Supporting information

Discovery of S-217622, a Non-Covalent Oral SARS-CoV-2 3CL Protease Inhibitor Clinical Candidate for Treating COVID-19

Yuto Unoh^{1†}, Shota Uehara^{1†}, Kenji Nakahara^{1†}, Haruaki Nobori¹, Yukiko Yamatsu¹, Shiho Yamamoto¹, Yuki Maruyama^{1,2}, Yoshiyuki Taoda¹, Koji Kasamatsu¹, Takahiro Suto¹, Kensuke Kouki¹, Atsufumi Nakahashi¹, Sho Kawashima¹, Takao Sanaki¹, Shinsuke Toba^{1,2}, Kentaro Uemura^{1,2}, Tohru Mizutare¹, Shigeru Ando¹, Michihito Sasaki², Yasuko Orba², Hirofumi Sawa², Akihiko Sato^{1,2}, Takafumi Sato¹, Teruhisa Kato¹, Yuki Tachibana^{1*}

¹Shionogi Pharmaceutical Research Center, 3-1-1 Futaba-cho, Toyonaka, Osaka, Japan

²International Institute for Zoonosis Control, Hokkaido University, Sapporo, Japan

*Corresponding author. Email: yuuki.tachibana@shionogi.co.jp

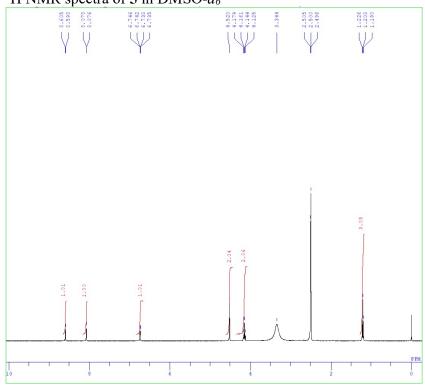
†These authors contributed equally to this work.

Contents

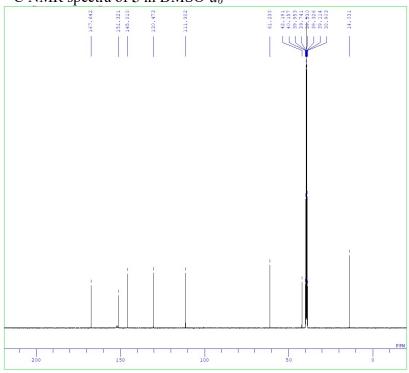
•	¹ H-NMR and ¹³ C-NMR spectra for synthesized compounds	1
•	HPLC traces of 1-3 (S-217622)	
•	Experimental Procedures for <i>in vitro</i> safety	17
•	Table S1. The calculated IC ₅₀ values for S-217622 to SARS-CoV-2 3CL protease activity in eac	h
	experiment	20
•	Table S2. EC ₅₀ values of S-217622 and remdesivir on cytopathic effects in SARS-CoV-2 3CL-	
	infected VeroE6/TMPRSS2 cells	20
•	Table S3. CC ₅₀ values of S-217622 and remdesivir on cytopathic effects in VeroE6/TMPRSS2	
	cells	21
•	Table S4. <i>In vitro</i> safety profiles of S-217622	21
•	Table S5. Diffraction data and refinement statistics for SARS-CoV-2 3CL ^{pro} complexed with	
	compound 1 and compound 3 (S-217622)	22

