1 **Atria: An Ultra-fast and Accurate Trimmer for Adapter and** 2 **Quality Trimming**

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21 **Abstract**

Background: As Next Generation Sequencing takes a dominant role in terms of output capacity and sequence length, adapters attached to the reads and low-quality bases hinder the performance of downstream analysis directly and implicitly, such as producing false-positive single nucleotide polymorphisms (SNP), and generating fragmented assemblies. A fast trimming algorithm is in demand to remove adapters precisely, especially in read tails with relatively low quality.

Findings: We present a trimming program named Atria. Atria matches the adapters in paired reads and finds possible overlapped regions with a super-fast and carefully designed byte-based matching algorithm ($O(n)$ time with $O(1)$ space). Atria also implements multi-threading in both sequence processing and file compression and supports single-end reads.

Conclusions: Atria performs favorably in various trimming and runtime benchmarks of both simulated and real data with other cutting-edge trimmers. We also provide an ultra-fast and lightweight byte-based matching algorithm. The algorithm can be used in a broad range of short-sequence matching applications, such as primer search and seed scanning before alignment.

Availability & Implementation: The Atria executables, source code, and benchmark scripts are available at <https://github.com/cihga39871/Atria> under the MIT license.

Research Area: Software and Workflows

Classifications: Bioinformatics, Software Engineering

43

44 **Statement of Need**

45 **Background**

46 Next generation sequencing (NGS) is a revolutionary new technology that produces
 47 massive, high-resolution genome sequence data to facilitate a broad range of biological
 48 applications. Illumina paired-end sequencing can read a DNA fragment from both ends
 49 and generate accurate reads for downstream bioinformatics analysis, such as assembly,
 50 resequencing, transcriptome profiling, variant calling, epigenome profiling, chromatin
 51 interaction, and chromosomal rearrangements [1, 2].

52 In paired-end library preparation, adapter sequences are the technical sequences
 53 ligated to both sides of inserts, which are the DNA fragments of interest. Then, DNA
 54 molecules with adapters are sequenced from both ends of the inserts so paired-end reads
 55 are generated. If insert sizes of paired-end reads are less than the read lengths, inserts
 56 are reversely complementary, and adapters are sequenced after reading through the
 57 inserts (**Fig. 1**). Thus, adapter contamination in the 3' end needs to be removed before
 58 downstream analysis.

59 Cleaning adapters can therefore be achieved by searching adapter sequences and/or
 60 aligning paired reads (**Fig. 1**). To date, some trimmers, such as AdapterRemoval [3],
 61 Trim Galore [4], and Trimmomatic [5], use both types of information to clean adapters.
 62 However, when the quality of sequencing reads decreases, the trimming process
 63 employing both types of information is likely to give different trimming suggestions.

Trimmers thus face a bottleneck when working on trimming adapters at accurate positions. Also, extremely short adapters at the low-quality 3' end are sometimes difficult to detect. Thus, trade-offs between trimming truncated adapters, and retaining inserts intact, become necessary.

These two issues hinder trimmers from cleaning adapter sequences and leaving DNA inserts intact. To combat this, we launch Atria, an integrated trimming program for NGS data. Atria uses a super-fast byte-based matching algorithm to detect adapters and reverse complementary regions of paired reads, and integrates carefully designed decision rules to infer true adapter positions. Thus, Atria can trim extremely short adapter sequences at accurate positions and not over-trim reads without adapters (**Fig. 1**).

In addition to adapter trimming, Atria integrated a set of trimming and filtering methods, such as consensus calling for overlapped regions, quality trimming, homopolymer trimming, N trimming, hard clipping from both ends, and read complexity filtration.

Implementation

The adapter finding algorithms used in Atria can be categorized in the following portions: DNA encoding, matching algorithm, matching and scoring, decision rules, consensus calling, quality trimming, and IO optimization (**Fig. 2**).

85 *DNA encoding*

86 The DNA encoding algorithm is developed based on BioSequences, a Julia package
 87 from BioJulia [6]. The original BioSequences package encodes DNA bases A, C, G, T
 88 as four-bit codes 0001, 0010, 0100, 1000, respectively. Extended codes are also
 89 supported, such as N (1111), S (0110), and gap (0000). DNA sequences are encoded
 90 and stored in a contiguous block of Random-Access Memory (RAM) as a dense array
 91 of unsigned 64-bit integers (UInt64) (**Fig. 2A**).

92 Atria makes use of the property of dense arrays to extract sequences as unsigned
 93 integers from available memory locations. When accessing the last several indices of a
 94 sequence, the extraction is illegal because operating systems do not allow the loading
 95 of data outside of sequence boundary. To solve the issue, Atria constructs a bit-safe
 96 sequence array, which elongates the sequence boundary by appending a UInt64 to the
 97 end of the original array, and setting all bits after the end of encoded DNA to 0 (**Fig.**
 98 **2A**).

99 It is noticeable that the smallest addressable unit of memory is one byte (8 bits)
 100 while each DNA is encoded in four bits, so only the even indices of sequence can be
 101 directly extracted (defining indices start from 0) (**Fig. 2A**). The extraction from odd
 102 indices requires extra operations, which is avoidable in many scenarios of a well-
 103 designed algorithm.

We denote a UInt64 extracted from the memory position n of sequence a by a_n . a_n is a 16-mer and represents the subsequence of a indexed from $2n$ to $2n+15$, which is denoted by $a[2n:2n+15]$ (**Fig. 2A**).

