Identification of a promiscuous conserved CTL epitope within the SARS-CoV-2 spike protein

Sheng Jiang^{a,b}, Shuting Wu^a, Gan Zhao^c, Yue He^c, Xinrong Guo^d, Zhiyu Zhang^c,
Jiawang Hou^c, Yuan Ding^c, Alex Cheng^c and Bin Wang^{a,b,c,*}

^aKey Laboratory of Medical Molecular Virology (MOE/NHC/CAMS), School of Basic Medical Sciences, Shanghai Medical College(SHMC), Fudan University & ^bNational Clinical Research Center for Aging and Medicine, Huashan Hospital, Fudan University, Shanghai, China.

^cAdvaccine Biopharmaceutics (Suzhou) Co. LTD, Jiangsu Province, China.

^dColby College, Waterville, Maine, USA

*Corresponding Author: Bin Wang, School of Basic Medical Sciences, Fudan University, 131 Dong An Road, 409 Fuxing Building, Shanghai, China 200032. E-mail: bwang3@fudan.edu.cn

1 **ABSTRACT**

2 The COVID-19 disease caused by infection with SARS-CoV-2 and its variants is devastating
3 to the global public health and economy. To date, over a hundred COVID-19 vaccines are known
4 to be under development and the few that have been approved to fight the disease are using the
5 spike protein as the primary target antigen. Although virus neutralizing epitopes are mainly
6 located within the RBD of the spike protein, the presence of T cell epitopes, particularly the CTL
7 epitopes that are likely to be needed for killing infected cells, has received comparatively little
8 attention. In this study, we predicted several potential T cell epitopes with web-based analytic
9 tools, and narrowed them down from several potential MHC-I and MHC-II epitopes by ELISpot
10 and cytolytic assays to a conserved MHC-I epitope. The epitope is highly conserved in current
11 viral variants including the most recent Omicron and compatible with presentation by most HLA
12 alleles worldwide. In conclusion, we identified a CTL epitope suitable for evaluating the CD8+ T
13 cell-mediated cellular response and potentially for addition into future COVID-19 vaccine
14 candidates to maximize CTL responses against SARS-CoV-2.

15 INTRODUCTION

16 Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) was first identified in
 17 Wuhan in early 2020, spread at unprecedented speed, and became a disaster to human beings
 18 worldwide¹. Effective vaccines, antiviral drugs, and treatments have high priorities to defend
 19 against such challenges. SARS-CoV-2 has four main structural proteins: the envelope, membrane,
 20 nucleocapsid, and spike protein that can be considered for inclusion in vaccines. The spike protein
 21 has a receptor-binding domain (RBD) that specifically binds to human angiotensin-converting
 22 enzyme 2 (hACE2) as a receptor and mediates virus entry into the host cell^{2,3}. Neutralizing
 23 antibodies recognizing the RBD can block the spike protein from binding to the hACE2 and
 24 inhibit virus entry^{4,5}. Therefore, spike protein has been the primary choice as the immunogen in
 25 candidate vaccines.

26 Although protection against disease via vaccine-induced neutralizing antibodies has been
 27 demonstrated, the elimination of SARS-CoV-2 infection within the host is also essential. The
 28 numbers of mild and asymptomatic cases are rising dramatically in recent years and such cases
 29 remain infective, prolonging viral dissemination. To eliminate the viral infection, induction of a
 30 potent antigen-specific CD8+ T cell response by vaccination is probably critical. To activate a
 31 viral-specific CD8+ T cell response, the vaccine must contain highly active major
 32 histocompatibility complex class I (MHC-I) epitopes that can be presented by MHC-I molecules
 33 to interact with CD8+ T cell receptors (TCR). The potentiation of viral-specific CD8+ T cell
 34 responses is dependent upon the high affinity and avidity of MHC-I and TCR binding. There is a
 35 lack of information on the CD8+ T cell-recognized epitopes within the spike antigen;
 36 consequently, only overlapping peptide pools that covered the whole region of spike antigen have
 37 been used routinely to evaluate cell-mediated immunity (CMI) of a vaccine candidate^{6,7,8}.
 38 However, a few reports had suggested that CTL epitopes are present within the spike protein⁹, but
 39 no detailed mapping information had been reported or potential sequences discovered. The
 40 identification of those CD8+ T cell epitopes would provide an important tool to evaluate the T cell
 41 immunity in vaccinated individuals or patients and was undertaken here.