¹H-NMR and ¹³C-NMR spectra for synthesized compounds

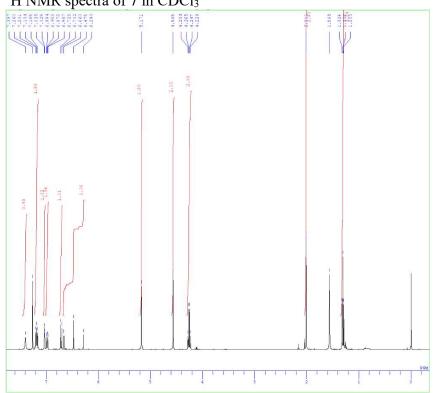
¹H NMR spectra of **5** in DMSO-*d*₆



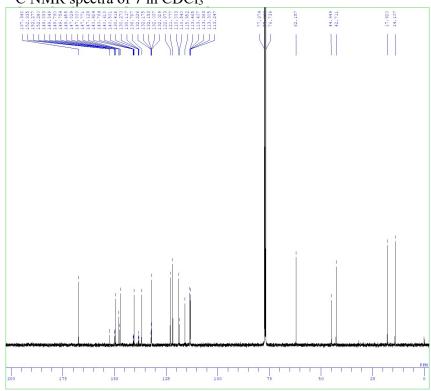
 13 C NMR spectra of **5** in DMSO- d_6



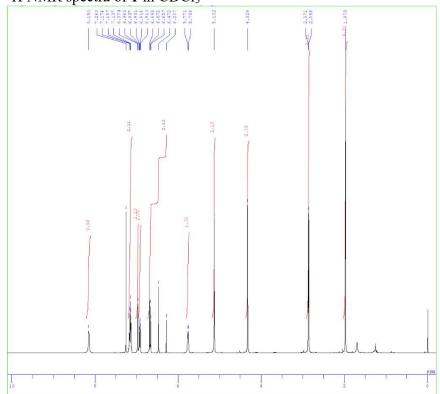
¹H NMR spectra of 7 in CDCl₃

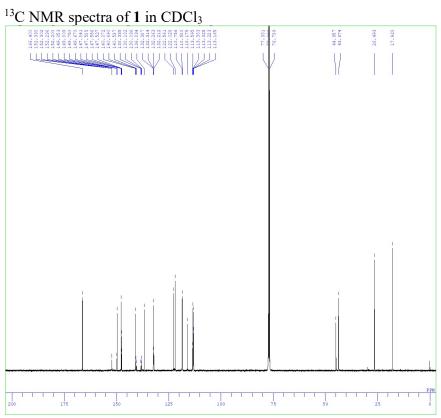


¹³C NMR spectra of 7 in CDCl₃

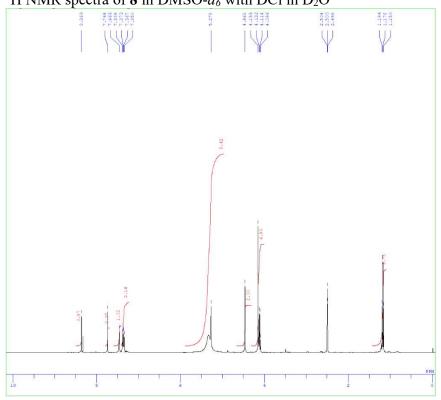


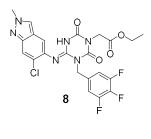
¹H NMR spectra of 1 in CDCl₃



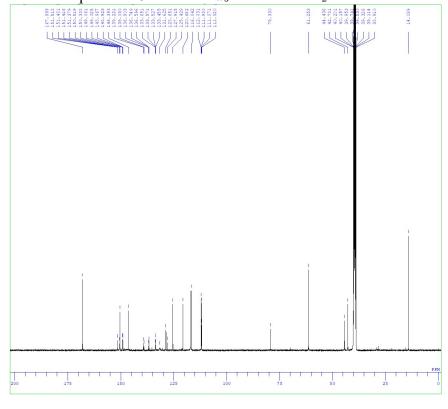


 1 H NMR spectra of **8** in DMSO- d_{6} with DCl in D $_{2}$ O

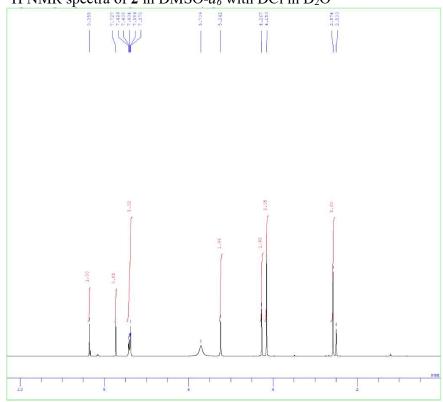




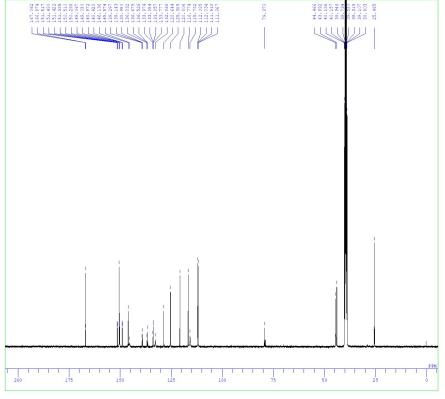
 13 C NMR spectra of **8** in DMSO- d_6 with DCl in D₂O