Matching algorithm

Given two sequences a and b , we plan to match the 16-base-long head of a to each index of b . However, only the even indices of b can be extracted from memory without bitwise operations, so we prepared two UInt64 of a : a_0 and a_- . a_0 is the 16-mer UInt64 loaded from the position 0 of a , and a_- can be computed from the following bitwise operations: $(a_0 \gg 4 | a_1 \ll 4)$. In this way, a_0 represents the subsequence of a indexed from 0 to 15 ($a[0:15]$), and a_- represents $a[1:16]$ (**Fig. 2B**).

In this way, the problem of matching the 16-base-long head of a to each index of b is converted to the problem of matching two 16-mers, a_0 , and a_- , to each addressable memory position of b . The latter requires less bitwise operations.

The number of mismatches K is computed in the formula:

$$K_{a_n, b_n} = 16 - \text{count_ones}(a_n \& b_n)$$

where *count_ones* counts the number of ones in the binary representation of the UInt64.

Let k denote the user-defined number of mismatches allowed in the 16-mer comparison of UInt64 a_n and b_n ($k = 2$ by default). After matching a_0 and a_- to each addressable memory position of b , if the minimum number of mismatches is not greater than k , the smallest index of b of the minimum mismatches is reported.

Therefore, the complexity of the matching algorithm is $O(n)$ time with $O(1)$ space, so its speed is extremely fast. One limitation is that when computing the number of mismatches of a_n and b_n , and if they have ambiguous bases in the same indices, the number of mismatches is underestimated. Another limitation is that the algorithm does not handle indels. Those limitations are compensated in the design of adapter matching, scoring, and decision rules.

Matching and scoring

We implement four pairs of matching to utilize properties of paired-end reads thoroughly: (1) matching adapter 1 head to read 1, (2) matching adapter 2 head to read 2, (3) matching read 1 head to reverse complement of read 2 and (4) matching read 2 head to reverse complement of read 1 (**Fig. 2C**). If the maximum number of bases matched of (1) and (2) is less than a user-defined cut-off (default is 9), (3) and (4) will be performed with a loosed k ($= k_{original} + 1$). If the largest number of matched bases of the four matches is greater than the cut-off, and some matches do not meet the requirement, we will re-run those matches with a loosed k ($= k_{original} + 3$) at the insert size indicated from the best match. If the new number of matched bases is greater than the cut-off, the old match will be discarded.

The scoring system measures the matching reliability of the whole 16-mer rather than each base. The Phred quality score Q of each base is converted to the probability P of that the corresponding base being correct using the formula:

$$P = 1 - 10^{\left(-\frac{Q}{10}\right)}$$

Then, the average base quality $\bar{P}$ of 16-mer sub-sequence a at the memory position n is computed:

$$\overline{P_{a_n}} = \frac{1}{16} \sum_{i=2n}^{2n+15} P_{a[i]}$$

Notably, if the read quality is too low, it would imply an invalid match. However, in reality, invalid matches are filtered out by the kmer-based algorithm. To solve the discordance, we limit the lower bound of $\bar{P}$ to 0.75 manually.

The matching score S between a_n and b_m is defined as:

$$S_{a_n, b_m} = \text{count_ones}(a_n \& b_m) \cdot \overline{P_{a_n}} \cdot \overline{P_{b_m}}$$

where *count_ones* counts the number of ones in the binary representation of the UInt64.

When sequence a is a user-defined adapter, $\bar{P}_a = 1$ is used. Generally, the matching score S is ranged from 0 to 16.

Pseudocode 1: Matching and scoring

```

r1_pos_adpt, r1_nmatch_adpt = match(adapter1, r1, k)
r2_pos_adpt, r2_nmatch_adpt = match(adapter1, r1, k)
k_extra = max(r1_nmatch_adpt, r2_nmatch_adpt) < 9 ? 1 : 0
r1_pos_pe, r1_nmatch_pe = match(reverse_complement(r2), r1, k + k_extra)
r2_pos_pe, r2_nmatch_pe = match(reverse_complement(r1), r2, k + k_extra)

max_nmatch = max(r1_nmatch_adpt, r2_nmatch_adpt, r1_nmatch_pe, r2_nmatch_pe)
max_pos = corresponding position of max_nmatch

if max_nmatch > 9

```



```

169         for matches with any nmatch < 9
170             redo match with loosed k = k + 3 at max_pos
171             replace old results if nmatch > 9
172
173     r1_prob_adpt = average_16mer_quality(r1, r1_pos_adpt)
174     r2_prob_adpt = average_16mer_quality(r2, r2_pos_adpt)
175     r1_prob_head = average_16mer_quality(r1, 1)
176     r2_prob_head = average_16mer_quality(r2, 1)
177     r1_prob_pe = average_16mer_quality(r1, r1_pos_pe)
178     r2_prob_pe = average_16mer_quality(r2, r2_pos_pe)
179     r*_prob_* = 0.75 if any r*_prob_* < 0.75
180
181     r1_score_adpt = r1_nmatch_adpt * r1_prob_adpt
182     r2_score_adpt = r2_nmatch_adpt * r2_prob_adpt
183     r1_score_pe = r1_nmatch_pe * r1_prob_pe * r2_prob_head
184     r2_score_pe = r2_nmatch_pe * r2_prob_pe * r1_prob_head

```

185

186 *Decision rules*

187 This module infers correct adapter positions from the four pairs of matching described
188 in the previous section. It is illustrated and self-explanatory in **Fig. 2D**. First, in each
189 read, the adapter and paired-end matches are compared. The one with the higher
190 matching score is chosen. If both matches support the same adapter position, the
191 matching score of the read is the sum of adapter and paired-end matching scores. Then,
192 the matches of the two paired-end reads are compared using the same strategy. If one
193 read finds an ideal adapter (matching score > 10 by default) while the other read is too

short to check or the average base accuracy of its 16-mer is less than 0.6 (Phred $Q < 5$), both reads will be trimmed. If the matching score of a given read pair is less than 10 (by default), the read pair will not be trimmed.

