How Our Cells Become Our Selves: The Cellular Phylodynamic Biology of Growth and Development

Philip Chodrow¹, Jessica Su², Daniel Lee³, Tahmid Ahmed³, Neil He³, Ruben, De Man³,
Ankur Tiwari³, William Mannherz⁴, Luca Citi⁵, Daniel DiCorpo⁶, James Selib Michaelson^{3,7,8}

¹ Department of Mathematics, University of California, Los Angeles, USA

² Department of Biomedical Engineering, Johns Hopkins University, USA

³ Department of Pathology, Massachusetts General Hospital, USA

⁴ Biological and Biomedical Sciences, Harvard Medical School, Boston MA 02115

⁵ School of Computer Science and Electronic Engineering, University of Essex, UK

⁶ Department of Biostatistics, Boston University School of Public Health, USA

⁷ Department of Surgery, Massachusetts General Hospital, USA

⁸ Department of Pathology, Harvard Medical School, USA

***For correspondence:**

JamesMichaelsonPhD@gmail.com (JSM)

Abstract Our lives begin with 1 cell, then 2, then 4, then the trillion cell adult, comprised of cell lineages, tissues, organs. How does this occur? Examination in numbers of cells, ***N***, ***Cellular Phylodynamics***, revealed two previously unappreciated processes: ***UNI-GROWTH***, the slowing of growth that occurs as we become larger, caused by fewer cells dividing, captured by the ***Universal Mitotic Fraction*** and ***Universal Growth Equations***, with accuracy confirmed for 13 species, including nematodes, mollusks, and vertebrates; and ***ALLO-GROWTH***, the creation of body parts from ***Founder Cells***, captured by the ***Cellular Allometric Growth Equation***, which describes mitotic expansion by ***Cell-Heritable*** change in the ***Cell Cycle Time***. These equations can generate cell lineage approximations, bringing the power of coalescent theory to developmental biology.

Introduction

Our lives begin with 1 fertilized cell, dividing to become 2 cells, then 4, then 8, growing to the trillion-or-so-celled human¹ or the thousand-or-so-celled nematode², comprised of cell lineages, tissues, organs, and anatomical structures, of recognizable size and shape. How does this occur? When considered in aggregate, this appears to us as growth, a process with practical implications in agriculture, aquaculture, and health.³⁻⁶ When considered in detail, this appears to us as embryonic development, whose mechanisms await our decipherment.

Examination of animal growth and development in continuous units of length/volume/mass has yielded quantitative insights, whose meanings have often been unclear. For example, while growth starts out exponentially, it is soon tempered by a decline in rate, and many **density dependent growth equations**, which form **S-shaped** growth curves, have been considered¹⁻⁹, none of which accurately fit the growth of real animals.¹⁰⁻¹² Within the body, the relative growth of tissues, organs, and anatomical structures often appears as straight lines on log-log graphs, although the cause of such **allometric growth** has long been a mystery.^{13,14} **Cellular-Selection**, that is, differential cellular proliferation and death, creates many anatomical structures by outgrowth from the undifferentiated body mass, and molds much of the fine scale detail of anatomy, although how cell division is harnessed to achieve this is obscure.^{15,16} Cell lineages grow from individual **Founder Cells**,^{2,17} although how embryos use mitosis to create these parts,^{18,19} and how these **Founder Cells** undergo **Cell-Heritable** change that characterized each clone,^{16,35} remains unknown. The analysis of cellular diversification, by single cell mRNA expression,^{50,53,54} cell marking,^{21,20,51} and high resolution 4D microscopy,^{18,19,47} has also wrestled with the problem of cell lineage formation.²¹

Here, we report the examination of growth and development in discrete units of numbers of cells, **N**. We call this approach “**Cellular Phylodynamics**”. Our frame of mind is inspired by Stadler, Pybus, and Stumpf,²² who have articulated how **Phylodynamics**, a powerful approach widely used in epidemiology, taxonomy, and population biology²³⁻³⁰ can also be brought to bear in the study of developmental biology. As they stated, “The term ‘phylodynamics’ describes the integration of phylogenetic and population dynamic models to study tree-generating processes”, and that such “**tree-thinking**” ... can be beneficial to the interpretation of empirical data pertaining to the individual cells of multicellular organisms.” As we shall see, the application of such a **Cellular Phylodynamic tree-thinking** approach to the analysis of data on numbers of cells in animals as a whole, **N_w**, and in their parts, **N_p**, does indeed provide new insight into how we grow and develop.

Results

UNI-GROWTH

The problem of body growth and development

For more than two centuries, biologists have appreciated that growth begins exponentially, but then declines in rate, and many expressions to capture such **density dependent growth** have been considered,³⁻¹² none of which accurately captures the growth of real animals.^{11,12} Our goal here is not to search for another equation that fits the growth of animals, but to carry out a **Cellular Phylodynamic Analysis** to examine what actual growth data, in units of numbers of cells, **N**, can tell us about the nature of the growth and development of the body.

The Cellular Phylodynamic Analysis of body growth and development

To carry out **Cellular Phylodynamic Analysis** of the growth of the body as a whole, we assembled values on the numbers of cells, **N_w**, from fertilization until maturity, for nematodes, frogs, chickens, cows, geese, quail, turkeys, zebrafish, seabass, clams, mice, rats, humans, and other animals (FIGURE 1[left], and FIGURES A2, A3, and A5 in APPENDIX). A detailed account of how these values were assembled can be found in the APPENDIX.

The Universal Growth Equation,

$$N_w = \int \ln 2/c * N_w * a^{N_w} b$$

The Universal Mitotic Equation,

$$m = a^{N_w} b$$

FIGURE 1: Growth data and the Universal Growth Equation and Universal Mitotic Fraction Equation for capturing growth from fertilization until maturity

Left: Growth, in units of numbers of cells, **N_w**, from fertilization until maturity, in units of days, **t**. Datapoints for animal size's (**N_x**) versus time (**t**), for humans, chickens, **C. elegans** nematode worms, and mice, and their fit to the numerically integrated form of the **Universal Growth Equation** (#5b). For graphs of the growth of other species, see FIGURE A1, A2, A5, and A6 in APPENDIX.

Right:

Estimate of the decline in the **Mitotic Fraction**, a measure of the fraction of cells dividing that occurs as animals increase in size, as estimated by the **Mitotic Fraction Method** and captured by the **Universal Mitotic Fraction Equation**.

Data points for animal size, **N_w**, in integer units of numbers of cells, vs. the **Mitotic Fraction**, **m**, from fertilization, until maturity, for humans, chickens, mice, and **C. elegans** nematode worms. For additional graphs of the change in the **Mitotic Fraction**, **m**, with size, **N_w**, see FIGURE A2, A3, and A5 in APPENDIX.