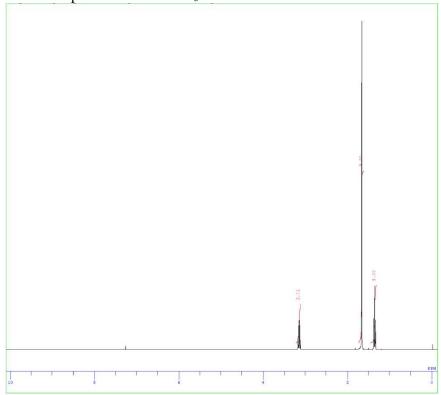
¹H NMR spectra of **2** in DMSO-*d*₆ with DCl in D₂O



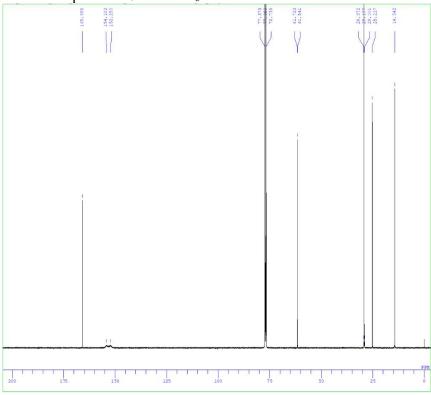
 13 C NMR spectra of **2** in DMSO- d_6 with DCl in D₂O