Other read pairs will be taken a further examination to reduce false positives, which are usually adapter matches at read tails. A read tail is defined as the last several bases (default is 12 bp) of each read. Reads are not trimmed if both statements are true: (1) In any paired read, the adapter is found at the tail, but the paired-end match is not; (2) In both paired reads, adapter and pair-end matches suggest different trimming positions.

Before the final trimming, one additional step is required for the accurate positioning of adapter sequences. The previous steps usually assume the read 1 and 2 have the same length of insert sizes, but indel in reads usually lead to over or under trim one base. To prevent this circumstance, Atria re-positions the adapter by matching one adjacent base with the first four bp of adapter sequences. The position of the highest number of bases matched is chosen to trim. This step is ignored when the inferred insert size is greater than the read length minus three because, in this situation, the adapter sequence is too short to check.

Pseudocode 2: Decision rules

```
function correct_insert_size(pos1, score1, pos2, score2)
    if pos1 == pos2
        return pos1, score1 + score2
    else
        score = max(score1, score2)
```

```

217         pos = corresponding pos of max score
218     return pos, score
219
220     r1_pos, r1_score = correct_insert_size(r1_pos_adpt, r1_score_adpt, r1_pos_pe,
221     r1_score_pe)
222     r2_pos, r2_score = correct_insert_size(r2_pos_adpt, r2_score_adpt, r2_pos_pe,
223     r2_score_pe)
224     r12_pos, r12_score = correct_insert_size(r1_pos, r1_score, r2_pos, r2_score)
225
226     if r1_pos != r2_pos
227         if r1_score > 10
228             r2_prob = average_16mer_quality(r2, r1_pos)
229             @goto "trim" if r2_prob < 0.6
230         elseif r2_score > 10
231             r1_prob = average_16mer_quality(r1, r2_pos)
232             @goto "trim" if r1_prob < 0.6
233
234     function check_read_tail(read)
235         E_adpt = whether adapter found at read tail
236         E_pe = whether pair-end match found at read tail
237         E = E_adpt & E_pe # both matches in read tail
238         R = rx_pos_adpt == rx_pos_pe # adapter and pair-end match at same position
239         return E, R
240
241     E1, R1 = check_read_tail(r1)
242     E2, R2 = check_read_tail(r2)
243     E = E1 | E2 # at least one read matching in read tail
244     R = R1 | R2 # at least one read matching at the same position

```

```

245  is_false_positive = E & !R
246
247  if r12_score > trim_score & !is_false_positive
248      @label "trim"
249      r1_pos_adjusted = adjacent_one_bp_check(r1, adapter1, r12_pos)
250      r2_pos_adjusted = adjacent_one_bp_check(r2, adapter2, r12_pos)
251      trim(r1, r1_pos_adjusted)
252      trim(r2, r2_pos_adjusted)

```

253

254 *Consensus calling*

255 In this module, the overlapped base pairs of read 1 and 2 are corrected to the
 256 corresponding bases with higher quality scores. It has three steps, prediction,
 257 assessment, and correction.

258 In the prediction step, Atria makes a preliminary estimate of whether a read pair
 259 contains an overlapped region. If adapters are trimmed and the remaining lengths of
 260 read 1 and 2 are the same, the prediction passes. If no adapter can be trimmed, two
 261 additional matching and scoring are required. The head of the reverse complement of
 262 read 2 is matched to read 1, and the head of the reverse complement of read 1 is matched
 263 to read 2. If the two matches reach a consensus, the prediction passes. Otherwise, the
 264 prediction fails and consensus calling is skipped.

265 In the assessment step, Atria compares the whole overlapped region using a similar
 266 matching algorithm, except that ambiguous bases (N, 1111) are converted to gaps (0000)

before matching. If the ratio of mismatch is greater than a user-defined value (28% by default), the assessment fails, and consensus calling is skipped.

In the correction step, each base pair in the overlapped region is corrected to the corresponding base with the highest quality score.

Quality trimming

Atria implements a traditional sliding window algorithm to remove the low-quality tail. The sliding window scans from the front of the read and computes the average Phred quality score of the sliding window. If the average quality is less than a given threshold, the read tail is removed.

IO optimization

Atria spends more time on reading and writing than matching and trimming, so the key to reducing runtime is to optimize IO usage. Considering that a large amount of RAM is easily accessible nowadays, Atria trades increased RAM usage with decreased time. A large block of memory is allocated for reading input files, which is then wrapped and encoded to FASTQ objects parallelly using multi-threading. On the contrary, in the writing process, Atria unboxes and decodes FASTQ objects to string vectors in parallel and writes sequentially to files. In addition, pigz (parallel gzip) and pbzip2 (parallel bzip2) are called for compression and decompression when needed [7, 8]. Atria also support running with a single thread.

288

289 **Comparison to related work**

290 **The performance of adapter trimming on a simulated dataset**

291 We simulated 8.9 G bases with 100 bp paired-end reads from the *Arabidopsis thaliana*
 292 reference genome using the Skewer modified ART, a public NGS read simulator to
 293 allow adapters in the reads [9, 10]. The simulation profile was trained from a 101 bp
 294 paired-end public dataset SRR330569, and the 33 bp adapter pair used in read
 295 simulation is AGATCGGAAGAGCACACGTCTGAACTCCAGTCA and
 296 AGATCGGAAGAGCGTCGTGTAGGGAAAGAGTGT [11].