Many studies have noted that mitosis occurs in the great majority of cells at the beginning of development, giving early embryos the capacity to increase in size, N_w , with age, t , such that the **rate of growth** is:

$$\frac{dN_w}{dt} = \frac{\ln(2)}{c} \cdot N_w \quad (1)$$

Where N_w is the number of cells in the body as a whole, and c is the **Cell Cycle Time**. Integration shows that when all cells are dividing, growth is exponential:

$$N_w = e^{\left(\frac{\ln(2)}{c}\right) \cdot t} \quad (2)$$

Such exponential growth can be visualized by graphing the log of the numbers of cells, N , against time, that is:

$$\text{Log}(N) = \frac{\ln(2)}{c} \cdot t \quad (2b)$$

Thus, data on the relationship between the log of cell number, $\text{Log}(N_w)$, and time, t , can be characterized at the very beginning of development, when all cells are dividing, to give one a practical measure of the **Cell Cycle Time**, c . (See APPENDIX for details). Values for c for each of the 13 animals noted above can be seen in the TABLE.

This early unrestricted growth soon gives way to a decrease in the fraction of cells dividing, (for references, see APPENDIX) driven by **mitotic quiescence**, the complex biochemical mechanism within cells that prevents mitosis from occurring in response to inhibitory and stimulatory signaling molecules that cells use to communicate.³¹⁻³³ This decrease in mitotic activity that occurs as we become larger, which we call the “**Mitotic Fraction**”, m , can be taken into account by inserting it into Equation #1:

$$\frac{dN_w}{dt} = \frac{\ln(2)}{c} \cdot N_w \cdot m \quad ; \quad \text{where } m = f(N_w) \quad (3)$$

Thus, converting the **exponential growth equation** (#1) into a **density dependent growth equation** (#3). This “**Swiss Army Knife**” of an expression (since $f(N_w)$ can accommodate any change in the rate of growth that might occur as an animal becomes larger) balances the unlimited quality of exponential growth that occurs at the beginning of development with the inevitable reduction in growth rate that occurs as we grow.⁵ The result is a classic **S-shaped** growth curve, such as can be seen with the data we have assembled on growth of the 13 animals listed above (FIGURE 1, and FIGURES A2, A3, and A5 in APPENDIX).

The **Mitotic Fraction Method**

Rearranging Equation #3 allows us to use growth data to characterize how the value of **Mitotic Fraction**, m , changes as animals increase in size, N_w :

$$m(N_w) = \frac{dN_w}{dt} / \left[\frac{\ln(2)}{c} \cdot N_w \right] \quad (3b)$$

We call this approach the “**Mitotic Fraction Method**”. In simple terms, Equation #3b gives us a rough measure how many cells would have to divide to account for the amount of growth that occurs over each period of time. Details for the practical execution of the method, and the limits of its approximation, can be found in the APPENDIX.

The **Universal Mitotic Fraction Equation**

We applied the **Mitotic Fraction Method** (Equation #3b) to the growth data for all of the 13 animals listed above. This revealed exponential growth during the first few mitoses ($m \approx 1$), smoothly transitioning to an accelerating decrease in the fraction of cells dividing as adulthood approaches ($m < 1$) (FIGURE 1[right], FIGURES A4-A6 in APPENDIX). The basis for the relationship between m and N_w was revealed by graphing $\ln(N_w)$ vs. $-\ln(-\ln(m))$, which yielded straight rows of datapoints (FIGURE A5 in APPENDIX), that is:

$$m = a^{(N_w^b)} \quad (4)$$

This expression was found to capture, with high r^2 values, the decrease in the value of the **Mitotic Fraction, m** , that occurs as animals increase in size, N_w , from fertilization until maturity, for all 13 of species noted above (TABLE). For this reason, we call it the “**Universal Mitotic Fraction Equation**” (#4).

The **Universal Growth Equation**

Combining Equations #3 and #4 leads to this expression for describing the rate of animal growth:

$$\frac{dN_w}{dt} = \frac{\ln(2)}{c} \cdot N_w \cdot a^{(N_w^b)} \quad (5a)$$

The relationship between size, N_w , and age, t , is captured by numerical integration of Equation #5, where the age, t , at fertilization is ~ 0 , and the age at death is d :

$$N_w = \int_{t=0}^{t=d} \frac{\ln(2)}{c} \cdot N_w \cdot a^{(N_w^b)} \quad (5b)$$

as well as by this closed-form integration:

$$t = \frac{c}{b \cdot \log(2)} \cdot \{ \text{Ei}[-N_w^b \cdot \log(a)] - \text{Ei}[-\log(a)] \} \quad (5c)$$

where **Ei** is the exponential integral function.³⁴

We tested the numerically integrated form of these expressions, Equation #5b, on body size, N_w , vs age, t , from fertilization until maturity, for all 13 species listed above, finding close fits, i.e., high r^2 values (FIGURE 1[left], TABLE). For this reason, we call these expressions “**Universal Growth Equations**”. Tests of other widely used density dependent growth equations found none to fit actual growth data as well as the **Universal Growth Equation** (FIGURES A3-A5 in APPENDIX). Thus, the **Universal Growth Equation** is not only more accurate than previous growth equations, it also succeeds, as previous equations did not, in capturing growth over its full span, from the first fertilized cell of the embryo up to mature size. We call the biological process captured by the **Universal Mitotic Fraction** and **Universal Growth Equations** “**UNI-GROWTH**”.

Possible mechanisms

The **a** and **b** parameters of the **Universal Growth Equation** determine the “curviness” of the “**S**” of the **S-shaped** growth curve, while its parameter **c**, the **Cell Cycle Time**, expands or contracts the “**S**” like an accordion. The **Universal Mitotic Fraction Equation**, acting within that expression, describes the decline in the **Mitotic Fraction**, **m**, that occurs as we grow, as made specific for each animal by the values of its **a** and **b** parameters. Fortunately, the forms of these expressions give us some hints of how the biochemistry of the cell may make such **UNI-GROWTH** possible.

The **Mitotic Fraction** and mitotic signaling

What might be the mechanism behind the change in the **Mitotic Fraction**, **m**, that occurs as we grow? A tempting possibility emerges from the mathematical analysis of the deceptively simple discrete allocation of ligand molecules among cells (Equations #700-#720, APPENDIX).^{35,36} We examined the consequences for cells growing in a confined space, such as an egg or uterus, if these cells produce inhibitory growth factor molecules. As the number of cells increase, the concentration of the inhibitory growth factor increases as well, and the fraction of cells that have bound one or more of these factors also increases. Most remarkably, the math reveals that the decline in the fraction of cells that had bound one or more of these inhibitory factors occurs by an expression of exactly the same form as the **Universal Mitotic Fraction Equation** (#4). Thus, little more than the binding of growth factor molecules to cells, driven by conventional thermodynamics, is sufficient to account for the decline in the fraction of cells dividing that occurs as we grow, a hypothesis-generating possibility, worthy of experimental analysis.