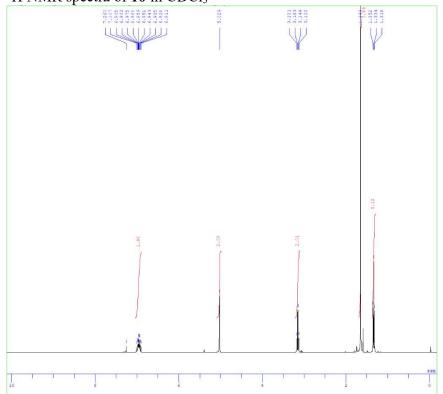


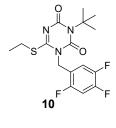


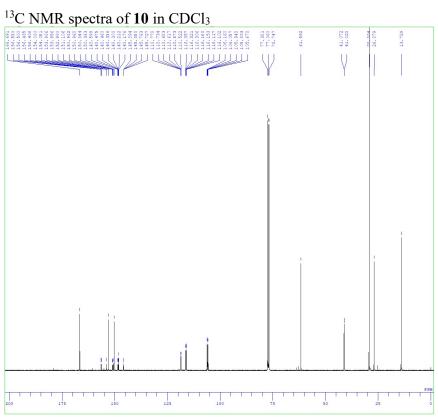
¹³C NMR spectra of **9** in CDCl₃



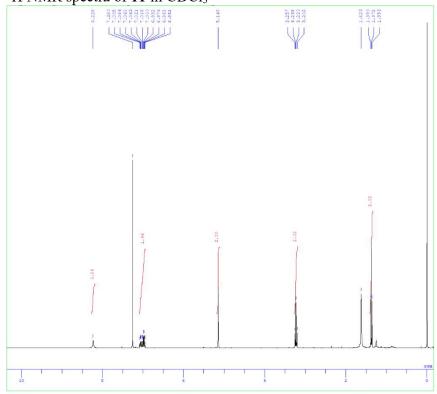
¹H NMR spectra of **10** in CDCl₃



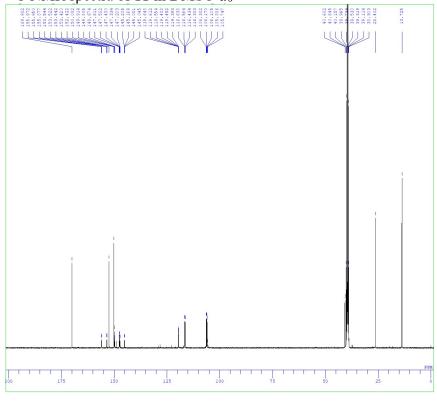




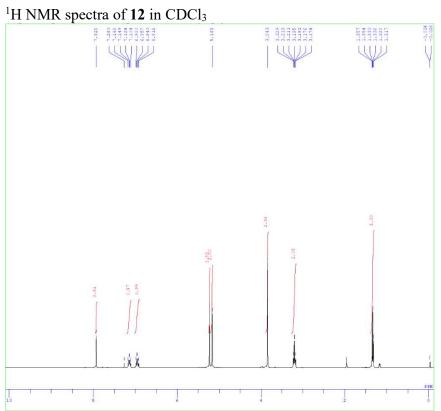
¹H NMR spectra of **11** in CDCl₃

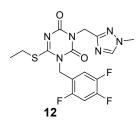


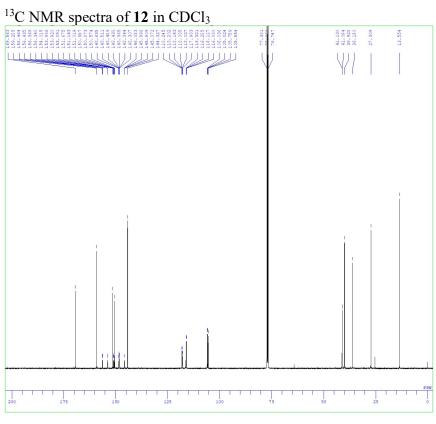
 13 C NMR spectra of **11** in DMSO- d_6



