Poly-gamma-glutamic acid secretion protects *Bacillus subtilis* from zinc and copper intoxication

Running title: *Bacillus subtilis* produces poly-gamma-glutamic acid in response to zinc and copper excess

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Abstract

Zinc and copper are essential micronutrients that serve as cofactors for numerous enzymes. However, when present at elevated concentrations, zinc and copper are highly toxic to bacteria. To combat the effects of zinc and copper excess, bacteria have evolved a wide array of defense mechanisms. Here, we show that the Gram positive soil bacterium, *Bacillus subtilis*, produces the extracellular polymeric substance, poly-gamma-glutamate (γ-PGA) as a protective mechanism in response to zinc and copper excess. Furthermore, we provide evidence that zinc and copper dependent γ-PGA production is independent of the DegS-DegQ two component regulatory system and likely occurs at a post-transcriptional level. These data provide new insight into bacterial metal resistance mechanisms and contribute to our understanding of the regulation of bacterial γ-PGA biosynthesis.

Importance

Zinc and copper are potent antimicrobial compounds. As such, bacteria have evolved a diverse range of tools to prevent metal intoxication. Here, we show that the Gram-positive model organism, *Bacillus subtilis*, produces poly-gamma-glutamic acid (γ-PGA) as a protective mechanism against zinc and copper intoxication and that zinc and copper dependent γ-PGA production occurs by a yet undefined mechanism independent of known γ-PGA regulation pathways.

Introduction
Transition metals ions, such as zinc and copper, are essential for life, yet toxic in excess. Metals have long been appreciated for their antimicrobial properties. Ancient Egyptians used copper salts as early as 2400 BCE as an astringent, food preservative, and disinfectant (1). Since zinc and copper disrupt antibiotic-resistant biofilms, display synergistic activity with other antimicrobials, and kill antibiotic-resistant bacteria, metal-based antimicrobials are currently used in industry, agriculture, and healthcare as metal-impregnated coatings and surfaces and in combination with traditional antibiotics (2). Furthermore, growing evidence suggest that zinc and copper intoxication play a key role in the host-microbe interaction as antibacterial agents (3).

At elevated concentrations, zinc and copper can outcompete other metals, including manganese and iron, for binding to metal containing proteins (4). Bacteria have evolved diverse mechanisms to prevent zinc and copper toxicity. Under zinc and copper excess, the intracellular level of these ions can be maintained by reduced uptake and/or increased efflux through the regulation of expression and activity of membrane associated metal transporters (5). Zinc and copper can also be detoxified by intracellular sequestration by small proteins or low molecular weight thiols, such as bacillithiol and glutathione (6, 7). The specific proteins and cellular processes poisoned by metals can also be repaired by chaperones or antioxidants, or bypassed through the expression of alternative metabolic pathways (8).

During the course of our investigations into the molecular mechanisms underlying bacterial zinc toxicity, we observed that B. subtilis colonies became highly mucoid when grown in the presence of zinc. Here, we demonstrate that the mucoid phenotype is due to the production of the secreted polymer, poly-gamma-glutamic acid (γ-PGA), show that γ-PGA
protects *B. subtilis* from zinc and copper intoxication, and provide evidence that zinc and copper induce γ-PGA production independently from known regulatory mechanisms.

Results

Zinc and copper induce *B. subtilis* γ-PGA production

While studying the mechanisms of zinc intoxication, we observed that *B. subtilis* colonies displayed a mucoid phenotype (Figure 1A). *Bacillus subtilis* serves as a model organism for the study of biofilm formation and extracellular polymeric substance (EPS) production (9).

There is a growing body of evidence suggesting a link between zinc and copper homeostasis and EPS production. In *B. subtilis*, zinc and copper decrease biofilm hydrophobicity, thereby increasing antibiotic effectiveness (10). Additionally, excess zinc decreases *Streptococcus pyogenes* hyaluronic acid capsule production by inhibiting phosphoglucomutase, a key enzyme in central carbon metabolism (11). Recent studies on *Escherichia coli* virulence suggest that the presence of capsule may exacerbate bile salt mediated zinc starvation (12).