Cell Cycle Time, **c**, growth, junk DNA.

The **Cell Cycle Time**, **c**, of the 13 animals we have examined here ranges from just 15 minutes for nematodes to about a day for humans (TABLE). Genome size is known to be the main determinant of **Cell Cycle Time**, as first reported by Van't Hof and Sparrow,³⁷ for plant cells, subsequently confirmed for cells from many types of organisms (see APPENDIX for references and details). Apparently, the more DNA a cell has to copy, the longer it takes to divide. Animal genomes are usually comprised principally non-coding DNA,³⁸ mostly transposable elements that are very susceptible to duplication and deletion,³⁹⁻⁴³ and thus capable of providing abundant genetic variation in genome size. This genetic variation in genome size should offer genetic variation in **Cell Cycle Time**, and thus variation in growth rate, which the bitter reality of Darwinian selection could draw upon to allow species to drift to growth rates that give them their greatest chance of survival. Could our much maligned “junk DNA” owe its existence to this important function?

ALLO-GROWTH

The problem of body part growth and development

For more than a century, biologists examining the growth of various parts of the body, p , in comparison to the size of the body as a whole, w , in continuous units of weight, volume, and length, have frequently found straight lines on log-log graphs. This observation is called **allometric growth**.^{13,44,45}

$$\log(w) = \frac{1}{s} \cdot \log(p) + \log(B) \quad (6)$$

The reasons why the growth of the parts of the body should take this form, and the meaning of the parameters of this **Allometric Growth Equation**, have long been mysteries.^{44,45} Our goal here is to carry out a **Cellular Phylodynamic Analysis** to examine what actual growth data, in units of numbers of cells, N , can tell us about the nature of the growth and development of the parts of the body.

The Cellular Phylodynamic Analysis of body part growth and development

To carry out **Cellular Phylodynamic Analysis** of the growth and development of the cell lineages, tissues, organs, and anatomical structures of the body, we assembled values, from the literature, on the numbers of cells in the body, N_w , and in its various parts, N_p . A detailed account of how these values were assembled can be found in the APPENDIX.

The anatomical structures of developing nematodes (*Caenorhabditis elegans*² and *Meloidogyne incognita*¹⁷) and tunicates (*Oikopleura dioica*⁴⁶), who are chordate quite closely related to ourselves, have been characterized for every cell from the first single cell zygote onward. From these cell lineage charts, we assembled values of N_p and N_w (FIGURES 2 and A8-A11 in the APPENDIX).

The tissues, organs, and anatomical structures seen later in development have usually been measured in units of weight or volume. From these batch values, we could generate useful approximations of N_p and N_w (see APPENDIX). We assembled values for chicks (gizzards, livers, hearts, kidneys), rats (livers, brains, kidneys, forelegs, ears, stomachs, spinal cords), humans (brains, livers, kidneys, lungs, pancreases, adrenals, thymuses, spleens, lower extremities, upper extremities, stomach, hearts, intestines), zebrafish (eye lens), mice (livers, brains, kidneys, forelegs), clams, and goldfish (FIGURES 2 and A12-A17 in the APPENDIX).

FIGURE 2: Cellular phylodynamic analysis of cell lineages, tissues organs, and anatomical structures, and their fit to the Cellular Allometric Growth Equation.
Cell Lineages: *M. incognita* (AB and P lineages and its cell lineage chart from which these values were derived). *C. elegans* (AB, E, MS and Germ Cell lineages). The AB lineage of nematodes forms the skin while the E lineage forms the intestine.
Tissues, organs, and anatomical structures of developing fetuses: Chick (heart), Zebrafish (lens), Human (brain), Human (leg).
 For references and additional values for growth of the **cell-lineages** of the nematodes, *M. incognita* and *C. elegans*, and the chordate tunicate *O. dioica*, and for the **tissues, organs, and anatomical structures** of the developing fetuses see FIGURES A9-17 in APPENDIX.

The **Cellular Allometric Growth Equation**

When we placed, on log-log graphs, the values we assembled on the number of cells in cell lineages, tissues, organs, and anatomical structures, of the body, N_p versus the number of cells in the body as a whole, N_w , in the great majority of cases, these datapoints appeared as straight rows of dots (FIGURE 2 and FIGURES A8-A17 in APPENDIX), as attested by high r^2 values, or:

$$\log(N_w) = \frac{1}{S_N} \cdot \log(N_p) + \log(B_N) \quad (7)$$

We call this expression the “**Cellular Allometric Growth Equation**” (FIGURE 2), and the biological process that it captures “**ALLO-GROWTH**”. We call the parameter B_N the “**Cellular Allometric Birth**” and S_N , the “**Cellular Allometric Slope**”.

FIGURE 3: The Cellular Allometric Growth Equation

Cells to Structures: The **Cellular Allometric Birth**, B_N .

The value of B_N , the **Cellular Allometric Birth**, can be seen easily on log-log graphs, as the place where the **Cellular Allometric Growth Equation** crosses the x -axis. Thus B_N corresponds to the value of N_w where $\log(N_p)=0$, which is where $N_p=1$ (FIGURE 3).

Should a part of the body come into existence as a single **Founder Cell**, the **Cellular Allometric Birth**, B_N , corresponds to the number of cells in the body of the embryo, N_w , when that single **Founder Cell** arose by mitosis (i.e., $B_N = N_w$ when $N_p = 1$). In fact, this is precisely what has been seen for those animals for which we have **cell lineage charts**, which have been collected to describe every cell in the embryo from the zygote onward: *Caenorhabditis elegans*² and *Meloidogyne incognita*¹⁷ nematode worms, and *Oikopleura dioica* tunicates,⁴⁶ which are chordates quite closely related to ourselves (FIGURE 2 and FIGURES A9-A11 in APPENDIX).

For the tissues, organs, and anatomical structures for which we have growth data later in development, we do not have data on their cellular origins. There has been much scholarship on this point (reviewed in the APPENDIX), but, fortunately, modern light sheet 4D microscopy should allow us to answer these questions,^{18,19,47} and the **Cellular Allometric Growth Equation** points us to where we should look. Furthermore, whether a body part arises from a single **Founder Cell**, or multiple **Founder Cells**, **Cellular Phylogenetic Analysis** allows us to distill the features of the process. For example, should a part of the body come existence from z **Founder Cells**, each of which is born at the same time, and has the same **Cell-Heritable Cell Cycle Time**, c_p , the part made of these z **Founder Cells** will also grow by the **Cellular Allometric Growth Equation**. Even more complex examples of this type, including the process of **ALLO-GROWTH** in *drosophila* development, are amenable to **Cellular Phylogenetic Analysis**, topics addressed in the APPENDIX.