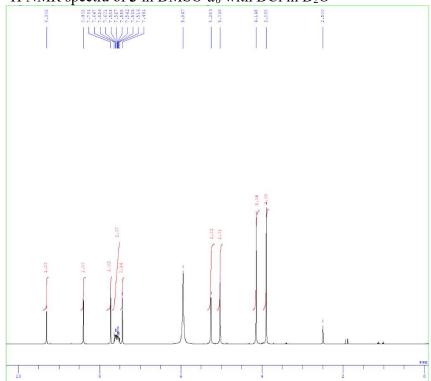




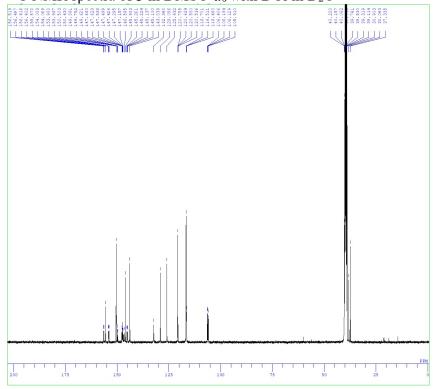




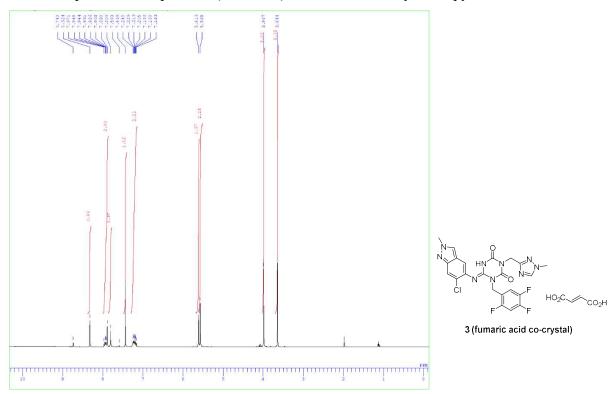




13 C NMR spectra of **3** in DMSO- d_6 with DCl in D₂O



 1 H NMR spectra of compound 3 (S-217622) fumaric acid co-crystal in pyridine- d_{5}

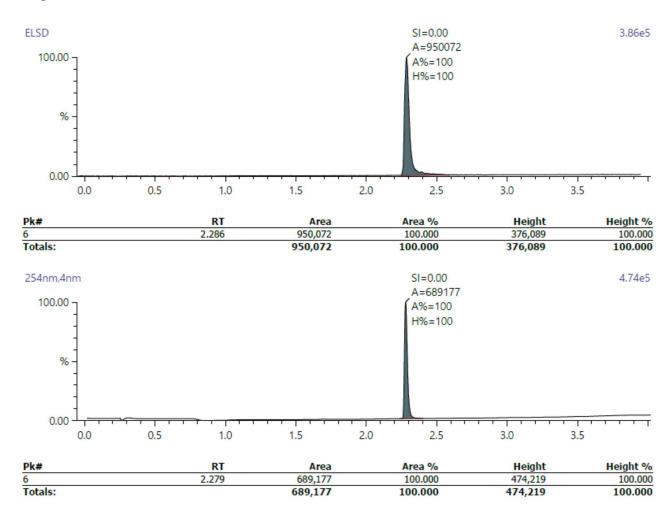


HPLC traces of compound 1-3

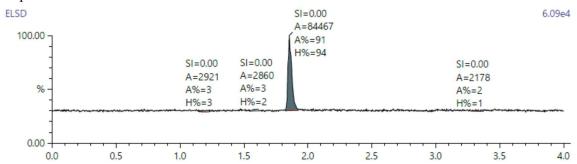
Procedure

Analytical liquid chromatography/mass spectroscopy (LC/MS) was performed on a Shimadzu Shimpack XR-ODS (C18, 2.2 μ m, 3.0 \times 50 mm, linear gradient from 10% to 100% B over 3 min, then 100% B for 1 min [A = water + 0.1% formic acid, B = MeCN + 0.1% formic acid], flow rate: 1.6 mL/min) using a Shimadzu UFLC system equipped with a LCMS-2020 mass spectrometer, LC-20AD binary gradient module, SPD-M20A photodiode array detector (detection at 254 nm), and SIL-20AC sample manager.

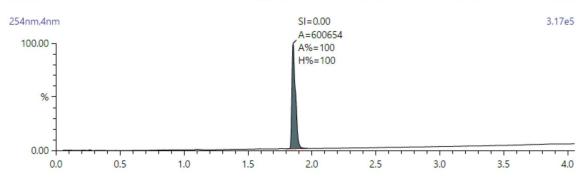
Compound 1



Compound 2

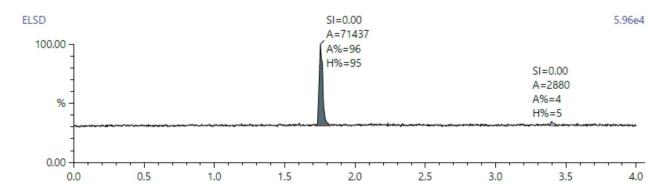


Pk#	RT	Area	Area %	Height	Height %
1	1.180	2,921	3.161	1,204	2.665
2	1.607	2,860	3.094	1,126	2.492
3	1.854	84,467	91.389	42,478	94.042
4	3.297	2,178	2.356	362	0.801
Totals:		92,426	100.000	45,169	100.000

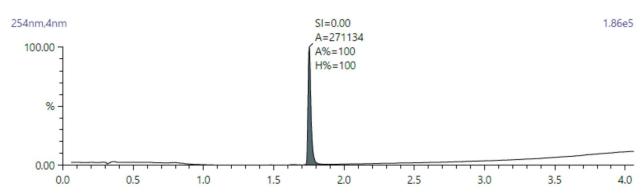


Pk#	RT	Area	Area %	Height	Height %
3	1.853	600,654	100.000	312,342	100.000
Totals:		600,654	100.000	312.342	100.000

Compound 3 (free form)