The EPS is composed of polysaccharides, proteins, nucleic acids, and poly-gamma-glutamic acid (γ-PGA) (13). γ-PGA is a biopolymer consisting of repeating units of L- or D-glutamic acid or a combination of both, and plays a role in virulence, survival during harsh conditions, and sequestration of toxic metal ions (14). In *B. subtilis*, the γ-PGA biosynthesis machinery is encoded by the *pgsBCAE* operon. In some bacteria, such as *B. subtilis*, γ-PGA is secreted and released from the cell surface, whereas in others like *B. anthracis* and *S. pneumoniae*, PGA remains attached to the cell as a capsule (15). When γ-PGA is made and
secreted by *B. subtilis*, the colonies produce a mucoid slime layer. Therefore, we hypothesized that mucoid material is the result of γ-PGA production.

To determine if the mucoidy observed in the presence of zinc is due to increased γ-PGA production, we compared the colony phenotype of wild-type and a *pgsB* deletion mutant in the presence of zinc (Figure 1B). We observed that in the presence of a sublethal concentration of zinc (250 µM ZnSO₄) wild-type colonies are mucoid and raised, whereas the *pgsB* mutant colonies are dull and flat. Furthermore, the mucoid phenotype is restored to the *pgsB* mutant when *pgsB* is expressed ectopically from an IPTG-inducible promoter (P$_{hyspank}$).

To determine if the increase in γ-PGA production is specific to zinc or a general response to metals, we tested for colony mucoidy in the presence of a number of physiologically relevant transition metals (iron, manganese, or copper). Previously reported sublethal metal concentrations (2mM FeSO₄, 500 µM MnCl₂, or 1 mM CuSO₄) were individually added to LB agar (16, 17). As previously reported, exposure to manganese leads to a wrinkled colony phenotype (18). Of the metals tested, only zinc and copper are able to induce colony mucoidy (Figure 1B) As observed with zinc, the colony mucoidy observed in the presence of copper is dependent on the presence of *pgsB*. Additionally, we utilized a spectrophotometric based assay to measure γ-PGA production in liquid media. Exposure to 250 µM ZnSO₄ or 100 µM CuSO₄ resulted in 3- or 5-fold increase in γ-PGA secretion (Figure 1C). From these data, we conclude that zinc and copper increase *B. subtilis* γ-PGA production.

γ-PGA protects *B. subtilis* from zinc and copper intoxication.
\(\gamma\)-PGA can directly bind to metals, such as zinc, copper, and silver, consistent with a proposed protective role for \(\gamma\)-PGA \((19, 20)\). To determine if \(\gamma\)-PGA production serves as a means of protecting \(B.\) subtilis from high levels of zinc and copper, we compared the zinc and copper sensitivity of wild-type and a \(\Delta pgsB\) mutant by a disk diffusion assay (Figure 2A). The \(\Delta pgsB\) mutant is more sensitive to zinc and copper than wild-type. Furthermore, the zinc and copper resistance of the \(\Delta pgsB\) mutant is restored when \(pgsB\) is expressed ectopically from an IPTG-inducible promoter.

We also assessed the contribution of \(\gamma\)-PGA to zinc and copper resistance by monitoring cell growth in LB broth in the presence or absence of zinc or copper (Figures 2B and C). Wild-type growth is impaired in the presence of 250 \(\mu\)M \(\text{ZnSO}_4\) or 100 \(\mu\)M \(\text{CuSO}_4\) as indicated by an extended lag phase, decreased growth rate or decreased final \(OD_{620}\). In the absence of metal, growth of the \(pgsB\) mutant is similar to wild-type. Upon the addition of zinc or copper, the \(pgsB\) mutant is unable to grow. When \(pgsB\) is expressed from an IPTG-inducible promoter, growth of the \(pgsB\) mutant in the presence of zinc or copper increases. Restoration of growth is to near wild type levels in the case of zinc. In the presence of copper, wild-type like growth is restored during exponential phase. However, the \(pgsB\) complemented strain is unable to achieve the same final \(OD_{600}\) as wild-type. Since metals are involved in numerous cellular processes, the partial rescue in the presence of copper may be indicative of differences is the physiological processes inhibited by zinc and copper. From the disk diffusion and growth data, we conclude that \(\gamma\)-PGA protects \(B.\) subtilis from zinc and copper intoxication on both solid and liquid media.

