

1 **Title: A general theory for temperature-dependence in biology**

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25
26 **Abstract**

27 At present, there is no simple, complete, and first principles-based model for quantita-
28 tively describing the full range of observed biological temperature responses. Here, we derive
29 a theory exhibiting these features based on the Eyring-Evans-Polanyi theory governing chem-
30 ical reaction rates, and which is applicable across all scales from the micro to the macro.
31 Assuming only that the conformational entropy of molecules changes with temperature, we
32 derive a theory for the temperature dependence which takes the form of an exponential
33 function modified by a power-law. Our framework leads to six deductions applicable to any
34 biological trait that depends on temperature, and elucidates novel aspects of universal tem-
35 perature responses across the tree of life, from quantum to classical scales. All predictions
36 are well supported by data for a wide variety of biological rates and steady states, from
37 molecular to ecological scales and across multiple taxonomic groups. In addition, we provide
38 novel explanations of several empirical relationships including optimal values in temperature
39 response curves.

40
41 **One-Sentence Summary:** We derive a simple and universal formulae to characterize
42 temperature responses of biological processes across the tree of life.

43
44 **Introduction: Temperature dependence models and the Eyring-Evans-Polanyi**
45 **(EEP) theory.** Temperature is a major determinant of reaction rates of enzymes, which

46 regulate processes that manifest at all levels of biological organization from molecules to
47 ecosystems [1-7]. Formulating a fundamental theory for the response of biological rates to
48 changes in temperature, especially in ecological systems, has become a matter of some ur-
49 gency with the intensification of the climate crisis, particularly since existing models are
50 unable to account for such responses across the entire range of temperatures that support
51 life. Here we address this challenge by developing a comprehensive theory that unifies several
52 key properties that have not been simultaneously included in past work. This is critical for
53 making accurate predictions of biological quantities that are relevant in industrial applica-
54 tions, food production, disease spread, and responses to climate warming, among others.
55 The model we derive is: i) based on first principles and fundamental chemical mechanisms;
56 ii) mathematically simple in form, yet efficient in that it generates many predictions with
57 very few free parameters; iii) general and applicable across multiple levels of biological orga-
58 nization and taxa, thereby manifesting a universal biophysical law. Among its many novel
59 results, our theory makes six significant categories of new deductions that are confirmed by
60 data and resolves unexplained observations in the temperature response of organisms.

61

62 Different models have been suggested to explain temperature dependence in biology,
63 among which the Arrhenius equation [8-9] has become the most used by biologists and
64 ecologists, as epitomized, for example, by the Metabolic Theory of Ecology (MTE), [7] and
65 is given by

$$k = ae^{-E/k_B T} \quad (1)$$

66 where k is some biological quantity (e.g. at the molecular level, enzyme reaction rate), k_B
67 is Boltzmann's constant, T is absolute temperature in Kelvin degrees (K), E is an effective
68 activation energy for the process of interest, and a is an overall normalization constant char-
69 acteristic of the process. Consequently, a plot of $\log k$ vs. $1/T$ should yield a straight line,
70 often referred to as an Arrhenius plot. This equation was originally an empirical formulation,
71 but was later motivated heuristically from chemical reaction theory [10, 11]. Although it
72 has been instrumental in explaining the approximately universal temperature dependence
73 across many diverse biological rates [5, 7], it cannot account for deviations that occur beyond
74 certain temperature ranges in, for example, the metabolic rates of endotherms, thermophiles
75 and hyperthermophiles [3, 5, 12]. Furthermore, experiments and observations have long es-
76 tablished that the form of the temperature response has an asymmetric concave upward or
77 downward pattern relative to the canonical straight-line Arrhenius plot. Consequently, there
78 are ranges of temperatures where the traditional Arrhenius expression, Eq. (1), even gives
79 the wrong sign for the observed changes in biological rates: they *decrease* with increasing
80 temperature rather than increase, as predicted by Eq. (1).