This study utilized web-based tools to analyze the potentials for transportation associated with antigen processing (TAP) in the human MHC-I epitopes that were predicted by the Immune Epitope Database analysis (IEDB) resource¹⁰ to be present in peptide pools covering the n-terminal domain (NTD) and receptor-binding domain (RBD) of the spike protein. We demonstrated that peptide 2 (YYVGYLQPRTFLLKY), although it did not give the highest score in the web-based analysis of immunogenicity, was the best epitope for inducing a robust antigen-specific IFN- γ producing CD8+ T response as defined by ELISpot assay. This epitope sequence is also highly conserved among currently discovered SARS-CoV-2 variants.

MATERIALS AND METHODS

Mice

Female BALB/c mice (6-8 weeks of age) were purchased from Beijing Vital Laboratory Animal Technology Co., Ltd. (Beijing, China) and Shanghai Jiesjie Laboratory Animal Co., Ltd. (Shanghai, China), and were kept in SPF conditions. All animal experiments were approved by the Experimental Animals Committee of SHMC, and all methods were carried out in accordance with relevant guidelines and regulations. This study was carried out in compliance with the ARRIVE guidelines. All mice was sacrificed after experiment under euthanasia with Isoflurane overdose.

Peptide pool derived from SARS-CoV-2 Spike protein

The spike receptor-binding domain (RBD) peptide pool (SARS-CoV-2 spike protein 258-518aa) published previously⁷ was used for the study (Table 1).

Immunization

The mice were injected twice, with a two-week interval, via the intramuscular route (i.m.) with 25 μ g pGX9501 expressing a synthetic, optimized sequence of the SARS-CoV-2 full-length spike glycoprotein⁷ and then electroporation was applied with the Celectro2000 device. Serum samples and spleens were collected 14 days after the second immunization.

IEDB analysis for SARS-CoV-2 MHC-I epitope identification

An explorative pannnel of SARS-CoV-2-derived epitopes with strongest predicted

affinity to MHC Class I molecules was defined by Immune Epitope Database analysis (www.IEDB.org). The selection was based on internal predictions using NetMHCpan Version EL4.1. all predicted epitopes with a percentile rank of <2 was selected for MHC-I processing analysis using MHC-NP methods. Simultaneously, those epitopes were analyzed in MHC-I immunogenicity for checking whether the peptide sequence are consistent to the site preference of this allele. Combining results of the above three analysis methods, the peptide with a percentile rank of <0.5, a TAP total score of > -1 and an immunogenicity score of >0 was picked up for the next ELISpot assay of T cell IFN- γ response.

Cytotoxic lymphocyte (CTL) killing ability

A single-cell suspension of splenocytes from naïve syngeneic mice was diluted to 1.5×10^8 /ml in RPMI1640 containing 10% FBS and 2% penicillin and streptomycin and pulsed at 37°C with or without 5 μ g/ml peptides as described previously²². After 4 h, eFlour450 (eBioscience, 65-0842-85) at 5 mM (high concentration) was used to label peptide-pulsed cells at room temperature in the dark. The non-peptide-pulsed cells were labeled with a low concentration of eFlour450 at 0.5 mM. After being rinsed three times with PBS, 4×10^6 labeled and peptide-pulsed cells and an equal number of labeled non-peptide-pulsed cells were adoptively transferred by tail vein injections into mice that had previously been immunized. Six hours later, the percentage of labeled cells in spleens was detected with LSRI Fortessa flow cytometry (BD) and analyzed by FlowJo (TreeStar). The following formula calculated the specific cell lysis: Specific cell lysis ability = $(1 - (\text{percentage of cells incubated with peptide} / \text{percentage of cells incubated without peptide})) \times 100\%$.

IFN- γ ELISpot

Spleens were collected from mice individually into RPMI1640 media supplemented with 10% FBS (Gibco) and penicillin/streptomycin and processed into single-cell suspensions. ELISpot assays were performed using Mouse IFN- γ ELISpot plates (Dakewe Biotech Co., Ltd, 2210006).

93 The ELISpot plates were repeat washed 5 times at RT with 100 μ L of PBS per well and incubated
 94 with 200 μ L of RPMI1640 10% FBS (R10) for 10 min before cell plating. Two hundred fifty
 95 thousand mouse splenocytes, CD4+, or CD8+ T cells were plated into each well and stimulated
 96 for 16 h with 15-mer peptides from the SARS CoV-2 Spike peptide pools that overlapped by nine
 97 amino acids as previously described⁷. Each peptide was present at a final concentration of 1 μ g in
 98 100 μ l R10 per well. The spots were developed based on the manufacturer's instructions. R10 and
 99 cell stimulation cocktails (Invitrogen) were used for negative and positive controls, respectively.
 100 Spots were scanned and quantified by AID ELISpot READER (AID, Germany). Spot-forming
 101 units (SFU) per million cells were calculated after subtracting the negative control wells.