Body part **ALLO-GROWTH** results from whole body **UNI-GROWTH**.

Recall that the **Cellular Allometric Growth Equation** captures the rate of growth of the whole embryo, while the rate of growth for each body-part is:

$$\frac{dN_p}{dt} = \frac{\ln(2)}{c_p} \cdot N_p \cdot (m_p) \quad (8)$$

where c_p is the **Cell Cycle Time**, and m_p the **Mitotic Fraction** of the cells of the body part. Thus, the ratio of the rate of growth of the body to the body-part is:

$$\frac{dN_w}{dN_p} = \left[\frac{\ln(2)}{c} \cdot N_w \cdot (m) \right] \cdot \left[\frac{\ln(2)}{c_p} \cdot N_p \cdot (m_p) \right]^{-1}, \quad \text{where } m = a^{(N_w^b)} \text{ and } m_p = a_p^{(N_p^{b_p})} \quad (9)$$

Consider the case where $m_p=m$ and $c_p=c$, that is, where a body part has the same **Mitotic Fraction** and **Cell Cycle Time** as the embryo, and starts sometime later than the first cell:

$$\frac{dN_w}{N_w} = \frac{dN_p}{N_p} \quad (10)$$

Integration reveals:

$$\log(N_w) = \log(N_p) + \log(B_N) \quad (7c)$$

which is the allometric relationship, with $S_N=1$. Thus, growth of a part of the body by the **Cellular Allometric Growth Equation** is the consequence of **Founder Cells** giving rise to cell lineages in animals growing by the **Universal Growth Equation**.

The **Cellular Allometric Slope**, S_N , captures **Cell-Heritable** change in the **Cell Cycle Time**, c_p , and body part size change by **Cellular-Selection**.

Now, let us consider the case where $m_p=m$ but $c_p \neq c$, that is, where a body part has the same **Mitotic Fraction** as the embryo but a different **Cell Cycle Time**:

$$\frac{dN_w}{N_w} = \frac{c_p}{c} \cdot \frac{dN_p}{N_p} \quad (11)$$

Integration reveals:

$$\log(N_w) = \frac{c_p}{c} \cdot \log(N_p) + \log(B_N) \quad (7d)$$

where:

$$S_N = \frac{c}{c_p} \quad (12)$$

Thus, S_N , the **Cellular Allometric Slope** of the **Cellular Allometric Growth Equation**, can be traced to the **Cell Cycle Time**, c_p in each part of the body (FIGURE 3)!

On the other hand, similar analysis revealed the **Mitotic Fraction**, m_p , that is, a change in the fraction of cell dividing, to be an unlikely source for allometric log-linearity. Indeed, changes in the value of either **a** or **b** of the **Universal Mitotic Fraction Equation** (#4), which sets the value of m_p , does not result in straight lines on log-log graphs but in curves (not shown).

The changes in the value of c_p seen by the **Cellular Phylodynamic Analysis** of body part growth are small (FIGURE 2 and FIGURES A8-A17 in APPENDIX). Thus, they have a minor impact on the average **Cell Cycle Time**, c , in the embryo as a whole, but can have dramatic impacts on the relative sizes of body parts.

The value of S_N , the **Cellular Allometric Slope** of the **Cellular Allometric Growth Equation**, appears on log-log graphs as a measure of whether a part of the body is growing faster ($S_N > 1$), slower ($S_N < 1$), or at the same speed ($S_N = 1$) as the body as a whole. Thus, **ALLO-GROWTH**, and its mathematical abstraction, the **Cellular Allometric Growth Equation** (#7), distills **Cellular-Selection**, that is, differential cellular proliferation,^{15,16} down to a single value, S_N . Embryos accomplish this by **Cell-Heritable** change in the **Cell Cycle Time**, c_p .

What might be the mechanism of such a **Cell-Heritable** change in the **Cell Cycle Time**, c_p ? DNA methylation has been found to induce **Cell-Heritable** changes in **Cell Cycle Time**⁴⁸, which one might imagine could occur at the **Cellular Allometric Birth**, B_N , of **Founder Cells**, a hypothesis worthy of experimental examination.

CELLULAR POPULATION TREE VISUALIZATION SIMULATION

The reader will note that we have extracted information on the relationship between the number of cells in the body as a whole, N_w , with age, t , from the cell lineage charts of nematodes (FIGURE 1). However, even without full lineage chart information, it is possible to reconstruct a plausible approximation of an animal's cell lineage chart by a simulation which uses the growth equations of **Cellular Phylodynamics** and their parameters a , b , c , B_N , and S_N , extracted from batch growth data (for method, see APPENDIX).

In the FIGURE 4 (left) we display the idealized case of the cell lineage chart of an animal growing by the **Universal Growth Equation**. Note how, as time goes on, an increasing fraction of cells, the **Mitotic Fraction**, m , don't divide, leading to some of the branches of the tree become extended in the absence of mitosis.

The **Universal Growth** and **Cellular Allometric Growth Equations** can also be taken to together, with their various parameters, to reconstruct an even more likely chart of an animal's estimated cell lineage. In FIGURE 4 (right) we display the idealized case of the cell heritable shortening of the **Cell Cycle Time** that occurs in one **Founder Cell** when the embryo is 16 cells in size (**Cellular Allometric Birth**, $B_N=16$); note how the progeny of the **Founder Cell** increased in number in comparison to the rest of the embryo.

FIGURE 4 Cellular Population Tree Visualization Simulations.

Left,

The idealized case of the cell lineage chart of an animal growing by the **Universal Growth Equation**. Note how, as time goes on, an increasing fraction of cells, the **Mitotic Fraction**, m , don't divide, and some of the branches of the tree become extended in the absence of mitosis.

Right,

The idealized case of the cell heritable shortening of the **Cell Cycle Time** that occurs in one **Founder Cell** when the embryo is 16 cells in size (**Cellular Allometric Birth**, $B_N=8$); note how the progeny of the **Founder Cell** increased in number in comparison to the rest of the embryo.

Code is executed in JAVA (see APPENDIX). Visualizations were made with IcyTree.⁵⁷ Future generations of such simulations may well benefit for the stochastics simulator,⁵⁸ located within BEAST.⁵⁹

Discussion

The **Cellular Phylodynamic Analysis** reported here, that is, the analysis of growth and development in units of numbers of cells, N , has revealed two striking, and previously unappreciated aspects of embryonic life: **UNI-GROWTH** and **ALLO-GROWTH**.