Zinc and copper dependent \(\gamma\)-PGA production is not controlled at the level of transcription
γ-PGA production can be regulated by altering the expression of the pgs operon, which encodes the PGA biosynthetic machinery, or of pgdS, which encodes a gamma-DL-glutamyl hydrolase involved in γ-PGA degradation (21, 22). Activation of pgsB operon expression involves the DegS-DegU two-component system, the ComP-ComA quorum sensing system, and the SwrA protein. (23). DegS-DegU control the expression of genes involved in many cellular behaviors, including biofilm formation, motility, and competence (24). In response to a variety of stimuli, the cytoplasmic DegS sensor histidine kinase phosphorylates the DegU response regulator (DegU-P), while DegQ enhances DegS-dependent phosphorylation of DegU (23, 25). Once phosphorylated, DegU-P activates pgs operon expression by binding upstream of the pgsB promoter (26, 27). ComP-ComA and SwrA indirectly activate pgsB operon expression by modulating DegU phosphorylation, in the case of SwrA or by regulating DegQ expression, in the case of ComP-ComA (27–29). Since DegS-DegU is downstream of ComA-ComP and SwrA, we decided to focus on DegS-DegU for further study.

To test if DegU is involved in the zinc or copper dependent production of PGA, we monitored the colony morphology of a degU mutant on LB agar in the presence of zinc or copper (Figure 3). Similar to wild-type, the degU mutant forms a highly mucoid colony in the presence zinc or copper. We infer that DegU is not required for zinc or copper dependent PGA production.

We next explored the possibility pgs operon or pgdS expression may be regulated in response to zinc or copper excess. To determine if expression of the pgs operon or pgdS is differentially regulated in response to zinc or copper, we used a β-galactosidase assay to measure pgs operon and pgdS promoter activity in the presence of increasing concentrations of
zinc or copper (Figure 3B). Even when grown in the presence of zinc (250 μM) and copper (100 μM) concentrations known to stimulate γ-PGA production, pgs operon and pgdS promoter activity did not substantially change. We conclude that zinc and copper dependent γ-PGA production does not occur through changes at the level of pgs or pgdS gene expression.

Discussion

Our studies are consistent with previous work showing that B. subtilis produces γ-PGA in response to excess zinc. We extend these previous studies to demonstrate that γ-PGA production is also induced by copper. We demonstrate that γ-PGA provides B. subtilis protection from zinc and copper intoxication to provide a more complete picture of bacterial metal resistance.