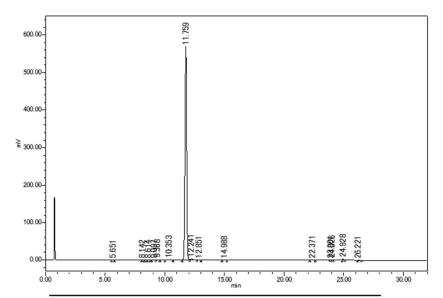
Pk#	RT	Area	Area %	Height	Height %
1	1.754	71,437	96.124	41,034	95.111
2	3.399	2,880	3.876	2,109	4.889
Totals:		74,317	100.000	43,144	100.000



Pk#	RT	Area	Area %	Height	Height %
1	1.752	271,134	100.000	189,771	100.000
Totals:		271,134	100.000	189,771	100.000

HPLC chromatogram of 3 (S-217622) fumaric acid co-crystal

Analytical liquid chromatography of S-217622 fumaric acid cocrystal was performed on a ACQUITY UPLC BEH C18 Column (1.7 $\mu m, 2.1 \ mm \ I.D.x100 \ mm,$ gradient from 10% to 100% B [A = 10 mmol/L HCOONH4 /H2O, B = MeCN], flow rate: 0.3 mL/min) using a Shimadzu Nexera system equipped with LC-20AD binary gradient module and SPD-20AV detector (detection at 255 nm).



Retention time Peak area Peak height % area (min) (μV·sec) (μV) 5.7 1179 320 0.02 8.1 212 0.02 1119 8.6 1316 219 0.02 9.1 1095 0.02 132 9.4 6948 987 0.12 10.4 14968 881 0.26 11.8 5559146 569193 98.21 12.2 45115 3668 0.80 12.9 1613 151 0.03 15.0 102 1242 0.02 22.4 4185 364 0.07 23.9 1721 436 0.03 24.0 4907 1408 0.09 24.9 13517 4116 0.24 26.2 2513 410 0.04

Time-program for gradient elution

	_
Time	Mobile
(min)	Phase B (%)
0	10
3	30
20	30
26	90
32	90
32.01	10
42	Stop

Experimental Procedures for in vitro safety.

Human ether-a-go-go-related gene inhibition assay

To evaluate an electrocardiogram QT interval prolongation of S-217622, effects on delayed rectifier K+ current (IKr) were evaluated using Chinese hamster ovary (CHO) cells expressing the human ether-a-go-go-related gene channel. The study was conducted in compliance with good laboratory practice regulations. CHO cells were retained at a membrane potential of –80 mV with a whole-cell patch-clamp system (EPC-10 amplifier/PatchMaster v2.8 software, HEKA Co., Ltd.), and IKr was elicited via repolarization pulse at –40 mV for 2 sec after a depolarization pulse at +20 mV for 1 sec. S-217622 (fumaric acid co-crystal) was dissolved in DMSO and diluted 200-fold with external solution to prepare an objective concentration. The S-217622 concentration levels were 10, 30 and 100 μM. E-4031 at 0.1 μM and the vehicle (0.5% DMSO) were applied as positive and negative controls, respectively.

From the recorded IKr, an absolute value of the tail peak current was measured based on the current value at the resting membrane potential using the whole-cell patch-clamp method. The percentages of the preapplication values in the test substance and control groups were compensated by the mean value of the percentage of the preapplication value in the negative-control group, and the compensated suppression rates were calculated.

Patch-clamp solutions were as follows. Internal solution: KCL: 130mmol/L, MgCl₂: 1 mmol/L, MgATP: 5 mmol/L, EGTA: 5 mmol/L, HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid): 10 mmol/L, pH=7.2. External solution: NaCl: 137 mmol/L, KCl: 4 mmol/L, CaCl₂: 1.8 mmol/L, MgCl₂: 1 mmol/L, glucose: 10 mmol/L, HEPES: 10 mmol/L, pH=7.4.

In vitro micronucleus test. To evaluate the clastogenic potential of S-217622, an *in vitro* micronucleus test was conducted using TK6 cells (human lymphoblast-derived) in a short-term treatment with and without a metabolic activation system (S9 mix) and continuous treatment without S9 mix. S9 mix containing 9,000 g of liver supernatant fraction was prepared from Sprague-Dawley rats treated with phenobarbital and 5,6-benzofravone. The study was conducted in compliance with good laboratory practice regulations.