81

82 The EEP transition state theory (TST) [13-14], which is the widely accepted theory of
83 enzyme chemical kinetics, offers the possibility of developing a fundamental theory for the
84 temperature dependence of biological processes that extends and generalises the heuristic
85 Arrhenius equation by grounding it in the underlying principles of thermodynamics, kinetic
86 theory and statistical physics [15]. The framework of the TST conceives of a chemical
87 reaction as a flux of molecules with a distribution of energies and a partition function given
88 by the Planck distribution, flowing through a potential energy surface (PES) which effectively

89 simulates molecular interactions. The configuration of molecules flowing through this surface
90 proceeds from i) a separate metabolite and enzyme to ii) an unstable metabolite-enzyme
91 complex, which, iii) after crossing a critical energy threshold barrier, or transition state, then
92 forms the final product (the transformed metabolite). EEP thereby derived the following
93 equation for the reaction rate [11]

$$k = \frac{k_B}{h} T e^{-\Delta G/RT} \quad (2)$$

94 where h is Planck's constant, ΔG is the change in Gibbs free energy or free enthalpy, $R =$
95 Nk_B is the universal gas constant and N is Avogadro's number. An overall coefficient of
96 transmission also is originally part of (2) but is usually taken to be 1. The change in Gibbs
97 free energy is the energy (heat) transferred from the environment to do chemical work. It can
98 be expressed in terms of enthalpy (ΔH) and the temperature-dependent change in entropy,
99 or dissipated energy (ΔS) [16], as $\Delta G = \Delta H - T\Delta S$. Eq. (2) can then be written as:

$$k = \frac{k_B}{h} T e^{\Delta S/R} e^{-\Delta H/RT} \quad (3)$$

100 Analogous to the Arrhenius expression, Eqs. (2) and (3) describe an exponential response
101 of the rate k to temperature provided, however, that there is no temperature dependence of
102 the thermodynamic parameters. Models have been developed for including this temperature
103 dependence, but they typically invoke several additional assumptions and new parameters
104 [11, 17-18]. Furthermore, unlike the widespread use of the Arrhenius equation in the MTE,
105 most models for temperature response have been conceived for a single level of biological
106 organization (primarily at the enzymatic/molecular level) [6, 18] or for specific taxonomic
107 groups; e.g. only for mesophilic ectotherms [19], endotherms [20], or thermophiles [21].

108 **Derivation of the Theory.** Temperature changes the conformational entropy of pro-
109 teins [23], which in turn determines the binding affinity of enzymes [24-25] and affects the
110 flexibility/rigidity and stability of the activated enzyme-substrate complex and hence the
111 reaction rate [25]. The resulting temperature dependence of the change in entropy, ΔS
112 (with enthalpy and heat capacity remaining constant), is the simplest mechanism for giving
113 rise to curvature in an Arrhenius plot and naturally leads, via Eq. (3), to power law devi-
114 ations from the simple exponential form [22]. Following [16], the change of entropy for a
115 given change in temperature can be expressed as $Td\Delta S/dT = \Delta C$, where ΔC is the heat
116 capacity of proteins. Integrating over temperature gives $\Delta S = \Delta S_0 + \Delta C \ln(T/T_0)$, where
117 ΔS_0 is the entropy when $T = T_0$, an arbitrary reference temperature, commonly taken to
118 be 298.15 K (25°C) [11]. Using this expression for ΔS in eq. (3), and after simplifying, we
119 straightforwardly obtain [11]

$$k = \left(\frac{k_B}{h}\right) \left[e^{\frac{\Delta S_0}{R}} T_0^{-\frac{\Delta C}{R}} \right] \left(\frac{1}{T}\right)^{-\left(\frac{\Delta C}{R}+1\right)} e^{-\frac{\Delta H}{RT}} \quad (4)$$

120 Eq. (4) has the form of a classic Arrhenius-like exponential term, modified by a power-
121 law, but with a different interpretation of the “effective activation energy” in terms of the
122 change in enthalpy. The pattern described by Eq. (4) is a curved temperature response in
123 an Arrhenius plot of $\ln k$ vs. T^{-1} :

$$\ln(k) = \ln\left(\frac{k_B}{h}\right) \left[e^{\frac{\Delta S_0}{R}} T_0^{-\frac{\Delta C}{R}} \right] - \left(\frac{\Delta H}{R}\right) T^{-1} - \left(\frac{\Delta C}{R} + 1\right) \ln T^{-1} \quad (5)$$

124 Consequently, $d \ln(k)/dT^{-1} = -\Delta H/R - (\Delta C/R + 1)/T^{-1}$, leading to the extrema of $\ln(k)$
 125 occurring at $T^{-1} = T_{opt}^{-1} = -(\Delta C + R)/\Delta H$ (see Supplementary Text S6). This is a mini-
 126 mum, i.e., the curve is concave upwards, or a “happy mouth”, if $\Delta C > -R$, whereas it is a
 127 maximum, or a convex downwards “sad mouth”, if $\Delta C < -R$. Furthermore, for T_{opt}^{-1} to be
 128 positive this requires $\Delta H < 0$ for a minimum or $\Delta H > 0$ for a maximum.

