Arid5a mediates an IL-17-dependent pathway that drives autoimmunity but not antifungal host defense

Tiffany C. Taylor¹, Yang Li¹, De-Dong Li¹, Saikat Majumder¹, Mandy McGeachy¹, Partha S. Biswas¹, Sebastien Gingras² and Sarah L. Gaffen^{1*}

¹ *Division of Rheumatology & Clinical Immunology, University of Pittsburgh, Pittsburgh, PA*

² *Department of Immunology, University of Pittsburgh, Pittsburgh, PA*

Running title: Arid5a functions selectively in IL-17 signaling

*Correspondence to:

Sarah L. Gaffen, University of Pittsburgh, Division of Rheumatology and Clinical Immunology, S702 BST, 200 Lothrop Street, Pittsburgh, PA 15261

Phone: 412-383-8903

Email: sarah.gaffen@pitt.edu

Conflict of interest statement: SLG has consulted for Aclaris Therapeutics and Eli Lilly.

Keywords: Arid5a, RNA Binding protein, IL-17, fungal infection, *Candida albicans*, autoimmunity

Abstract

IL-17 contributes to the pathogenesis of certain autoimmune diseases, but conversely is essential for host defense against fungi. Antibody-based biologic drugs that neutralize IL-17 are effective in autoimmunity but can be accompanied by adverse side effects. *Candida albicans* is a commensal fungus that is the primary causative agent of oropharyngeal and disseminated candidiasis. Defects in IL-17 signaling cause susceptibility to candidiasis in mice and humans. A key facet of IL-17 receptor signaling involves RNA binding proteins (RBP), which orchestrate the fate of target mRNA transcripts. In tissue culture models we showed that the RBP AT-rich interacting protein 5a (Arid5a) promotes the stability and/or translation of multiple IL-17-dependent mRNAs. Moreover, during OPC, Arid5a is elevated within the oral mucosa in an IL-17-dependent manner. However, the contribution of Arid5a to IL-17-driven events *in vivo* is poorly defined. Here, we used CRISPR/Cas9 to generate mice lacking Arid5a. *Arid5a*^{-/-} mice were fully resistant to experimental autoimmune encephalomyelitis (EAE), an autoimmune setting in which IL-17 signaling drives pathology. Surprisingly, *Arid5a*^{-/-} mice were resistant to OPC and systemic candidiasis, similar to immunocompetent WT mice and contrasting with mice defective in IL-17 signaling. Therefore, Arid5a-dependent signals mediate pathology in autoimmunity yet are not required for immunity to candidiasis, indicating that selective targeting of IL-17 signaling pathway components may be a viable strategy for development of therapeutics that spare IL-17-driven host defense.

Introduction

IL-17 (IL-17A) is a member of a subclass of cytokines that are structurally and functionally distinct from other inflammatory cytokines. Though produced mainly by T lymphocytes (“Type 17” cells, including $\gamma\delta$ - and $\alpha\beta$ -T cells, ILC3, NKT cells), IL-17 predominantly activates downstream signals in non-hematopoietic cells to protect against mucosal pathogens, notably the commensal fungus *Candida albicans* (1, 2). Conversely, IL-17 and Type 17 cells have been implicated in the pathogenesis of many autoimmune and inflammatory diseases (3), highlighted in the success of anti-cytokine drugs targeting IL-17 or the IL-17 receptor. Though unquestionably these drugs have improved patients’ disease activity and quality of life, their use is associated with adverse side effects including worsening bowel inflammation in patients with inflammatory bowel disease (IBD) and the frequent development of opportunistic fungal infections, especially oral and esophageal candidiasis (4-6).

IL-17 signals through a heterodimeric receptor composed of IL-17RA and IL-17RC, though some alternative configurations of the receptor have recently been described (7, 8). Binding of IL-17 to the IL-17R complex promotes recruitment of the adaptor Act1 (9-11). In turn, Act1 binds to multiple TRAF factors that promote alternative downstream signaling cascades (12). TRAF6 activates several transcription factors, including NF- κ B, I κ B ξ and C/EBP family members. Additionally, Act1 initiates recruitment of TRAF2 and TRAF5, which initiates a signaling cascade resulting in prolonged half-life of many IL-17-dependent mRNA transcripts. The effects of IL-17 signaling on mRNA fate is orchestrated by a complex array of RNA binding proteins (RBPs) (13). RBPs shape the output of inflammatory effector genes by stabilizing or degrading transcripts that encode inflammatory mediators. Some RBPs, including HuR (*Elavl1*) and IMP2 (*Igf2bp2*) act as positive regulators of mRNA expression, resulting in target mRNA stabilization or enhanced translation (14, 15). Alternatively, RBPs such as Regnase-1, Splicing factor 2 (SF2) and Roquins negatively regulate mRNA expression, facilitating degradation of unstable transcripts (16-18). These pathways are interlinked because many transcription factors operative in the IL-17 pathway are encoded by RNAs that are subject to post-transcriptional control, for example, I κ B ξ (*Nfkbiz*) and C/EBPs (*Cebpb*, *Cebpd*).

AT-rich interaction domain 5A (Arid5a) is another RBP implicated in IL-17 signaling and Th17-driven diseases. Arid5a enhances target mRNA transcript stability and translation, in part by offsetting the destabilizing activity of the endoribonuclease Regnase-1 (14, 19-21). Arid5a controls expression of genes downstream of the Th17/IL-17R axis through control of *Il6* and *Nfkbiz*, as well as Th17 cell responses via *Il6*, *STAT3*, *TBX21* (Tbet), and *OX40* transcripts (19, 21, 22). To date, the precise mechanisms by which IL-17 regulates Arid5a function in autoimmunity are not fully understood. Even less is known about Arid5a function in the context of host defense, though Arid5a mRNA is upregulated in an IL-17-dependent manner in oropharyngeal candidiasis (OPC) in the murine oral mucosa (14, 23). Here, we created a novel line of *Arid5a*^{-/-} mice to assess the role of Arid5a in autoimmunity versus fungal infections. These data reveal a surprising distinction used between signaling pathways critical in host defense compared to those required in autoimmunity.

