Data Release

Genomic features of *Mycobacterium avium* subsp. *hominissuis* isolated from pigs in Japan.

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72

73 Abstract

74 *Mycobacterium avium* subsp. *hominissuis* (MAH) is one of the most important agents causing
75 non-tuberculosis mycobacterial infection in humans and pigs. Genome analysis on MAH of
76 human isolates has been proceeding, however, those of pigs are limited despite its potential
77 source of infection to human. In the current study, we obtained 30 draft genome sequences

of MAH of pigs reared in Japan. The 30 draft genomes consisted of 4,848,678 – 5,620,788 bp length, 4,652 – 5,388 coding genes and 46 – 75 (Med: 47) tRNAs. All isolates had restriction modification associated genes and 185 – 222 predicted virulence genes. Two isolates had tRNA arrays and one isolate had a clustered regularly interspaced short palindromic repeat (CRISPR) region. Our results will be useful for evaluation of the ecology of MAH by providing a foundation for genome-based epidemiological studies.

Research Areas: Microbiology

Classification: Molecular Genetics, Microbial Ecology

Data description

Context

To date, incidence of infection caused by non-tuberculous mycobacteria (NTM) has been increasing all over the world [1]. Among NTMs, *Mycobacterium avium* complex (MAC) is one of the most critical agents. MAC has 4 subspecies, namely *M. avium* subsp. *avium* (MAA), *M. avium* subsp. *paratuberculosis* (MAP), *M. avium* subsp. *silvaticum* (MAS) and *M. avium* subsp. *hominissuis* (MAH). MAH is known as a major pathogen for humans, causing lung disease and sometimes disseminated infection in immune suppressed patients [2, 3]. MAH is also a main causative agent of mycobacteriosis in pigs [4], showing mesenteric and mandibular lymphadenitis [5] and sometimes systemic infection [6]. Swine mycobacteriosis exerts severe economic impact in affected farms. MAH infected pigs are suspected as potential risk for human infection [7, 8, 9, 10].

Recently, genomic epidemiological study of MAH has extensively progressed. In our recent studies, MAH is divided into 6 major lineages (MahEastAsia1, MahEastAsia2, SC1 - SC4) and each lineage is predominant in specific regions on a global scale [11, 12]. For example, the

104 MahEastAsia1 and MahEastAsia2 are frequently isolated from human lung disease in Japan
105 and Korea although SC1 – 4 are isolated from America and Europe [11, 12]. Japanese pig
106 isolates are mainly classified into 2 lineages, SC2 and SC4 [11, 12]. However, from the one
107 health point of view, to exactly clarify the ecology of MAH, the number of pig isolates used in
108 these studies was insufficient.
109 As stated above, genome-based analysis of MAH has been proceeding and the most
110 essential genes of MAH are thought to be mutual orthologues of genes in *Mycobacterium*
111 *tuberculosis* (MTB) [13]. Although components of virulence systems have been investigated
112 [14], reports about genome contents, even drug resistance genes are not available, despite
113 the increasing the incidence of MAH disease [1]. To understand MAH evolution, distribution
114 and to promote the identification of targets for antimicrobial drug discovery, the
115 characterization of the defining genomic features of MAH is essential.
116 Here we obtained draft genome sequences of 30 MAH isolates derived from pigs reared in
117 Japan, and identified genome features for bacterial defense systems, such as restriction
118 modification (RM) system, clustered regularly interspaced short palindromic repeat (CRISPR),
119 tRNA arrays, virulence factors and drug resistance genes. Our results in this study may
120 provide a way to understand the epidemiological relationship of MAH in human and pigs.
121

122 **Methods**

123 a) Sampling

124 MAH isolates were collected from pigs reared at two areas, Tokai and Hokuriku in Japan,
125 where about 10 % of pigs in Japan are reared. 48 mesenteric or mandibular lymph nodes of
126 pigs reared in Tokai area were collected from Gifu meat inspection center from July –
127 December, 2015. Samples (20: mesenteric lymph nodes, 1: mandibular lymph nodes, 1: liver)
128 of Tokai and Hokuriku area were collected between August, 1998 – Mar, 2018 and archived
129 in Toyama meat inspection center.

