

# 1 On the hindlimb biomechanics of the 2 avian take-off leap

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## 21 **Abstract**

22 Although extant land birds take to the air by leaping, generating the initial take-off velocity primarily  
23 from the hindlimbs, the detailed musculoskeletal mechanics remain largely unknown. We therefore  
24 simulated *in silico* the take-off leap of the zebra finch, *Taeniopygia guttata*, a model species of  
25 passerine, a class of bird which includes over half of all extant bird species. A 3D computational  
26 musculoskeletal model of the zebra finch hindlimb, comprising of 43 musculotendon units was  
27 developed and driven with previously published take-off ground reaction forces and kinematics. Using  
28 inverse dynamics, the external moments at the ankle, knee, and hip joints were calculated and  
29 contrasted to the cumulative muscle capability to balance these moments. Mean peak external flexion  
30 moments at the hip and ankle were 0.55 bodyweight times leg length (BWL) each whilst peak knee  
31 extension moments were about half that value (0.29 BWL). Muscles had the capacity to generate  
32 146%, 230%, and 212 % of the mean peak external moments at the hip, knee, and ankle, respectively.  
33 Similarities in hindlimb morphology and external loading across passerine species suggest that the  
34 effective take-off strategy employed by the zebra finch may be shared across the passerine clade and  
35 therefore half of all birds.

36

## 37 Introduction

38 Take-off is a crucial part of avian flight, requiring energy-intensive motion to accelerate into the air.  
39 Understanding how birds make the transition from standing statically to being airborne is one of the  
40 key components necessary for understanding how avian flight evolved. Previous work on the take-off  
41 of a variety of land birds demonstrated that the hindlimb plays a major role in propelling the bird in  
42 to the air, and suggests that the bipedal leap generates approximately 80-90% of the take-off velocity  
43 [1-6]. The group Passeriformes (passerines) includes over 5000 species of bird, making up  
44 approximately 60% of all bird species [7-9]. Among them, the zebra finch (*Taeniopygia guttata*), a  
45 frequently utilised model species, is also known to primarily use its hindlimbs to take to the air:  
46 previous studies showed that the hindlimb is responsible for producing 94% of their take-off velocity  
47 [2]. To date however, the detailed internal hindlimb mechanics necessary to produce a successful  
48 take-off remain largely unknown. This lack of knowledge in extant birds not only limits our  
49 understanding about how they master the feat of taking to the air but also presents an obstacle to  
50 accurately infer the capacity of fossil birds to be airborne, thus blurring our understanding of the origin  
51 of flight more generally.

52 Computational biomechanical modelling is a useful tool for calculating the internal mechanics  
53 occurring within a biomechanical system which are otherwise very difficult if not impossible to directly  
54 measure [10-15]. Moreover, the application of such computational analysis methods to extant animals  
55 is seen as a key strategy to systematically develop the sound biomechanical basis on which to further  
56 our understanding also of the conditions in extinct species [16-18]. The inverse analysis approach is  
57 one such computational method that takes measured kinematics and kinetics to drive a biomechanical  
58 model to calculate the internal moments at each degree of freedom of each joint [14, 19-22]. Detailed,  
59 3D models of the musculoskeletal anatomy allow relating these external moments to the moment  
60 generating capacity of the internal force generating structures, i.e. the muscles, and to develop a more  
61 detailed understanding of internal avian hindlimb kinetics during the take-off leap. However, few

62 studies have reported on the detailed external kinetics (ground reaction forces) [2, 4-6] associated  
63 with the avian take-off leap and even fewer studies have investigated 3D hindlimb kinematics for these  
64 activities [1]. The authors are aware of only a single study that has investigated the biomechanics of  
65 the avian jump, focussing on a predictive simulation of the ground dwelling elegant-crested tinamou  
66 *Eudromia elegans* [23]. However, due to the scarcity in ground reaction forces and kinematic data of  
67 the tinamou leap, data on which an inverse analysis would rely on, the authors opted for a forward  
68 approach to predict the leaping behaviour. In how far the simulations reflect kinetics and kinematics  
69 that are consistent with actual conditions that can be observed and measured in this bird therefore  
70 remains to be established. Although computational analyses in birds are available and have shown the  
71 value of such analyses to further our understanding of avian hindlimb biomechanics with respect to  
72 e.g. the critical role of the ankle muscles in the take-off leap of the tinamou and the function of the  
73 antitrochanter as a passive mechanism to stabilise the hip of the running ostrich [23, 24], to date no  
74 study has determined the internal hindlimb joint kinetics of a flying bird as it leaps into the air using  
75 detailed measurements of external forces and hind limb kinematics. With the application of X-ray  
76 reconstruction of moving morphology (XROMM) technology to capture detailed 3D bone kinematics  
77 of the avian take-off leap, in combination with computational biomechanical analyses, the technology  
78 is finally available to accurately simulate the internal mechanics of the extant avian take-off [1, 10, 16,  
79 25-28].

80 The current study therefore combines external forces and detailed bone kinematics that feed into  
81 computer simulations into the biomechanics of the hindlimb throughout the take-off leap of the zebra  
82 finch. In doing so we aim to address the following hypotheses that help to develop a comprehensive  
83 understanding of the mechanical requirements birds need to meet to propel themselves into the air.  
84 We firstly hypothesised that in order to generate the motion of the avian take-off leap, characterised  
85 by the hip, knee, and ankle joints all extending until the bird is airborne [1, 3], net external flexion  
86 moments of similar peak magnitudes act at all these joints which the muscles balance by exerting  
87 extension moments. Consistent with the observation that predictive simulations of the tinamou

88 predict the ankle to be most critical in determining the success of the take-off leap in a ground-  
89 dwelling bird [23], we further hypothesised that the ankle extensors of the zebra finch possess the  
90 largest capacity to balance the external moments. Finally, considering the avian hips' powerful  
91 capacity to generate internal/external rotation (IER) moments and a poor capacity to generate  
92 abduction/adduction (ABAD) moments [10, 16, 24] in the presence of the antitrochanter, we  
93 hypothesise that external moments of similar peak magnitudes act in IER/ABAD on the hip joint of the  
94 zebra finch and that the bird possess powerful ability to actively balance the IER moments.

95

## 96 **Materials and methods**

97 To address our hypotheses, we built upon previously published detailed kinetics and kinematics data  
98 of the take-off leap of the zebra finch as input to a biomechanical simulation of the avian take-off leap  
99 [1, 2] as explained in more detail below.

### 100 Overview

101 Key steps of our analysis methodology included the use of CT scans of the same individual, together  
102 with additional morphological data in the literature to characterise the bone and musculature  
103 respectively, from which a detailed 3D musculoskeletal model of the zebra finch hindlimb was  
104 developed. Here, muscles were mapped from a previously published magpie musculoskeletal model  
105 on to the zebra finch skeleton [10, 29, 30]. In addition, previously published 3D kinematics and ground  
106 reaction forces were used to drive the musculoskeletal model in inverse dynamics analyses to estimate  
107 external joint moments [1, 2]. The combination of these unique datasets, collected from the same  
108 species and even the same individual, offered a unique opportunity to generate an accurate simulation  
109 of the zebra finch take-off biomechanics.

