An acetylation-mediated chromatin switch governs H3K4 methylation read-write capability

Kanishk Jain^{1,2}, Matthew R. Marunde³, Jonathan M. Burg³, Susan L. Gloor³, Faith M. Joseph⁴, Zachary B. Gillespie³, Keli L. Rodriguez³, Sarah A. Howard³, Irina K. Popova³, Nathan W. Hall³, Anup Vaidya³, Spencer W. Cooke¹, Kevin E. W. Namitz⁵, Bethany C. Taylor⁴, Ellen N. Weinzapfel³, Marcus A. Cheek³, Matthew J. Meiners³, Krzysztof Krajewski^{1,2}, Michael S. Cosgrove⁶, Nicolas L. Young⁴, Michael-Christopher Keogh^{3*}, and Brian D. Strahl^{1,2,7*}

¹Department of Biochemistry and Biophysics, University of North Carolina at Chapel Hill, School of Medicine, Chapel Hill, NC;

²Lineberger Comprehensive Cancer Center, University of North Carolina at Chapel Hill, School of Medicine, Chapel Hill, NC;

³EpiCypher, Inc., Durham, NC;

⁴Verna & Marrs McLean Department of Biochemistry and Molecular Biology, Baylor College of Medicine, Houston, TX;

⁵Pennsylvania State University, State College, PA;

⁶Department of Biochemistry and Molecular Biology, Upstate Medical University, Syracuse, NY

⁷Lead Contact

*Correspondence: mkeogh@epicypher.com and brian_strahl@med.unc.edu

ABSTRACT

In nucleosomes, histone N-terminal tails exist in dynamic equilibrium between free/accessible and collapsed/DNA-bound states. The DNA-bound state is expected to impact histone N-terminal availability to the epigenetic machinery. Notably, H3 tail acetylation (K9ac, K14ac, K18ac) is linked to increased engagement of H3K4me3 by the BPTF PHD finger, but it is unknown if this mechanism has broader extension. Here we show that *cis* H3 tail acetylation promotes nucleosomal accessibility to other H3K4 methyl readers, and further extends to H3K4 writers, notably methyltransferase MLL1. This regulation is nucleosome-dependent and also observed *in vivo*, where H3 acetylation correlates with increased levels of *cis* H3K4me. These observations reveal an acetylation ‘chromatin switch’ on the H3 N-terminal tail that modulates the accessibility and function of H3K4 methylation in chromatin. Our findings also resolve the long-standing question of why H3K4me3 levels are linked with H3 acetylation.

INTRODUCTION

In the epigenetic landscape, histone proteins are often variably chemically modified by “writer” enzymes (Jenuwein and Allis, 2001; Strahl and Allis, 2000). The writer-installed post-translational modifications (PTMs) are recognized by “reader” proteins and removed by “eraser” enzymes. This interplay of PTMs comprises the “histone code,” and it has a central function in regulating chromatin organization and activity. For example, methylated/acetylated lysine or methylated arginine residues of histones can recruit transcription factors to activate or repress transcription (Strahl and Allis, 2000; Su and Denu, 2016); mitotically phosphorylated serine/threonine residues can regulate reader binding established at earlier stages of the cell cycle (Rossetto et al., 2012); or ubiquitinated lysine can impact the maintenance of DNA methylation (Vaughan et al., 2021). As the complex language of histone PTMs is being dissected, it has become clear that multivalent interactions with reader proteins can influence chromatin structure and DNA accessibility, thereby regulating gene transcription and other DNA-templated events (Su and Denu, 2016; Taylor and

Young, 2021; Young et al., 2010). In this manner, combinatorial PTMs can more effectively engage different chromatin-binding modules, promoting distinct outcomes versus either PTM alone.

The bulk of chromatin PTM research has used peptides, even though histones exist *in vivo* in a heteromeric complex with DNA (i.e., the nucleosome). It is becoming increasingly clear that studying histone PTM engagement in the nucleosome context provides a more accurate understanding of the histone code. Particularly, the highly charged histone tails interact directly with nucleosomal DNA, restricting access for PTM recognition by reader proteins (Ghoneim et al., 2021; Marunde et al., 2022a; Morrison et al., 2018). Studies with BPTF PHD suggest acetylation releases the H3 N-terminal tail from the nucleosome surface, such that H3K4me3 becomes more readily engaged by the PHD finger (Marunde et al., 2022a; Morrison et al., 2018).

