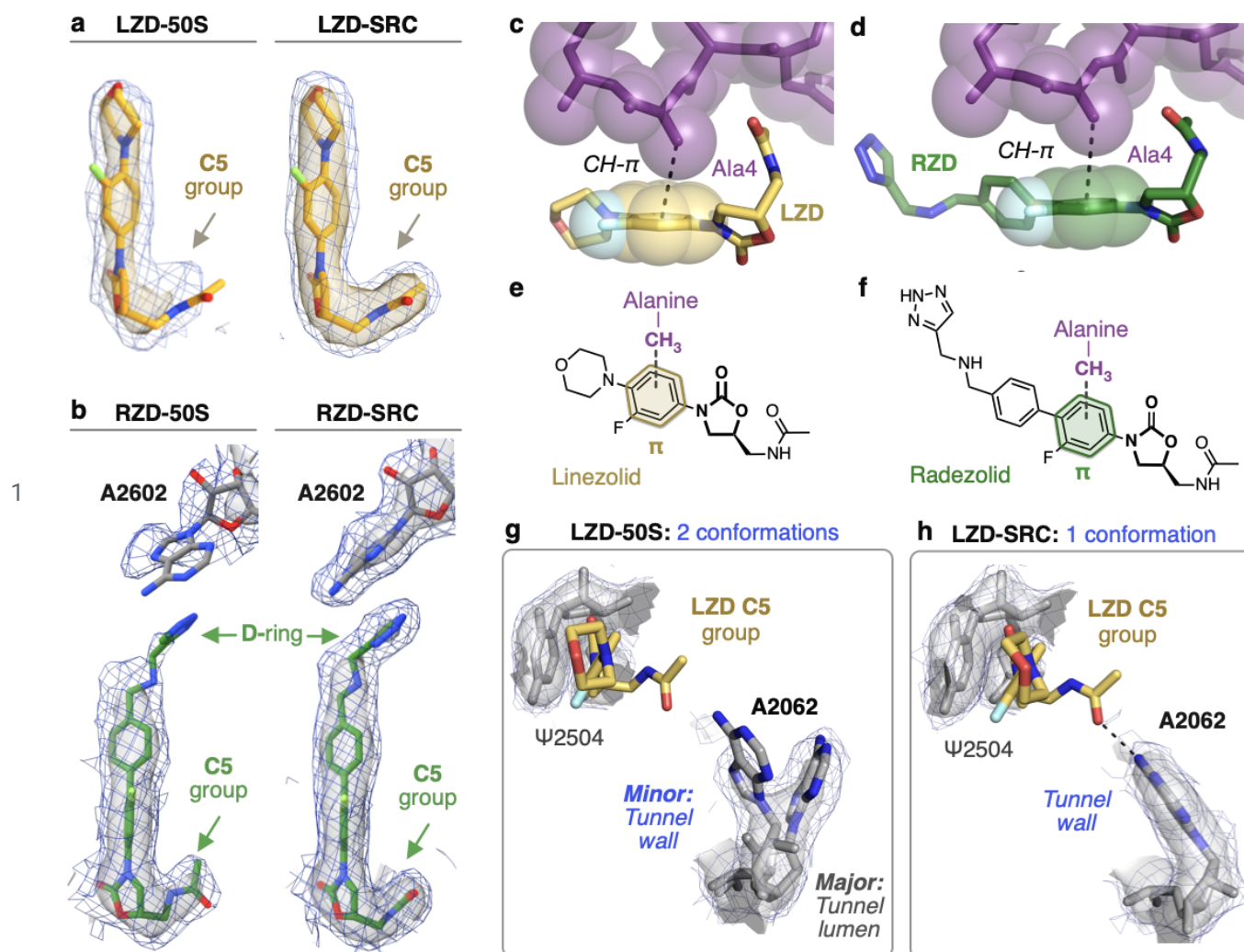
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20 **Dynamic nucleotides are stabilized to provide additional interactions with the oxazolidinone** 21 **antibiotics**