102 **Statistical Analysis**

103 The statistical analysis methods and sample sizes (n) are specified in the results section
 104 or figure legends for all quantitative data. All values are reported as means $\pm$ sem with the
 105 indicated sample size. No samples are excluded from the analysis. All relevant statistical tests
 106 are two-sided. P values less than 0.05 were considered statistically significant. All animal
 107 studies were performed with randomized animal selection. Statistics were performed using
 108 GraphPad Prism 7 software. In all data, *p<0.05, **p<0.01, ***p<0.001, and ****p<0.0001.

109

110 RESULTS

111 Strong CD8⁺ CTL epitope activity is embedded in an overlapping peptide pool 1 that covers 112 the NTD and RBD region of the spike protein

113 When Balb/c mice were immunized twice with the pGX9501 DNA vaccine expressing the
114 spike protein of SARS-CoV-2, a higher level of IFN- γ expression by splenocytes was often seen
115 by the ELISpot assay when the cells were stimulated in vitro with spike peptide pool 1 (Table 1)
116 compared with pool 2 (Figure 1A). In addition, when an in vivo CTL assay was done with
117 identically immunized animals, the same peptide pool 1 gave a strong CTL response in vivo
118 (Figure 1B), suggesting that MHC-I epitope(s) were present within pool 1.

119 Screening and identification of an MHC-I epitope in peptide pool 1.

120 We placed the entire 41 peptide sequences from peptide pool 1 into the Immune Epitope
121 Database analysis (IEDB) (<http://www.iedb.org/>) to seek critical epitopes. An evaluation method
122 was established by integrating MHC-I binding prediction, MHC-I immunogenicity, and MHC
123 natural processing (MHC-NP) prediction from three H-2d MHC-I alleles to improve the accuracy
124 of prediction results (Table 2). In the H-2D^d allele, peptide 2 showed good MHC-I binding ability,
125 immunogenicity, and TAP ability. In the H-2K^d allele, peptide 12 showed the strongest
126 immunogenicity, and peptide 2 presented the most potent MHC-I binding ability and TAP ability
127 among all peptides. In the H-2L^d allele, both peptide 2 and peptide 11 showed the strongest
128 immunogenicity, while peptide 12 emerged as having the most potent TAP ability (Figure
129 2A&2B). Consequently, we used peptide 2, peptide 11, and peptide 12 to conduct an ELISpot
130 assay of T cell IFN- γ response.

131 As shown in Figure 3A, peptide 2 from pool 1 presented the best stimulation to induce the
132 IFN- γ secretion compared to the other two selected peptides. Thus, peptide 2, consisting of 15
133 amino acids, stimulated CD8⁺ T cells via MHC-I or/and CD4⁺ T cells via MHC-II. To identify
134 which T cell type was stimulated by peptide 2, purified CD4⁺ T cells or CD8⁺ T cells were used

(sFigure 1). Peptide 2 stimulated CD8⁺ T cells but not CD4⁺ cells, indicating that it can only be presented by MHC-I (Figure 3B). To further investigate its sequence specificity, we mutated several predicted anchor amino acids of peptide 2 according to the preferences of the H-2d MHC-I allele¹¹⁻¹⁴. The mutated peptide 2 had a low MHC-I binding score in the IEDB prediction (Table 3) and showed a significantly reduced ability to stimulate IFN- γ secretion by CD8⁺ T cells (Figure 3C).

141 **Analysis of peptide 2 epitope conservation and HLA distribution**

142 We used the peptide 2 sequence, YYVGYLQPRTFLLKY (amino acid 264-278), to compare
143 with current posted SARS-CoV-2 variants including variant-of-interest (VOI) and variant-of-
144 concern (VOC) published by WHO including the latest Δ variant and observed that this sequence
145 is highly conserved (Figure 4A), and located at the end of the NTD of Spike protein and upstream
146 of the RBD (Figure 4B). The highly conserved epitopic sequence provides a useful tool for
147 evaluating the CD8⁺ T cell-mediated responses to vaccination in animals and humans.