Results

Characterization of Arid5a-deficient mice

To determine the role of Arid5a *in vivo*, CRISPR/Cas9 was used to create Arid5a-deficient mice with a deletion in exons 3-7 (**Fig 1A**, see Methods for details). Founders were validated by PCR (**Fig 1B**) and bred to homozygosity. Mice were born at expected Mendelian ratios and exhibited normal fertility and baseline body weights (data not shown). Arid5a is expressed at variable levels across tissues, with high expression in thymus, medium levels in spleen, and very low levels in kidney (**Fig 1C**). Confirming that mice lacked Arid5a protein expression, tissues from *Arid5a*^{-/-} mice displayed undetectable levels of Arid5a compared to *Arid5a*^{+/+} mice.

Previous studies using an independently generated Arid5a knockout mouse demonstrated a role for Arid5a in regulating several immune processes (19-22, 24-26). Therefore, we examined the baseline status of immune compartments in *Arid5a*^{+/+} and *Arid5a*^{-/-} littermates in spleen (**Fig 2A**) and thymus (**Fig 2B**). As shown, no observable differences between *Arid5a*^{+/+} and *Arid5a*^{-/-} mice were detected by flow cytometry in thymic or splenic hematopoietic cells, including TCRβ⁺ cells, B cells, neutrophils, monocytes or macrophages. Therefore, the absence of Arid5a does not appear to impact normal development of immune cells.

Arid5a is required for pathogenesis of EAE

Several reports have described a role for Arid5a in driving pathogenesis in mouse models of autoimmunity, notably through control of Th17 cell differentiation (19, 21). Accordingly, we subjected *Arid5a*^{-/-} mice and *Arid5a*^{+/+} littermate controls to a standard model of experimental autoimmune encephalomyelitis (EAE), an IL-17-dependent model of multiple sclerosis. Mice were injected with a myelin oligodendrocyte glycoprotein peptide (MOG 35-55) emulsified with complete Freund's Adjuvant (CFA). Mice were assessed daily for signs of ascending paralysis on a standard scoring system. *Arid5a*^{+/+} control mice developed a typical onset and clinical course of EAE, peaking at 14-16 days post immunization. As expected, mice lacking the IL-17 receptor adaptor, Act1 and *Arid5a*^{-/-} were resistant to disease (**Fig 3A**). Concomitant with reduced disease scores, there was marked reduction in EAE incidence in *Arid5a*^{-/-} mice (**Fig 3B**). These findings

confirm that *Arid5a* is required for development of EAE, and also functionally verify that this line of *Arid5a*-knockout mice has the expected phenotype with respect to IL-17-driven autoimmunity.

Arid5a is dispensable for immunity to oral and systemic *C. albicans* infections

IL-17R signaling is well established to drive oral immunity to the commensal fungus *C. albicans*, (2, 27, 28). Moreover, we previously observed that *Arid5a* mRNA is rapidly upregulated in an IL-17-dependent manner in the oral mucosa of mice during OPC (14). Because *Arid5a* drives cellular responses to IL-17 through post-transcriptional control of mRNA *in vitro* (14, 29), we predicted that *Arid5a*^{-/-} would likely be more susceptible to OPC than WT mice, which clear infection rapidly and do not exhibit overt signs of disease. To test this hypothesis, *Arid5a*^{-/-} mice were subjected to OPC by sublingual exposure with a cotton ball soaked with 2 x 10⁷ cells/ml *C. albicans* strain CAF2-1 (30). As a control, *Act1*^{-/-} mice (which lack the ability to respond to IL-17 signaling (31, 32)) showed high fungal loads at 5 days after infection (**Fig 4A**) and progressively lost weight throughout the time course of infection (**Fig 4B**). In contrast, *Arid5a*^{+/+} and *Arid5a*^{-/-} mice cleared the infection by day 5 and returned to normal body weights (**Fig 4A, B**).

Disseminated candidiasis is a serious nosocomial infection associated with high rates of morbidity and mortality in humans (2). In mouse models, IL-17R signaling mediates immunity to candidiasis (33, 34). Because *Arid5a* controls genes that drive Th17 differentiation as well as genes downstream of the IL-17 receptor (12, 29), we postulated that *Arid5a* deficiency would render mice susceptible to disseminated candidiasis. Therefore, we subjected *Arid5a*^{-/-} mice to disseminated candidiasis by intravenous inoculation of 2 x 10⁵ *C. albicans* yeast cells (strain SC5314). *Arid5a*^{-/-} mice had similar fungal loads as *Arid5a*^{+/+} mice (**Fig 4C**). Moreover, while *Act1*^{-/-} mice succumbed to disease by day 7, most *Arid5a*^{+/+} and *Arid5a*^{-/-} mice survived beyond day 10 and did not show statistically different survival patterns, indicating that *Arid5a* is not required for immunity to systemic *C. albicans* infections (**Fig 3D**). Collectively these data indicate that *Arid5a* is important for autoimmunity but does not play an important role in the response to *C. albicans*.

Discussion

Arid5a was originally described for its ability to interact with AT-rich DNA elements in DNA, modulating cellular proliferation, differentiation, and development (35-37). Arid5a belongs to the ARID family that consists of 15 proteins, which can be divided into seven sub-groups based on the degree of similarity within the ARID domain. The ARID domain is a helix-turn-helix motif essential for binding to DNA elements (35, 36). Recently, it was demonstrated that the ARID domain of Arid5a can also interact with cis-acting elements located in the 3' UTR of certain inflammatory RNAs, such as *Il6*, *Tbx21*, *Stat3*, *Ox40* (14, 19-22, 38). Binding of Arid5a to these transcripts promotes their longevity and/or facilitates their protein translation. Additionally, Arid5a offsets the action of a potent endoribonuclease, Regnase-1, which degrades target transcripts and thereby serves as a negative regulator of inflammation (19). Arid5a acts in hematopoietic cells (macrophages, T cells) as well as non-hematopoietic cells (fibroblasts, epithelial cells) (12, 29). Thus, Arid5a is a newly recognized RBP that is an important post-transcriptional modulator in multiple immune pathways.