22 In LZD and RZD antibiotic-only 50S structures, the exit tunnel nucleotide A2062 can adopt two distinct
23 conformations (**Fig. 3g**). In its dominant conformation the A2062 base projects into the lumen of the
24 exit tunnel, while in the minor conformation, A2062 is rotated and lays flat against the tunnel wall.
25 Strikingly, in the stalled ribosome complexes with the nascent protein chain occupying the tunnel, the
26 A2062 base is stabilized in the rotated state juxtaposed against the tunnel wall (**Fig. 3h, Supplementary**

1 **Fig. 7a**). The rotated state of A2062 is stabilized by a H-bond between the N1 atom of adenine and the
2 stalling peptide backbone, as well as a non-canonical A:A base-pair with m²A2503 (**Supplementary**
3 **Fig. 7b**). In this conformation, the exocyclic amine of A2062 is within H-bonding distance of the
4 acetamide carbonyl of LZD/RZD (2.7 Å/3.3 Å), which likely explains why we observe improved
5 density for the C5 group in the stalled complexes (**Fig. 3a,b**). This interaction has not been observed in
6 existing structures of oxazolidinone-bound vacant ribosomes due to the alternative orientation of A2062.
7 The peptide-induced interaction between A2062 and the oxazolidinone is distinct from that observed
8 with CHL. In structures of chloramphenicol without nascent peptide, A2062 is already in a rotated state
9 to form a H-bond with the antibiotic²⁰.

10 Compared to the antibiotic-only bound structures, we also observe stabilization of dynamic PTC
11 nucleotides in conformations that provide additional contacts with the antibiotics. Notably, the global
12 resolutions of the maps are similar and many residues show nearly identical fits to density at the same
13 map threshold (**Supplementary Fig. 8**). We observe improved density for U2585, which as the C4 enol
14 tautomer could provide a H-bonding interaction with the oxygen atom of the morpholine ring in LZD
15 (**Supplementary Fig. 7, 8**). Although U2506 does not make direct contact with the LZD, we observe
16 dramatically improved density for this nucleotide in a conformation analogous to an uninduced or
17 nonproductive state of the PTC²¹ (**Supplementary Fig. 7c, 8**). In the RZD-stalled complex, we observe
18 improved densities for U2506, U2585 and A2602 (**Supplementary Fig. 8**). Nucleotides U2506 and
19 A2602 provide π - π stacking interactions with the C- and D-ring, respectively, while nucleotide U2585
20 engages in a H-bond with the secondary amine of RZD (**Fig. 3b, Supplementary Fig. 7**). Interestingly,
21 the D-ring interaction with A2602 has not been observed in a previous ribosome structure with RZD¹⁷
22 but likely explains why RZD is a better inhibitor of translation compared to LZD, as the stabilized
23 D-ring would provide additional steric interference with aminoacyl tRNA binding (**Supplementary Fig.**
24 **9**). Together these results suggest that, in addition to favorable interactions with the penultimate alanine,
25 interactions between rRNA nucleotides and the oxazolidinone likely play a role in improving antibiotic
26 binding to the ribosome to facilitate stalling.



2 **Figure 3. Stalling peptide stabilizes oxazolidinone binding to the ribosome.** For this figure, coulomb
3 potential density is contoured at 4.0σ in surface representation and 1.0σ in mesh representation from
4 unsharpened maps generated with a soft 50S mask. **(a)** Comparison of linezolid density in the
5 linezolid-only bound (LLD-50S) and the linezolid-stalled complex (LLD-SRC). **(b)** Comparison of
6 radezolid density in the radezolid-only bound (RLD-50S) and the radezolid-stalled complex
7 (RLD-SRC). **(c)** Close up view of the CH- π interaction between the B-ring of LLD and the penultimate
8 alanine (Ala4). **(d)** Close up view of the CH- π interaction between the RLD B-ring and the penultimate
9 alanine. **(e,f)** Schematic of the CH- π interaction involved in stabilizing antibiotic binding in **(e)**
10 LLD-stalled and **(f)** RLD-stalled ribosome complexes. **(g)** Density of the exit tunnel rRNA nucleotide
11 A2062 in LLD-only bound structure, highlighting two conformations. **(h)** Density of A2062 in the
12 LLD-stalled structure, with only one observed conformation.

