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Population replacement gene drive characteristics for malaria elimination in a range of seasonal transmission settings: a modeling study

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20 Short title

21 Replacement gene drives for malaria elimination: a modeling study

22 Abstract

23 Genetically engineering mosquitoes is a promising new vector control strategy to reinvigorate the fight
24 against malaria in Sub-Saharan Africa. Using an agent-based model of malaria transmission with vector
25 genetics, we examine the impacts of releasing population-replacement gene drive mosquitoes on
26 malaria transmission and quantify the gene drive system parameters required to achieve local
27 elimination within a spatially-resolved, seasonal Sahelian setting. We evaluate the performance of two
28 different gene drive systems: “classic” and “integral.” Various transmission regimes (low, moderate, and
29 high - corresponding to annual entomological inoculation rates of 10, 30, and 80 infectious bites per
30 person) and other simultaneous interventions, including deployment of insecticide-treated nets (ITNs)
31 and passive healthcare seeking, are also simulated. Local elimination probabilities decreased with pre-
32 existing population target site resistance frequency, increased with transmission-blocking effectiveness
33 of the introduced antiparasitic gene and drive efficiency, and were context dependent with respect to
34 fitness costs associated with the introduced gene. Of the four parameters, transmission-blocking
35 effectiveness may be the most important to focus on for improvements to future gene drive strains
36 because a single release of classic gene drive mosquitoes is likely to locally eliminate malaria in low to
37 moderate transmission settings only when transmission-blocking effectiveness is very high (above ~80-
38 90%). However, simultaneously deploying ITNs and releasing integral rather than classic gene drive
39 mosquitoes significantly boosts elimination probabilities, such that elimination remains highly likely in
40 low to moderate transmission regimes down to transmission-blocking effectiveness values as low as
41 ~50% and in high transmission regimes with transmission-blocking effectiveness values above ~80-90%.
42 Thus, a single release of currently achievable population replacement gene drive mosquitoes, in
43 combination with traditional forms of vector control, can likely locally eliminate malaria in low to
44 moderate transmission regimes within the Sahel. In a high transmission regime, higher levels of
45 transmission-blocking effectiveness than are currently available may be required.

46 Author summary

47 Malaria remains a significant health burden in Sub-Saharan Africa. The mass deployment of insecticide-
48 treated nets and antimalarial drugs have drastically reduced malaria incidence, but insecticide and drug
49 resistance threaten to stall these efforts. The genetic engineering of mosquito populations is a promising
50 new vector control strategy to reinvigorate the fight against malaria. Releases of engineered gene drive
51 mosquitoes that can spread introduced antimalarial genes quickly throughout the mosquito population
52 may be a particularly effective new method for reducing malaria transmission. Important questions
53 arise, however, about how well these gene drive systems must work in order to deliver substantial
54 reductions in transmission. Here we use a spatial model of individual humans and vectors to simulate
55 the effects of releasing gene drive mosquitoes with antimalarial properties on malaria transmission in a
56 Sahelian setting. We quantify the gene drive system parameters required to achieve local elimination
57 and find that when deployed in combination with traditional forms of vector control, a single release of
58 gene drive mosquitoes with realistically achievable characteristics is highly likely to locally eliminate

59 malaria in low to moderate transmission regimes. In a high transmission regime, improved strains of
60 gene drive mosquitoes may be required. In all settings, releasing gene drive mosquitoes with
61 antimalarial properties helps create a window of opportunity during which malaria prevalence is
62 suppressed and other interventions can be ramped up to achieve elimination, even when a single gene
63 drive mosquito release by itself cannot.

64 Introduction

65 Malaria remains a significant health burden in Sub-Saharan Africa (SSA) despite many decades of effort
66 to eliminate the disease [1]. More recently, since the early 2000s, the scale up and mass deployment of
67 long-lasting insecticide-treated nets, indoor residual spraying of insecticides, and antimalarial drugs have
68 drastically reduced malaria incidence [2]. However, this existing set of tools is unlikely to bring about
69 eradication [[3]]. Drug and insecticide resistance further threaten to stall these malaria control efforts
70 [4–7]. New strategies and technologies will therefore be needed to achieve elimination in SSA. The
71 genetic engineering of mosquito populations is a promising new vector control strategy to reinvigorate
72 the fight against malaria and potentially lead to elimination.

73 Indeed, releases of genetically modified (GM) sterile male mosquitoes have been used to successfully
74 suppress *Aedes aegypti* vector populations [8–12]. This method is expensive, however, and requires
75 frequent, large-scale releases, known as inundation. Releases of GM gene drive mosquitoes, in contrast,
76 are predicted to be a cost-effective and longer-lasting alternative requiring far fewer and smaller
77 releases [13,14]. Mosquitoes engineered with gene drive systems can copy specified genes from one
78 chromosome to another in germline cells, ensuring that these genes are passed onto their offspring at
79 higher than Mendelian inheritance rates and therefore rapidly spread through a population even if there
80 are associated fitness costs [15].

81 Gene drive mosquito releases can either aim to reduce (population suppression) or to modify
82 (population replacement) a given vector population [15]. Population replacement gene drive systems
83 are the focus of this study and they consist of a driver gene that enables the copying of both itself and
84 an effector gene, which in turn confers desired phenotypic traits. The driver gene encodes a guide RNA
85 and an endonuclease, such as Cas9, that together recognize and cut specified DNA sequences present in
86 the wildtype mosquito population. Within mosquitoes that are heterozygous for the wildtype and drive
87 or effector alleles, the cut wildtype chromosome uses its intact drive or effector-containing sister
88 chromosome as a template for repairing itself, copying over the intact chromosome's drive or effector-
89 containing DNA in the process through homology-directed repair (HDR) [16].

90 Population replacement may be desirable in locations where the ecological effects of removing a
91 mosquito species are not well known. For population replacement applied to malaria reduction or
92 elimination, many potential effector genes have been shown to impair development of *Plasmodium*
93 parasites by *Anopheles* mosquitoes. These include genes that code for immune system activators,
94 peptides that neutralize *Plasmodium* parasites in the mosquito midgut or salivary glands, and others
95 [17–23].

96 A number of important questions about population replacement drives require further investigation.
97 How effective do these effector genes have to be in order to deliver substantial reductions in malaria
98 transmission? Can elimination be achieved even with imperfect transmission blocking traits? If there are

99 significant fitness costs associated with expressing the effector, can it nonetheless propagate quickly
100 within the vector population? Questions also arise around the required efficiency of the driver gene and
101 gene drive system itself to achieve elimination. For example, the process of copying the effector gene
102 from one chromosome to another is not always successful. After cutting, DNA can sometimes undergo
103 alternative repair pathways that do not result in accurate copying of the drive or effector-containing
104 DNA on the sister chromosome. Non-homologous end-joining (NHEJ), microhomology mediated end-
105 joining, or incomplete HDR may occur instead with different probabilities, generating “resistant” alleles
106 that do not contain the desired drive or effector gene but are no longer recognized by the driver
107 endonuclease [24–26]. These resistant alleles may also be present in the wild mosquito population even
108 before introduction of new drive or effector genes [27]. The extent to which the generation and pre-
109 existing presence of these resistant alleles affects the ability of introduced gene drive mosquitoes to
110 eliminate malaria must be better quantified.

111 Because the potential harms and possible ecological risks associated with releasing gene drive
112 mosquitoes into the wild have not yet been well established, it is not currently feasible or ethical to test
113 such releases in the field. Community understanding, support, and buy-in are also needed before gene
114 drive mosquito releases can proceed [28–30]. Modeling is therefore a key step needed to quantify both
115 the potential benefits and harmful impacts of gene drive mosquito release. Modeling can also help
116 inform the minimum efficacy and genetic parameters required of engineered mosquitoes to achieve
117 substantial public health impacts, thus driving efficient and targeted development of genetically
118 engineered mosquitoes in the laboratory [31].

119 Here we examine the impacts of releasing malaria transmission-blocking, gene drive mosquitoes in a
120 rural Sahelian setting and quantify the gene drive system characteristics required to achieve elimination.
121 We use an individual-based model of malaria transmission that also resolves agent-based vector
122 genetics and allows for many-to-many mappings of vector genotypes to phenotypes [32]. We quantify
123 the difference in malaria outcomes across a range of transmission settings between releasing two
124 different population replacement gene drive mosquitoes (classic and integral), as well as with and
125 without other forms of vector control. Previous modeling work has focused on understanding changes in
126 vector populations with release of GM mosquitoes without considering other types of vector control
127 and without also examining the downstream effects on malaria transmission within corresponding
128 human populations [31,33–37]. An advantage of our model [32,38] is that it can simulate the effects of
129 gene drive-induced vector population changes on malaria transmission within a realistic human
130 population directly. Because of this added ability, we are able to quantify the gene drive system and
131 other logistical release parameters needed to reach full malaria elimination.

132 Methods

133 Model overview

134 Simulations were carried out using EMOD v2.20 [39], which is a mechanistic, agent-based model of
135 *Plasmodium falciparum* malaria transmission that can individually track each mosquito’s movement and
136 feeding pattern as well as each human’s movement, infection, and immune dynamics. Mosquitoes
137 within EMOD go through four life cycle stages: eggs, larvae, immature adults that do not seek hosts or
138 reproduce, and mature adults that do seek hosts and reproduce [40]. While adult female mosquitoes

139 can complete their feeding cycle and lay eggs, the number of eggs that progress to the larval stage is
140 determined by the amount of larval habitat available at a given time, which in turn governs the number
141 of adult vectors that eventually emerge.

142 Mosquitoes within EMOD contain simulated genomes represented by up to 10 different loci or genes,
143 with up to 8 different alleles per gene. Various phenotypic traits can be assigned to different genotypes,
144 including changes in fecundity, malaria transmissibility, mortality, and insecticide resistance.

145 When an adult male and female mosquito mate in the model, they each contribute half of the genes
146 belonging to their offspring. During gametogenesis before meiosis is complete, the germline cells within
147 each parent mosquito undergo all necessary gene drive-related changes to their genomes. After
148 completion of all drive-related changes, each parent's germline cells undergo meiosis and gametes are
149 distributed to offspring according to Mendelian inheritance. Further details regarding the
150 implementation of vector genetics within EMOD are explained in See Selvaraj et al. (2020) [32].

151 Human agents within EMOD each have their own microsolver to track within-host parasite dynamics and
152 the associated parasitological and clinical immunity that arise from innate and adaptive responses to
153 specific antigens. Parameters associated with this microsolver have been calibrated to reflect
154 transmission in a range of scenarios in Sub-Saharan Africa under different transmission intensities and
155 with or without interventions [41].

156 Modeled region

157 To capture conditions representative of the Sahel region of SSA, simulations were conducted over a 300
158 square kilometer region of rural Burkina Faso (Figure 1A). This 300 square kilometer region was divided
159 into 1 km-by-1 km grid cells, each with its own simulated human and vector population. Human
160 population data from the region was obtained from the High Resolution Settlement Layer generated by
161 the Facebook Connectivity Lab and Columbia University's Center for International Earth Science
162 Information Network [42]. Only grid cells with more than 5 people were included in the simulations,
163 resulting in ~3,700 individuals simulated across 150 populated grid cells.

164 Vector carrying capacity and initial populations were scaled to human population within each node to
165 ensure that humans have the same probability of being bitten across all grid cells. Only one vector
166 species, *Anopheles gambiae*, was assumed to be present and responsible for all malaria transmission.
167 Characteristic Sahelian seasonality in vector populations was captured by appropriately varying the
168 amount of available larval habitat space over the year (Figure 1B) [41,43–45]. The same seasonal profile
169 of larval habitat space was used in all grid cells and all scenarios. The amplitude of the larval habitat, and
170 in turn mosquito density and biting, was varied to simulate different transmission intensities with annual
171 entomological inoculation rates (EIR) varying between 10 infectious bites per person (reflecting a low
172 transmission setting) to 80 infectious bites per person (reflecting a high transmission setting).

173 Human migration is simulated by assigning each individual person a daily probability of taking overnight
174 trips to other grid cells. This probability is governed by a gravity model dependent on population in and
175 distance between nodes [46]. The gravity model is calibrated to movements observed in geotagged
176 campaign data [47] and results in an average of 5 overnight trips per person per year. Similar to human
177 migration, vector migration is simulated by assigning each individual mosquito a daily probability of
178 migrating to another grid cell. This probability is governed by a negative exponential distance decay

179 function [48] (Supp. Figure 1). Neither humans nor vectors migrate into or out of the simulated region.
180 There is therefore no importation of malaria from outside of the modelled area.

181 All scenarios were simulated for 8 years and 20 stochastic realizations were run for each scenario.

182 Modeled interventions

183 All simulations included treatment with artemether-lumefantrine (AL) for symptomatic cases. Those
184 with severe malaria cases sought treatment 80% of the time within 2 days of symptom onset. Those
185 with clinical, but not severe, cases sought treatment 50% of the time within 3 days of symptom onset.
186 Health-seeking rates are assumed to be the same for all ages.

187 Some simulations included ITN deployments. Per WHO guidelines [49], ITNs were distributed (Figure 1A)
188 every 3 years at the beginning of the peak season on July 1, covering a random 70% of the population
189 per distribution. To reflect the effects of insecticide resistance, each ITN is set to have a reduced initial
190 vector blocking efficacy of only 60% and killing rate of only 70%. Both blocking and killing rates decay
191 exponentially over time with a decay constant of 2 years and 4 years, respectively. Simulations with ITN
192 deployment alone (that is, those without a gene drive mosquito release) result in elimination
193 probabilities of zero for all transmission regimes (low, moderate, and high) tested here (results not
194 shown).

195 Gene drives

196 The basic setup of a gene drive system for population replacement involves coupling a driver gene with
197 an anti-malaria effector gene that prevents the mosquito from transmitting malaria. There are,
198 however, multiple ways in which this can be implemented. In what we term “classic” gene drive
199 systems, the driver and effector genes are propagated as a single complex construct and inserted at an
200 arbitrary target site within the genome (Figure 2A). In a more recently conceived “integral” gene drive
201 system, the driver and one or multiple effector genes are separated into distinct molecularly simpler
202 constructs and are then inserted into essential genes [37] (Figure 2B). In this case, the endonuclease
203 produced from the driver mediates homing both of its own gene and the effector gene. A previous
204 compartmental vector model suggests that an integral gene drive system of this type can provide
205 longer-lasting protection from malaria within a vector population than a classical system by both slowing
206 down the generation of resistance alleles and allowing for the Mendelian inheritance of the effector
207 gene even when the driver gene is lost [37]. Here we use EMOD to simulate the release and spread of
208 both classic and integral gene drive mosquitoes.

209 In all simulations discussed below, we released 100 male gene drive mosquitoes in each of the 6 most
210 populous nodes (1 km-by-1 km grid cells), for a total of 600 released mosquitoes, on July 1 of the first
211 simulated year. These 6 most populous nodes account for ~23% of the total human population in the
212 simulated region. In our classic gene drive release simulations, we allow for the possibility of 3 allele
213 types at the target site locus: wild type, complete construct (drive and effector), and resistant (Figure
214 2A). Only expression of the complete construct confers anti-pathogenicity and increases fitness cost via
215 enhanced mortality, while only the wild type allele can be recognized and cut by the driver. Resistant
216 alleles may occur naturally in the initial vector population and/or may arise during errors in the
217 homologous copying process; they do not carry any anti-pathogenicity or fitness cost and cannot be
218 recognized by the driver. In our integral gene drive release simulations, we allow for the possibility of 4

219 allele types at both the driver and effector target site loci: wild type, nuclease (in the case of the driver
220 target site) or effector (in the case of the effector target site), resistant, and loss-of-function (Figure 2B).
221 As with the classic gene drive system, only expression of the nuclease or effector affects vector fitness,
222 while only the wild type alleles can be recognized and cut by the driver. Only expression of the effector
223 confers anti-parasitic properties to the vector. Specific to integral gene drive systems, driver target sites
224 are located within essential, recessive lethal genes; loss-of-function alleles that lead to non-viability in
225 homozygosity can therefore crop up when mutations arise during HDR. Because of conferred non-
226 viability, loss-of-function alleles are disproportionately lost from the population, which consequently
227 increases the proportion of intact, successfully-copied nuclease or effector alleles relative to the classic
228 setup. In all simulations, we assume negligible rates of random mutations at all target sites. Tables 1 and
229 2 summarize other important classic and integral gene drive system parameters, respectively.

230 In simulations of both classic and integral gene drive releases, we examine the effects of the following
231 parameters on likelihood of local malaria elimination (defined as malaria prevalence reaching and
232 staying at zero by the end of simulation year 7 within all spatial nodes): the probability of copying over
233 the driver and/or effector genes in the presence of the driver gene (also known as the efficiency of the
234 drive, d); the ability of the effector gene to prevent onward malaria transmission in mosquitoes (also
235 known as the transmission-blocking effectiveness of the effector, which is equivalent in either
236 heterozygosity or homozygosity, rc); the pre-existing frequency of target site resistance alleles in the
237 population ($rr0$ in the classic case; $rr20$ at the effector target site in the integral case); and the fitness
238 cost associated with expressing the introduced driver and effector genes, represented by an increase in
239 vector mortality (sne in the classic case; $se2$ associated with the effector in the integral case).

240 Because of the high dimensionality of the results, we also created a website with interactive
241 visualizations of simulation output to accompany the figures in this text (Supp. Figure 2), located here:
242 <https://gene-drive.bmgf.io>. Website users can interactively visualize the effects of all tested parameters
243 on elimination probabilities along with elimination timing, prevalence, vector populations, and allele
244 frequencies over all simulated combinations of gene drive release types, ITN deployments, and
245 transmission regimes. Though we focus primarily on understanding the effects of tested parameters on
246 local elimination probabilities in this text, we highly encourage website users to explore the effects of
247 tested parameters on additional malaria-related variables plotted on the website as well, particularly
248 reductions in prevalence even if elimination is not achieved.