The *Arid5a*^{-/-} mice described here are not identical to the original published *Arid5a*^{-/-} mice strain (19). The latter mouse lacks exons 1-3 and a major part of the DNA binding domain was replaced by a floxed Neo-cassette. In the *Arid5a*^{-/-} mice described here, LoxP sites were introduced in intron 2 and immediately following the stop codon, creating a 3,359 bp deletion. Despite these differences, both *Arid5a*^{-/-} lines are resistant to EAE (39), lending confidence that Arid5a plays an important role in driving Th17/IL-17-driven autoimmune inflammation.

IL-17 mediates effects through new gene expression, which can occur transcriptionally and post-transcriptionally (7). The data presented here indicate that *Arid5a*^{-/-} mice are, unexpectedly, resistant to oral and systemic candidiasis even though IL-17R signals are essential for immunity to this condition (33, 40, 41) and Arid5a functions downstream IL-17 to upregulate many IL-17 target genes (23, 42). Arid5a has been shown to act in T cells to stabilize genes such as *Il6*, *STAT3*, *Nfkbiz*, *OX40*, *Tbet* that drive Th17 differentiation and IL-17 production (20, 29). One explanation to reconcile these observations may be that the early responses to *C. albicans* in mice are innate in nature, as this fungus is not a commensal microbe in mice (43). Rather than conventional antigen-specific Th17 cells, IL-17 in these naïve settings comes primarily from a combination of $\gamma\delta$ -T cells and innate-acting CD4⁺ $\alpha\beta$ -T cells; additionally, adaptive T cell memory responses can be established in mice that appear to reflect human T cells responses (44-50). In line with this, *Il6*^{-/-}

and *Ccr6*^{-/-} mice are resistant to OPC, and Th1 cells (IL-12) are dispensable for protection against oral *C. albicans* (44, 51, 52). Thus, the genes regulated by *Arid5a* might not be important for protection against *Candida albicans* oral infections.

Anti-IL-17 therapy is not associated with risks to systemic candidiasis, likely because the major mechanisms of antifungal control at cutaneous/mucosal surfaces is through neutrophil recruitment and β -defensin production whereas circulating neutrophils are less IL-17-dependent (4, 41, 53, 54). In contrast, during systemic candidiasis IFN- γ , IL-12 as well as IL-17 signaling contribute to effective immunity against *C. albicans* (55-57). Previous studies showed that *Arid5a* regulates expression of IFN- γ in Th1 cells via control of the master Th1 transcription factor Tbet (22). Given this, we were surprised to find that *Arid5a*^{-/-} mice showed the same susceptibility to systemic candidiasis as their wild type littermate counterparts. It is possible that other mRNA stabilizing factors either in T cells or in IL-17-responsive cells [likely, renal tubular epithelial cells (34), though NK cells have been reported (58)] can compensate for the loss of *Arid5a* during *C. albicans* infection, but more work will be needed to dissect this phenomenon.

The use of biologics targeting IL-17 or IL-17 receptor was a major advance in the treatment of psoriasis, psoriatic arthritis, and ankylosing spondylitis (59). Unexpectedly, IL-17 blockade failed in Crohn's disease, which has been attributed to a vital tissue protective role of IL-17 in the intestine epithelium (5, 60-62). Intriguingly, IL-17 has been linked to controlling social behavior in mice (63), and blockade of IL-17RA was linked to a small risk of suicide in humans (64), though this is controversial (65). Thus, an ideal therapeutic approach to blocking IL-17 would inhibit its pathogenic effects while sparing host-defensive pathways. Based on our data, *Arid5a* may be an attractive candidate for selective blockade in IL-17-driven diseases. Indeed, the development and potential uses of RNA therapeutics is becoming increasingly appreciated (66). Furthermore, *Arid5a* was reported to be a target of the antipsychotic drug chlorpromazine (20), though this remains to be confirmed. Targeting individual pathway components such as *Arid5a* potentially allows for a more targeted approach to treat IL-17-mediated inflammatory/autoimmune diseases while preserving the protective role of IL-17 in host defense.

Material and Methods

Generation of *Arid5a*^{-/-} mice

The *Arid5a* knockout allele was fortuitously generated during an attempt to create a conditional deletion with CRISPR/Cas9 (67, 68). Briefly, the targeting strategy was to introduce a LoxP site in intron 2 and a Myc-tag at the C-terminal with another loxP site just after the stop codon. In the process of genotyping potential founders, we identified a mouse carrying a deletion between the two SpyCas9 target sites, with a deletion of 3,359 bp deletion on chromosome 1 between position 36,316,840 and 36,320,199. Fertilized embryos (C57BL/6J, The Jackson Laboratory) produced by natural mating were microinjected in the cytoplasm with a mixture of 0.33 μM EnGen Cas9 protein (New England Biolabs, M0646T), *Arid5a*-guide2 and *Arid5a*-guide5 (21.23 ng/μl (~0.66 μM) and two single stranded oligonucleotides: *Arid5a*-Myc-HDR and *Arid5a*-guide5-HDR (0.5 μM, “Ultramers” from IDT). Injected zygotes were cultured overnight, and 2-cell embryos were transferred to pseudopregnant CD1 mice. The sgRNA templates were generated by PCR (67) and used for synthesis of sgRNAs using NEB HiScribe™ T7 Quick High Yield RNA Synthesis Kit (New England Biolabs, E2050S). The sgRNAs purified using MEGAclear Kit (ThermoFisher Scientific). The target sequence of the sgRNAs are :ATAGGCTCTGGCCTACAGTTTGG for *Arid5a*-guide2 at chr1:36377037-36377060 and GTAAAAGCCAAATGCGCCCCAGG for *Arid5a*-guide5 at chr1:36373679-36373702 (coordinates from GRCm38/mm10 Assembly).