b) Bacterial isolation and DNA extraction

The method of bacterial isolation was available in protocols. io [15]. The mesenteric or mandibular lymph nodes with mycobacterial granulomatous lesions were mixed with 400ul of 2% NaOH and incubated at room temperature overnight. The samples were spread onto 2% Ogawa medium (Kyokuto Pharmaceutical, Tokyo, Japan) and incubated at 37 °C for 3 – 4 weeks. A single colony was inoculated onto 7H11 broth with 10% oleic acid-albumin-dextrose-catalase as a supplement. The isolates were stored with Microbank (Pro Lab Diagnostics Inc., Richmond Hill, ON, Canada) at -80°C. The method of extraction of genomic DNA was also available in protocols. io [16]. In brief, cells were delipidated by treatment with acetone, then lysed by lysozyme and Proteinase K. Genomic DNA was extracted by phenol/chloroform treatment of the lysates.

c) Identification of MAH and insertion sequence profile

PCR amplification of *M. avium* 16S rRNA genes (MAV) was conducted for screening [17]. Isolates positive for MAV were identified by sequencing *hsp65* and *rpoB* genes [18, 19]. Basic Local Alignment Search Tool (BLAST) analysis was conducted using partial sequences of *rpoB* gene. Phylogenetic analysis of both genes was conducted by maximum likelihood method using Molecular Evolutionary Genetics Analysis (MEGA) software ver. 7.0. Bootstrap values were calculated from 1,000 replications. Insertion sequence patterns of IS*900*, IS*901*, IS*902* and IS*1245* were performed as described previously [20, 21, 22]. IS*1311* and IS*1613* were searched for within draft genomes by using ISfinder (https://isfinder.biotoul.fr) with default parameters [23].

d) Draft genome sequences and genome annotation

Extraction of genomic DNA was described above. An average 350-bp paired-end libraries were prepared from extracted genomic DNA by TruSeq DNA PCR-Free High Throughput Library Prep Kit (Illumina, San Diego, CA, USA). Pair-end sequencing (2× 150-bp) was conducted using the HiSeq X Ten sequencing platform (Illumina) at the Beijing Genomics Institute (Shenzhen, China). Output reads were trimmed by TrimGalore! (https://github.com/FelixKrueger/TrimGalore) and were corrected its mismatched reads by SPAdes ver 3.12.0. [24]. The reads were assembled and polished using Pilon [25] and Unicycler [26], and then genome completeness was estimated by CheckM [27]. Taxonomic classification of contigs was carried out using Kaiju [28] and Anvi'o [29]. Draft genome sequences were annotated via the National Center for Biotechnology Information (NCBI) Prokaryotic Genome Annotation Pipeline (PGAP) [30].

e) Detection of bacterial defence systems (RM system and CRISPR CAS system) in MAH genome

RM systems were determined by online tool, Restriction-ModificationFinder version 1.1 (https://cge.cbs.dtu.dk/services/Restriction-ModificationFinder/) twice with the following settings (1: database: All incl. putative genes, threshold for %ID: 90%, minimum length: 80% to search the RM system of MAH and 2: database: All, threshold for %ID: 10%, minimum length: 20% to confirm the orthologue of MTB or the other Mycobacteria) [31]. CRISPR Cas systems were identified by the online tool CRISPRCasFinder program (https://crisprcas.i2bc.paris-saclay.fr/CrisprCasFinder/Index) with default setting [32, 33].

f) Detection of tRNA arrays in MAH genome

Total number of tRNAs in this study were retrieved from gb files annotated by PGAP. Draft genomes of GM17 and OCU479 isolates, which had more tRNAs than the others (Table 1), were inspected by tRNAscan-SE (http://lowelab.ucsc.edu/tRNAscan-SE/) to search tRNA arrays [34]. tRNA gene isotype synteny (expressed by the single-letter amino acid code) of both isolates and the reference strains were aligned and used for the maximum likelihood method by MEGA 7.0. Classification of both isolates was conducted as previously described [35].