249 Results

250 Elimination probability decreases with pre-existing resistance, increases with
251 transmission-blocking and drive efficiency, and is context dependent with respect
252 to fitness costs

253 When conducting a single release of classic gene drive mosquitoes, elimination probabilities increase
254 when transmission-blocking effectiveness (rc) increases, drive efficiency (d) increases, and pre-existing
255 population target site resistance ($rr0$) decreases over all tested parameter values (Figure 3). Holding all
256 other parameters constant, as transmission-blocking effectiveness increases, each individual mosquito
257 carrying the complete construct in the population is less likely to become infected by the malaria
258 parasite and pass it on to their human hosts. Thus, the higher the transmission-blocking effectiveness,

259 the lower the frequency of vectors that are infectious among the total vector population (Figure 4) and
260 the greater the chance of eliminating malaria within the local population. With all other parameters held
261 constant, an increase in drive efficiency leads to both an earlier and higher peak effector frequency, as
262 the introduced complete construct spreads at super-Mendelian rates through the vector population
263 (Figure 5). Earlier and higher peak effector frequencies at higher drive efficiencies reduce the fraction of
264 the mosquito population that can be infected by malaria parasites, thus increasing local malaria
265 elimination probabilities. The opposite occurs as pre-existing population target site resistance increases
266 with all other parameters held constant. Because target site resistance prevents the spread of the
267 introduced construct, peak effector frequency is reduced and more mosquitoes are able to transmit
268 malaria parasites at higher pre-existing target site resistances (Figure 6). As a result, an increase in pre-
269 existing target site resistance within a vector population reduces the chances of locally eliminating
270 malaria with a single gene drive release. To visualize elimination probabilities with the tested
271 parameters on different axes than those displayed in Figure 3, see <https://gene-drive.bmgf.io>.

272 In comparison to the above three parameters (rc , d , and $rr0$), the effects of mortality-enhancing fitness
273 costs (sne) associated with expression of the introduced gene drive construct have a more complex
274 relationship with the likelihood of elimination. Depending on the context, elimination probabilities can
275 either increase or decrease with increases in fitness cost. In some scenarios, an increase in fitness cost
276 leads to a decrease in elimination probability (Figure 3, columns with upward triangles). Here an
277 increase in fitness cost associated with construct expression both delays and reduces peak effector
278 frequency (Supp. Figure 3A). As fitness costs of expressing the complete construct increase, wild type
279 mosquitoes can more readily outcompete mosquitoes bearing the complete construct, slowing the
280 initial spread of the effector through the population. Resistant mosquitoes can also more readily
281 outcompete mosquitoes with the complete construct at higher fitness costs, such that resistant alleles
282 can increase more rapidly and to a higher frequency in comparison to effector alleles in these situations.
283 These two effects work together to reduce effector frequency at all times in the population and
284 therefore lead to lower elimination probabilities with higher fitness costs. In other scenarios, however,
285 an increase in fitness cost leads to an increase in elimination probability (Figure 3, columns with
286 downward triangles). Here an increase in fitness cost still reduces effector frequency as before;
287 however, a transient reduction in total vector population due to higher mortality rates with higher
288 fitness costs has a larger effect on reducing malaria prevalence than a decrease in effector frequency
289 has on increasing malaria prevalence (Supp. Figure 3B). It is also possible for a combination of the above
290 two situations to occur, such that elimination probabilities can first increase and then decrease with
291 fitness cost (Supp. Figure 3C). In this case, an increase in fitness cost from low initial values substantially
292 reduces the total vector population without significantly affecting effector frequencies; then an increase
293 in fitness cost at moderate to high initial values substantially lowers effector frequencies while only
294 somewhat reducing the total vector population in comparison. Notably, the first scenario (a decrease in
295 elimination probabilities with increasing fitness cost) typically occurs at higher transmission-blocking
296 effectiveness values, while the latter scenario (an increase in elimination probabilities with increasing
297 fitness cost) typically occurs at lower transmission-blocking effectiveness values (Figure 3). This is likely
298 because adult vector numbers matter more and effector frequencies matter less at lower transmission-
299 blocking effectiveness values, since the effector is already relatively pervasive. On the other hand, at
300 higher values of transmission-blocking effectiveness, decreases in effector frequency are more
301 detrimental to malaria suppression, as elimination is more dependent on the effector working well.
302 Importantly, all simulations assume that the target species *Anopheles gambiae* is the sole malaria vector

303 and no expansion of other malaria-transmitting species to fill the ecological niche left by a transient
304 decrease in the original number of vectors.

305 For simulations that result in elimination, trends in elimination timing (defined as the number of
306 simulated years required to reach elimination starting from simulation day 0) follow those of elimination
307 probability (Figure 7). That is, higher elimination probabilities are associated with faster times to
308 elimination and are driven in the same ways by the four tested parameters. Increasing drive efficiency
309 and transmission-blocking effectiveness reduce time to elimination; increasing pre-existing resistance
310 increases time to elimination; and increasing fitness cost can increase or decrease time to elimination
311 depending on the same factors described for elimination probability above. This association between
312 elimination probability and timing occurs because gene drive parameter spaces leading to higher and
313 longer-lasting peak effector frequencies also tend to lead to earlier peak effector frequencies as well.

314 The above-described patterns of elimination probability driven by transmission-blocking effectiveness,
315 drive efficiency, pre-existing target site resistance, and fitness cost associated with a single classic gene
316 drive mosquito release also hold when integral rather than classic gene drive mosquitoes are released,
317 as well as when ITNs are deployed in addition to a single release of either classic or integral gene drive
318 mosquitoes (Figures 8-9). In the case of additional ITN deployment, however, increases in drive
319 efficiency do not always lead to increases in elimination probability because of mismatches in
320 seasonality and timing of maximum net and gene drive efficacy within the setups simulated here (gene
321 drive mosquitoes released on July 1 of year 1 and ITNs deployed by July 1 of year 1, 4 and 7). As was the
322 case for gene drive only scenarios, when drive efficiency increases, the peak in effector frequency shifts
323 earlier. In some situations with additional ITN deployment, however, this earlier peak in effector
324 frequency then subsides by the time ITNs are re-deployed a second time (on July 1 of year 4 - after
325 waning efficacy of ITNs from the initial deployment), such that the overlapping maximum effects of ITNs
326 and gene drive mosquitoes during low season (when chances of eliminating are highest) are actually
327 smaller than if drive efficiency were lower and peak effector frequency were more delayed (Supp. Figure
328 4). Timing the release of gene drive mosquitoes such that peak effector frequency coincides with
329 maximum ITN efficacy and the smallest vector population size may therefore be key to eliminating
330 malaria in certain situations.

331 **A single release of classic gene drive mosquitoes with high transmission-blocking**
332 **effectiveness is likely to locally eliminate malaria in low to moderate transmission**
333 **settings**

334 A single release of classic gene drive mosquitoes can virtually guarantee elimination in a moderate
335 transmission regime (annual EIR = 30) if the transmission-blocking effectiveness is greater than or equal
336 to 90%, pre-existing target site resistance is less than or equal to 1%, drive efficiency is greater than or
337 equal to 95%, and the fitness cost to mortality of expressing the effector is less than or equal to 40%
338 (Figure 3). If transmission-blocking effectiveness is 100% and drive efficiency is greater than or equal to
339 95%, pre-existing target site resistance can be as high as 10% if fitness costs are below ~20% (Figure 3).
340 When drive efficiency is less than or equal to 95%, a transmission-blocking effectiveness of ~80% or less
341 makes elimination highly unlikely or virtually impossible (Figure 3). In a low transmission regime (annual
342 EIR = 10), elimination probabilities are appreciably higher across all parameter values, such that a

343 transmission-blocking effectiveness of 80% (rather than 90%) still leads to high probabilities of
344 elimination even when drive efficiency is less than or equal to 95% (Supp. Figure 5).

345 Deployment of ITNs in conjunction with a single classic gene drive mosquito 346 release boosts elimination probability

347 Although a single release of classic gene drive mosquitoes with high transmission-blocking effectiveness
348 is likely to locally eliminate malaria by itself in low to moderate transmission regimes, additional
349 deployment of ITNs greatly enhances elimination probabilities at lower values of transmission-blocking
350 effectiveness (Figure 8). By deploying ITNs in a moderate transmission regime (annual EIR = 30),
351 elimination goes from virtually impossible to highly likely when transmission-blocking effectiveness
352 drops down to ~70% and drive efficiency is less than or equal to 95% (Figure 8). In the absence of ITNs,
353 elimination probabilities are negligible when transmission-blocking effectiveness is less than or equal to
354 80% and drive efficiency is less than or equal to 95% (Figure 3). In a low transmission regime (annual EIR
355 = 10), additional ITN deployment leads to high probabilities of elimination at values of transmission-
356 blocking effectiveness as low as 50% (Supp. Figure 6). In a high transmission regime (annual EIR = 80),
357 ITN deployment and classic gene drive mosquito release in combination lead to high probabilities of
358 elimination when transmission-blocking effectiveness is ~80% or higher (Supp. Figure 7).

359 Release of integral, rather than classic, gene drive mosquitoes further boosts 360 elimination probabilities

361 In comparison to a release of classic gene drive mosquitoes, a release of integral gene drive mosquitoes
362 expands the parameter space over which elimination is highly likely (Figure 9). That is, an integral gene
363 drive mosquito release can achieve the same or better elimination outcomes as a classic gene drive
364 mosquito release even with a less potent effector, a lower drive efficiency, and in the presence of a
365 higher target site resistance frequency (here at the effector site) within the population. In a moderate
366 transmission regime (annual EIR = 30), releasing integral gene drive mosquitoes in conjunction with ITN
367 deployment leads to high and near certain elimination probabilities at transmission-blocking
368 effectiveness values as low as 50% and drive efficiencies as low as 90% (Figure 9), compared to
369 transmission-blocking effectiveness values around 70% and similar drive efficiencies when releasing
370 classic gene drive mosquitoes (Figure 8). An integral gene drive mosquito release in low and high
371 transmission regimes yields similar increases in elimination probabilities compared to a classic gene
372 drive mosquito release (Supp. Figures 8-9). To visualize elimination probabilities (along with elimination
373 timing, prevalence, vector population, and allele frequencies) for all simulated combinations of gene
374 drive release types (classic and integral), ITN deployments (with and without), and transmission regimes
375 (annual EIR = 10, 30, and 80), see <https://gene-drive.bmgf.io>.

376 Discussion

377 Transmission-blocking effectiveness and fitness cost may be the most important 378 parameters for future improvements and characterizations

379 Transmission-blocking effectiveness, drive efficiency, pre-existing target site resistance, and fitness cost
380 of expressing an introduced anti-pathogenic effector were all important parameters affecting local

381 malaria elimination probabilities across all simulated transmission intensities and scenarios. In general,
382 elimination probabilities were highest when transmission-blocking effectiveness was highest, drive
383 efficiency was highest, and pre-existing target site resistance was lowest (Figures 3, 8-9; Supp. Figures 5-
384 9). When deploying ITNs together with a gene drive release, however, increased drive efficiencies did
385 not always increase elimination probabilities due to mismatches in timing between maximum ITN
386 efficacy and peak effector frequency. To increase the chances of elimination, it is therefore important to
387 accurately quantify the genetic parameters associated with a given gene drive mosquito strain of
388 interest and time its release such that peak effector frequency coincides with both low mosquito season
389 and maximum efficacy of other forms of traditional vector control. Extensive vector surveillance before
390 gene drive mosquito release would also be needed to accurately quantify vector population seasonality,
391 along with pre-existing target site resistance.

392 Fitness cost affected elimination probability differently depending on transmission-blocking
393 effectiveness. At very high values of transmission-blocking effectiveness, higher fitness costs reduced
394 elimination probabilities, while at lower values of transmission-blocking effectiveness, this effect was
395 reversed (Figures 3, 8-9; Supp. Figures 5-9). Because fitness cost effects on elimination probabilities are
396 not uniform or easily predicted, researchers and public health workers should develop a good
397 understanding of both transmission-blocking effectiveness and fitness costs associated with their gene
398 drive mosquito strains of interest before release. Semi-field experiments and non-driving effector
399 releases could be instrumental in achieving this goal, as the translation of experimentally established
400 fitness parameters into actual fitness burden incurred by transgenic mosquitoes in the environment is
401 notoriously difficult. Extensive vector surveillance should also be conducted after all gene drive
402 mosquito releases, but especially for pilot releases, to validate models and better understand
403 complicated effects of gene drive system parameters such as fitness cost. Sufficient surveillance after
404 release can also be used to track failure rates and inform necessary adjustments to future gene drive
405 strains or release logistics.

406 Though all four parameters tested here had some measurable effect on elimination probabilities,
407 transmission-blocking effectiveness and fitness cost may be most important to focus on for future
408 improvements to new strains of gene drive mosquitoes, due both to their outsized influence on
409 elimination probability as well as their potentially limiting existing values.

410 Pre-existing target site resistance

411 Existing pre-existing target site resistances in wild *An. gambiae* populations will likely not adversely
412 affect the ability of population-replacing classic or integral gene drive mosquito releases to eliminate
413 malaria. Using a sample of ~1,000 wild-caught *An. gambiae s.l.* mosquitoes from natural populations
414 throughout Africa, Schmidt et al. (2020) [50] found that the vast majority (~90%) of all protein-coding
415 genes in *An. gambiae s.l.* contain at least one Cas9 target sequence with genetic variability less than or
416 equal to 1%. This sample of ~1,000 mosquitoes included both *An. coluzzii* and *An. gambiae s.s.*
417 specimens from the UC Davis Vector Genetics Laboratory and The *Anopheles gambiae* 1000 Genomes
418 Consortium. Furthermore, though genetic variability may be present, even when target sequences differ
419 by multiple nucleotides, efficient cleavage by a Cas9 driver enzyme may remain largely unimpaired
420 [51,52]. Existing population target site resistances are therefore likely to be low (less than or equal to
421 1%), given the ability of researchers to choose a favorable site with little Cas9-impairing genetic
422 variability.

423 Drive efficiency

424 Given realistic values of ~90-100% for drive efficiencies in *Anopheles* mosquitoes [51–55], elimination
425 probabilities are not substantially reduced when drive efficiency decreases within this range and
426 transmission-blocking effectiveness is sufficiently high (e.g., greater than or equal to ~90% when
427 releasing classic gene drive mosquitoes without ITNs in a transmission regime where annual EIR = 30,
428 and above ~60-70% in the same situation with ITNs). This is true for releases of classic or integral gene
429 drive mosquitoes, with or without vector control, though the exact values of transmission-blocking
430 effectiveness required differ depending on the gene drive system, transmission regime, and absence or
431 presence of other forms of vector control. Thus, even at the lower end of realistic *Anopheles* drive
432 efficiency values (~90%), elimination probabilities are generally not limited by drive transmission rates.
433 Though increasing drive efficiencies from 95% to 100% can boost elimination probabilities at lower
434 transmission-blocking effectiveness values (along with high fitness costs and low but realistic pre-
435 existing target site resistances), drive efficiencies of ~95% may be extremely difficult to improve upon
436 with conventional mosquito engineering efforts. Thus, drive transmission rate is not a high priority for
437 further improvements due to its already high efficiency and promising ability to enable elimination,
438 along with the likely difficulty associated with bringing efficiencies even higher.

439 Fitness cost

440 Fitness costs associated with expressing an anti-parasite effector have been reported to vary widely
441 among engineered strains of *An. gambiae* under laboratory conditions. Some anti-parasite effector
442 expressing strains have been created with negligible associated fitness costs [56,57], while others exhibit
443 measurable potential decreases in fecundity or lifespan [57,58]. It would be theoretically favorable to
444 create strains with effector expression fitness costs as low as possible, since lower fitness costs allow
445 introduced anti-pathogenic GM strains to more readily spread and compete against wild type
446 mosquitoes. However, fitness cost ranges required to achieve elimination are highly dependent on other
447 parameters. Assuming the presence of one primary malaria vector species and limited niche expansion
448 by another, a single release of gene drive mosquitoes with lower transmission-blocking effectiveness is
449 more likely to eliminate malaria when associated fitness costs of effector expression are higher (up to a
450 certain point). On the other hand, a release of more effective transmission-blocking gene drive
451 mosquitoes may be increasingly likely to eliminate malaria at lower fitness costs. Thus, rather than
452 universally seeking to generate strains with reduced fitness costs, researchers may opt to generate
453 strains with optimal combinations of drive efficiency, fitness cost, and transmission-blocking
454 effectiveness to increase the chances of elimination in their particular setting of interest. Though
455 transmission-blocking effectiveness must always be above some minimum threshold for any population
456 replacement gene drive release to achieve elimination, there is no equivalent maximum threshold that
457 fitness cost must be below. Here we simulated fitness cost as a uniform increase in vector mortality
458 across all ages and sexes, but future work could examine the outcomes of age or sex-specific fitness
459 effects, such as a reduction in the lifespan of females only.

460 Transmission-blocking effectiveness

461 Transgenic strains of *An. stephensi* with anti-*falciparum* transmission-blocking effectivenesses of 100%
462 or nearly 100% have been created [20,22,59,60]. However, there does not yet exist a transgenic strain of
463 *An. gambiae* that is able to inhibit *P. falciparum* parasite transmission as completely. Indeed, most

464 transgenic *An. gambiae* effectors show modest reductions in parasite transmission ability [57,58,61,62].
465 The most effective transgenic *An. gambiae* strain we were able to find in the published literature was
466 able to reduce the number of sporozoites per salivary gland by ~50% [57]. In the case of a single integral
467 gene drive mosquito release with ITNs in a high transmission setting (annual EIR = 80), elimination
468 probabilities drop precipitously below transmission-blocking effectiveness values of 60-70%, assuming
469 realistic ranges of other parameters. Thus, existing transmission-blocking effectivenesses in *An. gambiae*
470 are generally unlikely to be high enough to achieve elimination in high transmission settings even when
471 gene drive mosquito release is deployed in combination with ITNs. Transmission-blocking effectiveness
472 should therefore be the most important primary focus for future improvements in new strains of gene
473 drive mosquitoes.