1 Cfr methylation destabilizes the stalled ribosome complex

2 A prevalent resistance mechanism to LZD identified in multiple clinical isolates worldwide^{22–28} involves
3 methylation of rRNA by the Cfr enzyme, which adds a methyl group at the C8 atom of A2503
4 (m⁸A2503) in 23S rRNA^{29–33}. The Cfr modification disrupts LZD binding to the ribosome by
5 introducing a steric clash between the installed methyl mark and the C5 group of the antibiotic. While
6 this modification confers high levels of resistance to LZD, RZD retains modest efficacy against
7 Cfr-positive strains^{34,35}, likely due to retained interactions on the other side of the molecule involving the
8 D-ring (**Fig. 3b**). However, this suggests that when RZD binds to Cfr-modified ribosomes, the C5 group
9 must adopt an alternative conformation to accommodate C8 methyl group of m⁸A2503. Given that the
10 C5 group is an important component of the antibiotic pocket required to fit the alanine side chain, we
11 wanted to evaluate the ability of RZD to induce stalling of Cfr-modified ribosomes.

12 To perform *in vitro* assays, we expressed Cfr in *E. coli* and isolated ribosomes with near-complete
13 methylation of m⁸A2503 as described previously³³. As expected, RZD retains activity against the
14 m⁸A2503 ribosomes. *In vitro* translation of the sf-GFP by Cfr-modified ribosomes could not be
15 suppressed by LZD but was readily inhibited by RZD (IC₅₀ ~1 μM (**Supplementary Fig. 10a**)). As
16 expected, in toe-printing experiments the m⁸A2503 modification alleviated LZD-induced ribosome
17 arrest at the F5 codon of the MFKAF...-encoding ORF (**Fig. 4a**). Strikingly, RZD, while retaining its
18 general translation inhibitory activity, also failed to arrest the m⁸A2503 ribosome at the F5 codon of the
19 MFKAF template (**Fig. 4a**). Together, these results suggest that the presence of C8 methyl group at
20 A2503 does not prevent antibiotic binding but alters its ability to arrest translation in the penultimate
21 alanine-specific manner.

22 In search of a molecular explanation for this result, we determined the structure of RZD-bound to
23 Cfr-modified ribosome ‘forced-stalled’ with the MFKAF-tRNA in the P-site at 2.4 Å resolution. While
24 the overall binding of the antibiotic molecule was similar to that observed in the stalled *wildtype*
25 ribosome, the C5 group of RZD bound to the m⁸A2503 ribosome adopts a conformation distinct from
26 that of the *wildtype* complex (**Fig. 4b**). In this alternative conformation, the C5 group encroaches upon

1 the binding pocket of the penultimate alanine to introduce a mild steric clash (**Fig. 4c**). As a result, the
2 oxazolidinone cavity becomes a poor fit for the penultimate alanine of the nascent protein chain.
3 Destabilization of a properly stalled complex is exhibited by poor density for certain side chains of the
4 nascent peptide (**Supplementary Fig. 10b**). We also observe reversion of exit tunnel nucleotide A2062
5 to its lumen conformation which is now positioned too far away to engage in a H-bond with RZD. These
6 results suggest that the m⁸A2503 modification alters the RZD stalling landscape by changing the
7 interface between the antibiotic and the nascent peptide.

8 Our structural findings also provide important insights into how RZD retains efficacy against
9 LZD-resistant ribosomes. While the Cfr modification perturbs positioning of the C5 group, which likely
10 disrupts interaction with exit tunnel nucleotide A2062, we observe retention of other interactions.
11 Although somewhat diminished, we observe densities for the D-ring and A2602 engaged in a π - π
12 stacking interaction analogous to that observed in the *wildtype*, non-Cfr modified complex (**Fig. 4d**).
13 LZD does not contain the additional D-ring and is thus more dramatically impacted by lost interactions
14 due to m⁸A2503. Our results suggest that in addition to shortening of the C5 group, which has been
15 carried out with other oxazolidinones derivatives^{13,36}, extension of the ring system on the opposite end of
16 the molecule is an orthogonal, viable strategy for generating oxazolidinone antibiotics that overcome Cfr
17 resistance.