* **UNI-GROWTH** is the process by which growth slows as we increase in size. **UNI-GROWTH** is captured by the **Universal Mitotic Fraction** and **Universal Growth Equations**, and their parameters, a , b , and c , the average **Cell Cycle Time**. **UNI-GROWTH** occurs by the decline in the fraction of cells dividing, the **Mitotic Fraction**, occurring by the form of **Universal Mitotic Fraction Equation**, $m=a^{(N^b)}$. From this expression, we could then derive, and test, the **Universal Growth Equation**, whose accuracy was confirmed, from fertilization until maturity, for 13 species, including nematodes, mollusks, amphibians, fish, birds, rodents, cows, and humans.

* **ALLO-GROWTH** is the process by which body parts are created from **Founder Cells**. **ALLO-GROWTH** is captured by the **Cellular Allometric** and **Allometric Growth Equations**, and their parameters, S , B , S_N , the **Cellular Allometric Slope**, and B_N , the **Cellular Allometric Birth**. **ALLO-GROWTH** occurs by a **Founder Cell** acquiring a **Cell-Heritable, Cell Cycle Time**, c_p at the **Founder Cell's Cellular Allometric Birth**, B_N , and then undergoing mitotic expansion, as captured by the **Cellular Allometric Slope**, S_N , of the **Cellular Allometric Growth Equation**. Whether a body part arises from a single **Founder Cell**, or multiple **Founder Cells** arising at the same time with the same **Cell Cycle Time**, growth by the **Cellular Allometric** and **Allometric Growth Equations** will occur. For body parts arising from a single **Founder Cell**, the **Cellular Allometric Birth**, B_N , corresponds to the number of cells in the body when the **Founder Cell** arose.

For more than a century, biologists have appreciated that as we grow bigger, we also grow slower, that is, **density dependent growth**, and that, the relative growth of parts of the body often form straight lines on log-log graphs, that is, **allometric growth**.^{3,6,7,13} **Cellular Phylodynamic Analysis** captured these two observations in **UNI-GROWTH** and **ALLO-GROWTH**, and uncovered the surprising finding that they are really different manifestations of the same process, since the **Cellular Allometric Growth Equation** can be derived from the **Universal Growth Equation**.

UNI-GROWTH and **ALLO-GROWTH**, capture, in mathematical form, **Cellular-Selection**, that is, differential cellular proliferation.^{15,16} The **Cellular Allometric Slope**, S_N , of the **Cellular Allometric Growth Equation** captures **Cell-Selection** in the increase or decrease in the size of a body part, and traces its origin to the **Cell Cycle Time**, c_p . The **Mitotic Fraction**, m , of the **Universal Mitotic Fraction Equation** captures **Cell-Selection** in the restriction of cell division to a selected few cells. The **Cellular Allometric Birth**, B_N , of the **Cellular Allometric Growth Equation** captures **Cell-Selection** in the choice of **Founder Cells** among all the other cells of the embryo. Thus, these **Cellular Phylodynamic** equations, and their parameters, provide quantitative metrics for **Cellular-Selection** in its role in the creation of anatomical structures by outgrowth from the undifferentiated body mass, and in the molding of the fine scale detail of anatomy, processes that lie at the heart of embryonic development.^{15,16}

ALLO-GROWTH also captures, in mathematical form, **Cell-Heritability**, the persistence of phenotypic characteristics after cell division.^{15,16} The **Cellular Allometric Slope**, S_N , of the **Cellular Allometric Growth Equation** captures the **Cell-Heritable** change in the **Cell Cycle Time**, c_p , that occurs at the **Cellular Allometric Birth**, B_N , of the **Founder Cell**. However, the **Cellular Phylodynamic Analysis** of growth and development in units of numbers of cells, N , that we describe here has not yet told us how **Founder Cells** acquire such a **Cell-Heritable** change. Of course, other cellular processes also display **Cell-Heritability**, such as gene expression, cell size, and the geometry of each cell division, and how such programs of mitotic geometry contribute to the shape of the animal.⁴⁹ Details for how the **Cellular Phylodynamic Analysis** of such processes could be carried out computationally are presented in the APPENDIX, where we outline a technique, the **BinaryCellName** method, for analysis of cell lineages at the single cell level, and another technique, the **S_{N3} Method**, for analysis of the cellular basis of shape formation at the macroscopic level.

Growth is critical to the survival and evolution of individuals and species. Human fetuses that grow to average birthweight in 8 months, or 10 months, suffer a heart-breaking increase in its risk of disability and death. In the APPENDIX, we show how **Cellular Phylodynamic** mathematics can provide equations for the assessment and management of human fetal size and growth. **Life History Theory** teaches that in nature the struggle for existence includes growing faster or slower or bigger or smaller than competitors, causing animals to acquire growth curves molded by the cruel disciple of Darwinism.^{29,30} The **Universal Growth Equation**, being linked to the developmental biology and genetics of the creatures that must adapt, gives us a tool for achieving ever greater biological reality in our understanding of the impact of individual growth on populations.

Growth and development are not only crown fascinations of embryology, they also present practical challenges in agriculture and aquaculture. In the APPENDIX, we outline how the **Cellular Phylodynamic Analysis** can be put to work in breeding fish with optimal growth characteristics for use in the aquaculture industry.

We have seen that **Cellular Phylodynamic Analysis** can be carried out with cell numbers, N_p and N_w , counted from in cell lineage charts. From the number of these cells, we could identify the form and parameters of the **Universal Mitotic Fraction**, **Universal Growth**, and **Cellular Allometric Growth Equations**. We have also seen that we can reverse this line of reasoning, so that even in the absence of such lineage charts, useful approximations of cell lineage can be constructed by simulation, utilizing the parameters of these equations extracted from batch growth data. Following the thinking of Stadler, Pybus, and Stumpf,²² we call these “**Cellular Population Tree Visualization Simulations**”, which should bring the powerful mathematics of lineage formation, coalescent theory,²²⁻²⁷ to developmental biology. For example, there has been much interest in the past few years in extracting information on gene expression, differentiation, and lineage formation, from single cell RNA expression,^{50,53,54} cellular barcoding,^{21,51} and somatic mutation.⁵² However, as Telford²¹ and Klein^{53,54} have pointed out, without knowledge of the mitotic pattern of an animal's cell lineage, extraction of useful information from such single cell data is problematic. One might hope, and indeed expect, that the potential of **Cellular Population Tree Visualization Simulation**, driven by the parameters measured by **Cellular Phylodynamic Analysis**, could unburden single cell analysis from the requirement of deciphering ancestry, and unlock the much hoped-for treasure of how gene expression is linked to cell lineage formation.