110 The external joint moments were compared against the moment generating capacity of the muscles  
111 to document the biomechanical requirements and to assess the zebra finch's capability to actively  
112 balance the hindlimb joint moments experienced throughout the take-off leap. Here, moments about  
113 hip flex/extension (FE), ab/adduction (ABAD), int/external rotation (IER) and moments about FE at  
114 both the knee and ankle joints were considered. In addition, by comparing the hindlimb morphology  
115 and ground reaction forces of a variety of passerines ranging in mass (zebra finch *Taeniopygia guttata*,  
116 15.4 g, starling *Sturnis vulgaris*, (77.3 g), crow *Corvus corone* (440 g) and raven *Corvus corax*, 1.1 kg) it  
117 was possible to explore the take-off mechanics of passerines more generally.

### 118 Materials, model building, and musculoskeletal analysis approach

119 The computational biomechanical musculoskeletal model of a zebra finch (15.4 g) was developed  
120 based on CT scans and muscle data of corvids from previously published works [1, 10, 29]. The CT data

121 was used for establishing the 3D skeletal model. In order to describe the spatial relationships between  
122 bones, local coordinate systems were established based on shape fitting techniques and functional  
123 analyses of the joints [31], as detailed in the supplementary information (S1A). Muscles were modelled  
124 by 3D lines of action [12, 32, 33] by mapping muscles from the magpie skeleton to the zebra finch  
125 using elastic registration [34], while muscle maximum isometric force was scaled by body mass.  
126 Detailed kinematics derived from previously published XROMM data [1] were then used together with  
127 ground reaction forces [2] of zebra finch take-off leaps to drive the model in inverse dynamics  
128 simulations to calculate the external moments acting at the hindlimb joints. Nine sets of ground  
129 reaction forces obtained from 9 jumps of 4 birds ( $15.4 \pm 1.8$ g), were temporally synchronised with two  
130 sets of kinematics trials (taken from 1 bird with a mass of 15.4 g), resulting in 18 simulations analysed.  
131 Muscle moment arms were measured and used to calculate the maximum moment generating  
132 capacity about hip flexion/extension, ab/adduction and internal/external rotation, and knee and ankle  
133 flexion/extension. The muscle moment generating capacity was compared to the external joint  
134 moment to ascertain the zebra finch's ability to balance the external joint moments.

### 135 Skeletal model

136 The skeletal model was derived from CT scans (isotropic resolution 0.04mm) of a zebra finch  
137 *Taeniopygia guttata* specimen (15.4 g), obtained in previously published studies (for full details please  
138 refer to [1]). For the current study, bones were segmented using ITK-SNAP [35], and imported in to  
139 Rhino (v7; Robert McNeel & Associates, Seattle, USA) [36] where triangulated bone surfaces were  
140 obtained after fitting subdivision surfaces using the QuadRemesh function. Bones which were treated  
141 in this way were the pelvis, femur, patella, tibiotarsus and tarsometatarsus. The detailed definition of  
142 joint centres and axes and local bone coordinate systems are available in the supplementary  
143 information (S3).

144 A linked rigid body model with 4 segments including the pelvis, thigh, shank and tarsometatarsus was  
145 defined in OpenSim v4.1 [37]. Here, body segments were linked by 3 joints (hip, knee and ankle joints)

146 with 3 rotational degrees of freedom (DoF) at the hip (allowing flexion/extension, internal/external  
147 rotation and ab/adduction) and 2 DoF at each of the knee and ankle joints (allowing flexion/extension  
148 and internal/external rotation). Because the foot remained stationary on the perch and motion of the  
149 trunk was largely due to the extension of the hip, knee and ankle joints, the foot was not included as  
150 a dedicated structure of the musculoskeletal model. In order to better capture the mechanics of  
151 hindlimb extension, a biomechanical model of the patella-femoral joint was incorporated in to the  
152 model. Here, the motion of the patella was defined by a 3D spline curve following approximately the  
153 trochlear surface of the femur. The patella was allowed to move along that spline as a function of the  
154 knee angle, with details of the motion informed by a musculoskeletal model of the helmeted  
155 guineafowl *Numida meleagris* [38]. In order to map patellar motion from the guineafowl to the zebra  
156 finch model, the femoral surface of the guineafowl model was elastically registered to the zebra finch  
157 femur [39]. The parameters of the elastic registration were then used to map the spline defining  
158 patellofemoral motion from the guineafowl to the zebra finch model using the R-package MesheR  
159 [34].

## 160 Muscle geometry

161 The musculoskeletal model of the zebra finch included 43 muscles crossing the hip, knee and ankle,  
162 which were modelled as 3D polylines spanning origin and insertion using via points to fully describe  
163 their curved paths (Figure 1). Magpies (*Pica pica*) and zebra finch, share broadly similar hindlimb  
164 myology both following the characteristic hindlimb morphology of passerine birds [9]. Muscle  
165 attachment sites and via points were therefore mapped from a previously established model of the  
166 magpie [10] to the zebra finch hindlimb. Using a non-rigid iterative closest point (ICP) registration [39]  
167 the magpie femur, tibiotarsus and tarsometatarsus were elastically registered to the corresponding  
168 long bones of the zebra finch hindlimb [34] (Figure 2). Using 3D Slicer 4.11 [40], the pelvis of the  
169 magpie and zebra finch were first split in to the ilium, ischium and pubis before elastically registering  
170 them onto each other following the same approach as described for the long bones above. Following  
171 non-rigid registration, the rigid transformation and isotropic scaling parameters were recovered using



172 Ordinary Procrustes Analysis (OPA) [41] computed between the vertices of the original magpie bone  
173 surface and the vertices of the magpie bone surface that was elastically registered to the respective  
174 zebra finch bone (Matlab (2019b, The Mathworks, Nantucket, USA). This step was encoded in a 4x4  
175 homogeneous transformation matrix. The remaining difference between the positions of the OPA  
176 mapped vertices and their elastically registered counterparts was captured in a dense deformation  
177 field. The homogeneous transformation matrix and the dense deformation field were then applied to  
178 all attachment and via points of the magpie muscles associated to the respective bone surface using  
179 the R-package MesheR [34], and in so doing mapping muscles from the magpie hindlimb to the zebra  
180 finch (Figure 2). Wrapping cylinders and spheres were added with positions, orientations and radii  
181 individually adjusted to define the 3D muscle paths throughout the jumping RoM that avoided any  
182 intersection of muscles with bones.