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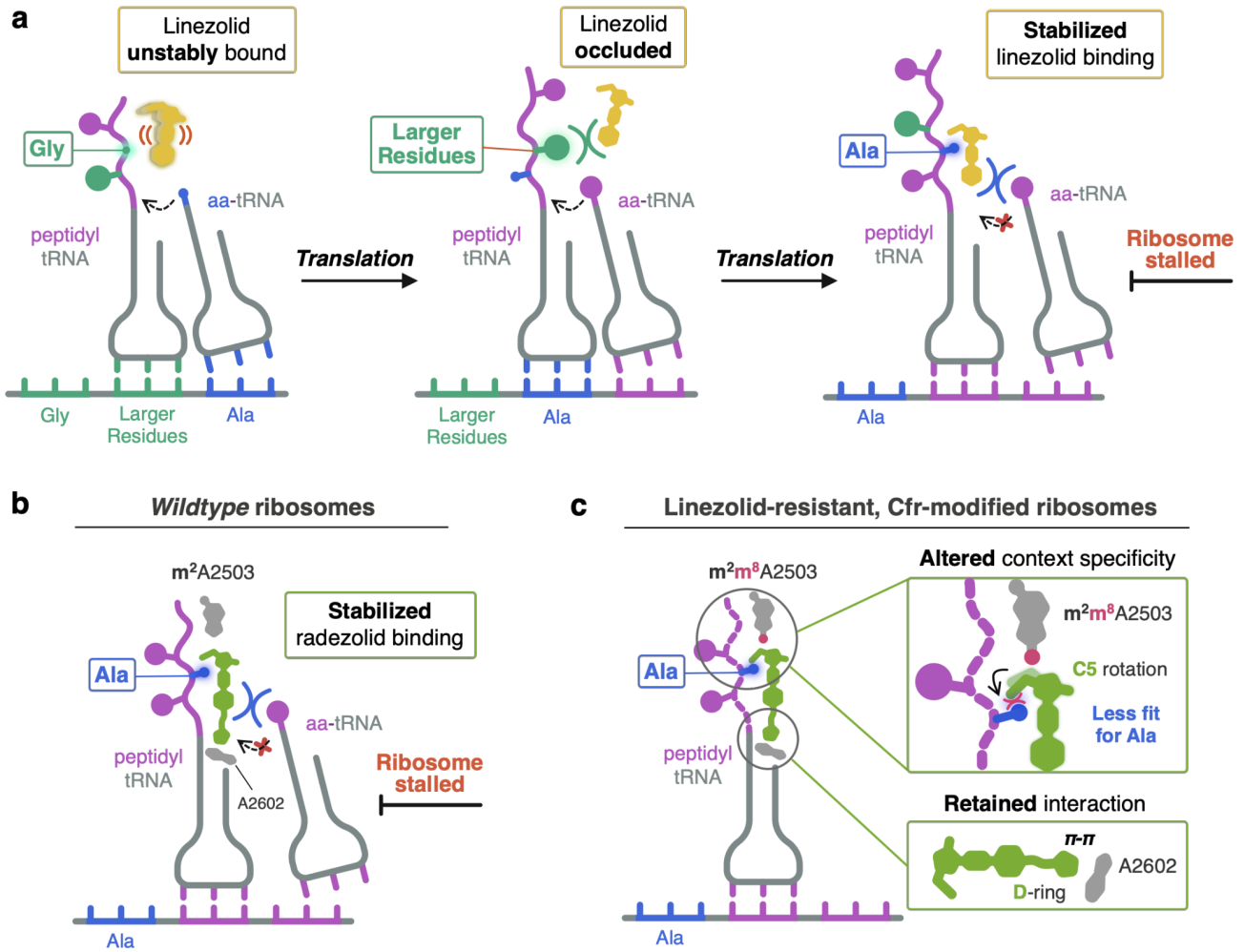
1 DISCUSSION

2 We identified two factors that contribute to context specificity of oxazolidinones LZD and RZD (**Fig. 5**).
3 While the antibiotic can still bind to the ribosome that lacks the nascent protein chain, as revealed by our
4 and previously published structures of the vacant ribosome complexed with LZD and RZD, the presence
5 of the nascent chain stabilizes binding of the drug by providing additional points of contact. Most
6 importantly, the side chain of the penultimate alanine residue is intercalated into the
7 complementary-shaped cavity formed by the drug molecule (**Fig. 2b, Fig. 3c,d**). Larger amino acids in
8 the penultimate position of the nascent peptide would clash with the antibiotic, preventing its binding. In
9 contrast, glycine cannot make a CH- π interaction with the drug, thereby making its binding less
10 favorable. As a secondary effect of the alanine interaction, we also observe stabilization of dynamic
11 nucleotides in conformations that provide additional contacts with the antibiotic (**Supplementary Fig.**
12 **7**). The culmination of these interactions leads to improved antibiotic binding to the ribosomal A-site in
13 the presence of nascent peptides containing a penultimate Ala, facilitating competition with incoming
14 aminoacyl tRNAs and thus resulting in ribosome stalling.