The ability of cells to slow down mitosis, so as to glide to ideal body size, and gear up mitosis, so as to create useful parts, are essential qualities of animal life. By the simple examination in units of numbers of cells, ***N***, we have seen these qualities in ***UNI-GROWTH*** and ***ALLO-GROWTH***, and their defining abstractions, the ***Universal Mitotic Fraction***, ***Universal Growth***, and ***Cellular Allometric Growth Equations***, in animals as diverse as tunicates, vertebrates, mollusks, and nematodes. The forms of these equations provide hints of their biochemical mechanisms, and their broad phylogeny suggest invention at the dawn of animal life.^{55,56} If so, perhaps ***Cellular Phylodynamic Analysis*** is giving us a peek at the mysterious billion-year-old spectacle of how our cells become our selves.

TABLE
Parameters and goodness-of-fit metrics to the
Universal Growth and ***Universal Mitotic Fraction Equations***.

Organism	Genus species	Fit of the data to the rate of growth $m=N^*a^*(N^b)$ (c =Cell Cycle Time, in days)					Fit of the data to growth $N=[\log(2)/c]^*N^*a^*(N^b)$
		c	a	b	r^2 (linear fit)	r^2 (log fit)	r^2 (log fit)
Humans (♂)	<i>Homo sapiens</i>	1.016	0.943	0.169	89.9%	95.0%	99.5%
Frogs	<i>Rana pipiens</i>	0.070	0.925	0.269	57.0%	82.3%	97.3%
Nematodes	<i>C. elegans</i>	0.011	0.946	0.680	66.7%	91.6%	99.8%
Chickens	<i>Gallus gallus</i>	0.127	0.906	0.166	80.8%	97.8%	99.8%
Cows	<i>Bos taurus</i>	0.921	0.882	0.128	76.8%	95.1%	100.0%
Geese	<i>Anser anser</i>	0.048	0.898	0.170	77.0%	92.9%	97.1%
Mice	<i>Mus musculus</i>	0.508	0.990	0.281	65.0%	96.2%	99.7%
Quail	<i>Colinus virginianus</i>	0.292	0.795	0.126	76.5%	95.3%	94.2%
Rats	<i>Rattus norvegicus</i>	0.417	0.950	0.197	66.5%	90.6%	99.5%
Clams	<i>M. mercenaria</i>	0.033	0.962	0.275	79.0%	91.0%	99.5%
Turkeys	<i>Meleagris gallopavo</i>	0.028	0.757	0.122	65.3%	92.2%	99.7%

Zebrafish (*Danio rerio*) and European sea bass (*Dicentrarchus labrax*) also show these features of growth (data not shown)

FIGURE LEGENDS

FIGURE 1: Growth data and the *Universal Growth Equation* and *Universal Mitotic Fraction Equation* for capturing growth from fertilization until maturity

Left: Growth, in units of numbers of cells, N_w , from fertilization until maturity, in units of days, t .

Datapoints for animal size's (N_x) versus time (t), for *C elegans*, chickens, mice, turkeys, quail, geese, frogs, and humans, and their fit to the numerically integrated form of the ***Universal Growth Equation*** (#5b). For graphs of the growth of other species, see FIGURE A1, A2, A5, and A6 in APPENDIX.

Right: Estimate of the decline in the ***Mitotic Fraction***, a measure of the fraction of cells dividing that occurs as animals increase in size, as estimated by the ***Mitotic Fraction Method*** and captured by the ***Universal Mitotic Fraction Equation***. Data points for animal size, N_w , in integer units of numbers of cells, vs. the ***Mitotic Fraction***, m , from fertilization, until maturity, for humans, chickens, *C elegans* nematode worms, and mice. For additional graphs of the change in the ***Mitotic Fraction***, m , with size, N_w , see FIGURE A2, A3, and A5 in APPENDIX.

FIGURE 2: *Cellular Phylodynamic Analysis* of cell lineages, tissues organs, and anatomical structures, and their fit to the *Cellular Allometric Growth Equation*.

Cell lineages: *M incognita* (AB and P lineages and its cell lineage chart from which these values were derived). *C elegans* (AB, M, E, MS and Germ Cell lineages). The AB lineage of nematodes forms the skin while the E lineage forms the intestine.

Tissues, organs, and anatomical structures of developing fetuses: Chick (heart), Zebrafish (lens), Human (brain), Human (leg)).

For references and additional values for growth of the cell lineages of the nematodes, *M incognita* and *C elegans*, and the chordate tunicate *O dioica*, and for the tissues, organs, and anatomical structures of the developing fetuses see FIGURES A9-17 in APPENDIX.

FIGURE 3: The *Cellular Allometric Growth Equation*.

FIGURE 4: Cellular *Population Tree Visualization Simulations*.

Left, the idealized case of the cell lineage chart of an animal growing by the ***Universal Growth Equation***. Note how, as time goes on, an increasing fraction of cells, the ***Mitotic Fraction***, m , don't divide, and some of the branches of the tree become extended in the absence of mitosis.

Right, the idealized case of the cell heritable shortening of the ***Cell Cycle Time*** that occurs in one ***Founder Cell*** when the embryo is 16 cells in size (***Cellular Allometric Birth***, $B_N=16$); note how the progeny of the ***Founder Cell*** increased in number in comparison to the rest of the embryo.

Code is executed in JAVA (see APPENDIX). Visualizations made with IcyTree⁵⁷. Future generations of such simulations may well benefit for the stochastics simulator⁵⁸ located within BEAST.⁵⁹

The Universal Growth Equation,

$$N_w = \int \ln 2 / c * N_w * a^{N_w} b$$

The Universal Mitotic Equation,

$$m = a^{N_w} b$$

FIGURE 1: Growth data and the *Universal Growth Equation* and *Universal Mitotic Fraction Equation* for capturing growth from fertilization until maturity

Left: Growth, in units of numbers of cells, N_w , from fertilization until maturity, in units of days, t .

Datapoints for animal size's (N_x) versus time (t), for *C elegans*, chickens, mice, turkeys, quail, geese, frogs, and humans, and their fit to the numerically integrated form of the *Universal Growth Equation* (#5b). For graphs of the growth of other species, see FIGURE A1, A2, A5, and A6 in APPENDIX.

Right: Estimate of the decline in the *Mitotic Fraction*, a measure of the fraction of cells dividing that occurs as animals increase in size, as estimated by the *Mitotic Fraction Method* and captured by the *Universal Mitotic Fraction Equation*. Data points for animal size, N_w , in integer units of numbers of cells, vs. the *Mitotic Fraction*, m , from fertilization, until maturity, for humans, chickens, *C elegans* nematode worms, and mice. For additional graphs of the change in the *Mitotic Fraction*, m , with size, N_w , see FIGURE A2, A3, and A5 in APPENDIX.