### 183 Biomechanical analysis

#### 184 Kinematics

185 Detailed 3D kinematics were derived from previously published X-ray reconstruction of moving  
186 morphology (XROMM) data of the left hindlimb throughout two autonomous take-off leaps of the  
187 zebra finch [1]. For these analyses three tantalum bead markers with an approximate diameter of 0.5  
188 mm were attached to the tarsometatarsus, while two markers were implanted to the tibia, one to the  
189 femur, and one at the pelvis. In order to reliably track 3D skeletal kinematics for the current study, the  
190 location of the pelvis was determined using the implanted pelvic marker and, while its orientation was  
191 determined using scientific roscoping, a reliable process in particular for bones with specific shapes  
192 such as the pelvis [42]. Owing to the generally higher precision of the 3D marker positions compared  
193 to 3D position and orientation data derived from scientific roscoping [1, 42], a method for  
194 reconstructing 3D skeletal motion that maximised usage of the marker data while minimising reliance  
195 on the manual process of scientific roscoping was developed. Here, 3D positions and orientations  
196 of the long bones of the hindlimb were obtained using the attached physical markers, supplemented

197 by functionally defined virtual markers, and detailed 3D bone surface models. A full detailed  
198 description of the methodology used to define these virtual markers and track 3D skeletal motion  
199 using a combination of XROMM data, bone surfaces and virtual markers is provided in the  
200 supplementary materials section (S1, S2).

201 An inverse kinematics analysis was then carried out in OpenSim [22, 32, 37] to map the kinematics  
202 from the XROMM data to the constrained biomechanical model using physical and virtual markers  
203 placed at their known locations on the model bones. Across the two kinematics trials, the ranges of  
204 motion for the skeletal kinematics with respect to hip flexion/extension (FE), abduction/adduction  
205 (ABAD) and internal/external rotation (IER) recovered in that manner were  $42^\circ$  ( $-4^\circ$  to  $38^\circ$ ),  $15^\circ$  ( $-37^\circ$   
206 to  $-22^\circ$ ), and  $20^\circ$  ( $8^\circ$  to  $28^\circ$ ), respectively. Knee and ankle FE range of motion were  $40^\circ$  ( $123^\circ$  to  $163^\circ$ )  
207 and  $74^\circ$  ( $61^\circ$  to  $135^\circ$ ), respectively (see supplementary figure SF1). Substantial internal/external  
208 rotation range of motion at the knee and ankle was also measured, with a range of motion of  $16^\circ$  ( $5^\circ$   
209 to  $21^\circ$ ) and  $30^\circ$  ( $-15^\circ$  to  $15^\circ$ ), respectively.

210 The take-off velocity of the bird was defined by measuring the velocity of its centre of mass at the  
211 instant the feet left the ground [2, 3, 6]. To that end, the centre of mass of the zebra finch was  
212 determined using the soft tissue envelope of the zebra finch and the RigidBodyParams [43] function  
213 under the assumption of a homogenous body soft tissue density [43]. The velocity of the centre of  
214 mass of the zebra finch was then measured by tracking its location throughout the take-off trials using  
215 the OpenSim 4.1 BodyKinematics Tool. Take-off velocity was determined as the velocity at the instant  
216 all toes on the hindlimb being tracked, left the perch. Take-off velocities for the two trials investigated  
217 were determined to be 1.39 and 1.08 m/s.

218 Kinetics

219 Previously reported ground reaction forces of the take-off leap of a zebra finch [1] were used as inputs  
220 to the current study in this study. Since full details for the collection of these data were previously  
221 reported, the experimental setup is only briefly summarized below. Vertical and horizontal ground

222 reaction forces were recorded at 400 Hz from a force platform (Squirrel force plate, Kistler France, Les  
223 Ulis, France; resolution  $\pm 0.01\text{N}$ ), to which a wooden perch, 1.5 cm in diameter, was attached. The  
224 ground reaction forces measured from nine trials of a zebra finch taking off from a perch [2] were  
225 used to drive the take-off simulations.

226 Each of the nine zebra finch kinetics trials were paired with the two sets of kinematics trials enabling  
227 analysis of 18 distinct take-off conditions. Here, the ground reaction forces were temporally aligned  
228 to the kinematic data such that at the instant at which the resultant ground force fell to zero was  
229 matched to the instant at which the toes left the ground. Ground reaction forces were assumed to act  
230 through the centre of the perch (Figure 3).

231 For use in the current study, the ground reaction force vectors were scaled in magnitude so that the  
232 impulse imparted by the leg resulted in 94% take-off velocity that was measured from the kinematics  
233 for the two trials considered here. This scaling was performed to reflect previously identified  
234 relationships for the take-off mechanics of the zebra finch [2]. Here, the method for calculating the  
235 velocity as a result of the impulse ( $V_{GRF}$ ) imparted by the hindlimb followed the approach by Provini  
236 and colleagues (2012) and is outlined below. Firstly, the acceleration resulting from the ground  
237 reaction forces,  $\dot{V}_{GRF}$ , were calculated as follows:

$$238 \quad \dot{V}_{GRF} = \frac{\sqrt{F_x^2 + F_y^2 + (F_z - mg)}}{m} \quad (2)$$

239 where  $F_x$  is forward force,  $F_y$  is lateral force,  $F_z$  is vertical force,  $m$  is the mass of the bird, and  $g$  is the  
240 acceleration due to gravity. The integral of the acceleration over the duration of the take-off then  
241 provides the take-off velocity as a result of the ground reaction forces,  $V_{GRF}$ :

$$242 \quad V_{GRF} = \int_{t_0}^{l_0} \frac{\sqrt{F_x^2 + F_y^2 + (F_z - mg)}}{m} dt \quad (3)$$

243 where  $t_0$  is the time when the jump starts and  $l_0$  the time at lift off.

244 External joint moments

245 Ground reaction forces were applied to the tarsometatarsus, with the centre of pressure placed in the  
246 centre of the perch (Figure 3). Using kinematics and ground reaction forces as input to the inverse  
247 dynamics analysis, external joint moments were calculated about hip FE, ABAD, IER, knee FE and ankle  
248 FE. Joint moments were normalised by the product of bodyweight and leg length L (defined as the  
249 sum of the segment lengths of femur, tibiotarsus, tarsometatarsus, and digit III) [44], and time  
250 normalised to the duration of the jump cycle. Mean and standard deviations of the external joint  
251 moments were calculated across all 18 take-off sequences to obtain a robust estimate of the envelope  
252 of zebra finch take-off biomechanics. Here, joint moments are identified by the direction of joint  
253 movement induced by the action of the muscle group activating the respective degree of freedom.