148 Since MHC-I-biased expression patterns in different populations globally are diversified and
149 variable, the peptide 2 sequence that can be recognized by one population may not be recognized
150 by others. To investigate if peptide 2 sequence could be recognized by most populations
151 worldwide, we performed an HLA allele analysis for different regions to assess binding to one or
152 more of the 27 prevalent MHC-I molecules, including HLA-A (01:01, 02:01, 02:03, 02:06, 03:01,
153 11:01, 23:01, 24:02, 26:01, 30:01, 30:02, 31:01, 32:01, 33:01, 68:01, 68:02), and HLA-B (07:02,
154 08:01, 15:01, 35:01, 40:01, 44:02, 44:03, 51:01, 53:01, 57:01, 58:01), as shown in Figure 4C &
155 Table 4. The frequency by HLAs was calculated by the online analysis tool at
156 <http://www.allelefrequencies.net/>. We also evaluated the MHC-I binding ability, immunogenicity,
157 and TAP potential of peptide 2 on different HLA alleles by IEDB (Figure 4D & Table 5). The
158 results indicated that peptide 2 could be recognizable by the HLA-A*02:01 allele (most in Europe
159 and America), HLA-B*08:01 allele (in Europe and Australia), HLA-A*23:01 allele (in North
160 Africa and Sub-Saharan Africa), HLA-A*02:03 allele (in Southeast Asia), HLA-A*24:02 allele

161 (in Oceania), HLA-A*02:06 allele (in North America, North-East Asia, and Oceania), HLA-
162 A*33:01 allele (in China and Pakistan), HLA-B*35:01 allele (in Oceania), and HLA-A*03:01
163 allele (in Europe). These findings suggest that peptide 2 could be well recognized by the most
164 frequent HLA alleles of the worldwide population and can therefore be considered to be
165 promiscuous.

166 **DISCUSSION**

167 In this study, we have defined and characterized a potential CTL epitope conserved and
168 established a BALB/c mice model to evaluate CTL responses to the spike protein of SARS-CoV-2.
169 Furthermore, we found the peptide 2 may be well recognized by HLA alleles in most populations
170 worldwide.

171 In recent studies, CD8⁺ T cell immunity was found to make major contributions to the
172 protective efficacy of SARS-CoV-2 vaccines¹⁵⁻¹⁷. Additionally, lymphopenia was more
173 accentuated in symptomatic COVID-19 patients with pneumonia than in those without pneumonia,
174 consistent with T cell immunity playing a protective role in pre-existing immunity against SARS-
175 CoV-2¹⁷⁻¹⁹. However, the role of T cell immunity in the pathology of COVID-19 has not been
176 clarified²⁰ and awaits further definition of T cell epitopes and their functions. Engineered
177 improvements in the peptide epitopes recognized by CTL have been an essential feature of the
178 advancements made in evaluating the cellular immunity induced by vaccines. Our work has
179 provided a tool to monitor virus-specific CD8⁺ T cells and assess the contribution of CTLs to the
180 control and the elimination of the virus.

181 The SARS-CoV-2 virus was found to mutate rapidly, the development of vaccines that could
182 protect people from different variants was seen to be urgent. The neutralizing antibodies induced
183 by vaccines were found to have variable efficacies against the different SARS-CoV-2 variants and
184 efficacies declined over time, whereas the protection represented by CD8⁺ T cell immunity
185 remained unchanged¹⁵. We showed here that one particular peptide from the spike protein, peptide
186 2 is highly conserved among variants and could be well presented by MHC-1 of all of the HLA
187 alleles that are most frequent worldwide. Thus, the conserved peptide 2 was found suitable for

188 developing peptide vaccines to induce cellular immunity against different variants of SARS-CoV-
189 2, including variant-of-interest (VOI) and variant-of-concern (VOC). A recent study confirmed
190 that peptide 2 probably has a strong cell-mediated immunological function in man; a 9-mer
191 (YLQPRTFLL) peptide that was overlapped by peptide 2 was able to induce a high level of IFN- γ
192 expression from PBMCs of patients who had recovered from COVID-19 and carried the HLA-
193 A*02:01 allele²¹.

194 In conclusion, our study utilized web-based tools to predict human MHC-I epitopes although
195 peptide 2 (YYVGYLQPRTFLLKY) did not give the strongest total TAP score in the prediction,
196 the simulations indicated particularly robust antigen-specific IFN- γ -expressing CD8+ T responses
197 overall compared to the other predicted epitope sequences. This epitope sequence is highly
198 conservative among currently known SARS-CoV-2 variants and recognizable by most world
199 populations. This critical MHC-I epitope sequence is located at the end of NTD of the spike
200 protein; it can be used to assess CMI induced by a COVID-19 vaccination; it may be strategically
201 incorporated into vaccine designs to enhance the prospect of viral elimination by vaccination.

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207 **COMPETING INTERESTS:** Authors declare that they have no competing interests.