474 Existing population replacement gene drive mosquitoes, in combination with 475 traditional forms of vector control, can likely eliminate malaria in low to moderate 476 transmission settings

477 A single release of a few hundred highly effective anti-pathogen, population-replacing classic gene drive
478 mosquitoes can locally eliminate malaria in a highly seasonal Sahelian setting with moderate
479 transmission rates (annual EIR of 30) when transmission-blocking effectiveness is very high (~90% or
480 higher) and other parameters (drive efficiency, pre-existing target site resistance, and fitness cost of
481 effector expression) are within realistic ranges (Figure 3). When paired with ITN deployment, a single
482 release of a few hundred classic gene drive mosquitoes increases the probability of elimination at all
483 values of drive efficiency, pre-existing target site resistance, transmission-blocking effectiveness, and
484 fitness cost (Figure 9). With ITNs, elimination probabilities are substantial even at transmission-blocking
485 effectiveness values down to ~50%. Thus, pairing a population replacement gene drive release with
486 provision of ITNs enhances rather than reduces the effect of the gene drive release, as was also shown
487 by Selvaraj et al. (2020) [32]. Utilizing integral gene drive mosquitoes with separate driver and effector
488 genes inserted at different loci also increases elimination probabilities across the board (Figure 9). In a
489 low transmission regime (annual EIR of 10), a single release of a few hundred classic gene drive
490 mosquitoes can eliminate malaria when transmission-blocking effectiveness is again very high, although
491 less so (~80% or higher) (Supp. Figure 5). To ensure high probabilities of elimination at lower
492 transmission-blocking effectiveness values (down to ~50%), integral gene drive mosquitoes, along with
493 ITNs and/or other forms of vector control, should again be utilized (Supp. Figure 6, 8). Thus, ITNs and
494 other traditional, non-gene drive vector control strategies are essential tools in the path towards
495 elimination because a single release of mosquitoes with currently achievable gene drive characteristics
496 is not likely to achieve elimination on its own, even in a low transmission regime. In a high transmission
497 setting (annual EIR = 80), additional non-gene drive interventions become even more important. In this
498 regime, transmission-blocking effectiveness values of ~50% lead to high probabilities of elimination only
499 when fitness costs are within a narrow range. Therefore, elimination in a high transmission Sahelian
500 setting will likely require a vast improvement to transmission-blocking effectiveness in future integral
501 gene drive mosquito strains and/or additional layering of non-gene drive interventions beyond ITNs.
502 These other interventions could include short term strategies such as indoor residual spraying (IRS),
503 attractive targeted sugar baits (ATSBs), long-acting injectable anti-malarials, and larviciding, as well as
504 longer term approaches including housing improvement, environmental management, and health
505 systems strengthening. Regardless of which other interventions are utilized, releasing population-

506 replacing gene drive mosquitoes helps create a window of opportunity during which prevalence may be
507 greatly suppressed and other tools can be ramped up to achieve elimination, even when a single gene
508 drive mosquito release by itself cannot.

509 Model limitations and future work

510 We sought to present as comprehensive and accurate an overview as possible of the effects of a single
511 gene drive mosquito release on malaria elimination within a spatially resolved and realistically seasonal
512 Sahelian setting. However, we made many necessary simplifying assumptions and were not able to
513 address all possible potentially relevant factors in this initial study. First, though releasing a larger
514 number of mosquitoes within the same 6 most populous nodes did not significantly affect our results,
515 we did not test whether altering the spatial pattern of mosquito release would measurably alter
516 elimination probabilities. Future work can optimize gene drive mosquito release locations and examine
517 factors contributing to optimal spatial planning for releases. Second, evolution and development of
518 parasite resistance to the anti-parasite effector molecules within the mosquitoes is not captured in our
519 model [63]. Future work can seek to better understand how this type of evolution could affect both
520 timing and probability of elimination probabilities, though parasite resistance can also be mitigated by
521 releasing a second set of mosquitoes with a different type of effector that would be new to the parasite.
522 Third, we only accounted for one species and pool of mosquitoes (*Anopheles gambiae*) in our
523 simulations and assumed that other species were either not present or did not play an appreciable role
524 in malaria transmission. If another malaria-transmitting *Anopheles* species were present to fill the
525 ecological niche of the single simulated species, elimination probabilities likely would not increase as
526 substantially with higher fitness costs and reduced vector populations at low values of transmission-
527 blocking effectiveness. Future work can examine elimination probabilities in the presence of multiple
528 malaria-transmitting *Anopheles* species with releases of gene drive mosquitoes corresponding to each
529 different species. Future work can also explore the effects of multiple gene drive mosquito releases over
530 several years compared to a one-time release. Lastly and perhaps most importantly, though we
531 simulated human and vector migration between 1 km-by-1 km nodes within the region, we did not
532 include migration of humans or vectors into or out of the simulated region. While the inner nodes from
533 the simulations serve as a proxy to study the effects of migration from outside regions to the simulated
534 area, we realize that continued importation of malaria via humans or vectors from outside of the
535 simulated region could have made elimination more difficult to achieve within the region across all
536 scenarios. However, because the gene drive systems simulated here are self-propagating, genes
537 introduced via these systems would gradually become established in surrounding regions as well,
538 spreading into all vector populations of the same species until a barrier to vector migration and
539 therefore gene flow is reached. Most or all vector populations migrating back into the simulated region
540 would therefore eventually have experienced their own introduction of gene drive mosquitoes as well.
541 Thus, it is not inconceivable that importation of malaria via migrating vectors and/or human travelers
542 into this relatively small region would gradually decrease and potentially become negligible over time. In
543 addition to the greatly reduced importation of malaria via gene drive mosquitoes from outside of the
544 simulated region, human importation of malaria into the simulated region could be greatly reduced if,
545 for example, travelers are required to be tested before entering or returning home. Future larger spatial
546 scale (and therefore lower resolution) simulations, along with incorporation of as yet unavailable
547 additional data on both human and vector migration distances, timings, and frequencies would allow us
548 to better resolve these dynamics. This additional data on vector movements would be invaluable for

549 better understanding the potential spatiotemporal evolution of gene drive mosquito frequencies and
550 the resultant effects on malaria transmission in SSA. Immediate future research should therefore
551 prioritize entomological surveillance efforts.

552

553

554 Tables and figures

555

| Parameter | Description | Default value | Range |
|------------|---------------------------------------------------------------|---------------|---------|
| <i>d</i> | Drive efficiency and transmission rate | 1 | 0.9 - 1 |
| <i>u</i> | Probability of resistance arising if drive transmission fails | 0.5 | - |
| <i>lne</i> | Probability of complete construct loss during homing | 3E-4 | - |
| <i>sd</i> | Fitness cost of target site disruption | 0 | - |
| <i>sne</i> | Fitness cost of complete construct expression | 0 | 0 - 0.5 |
| <i>hd</i> | Dominance coefficient for target site disruption | 0.5 | - |
| <i>hne</i> | Dominance coefficient for complete construct expression | 0.5 | - |
| <i>hrc</i> | Dominance coefficient for parasite refractoriness | 1 | - |
| <i>rc</i> | Homozygous degree of parasite refractoriness | 1 | 0.5 - 1 |
| <i>rr0</i> | Initial population target site resistance frequency | 0 | 0 - 0.1 |

556

557 Table 1 Classic gene drive system parameters.

558 All genetic parameters used in classic gene drive mosquito release simulations. Default values are
559 representative of and consistent with other published works [31,37,54]. Default values for tested
560 parameters (*d*, *sne*, *rc*, *rr0*) are used on accompanying website visualizations.

561

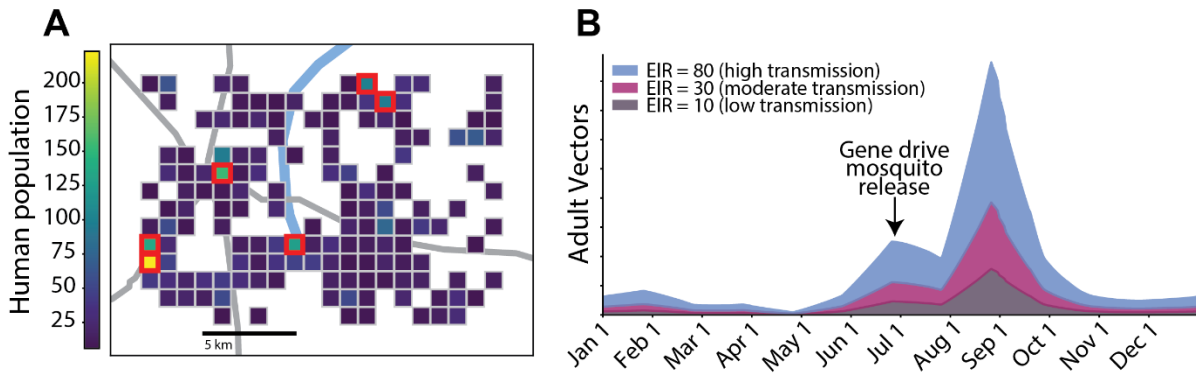
| Parameter | Description | Default value | Range |
|-----------------|------------------------------------------------------------------------------------------|---------------|---------|
| <i>d1</i> | Drive efficiency and transmission rate (applies to both driver and effector target loci) | 1 | 0.9 - 1 |
| <i>p_nhej</i> | Probability of NHEJ if drive transmission fails | 0.5 | - |
| <i>p_ihdr</i> | Probability of incomplete HDR during homing | 1E-4 | - |
| <i>p_r_nhej</i> | Probability of resistance arising from NHEJ | 1/3 | - |
| <i>p_r_ihdr</i> | Probability of resistance arising from incomplete HDR | 1/3 | - |
| $1 - p_r_nhej$ | Probability of loss of gene function from NHEJ | 2/3 | - |
| $1 - p_r_ihdr$ | Probability of loss gene function from incomplete HDR | 2/3 | - |
| <i>sd1</i> | Fitness cost of hijacking driver target locus | 0 | - |
| <i>sd2</i> | Fitness cost of hijacking effector target locus | 0 | - |
| <i>sn</i> | Fitness cost of driver expression | 0.05 | - |
| <i>se2</i> | Fitness cost of effector expression | 0 | 0 - 0.5 |
| <i>sm</i> | Fitness cost of loss of gene function | 1 | - |
| <i>hd1</i> | Dominance coefficient for hijacking at driver target locus | 0.5 | - |
| <i>hd2</i> | Dominance coefficient for hijacking at effector target locus | 0.5 | - |
| <i>hn</i> | Dominance coefficient for driver expression | 0.5 | - |
| <i>he2</i> | Dominance coefficient for effector expression | 0.5 | - |
| <i>hm</i> | Dominance coefficient for loss of gene function | 0.2 | - |
| <i>hrc1</i> | Dominance coefficient for parasite refractoriness | 1 | - |
| <i>rc</i> | Homozygous degree of parasite refractoriness | 1 | 0.5 - 1 |
| <i>rr10</i> | Initial population driver target site resistance frequency | 0 | - |
| <i>rr20</i> | Initial population effector target site resistance frequency | 0 | 0 - 0.1 |

562

563 Table 2 Integral gene drive system parameters.

564 All genetic parameters used in integral gene drive mosquito release simulations. Default values are
 565 representative of and consistent with other published works [31,37,54]. Default values for tested
 566 parameters (*d1*, *se2*, *rc*, *rr20*) are used on accompanying website visualizations.

567

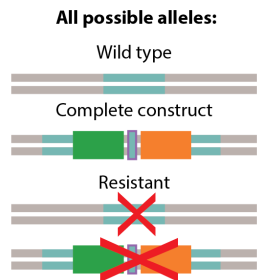
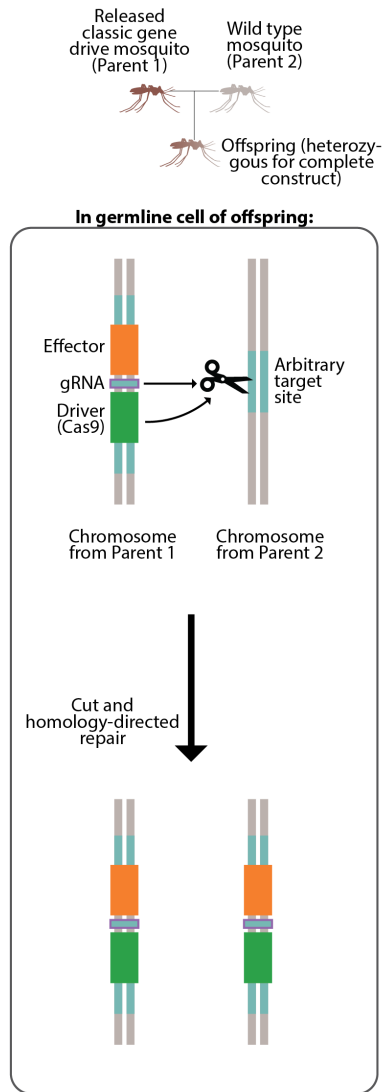


569 Figure 1 Simulated spatial region and seasonality.

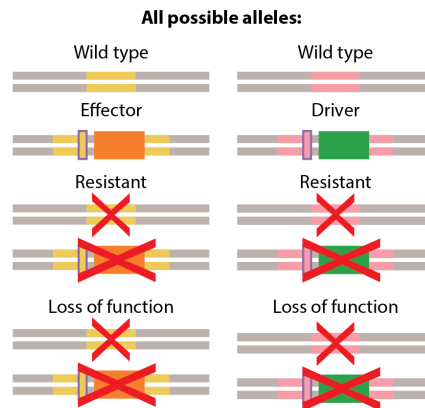
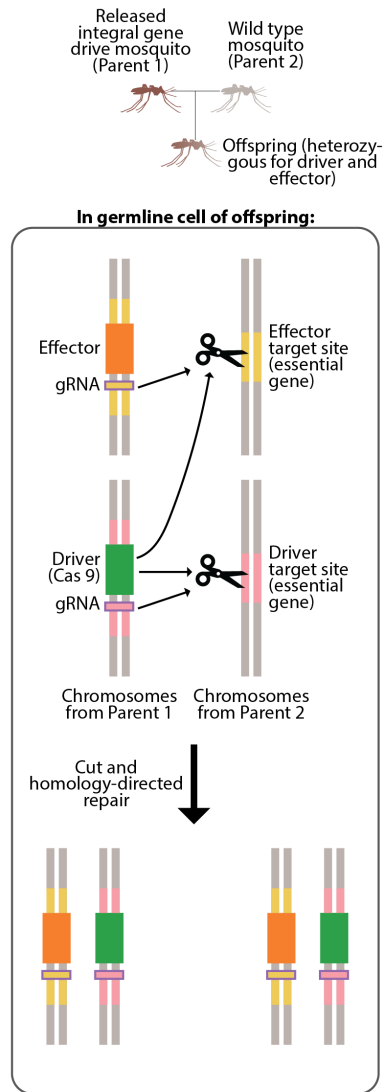
570 (A) Spatial region and grid composed of 150 1 km-by-1 km nodes used for all simulations. Colors denote
571 the human population within each node. In all simulation, 100 male gene drive mosquitoes were
572 released in each of the six most populous nodes (outlined in red), which account for ~23% of the human
573 population in the region. (B) Baseline seasonal cycle of adult vector populations within the simulated
574 area before gene drive releases in the three low (annual EIR = 10 infectious bites per person), moderate
575 (annual EIR = 30 infectious bites per person), and high (annual EIR = 80 infectious bites per person)
576 Sahelian transmission regimes simulated here. Gene drive mosquitoes were released on July 1 of the
577 first simulation year in all simulations. ITNS were also deployed on July 1 of the first, fourth, and seventh
578 simulation years in simulations with ITNs.