129

130 Several important points should be noted about our result:

131

132 1) Its simple mathematical form, namely an exponential modified by a power law, coin-
 133 cides with an empirical phenomenological equation suggested by Kooij in 1893 [26]. However,
 134 our derivation provides an underlying mechanism for the origin of the expression and, con-
 135 sequently, for how its parameters are related to the thermodynamic variables. Our approach
 136 differs from previous expressions derived from considerations of chemical kinetics [11]. For
 137 instance, a heuristic derivation inspired by a Maxwell-Boltzmann distribution predicts a
 138 similar expression but with a power law modification of $T^{1/2}$ rather than $T^{\frac{\Delta C}{R}+1}$ [13, 14],
 139 which apart from not having a mechanistic basis, is also unable to explain concave deviations.

140

141 2) An important consequence of our derivation is that it shows that a change of entropy
 142 with temperature is both sufficient and necessary for simultaneously explaining both the
 143 convex and concave curvatures commonly observed in temperature-response plots. Under a
 144 thermodynamic interpretation, the decrease in enzymatic rate with increasing entropy due
 145 to increasing temperature beyond the optimal, means that the disorder of the enzyme, and
 146 particularly of the active site, has reached a state that causes a decrease in the binding affin-
 147 ity to the ligands. In contrast, changes in enthalpy alone can only explain convex curvature
 148 but not concave. To see this explicitly, we express ΔH in terms of heat capacity in eq. (3),
 149 $\Delta H = \Delta H_0 - \Delta C(T - T_0)$, to obtain $k = \frac{k_B}{h} e^{\Delta S/R} \left(\frac{1}{T}\right)^{-1} e^{\left[\frac{\Delta H_0 - \Delta C(T - T_0)}{R}\right] \left(\frac{1}{T}\right)}$, which leads to
 150 $\ln k \propto \ln\left(\frac{1}{T}\right) - \left[\frac{\Delta H_0 + T_0 \Delta C}{R}\right] \left(\frac{1}{T}\right)$. Regardless of the sign of both ΔC and/or ΔH_0 , this always
 151 results in a convex downwards curve and so cannot explain, nor accommodate, concavity.
 152 Hobbs et al. [27] included changes in both enthalpy and entropy with temperature and de-
 153 rived a significantly more complicated expression than ours based on TST. In contrast, the
 154 minimalist scenario developed here is one in which only changes in entropy with temperature
 155 need be considered.

156

157 3) The above derivation was for reaction rates at the microscopic enzymatic scale. Fol-
 158 lowing the argument in the MTE we now show how it can be extended to biological variables
 159 at multiple scales up through multicellular organisms to ecosystems. The most salient exam-
 160 ple is metabolic rate, B . In general, this is derived by appropriately summing and averaging
 161 over all enzymatic reaction rates contributing to metabolism - some connected in series, some
 162 in parallel - and then summing and averaging over all cells: symbolically, $B \propto \overline{\sum k} \approx \bar{k}$.
 163 Assuming there is a dominant set of rate limiting reactions contributing to the production
 164 of ATP [19], then the temperature dependence of \bar{k} , and therefore B , can be approximated

165 by an equation of the form of Eq. (4), but with the parameters being interpreted as corre-
 166 sponding averages, $\overline{\Delta H}$ and $\overline{\Delta C}$. This results in: $B \approx B_0 \left(\frac{T_0}{T}\right)^{\frac{-\overline{\Delta C}}{R}-1} e^{\frac{-\overline{\Delta H}}{RT_0} \left(\frac{T_0}{T}\right)}$, where B_0 is
 167 a normalization constant (see Supplementary Material Eq. (S3.3) and Text S8).