254 Muscle moment generating capacity

255 The methodology for calculating muscle moment generating capacity followed the approach  
256 described by Meilak and colleagues (2021) and is outlined below. The moment generating capacity for  
257 each muscle about each joint degree of freedom (DoF) being considered was calculated by multiplying  
258 the muscle maximum isometric force ( $F_{max}$ ) by the instantaneous moment arm ( $MA_{t,i,j}$ ), evaluated  
259 at each time increment ( $t$ ) throughout the kinematics trials. These moments were evaluated for each  
260 muscle  $i$  (where  $i=1..43$ ) as the product of the maximum isometric muscle force ( $F_{max}$ ) and the  
261 instantaneous moment arm ( $MA_{t,i,j}$ ) determined throughout the take-off kinematics for each  
262 rotational DoF  $j$  (where  $j=1..5$  for hip FE, ABAD, IER, knee FE and ankle FE):

$$263 \quad M_{t,i,j} = F_{max,i} \cdot MA_{t,i,j} \quad (4)$$

264 The moments of muscles acting in the same direction (i.e. their moments have the same sign) were  
265 summed for each degree of freedom to provide the total joint moment generating capacity in relation  
266 to that specific action ( $M_{t,j}$ ):

$$267 \quad M_{t,j} = \sum_{i=1}^{43} M_{t,i,j} \quad (5)$$

268 Comparing biomechanical conditions across the group of Passerine birds  
269 To allow comparative analysis of conditions across the Passerine clade, the ground reaction force data  
270 available for the zebra finch was amended with similar data from further animals within the group of  
271 passerines. In a study approved by the local ethics committee of the University of Southampton (ERGO  
272 II ID 32207), a crow (*Corvus corone*) and raven (*Corvus corax*) took off 6 times each from a force  
273 platform (9260AA, Kistler) while 3D ground reaction forces (GRFs) were sampled at 10 kHz. For all  
274 species, vertical and horizontal forces were filtered using a zero-phase low-pass (40Hz) custom filter  
275 in Matlab. The peak ground reaction forces of the zebra finch, starling [3], crow and raven were all  
276 normalised by bodyweight and, together with key measures of hind limb geometry (Table 2) to  
277 compare take-off conditions within the group of passerines.  
278

## 279 **Results**

280 The duration of the take-off leap ranged between 62-65ms (Figure 4). Mean resultant peak ground  
281 reaction forces per leg were  $1.81 \pm 0.21$  BW (range: 1.42 to 2.15 BW, c.f. supplementary figure SF1) and  
282 occurred at about 62 % leap cycle time. Across the two kinematics trials, the largest ranges of motions  
283 were  $74^\circ$ ,  $42^\circ$  and  $40^\circ$  about the ankle, hip, and knee FE axes, respectively. The ranges of motion about  
284 the joints' secondary degrees of freedom were similar, with hip IER and ABAD RoMs of  $20^\circ$  and  $15^\circ$ ,  
285 respectively and knee and ankle IER RoMs of  $16^\circ$  and  $30^\circ$ , respectively (see supplementary figure SF2).

286 The largest external moments observed were the moments around the flexion/extension axes. Here,  
287 the mean peak external joint flexion moments at the hip and ankle were of similar magnitude  
288 ( $0.66 \pm 0.04$  and  $0.68 \pm 0.05$  BWL, respectively) while the mean knee extension moment was only about  
289 half that value ( $0.38 \pm 0.05$  BWL, Figure 5, Table 3). The smallest peak moments were computed about  
290 hip IER and ABAD with mean peak moments of  $0.27 \pm 0.07$  and  $0.20 \pm 0.07$  BWL, respectively (Figure 5,  
291 Table 3). At the instances that these peak moments occurred, the muscles had the ability to generate  
292 120%, 282% and 61% of the mean peak hip FE, IER and ABAD joint moments, respectively (Figure 5  
293 Table 3). The muscles were able to generate 177% and 176% of the mean peak joint moments about  
294 the FE axes of the ankle and knee, respectively (Figure 5 B and C, Table 3).

295 Comparing conditions across the group of passerines, the zebra finch as bird with the smallest mass  
296 by far (15.4 g) had the smallest normalised peak ground reaction forces per leg of  $1.94 \pm 0.25$  BW, while  
297 starling (77.3 g), crow (440 g), and raven (1.1 kg) exerted very similar peak GRFs with values of  
298  $2.15 \pm 0.14$  BW [3],  $2.20 \pm 0.29$  BW, and  $2.25 \pm 0.14$  BW, respectively (see supplementary figure SF3).

## 299 Discussion

300 This is the first detailed investigation into the hindlimb mechanics of a flying bird as it takes to the air  
301 with a take-off leap using modern biomechanical analyses. The model presented here simulated 18  
302 distinct take-off trials of a zebra finch's take-off leap from a perch covering a range of take-off  
303 conditions in a passerine bird, the group that makes up approximately 60% of all extant bird species  
304 [7-9]. In doing so our study revealed a consistent pattern of internal mechanics across the trials that  
305 was characterised by the largest peak external moments occurring around the FE axis of the hip and  
306 ankle (requirements, 0.68 and 0.66 BWL, respectively), while considerably smaller external moments  
307 were observed about the FE axis of the knee, amounting to only 56% of the respective peak moments  
308 at the hip. On the other hand, peak hip IER and ABAD moments were 40% and 29% of peak hip FE  
309 moments respectively. Together with previous findings regarding the substantial extent of joint range  
310 of motion in IER exercised by birds [1, 27] and the substantial moment generating capacity of the hip  
311 joint muscles around that axis [10], these novel data on the internal hind limb mechanics during the  
312 take-off leap strongly support the notion that the execution and control of hindlimb motion in birds is  
313 not limited to a single (sagittal) plane but is essentially 3D in nature [16].

314 Across the hip, knee and ankle FE degrees of freedom, the muscles had the ability to generate 120%,  
315 176%, and 177% of the mean peak external joint moments, respectively. In relative terms therefore,  
316 muscles spanning the hip were closest to reaching the capacity limit whilst muscles spanning the knee  
317 and ankle joints had the largest reserves. Although for the most typical (mean) of the conditions  
318 studied here the capacity of the muscles to generate moments was always larger than the  
319 requirement (Figure 5), the closing gap between the upper limit of the 2.5 SD range of the external  
320 hip flexion moments and available hip muscle extension capability suggests that more powerful take-  
321 off leaps than those observed here would likely require further activation of the more distal, knee and  
322 ankle joint spanning muscles for which the requirements remained more comfortably within their  
323 capability [23]. An explanation as to why capacity to balance the moments at the more distal joints

324 retains a larger reserve may be found in the design of the avian hindlimb and specifically the biarticular  
325 muscles [45-47]. Such biarticular muscles include in particular the m. caudofemoralis pars caudalis  
326 (MFCLP), m. flexor cruris medialis (MFCM) and m. iliofibularis (MIF) which span the caudal side of the  
327 hip and knee (Figure 1, Table 1). MFCLP and MFCM form a major part of the hip extensor moment  
328 generating capacity, together generating 35% of the total hip extensor moments whereas the MIF has  
329 a greater role as a hip abductor, generating 20% of the muscle abduction generating capacity [10].  
330 Whilst during the take take-off leap of the zebra finch the hip, knee and ankle are all extending (Figure  
331 SF1), the muscles need to generate a net knee flexion moment, suggesting that knee flexors may be  
332 activated. Biarticular muscles, which if activated generate an extension moment at the hip but a  
333 flexion moment at the knee such as the MFCLP and MFCM, would thus appear to be prime candidates  
334 for meeting the demands during the take-off leap. Moreover, co-activation of knee flexors and  
335 extensor muscles (muscle co-contraction) though energetically less optimal, may help to increase  
336 compressive forces across the knee joint and help to lock or at least minimise the extent of  
337 internal/external rotation [27, 28, 48]. Further analysis to examine muscle activation patterns, though  
338 beyond the scope of the current work, could corroborate whether activation of these biarticular  
339 muscles to extend the hip and generate the extension moment applied to the knee suggested by the  
340 analyses here does indeed occur and help to further elucidate how avian hindlimb muscles are  
341 orchestrated during the take-off leap.