Since γ-PGA is a non-toxic, biodegradable compound with utility in medical and commercial applications, microbial biosynthesis of γ-PGA is an extensively studied process (30, 31). Given its anionic charge and affinity for metals, γ-PGA is particular interest as a tool for the bioremediation of toxic metal ions from contaminated water (32). Thus, there is great interest in optimizing growth conditions and understanding the genetic circuitry underlying γ-PGA production. Based on our results, the mechanism of zinc and copper induction of γ-PGA production is independent of the DegS/U two component system known to activate pgs operon expression. Furthermore, our data suggest that regulation may be post-transcriptional, since neither γ-PGA biosynthesis (pgsBCAE) nor degradation (pgdS) gene expression is affected by the presence of excess zinc or copper.
Future investigations will explore the post-transcriptional regulation of γ-PGA biosynthesis. An intriguing candidate is PgsE, a small, 55 amino acid protein encoded last gene in the pgs operon. The *B. anthracis* PgsE homolog (CapE) is required for γ-PGA synthesis and is believed to play a structural role (33). Interestingly, when PgsE is overexpressed from an inducible promoter on high copy plasmid in the presence of zinc, γ-PGA production by *B. subtilis (chungkookjang)* increases 3-fold compared to the absence of PgsE overexpression (34). However, its role under physiologically relevant conditions is unclear. PgsE (*B. anthracis* CapE) is thought to associate with the PgsBCA (*B. anthracis* CapBCA) biosynthetic machinery in *B. anthracis* and when heterologously expressed in *E. coli* (33, 35). It is tempting to speculate that zinc and copper may be modulating PGA levels by affecting the interaction between PgsE and the PgsBCA protein complex. In total, our continuing work will provide further insight into the molecular mechanism underlying zinc and copper dependent γ-PGA production.

**Materials and Methods**

**Strain and growth conditions**

Strains were grown in Luria-Bertani broth (10 g tryptone, 5 g yeast extract, 5 g/l NaCl) or plates containing 1.5% agar at 37°C. When necessary, antibiotics were added to the growth media at the following concentrations: 1 μg/ml erythromycin plus 25 μg/ml lincomycin (mls), 100 μg/ml spectinomycin, 5 μg/ml kanamycin, 5 μg/ml chloramphenicol or 100 μg/ml ampicillin. One mM of isopropyl β-D-thiogalactopyranoside (IPTG) was added to the growth medium when appropriate.
Strain construction

All strains and primers used in this study are listed in Tables 1 and 2, respectively. To facilitate genetic manipulation, all experiments were performed using the highly competent derivative of Bacillus subtilis 3610 carrying a comI deletion (3610 comI) (36). Chromosomal DNA was used for transformation as previously described (37). To induce natural competence, cells were grown in minimal competence media (1.7 g K$_2$HPO$_4$, 0.52 g KH$_2$PO$_4$, 2 g dextrose, 0.088 g sodium citrate dehydrate, 0.2 g l-glutamic acid monopotassium salt, 1 ml of 1,000× ferric ammonium citrate, and 1 g/l casein hydrolysate and 1% 300 mM MgSO$_4$).

Colony mucoidy assay

To evaluate the colony mucoidy phenotype, strains were grown to mid-exponential phase (OD$_{600}$~0.4) in LB broth. Five µl of the culture were then spotted on to freshly poured LB plates containing 1.5% agar that had been dried in the laminar flow hood for 10 minutes. The spots were allowed to dry an additional 10 minutes in the laminar flow hood. The plates were then incubated at 37°C for 16-18 hours. Images of the colonies were recorded using an iPhone 12 held in a tripod 36 inches from the agar surface.

PGA quantification

B. subtilis PGA production was measured using a cetyltrimethylammonium bromide (CTAB) assay as previously described (38, 39). Cells were grown in LB broth containing various concentrations of ZnSO$_4$ or CuSO$_4$ in LB media overnight at 37°C for 16-18 hours. Cells were then pelleted from 15 ml of culture by centrifugation at 10000 x g for 30 minutes at 4°C and the
supernatant was retained. To precipitate the PGA, thirty ml of ethanol (96%) was added to the
supernatant and incubated at 4°C overnight. The precipitated PGA was harvested by
centrifugation at 10000 x g for 10 minutes at 4°C. The resulting pellet was resuspended in 1 ml
of distilled water. In a 96 well flat bottom microtiter plate, one hundred µl of 0.07 M CTAB in
2% (w/v) NaOH was added to 100 µl of the resuspended PGA and mixed gently by pipetting to
avoid bubble formation. The OD_{400} of the resulting solution was measured in a Tecan Sunrise
microplate reader. A standard curve using purified PGA (Sigma, G1049) was used to calculate
the PGA concentration in the broth.

pgsB complementation

The plasmid pDR111 was used to generate the IPTG-inducible amyE::PhySpang- pgsB construct.
Briefly, a PCR product containing the pgsB coding region was amplified from B. subtilis 3610
comI chromosomal DNA using primers 1003/1004, digested with SalI and SphI, and cloned into
the SalI and SphI sites of the pDR111 plasmid. The plasmid was then transformed in to B.
subtilis 3610 comI. Successful transformation was indicated by spectinomycin resistance and
verified by PCR.