15 Our structural investigations into RZD binding to a Cfr-modified ribosome revealed the importance of
16 the D-ring in stabilizing ribosome engagement, providing unprecedented insight into how RZD retains
17 efficacy against Cfr modified ribosomes. Surprisingly, we found that while RZD retains inhibitory
18 activity against Cfr-modified ribosomes, the m⁸A2503 modification disrupts RZD stalling on the tested
19 alanine-containing peptide. Our structural findings suggest the Cfr modification likely changes the
20 context specificity of RZD by remodeling the hydrophobic pocket engaged by alanine in complexes
21 with *wildtype* ribosomes. In summary, our findings provide a unifying model for context-specific
22 inhibition of translation by oxazolidinone antibiotics. Our observation that the antibiotic binding pocket
23 is formed, in part, by the nascent peptide, has revealed the “missing” component of translation
24 machinery involved in antibiotic binding from previous cross-linking experiments. The importance of
25 Ala in the penultimate position of the nascent peptide for stabilizing the antibiotic provides the structural
26 basis for context specificity observed in ribosome profiling and single molecule studies. Our structural

1 insights into nascent-peptide specific inhibition of translation by oxazolidinones, and complementary
2 work in related systems^{19,37-39}, suggests prospects for the development of drugs that can modulate
3 activity of the ribosomes in a protein-selective manner.

4



2 **Figure 5. Model for oxazolidinone context-specific inhibition of translation.** (a) Linezolid (yellow)
3 is either unstably bound or sterically occluded from binding to the ribosome when glycine (Gly) or
4 larger residues occupy the penultimate position within the nascent chain. When alanine (Ala, blue)
5 occupies the penultimate position, linezolid becomes stably bound to the ribosomal PTC A-site,
6 enabling linezolid to better compete away incoming aminoacyl tRNAs (aa-tRNAs) and resulting in
7 ribosome stalling. (b) Radezolid (green) also facilitates stalling of ribosomes with alanine in the
8 penultimate position by enhancing radezolid binding to the PTC A-site. (c) Radezolid context-specificity
9 is altered in Cfr-modified ribosomes due to repositioning of the C5 group required to accommodate the
10 m⁸A2503 modification (pink circle). Rotation of the radezolid C5 group narrows the antibiotic pocket
11 that normally houses the alanine side chain. Retained interaction between radezolid D-ring and A2602
12 (grey) explains why RZD can overcome Cfr-mediated resistance.

1 **DATA AVAILABILITY**

2 Atomic coordinates for all presented structures have been deposited in the Protein Data Bank and
3 EMDB under the following accession numbers:

4 LZD-50S	PDB ID: 7RKQ	EMDB: 24503
5 LZD-SRC	PDB ID: 7RJG	EMDB: 24487
6 RZD-50S	PDB ID: 7RKO	EMDB: 24502
7 RZD-SRC	PDB ID: 7RJH	EMDB: 24488
8 RZD-SRC*	PDB ID: 7RKJ	EMDB: 24498