208 **AUTHOR CONTRIBUTIONS:** BW and SJ conceived, designed the study and drafted
209 manuscript. BW supervised the study. SJ, SW, XG, JH, YD and ZZ performed the
210 experiments. SJ, YH, GZ and AC analyzed the data. All authors reviewed the manuscript.

Tables

Table 1. Overlapping Peptide Pool 1.

Peptide number	Sequence	Start	End	aa
1	TAGAAAYYVGYLQPR	258	272	15
2	YYVGYLQPRTFLLKY	264	278	15
3	QPRTFLLKYNENGTI	270	284	15
4	LKYNENGTITDAVDC	276	290	15
5	GTITDAVDCALDPLS	282	296	15
6	VDCALDPLSETKCTL	288	302	15
7	PLSETKCTLKSFTVE	294	308	15
8	CTLKSFTVEKGIYQT	300	314	15
9	TVEKGIYQTSNFRVQ	306	320	15
10	YQTSNFRVQPTESIV	312	326	15
11	RVQPTESIVRFPNIT	318	332	15
12	SIVRFPNITNLCPPG	324	338	15
13	NITNLCPPGGEVFNAT	330	344	15
14	PFGEVFNATRFASVY	336	350	15
15	NATRFASVYAWNRKR	342	356	15
16	SVYAWNRKRISNCVA	348	362	15
17	RKRISNCVADYSVLY	354	368	15
18	CVADYSVLYNSASFS	360	374	15
19	VLYNSASFSTFKCYG	366	380	15
20	SFSTFKCYGVSP TKL	372	386	15
21	CYGVSP TKLNDLCFT	378	392	15
22	TKLNDLCFTNVYADS	384	398	15
23	CFTNVYADSFVIRGD	390	404	15
24	ADSFVIRGDEV RQIA	396	410	15
25	RGDEV RQIAPGQTGK	402	416	15
26	QIAPGQTGKIADYNY	408	422	15
27	TGKIADYNYKLPDDF	414	428	15
28	YNYKLPDDFTGCVIA	420	434	15
29	DDFTGCVIAWNSNNL	426	440	15
30	VIAWNSNNLDSKVGG	432	446	15
31	NNLDSKVGGNYNYLY	438	452	15
32	VGGNYNYLYRLFRKS	444	458	15
33	YLYRLFRKSNLKPFE	450	464	15
34	RKSNLKPFE RDI STE	456	470	15
35	PFE RDI STEIYQAGS	462	476	15

36	STEIQAGSTPCNGV	468	482	15
37	AGSTPCNGVEGFNCY	474	488	15
38	NGVEGFNCYFPLQSY	480	494	15
39	NCYFPLQSYGFQPTN	486	500	15
40	QSYGFQPTNGVGYP	492	506	15
41	PTNGVGYPYRVVVL	498	512	15

Notes:

1. The peptides covered the entire sequence of 258-512 amino acids in the spike protein;
2. The peptides were synthesized with an average 15 amino acid length and with nine amino acids overlapping each other.

Table 2. MHC-I epitope analysis for Overlapping Peptide Pool 1.

allele	peptide number	MHC-I binding	immunogenicity	Proteasome Score	TAP Score	MHC Score	Processing Score	TAP Total Score
H-2-Dd	21	0.01	0.03612	1.44	1.12	-3.49	2.57	-0.92
H-2-Dd	41	0.04	0.13706	1.74	0.4	-2.56	2.14	-0.42
H-2-Dd	2	0.05	0.1573	1.36	1.07	-3.26	2.43	-0.83
H-2-Dd	26	0.13	-0.02676	1.38	1.27	-4.51	2.65	-1.86
H-2-Dd	9	0.19	-0.11058	1.27	0.99	-3.48	2.26	-1.22
H-2-Dd	19	0.25	0.03263	1.33	1.04	-3.45	2.37	-1.08
H-2-Dd	12	0.29	0.1431	1.05	1.29	-3.9	2.35	-1.55
H-2-Dd	11	0.49	0.1386	1.36	1.21	-4.02	2.57	-1.45
H-2-Dd	32	0.62	0.0966	1.19	1.17	-3.83	2.36	-1.46
H-2-Dd	37	0.65	0.12191	1.38	1.09	-3.76	2.47	-1.29
H-2-Dd	14	0.67	0.08562	1.39	0.82	-3.76	2.21	-1.55
H-2-Dd	31	0.7	0.023	1.31	1.15	-4.17	2.46	-1.71
H-2-Dd	20	0.7	-0.31841	1.47	1.3	-4.6	2.77	-1.83
H-2-Dd	27	0.89	-0.11289	1.4	1.21	-4.54	2.61	-1.92
H-2-Dd	28	1.1	-0.19576	0.98	1.12	-4.22	2.1	-2.12
H-2-Dd	24	1.2	0.12947	1.27	0.71	-4.63	1.98	-2.64
H-2-Dd	25	1.2	-0.11559	1.43	0.15	-4.39	1.58	-2.81
H-2-Dd	18	1.2	-0.22309	1.33	1.17	-4.05	2.5	-1.55
H-2-Dd	40	1.3	0.1256	1.38	1.34	-4.5	2.71	-1.79
H-2-Dd	1	1.3	0.06158	1.24	1.23	-4.32	2.47	-1.85
H-2-Dd	33	1.5	-0.21085	1.03	1.24	-4.23	2.27	-1.96
H-2-Dd	29	1.6	0.05792	1.52	0.5	-4.17	2.02	-2.15
H-2-Dd	30	1.6	0.05792	1.35	0.46	-4.17	1.8	-2.36
H-2-Dd	39	1.6	-0.15021	1.25	1.15	-3.84	2.41	-1.44
H-2-Dd	10	2	0.01977	1.1	0.24	-4.23	1.34	-2.89
H-2-Kd	2	0.01	0.06572	1.24	0.48	-1.78	1.72	-0.06
H-2-Kd	41	0.05	0.04196	1.74	0.46	-3.43	2.2	-1.22