579

A Classic gene drive system

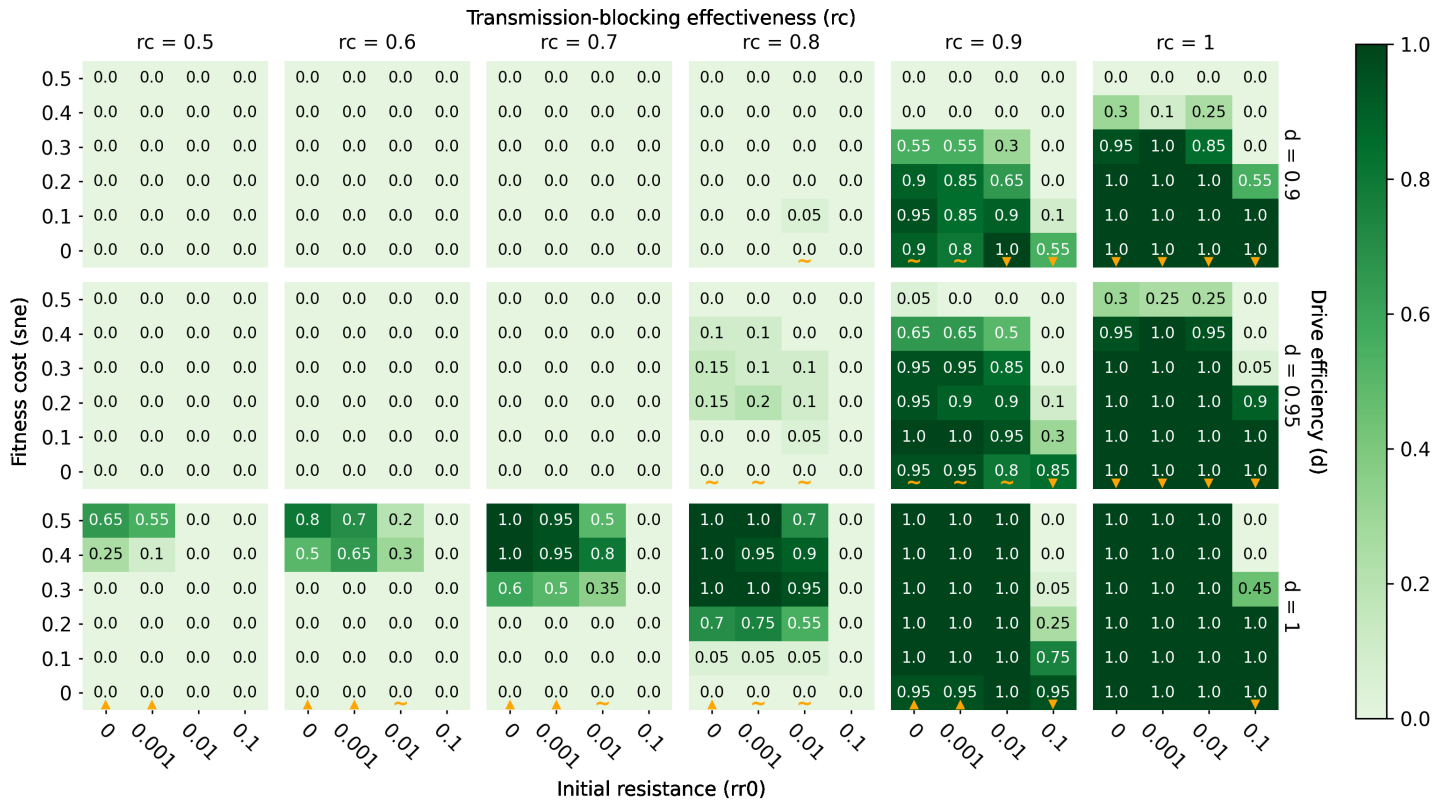


B Integral gene drive system



581 Figure 2 Classic and integral gene drive systems.

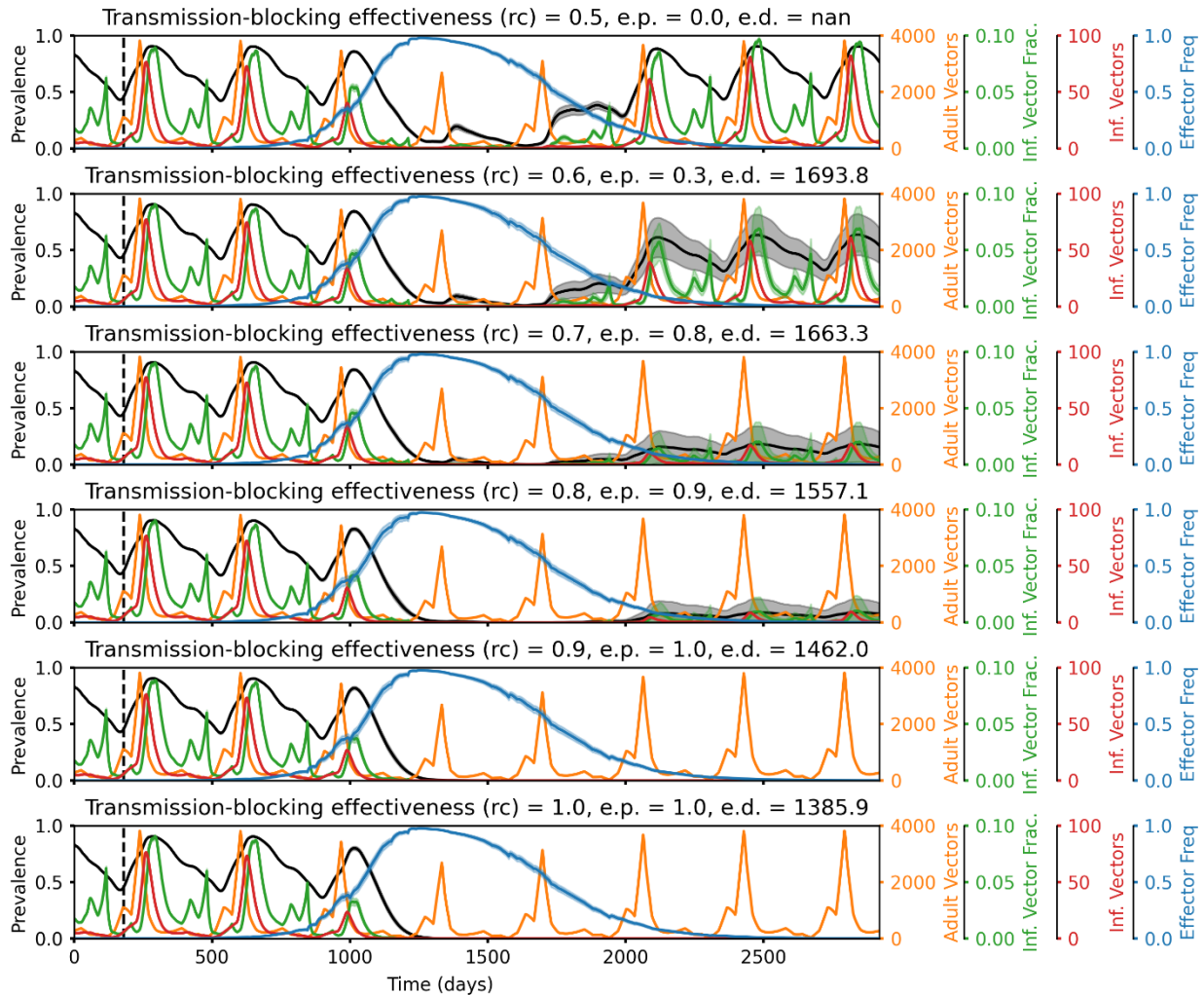
582 (A) Classic gene drive system and possible alleles. (B) Integral gene drive system and possible alleles.



584 Figure 3 Elimination probabilities after a single release of classic gene drive
 585 mosquitoes only in a moderate transmission (annual EIR = 30) regime.

586 Elimination probabilities (computed as the fraction of 20 model realizations in which malaria prevalence
 587 reaches and remains at zero by the end of simulation year 7) over a range of transmission-blocking
 588 effectiveness (rc), drive efficiency (d), pre-existing population target site resistance frequency ($rr0$), and
 589 mortality-enhancing effector expression fitness cost (sne) values. Orange upward and downward-
 590 pointing orange triangles denote columns along which elimination probabilities increase and decrease
 591 with increasing fitness cost, respectively. Orange tildes denote columns along which elimination
 592 probabilities first increase and then decrease with increasing fitness cost.

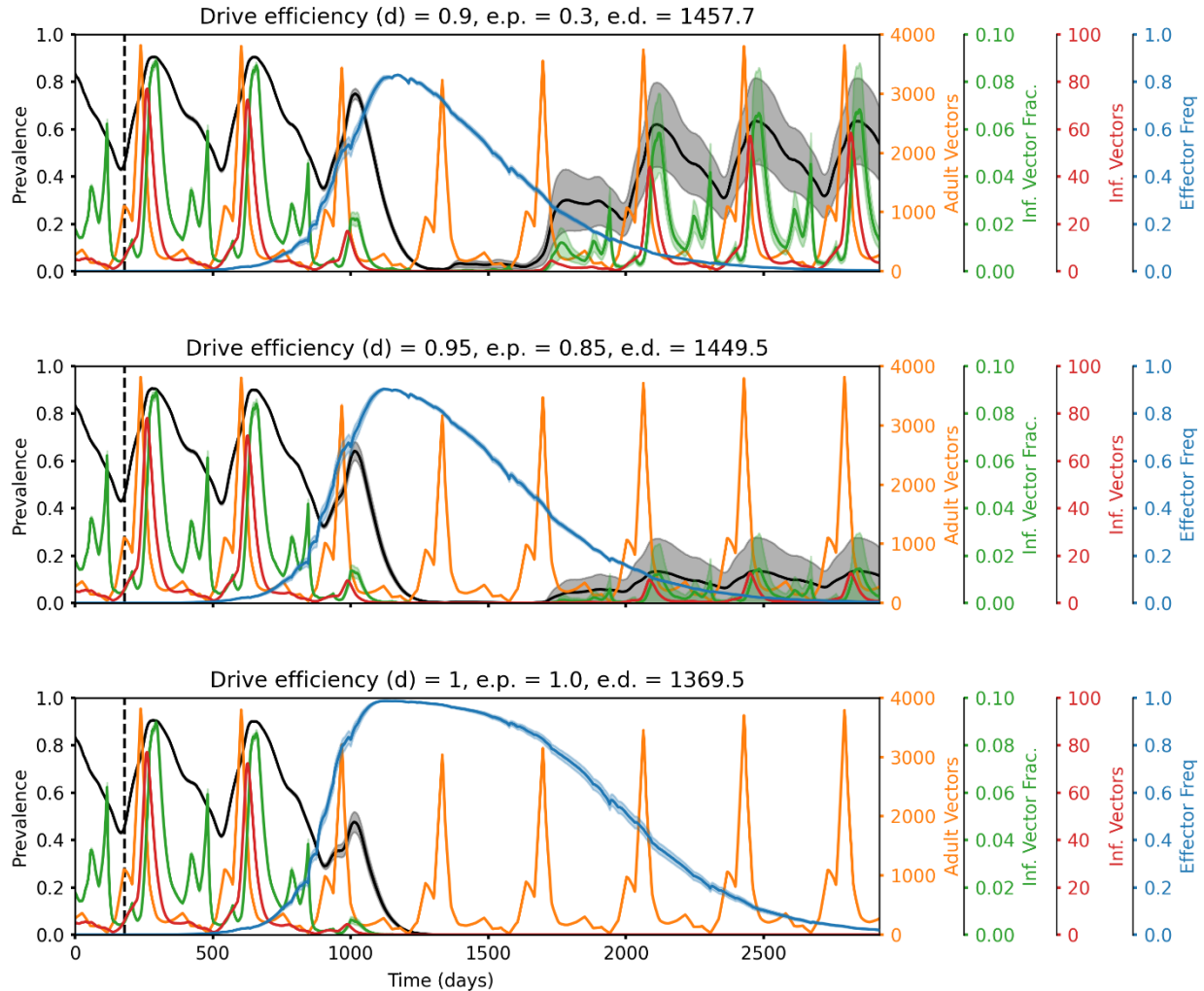
593



595 Figure 4 Representative time series illustrating how elimination probabilities
596 increase with increasing transmission-blocking effectiveness.

597 Time series of malaria prevalence, total adult vector population, infectious vector fraction, total
598 infectious adult vector population, and adult vector effector frequency over increasing values of
599 transmission-blocking effectiveness (rc). Elimination probabilities (e.p.) and number of days to
600 elimination (e.d.) are denoted in the subplot titles. In the simulations corresponding to these time
601 series, classic gene drive mosquitoes were released in a moderate transmission setting (annual EIR = 30)
602 with non- rc parameters set equal to the following values: drive efficiency (d) = 1, pre-existing resistance
603 ($rr0$) = 0.01, and fitness cost (sne) = 0.4. The higher the transmission-blocking effectiveness, the lower
604 the frequency of vectors that are infectious among the total vector population and the greater the
605 chance of locally eliminating malaria.

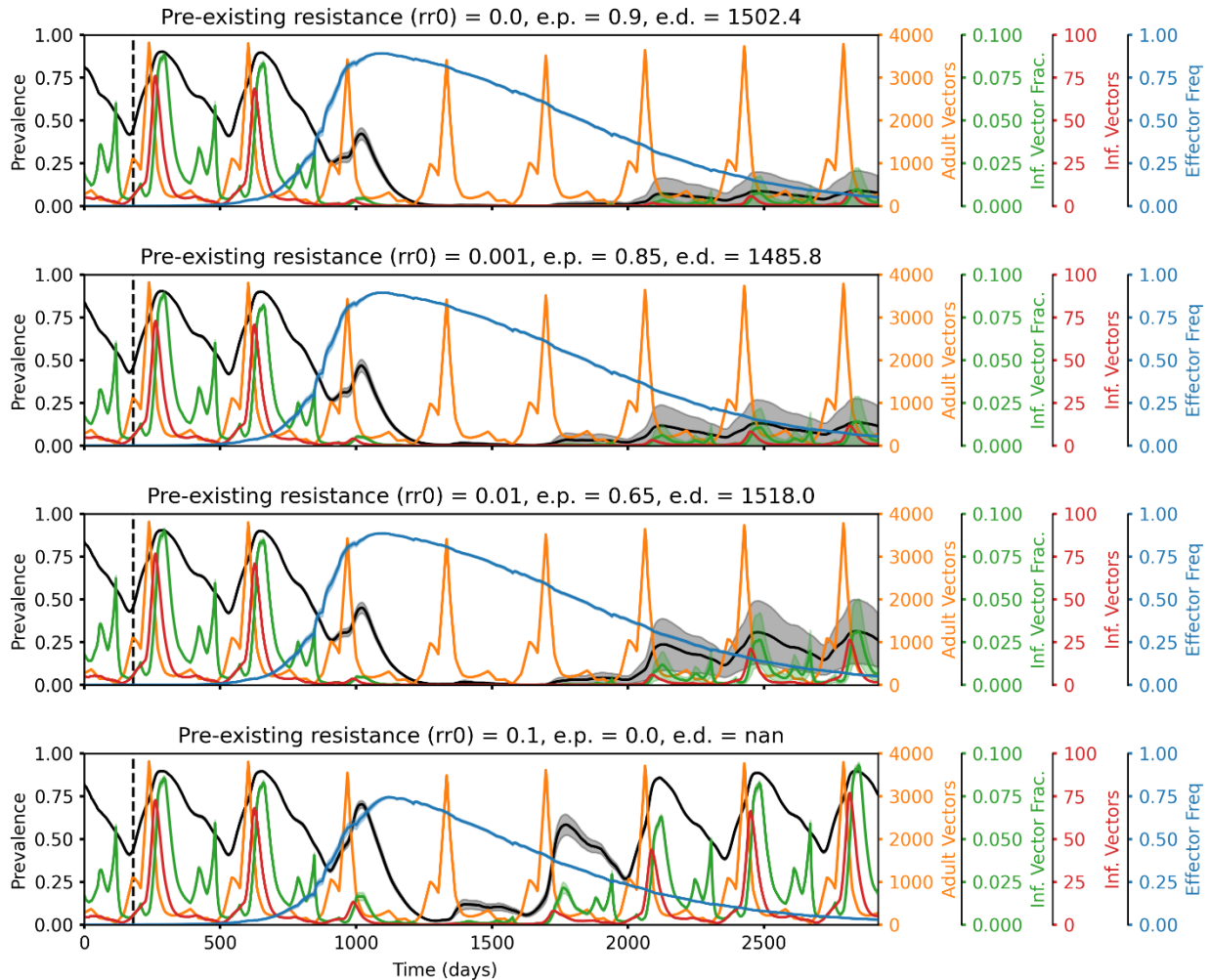
606



608 Figure 5 Representative time series illustrating how elimination probabilities
609 increase with increasing drive efficiency.

610 Time series of malaria prevalence, total adult vector population, infectious vector fraction, total
611 infectious adult vector population, and adult vector effector frequency over increasing values of drive
612 efficiency (d). Elimination probabilities (e.p.) and number of days to elimination (e.d.) are denoted in the
613 subplot titles. In the simulations corresponding to these time series, classic gene drive mosquitoes were
614 released in a moderate transmission setting (annual EIR = 30) with non- d parameters set equal to the
615 following values: transmission-blocking effectiveness (rc) = 0.9, pre-existing resistance ($rr0$) = 0.01, and
616 fitness cost (sne) = 0.3. The higher the drive efficiency, the greater the peak effector frequency, the
617 lower the infectious vector fraction, and the greater the chance of locally eliminating malaria.