In frame markerless gene deletions

Gene deletion mutants were constructed from the BKE strains as previously described (40). BKE
strains carrying the gene deletion of interest marked by a kanamycin or erythromycin
resistance cassette were acquired from the Bacillus Genetic Stock Center (www.bgsc.org).
Chromosomal DNA was isolated and transformed in to our wild-type 3610 comI strain. To
remove the antibiotic resistance cassette, the temperature sensitive pDR244 plasmid encoding
the cre recombinase was introduced into the transformants by selecting for spectinomycin
resistance at 30°C. The strains was subsequently cured of the plasmid by growth at 42°C and
screened for sensitivity to spectinomycin (loss of the plasmid) and kanamycin or erthyromycin
(loss of the antibiotic resistance cassette). Successful gene deletion was verified by PCR using
primers 1001/1002.

β-galactosidase assay

A β-galactosidase assay was used to measure pgs operon and P_{pdgS} promoter activity, as
previously described (41). Cells were grown in LB broth in the presence or absence of ZnSO_{4}
(62.5, 125, or 250 µM) or CuSO_{4} (25, 50, 100 µM) at 37°C. One ml of cells were harvested by
centrifugation at early stationary phase (OD_{600}~1) and resuspended in an equal volume of Z-
buffer (40 mM NaH_{2}PO_{4}, 60 mM Na_{2}HPO_{4}, 1 mM MgSO_{4}, 10 mM KCl, and 38 mM 2-
mercaptoethanol. Cells were lysed by addition of lysozyme (0.2 mg/ml) and incubation at 30°C
for 15 minutes. Each sample was then diluted in Z-buffer to a final volume of 500 µl. The
reaction was started with the addition of 100 µl of 4 mg/ml of 2-nitrophenyl β-D-
galactopyranoside (ONPG). Once the sample began to turn yellow, the reaction was stopped
with 250 µl of 1 M Na_{2}CO_{3}. The OD_{420} was measured, and the specific activity of β-galactosidase
was calculated using the equation \[ \text{specific activity} = \frac{\text{OD}_{420}}{(\text{reaction time} \times \text{OD}_{600})} \times \text{dilution factor} \times 1000. \]

Growth curves
To measure the effect of zinc and copper on cell growth, strains were grown to mid-exponential phase ($OD_{600} \sim 0.4$) and subsequently diluted 1:100 into fresh LB media in the presence or absence of ZnSO$_4$ (250 µM) and CuSO$_4$ (100 µM) in a 96 well flat bottom microtiter plate. The plate was then incubated in a Tecan Sunrise microplate reader at 37°C with continuous shaking (high setting) and the OD$_{620}$ was measured every 15 minutes for 16 hours.

Disk diffusion assay

Disk diffusion assays were modified from (42). Briefly, a 1:100 dilution of an overnight culture was grown in fresh LB medium at 37°C with 250 rpm shaking until mid-exponential phase ($OD_{600} \sim 0.4$). A 100 µl aliquot of cells was then spread on the surface of a LB agar plate (15 ml of 1.5% agar) that had been dried in a laminar flow hood for 10 minutes. The inoculum was allowed to dry for an additional 10 minutes in the laminar flow hood. A sterile 6.5 mm Whatman filter disk was then placed on the surface of the agar and ZnSO$_4$ (5 µl of a 500 mm solution) or CuSO$_4$ (5 µl of a 100 mm solution) was added to filter disk and allowed to absorb into the disk for 5 minutes. The plates were incubated at 37°C for 16-18 hrs, after which the diameter of the zone of inhibition was measured.