g) Detection of virulence factors and drug resistance genes

Virulence genes were identified by using VFanalyzer (http://www.mgc.ac.cn/VFs/main.htm) [36]. We selected the following settings, genus: Mycobacterium, specify a representative genome: *M. avium* 104 and choose genomes for comparison: blank and draft genome fasta files were uploaded. Drug resistance genes were identified by Resistance Gene Identifier (RGI) version 5.1.0 (https://card.mcmaster.ca/analyze/rgi) with the following settings, Select Data Type: DNA sequence, Select Criteria: Perfect and Strict hit only, Nudge ≥95% identity Loose hits to Strict: Exclude nudge, Sequence Quality: high quality/coverage [37]. To confirm the existence of mutations detected by RGI, we retrieved the respective drug resistance associated genes from draft genome sequences, aligned by MEGA 7.0., and then manually checked for mutations in the nucleotide sequences.

Data Validation and quality control

199 Identification of MAH

200 The experimental workflow from sampling to identification is shown in Fig. 1. We
201 successfully obtained 13 MAH isolates derived from Tokai area and 8 out of 13 isolates (GM5
202 – GM44) with 22 isolates of Tokai and Hokuriku area (OCU467 – OCU486, Toy194, Toy195)
203 were used for draft genome sequence analysis. We conducted multiple examinations to
204 determine the isolates as MAH, IS possession patterns, sequence analysis of *hsp65*
205 (Supplementary Table 1). Among MAH subspecies, the patterns of IS possession is different
206 and is used for subspecies identification [38]. IS*900* and IS*901* are known as the indicator of
207 MAP and MAA, respectively [21, 22]. MAH is usually positive for IS*1245* [39], and is negative
208 for IS*900*, IS*901* and IS*902* [20], however, MAH strains without IS*1245* are frequently
209 distributed in Japan [39, 40]. In our study, 10/30 isolates were negative for IS*1245* (33.3%)
210 and none had IS*900*, IS*901* and IS*902* (Supplementary Table 1). In general, subspecies of *M.*
211 *avium* is also identified by *hsp65* gene analysis, which had 17 variations of SNP among
212 subspecies [19]. MAH has usually 1, 2, 3, 7, 8 or 9 *hsp* code [19], however, five isolates had
213 unclassified *hsp* code (indicated by N) in this study (Supplementary Table 1). Therefore, we
214 also conducted partial sequence analysis of the *rpoB* gene and the isolates were identified as
215 MAH by BLAST analysis. In addition, we conducted phylogenetic analysis based on *hsp65* and
216 *rpoB* genes retrieved from draft genome and all isolates in this study were also classified into
217 MAH (Fig 2). All of these examinations confirmed that our isolates were MAH.

218

219 Draft genome data

220 All of our draft genome sequences had a total length between 4.85 – 5.62 Mb, similar to
221 complete MAH genomes [41, 42]. All isolates had over 24kb N50 and over 40 fold genome
222 coverage (average 233) (Table 1).

223

224 Genome content analysis

In total, we identified 73 putative RM systems, including 24 type I RM systems, 48 type II RM systems, and 1 type III RM systems (Supplementary Table 2). All isolates had at least one Type II RM system and GM5, GM16, GM17, OCU468 – OCU470, OCU472, OCU473, OCU475, OCU476, OCU479, OCU483 and OCU484 had Type I, Type II RM systems, and GM44 had 3 types of RM systems. In these RM systems, 7 RM systems had homologues in MTB and 30 RM systems had homologues in *M. kansasii*. Orphan methyltransferase was detected in OCU473 and OCU479. CRISPR was detected only in GM44 (Supplementary Table 3). The sequences of the region were identical to MAH 104 (Query Cover: 100%, E value: 0.0, Per. Ident: 99.99%) which is the only MAH strain that had an intact CRISPR in the database (https://crisprcas.i2bc.paris-saclay.fr/MainDb/StrainList). The isolates had 185 – 222 virulence factors and 141 factors were common in all isolates (Supplementary Table 4). All isolates shared the same 2 drug resistance genes, *mtrA* which is associated with cell division and cell wall integrity [43] and resistance to macrolide antibiotics, and *RbpA* which regulates bacterial transcription and is associated with rifampicin resistance (Supplementary Table 5) [44]. In addition, single nucleotide polymorphisms (SNP) associated with drug resistance were found. All isolates had a C117D change in the *murA* gene conferring resistance to fosfomycin. A A2274G mutation in the *Mycobacterium avium* 23S rRNA which contributes to macrolide resistance was also detected by RGI, but when we examined the aligned nucleotide sequence, no point mutation was found in all isolates (Supplementary Table 5). CRISPR, virulence factor and drug resistance genes were selected from online tools. Original databases of each tool used in this study were updated in 2020, suggesting our data are based on the forefront of existing knowledge.