A TK6 cell suspension was used in the absence of metabolic activation in the 3-h and 24-h treatment groups or in the presence of metabolic activation in the 3-h treatment group. Suspensions were mixed with S-217622 (fumaric acid co-crystal) in DMSO solution with S9 mix in the presence of metabolic activation

in the 4-h treatment group, then incubated at 37°C. The negative-control (DMSO) and positive-control (mitomycin C, cyclophosphamide monohydrate, or colchicine) substances were prepared concurrently. After the end of the short-term treatment, cells were washed and then incubated in fresh culture medium for 21 h. After incubation, the cells were counted to evaluate cytotoxicity, and the *in vitro* micronucleus test was conducted with S-217622 (fumaric acid co-crystal) at doses of 150–250 µg/mL for short-term treatment and 75–125 µg/mL for continuous treatment based on cytotoxicity. The nucleic acid was stained with acridine orange, and the micronuclear frequency of the specimens was observed under a fluorescence microscope. The test was considered positive when a significant and dose-dependent increase was noted in the number of cells with micronuclei in the test-substance groups compared with the negative-control group under any treatment condition.

Ames test. To evaluate the mutagenic potential of S-217622, a bacterial reverse-mutation test was conducted via the preincubation method using five bacterial strains, including *Salmonella typhimurium* (TA98, TA100, TA1535, and TA1537) and *Escherichia coli* (WP2*uvrA*) in the presence or absence of a metabolic activation system (S9 mix). S9 mix containing 9,000 g of liver supernatant fraction was prepared from Sprague-Dawley rats treated with phenobarbital and 5,6-benzofravone. The study was conducted in compliance with good laboratory practice regulations.

S-217622 (fumaric acid co-crystal, DMSO solution) was mixed with S9 mix in the presence of metabolic activation or phosphate buffer in the absence of metabolic activation and 0.1 mL of test strain suspension (1 × 10⁹ cells/mL or greater) and incubated at 37°C for 20 min. Then, the mixture with a layer of soft agar containing histidine and biotin or tryptophan was overlaid on minimal glucose agar plates. The negative-control (DMSO) and positive-control (4-nitroquinoline 1-oxide, sodium azide, 9-aminoacridine hydrochloride monohydrate, or 2-aminoanthracene) substances were prepared concurrently. The mutation test was conducted with S-217622 (fumaric acid co-crystal) at 156–5000 μg/plate in TA98, TA100, TA1535, and WP2*uvrA* and at 39.1–5000 μg/plate in TA1537. After incubation at 37°C for 48 h, the revertant colonies were counted and evaluated by comparing them with the negative-control group. The test was considered positive when the number of revertant colonies was concentration-dependently increased and twofold or greater increased over the number of colonies of the negative-control group.

In vitro **3T3.** To evaluate the phototoxicity potential of S-217622, an *in vitro* phototoxicity study was conducted with cultured mammalian cells. The study was conducted in compliance with good laboratory

practice regulations. A fibroblastic cell line derived from BALB/c mice (BALB/3T3 cells) was cultured

in 96-well plates and treated with S-217622 (fumaric acid co-crystal) for 1 h followed by UV-A (5 J/cm²)

and UV-B (68.7 mJ/cm²) irradiation. For the comparator, no irradiation was conducted. The phototoxicity

test was performed at 0.781-100 µg/mL. A vehicle (DMSO)-treated group and a chlorpromazine

hydrochloride-treated group were set as the negative and positive controls, respectively. Cell viability was

determined by neutral-red extraction from cells (measurement of absorbance at 540 nm). When the IC₅₀

could be determined for both the irradiated and nonirradiated plates, the result was determined from the

PIF. When the IC₅₀ could not be determined for either the irradiated or nonirradiated plates, the result was

determined from the MPE. The judgment criteria are shown below.

No phototoxicity: PIF < 5 or MPE < 0.15

Phototoxicity:

 $5 \le PIF$ or $0.15 \le MPE$

19

Table S1. The calculated IC₅₀ values for S-217622 (fumaric acid co-crystal) to SARS-CoV-2 3CL protease activity in each experiment.