The sequence of the single-stranded oligonucleotides are: 5’-

tcgaactcagagagttctgcctgcttctcctgagtctaggattaaaggtgtgtgccaccactgcctgggAATTCATAACTTCG
TATAATGTATGCTATACGAAGTTATgcgcatgttgctttaccatacgggtgagggactctgaccctgccttcagga
actgagttggataa-3’ for *Arid5a*-guide5-HDR

5’-

TGGCACATGCCACCCGTCACAACCTATGCGGCACCTCACTTCTTCCACCTCAACACC
AAACTGGAGCAGAACTCATCTCAGAAGAGGATCTGTAAGAATTCATAACTTCGTA
TAATGTATGCTATACGAAGTTATAGAGCCTATCCTGCTATGCTGTGGAGGATTTGAT
GGGCAGCTGCCGCCATTATCTCAGGCC-3’ for *Arid5a*-Myc-HDR.

The potential founders were identified by PCR and the sequence of the knockout allele was determined by sequencing. *Arid5a*^{-/-} mice or littermates were genotyped with the following primers:

F31-5'-ACCTTCTGGCTACAACAGGC-3'

R31-5'-ACCTACACACTTGCTCCTGC-3'

F51-5'-ATATGGGTGCTAGGCAAGGC-3'

The WT allele is detected with primer F31 and R31 and produces a product of 564 bp. The knockout allele is detected with primer F51 and R31 and produces a product of 595 bp.

Candidiasis Models

OPC was induced in 7-9 week old mice of both sexes by sublingual inoculation with *C. albicans* strain CAF2-1-saturated cotton balls for 75 minutes under anesthesia, as described (30, 51). Tongue homogenates were prepared on GentleMACS homogenizers (Miltenyi Biotec) with C-tubes, and CFU was determined by serial dilution plating on Yeast extract-peptone-dextrose (YDP) with ampicillin. Limit of detection is ~30 CFU/g. For systemic candidiasis, mice were injected i.v. with $2-7 \times 10^5$ *C. albicans* strain SC5412 in PBS. Three days post-infection, kidney tissues were homogenized in C-tubes, and CFU was determined as above.

Experimental autoimmune encephalomyelitis

Mice (males age 6-9 weeks) were immunized subcutaneously in 4 sites on the back with 100 µg MOG peptide 35-55 (Biosynthesis) emulsified with Complete Freund's Adjuvant (CFA) with *M. tuberculosis* strain H37Ra (DIFCO laboratories) and administered 100 ng pertussis toxin (List Biological Laboratories) i.p. on day 0 and day 2, as described (69). Mice were scored daily as follows: 1. Flaccid tail; 2. Impaired righting reflex and hindlimb weakness; 3. Partial hindlimb paralysis; 4. Complete hindlimb paralysis; 5. Hindlimb paralysis with partial forelimb paralysis; 6. Moribund.

Flow Cytometry

Tongues were harvested and digested with Collagenase IV (0.7 mg/ml) in Hanks' balanced salt solution for 30-45 mins at 37°C. Cells were separated by Percoll gradient centrifugation. Abs were from the following sources: anti-CD45 and anti-CD11b (BioLegend), anti-CD4 and anti-F4/80 (Invitrogen), anti-TCRβ, anti-CD8, and anti-CD19 (eBioscience), and anti-Ly6G and anti-Ly6C (BD Biosciences). Dead cells were excluded using Ghost Dye (eBioscience). Data were acquired with an LSRFortessa and analyzed using FlowJo software (TreeStar).

Immune Cell Isolation

Naïve splenocytes and thymocytes were isolated from 6 week old *Arid5a*^{-/-} and *Arid5a*^{+/+} male mice. Flow cytometry was performed on single cell suspensions.

Western Blotting

Western blotting was performed as described (42). Abs against Arid5a were from Abcam (ab81149). Blots were imaged with a FluorChem E imager (ProteinSimple, Santa Clara CA).

Statistics

Data were analyzed on Graphpad Prism. Each symbol represents one mouse unless indicated. **P*<0.05, **<0.01, ***<0.001, ****<0.0001. *P* values less than 0.05 were considered significant.

Study approval

All the experiments were conducted following NIH guidelines under protocols approved by the University of Pittsburgh IACUC.

Acknowledgements

This work was supported by NIH grants to SLG (AI144436, AI147383, DE022550). PSB was supported by AI142354 and AI159058, and MJM by AI148356. TCT was supported by T32-AI089443. We thank B.M. Coleman, N. Amatya and R. Bechara for valuable input.

Conflicts of interest: SLG has consulted for Aclaris Therapeutics and Eli Lilly. There are no other conflicts of interest.