168

169 4) Care, however, has to be taken with the normalization constants, such as B_0 in the
 170 case of metabolic rate, since from Eq. (4), these would naively be proportional to the ratio
 171 of the two fundamental constants, k_B and h . The presence of Planck's constant, h , for mi-
 172 croscopic enzymatic reactions appropriately reflects the essential role of quantum mechanics
 173 in molecular dynamics. On the other hand, for macroscopic processes, such as whole body
 174 metabolic rate, the averaging and summing over macroscopic spatio-temporal scales which
 175 are much larger than microscopic molecular scales must lead to a classical description de-
 176 coupled from the underlying quantum mechanics and, therefore, must be independent of h .
 177 This is analogous to the way that the motion of macroscopic objects, such as animals or
 178 planets, are determined by Newton's laws and not by quantum mechanics, and therefore do
 179 not involve h . Formally, the macroscopic classical limit is, in fact, realised when $h \rightarrow 0$. The
 180 situation here is resolved by recognising that the partition function for the distribution of
 181 energies in the transition state of the reaction has not been explicitly included in Eq. (2).
 182 This is given by a Planck distribution which leads to an additional factor $(1 - e^{-h\nu/k_B T})$
 183 where ν is the vibrational frequency of the bond, as first pointed out by Herzfeld [28]. For
 184 purely enzymatic reactions discussed above this has no significant effect since $k_B T \ll h\nu$,
 185 and thus $(1 - e^{-h\nu/k_B T}) \rightarrow 1$, resulting in Eq. (2). Multicellular organisms, however, cor-
 186 respond to the classical limit where $h \rightarrow 0$ so $k_B T \gg h\nu$ and $(1 - e^{-h\nu/k_B T}) \rightarrow h\nu/k_B T$,
 187 thereby cancelling the h in the denominator of Eq. (4).

188 Consequently, the resulting temperature dependence of macroscopic processes, such as
 189 metabolic rate, become independent of h , as they must, but lose a factor of T relative to the
 190 microscopic result, Eq. (4), so for metabolic rate, B , this is:

$$B \approx \tilde{B}_0 \left(\frac{1}{T}\right)^{\frac{-\overline{\Delta C}}{R}} e^{\frac{-\overline{\Delta H}}{RT}} \quad (6)$$

191 with the normalization constant, \tilde{B}_0 , no longer depending on h . Note that the above correc-
 192 tion for the enzyme level can also be applied to Eyring Eqs. (2) and (3), in which case they
 193 become mathematically identical to the Arrhenius relationship.

194

195 5) The micro and macro results, Eqs. (4) and (6), can be combined into a single expression
 196 for the temperature dependence of any variable, $Y(T)$:

$$Y(T) \approx Y_0 \left(\frac{1}{T}\right)^{\frac{-\overline{\Delta C}}{R} - \alpha} e^{\frac{-\overline{\Delta H}}{RT}} \quad (7)$$

197 where $\alpha = 1$ for the molecular level and 0 otherwise. $Y(T)$ represents either a rate or various
 198 steady-state quantities [11] including variables that have been explicitly derived theoretically,
 199 such as in the MTE. For reaction rates at the molecular level Y_0 is determined by Eq. (4).
 200 The corresponding extrema (either minima or maxima) in an Arrhenius plot now occur at
 201 $T^{-1} = T_{opt}^{-1} = -(\overline{\Delta C} + \alpha R) / \overline{\Delta H}$. It should be noted that the thermodynamic parameters

202 may have additional dependencies that make the forms of Eqs. (6) and (7) more complicated
203 under certain conditions [11].

204

205 In addition to quantitatively explaining the origin and systematic curvature of the Ar-
206 rhenius plot, our theory makes several further testable deductions that interrelate the key
207 features of thermodynamic parameters (e.g. enthalpy and heat capacity), biological traits
208 (e.g. growth and metabolic rates), classic thermal traits (e.g. thermal range and optimum
209 temperature). These various deductions, exhibited in Fig. S1, are summarized as follows:

- 210 i. The concave or convex form of the relationship between any biological trait and tem-
211 perature (Eq. (4)-(7); Fig. 1).
- 212 ii. The relationship between differences in rates (e.g., $Y(T_2)/Y(T_1)$) and differences in
213 temperatures ($T_2 - T_1$) (Eq. (S5.2)-(S5.3); Fig. S2).
- 214 iii. A linear relationship between heat capacity and enthalpy resulting from optimization
215 of the rate (i.e. when the rate of change of k respect to temperature is zero), and where
216 the slope of the relationship is the optimum temperature of the temperature response
217 curve (Eq. (S6.2); Fig. S4).
- 218 iv. The linear relationship amongst all pairs of the key thermal traits of the temperature
219 response curve such as the minimum, maximum, and optimum temperatures or thermal
220 range (Eq. (S6.5.1-3); Fig. S6).
- 221 v. The linear relationships between a given thermal trait and fundamental thermodynamic
222 parameters such as enthalpy (Eq. (S6.6.3-5); Fig. S7).
- 223 vi. The collapse, onto a single universal curve, of all temperature response curves after
224 the appropriate re-scaling given by our theoretical framework (see discussion below
225 and [11]; Eq. (9), (10); Fig. 2, Fig. S10). In particular our theory predicts that the
226 optimum of this curve should be located at a rescaled temperature of 1.