H-2-Kd	9	0.06	0.12441	1	0.23	-2.55	1.23	-1.32
H-2-Kd	4	0.07	0.28634	1.34	0.37	-2.34	1.71	-0.63
H-2-Kd	3	0.07	0.05892	1.16	0.37	-2.34	1.54	-0.8
H-2-Kd	20	0.08	0.25644	1.45	0.49	-2.65	1.93	-0.71
H-2-Kd	21	0.08	0.05832	1.75	0.36	-2.65	2.12	-0.53
H-2-Kd	16	0.11	0.16858	1.36	0.4	-2.32	1.77	-0.55
H-2-Kd	40	0.19	0.0905	1.38	1.32	-3.9	2.69	-1.21
H-2-Kd	23	0.26	0.0573	1.31	0.44	-3.18	1.75	-1.43
H-2-Kd	24	0.61	-0.0378	1.07	0.2	-3.02	1.27	-1.75
H-2-Kd	19	0.68	0.0279	1.33	1.2	-3.27	2.52	-0.75
H-2-Kd	10	0.79	0.034	1.1	0.23	-3.35	1.33	-2.02
H-2-Kd	12	0.92	0.34063	1.45	0.59	-3.13	2.03	-1.1
H-2-Kd	1	0.96	-0.04018	1.24	1.29	-4.51	2.53	-1.98
H-2-Kd	32	0.99	0.13255	1.19	1.18	-3.56	2.37	-1.19
H-2-Kd	39	1.1	0.0801	1.25	1.31	-3.16	2.56	-0.6
H-2-Kd	31	1.2	0.1811	1.48	0.48	-3.32	1.96	-1.36
H-2-Kd	29	2	0.07062	1.52	0.5	-3.5	2.02	-1.48
H-2-Kd	30	2	0.07062	1.35	0.46	-3.5	1.8	-1.7
H-2-Kd	35	2	-0.15381	1.42	1.16	-4.52	2.57	-1.95
H-2-Ld	11	0.06	0.30371	1.36	0.98	-3.5	2.34	-1.16
H-2-Ld	41	0.12	-0.07228	1.74	0.35	-3.54	2.09	-1.45
H-2-Ld	21	0.2	0.05832	1.44	0.99	-3.65	2.44	-1.21
H-2-Ld	39	0.27	-0.19696	1.25	0.91	-3	2.16	-0.84
H-2-Ld	12	0.28	0.09851	1.05	0.94	-2.75	2	-0.75
H-2-Ld	37	0.44	0.0801	1.38	0.99	-4.01	2.37	-1.65
H-2-Ld	2	0.54	0.28634	1.41	1.19	-3.97	2.6	-1.38
H-2-Ld	3	0.57	-0.08994	1.52	1.13	-3.97	2.65	-1.32
H-2-Ld	27	0.77	0.0573	1.4	1.21	-4.39	2.61	-1.78
H-2-Ld	28	0.77	0.03448	0.98	1.12	-4.18	2.1	-2.07
H-2-Ld	19	0.95	0.11915	1.53	1.42	-4.32	2.95	-1.37
H-2-Ld	18	1.1	-0.22669	1.33	1.17	-3.82	2.5	-1.32
H-2-Ld	34	1.9	0.1811	0.91	1.16	-4.58	2.07	-2.5
H-2-Ld	9	1.9	-0.08994	1.27	0.99	-4.13	2.26	-1.87
H-2-Ld	38	2	-0.19696	0.99	0.28	-2.8	1.27	-1.53

Notes:

1. MHC-I binding score was between 0 and 2. <0.5 strong binder, 0.5-2 weak binder, >2 non-binder.
2. A high Immunogenicity score indicates that the degree of the peptide conformity to sequence preference was good.
3. The higher the TAP total score, the higher the likelihood that the peptide will be presented after being swallowed by DCs.