9

10 **CONFLICTS OF INTEREST**

11 The authors would like to report no conflicts of interest.

12

13 **ACKNOWLEDGEMENTS**

14 We thank D. Bulkley and G. Gilbert for technical support at the UCSF Center for Advanced CryoEM,
15 which is supported by the National Institutes of Health (S10OD020054 and S10OD021741) and the
16 Howard Hughes Medical Institute (HHMI). Some of this work was performed at the Stanford-SLAC
17 Cryo-EM Center (S²C²), which is supported by the National Institutes of Health Common Fund
18 Transformative High-Resolution Cryo-Electron Microscopy program (U24 GM129541). The content is
19 solely the responsibility of the authors and does not necessarily represent the official views of the
20 National Institutes of Health. We thank E. Eng, E. Kopylov, and the rest of the staff for technical support
21 at the National Center for CryoEM Access and Training (NCCAT) and the Simons Electron Microscopy
22 Center located at the New York Structural Biology Center, which is supported by the NIH Common
23 Fund Transformative High Resolution Cryo-Electron Microscopy program (U24 GM129539) and by
24 grants from the Simons Foundation (SF349247) and NY State. We acknowledge support from NIAID
25 (R01AI137270 to D.G.F and F32AI148120 to D.J.L.), W.M. Keck Foundation Medical Research Grant
26 (to J.S.F. and D.G.F.), a Sanghvi-Agarwal Innovation Award (J.S.F.), NSF GRFP (1650113 to K.T.), and
27 the UCSF Discovery Fellowship (to K.T).

28

29 **AUTHOR CONTRIBUTIONS**

30 K.T. performed structural analysis, assisted with model refinement, prepared figures, and wrote the
31 manuscript. V.S. prepared ribosome samples, performed model refinement, and edited the manuscript.
32 D.J.L. performed cryo-EM analysis, performed model refinement, prepared figures, and edited the
33 manuscript. I.D.Y. performed structural analysis, prepared figures, and assisted with model refinement.
34 T.S. performed *in vitro* translation experiments. N.V.L. and A.M. provided data interpretation and edited
35 the manuscript. J.S.F. and D.G.F. conceived and supervised the research, assisted in data interpretation,
36 and edited the manuscript.

1 **METHODS**

2

3 **Generation of DNA templates for PURExpress system**

4 PCR reactions were conducted with AccuPrime Taq DNA Polymerase (Thermo Fisher). DNA templates
5 used for toeprinting analysis were prepared as described previously¹¹. DNA templates used for the
6 generation of stalled ribosome complexes were prepared using by combining the following primers: 100
7 μ M T7, 100 μ M ORF_SD, 10 μ M T7_MFKAF_Fwd, and 10 μ M SD_MFKAF_Rev. Primer sequences
8 are listed in **Supplementary Table 2**. The PCR product was purified using the MiniElute PCR kit
9 (Qiagen) following manufacturer's instructions and quality was confirmed using a 8% TBE (Novagen)
10 gel. Sequence architecture of the resulting DNA product is outlined in **Supplementary Fig. 2**.

11

12 **Purification of 70S ribosomes**

13 *Wildtype E. coli* 70S ribosomes were purified from the MRE600 strain as described previously⁴⁰.
14 Cfr-modified 70S ribosomes were prepared using previously published protocol with modification³³. In
15 short, *E. coli* BW25113 expressing the evolved variant CfrV7 were grown to an OD₆₀₀ of ~0.7 in LB
16 media containing ampicillin (100 μ g/mL) and AHT inducer (30 ng/mL) at 37°C. After lysis using a
17 microfluidizer, clarified lysates were applied to a 32% w/v sucrose cushion. Tight-coupled 70S
18 ribosomes were purified on a 15-30% w/v sucrose gradient. To eliminate sucrose, ribosomes were
19 precipitated by the addition of PEG 20,000 and subsequently resuspended in buffer containing 50 mM
20 Hepes-KOH (pH 7.5), 150 mM KOAc, 20 mM Mg(OAc)₂, 7 mM β -mercaptoethanol, 20 U/mL
21 SuperASE-In.

22

23 ***In vitro* toeprinting assay**

24 Toeprinting assays were conducted as previously described^{11,38,41}. Briefly, reactions were prepared using
25 the PURExpress Δ Ribosome Kit (New England Biolabs) in volumes of 5 μ L and were allowed to
26 proceed for 15 min at 37°C. Primer extension was initiated by the addition of AMV reverse
27 transcriptase (New England Biolabs) and allowed to proceed for 10 min. All reactions contained 50 μ M
28 mupirocin.