Author Contributions

Conceptualization: SLG

Methodology, SG, SLG

Investigation: TCT, YL, DL, SM, PSB, SG

Writing – original draft, TCT, SLG

writing- review and editing, PSB, SG, SLG, MJM

Funding Acquisition, SLG, PSB, MJM

Supervision, SLG, PSB, MJM

References

1. Conti, H. R., and S. L. Gaffen. 2015. IL-17-Mediated Immunity to the Opportunistic Fungal Pathogen *Candida albicans*. *J Immunol* 195: 780-788.
2. Lionakis, M. S., and S. M. Levitz. 2018. Host Control of Fungal Infections: Lessons from Basic Studies and Human Cohorts. *Annual review of immunology* 36: 157-191.
3. McGeachy, M. J., D. J. Cua, and S. L. Gaffen. 2019. The IL-17 Family of Cytokines in Health and Disease. *Immunity* 50: 892-906.
4. Davidson, L., J. van den Reek, M. Bruno, F. van Hunsel, R. M. C. Herings, V. Matzaraki, C. K. Boahen, V. Kumar, H. M. M. Groenewoud, F. L. van de Veerdonk, M. G. Netea, E. de Jong, and B. J. Kullberg. 2022. Risk of candidiasis associated with interleukin-17 inhibitors: A real-world observational study of multiple independent sources. *Lancet Reg Health Eur* 13: 100266.
5. Fauny, M., D. Moulin, F. D'Amico, P. Netter, N. Petitpain, D. Arnone, J. Y. Jouzeau, D. Loeuille, and L. Peyrin-Biroulet. 2020. Paradoxical gastrointestinal effects of interleukin-17 blockers. *Ann Rheum Dis* 79: 1132-1138.
6. Smith, M. K., J. Pai, R. Panaccione, P. Beck, J. G. Ferraz, and H. Jijon. 2019. Crohn's-like disease in a patient exposed to anti-Interleukin-17 blockade (Ixekizumab) for the treatment of chronic plaque psoriasis: a case report. *BMC Gastroenterol* 19: 162.
7. Li, X., R. Bechara, J. Zhao, M. J. McGeachy, and S. L. Gaffen. 2019. IL-17 receptor-based signaling and implications for disease. *Nat Immunol* 20: 1594-1602.
8. Chung, S. H., X. Q. Ye, and Y. Iwakura. 2021. Interleukin-17 family members in health and disease. *Int Immunol* 33: 723-729.
9. Sonder, S. U., S. Saret, W. Tang, D. E. Sturdevant, S. F. Porcella, and U. Siebenlist. 2011. IL-17-induced NF-kappaB activation via CIKS/Act1: physiologic significance and signaling mechanisms. *The Journal of biological chemistry* 286: 12881-12890.
10. Chang, S. H., H. Park, and C. Dong. 2006. Act1 adaptor protein is an immediate and essential signaling component of interleukin-17 receptor. *The Journal of biological chemistry* 281: 35603-35607.
11. Qian, Y., C. Liu, J. Hartuppee, C. Z. Altuntas, M. F. Gulen, D. Jane-Wit, J. Xiao, Y. Lu, N. Giltiy, J. Liu, T. Kordula, Q. W. Zhang, B. Vallance, S. Swaidani, M. Aronica, V. K. Tuohy, T. Hamilton, and X. Li. 2007. The adaptor Act1 is required for interleukin 17-dependent signaling associated with autoimmune and inflammatory disease. *Nature immunology* 8: 247-256.
12. Amatya, N., A. V. Garg, and S. L. Gaffen. 2017. IL-17 Signaling: The Yin and the Yang. *Trends Immunol* 38: 310-322.
13. Bulek, K., C. Liu, S. Swaidani, L. Wang, R. C. Page, M. F. Gulen, T. Herjan, A. Abbadi, W. Qian, D. Sun, M. Lauer, V. Hascall, S. Misra, M. R. Chance, M. Aronica, T. Hamilton, and X. Li. 2011. The inducible kinase IKKi is required for IL-17-dependent signaling associated with neutrophilia and pulmonary inflammation. *Nat Immunol* 12: 844-852.
14. Amatya, N., E. E. Childs, J. A. Cruz, F. E. Y. Aggor, A. V. Garg, A. J. Berman, J. E. Gudjonsson, U. Atasoy, and S. L. Gaffen. 2018. IL-17 integrates multiple self-reinforcing, feed-forward mechanisms through the RNA binding protein Arid5a. *Sci Signal* 11.
15. Herjan, T., P. Yao, W. Qian, X. Li, C. Liu, K. Bulek, D. Sun, W. P. Yang, J. Zhu, A. He, J. A. Carman, S. C. Erzurum, H. D. Lipshitz, P. L. Fox, T. A. Hamilton, and X. Li. 2013.

- HuR is required for IL-17-induced Act1-mediated CXCL1 and CXCL5 mRNA stabilization. *J Immunol* 191: 640-649.
16. Sun, D., M. Novotny, K. Bulek, C. Liu, X. Li, and T. Hamilton. 2011. Treatment with IL-17 prolongs the half-life of chemokine CXCL1 mRNA via the adaptor TRAF5 and the splicing-regulatory factor SF2 (ASF). *Nat Immunol* 12: 853-860.
17. Garg, A. V., N. Amatya, K. Chen, J. A. Cruz, P. Grover, N. Whibley, H. R. Conti, G. Hernandez Mir, T. Sirakova, E. C. Childs, T. E. Smithgall, P. S. Biswas, J. K. Kolls, M. J. McGeachy, P. E. Kolattukudy, and S. L. Gaffen. 2015. MCP1 Endoribonuclease Activity Negatively Regulates Interleukin-17-Mediated Signaling and Inflammation. *Immunity* 43: 475-487.
18. Jeltsch, K. M., D. Hu, S. Brenner, J. Zoller, G. A. Heinz, D. Nagel, K. U. Vogel, N. Rehage, S. C. Warth, S. L. Edelmann, R. Gloury, N. Martin, C. Lohs, M. Lech, J. E. Stehklein, A. Geerlof, E. Kremmer, A. Weber, H. J. Anders, I. Schmitz, M. Schmidt-Supprian, M. Fu, H. Holtmann, D. Krappmann, J. Ruland, A. Kallies, M. Heikenwalder, and V. Heissmeyer. 2014. Cleavage of roquin and regnase-1 by the paracaspase MALT1 releases their cooperatively repressed targets to promote T(H)17 differentiation. *Nature immunology* 15: 1079-1089.
19. Masuda, K., B. Ripley, R. Nishimura, T. Mino, O. Takeuchi, G. Shioi, H. Kiyonari, and T. Kishimoto. 2013. Arid5a controls IL-6 mRNA stability, which contributes to elevation of IL-6 level in vivo. *Proc Natl Acad Sci U S A* 110: 9409-9414.
20. Masuda, K., B. Ripley, K. K. Nyati, P. K. Dubey, M. M. Zaman, H. Hanieh, M. Higa, K. Yamashita, D. M. Standley, T. Mashima, M. Katahira, T. Okamoto, Y. Matsuura, O. Takeuchi, and T. Kishimoto. 2016. Arid5a regulates naive CD4⁺ T cell fate through selective stabilization of Stat3 mRNA. *The Journal of experimental medicine* 213: 605-619.
21. Hanieh, H., K. Masuda, H. Metwally, J. P. Chalise, M. Mohamed, K. K. Nyati, D. M. Standley, S. Li, M. Higa, M. M. Zaman, and T. Kishimoto. 2018. Arid5a stabilizes OX40 mRNA in murine CD4⁺ T cells by recognizing a stem-loop structure in its 3'UTR. *Eur J Immunol* 48: 593-604.
22. Zaman, M. M., K. Masuda, K. K. Nyati, P. K. Dubey, B. Ripley, K. Wang, J. P. Chalise, M. Higa, H. Hanieh, and T. Kishimoto. 2016. Arid5a exacerbates IFN-gamma-mediated septic shock by stabilizing T-bet mRNA. *Proc Natl Acad Sci U S A* 113: 11543-11548.
23. Puel, A., and J.-L. Casanova. 2018. Arid5a makes the IL-17A/F-responsive pathway less arid. *Science Signaling* 11: eaau8876.
24. Dubey, P. K., K. Masuda, K. K. Nyati, M. M. Uz Zaman, J. P. Chalise, D. Millrine, W. Kai, B. Ripley, and T. Kishimoto. 2017. Arid5a-deficient mice are highly resistant to bleomycin-induced lung injury. *Int Immunol* 29: 79-85.
25. Nyati, K. K., K. Masuda, M. M. Zaman, P. K. Dubey, D. Millrine, J. P. Chalise, M. Higa, S. Li, D. M. Standley, K. Saito, H. Hanieh, and T. Kishimoto. 2017. TLR4-induced NF-kappaB and MAPK signaling regulate the IL-6 mRNA stabilizing protein Arid5a. *Nucleic Acids Res* 45: 2687-2703.
26. Higa, M., M. Oka, Y. Fujihara, K. Masuda, Y. Yoneda, and T. Kishimoto. 2018. Regulation of inflammatory responses by dynamic subcellular localization of RNA-binding protein Arid5a. *Proc Natl Acad Sci U S A* 115: E1214-E1220.
27. Conti, H. R., F. Shen, N. Nayyar, E. Stocum, J. N. Sun, M. J. Lindemann, A. W. Ho, J. H. Hai, J. J. Yu, J. W. Jung, S. G. Filler, P. Masso-Welch, M. Edgerton, and S. L. Gaffen.