Considerable research effort has focused on dissecting the direct (and multivalent) engagement of chromatin via histone PTM-reader protein interactions. However, less appreciated are any indirect effects of PTMs on histone tail accessibility/nucleosome dynamics (e.g., via charge neutralization). In this report, we demonstrate enhanced nucleosome binding by a range of H3K4 readers when the histone tail is concomitantly acetylated (one or more of K9ac, K14ac, and K18ac). Furthermore, from *in vitro* enzymatic assays, we found that neighboring acetylation of the H3 tail was a prerequisite switch that enabled MLL1 complex (MLL1C) (Rao and Dou, 2015) to robustly methylate H3K4. Interestingly, mass spectrometric proteomic analyses of mammalian cells in a timed response to sodium butyrate (a broad-spectrum lysine deacetylase (KDAC) inhibitor) revealed a tight correlation of H3K4 methylation with *cis* acetylation. Our findings define a critical aspect of chromatin regulation: *i.e.*, crosstalk by histone tail accessibility. The findings also provide a molecular basis for the long-standing connection between H3K4 methylation and H3 acetylation (Garcia et al., 2007; Taverna et al., 2007), and resolve the directionality of these correlations: *cis* hyperacetylation of the H3 tail precedes, and is largely a prerequisite for, H3K4

methylation. Thus, the establishment of sites of H3K4me3 and activation of transcription occur by a sequence of modifications of the same histone molecule.

RESULTS

PHD finger readers show narrowed selectivity for histone tail PTMs on mononucleosomes versus peptides

How histone readers engage nucleosomes is an extensively researched area of chromatin biology. Most investigators have used PTM-defined histone peptides to characterize reader binding, although these often display a refined preference to similarly modified nucleosomes (Marunde et al., 2022a, 2022b; Morgan et al., 2021; Morrison et al., 2018). To further assess this potential, we used the dCypher[®] approach (Jain et al., 2020; Marunde et al., 2022b; Morgan et al., 2021; Weinberg et al., 2021) to measure the interactions of three PHD readers (from KDM7A, DIDO1, and MLL5: the queries) with PTM-defined peptides and nucleosomes (the potential targets). As expected (Jain et al., 2020), each GST-PHD fusion showed a preference for H3K4-methylated peptides and, particularly, to higher methyl states (*i.e.*, KDM7A bound me2/me3, DIDO1 bound me1/me2/me3, and MLL1 bound me1/me2/me3; **Figure 1A**). We further observed no impact of co-incident acetylation on H3K4me3 binding in the peptide context (compare H3K4me3 to H3K4me3K9acK14acK18ac [hereafter H3K4me3tri^{ac}]). However on nucleosomes each GST-PHD showed reduced and restricted binding to H3K4me3 over the lower methyl states, and this interaction was dramatically enhanced (~10-15-fold) by co-incident acetylation (as at H3K4me3tri^{ac}; **Figure 1B**). The notable impact of acetylation state on binding of these readers in the nucleosome context (also recently observed for the BPTF PHD domain (Marunde et al., 2022a; Morrison et al., 2018)) led us to consider additional implications of this interplay.

Histone tail lysine acetylation (Kac) can directly recruit residue-specific readers, *e.g.*, bromodomains (Musselman et al., 2012), but acetylation also neutralizes the positive charge on lysine residues and relieves their interaction with negatively charged DNA (*i.e.*, altering the

histone tail-DNA binding equilibrium) (Marunde et al., 2022a; Morrison et al., 2018). In the nucleosome context, this decreased histone-DNA binding supports the increased engagement of reader domains that have no direct affinity for the Kac. Conversely, isolated histone tail peptides are 'constitutively open' (no DNA to engage), and thus not subject to this mode of regulation. We surmised that the effect of charge neutralization to increase H3K4me3 accessibility could have application beyond reader domains, perhaps also applying to enzymes that target H3K4.

H3 N-terminal acetylation significantly enhances MLL1C methylation of nucleosomal H3K4

To investigate if acetylation makes the H3 N-terminus more accessible for modification, we performed enzymatic assays with the MLL1 core complex (MLL1C) responsible for H3K4me3 (Rao and Dou, 2015; Sha et al., 2020) and nucleosome substrates $\pm$ accompanying polyacetylation (H3K9acK14acK18ac; hereafter H3tri^{ac}) (see **Methods**). In an endpoint assay at constant enzyme and substrate concentrations (and [*methyl*-³H]-SAM donor), we observed a significant increase in net methylation when the H3 tail was also acetylated (**Figure 2A**). As expected, methylation by MLL1C sequentially decreased towards H3K4 mono-, di-, and trimethylated nucleosomal substrates, being undetectable on H3K4me3 (which also confirmed MLL1C targeting of this specific residue). Despite this, methyl group incorporation to each H3K4 methyl state substrate was consistently enhanced by accompanying H3tri^{ac} (**Figure 2A**). Of note, we further tested the viability of synthetic methyllysine analogs (MLAs) (Simon and Shokat, 2012) $\pm$ H3tri^{ac} as MLL1C substrates and observed no activity, indicating the unsuitability of MLAs for such studies (**Figure S2D**).

We also measured the steady-state kinetics of MLL1C methylation of nucleosomes with each H3K4 methyl state $\pm$ H3tri^{ac}, and again observed that acetylation increased methyltransferase activity (**Figure 2B**). Using an extra sum-of-squares F-test, methylation data for the H3tri^{ac} nucleosomes were indicative of positive cooperativity because of better fit ($p = 0.0216$) to the Hill equation (Weiss, 1997) (**Figure 2B v. Figure S2C; Table 1 v. Table S1**).