227 Importantly, data fitting to deductions iii), iv), and v) all reveal universal relationships
228 and constants. For example, the relationship between ΔC and ΔH holds across all data (fig
229 S4) and is driven by a slope that is the optimum temperature associated to response curves.

230

231 **Comparing the theory to temperature response curve data across levels of**
232 **biological organization and taxa.** To assess the model performance, we compiled a
233 database of 65 studies encompassing 128 temperature-response curves including those which
234 are explicitly predicted by biological theories such as the MTE. Our survey included data of
235 different rates/times/properties in different environments ranging from psychrophilic to hy-
236 perthermophilic organisms and across all domains of life, including viruses, bacteria, archaea
237 and unicellular and multicellular eukaryotes covering both ectotherms and endotherms (see
238 [11]).

239

240 We found that our theory provides an excellent fit to a wide variety of temperature re-
241 sponse data for rates and times, spanning individual to ecosystem-level traits across viruses,

242 unicellular prokaryotes, and mammals (see Supplementary Table S2). Fig. 1 shows some rep-
243 resentative examples of fits to concave patterns with long tails at low and high temperatures
244 (Fig. 1a-c) as well as convex patterns (such as the temperature dependence of endotherm
245 metabolism and biological times, Fig. 1d-f) also with tails at both ends. Prediction ii) also
246 fits the data well showing that curved temperature responses can be transformed into a linear
247 relationship for discrete measures of both rates and temperatures (Fig. S2). As predicted in
248 Eq. S6.2 we found a relationship between the estimated thermodynamic parameters $-\Delta C$
249 and ΔH (fig. S4) for all the (128) curve from our database.

250 Deductions iv-v) — the relationships among thermal traits and between thermal traits
251 and parameters — are well supported by a subset of the overall data (Figs. S6 and S7).

252 **Universal scaling and data collapse.** A powerful, but classic, method for exhibiting
253 and testing the generality of a theory is to express it in terms of dimensionless variables
254 which collapse the data onto a single “universal” curve across all scales [e.g. 30]. To do so
255 here, we introduce dimensionless rates, Y^* , and temperatures, T^* , by rescaling them by T_{opt} ,
256 where Y takes on either its minimum or maximum value, $Y_{opt} = Y(T_{opt})$:

$$Y^*(T^*) = \frac{Y(T)}{Y_{opt}}; \quad T^* = \frac{T}{T_{opt}} \quad (8)$$

257 In terms of these rescaled variables, Eq. (7) reduces to the simple dimensionless form

$$Y^{*1/a} = T^* e^{1/T^* - 1} \quad (9)$$

258 where $a = \overline{\Delta C}/R + \alpha$ with $\alpha = 0$ or 1 , depending on whether the system is macro- or mi-
259 croscopic. Note that the optimum is given by $Y_{opt} = Y_0 T_{opt}^a e^{-b/T_{opt}}$ and $T_{opt} = -b/a$, where
260 $b = \overline{\Delta H}/R$ [11].

261
262 Our theory therefore predicts that when $Y^{*1/a}$ is plotted against $1/T^*$ all of the various
263 rates regardless of the specific processes collapse onto a single parameterless curve whose
264 simple functional form is given by Eq. (9). Notice that this optimises at $T^* = 1$ and
265 encompasses in the same curve both the convex and concave behaviours predicted in the
266 original Arrhenius plot as a function of T . In that regard, note also that the function

$$\hat{Y}^*(T^*) \equiv (e/T^*)^a Y^*(T^*) = e^{a/T^*} \quad (10)$$

267 is predicted to be of a “pure” exponential Arrhenius form as a function of T^* . Thus, a plot
268 of $\ln(\hat{Y}^*(T^*))$ vs. $1/T^*$ should yield a straight line with slope a (see [11]).