297 Atria v3.0.0 was benchmarked with cutting-edge and popular trimmers, including
 298 AdapterRemoval v2.3.1 [3], Skewer v0.2.2 [10], Fastp v0.21.0 [12], Ktrim v1.2.1 [13],
 299 Atropos v1.1.29 [14], SeqPurge v2012_12 [15], Trim Galore v0.6.5 [4] and
 300 Trimmomatic v0.39 [5]. Only adapter trimming was used, and other trimming and
 301 filtration were disabled. Detailed command line arguments are listed in **Table S1**. Each
 302 trimming software was running on an idle Ubuntu 19.10 server with a 32-thread Intel
 303 i9-9960X Central Processing Unit (CPU) @ 3.10 GHz, 128 gigabyte (GB) DDR4-3200
 304 RAM, and a 2 terabyte (TB) Samsung 970 EVO Solid State Drive (SSD) (sequential
 305 reads and writes up to 3.5 and 2.5 TB/s).

306 The trimming performance was evaluated based on the following metrics: positive
 307 predictive value (PPV), as the fraction of the number of correctly trimmed reads to all
 308 trimmed reads; sensitivity, as the fraction of the number of correctly trimmed reads to

the reads with adapters; specificity, as the fraction of the number of untrimmed reads without adapters to all reads without adapters; and Matthew's correlation coefficient (MCC) measuring overall quality of pattern recognition, as

$$MCC = \frac{TP \cdot TN - FP \cdot FN}{\sqrt{(TP + FP)(TP + FN)(TN + FN)(TN + FP)}}$$

where TP is the number of reads trimmed correctly, TN is the number of untrimmed reads without adapters, FP is the number of over-trimmed reads, and FN is the number of under-trimmed reads [3, 10].

Table 1 Adapter trimming performance on the 8.9 G bases with 100 bp paired-end simulated data

Trimmer	PPV (%)	Sensitivity (%)	Specificity (%)	MCC (%)
Atria	99.35	99.81	99.82	99.51
AdapterRemoval	99.42	99.94	99.83	99.61
Atropos	99.57	97.34	99.88	98.00
Fastp	98.73	99.58	99.61	98.92
Ktrim	91.51	85.85	98.84	87.84
SeqPurge	57.92	99.80	76.84	66.62
Skewer	99.58	99.53	99.88	99.44
Trim Galore	40.05	82.98	62.39	38.96
Trimmomatic	99.29	57.86	99.88	71.05

The adapter trimming performance is shown in **Table 1**. AdapterRemoval, Atria and Skewer were the top-class adapter trimmers in terms of MCC (99.61%, 99.51%, 99.44%, respectively) (**Table 1**). Fastp (98.92%) and Atropos (98.00%) were in the second tier (**Table 1**). Ktrim obtained a good specificity (98.84%) but sacrificed its

sensitivity (85.85%), and Trimmomatic achieved an exceptional specificity (99.88%) by trading off its sensitivity (57.86%) (**Table 1**).

To compare speed and efficiency, elapsed time (wall time) and average CPU consumption of each trimmer were recorded in different threading (1-32 threads) for uncompressed and gzip compressed data formats (**Fig. 3, Table S1**). Efficiency was defined as the fraction of processing speed to the percent of CPU utilized, so it was a better measurement, especially in CPU-intensive scenarios, such as running on a server with a job scheduling system or trimming multiple samples at the same time. Ktrim and Atria were two of the fastest trimmers in terms of speed and efficiency, from one to 16 threads (**Fig. 3, Table S1**). For uncompressed data, Trimmomatic was faster than Atria using 8-32 threads, but its real CPU usage was much greater than Atria (**Fig. 3, Table S1**). The speed and efficiency of AdapterRemoval and Skewer were generally 2-4 times less than Atria, and Atropos was the slowest one (**Fig. 3, Table S1**). SeqPurge did not support the output of uncompressed data, so it was only tested in the compressed benchmark.

When trimming compressed data, the speed of AdapterRemoval, Skewer, Fastp, Atropos and Trimmomatic kept constant when the number of threads increased from 4 to 32, because they failed to utilize more than four CPU in the IO process, while Atria and Trim Galore did not have the limitation (**Fig. 3, Table S1**). Atria was faster than Trim Galore, and the efficiency of Atria was constantly two to three times greater than Trim Galore (**Fig. 3, Table S1**). SeqPurge showed strange speed curves; when

assigning a single thread to SeqPurge, the average CPU usage was 300%, and the speed and average CPU usage dropped when assigning 8 to 32 threads (**Fig. 3, Table S1**). In addition, Ktrim did not support output compressed files, so we ignored it. In general, Atria was the fastest trimmer when trimming compressed files.

The detailed statistics of adapter trimming accuracy on a simulated dataset

The previous portion benchmarks on a whole dataset. This section evaluates trimming accuracy regarding different read properties, including adapter presence or absence, base error, and adapter length. To achieve the goal, Atria integrates a benchmarking toolkit for read simulation and trimming analysis.

The read simulation method was inspired by how sequencers read DNA. First, an original DNA fragment (insert) with a given original insert size is simulated base by base. Adenine, thymine, cytosine, and guanine are randomly chosen repetitively. Then, the insert and adapter sequences are copied base by base with an error profile, which simulates the procedure of sequencing by synthesis. The error profile defines substitution rate, insertion rate, and deletion rate.

Twenty-one million read pairs were simulated with a uniform read length (100 bp), different error profiles, adapter length, and original insert sizes. The baseline error profile comprises a 0.1% substitution rate, 0.001% insertion rate, and 0.001% deletion rate, inspired by an Illumina error profile analysis [16]. 1x, 2x, 3x, 4x, and 5x baseline error profile, 16, 20, 24, 28, and 33 adapter lengths, and 66 to 120 even insert sizes are

chosen. In this way, the reads with the least insert size have full lengths of adapters. The reads with 66-98 original insert sizes contain adapters, and the reads with 100-120 original insert sizes are free from adapter contamination, except for few reads with a 100 bp insert size containing indels. Therefore, in each condition combination, 30 thousand read pairs were simulated to avoid random errors. The reads were trimmed with the same method described in the last section.