2009. Th17 cells and IL-17 receptor signaling are essential for mucosal host defense against oral candidiasis. *J Exp Med* 206: 299-311.
28. Li, J., D. C. Vinh, J. L. Casanova, and A. Puel. 2017. Inborn errors of immunity underlying fungal diseases in otherwise healthy individuals. *Curr Opin Microbiol* 40: 46-57.
29. Nyati, K. K., M. M. Zaman, P. Sharma, and T. Kishimoto. 2020. Arid5a, an RNA-Binding Protein in Immune Regulation: RNA Stability, Inflammation, and Autoimmunity. *Trends Immunol* 41: 255-268.
30. Solis, N. V., and S. G. Filler. 2012. Mouse model of oropharyngeal candidiasis. *Nat Protoc* 7: 637-642.
31. Leonardi, A., A. Chariot, E. Claudio, K. Cunningham, and U. Siebenlist. 2000. CIKS, a connection to Ikappa B kinase and stress-activated protein kinase. *Proc Natl Acad Sci U S A* 97: 10494-10499.
32. Claudio, E., S. U. Sonder, S. Saret, G. Carvalho, T. R. Ramalingam, T. A. Wynn, A. Chariot, A. Garcia-Perganeda, A. Leonardi, A. Paun, A. Chen, N. Y. Ren, H. Wang, and U. Siebenlist. 2009. The adaptor protein CIKS/Act1 is essential for IL-25-mediated allergic airway inflammation. *J Immunol* 182: 1617-1630.
33. Huang, W., L. Na, P. L. Fidel, and P. Schwarzenberger. 2004. Requirement of interleukin-17A for systemic anti-Candida albicans host defense in mice. *J Infect Dis* 190: 624-631.
34. Ramani, K., C. V. Jawale, A. H. Verma, B. M. Coleman, J. K. Kolls, and P. S. Biswas. 2018. Unexpected kidney-restricted role for IL-17 receptor signaling in defense against systemic Candida albicans infection. *JCI Insight* 3.
35. Wilsker, D., A. Patsialou, P. B. Dallas, and E. Moran. 2002. ARID proteins: a diverse family of DNA binding proteins implicated in the control of cell growth, differentiation, and development. *Cell Growth Differ* 13: 95-106.
36. Patsialou, A., D. Wilsker, and E. Moran. 2005. DNA-binding properties of ARID family proteins. *Nucleic Acids Res* 33: 66-80.
37. Wilsker, D., L. Probst, H. M. Wain, L. Maltais, P. W. Tucker, and E. Moran. 2005. Nomenclature of the ARID family of DNA-binding proteins. *Genomics* 86: 242-251.
38. Masuda, K., and T. Kishimoto. 2018. A Potential Therapeutic Target RNA-binding Protein, Arid5a for the Treatment of Inflammatory Disease Associated with Aberrant Cytokine Expression. *Curr Pharm Des* 24: 1766-1771.
39. Masuda, K., B. Ripley, R. Nishimura, T. Mino, O. Takeuchi, G. Shioi, H. Kiyonari, and T. Kishimoto. 2013. Arid5a controls IL-6 mRNA stability, which contributes to elevation of IL-6 level in vivo. *Proc Natl Acad Sci U S A*.
40. Puel, A., S. Cypowji, J. Bustamante, J. Wright, L. Liu, H. Lim, M. Migaud, L. Israel, M. Chrabieh, M. Audry, M. Gumbleton, A. Toulon, C. Bodemer, J. El-Baghdadi, M. Whitters, T. Paradis, J. Brooks, M. Collins, N. Wolfman, S. Al-Muhsen, M. Galicchio, L. Abel, C. Picard, and J.-L. Casanova. 2011. Chronic mucocutaneous candidiasis in humans with inborn errors of interleukin-17 immunity. *Science* 332: 65-68.
41. Conti, H., V. Bruno, E. Childs, S. Daugherty, J. Hunter, B. Mengesha, D. Saevig, M. Hendricks, B. M. Coleman, L. Brane, N. V. Solis, J. A. Cruz, A. Verma, A. Garg, A. G. Hise, J. Naglik, J. R. Naglik, S. G. Filler, J. K. Kolls, S. Sinha, and S. L. Gaffen. 2016. IL-17RA signaling in oral epithelium is critical for protection against oropharyngeal candidiasis. *Cell Host Microbe* 20: 606-617.