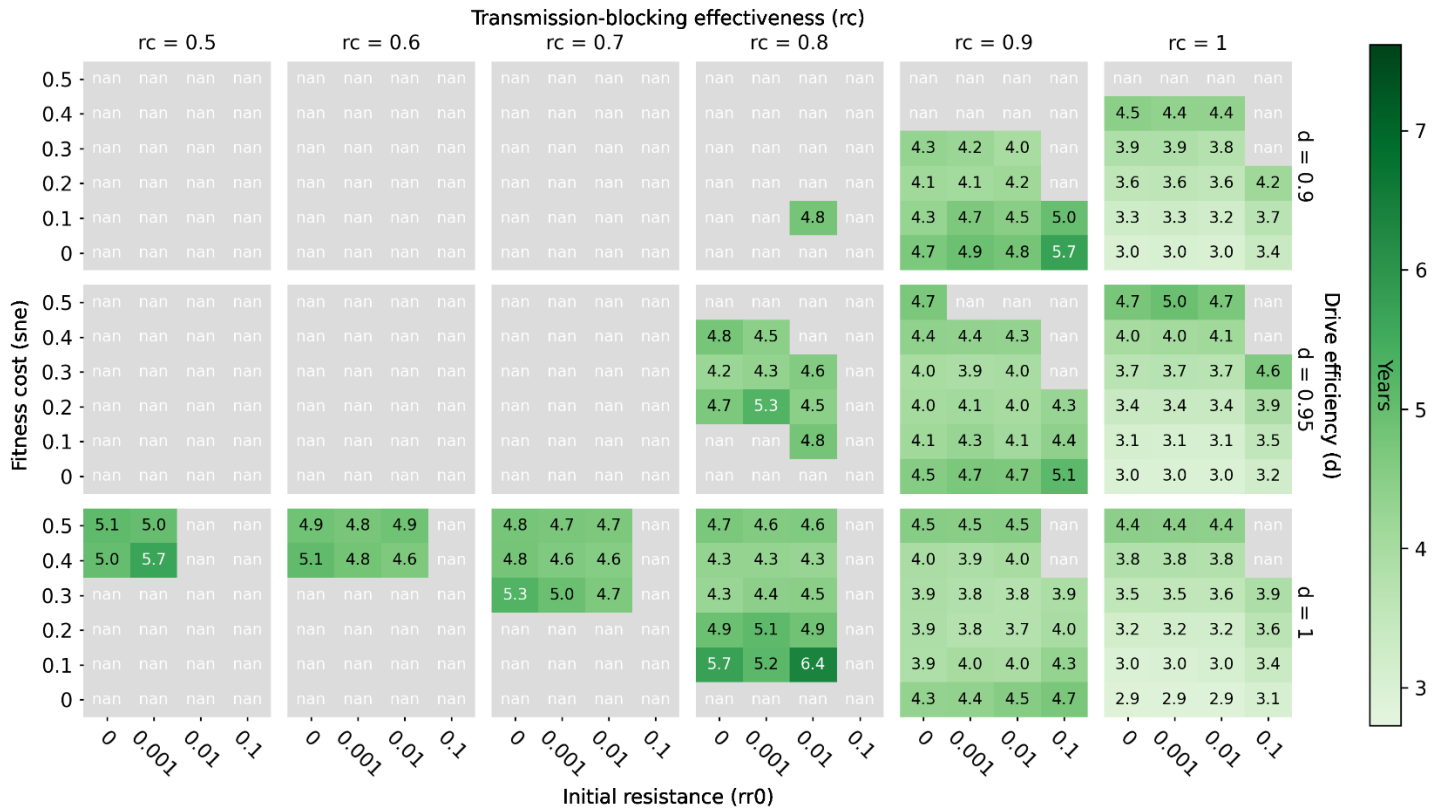
618



620 Figure 6 Representative time series illustrating how elimination probabilities
621 decrease with increasing pre-existing population target site resistance.

622 Time series of malaria prevalence, total adult vector population, infectious vector fraction, total
623 infectious adult vector population, and adult vector effector frequency over increasing values of pre-
624 existing population target site resistance frequency ($rr0$). Elimination probabilities (e.p.) and number of
625 days to elimination (e.d.) are denoted in the subplot titles. In the simulations corresponding to these
626 time series, classic gene drive mosquitoes were released in a moderate transmission setting (annual EIR
627 = 30) with non- $rr0$ parameters set equal to the following values: drive efficiency (d) = 0.9, transmission-
628 blocking effectiveness (rc) = 0.9, and fitness cost (sne) = 0.2. The higher the pre-existing resistance, the
629 lower the peak effector frequency, the higher the infectious vector fraction, and the lower the chance of
630 locally eliminating malaria.

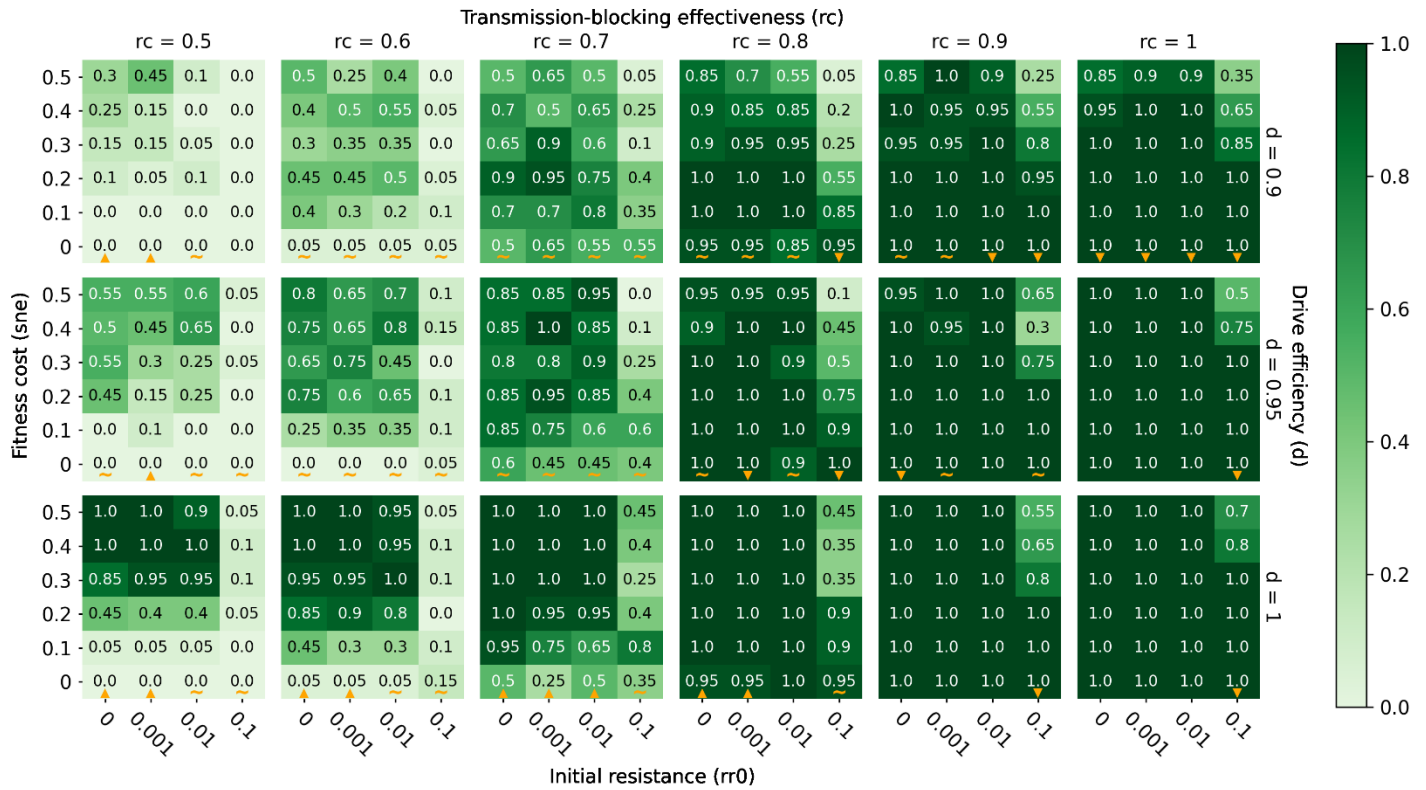
631



633 Figure 7 Elimination timing after a single release of classic gene drive mosquitoes
 634 only in a moderate transmission (annual EIR = 30) regime.

635 Elimination timing (computed as the number of years taken to reach elimination starting from
 636 simulation day 0, averaged over all realizations that eliminate) over a range of transmission-blocking
 637 effectiveness (rc), drive efficiency (d), pre-existing population target site resistance frequency ($rr0$), and
 638 mortality-enhancing effector expression fitness cost (sne) values.

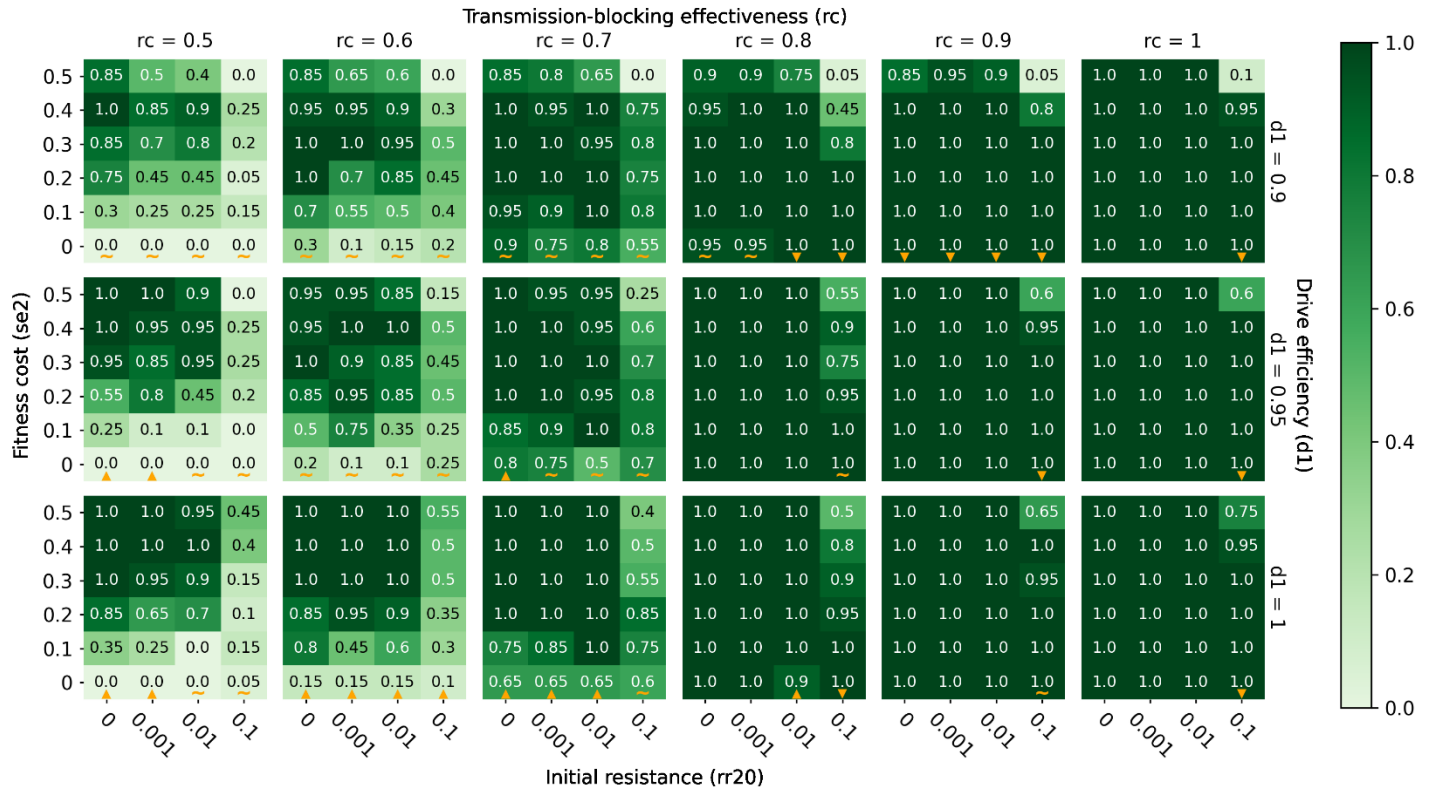
639



641 Figure 8 Elimination probabilities after a single release of classic gene drive
 642 mosquitoes and ITN deployment in a moderate transmission (annual EIR = 30)
 643 regime.

644 Same as Figure 3.

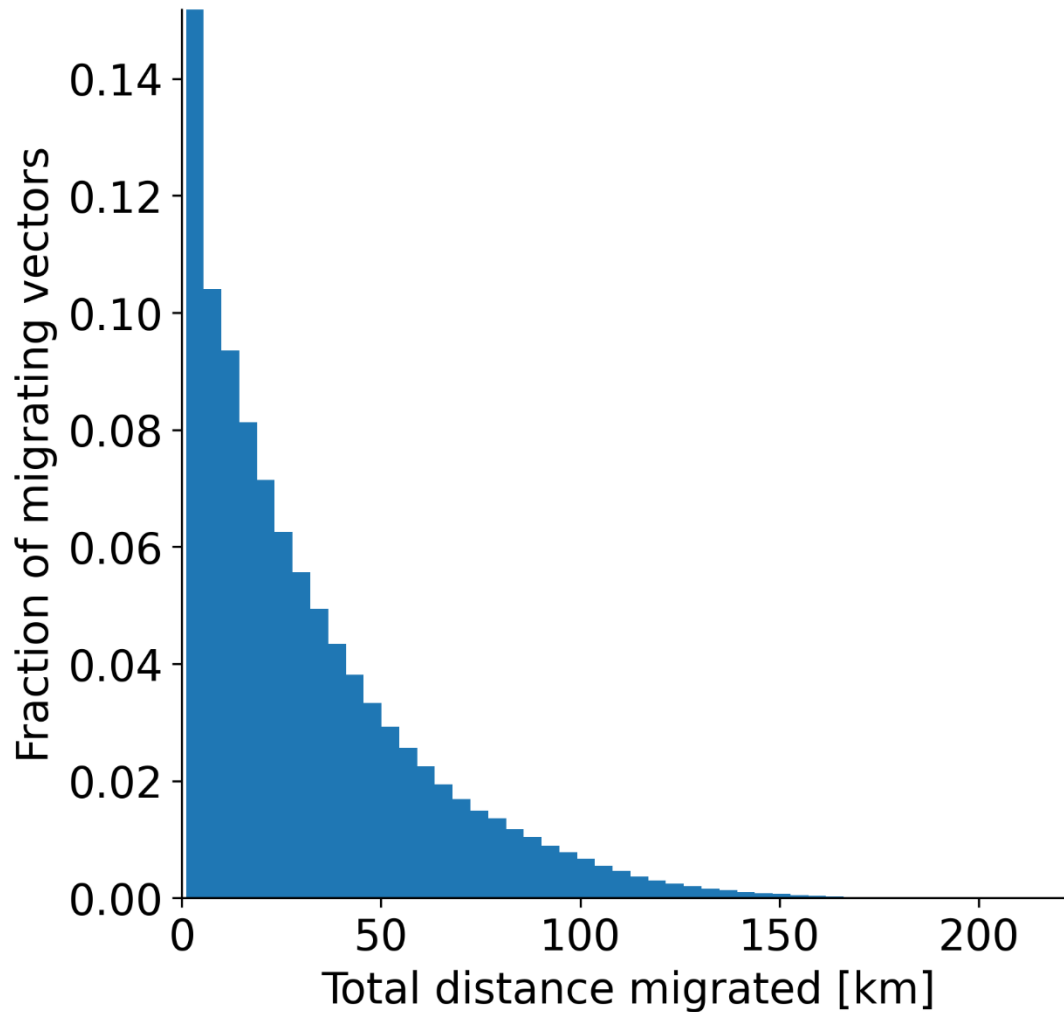
645



647 Figure 9 Elimination probabilities after a single release of integral gene drive
 648 mosquitoes and ITN deployment in a moderate transmission (annual EIR = 30)
 649 regime.

650 Same as Figure 3.

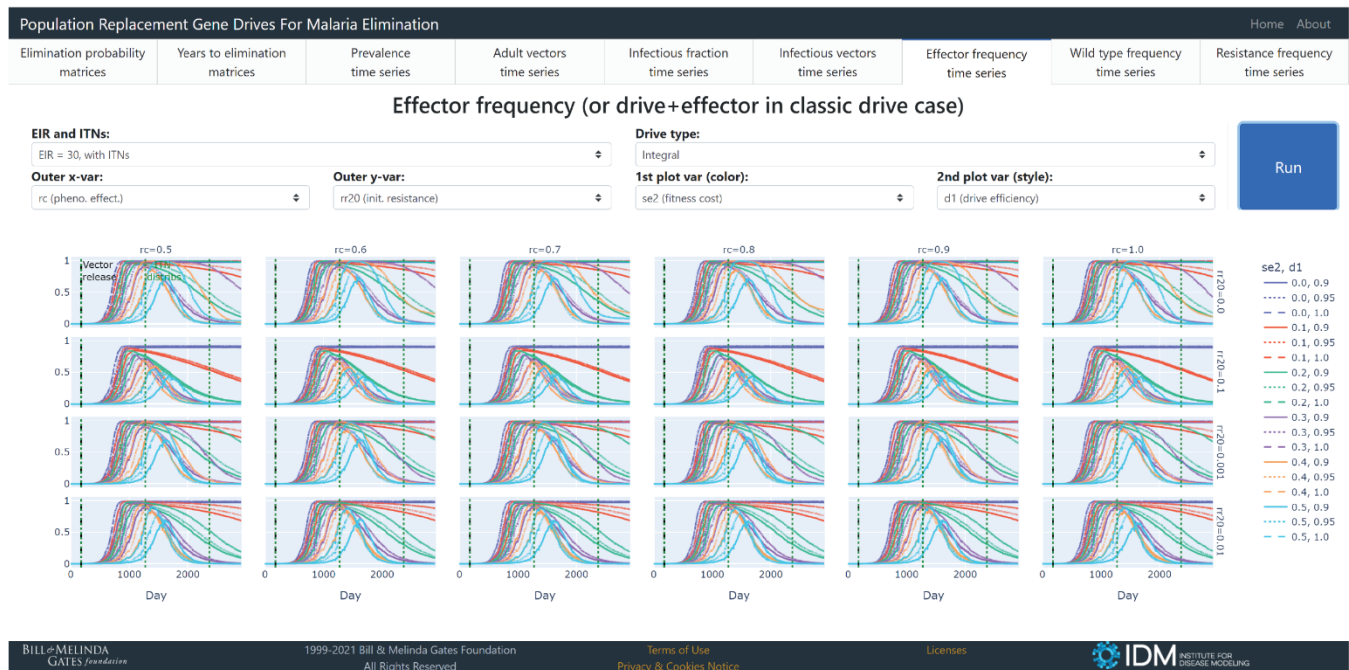
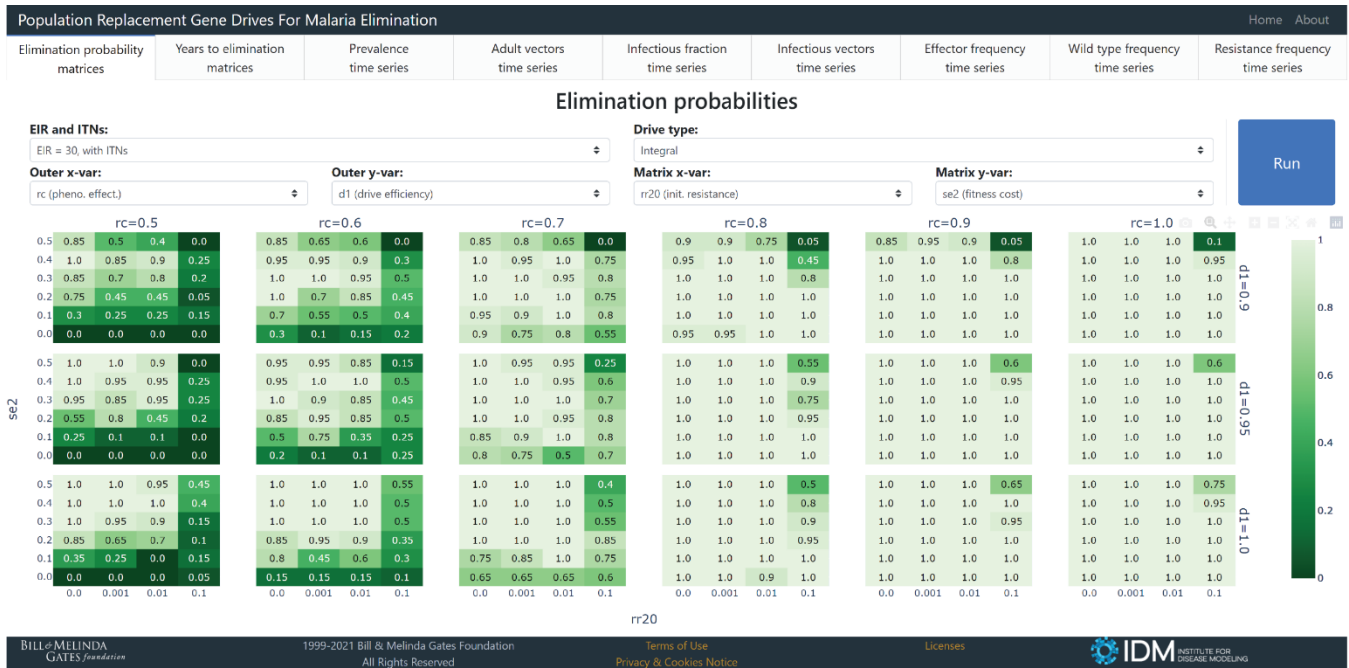
651