29

30 **Preparation of linezolid and radezolid-only bound ribosomes for cryo-EM**

31 Purified 70S *wildtype* or Cfr-modified ribosomes were diluted to 1 pmol/ μ L in buffer containing 50 mM
32 Hepes-KOH (pH 7.5), 150 mM KOAc, 20 mM Mg(OAc)₂, 7 mM β -mercaptoethanol, 20 U/mL
33 SuperASE-In. Diluted ribosomes were then incubated at 4°C for 1.5 h after the addition of either
34 linezolid (Med Chem Express) or radezolid (Med Chem Express) in 40-60X molar excess. Samples
35 were then filtered for 5 min at 14,000 x g at 4°C using a 0.22 μ M low-binding Durapore PVDF filter
36 (Millipore).

37

38

1 **Preparation of stalled ribosome complexes for cryo-EM**

2 Stalled ribosome complexes containing the stalling peptide and corresponding oxazolidinone antibiotic
3 were prepared by *in vitro* transcription-translation. Reactions of 100 μ L volume were prepared using the
4 PURExpress Δ Ribosome Kit (New England Biolabs, E3313S) containing 0.8 U/ μ L of SuperASE-In,
5 ~1100 ng of DNA template encoding the stalling peptide sequence (**Supplementary Fig. 2**), 5000 pmol
6 of linezolid (Med Chem Express) or radezolid (Med Chem Express), and 360 pmol of *wildtype*
7 ribosomes or 250 pmol of Cfr-modified ribosomes. The reaction was halted by placing reactions on ice
8 after incubation at 37 °C for 1 h. The reaction was diluted to 190 μ L by the addition of Buffer C (50 mM
9 Hepes-KOH-pH 7.5, 150 mM KOAc, 20 mM Mg(OAc)₂, 7 mM β -mercaptoethanol, 20 U/mL
10 SuperASE-In) and purified by a 10-50% sucrose gradient also prepared in Buffer C. Ultracentrifugation
11 was performed using a SW Ti41 rotor (Beckman Coulter) at 22,000 rpm for 16 h at 4°C. Gradients were
12 fractionated using a Bio-Comp Fractionator in 20 fractions where absorbance at 260 nm was
13 continuously monitored. Fractions corresponding to stalled ribosome complexes were precipitated by
14 the slow addition of PEG 20,000 in Buffer C at 4°C to a final concentration of 8% w/v. Stalled
15 complexes were isolated by centrifugation for 10 min at 17,500 x g at 4°C. After removing the
16 supernatant, samples were slowly resuspended in Buffer C at 4°C. Sample concentration was determined
17 by NanoDrop UV spectrophotometer (Thermo), where $A_{260}=1$ corresponds to 24 pmol of 70S ribosome.
18 Purified stalled ribosome complexes were then incubated with 20-30X molar excess of the
19 corresponding oxazolidinone antibiotic (linezolid or radezolid) for 1 h at 4°C. Prior to freezing grids,
20 stalled ribosome complexes were filtered for 5 min at 14,000 x g at 4°C using a 0.22 μ M low-binding
21 Durapore PVDF filter (Millipore).

22 23 **Cryo-EM analysis**

24 Samples described above were diluted in Buffer C and deposited onto freshly glow-discharged
25 (EMS-100 Glow Discharge System, Electron Microscopy Sciences, 30 s at 15 mA) copper Quantifoil
26 (Quantifoil Micro Tools GmbH) grids with 2 nm thick amorphous carbon on top. Grids were incubated
27 for 30 s at 10 °C and 95% humidity, before blotting and vitrification by plunging into liquid ethane using
28 a FEI Vitrobot Mark IV (ThermoFisher). Ice thickness was controlled by varying the blot time, using
29 Whatman #1 filter paper for blotting. Grids were screened for ice quality using a FEI Talos Arctica
30 electron microscope (ThermoFisher, 200 kV, at UCSF) before grids were transported via dry shipper to
31 other facilities or loaded into a UCSF FEI Titan Krios (ThermoFisher).