42. Amatya, N., C. EE, J. A. Cruz, F. Aggor, A. Garg, A. Berman, J. E. Gudjonsson, U. Atasoy, and S. L. Gaffen. 2018. IL-17 integrates multiple self-reinforcing, feed-forward mechanisms through the RNA-binding protein Arid5a. *Science Signaling* 11: eaat4617.
43. Suegara, N., J. E. Siegel, and D. C. Savage. 1979. Ecological determinants in microbial colonization of the murine gastrointestinal tract: adherence of *Torulopsis pintolopesii* to epithelial surfaces. *Infect Immun* 25: 139-145.
44. Conti, H., A. Peterson, A. Huppler, L. Brane, N. Hernández-Santos, N. Whibley, A. Garg, M. Simpson-Abelson, G. Gibson, A. Mamo, L. Osborne, S. Bishu, N. Ghilardi, U. Siebenlist, S. Watkins, D. Artis, M. McGeachy, and S. Gaffen. 2014. Oral-resident 'natural' Th17 cells and $\gamma\delta$ -T cells control opportunistic *Candida albicans* infections. *The Journal of experimental medicine* 211: 2075-2084.
45. Verma, A., J. Richardson, C. Zhou, B. M. Coleman, D. Moyes, J. Ho, A. R. Huppler, K. Ramani, M. J. McGeachy, I. A. Mufazalov, A. Waisman, L. P. Kane, P. Biswas, B. Hube, J. Naglik, and S. L. Gaffen. 2017. Oral epithelial cells orchestrate innate Type 17 responses to *Candida albicans* through the virulence factor Candidalysin. *Sci Immunol* 2: eam8834.
46. Bär, E., A. Gladiator, S. Bastidas, B. Roschitzki, H. Acha-Orbea, A. Oxenius, and S. LeibundGut-Landmann. 2012. A novel Th cell epitope of *Candida albicans* mediates protection from fungal infection. *J Immunol* 188: 5636-5643.
47. Hernández-Santos, N., A. R. Huppler, A. C. Peterson, S. A. Khader, M. KC, and S. L. Gaffen. 2013. Th17 cells confer long term adaptive immunity to oral mucosal *Candida albicans* infections. *Mucosal Immunol* 6: 900-910.
48. Sparber, F., T. Dolowschiak, S. Mertens, L. Lauener, B. E. Clausen, N. Joller, P. Stoitzner, R. Tussiwand, and S. LeibundGut-Landmann. 2018. Langerin⁺ DCs regulate innate IL-17 production in the oral mucosa during *Candida albicans*-mediated infection. *PLoS Pathog* 14: e1007069.
49. Kirchner, F. R., and S. LeibundGut-Landmann. 2021. Tissue-resident memory Th17 cells maintain stable fungal commensalism in the oral mucosa. *Mucosal Immunol* 14: 455-467.
50. Break, T. J., V. Oikonomou, N. Dutzan, J. V. Desai, M. Swidergall, T. Freiwald, D. Chauss, O. J. Harrison, J. Alejo, D. W. Williams, S. Pittaluga, C. R. Lee, N. Bouladoux, M. Swamydas, K. W. Hoffman, T. Greenwell-Wild, V. M. Bruno, L. B. Rosen, W. Lwin, A. Renteria, S. M. Pontejo, J. P. Shannon, I. A. Myles, P. Olbrich, E. M. N. Ferre, M. Schmitt, D. Martin, Genomics, C. Computational Biology, D. L. Barber, N. V. Solis, L. D. Notarangelo, D. V. Serreze, M. Matsumoto, H. D. Hickman, P. M. Murphy, M. S. Anderson, J. K. Lim, S. M. Holland, S. G. Filler, B. Afzali, Y. Belkaid, N. M. Moutsopoulos, and M. S. Lionakis. 2021. Aberrant type 1 immunity drives susceptibility to mucosal fungal infections. *Science* 371.
51. Conti, H., F. Shen, N. Nayyar, E. Stocum, S. JN, M. Lindemann, A. Ho, J. Hai, J. Yu, J. Jung, S. Filler, P. Masso-Welch, M. Edgerton, and S. Gaffen. 2009. Th17 cells and IL-17 receptor signaling are essential for mucosal host defense against oral candidiasis. *The Journal of experimental medicine* 206: 299-311.
52. Farah, C., Y. Hu, S. Riminton, and R. Ashman. 2006. Distinct roles for interleukin-12p40 and tumour necrosis factor in resistance to oral candidiasis defined by gene targeting. *Oral Microbiol Immunol* 21: 252-255.
53. Conti, H., O. Baker, A. Freeman, W. Jang, R. Li, S. Holland, M. Edgerton, and S. Gaffen. 2011. New mechanism of oral immunity to mucosal candidiasis in hyper-IgE syndrome. *Mucosal Immunol* 4: 448-455.