The average trimming performance among different conditions is shown in **Fig. 4 A**. When adapters are present, Atria trims 99.9% adapters accurately, and SeqPurge, Fastp, and Atropos follow closely with an accuracy of 99.7% (**Fig. 4 A1**). When adapters are absent, AdapterRemoval, Skewer, Trimmomatic, Atropos, and Atria successfully leave 100.0% reads intact, and Fastp falls behind with 99.8% accuracy (**Fig. 4 A2**).

Fig. 4 B illustrates the trimming accuracy on different read error profiles. When adapters are present, the accuracy of all trimmers drops as error rates increase (**Fig. 4 B1**). Atria keeps the highest accuracy from 100.0% to 99.9%, and is almost not affected by different error rates (**Fig. 4 B1**). The accuracy of SeqPurge, Fastp, and Atropos decrease from 99.9% to 99.6%, 99.5%, and 99.4%, respectively (**Fig. 4 B1**). With no adapter present in reads, the accuracy is hardly influenced by error profiles (**Fig. 4 B2**), so the performance is similar to **Fig. 4 A2**. In addition, adapter lengths ranging from 16 to 33 bp are not relevant to most trimmers' accuracy, including Atria (**Fig. 4 C**).

The performance of adapter trimming on real sequencing dataset

RNA-Seq paired-end dataset (SRR330569)

SRR330569 is a real RNA-Seq dataset sequenced from *Drosophila simulans* with 5.46 G bases and 2 x 101 bp read length. It contains 38 bp adapter sequences AGATCGGAAGAGCGGTTCAGCAGGAATGCCGAGACCG and AGATCGGAAGAGCGTCGTGTAGGGAAAGAGTGTAGAT in read 1 and read 2, respectively. Adapter trimming was performed by different trimmers without other trimming or filtering methods. Then, a sliding-window based quality trimming was performed to remove low-quality tails (sliding window size = 5 and average Q score $\geq$ 15). The adapter-trimmed reads and adapter-and-quality-trimmed reads were mapped to the *Drosophila simulans* genome version 2.02 from FlyBase using Hisat2 v2.2.1, respectively [17, 18]. Mapping statistics were collected using SAMTools Stat v1.10 [19]. Skewer did consensus calling after adapter trimming, and no option was provided to disable it. To achieve benchmark parity, Skewer was compared to Atria with consensus calling enabled, and other trimmers were compared to Atria without consensus calling. Time trimming was recorded in accordance with a common scenario: inputs were gzip-compressed and trimmed with eight threads, and outputs were also gzip-compressed to reduce massive disk use. All tested trimmers worked in the scenario except that Ktrim could not output gzip files (**Table 2**).

Atria was the fastest program to process and output compressed data in terms of wall time (**Table 2**). It also achieved the highest number of reads mapped and paired,

and the percent of properly paired reads with or without quality trimming. Generally, higher base mapped is accompanied with higher error rate in the mapping process, so it is important to interpret the two metrics together. Atria had the lowest mapping error rate of 8.1833‰ and the forth highest base mapped (**Table 2**). The trimmers (AdapterRemoval, Fastp, and Atropos) with the highest three error rates has the highest base mapped (**Table 2**). Our program generally improved more than 5% compared to other trimmers for the data without quality trimming (**Table 2**). The mapping statistics of data without quality trimming were generally worse than with quality trimming except for Atria. Specifically, the properly paired rates of other trimmers without quality trimming were 0.5 to 4% less than with quality trimming (**Table 2**). Quality trimming also increased the number of mapped and paired reads and reduced the number of unmapped reads (**Table 2**).