tRNA arrays

tRNA arrays were detected in isolates GM17 and OCU479 (Supplementary Table 6). tRNA array was discovered in some MAH isolates in the past study, and phylogenetic analysis

based on nucleotide sequences of tRNA array showed that tRNA array of MAH was classified into a specific group [35]. To confirm tRNA arrays in this study as authentic tRNA arrays, phylogenetic analysis was performed. Our tRNA arrays were classified into the group 3, as defined in a previous study (Fig. 3) [35].

Re-use potential

MAH is known as one of the most critical *M. avium* subspecies causing non-tuberculosis mycobacterial infection in human and pigs. Pigs are suspected to be the most dominant host of MAH in animal and a potential source of infection for human [7, 8, 9, 10]. However, the study about relationship with human and pig MAH isolates based on genome is limited [11, 12]. Our study provides 30 draft genome sequences of MAH isolated from pigs. We believe that these data will be useful for genome-based epidemiological studies to evaluate the importance of pigs as a source of infection. In addition, we provide molecular identification of defense systems, tRNA arrays, virulence factors and drug resistance genes. These data are expected to be used in future research of MAH classification, pathogenicity, and identification of antimicrobial drug targets. Principally, our draft genomes were derived from both cases of systemic and lymph node limited infection of MAH. Thus, the provided virulence factors can be included in the important candidate genes associated with systemic infection of pigs.

Data Availability

The summary information of draft genomes of the 30 MAH isolates are shown in Table 1. The genome sizes ranged to from approximately 4.8Mbps to 5.6Mbps. GC content was from 68.77% to 69.26%. All genome sequences have been deposited in GenBank under accession numbers VRUQ00000000, WEGO00000000 to WEGZ00000000 and WEHA00000000 to

WEHQ00000000, and SRA under accession numbers SRR13521605, SRR13556487 to SRR13556515.

Declarations

List of abbreviations

NTM: non-tuberculous mycobacteria; MAC: *Mycobacterium avium* complex; MAA: *M. avium* subsp. *avium*; MAP: *M. avium* subsp. *paratuberculosis*; MAS: *M. avium* subsp. *silvaticum*; MAH: *Mycobacterium avium* subsp. *hominissuis*; MTB: *Mycobacterium tuberculosis*; RM: restriction modification; CRISPR: clustered regularly interspaced short palindromic repeat; BLAST: Basic Local Alignment Search Tool; MEGA: Molecular Evolutionary Genetics Analysis; NCBI: National Center for Biotechnology Information; PGAP: Prokaryotic Genome Annotation Pipeline; SNP: single nucleotide polymorphism

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

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Author's contributions

T.K, K.O and H.Y wrote the manuscript. K.M, A.H, S.S and K.S collected samples. K.O, J.O.O, S.S and K.S performed laboratory works. T.K, K.O, A.O, H.Y, J.O.O, T.Ito and M.K conducted computational analysis. Y.N, T.A, T.Y, H.F, T.W, S.Y, K.A designed methods. M.A, A.D.B, K.O, N.Y, T.Iwamoto and F.M designed whole research and advised on the interpretation of the study's findings. All authors reviewed the manuscript.