342 Here, it is also informative to consider absolute moments where indeed the ankle extensors had the  
343 largest joint moment generating capacity, capable of generating peak total extension moments of up  
344 to 1.18 BWL at the ankle, followed by the hip extensors, with a peak total capacity of 0.81 BWL. The  
345 passerine bird ankle muscles' capacity to produce the largest moments indicates the importance of  
346 the ankle joint throughout the take-off, a finding in line with previous research on the ground dwelling  
347 elegant-crested tinamou *Eudromia elegans* [23] where the take-off heavily depended on the  
348 parameters and activation of the ankle extensors. Though to the authors knowledge no similar studies  
349 reporting internal mechanics during a take-off leap of a flying bird are available for comparison, similar



350 analyses have been performed in ground-dwelling birds such as the emu and the ostrich. Here, data  
351 on the running of the ostrich obtained using similar analysis methods revealed that the peak  
352 normalised FE moments across all of the hip, knee, and ankle joint of the ostrich were considerably  
353 smaller than those reported during the take-off leap of the zebra finch reported here. The largest  
354 differences in normalised moment magnitudes were observed for the ankle and hip joints where the  
355 peak moments during running in the ostrich amounted to only 24% (0.16 BWL) and 25% (0.17 BWL)  
356 of the values for the zebra finch leap. The external moment at the knee during running in the ostrich  
357 was the largest of all 3 hindlimb joints with 0.20 BWL yet amounted to only about 53% of the value  
358 the current study calculated for the take-off leap of the zebra finch. In the comparison of the absolute  
359 moment magnitudes it is important to consider that speed at which the ostriches were running was  
360 rather slow (3.24 m/s) compared to the maximum speed ostriches can achieve (about 13.9 m/s) and  
361 higher speeds will be associated with higher external forces and moments. However, not only absolute  
362 magnitudes but also the ratios of their magnitudes at the hip, knee, and ankle differed between the  
363 zebra finch take-off leap and ostrich running. While the largest external moment during running of the  
364 ostrich was computed at the knee, suggesting that that ostrich running is knee driven, the external  
365 knee moment during the take-off leap of the zebra finch was the smallest of all the hind limb joints.  
366 For the zebra finch leap, peak moments were observed at the hip and ankle suggesting that this  
367 motion is hip and ankle driven instead and signifying that an interesting avenue for future work would  
368 be to investigate whether the different motor behaviours are indeed associated with different muscle  
369 coordination patterns and that care must be taken when speculating about design principles and  
370 interpreting optimality of the musculoskeletal system based on a limited repertoire of motor  
371 behaviours.

372 Previous studies have demonstrated that avian pelvic muscles have a considerable capacity to produce  
373 IER moments at the hip [10, 16, 49]. The current study revealed not only that substantial external  
374 moments about the IER axis occur during the take-off leap of the zebra finch, approaching 40% of the  
375 hip extension moment, but also demonstrated that the hip muscles had the greatest relative capacity

376 to actively balance these moments, with muscles capable of generating up to 280% of the mean peak  
377 external IER moments. Together these data provide further evidence that IER rotation is actively  
378 controlled during routine, straight line take-off leap of passerine birds. The ample capacity of the  
379 muscles to enable IER during such jumps further allows for take-off leaps to occur with substantially  
380 more axial turning/rotation while the bird remains on the ground, offering the bird a wider repertoire  
381 of take-off and escape behaviours. On the other hand, foraging behaviours have been shown to be  
382 linked with substantial extents of hindlimb IER [27] and it may well be that substantial IER capability  
383 of the hip muscles are particularly crucial in supporting those behaviours (particularly for single limb  
384 support) in addition to allowing variation in the take-off leap.

385 In contrast to the well powered hip IER DoF during the take-off leap, the pelvic muscles of the zebra  
386 finch were only able to balance 61% of the peak mean external adduction moments. Analysis of the  
387 relationship of the relative moment generating capacity of the avian hip muscles do indeed  
388 demonstrate that the smallest capacity to generate a moment exists with respect to the ABAD axis.  
389 However, even though the avian hip has a relatively limited capacity to actively produce hip ABAD  
390 moments, birds can rely on a passive mechanism using the antitrochanter and associated ligamentous  
391 structures to balance external abduction moments [49]. The utilisation of the antitrochanter is also a  
392 feature in the hip mechanics of the ostrich: though substantial external abduction moments are  
393 generated during walking and running in these flightless birds, the design of the hip allows to stabilise  
394 the joint passively such that ostriches neither require nor possess muscles to do so actively [49, 50].  
395 The antitrochanter is indeed a feature shared across all extant birds that was not present in some of  
396 the very earliest birds such as *Archaeopteryx*, who likely relied on powerful hip adductors to generate  
397 the required adduction moments [16].

398 Maintaining substantial lever arms of the muscles throughout the functional range of motion of a joint  
399 is a prerequisite to maintain high levels of muscle capacity to generate moments. Passive structures  
400 can play a key role to help maintaining muscle lever arms include sesamoid bones such as the patella

401 which is key to enable joint function in flexion at the knees in human and avian bipeds [33, 46] The  
402 hypotarsus is another anatomical feature of the avian hindlimb that helps to guide tendons around  
403 the ankle joint and to and maintain their lever arms [51]. The posterior side of the ankle further  
404 includes the cartilago tibialis which constrains the muscle line of action to act further away from the  
405 joint axis of rotation thus helping to maintain its moment [29]. The model presented here therefore  
406 modelled the patellofemoral joint in an approach informed by the birds' bone surface anatomy and  
407 kinematic model data from the literature [38] and in incorporated these passive structures of the ankle  
408 with wrapping objects and via points informed by CT scans.