32
33 All datasets used for reconstruction were imaged on FEI Titan Krios microscopes (ThermoFisher, 300
34 kV). The LZD-50S and LZD-SRC datasets were collected at the Stanford-SLAC CryoEM Center (S²C²)
35 using SerialEM on a microscope equipped with a Gatan K3 direct electron detector (DED) but without
36 an imaging filter. The RZD-50S and RZD-SRC datasets were collected at the National Center for
37 CryoEM Access and Training (NCCAT) using Legimon/Appion on a microscope equipped with a Gatan
38 K2 Summit DED and an imaging filter (20 eV slit). The RZD-SRC* dataset was collected at UCSF on a
39 microscope equipped with a Gatan K3 DED and an imaging filter (20 eV slit). The RZD-SRC* dataset
40 was collected on-axis; all other datasets were collected using a nine-shot beam-image shift approach
41 with coma compensation. All image stacks were collected in super-resolution mode. Pixel sizes,

1 micrograph count, defocus values, and exposures varied slightly between facilities and are reported in
2 **Supplementary Table 1.**

3

4 All image stacks were binned by a factor of 2, motion corrected, and dose-weighted using UCSF
5 MotionCor2⁴². All reconstructions used dose-weighted micrographs. Initial CTF parameters were
6 determined using CTFFIND4 within the cisTEM (v1.0.0-beta)¹⁶ software suite. Micrographs with poor
7 CTF fits or crystalline ice were excluded. Unsupervised particle picking used a soft-edged disk template
8 was followed by 2D classification in cisTEM. Initial and final particle counts are reported in
9 **Supplementary Table 1.** Only classes that clearly contained ice were omitted. An *ab initio*
10 reconstruction was carried out in cisTEM on the RZD-SRC* dataset, which yielded a starting reference
11 which was lowpass filtered and used as the initial reference for all five datasets. For SRC datasets,
12 multi-class Auto refinement in cisTEM was used to select for all particles that had tRNA present. After
13 this, and all non-SRC datasets, were subjected to a two-class Auto refinement in cisTEM to classify
14 between “good” particles and damaged “garbage” particles and high-frequency noise. The good classes
15 were carried forward into single class Auto and manual refinement efforts, including per-particle CTF
16 estimation. Care was taken not to increment the high-resolution cutoff in refinement to prevent
17 overfitting. Unsharpened maps were used in model refinement and figure preparation. Pixel size was
18 confirmed by comparison and cross-correlation between the resulting map and a
19 crystallographically-derived ribosome structure. 70S maps were used for model building, but 50S
20 focused refinement (using a binary mask to select for the 50S in cisTEM) was carried out to aid in figure
21 preparation. Map resolution values are reported as particle Fourier Shell Correlation (FSC) at 0.143.

22

23 **Atomic model building and refinement**

24 Atomic models of 50S ribosomal subunit with antibiotics and 70S stalled ribosome complexes were
25 generated by rounds of model building in Coot⁴³ and refinement in PHENIX⁴⁴. The atomic models of the
26 *wildtype E. coli* 50S subunit (PDB 6PJ6) was used as the starting point. Initial models for the three 70S
27 stalled ribosome complexes were obtained by combining: (i) a model of *wildtype E. coli* 50S subunit
28 (PDB 6PJ6); (ii) a model of the 30S subunit from *wildtype E. coli* ErmBL-stalled ribosome structure
29 (PDB: 5JU8)⁴⁵; (iii) P-tRNA and mRNA extracted from the ErmBL-stalled ribosome structure (PDB:
30 5JU8, mutated and remodeled in Coot⁴³ to yield fully modified *E. coli* tRNA^{Phe} and a short mRNA,
31 respectively); and (iv) the nascent peptide, which was modelled in Coot. Model refinement against the
32 acquired cryo-EM map was performed by multiple rounds of manual model building and restrained
33 parameter-refinement (real-space refinement, positional refinement, and simulated annealing). Modified
34 nucleotides were generated using eLBOW⁴⁶ within Phenix⁴⁰. The L3, L10 and L31 proteins were not
35 modelled in any of the cryo-EM structures. Prior to running MolProbity⁴⁷ analysis, nucleotides 76-94,
36 998-1009, and 1020-1040 of 16S rRNA, nucleotides 1053–1107, 2100–2189 of 23S rRNA, and
37 ribosomal proteins L9 and L11 were removed, due to their high degree of disorder. Overall, protein
38 residues and rRNA nucleotides show well refined geometrical parameters (**Supplementary Table 1**).
39 Figures were prepared using Pymol Molecular Graphics System Version 2.4.1 Schrödinger, LLC or
40 UCSF ChimeraX Version 1.2.5⁴⁸.

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