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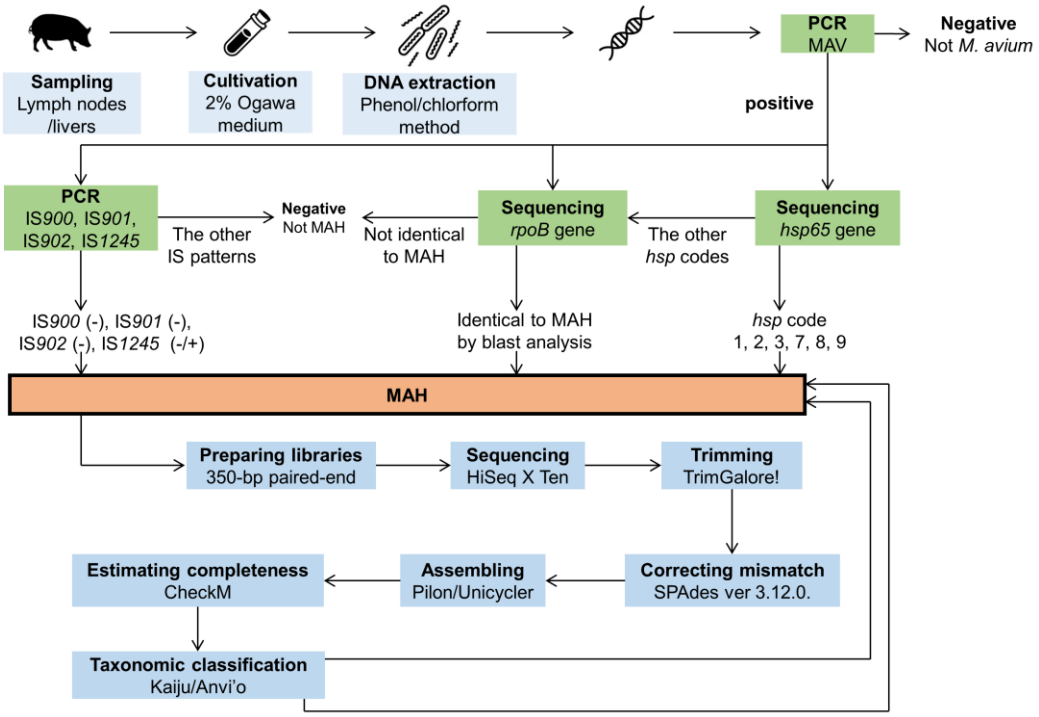
Figure Legends



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319 **Figure 1. The experimental workflows in this study.**