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References
History of Medicine 1:237–257.

Copper Beds Sustain Terminal Cleaning and Disinfection Effects throughout Patient Care.

3. Djoko KY, Ong CY, Walker MJ, McEwan AG. 2015. The Role of Copper and Zinc Toxicity in


5. Braymer JJ, Giedroc DP. 2014. Recent developments in copper and zinc homeostasis in


Copper in Streptococcus pyogenes. mBio 11:e02804-20.


374    and nattokinase in Bacillus subtilis natto based on RNA-seq analysis. Microb Cell Fact
375    20:83.
376  39. Halmschlag B, Putri SP, Fukusaki E, Blank LM. 2020. Identification of Key Metabolites in
377    Poly-γ-Glutamic Acid Production by Tuning γ-PGA Synthetase Expression. Front Bioeng
378    Biotechnol 8:38.
381    Analysis of Two Genome-scale Deletion Libraries for Bacillus subtilis. Cell Syst 4:291-
382    305.e7.
383  41. Chan JM, Guttenplan SB, Kearns DB. 2014. Defects in the Flagellar Motor Increase
385  42. Chandrangsu P, Helmann JD. 2016. Intracellular Zn(II) Intoxication Leads to Dysregulation
386    of the PerR Regulon Resulting in Heme Toxicity in Bacillus subtilis. PLoS Genet
387    12:e1006515.
388
389  Strains used in this study

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### Primer table

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Figure 1: Zinc and copper induce PGA production. (A) Colony morphology of WT *B. subtilis* 3610 comI in the presence of 250 µM ZnSO₄, 1 mM CuSO₄, 2 mM FeSO₄, 500 µM MnCl₂. (B) Colony morphology of WT, ΔpgsB, and ΔpgsB P<sub>hyspank</sub>−pgsB grown on LB agar in the presence of 250 µM ZnSO₄ or 100 µM CuSO₄. IPTG (1 mM) was also added to ΔpgsB P<sub>hyspank</sub>−pgsB to induce pgsB expression. (C) Concentration of γ-PGA produced by cells grown in the presence of 250 µM ZnSO₄ or 100 µM CuSO₄. The γ-PGA concentration is expressed as mg of γ-PGA per gram of cell wet weight. Data shown are the mean and standard deviation from three independent experiments. *P* values were calculated by an ANOVA. **, *P* ≤ 0.01; ***, *P* ≤ 0.001
Figure 2: PGA protects *B. subtilis* from zinc and copper intoxication. (A) Disk diffusion susceptibility test performed on WT, ΔpgsB, and ΔpgsB *P* _hyspangk*-pgsB grown on LB agar. Five µl of 500 µM ZnSO₄ or 5 µl of 100 µM CuSO₄ was added to a filter disk. The zone of inhibition was determined by measuring the diameter of the zone of clearing. Data shown are the mean and standard deviation from three different experiments. *P* values were calculated by an ANOVA. n.s., nonsignificant; **, *P* ≤ 0.01. (B and C) Growth curves of WT, ΔpgsB, and ΔpgsB *P* _hyspangk*-pgsB in the presence and absence of either (B) 250 µM ZnSO₄ or (C) 100 µM CuSO₄. IPTG (1 mM) was also added to ΔpgsB *P* _hyspangk*-pgsB to induce pggB expression. Data shown are the mean and standard deviation from three independent experiments.
Figure 3: Zinc and copper dependent PGA production is not controlled at the level of transcription. (A) Colony morphology of WT and ΔdegU grown in LB agar in the presence of 250 µM ZnSO$_4$ or 1 mM CuSO$_4$. (B) $pgs$ operon ($pgsEΩlacZ$) and $P_{pgdS}$ promoter activity in cells grown in LB media amended with varying concentrations of ZnSO$_4$ or CuSO$_4$. Data shown are the mean and standard deviation from three independent experiments. No significant difference was detected between samples by ANOVA.