Table 2 Performance of trimmers on real data

Metric	Trimming and consensus		Trimming only							
	Atria	Skewer	Atria	AR	Atropos	Fastp	Ktrim*	SeqPurge	Trim Galore	Trimmomatic
Low-quality dataset (SRR330569, RNA, Hisat2 mapping)										
Elapsed time (min:sec)*	2:38	9:19	2:32	11:29	10:08	9:17	1:34 + GZ	3:53	<u>3:39</u>	9:38
No quality trimming										
Reads mapped and paired	26,126,804	24,694,330	25,781,268	<u>24,539,060</u>	24,505,656	24,545,646	24,196,658	24,240,072	24,046,542	22,797,620
Reads unmapped	27,276,761	28,254,804	27,379,299	28,338,455	28,410,022	<u>28,350,294</u>	28,747,248	28,591,442	28,647,873	29,649,287
Properly paired reads (%)	48.3	45.6	47.6	<u>45.3</u>	45.2	<u>45.3</u>	44.5	44.7	44.2	38.3
Base mapped (cigar)	2,387,212,225	2,354,164,041	2,322,436,545	2,346,791,204	2,341,355,822	<u>2,344,847,438</u>	2,316,915,673	2,304,776,514	2,317,321,510	2,237,846,534
Error rate (%)	7.3952	9.5897	8.1833	9.8902	9.8536	9.8683	9.7920	9.7904	9.6994	<u>9.3106</u>
With quality trimming										
Reads mapped and paired	25,942,092	25,787,464	25,728,206	25,721,788	25,714,956	<u>25,725,480</u>	25,473,670	25,364,392	25,654,498	24,744,754
Reads unmapped	27,245,720	27,364,827	27,361,655	27,369,773	27,373,854	<u>27,360,527</u>	27,556,820	27,736,292	27,400,932	28,064,739
Properly paired reads (%)	47.9	47.6	47.5	<u>47.5</u>	47.5	47.5	46.9	46.8	47.3	42.3
Base mapped (cigar)	2,317,238,536	2,316,981,456	2,302,740,463	2,304,584,269	2,304,325,743	<u>2,304,437,244</u>	2,292,034,762	2,263,465,110	2,297,815,439	2,246,076,618
Error rate (%)	7.1114	7.7882	<u>7.8902</u>	7.9160	7.9141	7.9149	7.9059	7.8787	7.8921	7.5649
High-quality dataset (ERR4695159, cell-free DNA, Bowtie2 mapping)										
Elapsed time (min:sec)*	3:08	11:34	3:03	13:48	13:41	11:29	1:41 + GZ	<u>4:05</u>	4:34	11:44
No quality trimming										
Reads mapped and paired	54,367,548	54,287,616	<u>54,324,964</u>	54,319,438	54,299,922	54,322,088	53,087,420	54,446,104	54,218,344	54,128,760
Reads unmapped	1,094,145	1,016,244	1,119,103	<u>989,335</u>	1,002,745	978,665	2,317,968	999,099	1,041,005	1,094,752
Properly paired reads (%)	96.8	96.7	<u>96.7</u>	<u>96.7</u>	<u>96.7</u>	<u>96.7</u>	94.1	97.0	96.4	88.6
Base mapped (cigar)	7,703,820,585	7,700,134,673	7,700,298,217	<u>7,701,482,302</u>	7,700,749,164	7,699,298,008	7,607,799,306	7,512,839,360	7,677,087,845	7,720,352,493
Error rate (%)	3.3082	3.8388	3.8724	3.8771	3.8834	<u>3.8564</u>	4.3239	3.8007	3.9173	6.1984
With quality trimming										
Reads mapped and paired	54,553,566	54,526,276	54,546,192	54,541,948	54,539,502	<u>54,549,462</u>	53,335,674	54,608,308	54,482,002	54,403,982
Reads unmapped	965,447	984,845	967,917	970,869	973,217	826,424	2,136,081	<u>890,884</u>	999,918	914,003
Properly paired reads (%)	97.0	97.0	<u>97.0</u>	<u>97.0</u>	<u>97.0</u>	<u>97.0</u>	94.4	97.1	96.8	89.0
Base mapped (cigar)	7,653,879,312	7,649,380,218	7,646,989,362	7,647,893,624	<u>7,648,184,196</u>	7,646,574,606	7,556,468,882	7,461,588,482	7,625,484,706	7,668,777,971
Error rate (%)	2.9547	3.2535	3.2678	3.2698	3.2792	<u>3.2634</u>	3.7183	3.2117	3.3109	5.5798

Note: AR = AdapterRemoval. In the trimming-only benchmark, bold and underline formats indicate the first and second trimmers (including tie) in terms of each metric, respectively. *Elapsed time (wall time) is benchmarked based on trimming and output gzip files with 8 threads, except that Ktrim cannot output gzip files (marked with time + GZ).

Genome-wide human cell-free DNA dataset (ERR4695159)

Generally, plasma cell-free DNA is short in length [20], and trimming is extremely important in medical diagnosis. Here, we chose a human genome-wide cell-free DNA dataset ERR4695159. It has 8.4 G bases with 2 x 150 bp read length with 33 bp adapter sequences AGATCGGAAGAGCACACGTCTGAACTCCAGTCA in read 1 and AGATCGGAAGAGCGTCGTGTAGGGAAAGAGTGT in read 2. The benchmark workflow was the same as the RNA-Seq analysis, except that the clean reads were

mapped to the human reference genome hg38 (GRCh38.p13) using Bowtie2 v2.3.5.1 [21].

Atria was also the fastest trimmer in the scenario (3 min 3 s) (**Table 2**). SeqPurge and Trim Galore finished the task in more than 4 minutes, while others spent more than 11 minutes (**Table 2**).

In adapter-trimming-only statistics, SeqPurge had the highest mapped and paired reads (54,446,104) and the highest properly paired reads (97.0%) (**Table 2**). Atria followed with 54,324,964 mapped and paired reads. Atria, AdapterRemoval, Atropos, and Fastp all had 96.7% properly paired reads (**Table 2**).

With quality trimming, the overall performance increased, and properly paired reads were closer; SeqPurge had 97.1% properly paired reads, with Atria, AdapterRemoval, Atropos, and Fastp close behind at 97.0% (**Table 2**). Only 89.0% of reads were properly paired with Trimmomatic (**Table 2**).

Discussion

Atria performs favorably with other cutting-edge adapter trimmers in accuracy, robustness, speed, and efficiency. Its performance is ascribed to the byte-based matching algorithm. The design concept of the algorithm is to minimize any unnecessary CPU operations by taking advantage of the data structure of dense arrays.

Matrix-based algorithms, such as the Needleman-Wunsch algorithm and the Smith-Waterman algorithm, allocate and update a matrix and perform base-to-base

comparison [22, 23]. They report every mismatch and gap between two sequences while Atria skips this step since it is focussed on the start positions of successful matches. Despite that, the matching algorithm used in Atria is able to identify mismatch loci when needed.

The byte-based matching algorithm is lightweight and designed for short sequence scanning. Each DNA is encoded in four bits and stores continuously in RAM. A sub-sequence can be extracted as an unsigned integer from a given memory position. For example, a 64-bit unsigned integer (UInt) represents a 16-mer, and a 128-bit UInt represents a 32-mer. The comparison between two sub-sequences is completed within the accumulator register, a CPU unit for arithmetic or logical operation. It does not require addressing or updating a scoring matrix from RAM. When comparing a short sequence, such as an adapter, to a long sequence, such as the read, the 16-mer of the short sequence is compared to every position of the long sequence. Hence, the byte-based matching algorithm has $O(n)$ expected time complexity and $O(1)$ space complexity in adapter matching, where n is the length of the long sequence.