409 The relative peak ground reaction forces during the take-off leap of all passerines considered here  
410 were rather similar in magnitude, despite the large range in body mass. During the take-off leap the  
411 zebra finch (15.4 g), starling (77.3 g), crow (440 g) and raven (1.1 kg) generated peak ground reaction  
412 forces per leg of  $1.94 \pm 0.25$  BW,  $2.15 \pm 0.14$  BW [3],  $2.2 \pm 0.29$  BW, and  $2.25 \pm 0.14$  BW, respectively.  
413 Similarities do not stop with the ground reaction forces but are also apparent in the morphology of  
414 their hindlimbs. The passerines considered here, the zebra finch, starling, crow, and raven, possess  
415 very similar hindlimb indices, a measure of relative segment lengths of the hindlimb ((tarsometatarsus  
416 length + tibiotarsus length) / femur length [52]) with values of 2.64, 2.84, 2.75, and 2.63, respectively  
417 [53] (Table 2). The similarities in normalised leg segment lengths and ground reaction forces spanning  
418 a range of passerines, which differ in mass by approximately two orders of magnitude, support the  
419 hypothesis that passerines share a similar take-off behaviour, as reported here. Forward dynamics  
420 simulations of the tinamou (0.55 kg) leaping predict similar but somewhat higher peak ground reaction  
421 forces of 2.62 BW per leg during the jump [23]. On the other hand, predicted joint kinematics profiles  
422 for the take-off leap of the tinamou demonstrated ranges of motion at the hip knee and ankle of  $65^\circ$ ,  
423  $91^\circ$  and  $109^\circ$  respectively, consistently greater than the range of motion measured in the zebra finch  
424 ( $42^\circ$ ,  $40^\circ$  and  $74^\circ$  respectively), while the hindlimb index of the tinamou *Eudromia elegans* (belonging  
425 to the group Tinamiformes) was 2.21, substantially smaller than that of the passerines considered  
426 here. The relative difference in relative leg morphology between the passerines and tinamou could be

427 one of the determining factors resulting in the variability in take-off mechanics between the clades of  
428 birds.

429 The kinematics used to drive the take-off simulation were informed by previously obtained XROMM  
430 data of the zebra finch take-off leaps [1]. In this study, the use of the tantalum bead markers, detailed  
431 bone surface geometry from high resolution  $\mu$ CT, and anatomical-functional relationships [31, 54]  
432 were all used to increase repeatability in tracking 3D skeletal kinematics and reduce the influence of  
433 the user during scientific roto-scoping [42]. Across the two kinematics trials, the difference between  
434 the originally published orientation angles of the femur, tibiotarsus and tarsometatarsus was smallest  
435 about FE and ABAD axes of the bones, with mean differences ranging from  $0.6 \pm 0.5^\circ$  to  $3.3 \pm 2.7^\circ$ . The  
436 degree of freedom most difficult to determine during scientific roto-scoping was IER due to the  
437 cylindrical shape of the long bones. Predictably, the largest difference was observed when comparing  
438 the bone IER orientations, with mean differences in IER orientations across the three long bones  
439 ranging from  $7.3 \pm 4.8^\circ$  to  $11.5 \pm 6.3^\circ$ . Differences in bone locations between the two methods were  
440 minimal, mean differences across all three long bones ranged between 185 – 333 microns.

441 Due to the limited number of specimens in which sufficient bones had had a minimum of three  
442 tantalum markers attached, only two sets of kinematics trials were used in this study. However, by  
443 pairing each set of kinematic trials with nine sets of kinetics trials, we maximised the variability in take-  
444 off conditions studied here. Future studies, using XROMM to capture detailed kinematics of passerines  
445 should ensure that at least one of the long bones includes at least 3 markers to reduce the reliance on  
446 the user during scientific roto-scoping. The simulations here included IER motion of the knee and ankle  
447 joints, however the active muscle actuation of these degrees of freedom was not considered in line  
448 with previous studies and under the assumption that typically small moments are balanced by passive  
449 structures such as ligaments [37, 55]. Previous studies reported the take-off velocity of the zebra finch  
450 to be around 1.7m/s [2, 6] which is faster than the take-off velocity, measured here from the XROMM  
451 data which ranged between 1.08-1.39 m/s. Previously published studies measuring the kinematics of

452 animal subjects using implanted markers have reported the markers causing a limp [14]. The  
453 comparatively slower take-off velocities reported here could thus be attributed to a response to the  
454 implantation of the tantalum bead markers on the hindlimb bones. However, varying levels of  
455 motivation between experimental conditions may also explain the observed difference and the similar  
456 protocols using tantalum bead markers have been used to study a range of motor behaviours in birds  
457 [46, 56, 57]. This study did not include the mechanics of phalanges in the analysis, as the take-off  
458 trajectories of the hindlimb are defined primarily by the motion at the more proximal hindlimb joints  
459 where also more substantial joint moments are generated. Future studies which may consider hind  
460 limb mechanics during landing, when the detailed mechanics of foot are likely to play a more  
461 important role, should aim to capture the detailed kinematics of the phalanges.

462 This study considered the maximal moment generating capacity of the muscles, taking into account  
463 the muscle maximum isometric force and instantaneous moment arm throughout the take-off leap  
464 and contrasted these to the external moments applied to the joints of the hind limb. In this way  
465 general patterns of mechanical requirement and hindlimb muscles capability to meet the  
466 requirements of a take-off leap were analysed. Although the sensitivity of muscle moment generating  
467 capacity with respect to uncertainty in the definition of muscle geometry was not directly investigated  
468 in this study, a thorough sensitivity analysis was performed for pelvic muscles in the magpie hindlimb  
469 model that was the basis for the current study [10], which, given the similarity in overall hindlimb  
470 design and specific hindlimb bone morphology, can reasonably be expected to remain valid for the  
471 musculature of the zebra finch model here. Though the determination of the detailed muscle  
472 activation patterns to balance the external moments was not within the scope of the current study,  
473 further analyses of the biomechanical model (such as static optimisation [15, 32, 37]) would help to  
474 further elucidate the detailed activation patterns of individual muscles as well as providing estimates  
475 for the likely bone-on-bone joint contact forces being transferred at the large joints of the avian hind limb  
476 during a take-off leap.

477 This study is the first to establish the biomechanical requirements of the hindlimb of a flying bird as it  
478 takes to the air. We present biomechanical conditions that hindlimbs of passerines, a clade of birds  
479 that include over half of all avian species, experience during take-off. The zebra finch take-off leap is  
480 primarily hip and ankle driven, which is in direct contrast to ostrich running mechanics which indicates  
481 avian running is a primarily knee driven activity. The ability of the zebra finch muscles to produce over  
482 double the mechanical requirements at the ankle and knee axes and about 20% more than the  
483 requirements about the hip FE axis along with the suspected use of biarticular muscles and passive  
484 structures is consistent with the hypothesis that the take-off leap as reported here is an optimized  
485 way for the zebra finch to take to the air. Striking similarities in ground reaction forces and relative leg  
486 morphology of multiple passerines suggest that the take-off behaviour described here could be shared  
487 across all passerines, despite differences in mass by two orders of magnitude.