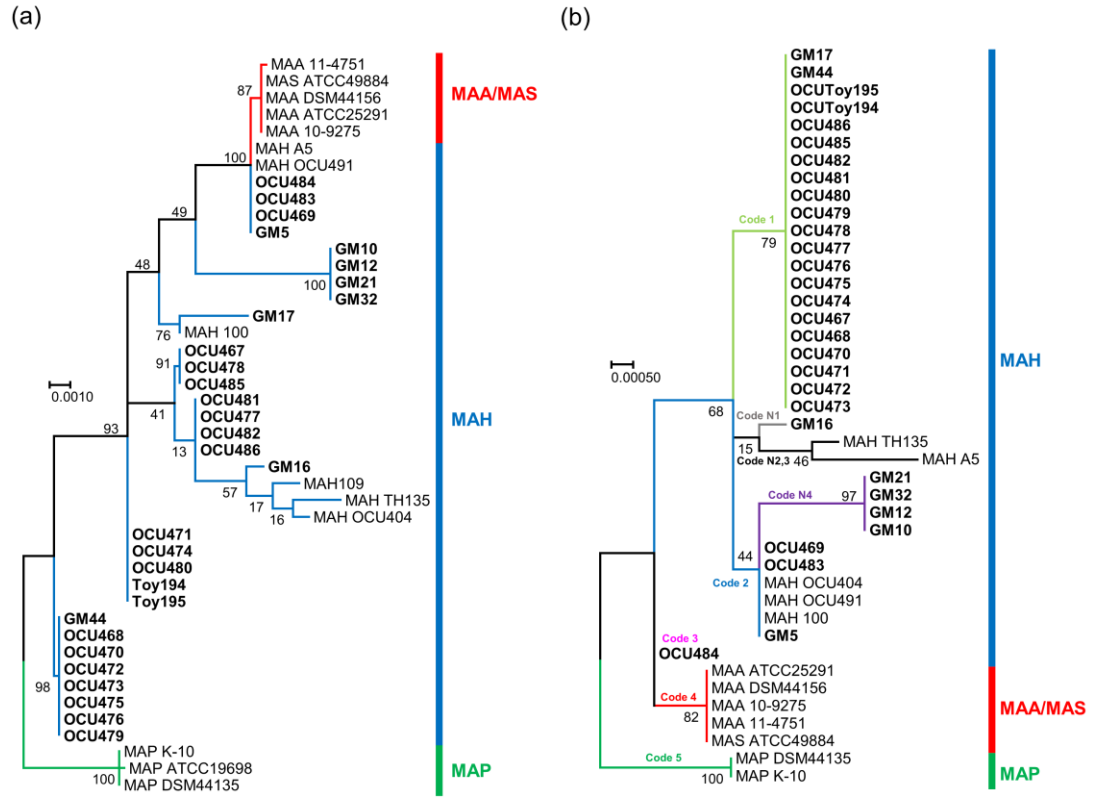

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Figure 2. Phylogenetic analysis based on *rpoB* gene and *hsp65* gene. Phylogenetic tree was generated by maximum likelihood method using MEGA 7.0. All isolates in this study are indicated in bold font. **(a)** 30 MAH isolates in this study were classified as MAH and were differentiated from MAP and MAA/MAS node. **(b)** All the isolates in this study were classified into 5 *hsp* code, code 1, 2, 3, N1 and N4. These isolates were differentiated from MAP and MAA/MAS nodes. The bootstrap values were determined from 1,000 replications. The scale bar indicates genetic distances among strains.