Table 3. Prediction scores of peptide 2 and mutated peptide 2

	Sequence	Score	MHC-I binding
Peptide 2	YYVGYLQPRTFLLKY	0.91863	0.01
Peptide 2 mut	Y <u>E</u> V <u>G</u> E <u>L</u> Q <u>D</u> R <u>T</u> F <u>E</u> LKY	0.025669	>2

Notes:

1. MHC-I binding score was between 0 and 2, where, the <0.5 represents a strong binder, 0.5-2 weak binder, >2 non-binder.
2. Underlined amino acids were the mutated residues in the Peptide 2 mut.

Table 4. Geographic Distribution of HLA allele

Continent	Allele	Frequency	Allele	Frequency
Australia	HLA-A*24:02	0.2	HLA-B*07:02	0.08
	HLA-A*02:01	0.11	HLA-B*40:01	0.07
	HLA-A*11:01	0.08		
Europe	HLA-A*02:01	0.26	HLA-B*07:02	0.08
	HLA-A*01:01	0.12	HLA-B*08:01	0.07
	HLA-A*03:01	0.12	HLA-B*51:01	0.07
	HLA-A*24:02	0.1		
North Africa	HLA-A*02:01	0.12	HLA-B*51:01	0.07
			HLA-B*08:01	0.05
			HLA-B*35:01	0.05
North America	HLA-A*02:01	0.2	HLA-B*35:01	0.08
	HLA-A*24:02	0.12	HLA-B*07:02	0.07
			HLA-B*08:01	0.05
			HLA-B*15:01	0.05
			HLA-B*44:03	0.05
			HLA-B*51:01	0.05
North-East Asia	HLA-A*24:02	0.22	HLA-B*51:01	0.08
	HLA-A*02:01	0.14	HLA-B*35:01	0.07
			HLA-B*15:01	0.07
			HLA-B*44:03	0.06
			HLA-B*07:02	0.05
Oceania	HLA-A*24:02	0.3	HLA-B*35:01	0.15
	HLA-A*11:01	0.15		
South and Central America	HLA-A*02:01	0.2	HLA-B*40:01	0.25
	HLA-A*24:02	0.14		
South Asia	HLA-A*11:01	0.13		
	HLA-A*01:01	0.1		
	HLA-A*24:02	0.1		

South-East Asia	HLA-A*11:01	0.2	HLA-B*40:01	0.1
	HLA-A*24:02	0.15	HLA-B*58:01	0.06
Sub-Saharan Africa	HLA-A*23:01	0.11	HLA-B*07:02	0.06
	HLA-A*02:01	0.1	HLA-B*08:01	0.05
Western Asia	HLA-A*02:01	0.15		
	HLA-A*01:01	0.1		

Notes

1. Frequency means the proportion of HLA allele present in the survey population. This table only shows the alleles for which frequency was higher than 0.05 in the continent.