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496

## 497 Tables

498 Table 1 Musculotendon units included in the musculoskeletal model of the zebra finch, grouped by  
 499 which joints they cross; H, K and A denote the hip, knee and ankle joints, respectively. Muscles which  
 500 are categorised under two joints are biarticular. Maximum isometric force was calculated by scaling  
 501 the maximum isometric force of the corresponding muscles of the magpie by mass [10].

Abbreviation	Muscle name	Joints	Maximum isometric force [N]
MCFC	<i>M. caudofemoralis pars caudalis</i>	H	0.154
MFCLA	<i>M. flexor cruris lateralis pars accessoria</i>	H	0.250
MISF1	<i>M. ischiofemoralis 1</i>	H	0.259
MISF2	<i>M. ischiofemoralis 2</i>	H	0.259
MISF3	<i>M. ischiofemoralis 3</i>	H	0.259
MITC1	<i>M. iliotrochantericus caudalis 1</i>	H	0.396
MITC2	<i>M. iliotrochantericus caudalis 2</i>	H	0.396
MITC3	<i>M. iliotrochantericus caudalis 2</i>	H	0.396
MITCR	<i>M. iliotrochantericus cranialis</i>	H	0.003
MITM	<i>M. iliotrochantericus medius</i>	H	0.057
MOL	<i>M. obturatorius lateralis</i>	H	0.112
MOM	<i>M. obturatorius medialis</i>	H	0.431
MPIFL	<i>M. puboischiofemoralis pars lateralis</i>	H	0.388
MPIFM	<i>M. puboischiofemoralis pars medialis</i>	H	0.448
MFCLP	<i>M. flexor cruris lateralis pars pelvica</i>	H, K	0.346
MFCM	<i>M. flexor cruris medialis</i>	H, K	0.238
MIC	<i>M. iliotibialis cranialis</i>	H, K	0.218
MIF	<i>M. iliofibularis</i>	H, K	0.458
MILcaudal1	<i>M. iliotibialis lateralis caudalis 1</i>	H, K	0.254
MILcaudal2	<i>M. iliotibialis lateralis caudalis 2</i>	H, K	0.254
MILcaudal3	<i>M. iliotibialis lateralis caudalis 3</i>	H, K	0.254
MILcranial1	<i>M. iliotibialis lateralis cranialis 1</i>	H, K	0.036
MILcranial2	<i>M. iliotibialis lateralis cranialis 2</i>	H, K	0.036
MILcranial3	<i>M. iliotibialis lateralis cranialis 3</i>	H, K	0.036
MFTI	<i>M. femorotibialis intermedius</i>	K	0.614
MFTL	<i>M. musculus femorotibialis lateralis</i>	K	0.013
MFTM	<i>M. femorotibialis medialis</i>	K	0.490
MFHL	<i>M. flexor hallucis longus</i>	K, A	0.843
MFPD2	<i>M. flexor perforates digiti 2</i>	K, A	0.357
MFPD3	<i>M. flexor perforates digiti 3</i>	K, A	0.362
MFPD4	<i>M. flexor perforates digiti 4</i>	K, A	0.483
MFPD2	<i>M. flexor perforans et perforates digiti 2</i>	K, A	0.236

MGI	<i>M. gastrocnemius pars intermedia</i>	K, A	0.621
MGL	<i>M. gastrocnemius pars lateralis</i>	K, A	1.824
MTCF	<i>M. tibialis cranialis caput femorale</i>	K, A	0.385
MEDL	<i>M. extensor digitorum longus</i>	A	0.031
MFB	<i>M. fibularis brevis</i>	A	0.366
MFDL	<i>M. flexor digitorum longus</i>	A	1.003
MFL	<i>M. fibularis longus</i>	A	0.861
MFPPD3	<i>M. flexor perforans et perforates digiti 3</i>	A	0.894
MGM	<i>M. gastrocnemius pars medialis</i>	A	1.153
MP	<i>M. plantaris</i>	A	0.079
MTCT	<i>M. tibialis cranialis caput tibiale</i>	A	0.516

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502



503 Table 2. Key morphometric parameters of zebra finch (*Taeniopygia guttata*), starling (*Sturnis*  
504 *vulgaris*), crow (*Corvus corone*), and raven (*Corvus corax*).

species	mass [g]	femur length ( $L_{fem}$ ) [cm]	tibiotarsus length ( $L_{tib}$ ) [cm]	tarsometatarsus length ( $L_{tars}$ ) [cm]	digit III length [cm]	hindlimb length [cm]	hindlimb index $((L_{tars} + L_{tib}) / L_{fem})$
zebra finch	15.4	1.40	2.24	1.46	1.10	6.2	2.64
starling [3]	77.3	2.53	4.37	2.82			2.84
crow	440	5.28	8.74	5.77			2.75
raven	1100	6.92	11.42	6.80			2.63

505

506 Table 3 External joint moments and total muscle moment generating capacities of the zebra finch at  
507 the instances at which the peak external joint moments occur. To actively power the take-off leap by  
508 muscle action, the combined(total) moment generating capacity of all muscles must at least reach if  
509 not exceed the level of the external joint moments . Positive moments act in extension, external  
510 rotation, and adduction, while negative moments act in flexion, internal rotation, and abduction.

joint DoF	joint moment [BWL] (requirement)	total muscle moment generating capacity [BWL] (capacity to meet requirement)
ankle extension	0.66±0.04	1.18
knee extension	-0.38±0.05	-0.67
hip extension	0.68±0.05	0.81
hip internal rotation	-0.27±0.07	-0.75
hip abduction	-0.20±0.07	-0.12

511

## 512 **Figure captions**

513 Figure 1 The musculoskeletal model of the left zebra finch hindlimb included 43 musculotendon  
514 units. A) Muscles that either cross the hip or knee joint and biarticular muscles crossing both hip and  
515 knee joints are shown in a caudo-lateral view. B) Anterior and C) posterior view of muscles crossing  
516 the ankle joint and biarticular muscles crossing both knee and ankle joints. For an explanation of the  
517 abbreviations of the muscles used here, please refer to Table 1.

518 Figure 2 Schematic demonstrating the process of morphing the musculature from the magpie to the  
519 zebra finch hindlimb. To allow for a better visual comparison of the bones which differ by almost a  
520 factor of two in size, the bone surfaces depicted here are isotopically scaled by the reciprocal of the  
521 square root of their respective surface area. **A** Establishing an elastic mapping by comparing magpie  
522 and zebra finch bone surfaces. Here, magpie bones were first registered to the corresponding zebra  
523 finch bone using non-rigid iterative closest point (ICP) registration [39]. Following non-rigid  
524 registration, the rigid transformation and isotropic scaling parameters were recovered using  
525 Ordinary Procrustes Analysis (OPA) computed between the vertices of the original magpie bone  
526 surface and the vertices of the magpie bone surface that was elastically registered to the respective  
527 zebra finch bone. This step was encoded in a 4x4 homogeneous transformation matrix. The  
528 remaining difference between the positions of the OPA mapped vertices and their elastically  
529 registered counterparts was captured in a dense deformation field. **B** Application of the elastic  
530 mapping to muscle attachment and via point data. The homogeneous transformation matrix and the  
531 dense deformation field were then applied to all attachment and via points of the magpie muscles  
532 associated to the respective bone surface, resulting in an elastic registration of these structures to  
533 the zebra finch model.