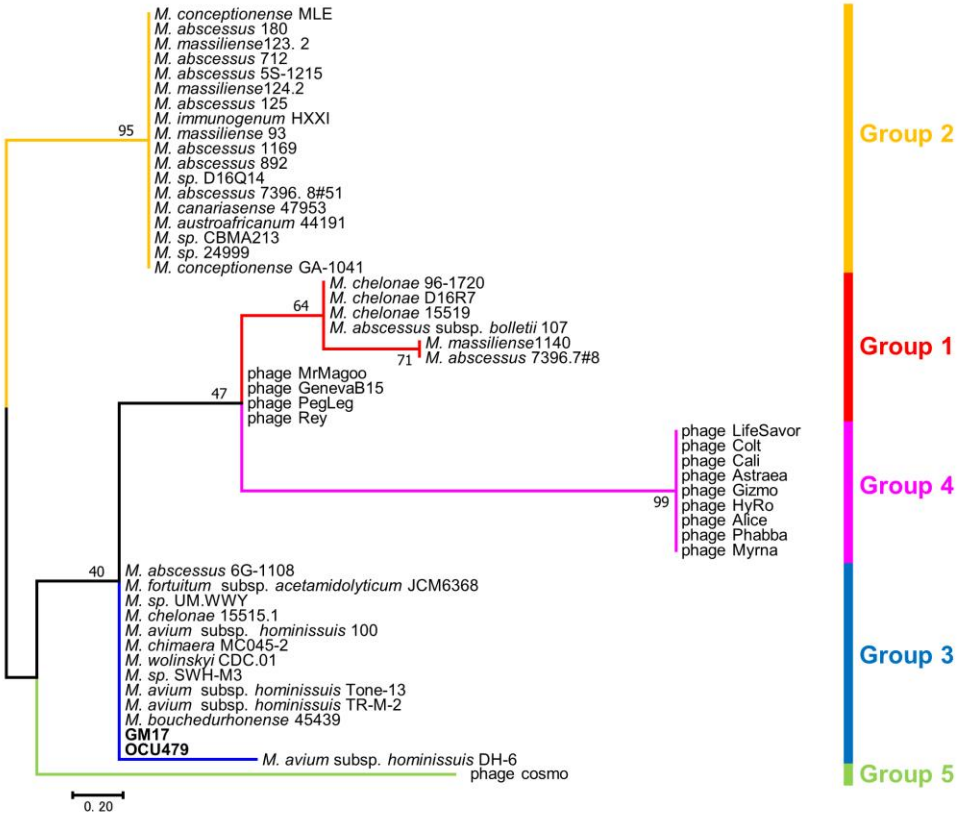

Figure 3. Phylogenetic tree based on the sequence of tRNA isotype located in tRNA array. Phylogenetic tree was generated by maximum likelihood method using MEGA 7.0. Two isolates (GM17 and OCU479 indicated in bold) were classified in Group 3. The bootstrap values were determined from 1,000 replications. The scale bar indicates genetic distances among strains.

Tables

336 **Table 1. Summary information for the draft genome sequences of 30 MAH isolates in this**

337 **study.** * CDSs: coding sequences.

Isolate	Genome size (bp)	N50 (bp)	Coverage	No. of contig	G+C content (%)	No. of CDSs*	No. of tRNAs
GM5	5,037,010	35,760	277	224	69.06	4,877	47
GM10	4,858,055	33,212	277	248	69.16	4,708	47
GM12	4,848,678	33,219	253	261	69.17	4,732	47
GM16	5,012,047	24,262	274	346	68.84	4,981	46
GM17	5,265,075	30,906	355	289	68.77	5,190	75
GM21	4,899,737	45,080	411	216	69.20	4,734	47
GM32	4,897,271	47,147	292	208	69.20	4,712	47
GM44	5,086,547	26,307	251	316	68.95	4,780	46
OCU467	5,110,693	243,182	207	75	69.16	4,803	46
OCU468	5,459,638	137,464	198	132	68.96	5,176	46
OCU469	5,167,480	190,329	191	57	69.19	4,886	47
OCU470	5,388,572	124,661	220	132	68.98	5,103	46
OCU471	4,990,913	193,095	237	70	69.24	4,713	47
OCU472	5,410,552	119,264	180	139	68.97	5,163	47
OCU473	5,237,229	105,027	232	118	69.11	4,981	47
OCU474	5,087,878	168,670	213	81	69.26	4,817	47
OCU475	5,376,580	113,114	243	130	68.99	5,121	46
OCU476	5,359,545	133,302	268	132	69.00	5,094	46
OCU477	5,087,664	218,065	221	85	69.22	4,779	47
OCU478	5,108,303	272,265	230	73	69.17	4,803	46
OCU479	5,620,788	112,152	167	143	68.78	5,388	75

OCU480	5,088,946	195,446	53	73	69.24	4,820	47
OCU481	5,100,722	163,519	247	101	69.19	4,802	47
OCU482	5,100,769	163,705	244	99	69.19	4,800	47
OCU483	4,943,024	200,611	228	68	69.24	4,652	47
OCU484	5,096,430	141,792	249	104	69.20	4,811	47
OCU485	5,109,020	243,182	258	80	69.16	4,805	46
OCU486	5,023,805	234,302	40	52	69.23	4,722	47
Toy194	5,347,524	216,164	273	93	68.97	5,018	47
Toy195	5,346,468	168,809	192	103	68.97	5,029	47

Supplementary Table 1. Isolates information and molecular characteristics of 30 MAH in this study. a: Detected IS was 1213bp and shared 83% identity with IS*900*.

Supplementary Table 2. Restriction modification system detected in 30 MAH isolates in this study. *1: These genes include the function of restriction enzyme/methyltransferase. *2: These genes could be orphan methyltransferase. Yellow background: putative genes.

Supplementary Table 3. Detected CRISPR-Cas systems in MAH GM44.

Supplementary Table 4. Virulence factors detected in 30 MAH isolates in this study.

Supplementary Table 5. Drug resistance genes detected in 30 MAH isolates in this study.

Supplementary Table 6. The information about tRNA array detected in MAH isolates GM17 and OCU479.

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