653 Supp. Figure 1 Distribution of vector migration distances within the simulated
654 area.

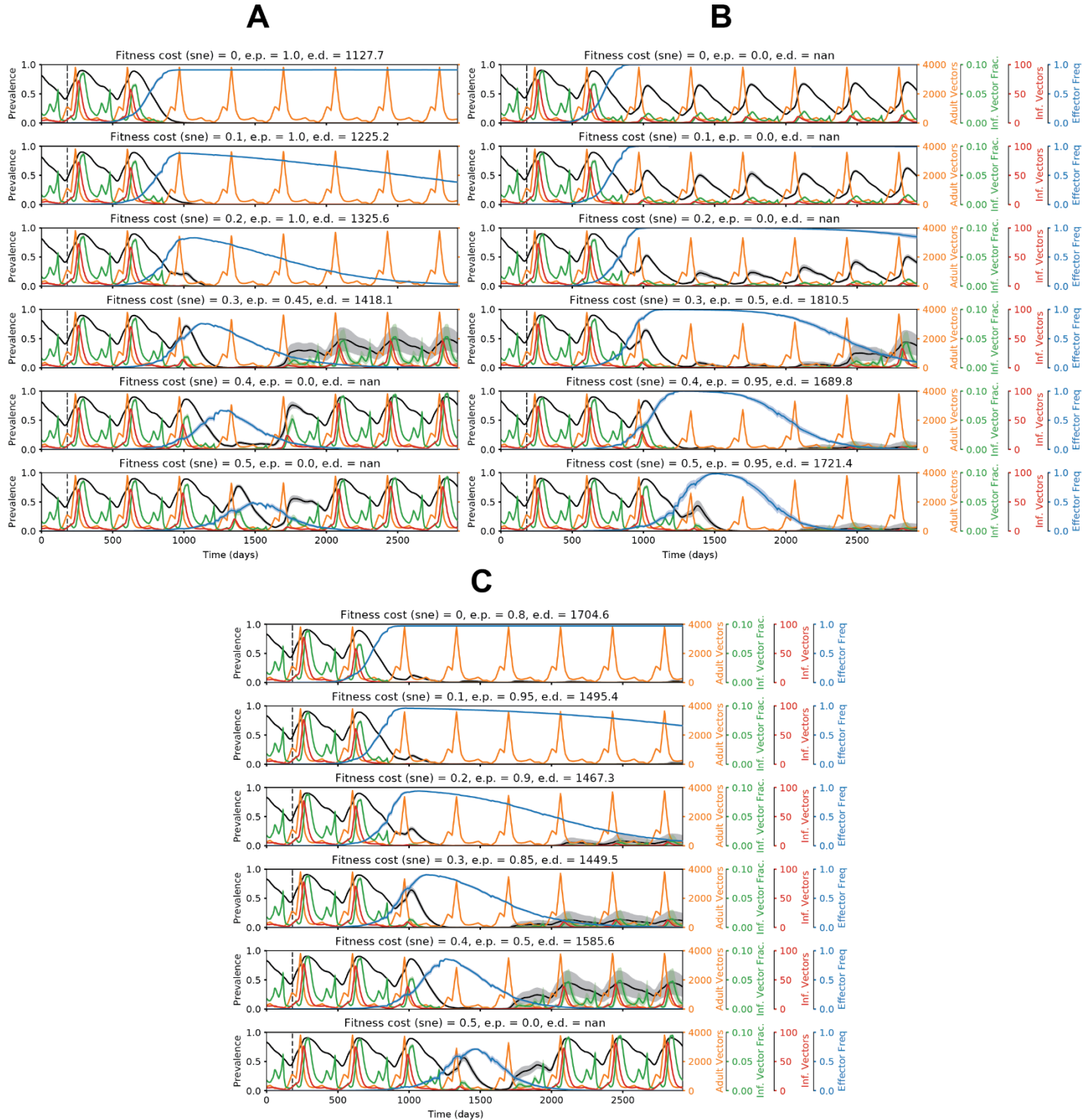
655 Fraction of vector migrations versus distance, computed by summing total migration distance over each
656 migrating vector's existence within a 2-month period (August 1 to October 1 in the first simulation year
657 with annual EIR = 30 and no ITNs or gene drive release), counting the number of total migration
658 distances within each histogram distance bin, and then dividing by the total number of migrating vectors
659 in the 2-month period. Total migration distance as plotted here does not necessarily represent the
660 distance between a vector's starting and ending point (i.e, its displacement), but instead represents the
661 total distance traveled. Migration probabilities are governed by an empirical negative exponential
662 distance decay function [48].

663



665 Supp. Figure 2 Screenshots of accompanying website for interactive visualization
666 of simulation output.

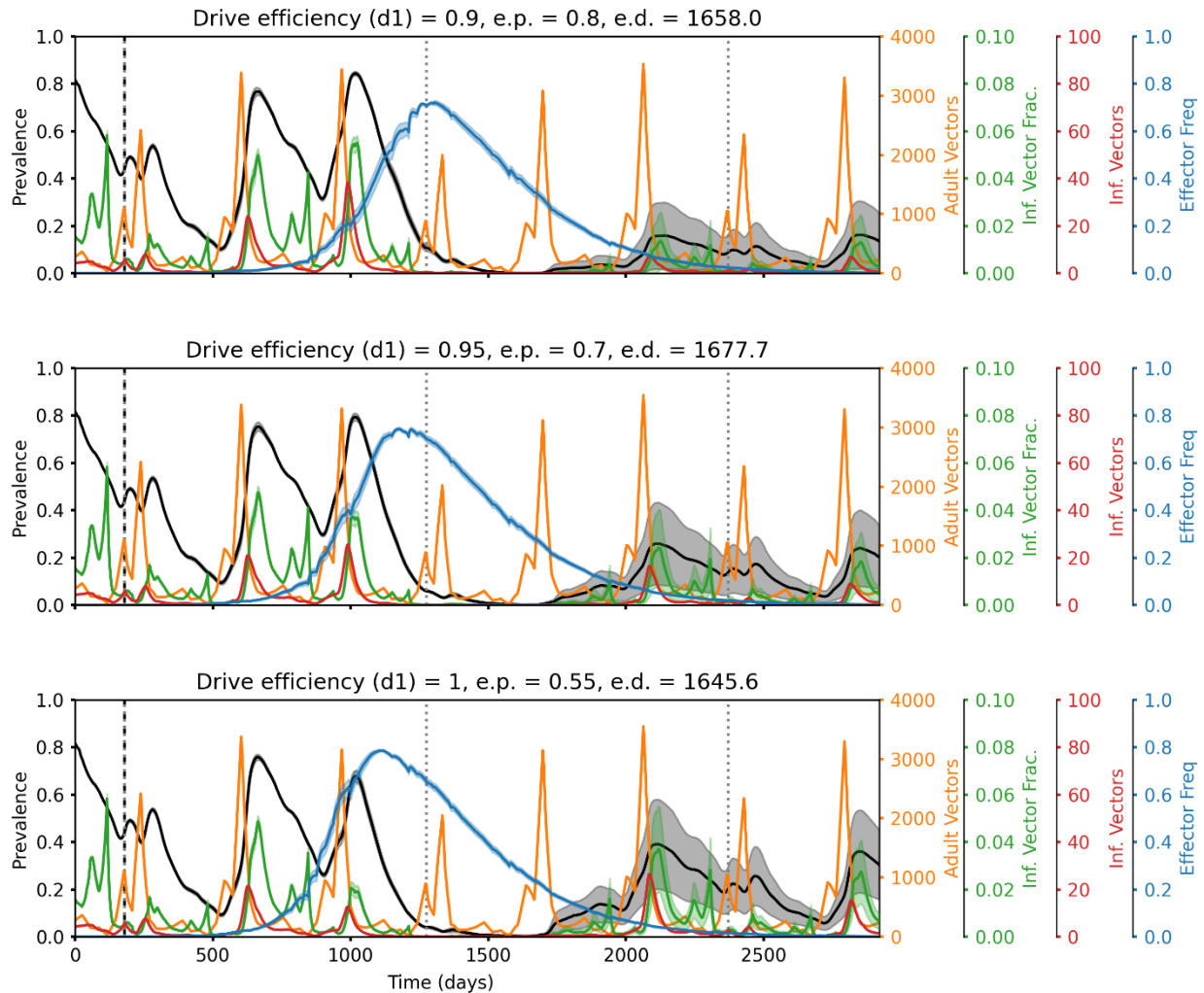
667 Screenshots of two different tabs on the website located here: <https://gene-drive.bmgf.io>. Website
668 users can interactively visualize the effects of tested gene drive parameters on elimination probabilities,
669 elimination timing, prevalence, vector populations, and allele frequencies over all simulated
670 combinations of gene drive release types, ITN deployments, and transmission regimes.



672 Supp. Figure 3 Representative time series illustrating how elimination
673 probabilities can either increase or decrease with increasing fitness costs of
674 complete construct expression.

675 Time series of malaria prevalence, total adult vector population, infectious vector fraction, total
676 infectious adult vector population, and adult vector effector frequency over increasing values of fitness
677 costs associated with complete construct expression (*sne*). Elimination probabilities (e.p.) and number of
678 days to elimination (e.d.) are denoted in the subplot titles. In the simulations corresponding to these
679 time series, classic gene drive mosquitoes were released in a moderate transmission setting (annual EIR
680 = 30). In column A, representing the case in which increasing fitness costs increase elimination
681 probabilities, non-*sne* parameters were set equal to the following values: drive efficiency (d) = 1, pre-
682 existing resistance ($rr0$) = 0.001, and transmission-blocking effectiveness (rc) = 0.7. In column B,
683 representing the case in which increasing fitness costs decrease elimination probabilities, non-*sne*
684 parameters were set equal to the following values: $d = 1$, $rr0 = 0.1$, and $rc = 1$. In column C, representing
685 the case in which increasing fitness costs increase and then decrease elimination probabilities, non-*sne*
686 parameters were set equal to the following values: $d = 0.95$, $rr0 = 0.01$, and $rc = 0.9$. In column A, the
687 higher the fitness costs, the lower the total vector population, and the greater the chance of locally
688 eliminating malaria. In column B, the higher the fitness costs, the lower the peak effector frequency, and
689 the lower the chance of elimination. In column C, the effects described for columns A and B are both at
690 play.

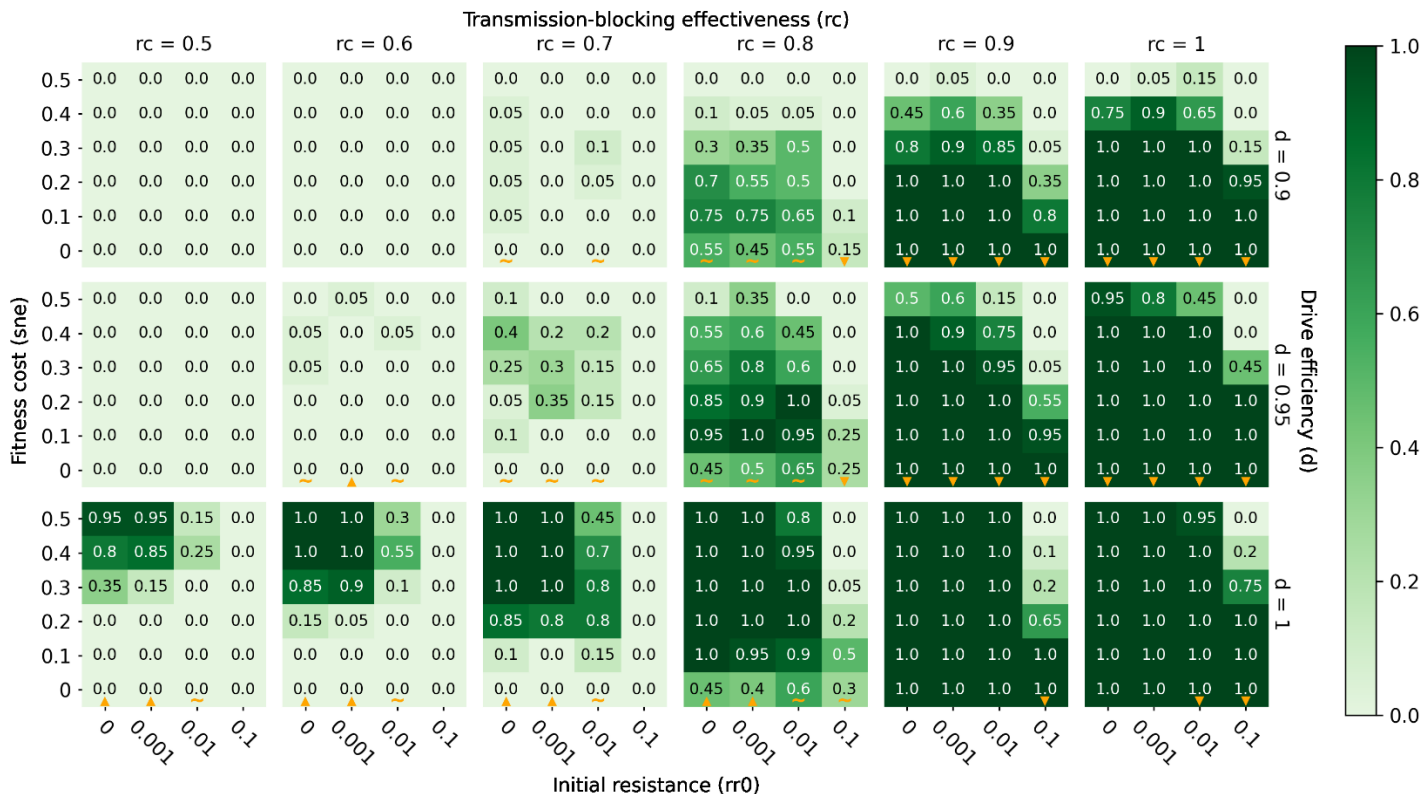
691



693 Supp. Figure 4 Representative time series illustrating how elimination
694 probabilities can sometimes decrease with increasing drive efficiency.

695 Time series of malaria prevalence, total adult vector population, infectious vector fraction, total
696 infectious adult vector population, and adult vector effector frequency over increasing values of drive
697 efficiency ($d1$). Elimination probabilities (e.p.) and number of days to elimination (e.d.) are denoted in
698 the subplot titles. In the simulations corresponding to these time series, integral gene drive mosquitoes
699 were released and ITNs were deployed in a moderate transmission setting (annual EIR = 30) with non- $d1$
700 parameters set equal to the following values: transmission-blocking effectiveness (rc) = 0.7, pre-existing
701 resistance at the effector target site ($rr20$) = 0.1, and fitness cost of expressing the effector ($se2$) = 0.3.
702 The higher the drive efficiency, the earlier the peak in effector frequency, and the lower the chance of
703 locally eliminating malaria when this earlier peak does not match up with maximum ITN efficacy.