Table 5. Human MHC-I epitope analysis for peptide 2

Allele	Peptide number	MHC-I binding	Immunogenicity	Proteasome Score	TAP Score	MHC Score	Processing Score	TAP Total Score
HLA-A*02:01	2	0.02	0.18048	1.39	0.39	-0.66	1.78	1.12
HLA-B*08:01	2	0.02	0.14268	1.39	0.39	-1.31	1.78	0.47
HLA-A*23:01	2	0.03	0.19092	1.36	1.13	-1.38	2.5	1.12
HLA-A*02:03	2	0.03	0.18064	1.39	0.39	-0.91	1.78	0.87
HLA-A*24:02	2	0.04	0.19092	1.36	1.13	-1.67	2.5	0.83
HLA-A*02:06	2	0.04	0.18064	1.39	0.39	-0.96	1.78	0.82
HLA-A*33:01	2	0.09	0.18048	0.76	0.68	-1.13	1.43	0.3
HLA-B*35:01	2	0.1	0.18064	1.41	1.19	-2.35	2.6	0.25
HLA-A*03:01	2	0.12	0.18064	0.89	0.18	-1.35	1.06	-0.29
HLA-A*32:01	2	0.21	0.18064	1.39	0.39	-2.21	1.78	-0.43
HLA-B*53:01	2	0.21	0.18064	1.41	1.19	-3.4	2.6	-0.8
HLA-A*01:01	2	0.23	0.18424	1.41	1.24	-2.91	2.65	-0.26
HLA-B*15:01	2	0.29	0.18064	1.41	1.28	-1.79	2.69	0.89
HLA-A*30:02	2	0.35	0.19092	1.41	1.28	-2.29	2.69	0.4
HLA-A*31:01	2	0.45	0.18064	0.76	0.68	-1.9	1.43	-0.47
HLA-A*26:01	2	0.52	0.18048	1.41	1.28	-4.14	2.69	-1.45
HLA-A*11:01	2	0.53	0.18064	0.89	0.18	-1.63	1.06	-0.57
HLA-B*58:01	2	0.55	0.18064	1.36	1.06	-3.33	2.42	-0.9
HLA-B*57:01	2	0.56	0.18064	1.36	1.07	-3.39	2.43	-0.95
HLA-B*51:01	2	0.65	0.18064	1.41	1.28	-4.36	2.69	-1.67
HLA-B*07:02	2	0.72	0.18064	1.41	1.19	-3.75	2.6	-1.15
HLA-A*30:01	2	0.73	0.18424	1.41	1.28	-3.73	2.69	-1.05
HLA-A*68:02	2	1.4	0.18064	1.39	0.39	-3.11	1.78	-1.33
HLA-B*44:03	2	1.6	0.18424	1.41	1.28	-4.01	2.69	-1.33
HLA-B*44:02	2	1.9	0.18424	1.41	1.28	-4.16	2.69	-1.47
HLA-A*68:01	2	2.4	0.18064	0.76	0.68	-2.33	1.43	-0.9
HLA-B*40:01	2	3.1	0.18048	1.41	1.28	-4.44	2.69	-1.75

Notes

4. MHC-I binding score was between 0 and 2. <0.5 strong binder, 0.5-2 weak binder, >2 non-binder.
5. The high Immunogenicity score means the degree of the peptide conformity to sequence preference was good.
6. The higher the TAP total score, the higher the likelihood that the peptide will be presented after being swallowed by DCs.

Figure Legends

Figure 1. Peptide pool 1 Induced strong T cell responses in BALB/c mice post-administration of pGX9501

Balb/c mice (n = 5/group) were immunized twice two weeks apart with 25 µg pGX9501 or pVAX1 (empty vector). T cell responses were analyzed on day 14 after the second injection. (A) Splenocytes were harvested and T cell responses were measured by IFN-γ ELISpot after stimulation for 20 h with overlapping peptide pools 1 or 2. (B) Antigen-specific cytotoxic lymphocyte (CTL) killing activity was evaluated by an in vivo CTL assay. Target cells at 4×10^6 /ml from naïve mice were peptide-pulsed with pool 1 then labeled with a high concentration of eFluor450 in vitro. Control cells were non-peptide-pulsed cells and labeled with a low concentration of eFluor450. The cells were mixed and transferred i.v. into immunized mice. After 5 h, splenocytes were harvested and the intensity of eFluor450 peptide labeled target cells was compared with the non-peptide-labeled negative control cells by flow cytometry.

Figure 2. IEDB prediction scores of peptides in pool 1

CTL epitope peptides were screened by integrating MHC-I binding prediction, MHC-I immunogenicity (A), and MHC-NP (B) prediction from three H-2d MHC-I alleles.

Figure 3. Peptide 2 is identified as the CD8+ CTL epitope

BALB/c mice were immunized with the pGX9501. Splenocytes were obtained and specific T-cell induction was analyzed with IFN-γ ELISpot, using stimulation with the indicated peptides.

(A) Splenocytes, (B) CD4+, and (C) CD8+ T cells. .

Figure 4. Conserved Sequence & MHC-I HLA Analysis of peptide 2

(A) The sequence of peptide 2 was highly conserved in the 11 virus variants that have been identified as the variants of interest and the variants of concern, as published by WHO.

(B) The position of the peptide 2 (yellow marked) in the stereoscopic structure of the spike protein. (C) The global distribution of HLA alleles. (D) Analysis of peptide 2 by integration of MHC-I binding prediction, MHC-I immunogenicity, and MHC-NP prediction from the HLA alleles.

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Figure 1

Figure 2

A

B

Figure 3

A

B

C

A

B