FIGURE 2: Cellular Phylodynamic Analysis of cell lineages, tissues organs, and anatomical structures, and their fit to the **Cellular Allometric Growth Equation**.

Cell lineages: ***M incognita*** (AB and P lineages and its cell lineage chart from which these values were derived). ***C elegans*** (AB, M, E, MS and Germ Cell lineages). The AB lineage of nematodes forms the skin while the E lineage forms the intestine.

Tissues, organs, and anatomical structures of developing fetuses: Chick (heart), Zebrafish (lens), Human (brain), Human (leg)).

For references and additional values for growth of the cell lineages of the nematodes, ***M incognita*** and ***C elegans***, and the chordate tunicate ***O dioica***, and for the tissues, organs, and anatomical structures of the developing fetuses see FIGURES A9-17 in APPENDIX.

FIGURE 4 Cellular Population Tree Visualization Simulations.

Left,

The idealized case of the cell lineage chart of an animal growing by the **Universal Growth Equation**. Note how, as time goes on, an increasing fraction of cells, the **Mitotic Fraction, m** , don't divide, and some of the branches of the tree become extended in the absence of mitosis. We have treated the **Mitotic Fraction, m** , as a random variable, a plausible assumption simply for not knowing which cells are dividing at each moment in time, and a possible biological process, if the **Mitotic Fraction** is actually found to be the result of the discrete allocation of ligand molecule among cells, as indicated by the mathematical analysis of such a process (Equations #700-#720, APPENDIX).

Right,

The idealized case of the cell heritable shortening of the **Cell Cycle Time** that occurs in one **Founder Cell** when the embryo is 16 cells in size (**Cellular Allometric Birth, $B_N=8$**); note how the progeny of the **Founder Cell** increased in number in comparison to the rest of the embryo.

Code is executed in JAVA (see APPENDIX). Visualizations were made with IcyTree.⁵⁷ Future generations of such simulations may well benefit for the stochastics simulator,⁵⁸ located within BEAST.⁵⁹

Acknowledgments

We thank Alain Goriely, Derek Moulton, Aris Papageorgiou, Jose Villar, Stephen Kennedy, Eleonora Staines Urias, Henning Tiemeier, and Hari Shroff for many useful suggestions and discussions, and Alex Gould for kindly sharing data.

References

- 1 Sender R, Fuchs S, Milo R. Revised Estimates for the Number of Human and Bacteria Cells in the Body. PLoS Biol. 2016 Aug 19;14(8):e1002533
- 2 Sulston JE, Schierenberg E, White JG, Thomson JN. The embryonic cell lineage of the nematode *Caenorhabditis elegans*. Dev Biol. 1983 Nov;100(1):64-119.
- 3 Verhulst, P.-F. "Deuxième mémoire sur la loi d'accroissement de la population." Mém. de l'Académie Royale des Sci., des Lettres et des Beaux-Arts de Belgique 20, 1-32, 1847
- 4 Smil, V., Growth: From Microorganisms to Megacities 2019, The MIT Press
- 5 Goriely A The mathematics and mechanics of biological growth New York Springer 2017.
- 6 Wright S. Book review. Jour AmStat Assoc. 1926;21:494
- 7 Bertalanffy, L. von, (1934). Untersuchungen über die Gesetzlichkeit des Wachstums. I. Allgemeine Grundlagen der Theorie; mathematische und physiologische Gesetzlichkeiten des Wachstums bei Wassertieren. Arch. Entwicklungsmech., 131:613-652
- 8 Richards, F. J. (1959). A Flexible Growth Function for Empirical Use. Journal of Experimental Botany 10 (2): 290–300
- 9 West GB, Brown JH, Enquist BJ. A general model for ontogenetic growth. Nature. 2001 Oct 11;413(6856):628-31.
- 10 Knight W. Asymptotic Growth: An Example of Nonsense Disguised as Mathematics Journal of the Fisheries Research Board of Canada, 1968, 25(6): 1303-1307, 10.1139/f68-114
- 11 Pardo, S. A., Cooper, A. B., Dulvy, N. K. Avoiding fishy growth curves. Methods in Ecology and Evolution. Volume 4, Issue 4, pages 353–360, April 2013
- 12 Hirst AG, Forster J. When growth models are not universal: evidence from marine invertebrates. Proc Biol Sci. 2013 Aug 14;280(1768):20131546.
- 13 Huxley, Julian S. Problems of Relative Growth. Methuen And Company Limited. 1932.
- 14 Nijhout HF, McKenna KZ. Allometry, Scaling and Ontogeny of Form-An Introduction to the Symposium. Integr Comp Biol. (2019)
- 15 Michaelson J. Cell Selection in the Genesis of Multicellular Organization Lab Invest. 69: 136-150 1993.
- 16 Michaelson J. Cell selection in development. Biological Reviews 1987; 62:115-139.
- 17 Calderón-Urrea A, Vanholme B, Vangestel S, Kane SM, Bahaji A, Pha K, Garcia M, Snider A, Gheysen G. Early development of the root-knot nematode *Meloidogyne incognita*. BMC Dev Biol. 2016 Apr 28;16:10
- 18 Christensen RP, Bokinsky A, Santella A, Wu Y, Marquina-Solis J, Guo M, Kovacevic I, Kumar A, Winter PW, Tashakkori N, McCreedy E, Liu H, McAuliffe M, Mohler W, Colón-Ramos DA, Bao Z, Shroff H Untwisting the *Caenorhabditis elegans* embryo. Elife. 2015 Dec 3;4. pii: e10070