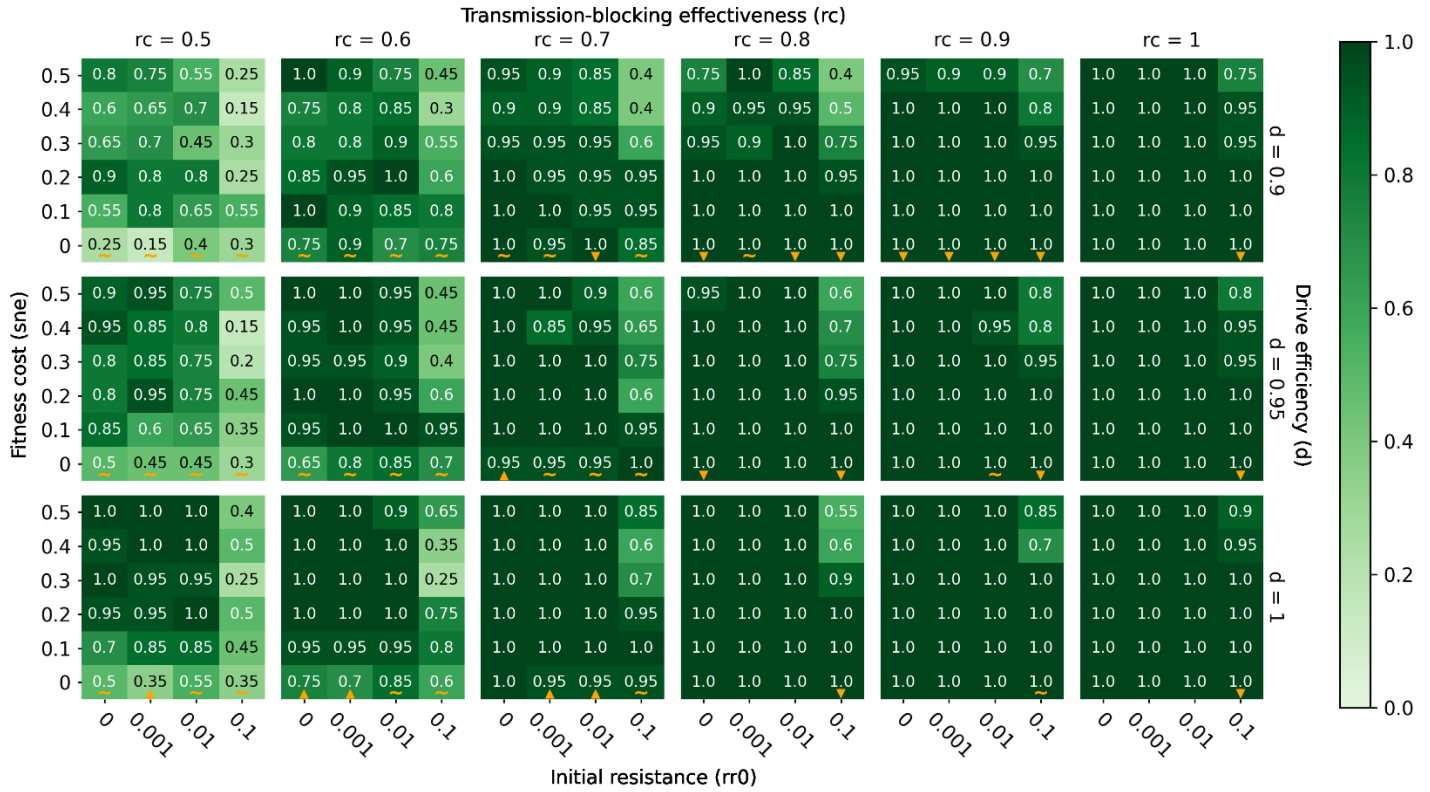
704



706 Supp. Figure 5 Elimination probabilities after a single release of classic gene drive
707 mosquitoes only in a low transmission (annual EIR = 10) regime.

708 Same as Figure 3.

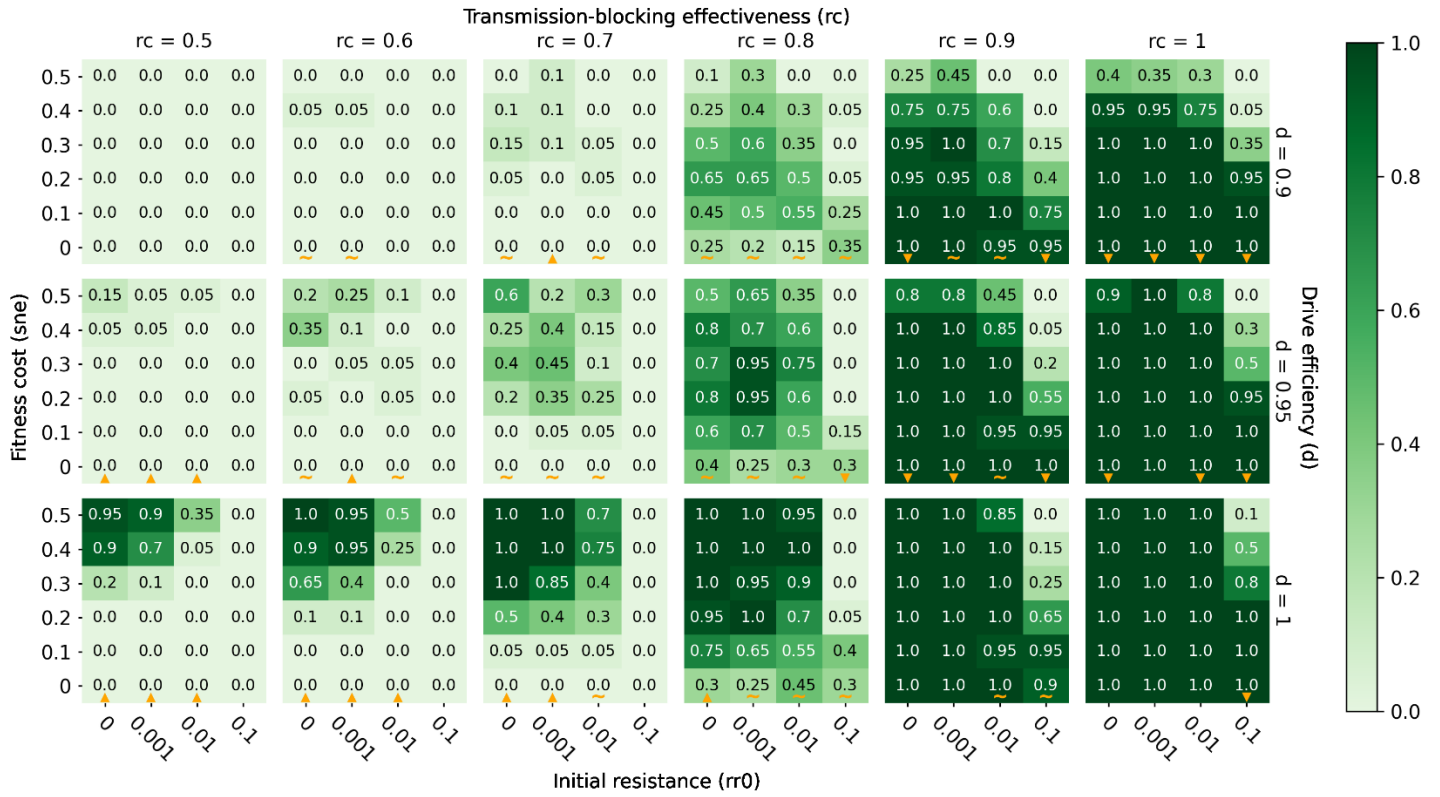
709



711 Supp. Figure 6 Elimination probabilities after a single release of classic gene drive
 712 mosquitoes and ITN deployment in a low transmission (annual EIR = 10) regime.

713 Same as Figure 3.

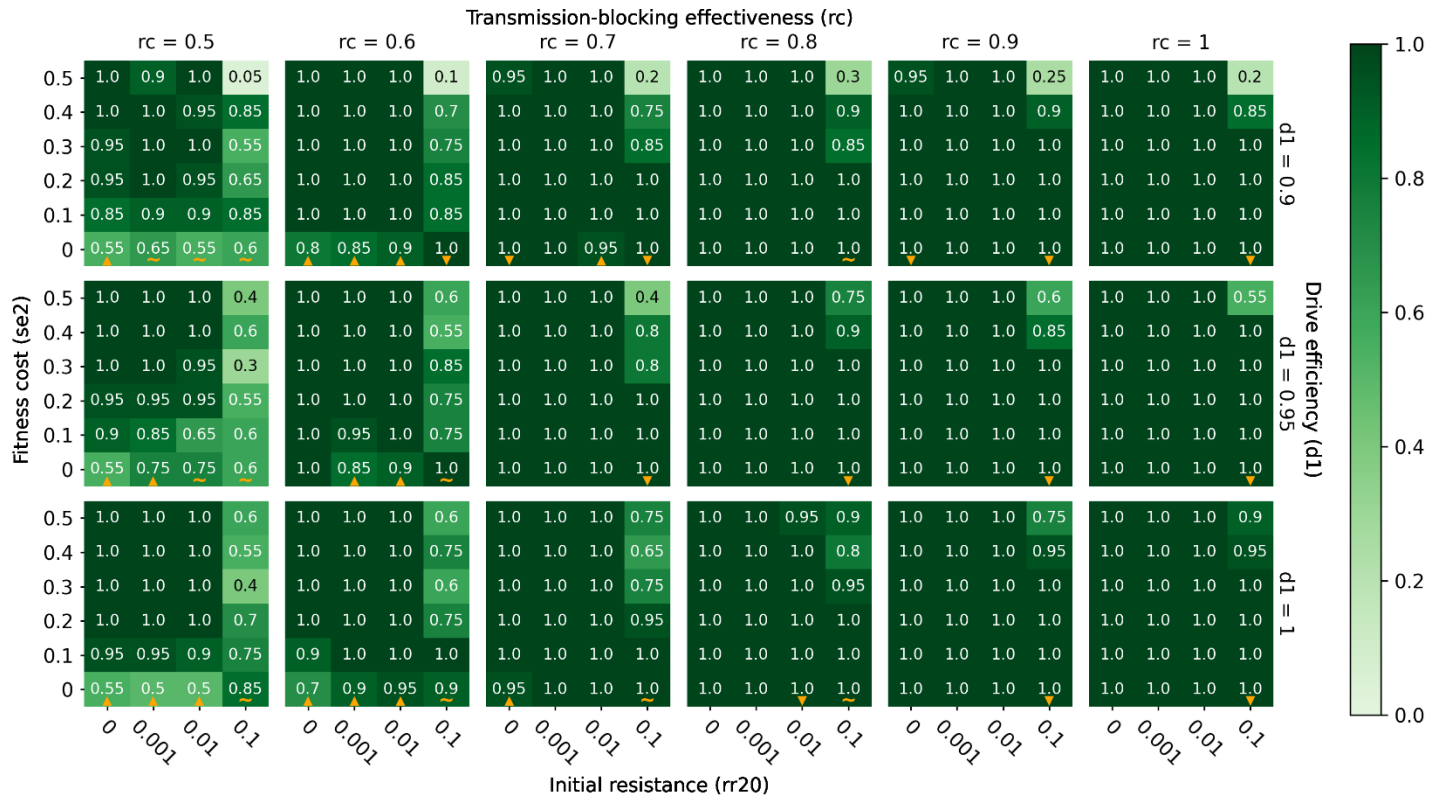
714



716 Supp. Figure 7 Elimination probabilities after a single release of classic gene drive
 717 mosquitoes and ITN deployment in a high transmission (annual EIR = 80) regime.

718 Same as Figure 3.

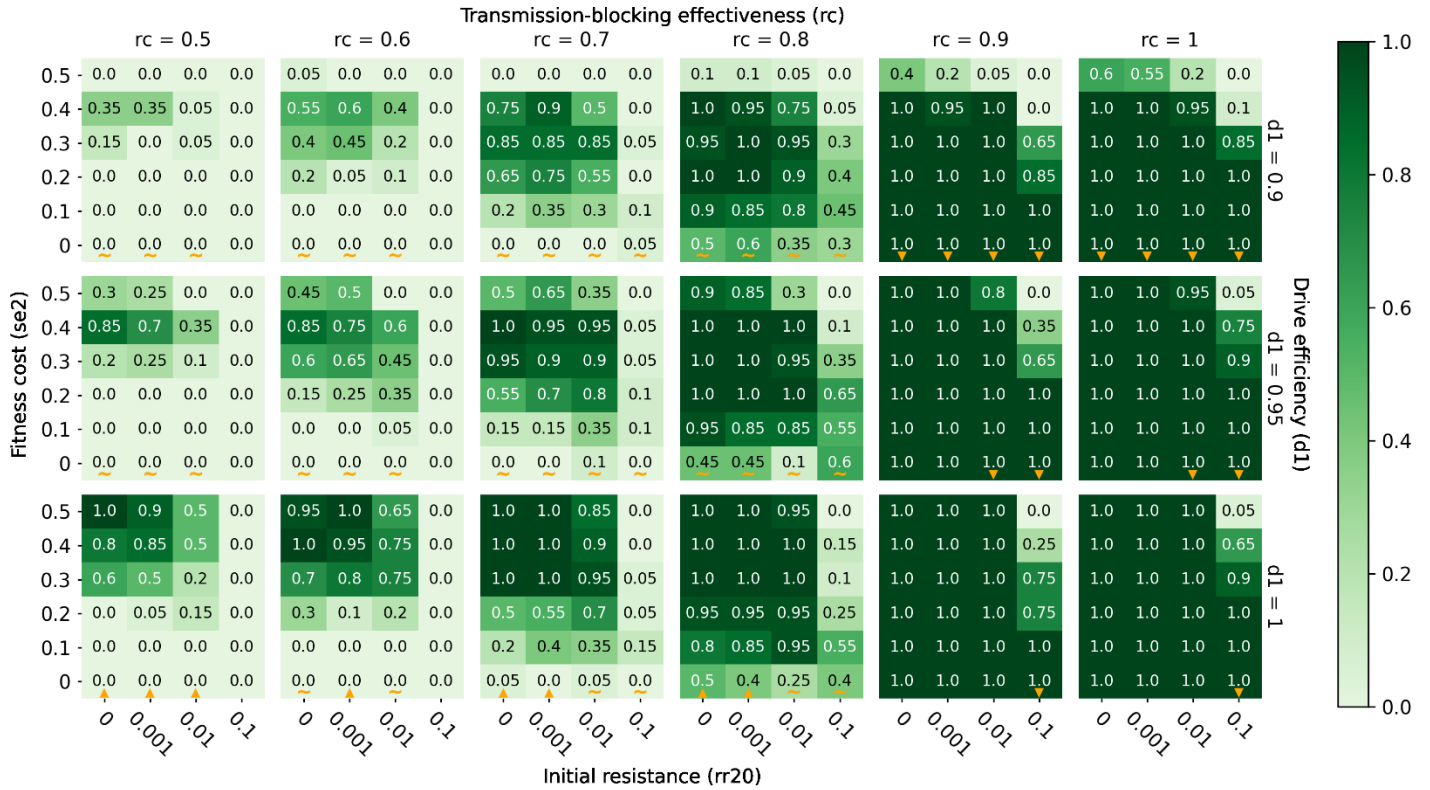
719



721 Supp. Figure 8 Elimination probabilities after a single release of integral gene
722 drive mosquitoes and ITN deployment in a low transmission (annual EIR = 10)
723 regime.

724 Same as Figure 3.

725



727 Supp. Figure 9 Elimination probabilities after a single release of integral gene
 728 drive mosquitoes and ITN deployment in a high transmission (annual EIR = 80)
 729 regime.

730 Same as Figure 3.

731

732 Acknowledgments

733 The authors would like to thank Svetlana Titova, Daniel Bridenbecker, and Clinton Collins for model and
734 software support. We would also like to thank Bee Workeneh, David Kong, Dejan Lukacevic, and Clinton
735 Collins for the tremendous amount of work they did to help us set up the interactive visualization
736 website.

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738 SL: Conceptualization, Formal analysis, Investigation, Methodology, Project administration, Software,
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745 Data availability

746 The input files, model executable, and code for running simulations as well as analyzing and plotting
747 model output can be found on Github (<https://github.com/InstituteForDiseaseModeling/leung-gene-drive-2021>). Software dependencies such as dtk-tools, dtk-tools-malaria, and the malaria-toolbox
748 packages are available upon request from support@idmod.org.
749

750 Funding

751 This work was supported by the Gates Foundation. The funders had no role in study design, data
752 collection and analysis, decision to publish, or preparation of the manuscript.