The algorithm also has its limitations. It only reports the number of matched bases and does not report the positions of mismatches, so it cannot be used for sequence alignment solely. Besides, the algorithm does not handle insert and deletion. However, the average indel rate of Illumina library is 10^{-6} to 10^{-5} [16], and the low indel rate is almost negligible in real data analysis. In addition, Atria does four pairs of matches in

different locations to compensate for the limitation. If one match is failed because of indel, other matches will suggest the real adapter positions.

In the runtime benchmark, we compared how trimmers performed using extremely high CPU cores. In general, efficiency marginally decreased as CPU usage increased due to the trimmers' parallel implementation and the inevitable cost of multi-threading, such as task scheduling and context switching. In addition, IO could be the main bottleneck for most hard disk drives and some solid-state drives. Thus, if the system IO reaches a bottleneck, an efficiency plateau would be expected sooner.

Conclusions

We introduce not only Atria, a cutting-edge trimming software for sequence data, but also the ultra-fast and lightweight byte-based matching algorithm. The algorithm can be used in a broad range of short-sequence matching applications, such as primer search and seed scanning before alignment. Atria is implemented in Julia, a programming language designed specifically for high performance. The source code, executables, and benchmark scripts are available on Atria's Github page [24].

Availability and requirements

Project name: Atria

Project home page: <https://github.com/cihga39871/Atria>

Operating system(s): Linux, OSX

498 Programming language: Julia

499 Other requirements: Julia v1.4, Pigz v2.4 or higher, Pbzip v1.1.13 or higher

500 License: MIT

501 Research Resource Identification Initiative ID: SCR_021313

502

503 **Data Availability**

504 The datasets SRR330569, and ERR4695159 analyzed during the current study are
505 available in the Sequence Read Archive from the National Center for Biotechnology
506 Information [11, 25].

507 The Atria source codes, releases, documents, and benchmark scripts can be
508 downloaded from Atria's Github page [24].

509

510 **Abbreviations**

511 CPU: Central processing unit; DNA: Deoxyribonucleic acid; GB: Gigabyte; MCC:
512 Matthew's correlation coefficient; NGS: Next-generation sequencing; PPV: Positive
513 predictive value; RAM: Random-access memory; RNA: Ribonucleic acid; SNP: Single
514 nucleotide polymorphism; SSD: Solid-state drive; TB: Terabyte; UInt: Unsigned
515 integer; UInt64: Unsigned 64-bit integer; WGS: Whole-genome sequencing.

516

517 **Competing interests**

518 The authors declare that they have no competing interests.

519

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523

524 **Authors' contributions**

525 JC developed Atria software, performed benchmark experiments under the supervision
526 of XL. Both XL and LH serve as co-supervisors and participates in the design of the
527 study. MH contributes to the optimization of the algorithm. AZ participates in
528 benchmark validation. JC, LH, and XL drafted the manuscript. All authors read and
529 approved the final version of the manuscript.

530

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536

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Figure legends

Figure 1 Overview of Atria workflow

Figure 2 Adapter trimming algorithms

Figure 3 Benchmark of adapter-trimming speed for uncompressed and compressed files on different threading options

The 8.9 G bases simulated paired-end data with a 100 bp read length was trimmed in both uncompressed and compressed format using up to 32 threads. Speed is the ratio of

the number of bases to elapsed time (wall time). SeqPurge does not support uncompressed outputs, so it is not shown in the uncompressed benchmark. In the trimming for compressed data, the speed of AdapterRemoval, Skewer, Fastp, Atropos, and Trimmomatic kept constant when the number of threads increased from 4 to 32, so we only benchmark those trimmers using 1, 2, and 4 threads. Ktrim does not support output compressed files, so it is not shown in the compressed benchmark.

Figure 4 Adapter trimming accuracy on adapter presence and absence, different base errors, and adapter lengths

A1, B1, and C1 are statistics for reads with adapter contamination, while A2, B2, C2 for reads without adapters. A1 and A2 show the accumulated rates of accurate trim, one bp over trim, one bp under trim, multiple bp over trim, and multiple bp under trim. In A1, the accuracy of Trimmomatic is 41.0%. In A2, the accuracy of SeqPurge is 78.8%, the accuracy of Trim Galore is 68.3%. B1 and B2 show the trimming accuracy on different error profiles. In B1, the accuracy of Trimmomatic drops from 41.9% to 40.1%. In B2, the accuracy of SeqPurge is 78.8%, and the accuracy of Trim Galore is 68.2 - 68.3%. C1 and C2 show the trimming accuracy on different adapter lengths. In C1, the accuracy of Trimmomatic is 0.0% at 16 bp adapter length, 50.7% to 51.6% at adapter lengths from 20 to 33 bp. In C2, the accuracy of SeqPurge ranges from 78.7% at 16 bp to 78.9% at 33 bp, and the accuracy of Trim Galore ranges in 68.2 - 68.3% from 16 to 33 bp.

Supplementary material

Table S1 Trimming speed on the 8.9 G bases 100 bp paired-end simulated data

Atria (consensus) does both adapter trimming and paired-end consensus call (base correction of overlapped regions). In the trimming for uncompressed data, SeqPurge does not support uncompressed outputs, so it is not shown in the uncompressed benchmark. Fastp does not support 32 threads, so only 1-16 threads were tested. In the trimming for compressed data, the speed of AdapterRemoval, Skewer, Fastp, and Trimmomatic kept constant when the number of threads increased from 4 to 32, so we only benchmarked those trimmers using 1, 2, and 4 threads. Atropos was too slow to trim compressed data, and Ktrim did not support compressed outputs, so they are not shown in the compressed benchmark.