269
270 Our prediction of the universal curve is very well supported by data, as illustrated in Fig.
271 2 where the collapse of all the data from this study for both convex and concave patterns
272 regardless of organizational level, temperature range or taxa are shown. This result strongly
273 supports the idea that our theory captures all of the meaningful dimensions of thermody-
274 namic and temperature variation for diverse biological properties, which can ultimately be
275 viewed as a single exponential relationship, Eq. (10). (See also Supplementary Material S8
276 and Fig. S8 for an alternative formulation for data collapse).

277

278 **Conclusion.** In conclusion, we have derived a mechanistic yet simple theory for bio-
279 logical temperature responses. Our model is a general extension of the EEP equation, but
280 unlike previous models requires only entropy to vary with temperature. From this single
281 assumption, we derived six novel general predictions that include not only a formula for
282 the temperature-dependence but also for explaining the parameters and relationships among
283 thermodynamic properties, thermal traits, and between the two. This set of predictions
284 leads to the discovery of universal constants, such an average global optimum for tempera-
285 ture response curves. We also derive a formula that expresses temperature dependence as
286 a universal law that leads to data collapse across all levels of biological organization, taxa,
287 and the whole range of temperature within which life can operate (-25 to 125°C). We do not
288 imply that temperature is the only variable determining biological rates. We acknowledge
289 the importance, and have included here, other variables that could be more limiting than
290 temperature in certain environments, such as pH, which also determine enzymatic and other
291 rates at higher levels of organization [31]. This framework allows us to make predictions for
292 scenarios of global warming, disease spread, and industrial applications. Further extensions
293 of this theory could incorporate time and other variables to predict the thermodynamic
294 parameters or vice versa (i.e. the parameters could explain biological traits), and future
295 connections could and should be made with non-equilibrium thermodynamics [32]. Finally,
296 our framework allow us to better understand the diverse impacts of climate change upon
297 processes at global scales, suggesting that processes such as mutation rates of viruses and
298 mortality will likely increase, given their convex temperature response curves , but other such
299 germination and growth rates will likely decrease given their concave temperature response
300 curves (Fig. 1).

301

302 **References and Notes**

303

- 304 1. Noll P, Lilge L, Hausmann R, Henkel M. *Processes* **8**, 121 (2020).
- 305 2. Rezende EL, Bozinovic F, Szilágyi A, Santos M. *Science* **369**, 1242-1245 (2020).
- 306 3. Schulte PM, Healy TM, Fanguie NA. *Integr Comp Biol* **51**, 691-702 (2011).
- 307 4. Yap TF, Liu Z, Shveda RA, Preston DJ. *Appl Phys Let* **117**, 060601 (2020).
- 308 5. Dell AI, Pawar S, Savage VM. *Proc Natl Acad Sci U S A*. **108**, 10591-6 (2011).
- 309 6. Ghjuvan Grimaud, Francis Mairet, Antoine Sciandra, Olivier Bernard. *Rev Environ*
310 *Sci Biotechnol* **16**, 625–645 (2017).
- 311 7. Brown JH, Gillooly JF, Allen AP, Savage VM, West GB. *Ecology* **85**,1771-1789 (2004).
- 312 8. Arrhenius SA. *Z Phys Chem* **4**, 226–248 (1889).
- 313 9. Laidler KJ. *J Chem Educ* **61**:494 (1984).
- 314 10. Laidler KJ, and King MC. *J Phys Chem* **87**, 2657–2664 (1983).
- 315 11. See Supplementary Text
- 316 12. Price CA, Weitz JS, Savage VM, Stegen J, Clarke A, Coomes DA, et al. *Ecol Lett*
317 **15**, 1465-74 (2012).
- 318 13. Eyring H. *J Chem Phys* **3**, 107 (1935).
- 319 14. Evans MG, Polanyi M. *Trans Faraday Soc* **31**, 875 (1935).
- 320 15. Zhou HX. *Q Rev Biophys* **43**, 219–293.
- 321 16. Prabhu NV and Sharp KA. *Annu Rev Phys Chem* **56**, 521-48.
- 322 17. Daniel RM, Danson MJ, Eisenthal R *Trends Biochem Sci* **26**, 223–225 (2001).