753 Competing interests

754 The authors have declared that no competing interests exist.

755

756 References

- 757 1. World malaria report 2020 - 20 years of global progress & challenges. : 300.
- 758 2. Bhatt S, Weiss DJ, Cameron E, Bisanzio D, Mappin B, Dalrymple U, et al. The effect of malaria
759 control on *Plasmodium falciparum* in Africa between 2000 and 2015. *Nature*. 2015;526: 207–211.
760 doi:10.1038/nature15535
- 761 3. Feachem RGA, Chen I, Akbari O, Bertozzi-Villa A, Bhatt S, Binka F, et al. Malaria eradication within a
762 generation: ambitious, achievable, and necessary. *The Lancet*. 2019;394: 1056–1112.
763 doi:10.1016/S0140-6736(19)31139-0
- 764 4. Cook J, Tomlinson S, Kleinschmidt I, Donnelly MJ, Akogbeto M, Adechoubou A, et al. Implications
765 of insecticide resistance for malaria vector control with long-lasting insecticidal nets: trends in
766 pyrethroid resistance during a WHO-coordinated multi-country prospective study. *Parasites
767 Vectors*. 2018;11: 550. doi:10.1186/s13071-018-3101-4
- 768 5. Hancock PA, Hendriks CJM, Tangena J-A, Gibson H, Hemingway J, Coleman M, et al. Mapping
769 trends in insecticide resistance phenotypes in African malaria vectors. *PLOS Biology*. 2020;18:
770 e3000633. doi:10.1371/journal.pbio.3000633
- 771 6. Hemingway J, Ranson H, Magill A, Kolaczinski J, Fornadel C, Gimnig J, et al. Averting a malaria
772 disaster: will insecticide resistance derail malaria control? *The Lancet*. 2016;387: 1785–1788.
773 doi:10.1016/S0140-6736(15)00417-1
- 774 7. Sougoufara S, Doucoure S, Sembéne P, Harry M, Sokhna C. Challenges for malaria vector control in
775 sub-Saharan Africa: Resistance and behavioral adaptations in *Anopheles* populations. *Journal of
776 Vector Borne Diseases*. 2017;54: 4–15.
- 777 8. Carvalho DO, McKemey AR, Garziera L, Lacroix R, Donnelly CA, Alphey L, et al. Suppression of a
778 field population of *Aedes aegypti* in Brazil by Sustained Release of Transgenic Male Mosquitoes.
779 *PLOS Neglected Tropical Diseases*. 2015;9: e0003864. doi:10.1371/journal.pntd.0003864
- 780 9. Alphey L, McKemey A, Nimmo D, Neira Oviedo M, Lacroix R, Matzen K, et al. Genetic control of
781 *Aedes* mosquitoes. *Pathogens and Global Health*. 2013;107: 170–179.
782 doi:10.1179/2047773213Y.0000000095
- 783 10. Harris AF, McKemey AR, Nimmo D, Curtis Z, Black I, Morgan SA, et al. Successful suppression of a
784 field mosquito population by sustained release of engineered male mosquitoes. *Nat Biotechnol*.
785 2012;30: 828–830. doi:10.1038/nbt.2350
- 786 11. Lacroix R, McKemey AR, Raduan N, Wee LK, Ming WH, Ney TG, et al. Open Field Release of
787 Genetically Engineered Sterile Male *Aedes aegypti* in Malaysia. *PLOS ONE*. 2012;7: e42771.
788 doi:10.1371/journal.pone.0042771
- 789 12. Harris AF, Nimmo D, McKemey AR, Kelly N, Scaife S, Donnelly CA, et al. Field performance of
790 engineered male mosquitoes. *Nat Biotechnol*. 2011;29: 1034–1037. doi:10.1038/nbt.2019
- 791 13. Adolfi A, Gantz VM, Jasinskiene N, Lee H-F, Hwang K, Terradas G, et al. Efficient population
792 modification gene-drive rescue system in the malaria mosquito *Anopheles stephensi*. *Nat
793 Commun*. 2020;11: 5553. doi:10.1038/s41467-020-19426-0
- 794 14. Carballar-Lejarazú R, James AA. Population modification of Anopheline species to control malaria
795 transmission. *Pathogens and Global Health*. 2017;111: 424–435.
796 doi:10.1080/20477724.2018.1427192
- 797 15. Champer J, Buchman A, Akbari OS. Cheating evolution: engineering gene drives to manipulate the
798 fate of wild populations. *Nat Rev Genet*. 2016;17: 146–159. doi:10.1038/nrg.2015.34
- 799 16. Burt A. Site-specific selfish genes as tools for the control and genetic engineering of natural
800 populations. *Proceedings of the Royal Society of London Series B: Biological Sciences*. 2003 [cited
801 16 Aug 2021]. doi:10.1098/rspb.2002.2319

- 802 17. James A, Beerntsen B, Capurro M, Coates C, Coleman J, Jasinskiene N, et al. Controlling malaria
803 transmission with genetically-engineered, Plasmodium-resistant mosquitoes: Milestones in a
804 model system. *Parassitologia*. 1999;41: 461–71.
- 805 18. Wang S, Jacobs-Lorena M. Genetic approaches to interfere with malaria transmission by vector
806 mosquitoes. *Trends in Biotechnology*. 2013;31: 185–193. doi:10.1016/j.tibtech.2013.01.001
- 807 19. Isaacs AT, Li F, Jasinskiene N, Chen X, Nirmala X, Marinotti O, et al. Engineered Resistance to
808 Plasmodium falciparum Development in Transgenic Anopheles stephensi. *PLOS Pathogens*. 2011;7:
809 e1002017. doi:10.1371/journal.ppat.1002017
- 810 20. Isaacs AT, Jasinskiene N, Tretiakov M, Thiery I, Zettor A, Bourgouin C, et al. Transgenic Anopheles
811 stephensi coexpressing single-chain antibodies resist Plasmodium falciparum development. *PNAS*.
812 2012;109: E1922–E1930. doi:10.1073/pnas.1207738109
- 813 21. Ito J, Ghosh A, Moreira LA, Wimmer EA, Jacobs-Lorena M. Transgenic anopheline mosquitoes
814 impaired in transmission of a malaria parasite. *Nature*. 2002;417: 452–455. doi:10.1038/417452a
- 815 22. Corby-Harris V, Drexler A, Jong LW de, Antonova Y, Pakpour N, Ziegler R, et al. Activation of Akt
816 Signaling Reduces the Prevalence and Intensity of Malaria Parasite Infection and Lifespan in
817 Anopheles stephensi Mosquitoes. *PLOS Pathogens*. 2010;6: e1001003.
818 doi:10.1371/journal.ppat.1001003
- 819 23. Adelman ZN, Kojin BB. Malaria-Resistant Mosquitoes (Diptera: Culicidae); The Principle is Proven,
820 But Will the Effectors Be Effective? *Journal of Medical Entomology*. 2021 [cited 28 Aug 2021].
821 doi:10.1093/jme/tjab090
- 822 24. Champer J, Reeves R, Oh SY, Liu C, Liu J, Clark AG, et al. Novel CRISPR/Cas9 gene drive constructs
823 reveal insights into mechanisms of resistance allele formation and drive efficiency in genetically
824 diverse populations. *PLOS Genetics*. 2017;13: e1006796. doi:10.1371/journal.pgen.1006796
- 825 25. Hammond AM, Kyrou K, Bruttini M, North A, Galizi R, Karlsson X, et al. The creation and selection
826 of mutations resistant to a gene drive over multiple generations in the malaria mosquito. *PLOS*
827 *Genetics*. 2017;13: e1007039. doi:10.1371/journal.pgen.1007039
- 828 26. Champer J, Liu J, Oh SY, Reeves R, Luthra A, Oakes N, et al. Reducing resistance allele formation in
829 CRISPR gene drive. *PNAS*. 2018;115: 5522–5527. doi:10.1073/pnas.1720354115
- 830 27. Kranjc N, Crisanti A, Nolan T, Bernardini F. Anopheles gambiae Genome Conservation as a
831 Resource for Rational Gene Drive Target Site Selection. *Insects*. 2021;12: 97.
832 doi:10.3390/insects12020097
- 833 28. James S, Collins FH, Welkhoff PA, Emerson C, Godfray HCJ, Gottlieb M, et al. Pathway to
834 Deployment of Gene Drive Mosquitoes as a Potential Biocontrol Tool for Elimination of Malaria in
835 Sub-Saharan Africa: Recommendations of a Scientific Working Group. *The American Journal of*
836 *Tropical Medicine and Hygiene*. 2018;98: 1–49. doi:10.4269/ajtmh.18-0083
- 837 29. Brossard D, Belluck P, Gould F, Wirz CD. Promises and perils of gene drives: Navigating the
838 communication of complex, post-normal science. *PNAS*. 2019;116: 7692–7697.
839 doi:10.1073/pnas.1805874115
- 840 30. Connolly JB, Mumford JD, Fuchs S, Turner G, Beech C, North AR, et al. Systematic identification of
841 plausible pathways to potential harm via problem formulation for investigational releases of a
842 population suppression gene drive to control the human malaria vector Anopheles gambiae in
843 West Africa. *Malaria Journal*. 2021;20: 170. doi:10.1186/s12936-021-03674-6
- 844 31. Beaghton A, Hammond A, Nolan T, Crisanti A, Godfray HCJ, Burt A. Requirements for Driving
845 Antipathogen Effector Genes into Populations of Disease Vectors by Homing. *Genetics*. 2017;205:
846 1587–1596. doi:10.1534/genetics.116.197632
- 847 32. Selvaraj P, Wenger EA, Bridenbecker D, Windbichler N, Russell JR, Gerardin J, et al. Vector genetics,
848 insecticide resistance and gene drives: An agent-based modeling approach to evaluate malaria
849 transmission and elimination. Davenport MP, editor. *PLoS Comput Biol*. 2020;16: e1008121.

- 850 doi:10.1371/journal.pcbi.1008121
- 851 33. North AR, Burt A, Godfray HCJ. Modelling the suppression of a malaria vector using a CRISPR-Cas9
852 gene drive to reduce female fertility. *BMC Biology*. 2020;18: 98. doi:10.1186/s12915-020-00834-z
- 853 34. North A, Burt A, Godfray HCJ. Modelling the spatial spread of a homing endonuclease gene in a
854 mosquito population. *Journal of Applied Ecology*. 2013;50: 1216–1225. doi:10.1111/1365-
855 2664.12133
- 856 35. C HMS, Wu SL, Bennett JB, Marshall JM. MGDriVE: A modular simulation framework for the spread
857 of gene drives through spatially explicit mosquito populations. *Methods in Ecology and Evolution*.
858 2020;11: 229–239. doi:10.1111/2041-210X.13318
- 859 36. Marshall JM, Buchman A, Sánchez C. HM, Akbari OS. Overcoming evolved resistance to population-
860 suppressing homing-based gene drives. *Sci Rep*. 2017;7: 3776. doi:10.1038/s41598-017-02744-7
- 861 37. Nash A, Urdaneta GM, Beaghton AK, Hoermann A, Papathanos PA, Christophides GK, et al. Integral
862 gene drives for population replacement. *Biology Open*. 2018; bio.037762. doi:10.1242/bio.037762
- 863 38. Eckhoff PA, Wenger EA, Godfray HCJ, Burt A. Impact of mosquito gene drive on malaria elimination
864 in a computational model with explicit spatial and temporal dynamics. *Proc Natl Acad Sci USA*.
865 2017;114: E255–E264. doi:10.1073/pnas.1611064114
- 866 39. Epidemiological Modeling Software. Institute for Disease Modeling; 2021. Available:
867 <http://idmod.org>
- 868 40. Eckhoff PA. A malaria transmission-directed model of mosquito life cycle and ecology. *Malar J*.
869 2011;10: 303. doi:10.1186/1475-2875-10-303
- 870 41. Selvaraj P, Wenger EA, Gerardin J. Seasonality and heterogeneity of malaria transmission
871 determine success of interventions in high-endemic settings: a modeling study. *BMC Infectious*
872 *Diseases*. 2018;18: 413. doi:10.1186/s12879-018-3319-y
- 873 42. Center for International Earth Science Information Network. [cited 16 Aug 2021]. Available:
874 <https://www.ciesin.columbia.edu/data/hrsl/>
- 875 43. Molineaux L, Gramiccia G. The Garki Project. Research on the epidemiology and control of malaria
876 in the Sudan savanna of West Africa. *Transactions of the Royal Society of Tropical Medicine and*
877 *Hygiene*. 1981;75: 190–191. doi:10.1016/0035-9203(81)90085-7
- 878 44. Huestis DL, Dao A, Diallo M, Sanogo ZL, Samake D, Yaro AS, et al. Windborne long-distance
879 migration of malaria mosquitoes in the Sahel. *Nature*. 2019;574: 404–408. doi:10.1038/s41586-
880 019-1622-4
- 881 45. Collins KA, Ouedraogo A, Guelbeogo WM, Awandu SS, Stone W, Soulama I, et al. Investigating the
882 impact of enhanced community case management and monthly screening and treatment on the
883 transmissibility of malaria infections in Burkina Faso: study protocol for a cluster-randomised trial.
884 *BMJ Open*. 2019;9: e030598. doi:10.1136/bmjopen-2019-030598
- 885 46. Selvaraj P, Suresh J, Wenger EA, Bever CA, Gerardin J. Reducing malaria burden and accelerating
886 elimination with long-lasting systemic insecticides: a modelling study of three potential use cases.
887 *Malaria Journal*. 2019;18: 307. doi:10.1186/s12936-019-2942-4
- 888 47. Eisele TP, Bennett A, Silumbe K, Finn TP, Chalwe V, Kamuliwo M, et al. Short-term Impact of Mass
889 Drug Administration With Dihydroartemisinin Plus Piperaquine on Malaria in Southern Province
890 Zambia: A Cluster-Randomized Controlled Trial. *The Journal of Infectious Diseases*. 2016;214:
891 1831–1839. doi:10.1093/infdis/jiw416
- 892 48. Thomas CJ, Cross DE, Bøgh C. Landscape Movements of *Anopheles gambiae* Malaria Vector
893 Mosquitoes in Rural Gambia. Shiff C, editor. *PLoS ONE*. 2013;8: e68679.
894 doi:10.1371/journal.pone.0068679
- 895 49. Achieving and maintaining universal coverage with long-lasting insecticidal nets for malaria
896 control. : 4.
- 897 50. Schmidt H, Collier TC, Hanemaaijer MJ, Houston PD, Lee Y, Lanzaro GC. Abundance of conserved

- 898 CRISPR-Cas9 target sites within the highly polymorphic genomes of *Anopheles* and *Aedes*
899 mosquitoes. *Nat Commun.* 2020;11: 1425. doi:10.1038/s41467-020-15204-0
- 900 51. Carballar-Lejarazú R, Ogaugwu C, Tushar T, Kelsey A, Pham TB, Murphy J, et al. Next-generation
901 gene drive for population modification of the malaria vector mosquito, *Anopheles gambiae*. *PNAS.*
902 2020;117: 22805–22814. doi:10.1073/pnas.2010214117
- 903 52. Kyrou K, Hammond AM, Galizi R, Kranjc N, Burt A, Beaghton AK, et al. A CRISPR–Cas9 gene drive
904 targeting doublesex causes complete population suppression in caged *Anopheles gambiae*
905 mosquitoes. *Nat Biotechnol.* 2018;36: 1062–1066. doi:10.1038/nbt.4245
- 906 53. Gantz VM, Jasinskiene N, Tatarenkova O, Fazekas A, Macias VM, Bier E, et al. Highly efficient Cas9-
907 mediated gene drive for population modification of the malaria vector mosquito *Anopheles*
908 *stephensi*. *PNAS.* 2015;112: E6736–E6743. doi:10.1073/pnas.1521077112
- 909 54. Hammond A, Galizi R, Kyrou K, Simoni A, Siniscalchi C, Katsanos D, et al. A CRISPR-Cas9 gene drive
910 system targeting female reproduction in the malaria mosquito vector *Anopheles gambiae*. *Nat*
911 *Biotechnol.* 2016;34: 78–83. doi:10.1038/nbt.3439
- 912 55. Pham TB, Phong CH, Bennett JB, Hwang K, Jasinskiene N, Parker K, et al. Experimental population
913 modification of the malaria vector mosquito, *Anopheles stephensi*. *PLOS Genetics.* 2019;15:
914 e1008440. doi:10.1371/journal.pgen.1008440
- 915 56. McArthur CC, Meredith JM, Eggleston P. Transgenic *Anopheles gambiae* Expressing an Antimalarial
916 Peptide Suffer No Significant Fitness Cost. *PLOS ONE.* 2014;9: e88625.
917 doi:10.1371/journal.pone.0088625
- 918 57. Dong S, Fu X, Dong Y, Simões ML, Zhu J, Dimopoulos G. Broad spectrum immunomodulatory
919 effects of *Anopheles gambiae* microRNAs and their use for transgenic suppression of *Plasmodium*.
920 *PLOS Pathogens.* 2020;16: e1008453. doi:10.1371/journal.ppat.1008453
- 921 58. Hoermann A, Tapanelli S, Capriotti P, Del Corsano G, Masters EK, Habtewold T, et al. Converting
922 endogenous genes of the malaria mosquito into simple non-autonomous gene drives for
923 population replacement. Messer PW, Tautz D, Lawniczak M, Marois E, editors. *eLife.* 2021;10:
924 e58791. doi:10.7554/eLife.58791
- 925 59. Dong Y, Simões ML, Dimopoulos G. Versatile transgenic multistage effector-gene combinations for
926 *Plasmodium falciparum* suppression in *Anopheles*. *Science Advances.* 2020 [cited 1 Sep 2021].
927 Available: <https://www.science.org/doi/abs/10.1126/sciadv.aay5898>
- 928 60. Dong Y, Das S, Cirimotich C, Souza-Neto JA, McLean KJ, Dimopoulos G. Engineered *Anopheles*
929 Immunity to *Plasmodium* Infection. *PLOS Pathogens.* 2011;7: e1002458.
930 doi:10.1371/journal.ppat.1002458
- 931 61. Meredith JM, Basu S, Nimmo DD, Larget-Thierry I, Warr EL, Underhill A, et al. Site-Specific
932 Integration and Expression of an Anti-Malarial Gene in Transgenic *Anopheles gambiae* Significantly
933 Reduces *Plasmodium* Infections. *PLOS ONE.* 2011;6: e14587. doi:10.1371/journal.pone.0014587
- 934 62. Volohonsky G, Hopp A-K, Saenger M, Soichot J, Scholze H, Boch J, et al. Transgenic Expression of
935 the Anti-parasitic Factor TEP1 in the Malaria Mosquito *Anopheles gambiae*. *PLOS Pathogens.*
936 2017;13: e1006113. doi:10.1371/journal.ppat.1006113
- 937 63. James SL, Marshall JM, Christophides GK, Okumu FO, Nolan T. Toward the Definition of Efficacy
938 and Safety Criteria for Advancing Gene Drive-Modified Mosquitoes to Field Testing. *Vector-Borne*
939 *and Zoonotic Diseases.* 2020;20: 237–251. doi:10.1089/vbz.2019.2606