534 Figure 3 Force and moment diagram for the zebra finch hindlimb, including the pelvis, femur,  
535 tibiotarsus and tarsometatarsus in a lateral view. The musculoskeletal model described the hip, knee  
536 and ankle as three, two and two degree of freedom joints respectively. The straight black arrow on  
537 the perch represents the ground reaction force acting through the centre of the perch. Curved  
538 arrows reflect external moments acting at the joints and are colour coded such that the orange  
539 arrows identify the moment about the joint's flexion/extension axes, blue arrows identify moments  
540 about a joints' internal/external rotation axis, and green arrow identifies the moments about the  
541 joints' ab/adduction axis. At the hip, the ground reaction forces consistently result in flexion,  
542 abduction and an internal rotation moments throughout the leap cycles. At the knee and ankle  
543 joints, the ground reaction forces result in extension, and flexion moments, respectively.

544 Figure 4 Lateral view of right zebra finch hindlimb throughout the normalised take-off leap cycle. The  
545 duration of the take-off jump duration ranged from 62 to 67 ms while all of the ankle, knee, and hip  
546 joint undergo substantial extension.

547 Figure 5 External joint moments at the hip, knee and ankle together with the total moment  
548 generating capacity of the muscles, plotted over normalised leap cycle time. Solid lines depict the  
549 mean external joint moments (requirements), bands depict  $\pm 2.5$  standard deviations, while dashed  
550 lines depict the total moment generating capacity of the muscles (capacity). Colours are used to  
551 differentiate the axis around which the moments act, with orange representing flexion/extension,  
552 green ab/adduction, and blue internal/external rotation. Positive moments represent  
553 (internal/external) moments that act to extend, adduct, and internally rotate, respectively.

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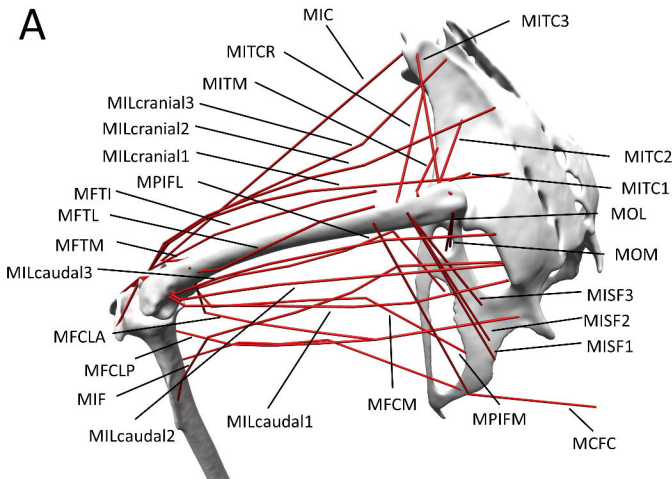
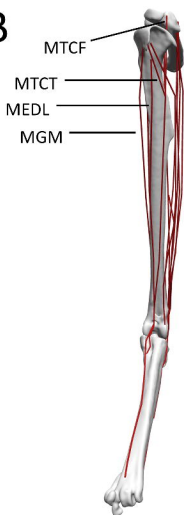
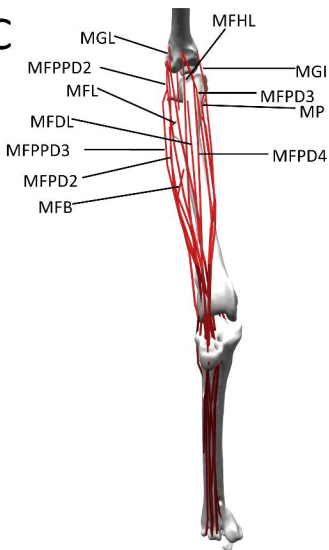
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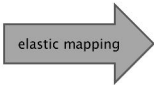
709

**A****B****C**

**A** establish elastic mapping by comparing magpie and zebra finch bone surfaces

magpie bone

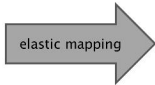
zebra finch bone



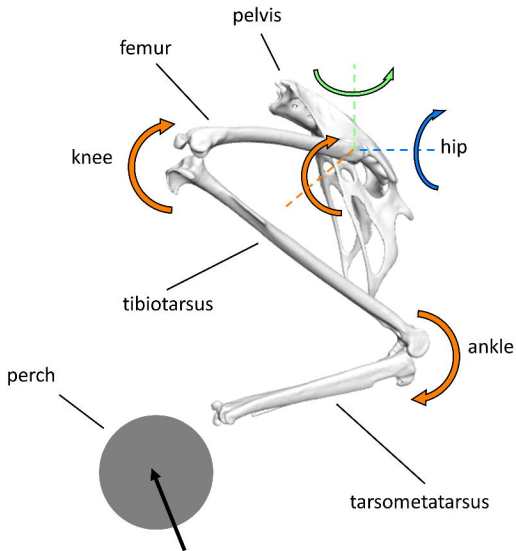
**B** apply elastic mapping to muscle attachment and via point data

magpie bone

zebra finch bone









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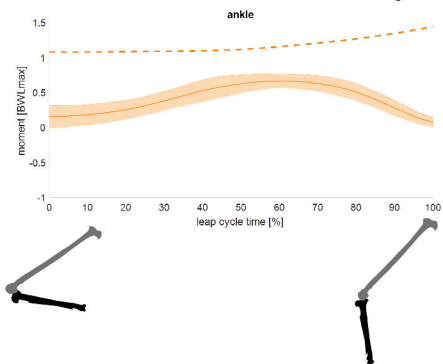
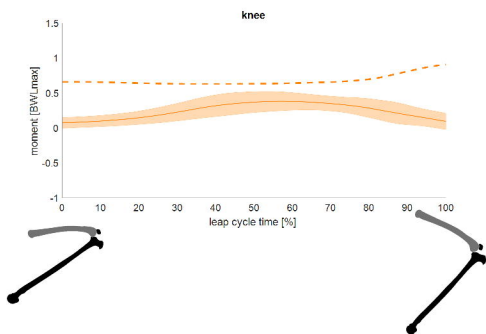
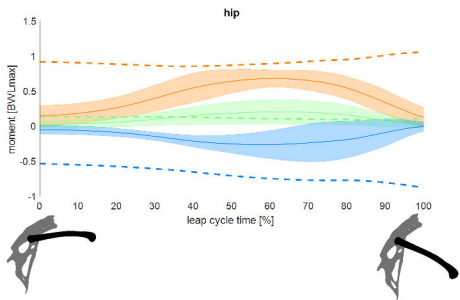


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— FE    - - - muscle FE  
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