- 19 Santella A, Catena R, Kovacevic I, Shah P, Yu Z, Marquina-Solis J, Kumar A, Wu Y, Schaff J, Colón-Ramos D, Shroff H, Mohler WA, Bao Z. WormGUIDES: an interactive single cell developmental atlas and tool for collaborative multidimensional data exploration. BMC Bioinformatics. 2015 Jun 9;16:189.
- 20 Griffin R, Binari R, Perrimon N. Genetic odyssey to generate marked clones in Drosophila mosaics. Proc Natl Acad Sci U S A. Apr 1;111(13):4756-63 (2014)
- 21 Salvador-Martínez I, Grillo M, Averof M, Telford MJ. Is it possible to reconstruct an accurate cell lineage using CRISPR recorders? Elife. 2019 Jan 28;8:e40292
- 22 Stadler T, Pybus OG, Stumpf MPH. Phylodynamics for cell biologists. Science. 2021 Jan 15;371(6526)
- 23 Grenfell BT, Pybus OG, Gog JR, Wood JLN, Daly JM, Mumford JA, Holmes EC. Unifying the epidemiological and evolutionary dynamics of pathogens. Science. 2004 Jan 16;303(5656):327-32.
- 24 Smadbeck P, Stumpf MPH. Coalescent models for developmental biology and the spatio-temporal dynamics of growing tissues J R Soc Interface. 2016 Apr;13(117):20160112.
- 25 Kingman J F. Origins of the coalescent. 1974-1982 Genetics. 2000 Dec;156(4):1461-3.
- 26 N. F. Müller, D. A. Rasmussen, T. Stadler. The structured coalescent and its approximations. Mol. Biol. Evol. 34, 2970–2981 (2017).
- 27 V. Boskova, S. Bonhoeffer, T. Stadler, Inference of epidemiological dynamics based on simulated phylogenies using birth-death and coalescent models. PLOS Comput. Biol. 10, e1003913 2014
- 28 Edward O. Wilson and William H. Bossert A Primer of Population Biology 1971
- 29 May RM Limit cycles in predator-prey communities. Science. 1972 Sep 8;177(4052):900-2
- 30 Neal D. Introduction to Population Biology, 2nd Edition Cambridge University Press; 2nd edition 2019
- 31 O'Farrell PH. Quiescence: early evolutionary origins and universality do not imply uniformity. Philos Trans R Soc Lond B Biol Sci. 2011 Dec 27;366 (1584):3498-507.
- 32 Hartwell LH- Nobel Lecture: Yeast and Cancer. Nobelprize.org. Nobel Media AB 2014. Web. 13 Sep 2015. <http://www.nobelprize.org/nobel_prizes/medicine/laureates/2001/hartwell-lecture.html>
- 33 Bertoli C, Skotheim JM, de Bruin RA. Control of cell cycle transcription during G1 and S phases. Nat Rev Mol Cell Biol. 2013 Aug;14(8):518-28. doi: 10.1038/nrm3629.
- 34 https://en.wikipedia.org/wiki/Exponential_integral
- 35 Guillemin A, Stumpf MPH Noise and the molecular processes underlying cell fate decision-making Phys Biol 2021 Jan 7;18(1):011002
- 36 Michaelson, J. The Role of Molecular Discreteness in Normal and Cancerous Growth. Anticancer Research 19: 4853-4868 (1999)
- 37 J. Van't Hof and A. H. Sparrow A relationship between DNA content, nuclear volume, and minimum mitotic cycle time Proc Natl Acad Sci U S A. 1963 Jun; 49(6): 897–902
- 38 Orgel LE, Crick F. Selfish DNA: the ultimate parasite. Nature. 1980 Apr 17;284(5757):604-7.
- 39 Moffat AS. Transposons help sculpt a dynamic genome. Science. 2000 Sep 1;289(5484):1455-7.
- 40 Barabas O. Snapshots of a genetic cut-and-paste. Nature. 2019 Nov;575(7783):447-448.
- 41 Bourque G., Burns KH, Gehring M, Gorbunova V, Seluanov A, Hammell M, Imbeault M, Izsvák Z, Levin HL, Macfarlan TS, Mager DL, Feschotte C. Ten things you should know about transposable elements. Genome Biol. 2018 Nov 19;19(1):199.

- 42 Petrov DA, Sangster TA, Johnston JS, Hartl DL, Shaw KL. Evidence for DNA loss as a determinant of genome size. *Science*. 2000 Feb 11;287(5455):1060-2.
- 43 Nam K, Ellegren H. Recombination drives vertebrate genome contraction. *PLoS Genet*. 2012;8(5):e1002680. 2012.
- 44 Gould SJ. Geometric Similarity in Allometric Growth: A Contribution to the Problem of Scaling in the Evolution of Size. *The American Naturalist*, Vol. 105, No. 942 pp. 113-136 Mar - Apr. 1971
- 45 White JF Gould SJ. Interpretation of the Coefficient in the Allometric Equation *The American Naturalist* Vol. 99, No. 904 pp. 5-18 Jan. - Feb., 1965
- 46 Stach T, Anselmi C. High-precision morphology: bifocal 4D-microscopy enables the comparison of detailed cell lineages of two chordate species separated for more than 525 million years. *BMC Biol*. 2015 Dec 23;13:113.
- 47 Wan Y, McDole K, Keller PJ. Light-Sheet Microscopy and Its Potential for Understanding Developmental Processes. *Annu Rev Cell Dev Biol*. 2019 Oct 6;35:655-681.
- 48 Desjobert C, El Maï M, Gérard-Hirne T, Guianvarc'h D, Carrier A, Pottier C, Arimondo PB, Riond J. Combined analysis of DNA methylation and cell cycle in cancer cells. *Epigenetics*. 2015;10(1):82-91.
- 49 Leung, B., Hermann, G.J., and Priess, J.R. (1999). Organogenesis of the *Caenorhabditis elegans* intestine. *Dev. Biol*. 216, 114–134
- 50 Cao J et al. A human cell atlas of fetal gene expression. *Science*. 2020 Nov 13;370(6518): eaba7721.
- 51 Leeper K et al. Lineage barcoding in mice with homing CRISPR. *Nat Protoc* . 2021 Apr;16(4):2088-2108
- 52 Chapman MS et al. Lineage tracing of human development through somatic mutations *Nature*. 2021 Jul;595 (7865):85-90.
- 53 Weinreb C, Klein AM. Lineage reconstruction from clonal correlations. *Proc Natl Acad Sci U S A*. 2020 Jul 21;117(29):17041-17048.
- 54 Weinreb C, Wolock S, Tusi BK, Socolovsky M, Klein AM. Fundamental limits on dynamic inference from single-cell snapshots. *Proc Natl Acad Sci U S A*. 2018 Mar 6;115(10):E2467-E2476.
- 55 Turner EC. Possible poriferan body fossils in early Neoproterozoic microbial reefs. *Nature*. 2021 Aug;596(7870):87-91. Epub 2021 Jul 28.
- 56 Watson T. These bizarre ancient species are rewriting animal evolution. *Nature*. 2020 Oct;586(7831):662-665.
- 57 Vaughan TG. IcyTree: rapid browser-based visualization for phylogenetic trees and networks. *Bioinformatics*. 2017 Aug 1;33(15):2392-2394.
- 58 Vaughan TG, Drummond AJ. A stochastic simulator of birth-death master equations with application to phylodynamics. *Mol Biol Evol*. 2013 Jun;30(6):1480-93.
- 59 Bouckaert R et al. BEAST 2.5: An advanced software platform for Bayesian evolutionary analysis *PLoS Comput Biol*. 2019 Apr 8;15(4):e1006650