Experiment	IC ₅₀ (μM)
Exp. No.1	0.0122
Exp. No.2	0.0130
Exp. No.3	0.0143
Mean	0.0132
Standard deviation	0.0011

Table S2. EC₅₀ values of S-217622 (fumaric acid co-crystal) and remdesivir on cytopathic effects in SARS-CoV-2 3CL-infected VeroE6/TMPRSS2 cells. ^astandard deviation. The mean and SD were calculated from three independent experiments.

	D.	EC ₅₀ (μM)			
Strains	Pango -	S-217622	2	Remdesivir	
	Lineage	Mean	SDa	Mean	SDa
hCoV-19/Japan/TY/WK-521/2020	A	0.37	0.060	1.9	0.14
hCoV-19/Japan/QK002/2020	B.1.1.7	0.33	0.050	0.87	0.027
hCoV-19/Japan/QHN001/2020	B.1.1.7	0.31	0.070	0.97	0.14
hCoV-19/Japan/QHN002/2020	B.1.1.7	0.46	0.044	0.99	0.18
hCoV-19/Japan/TY7-501/2021	P.1	0.50	0.048	2.1	0.39
hCoV-19/Japan/TY7-503/2021	P.1	0.43	0.00085	1.0	0.16
hCoV-19/Japan/TY8-612/2021	B.1.351	0.40	0.048	1.2	0.30
hCoV-19/Japan/TY11-927-P1/2021	B.1.617.2	0.41	0.014	1.6	0.22
hCoV-19/Japan/TY38-873/2021	B.1.1.529	0.29	0.054	1.1	0.28

Table S3. CC₅₀ values of S-217622 (fumaric acid co-crystal) and remdesivir against VeroE6/TMPRSS2 cells. ^astandard deviation. The mean and SD were calculated from three independent experiments.

Substance	CC_{50} (μM	()					
Substance	Exp.1	Exp.2	Exp.3	Mean	SD^a	SDa	
S-217622	>100	>100	>100	>100	-		
Remdesivir	>100	>100	>100	>100	-		

Table S4. *In vitro* safety profiles of S-217622 (fumaric acid co-crystal)

In vitro safety assays	Results
hERG inhibition assay	>100 μM
Bacterial reverse mutation test (Ames test)	Negative
Micronucleus test	Negative
3T3 assay	No Phototoxicity

Table S5. Diffraction data and refinement statistics for SARS-CoV-2 3CL^{pro} complexed with compound 1 and compound 3 (S-217622).

3CL^{pro} – Compound 1 3CL^{pro} – Compound 3 (S-217622) PDB code 7VTH 7VU6 Data collection Space group P21 P21 Cell dimensions a, b, c (Å) 44.41 54.27 114.50 55.47 99.23 58.88 90.00 99.42 90.00 90.00 108.05 90.00 α , β , γ (°) Wavelength (Å) 1.54178 1.54178 Resolution (Å) 28.24 - 2.00(2.05 - 2.00)28.68 - 1.80(1.84 -1.80) Completeness (%) 99.5 (98.9) 99.8(100.0) R_{merge} (%) a,b 9.0(27.7) 6.0(42.9) $I/\sigma(I)^a$ 8.3(3.3) 20.5(2.6) Refinement Resolution (Å) 112.952 - 2.00155.986-1.800 No. of reflections 34366 52687 $R_{\text{work}}/R_{\text{free}}$ 0.2050/0.2568 0.2241/0.2790 B factor (Å2) 16.2 17.8 Protein Ligand 27.4 14.3 Water 22.2 25.3 R.m.s deviations Bond length (Å) 0.0128 0.0175 Bond angles (°) 1.576 1.796 Ramachandran plot (%) Favored 97.2 97.5 Allowed 2.1 2.2 0.7 0.3 Outliers

Values in parentheses are for the highest resolution shell.

^b $R_{\text{merge}} = \Sigma |I - \langle I \rangle|/\Sigma I$, where *I* is the intensity of observation *I* and $\langle I \rangle$ is the mean intensity of the reflection.