- 323 18. DeLong JP, Gibert JP, Luhring TM, Bachman G, Reed B, Neyer A et al. *Ecol Evol*
324 **7**, 3940–3950 (2017).
- 325 19. Gillooly JF, Brown JH, West GB, Savage VM, Charnov EL. *Science* **293**, 2248-51
326 (2001).
- 327 20. Porter WP, Kearney M. *Proc Natl Acad Sci U S A* **106**, 19666-72 (2009).
- 328 21. Klales A, Duncan J, Nett EJ, Kane SA. *Phys Rev E Stat Nonlin Soft Matter Phys*
329 **85**, 021911 (2012).
- 330 22. Sturtevant JM. *Proc Natl Acad Sci U S A* **74**, 2236–2240 (1977).
- 331 23. Wallin S and Chan HS. *J Phys Condens Matter* **21**, 329801 (2009).
- 332 24. Frederick KK, Marlow MS, Valentine KG, Wand AJ. *Nature* **448**, 325–329 (2007).
- 333 25. Tzeng S-R and Kalodimos CG. *Nature* **488**, 236-40 (2012).
- 334 26. Kooij DM. *Z Phys ChemAbt B* **12**, 155 (1893).
- 335 27. Hobbs JK, Jiao W, Easter AD, Parker EJ, Schipper LA and Arcus VL. *ACS Chem*
336 *Biol* **8**, 23882393 (2013).
- 337 28. Herzfeld KF. *Ann Phys* **59**, 635–667 (1919).
- 338 29. Park C and Raines RT. *J Am Chem Soc* **123**, 11472-11479 (2001).
- 339 30. West, GB, Brown JH, Enquist BJ. *Nature* **413**, 628 (2001).
- 340 31. Rektorschek M, Weeks D, Sachs G, Melchers K. *Gastroenterology* **115**, 628-41 (1998).
- 341 32. Demirel Y, Gerbaud V. 4th Edition (2018). Elsevier

342

343 **Methods**

344

Details on mathematical derivation, database compilation and estimation of parameters for
345 the models are in Supplementary Methods.

346

346 **Data and code availability**

347

The database and (R) code will be available in a public repository after acceptance. During
348 the review process, data and code can be provided upon request.

349

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350

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358 **Author contributions.**

359

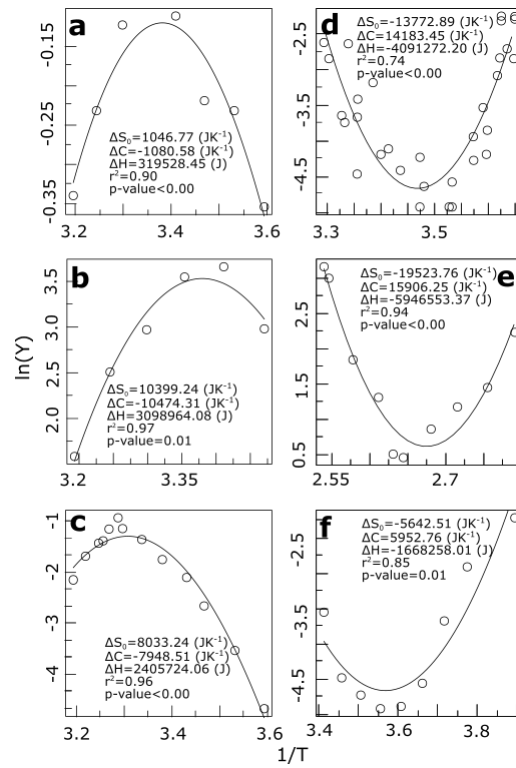
JIA and PM conceived the paper. JIA, CK, GW, and PM, derived the model. JIA compiled
360 the database and made the statistical analysis. JIA, BD, CK, GW, PM wrote the paper.