54. Lionakis, M. S., J. K. Lim, C. C. Lee, and P. M. Murphy. 2011. Organ-specific innate immune responses in a mouse model of invasive candidiasis. *J Innate Immun* 3: 180-199.
55. Cenci, E., A. Mencacci, R. Spaccapelo, L. Tonnetti, P. Mosci, K. H. Enssle, P. Puccetti, L. Romani, and F. Bistoni. 1995. T helper cell type 1 (Th1)- and Th2-like responses are present in mice with gastric candidiasis but protective immunity is associated with Th1 development. *J Infect Dis* 171: 1279-1288.
56. Kaposzta, R., P. Tree, L. Marodi, and S. Gordon. 1998. Characteristics of invasive candidiasis in gamma interferon- and interleukin-4-deficient mice: role of macrophages in host defense against *Candida albicans*. *Infect Immun* 66: 1708-1717.
57. Balish, E., R. D. Wagner, A. Vazquez-Torres, C. Pierson, and T. Warner. 1998. Candidiasis in interferon-gamma knockout (IFN-gamma^{-/-}) mice. *J Infect Dis* 178: 478-487.
58. Bar, E., P. G. Whitney, K. Moor, C. Reis e Sousa, and S. LeibundGut-Landmann. 2014. IL-17 regulates systemic fungal immunity by controlling the functional competence of NK cells. *Immunity* 40: 117-127.
59. Ruiz de Morales, J. M. G., L. Puig, E. Dauden, J. D. Canete, J. L. Pablos, A. O. Martin, C. G. Juanatey, A. Adan, X. Montalban, N. Borruel, G. Orti, E. Holgado-Martin, C. Garcia-Vidal, C. Vizcaya-Morales, V. Martin-Vazquez, and M. A. Gonzalez-Gay. 2020. Critical role of interleukin (IL)-17 in inflammatory and immune disorders: An updated review of the evidence focusing in controversies. *Autoimmun Rev* 19: 102429.
60. Lin, X., S. J. Gaudino, K. K. Jang, T. Bahadur, A. Singh, A. Banerjee, M. Beaupre, T. Chu, H. T. Wong, C. K. Kim, C. Kempen, J. Axelrad, H. Huang, S. Khalid, V. Shah, O. Eskiocak, O. B. Parks, A. Berisha, J. P. McAleer, M. Good, M. Hoshino, R. Blumberg, A. B. Bialkowska, S. L. Gaffen, J. K. Kolls, V. W. Yang, S. Beyaz, K. Cadwell, and P. Kumar. 2022. IL-17RA-signaling in Lgr5(+) intestinal stem cells induces expression of transcription factor ATOH1 to promote secretory cell lineage commitment. *Immunity* 55: 237-253 e238.
61. Lee, J., C. Tato, B. Joyce-Shaikh, F. Gulan, C. Cayatte, Y. Chen, W. Blumenschein, M. Judo, G. Ayanoglu, T. McClanahan, X. Li, and D. Cua. 2015. Interleukin-23-dependent IL-17 production regulates intestinal epithelial permeability. *Immunity* 43: 727-738.
62. Maxwell, J., Y. Zhang, W. Brown, C. Smith, F. Byrne, M. Florino, E. Stevens, J. Bigler, J. Davis, J. Rottman, A. Budelsky, A. Symons, and J. Towne. 2015. Differential roles for interleukin-23 and interleukin-17 in intestinal immunoregulation. *Immunity* 43: 739-750.
63. Reed, M. D., Y. S. Yim, R. D. Wimmer, H. Kim, C. Ryu, G. M. Welch, M. Andina, H. O. King, A. Waisman, M. M. Halassa, J. R. Huh, and G. B. Choi. 2020. IL-17a promotes sociability in mouse models of neurodevelopmental disorders. *Nature* 577: 249-253.
64. Mullard, A. 2017. New plaque psoriasis approval carries suicide warning. *Nat Rev Drug Discov* 16: 155.
65. Chiricozzi, A., M. Romanelli, R. Saraceno, and T. Torres. 2016. No meaningful association between suicidal behavior and the use of IL-17A-neutralizing or IL-17RA-blocking agents. *Expert Opin Drug Saf* 15: 1653-1659.
66. Damase, T. R., R. Sukhovshin, C. Boada, F. Taraballi, R. I. Pettigrew, and J. P. Cooke. 2021. The Limitless Future of RNA Therapeutics. *Front Bioeng Biotechnol* 9: 628137.
67. Pelletier, S., S. Gingras, and D. R. Green. 2015. Mouse genome engineering via CRISPR-Cas9 for study of immune function. *Immunity* 42: 18-27.

68. Wang, H., H. Yang, C. S. Shivalila, M. M. Dawlaty, A. W. Cheng, F. Zhang, and R. Jaenisch. 2013. One-step generation of mice carrying mutations in multiple genes by CRISPR/Cas-mediated genome engineering. *Cell* 153: 910-918.
69. McGeachy, M. J., Y. Chen, C. M. Tato, A. Laurence, B. Joyce-Shaikh, W. M. Blumenschein, T. K. McClanahan, J. J. O'Shea, and D. J. Cua. 2009. The interleukin 23 receptor is essential for the terminal differentiation of interleukin 17-producing effector T helper cells in vivo. *Nature immunology* 10: 314-324.

Figure Legends

Figure 1. Creation of *Arid5a*^{-/-} mice by CRISPR/Cas9. (A) Schematic diagram of *Arid5a* gene locus and area of deletion. Approximate locations of guide RNA sites are indicated. (B) Genotyping validation of control littermates, heterozygous and homozygous *Arid5a*-knockout mice. (C) Western blotting of spleen, thymus and kidney tissues from *Arid5a*^{-/-} and *Arid5a*^{+/+} littermate controls. Top: *Arid5a*. Bottom: β -actin loading control.

Figure 2. Immunophenotyping of *Arid5a*^{-/-} mice. Spleen (A) or thymus (B) from *Arid5a*^{-/-} or *Arid5a*^{+/+} littermate controls were stained with the indicated cellular markers and analyzed by flow cytometry. Y-axis indicates total cell numbers. Each dot indicates one mouse. Bars indicate mean + SEM. No significant differences between groups were identified by Student's t-test.

Figure 3. *Arid5a*^{-/-} mice are resistant to EAE. The indicated mice (n=4) were subjected to EAE by injection of MOG peptide. (A) Clinical score was assessed daily by investigators blinded to sample identity. *** P<0.001 by ANOVA with Mann-Whitney U analysis. (B) Percent incidence of EAE in each group is indicated.

Figure 4. *Arid5a*^{-/-} mice show no alterations in susceptibility to mucosal or systemic candidiasis. (A) The indicated mice were infected orally with PBS (Sham) or *C. albicans* strain CAF2-1 for 75 min. After 5 days, tongues were assessed for CFU by plating. Dashed line = limit of detection. Each dot indicates one mouse. Bars indicate geometric mean. *P<0.05, ****P<.0001 by ANOVA with Kruskal-Wallis test (B) The mice in panel A were weighed daily and percent weight loss relative to day 0 is indicated. (C) The indicated mice were infected i.v. with *C. albicans* strain SC5314 and on day 3 fungal loads in kidney homogenates were assessed by plating. (D) The indicated mice were infected i.v. with *C. albicans* strain SC5314 and survival was monitored up to 15 days. Data were analyzed by log-rank (Mantel-Cox) test. *P<0.05 **P<0.001. ns, not significant.

A. Spleen

B. Thymus

Figure 3 (Taylor et al.)

A

B

A

B

C

D