Table 2 Performance of trimmers on real data (larger than A4)

Metric	Trimming and consensus		Trimming only							
	Atria	Skewer	Atria	AR	Atropos	Fastp	Ktrim	SeqPurge	Trim Galore	Trimmomatic
Low-quality dataset (SRR330569, RNA, Hisat2 mapping)										
Elapsed time							1:34 +			
(min:sec)*	2:38	9:19	2:32	11:29	10:08	9:17	GZ	3:53	3:39	9:38

No quality

trimming

Reads

mapped and	26,126	24,694	25,781	<u>24,559</u>	24,505	24,545	24,196	24,240	24,046	22,797
paired	,804	,330	,268	<u>,060</u>	,656	,646	,658	,072	,542	,620
Reads	27,276	28,254	27,379	28,338	28,410	<u>28,350</u>	28,747	28,591	28,647	29,649
unmapped	,761	,804	,299	,455	,022	<u>,294</u>	,248	,442	,873	,287
Properly										
paired reads										
(%)	48.3	45.6	47.6	<u>45.3</u>	45.2	<u>45.3</u>	44.5	44.7	44.2	38.3
Base	2,387,	2,354,	2,322,	2,346,	2,341,	<u>2,344,</u>	2,316,	2,304,	2,317,	2,237,
mapped	212,22	164,04	436,54	791,20	355,82	<u>847,43</u>	915,67	776,51	321,51	846,53
(cigar)	5	1	5	4	2	<u>8</u>	3	4	0	4
Error rate				9.8902	9.8536	9.8683	9.7920	9.7904	9.6994	<u>9.3106</u>
(‰)	7.3952	9.5897	8.1833							

With quality trimming

Reads

mapped and	25,942	25,787	25,728	25,721	25,714	<u>25,725</u>	25,473	25,364	25,654	24,744
paired	,092	,464	,206	,788	,956	<u>,480</u>	,670	,392	,498	,754
Reads	27,245	27,364	27,361	27,369	27,373	<u>27,360</u>	27,556	27,736	27,400	28,064
unmapped	,720	,827	,655	,773	,854	<u>,527</u>	,820	,292	,932	,739
Properly										
paired reads										
(%)	47.9	47.6	47.5	47.5	47.5	47.5	46.9	46.8	47.3	42.3
Base	2,317,	2,316,	2,302,	2,304,	2,304,	<u>2,304,</u>	2,292,	2,263,	2,297,	2,246,
mapped	238,53	981,45	740,46	584,26	325,74	<u>437,24</u>	034,76	465,11	815,43	076,61
(cigar)	6	6	3	9	3	<u>4</u>	2	0	9	8
Error rate				7.9160	7.9141	7.9149	7.9059	7.8787	7.8921	7.5649
(‰)	7.1114	7.7882	<u>7.8902</u>							

High-quality dataset (ERR4695159, cell-free DNA, Bowtie2 mapping)

Elapsed time

1:41 +

(min:sec)*	3:08	11:34	3:03	13:48	13:41	11:29	GZ	<u>4:05</u>	4:34	11:44
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No quality trimming

Reads

mapped and	54,367	54,287	<u>54,324</u>	54,319	54,299	54,322	53,087	54,446	54,218	54,128
paired	,548	,616	<u>,964</u>	,438	,922	,088	,420	,104	,344	,760
Reads	1,094,	1,016,	1,119,	<u>989,33</u>	1,002,	978,66	2,317,	999,09	1,041,	1,094,
unmapped	145	244	103	<u>5</u>	745	5	968	9	005	752
Properly										
paired reads										
(%)	96.8	96.7	<u>96.7</u>	<u>96.7</u>	<u>96.7</u>	<u>96.7</u>	94.1	97.0	96.4	88.6

Base	7,703,	7,700,	7,700,	<u>7,701,</u>	7,700,	7,699,	7,607,	7,512,	7,677,	7,720,
mapped	820,58	134,67	298,21	<u>482,30</u>	749,16	298,00	799,30	839,36	087,84	352,49
(cigar)	5	3	7	<u>2</u>	4	8	6	0	5	3
Error rate				3.8771	3.8834	<u>3.8564</u>	4.3239	3.8007	3.9173	6.1984
(‰)	3.3082	3.8388	3.8724							
With quality trimming										
Reads										
mapped and	54,553	54,526	54,546	54,541	54,539	<u>54,549</u>	53,335	54,608	54,482	54,403
paired	,566	,276	,192	,948	,502	<u>,462</u>	,674	,308	,002	,982
Reads										
unmapped	965,44	984,84	967,91	970,86	973,21	826,42	2,136,	<u>890,88</u>	999,91	914,00
	7	5	7	9	7	4	081	<u>4</u>	8	3
Properly paired reads										
(%)	97.0	97.0	<u>97.0</u>	<u>97.0</u>	<u>97.0</u>	<u>97.0</u>	94.4	97.1	96.8	89.0
Base	7,653,	7,649,	7,646,	7,647,	<u>7,648,</u>	7,646,	7,556,	7,461,	7,625,	7,668,
mapped	879,31	380,21	989,36	893,62	<u>184,19</u>	574,60	468,88	588,48	484,70	777,97
(cigar)	2	8	2	4	<u>6</u>	6	2	2	6	1
Error rate				3.2698	3.2792	<u>3.2634</u>	3.7183	3.2117	3.3109	5.5798
(‰)	2.9547	3.2535	3.2678							

Adapter
Trimming

Consensus Calling of
Overlapped Region

Quality
Trimming

— Atria (consensus)
 — Atria
 — AdapterRemoval
 — Skewer
 — Trim Galore
 — Trimmomatic
— Ktrim
— Atropos
— Fastp
— SeqPurge