361

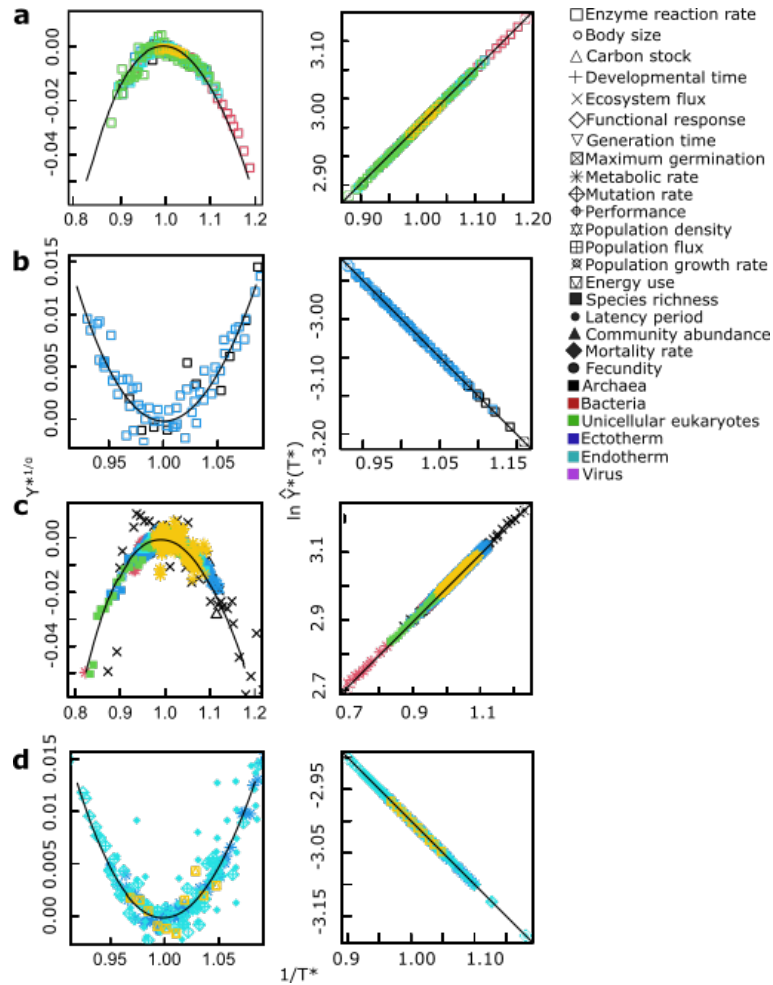
361 **Competing interests.**

362

The authors declare no competing interests.



363 Figure 1. Temperature response curves compared to the predictions of Eqs. (5) and (7) for
 364 a wide diversity of biological examples. Plotted are $\ln(Y)$ vs. $1/T$ (in $1/K$; where K is Kelvin
 365 degrees) showing (a)-(c) convex patterns and (d)-(f) concave patterns: (a) metabolic rate
 366 in the multicellular insect *Blatella germanica*, (b) maximum relative germination in alfalfa
 367 (for a conductivity of 32.1 dS/m), (c) growth rate in *Saccharomyces cerevisiae*, (d) mortality
 368 rate in the fruit fly (*Drosophila sukuzii*), (e) generation time in strain 121, (f) metabolic rate
 369 in the rodent *Spermophilus parryii*. For references see Supplementary Methods. The x-axis
 370 is in units of $(1/K) \times 10^3$.



371 Fig. 2. Universal patterns of temperature response predicted by Eqs. (9) and (10). The
 372 left panels show the convex and concave non-linear patterns predicted when $\ln Y^*$ is plotted
 373 vs. $1/T^*$, [Eq. (9)], whereas the right panels show the straight lines predicted when $\ln \hat{Y}^*$ is
 374 plotted vs. $1/T^*$, [Eq. (10)]. All curves regardless of variable, environment and taxa collapse
 375 onto a single curve when plotted in either of these ways. These rescalings explicitly show
 376 the universal temperature-dependence of the data used in Fig. 1, as well as additional data
 377 from compiled studies. Panels (a) and (b) show molecular (enzymatic) data exhibiting the
 378 predicted concave and convex patterns on the left, while (c) and (d) show corresponding
 379 concave and convex patterns for data above the molecular level. Note that there appears
 380 to be no variance in the fits to the linear predictions (the right-hand set of graphs) whereas
 381 there is significant variation in the non-linear ones (the left-hand set of graphs). This is
 382 basically because $\ln(\hat{Y}^*) \gg \ln(Y^*)$. The value of $\ln(Y^*)$ is typically around 0.01 with a
 383 variance much smaller than 0.005. Since $\ln(\hat{Y}^*) = \ln(Y^*) + a \ln(e/T^*)$ and $\ln(\hat{Y}^*)$ is typically
 384 around 3, fluctuations in $\ln(Y^*)$ are very much smaller and consequently completely lost. The
 385 point is that the difference between what is plotted in the left panels vs. that on the right,
 386 namely $a \ln(e/T^*)$, is in absolute value very large (more than 10 times the value of $\ln(\hat{Y}^*)$);
 387 furthermore, it is almost a constant over the range of temperatures since it is logarithmic,
 388 whereas all of the temperature variation is in the much smaller term $\ln(\hat{Y}^*)$.