Because of the low level of enzymatic activity towards the unacetylated nucleosomes we could not make a statistically significant comparison between the Hill and Michaelis-Menten fits. There have been limited studies of MLL1C activity on nucleosomes (Park et al., 2019; Patel et al., 2011; Xue et al., 2019), so an overlooked potential allostery is understandable given the complex possible interactions between this enzyme and substrate (Lee et al., 2021; Park et al., 2019)

Interestingly, although overall k_{cat} was ~17-fold greater for the H3tri^{ac} nucleosomes, the $K_{0.5}$ values (substrate concentration at half-maximal velocity/half-saturation for an allosterically regulated enzyme) were indistinguishable (**Table 1**). Therefore, although MLL1C catalytic efficiency toward H3tri^{ac} nucleosomes was enhanced by an increase in k_{cat} , this catalytic efficiency was not driven by $K_{0.5}$, which suggested no increase in relative binding affinity. To further examine this, we used dCypher to examine potential binding between MLL1C query and a selection of nucleosome targets: unmodified, H3N Δ 32 (lacking the first 32 residues of H3), Tailless (trypsin-digested nucleosomes to remove N- and C-terminal histone tails), and H3tri^{ac}. At 50 mM NaCl, we observed no compelling difference in MLL1C binding between any of these nucleosomes (**Figure 2C**). This finding agreed with structural studies where binding between MLL1C and the nucleosome occurs primarily through interactions with nucleosomal DNA and, to a lesser degree, the H4 N-terminal tail (Lee et al., 2021; Park et al., 2019). Thus, the increased H3K4 methylation observed when the H3 N-terminal tail was acetylated was not due to enhanced MLL1C – nucleosome binding. Instead, H3 acetylation likely released the histone tail from the nucleosome, thereby increasing the apparent H3K4 concentration and enhancing methylation (**Figure 2D**).

Cellular level of H3K4 methylation is coupled to H3 N-terminal tail hyperacetylation

The above data suggested a molecular model for how the H3 N-terminal tail, via *cis* acetylation, becomes available for H3K4 reader binding or enzymatic modification *in vitro*. To determine if such acetylation could function as an accessibility switch *in vivo*, we used middle-down mass spectrometry on acid-extracted histones from asynchronous MCF-7 breast cancer cells to

measure the relationship between H3K4 methylation and tail acetylation on the same H3 proteoforms (Holt et al., 2021; Smith and Kelleher, 2013). As expected (Garcia et al., 2007; Peach et al., 2012; Young et al., 2009), the absolute amounts of H3K4me3 and higher Kac states (3ac, 4ac, and 5ac) across adjacent lysine residues were extremely low (**Figure S3A**). Nonetheless, H3K4me3 (<1% of total H3) was strictly associated with molecules that contained multiple acetylations (also <1% of total H3; **Figure 3A**). Given this relationship, we next addressed the hypothesis that increased lysine acetylation may release the H3 tail for more effective H3K4 methylation (*i.e.*, acetylation precedes methylation). We treated MCF-7 cells with the KDAC inhibitor sodium butyrate and collected samples at multiple timepoints to measure the levels of H3K4methylation with *cis* acetylation. H3 poly-acetylation rapidly increased upon butyrate treatment (**Figure S3B** and **Table S2**), as would be predicted (Holt et al., 2019; Young et al., 2009). While H3K4me3 levels most dramatically increased in tandem with the 5ac H3 proteoform (**Figure 3B**), we observed an associated increase in all H3K4 methyl states as the H3 tail was increasingly acetylated (**Figures 3C** and **S3C**; **Table S2**). These findings support a direct link whereby acetylation releases nucleosome-bound H3 tails to available H3K4 methyltransferases for subsequent methylation (see model in **Figure 2D**). Such a chromatin-switch may also help cells to translate short-term acetylation signals at gene promoters to longer-term heritable marks of epigenetic memory (Greer et al., 2014; Hörmanseder et al., 2017; Muramoto et al., 2010; Ng et al., 2003).

DISCUSSION

While previous investigations identified a link between H3K4 methylation and H3 acetylation in diverse species (Garcia et al., 2007; Strahl et al., 1999; Taverna et al., 2007; Young et al., 2009), the molecular basis for this link was unknown, and we posit the H3 acetyl ‘chromatin switch’ is conserved across eukaryotes. Our new understanding of the dynamic structure of nucleosome histone tails, alternating between collapsed (*i.e.*, nucleosome-bound) and accessible forms

(Marunde et al., 2022a; Morrison et al., 2018), has made more plausible the notion that tail availability could be driven by combinatorial *cis* acetylation to directly promote H3K4 methylation. In this study, we showed that hyperacetylation of the H3 N-terminal tail promoted rapid accumulation of H3K4 methylation in *cis*, most likely by increasing availability of the substrate residue to the MLL1C active site. This finding was supported by our *in vitro* enzymatic and *in vivo* mass spectrometric analyses. Methylation assays with MLL1C revealed significantly enhanced enzyme activity towards nucleosome substrates with co-incident acetylated (H3tri^{ac}) over unmodified H3 (**Figure 2A-B; Table 1**). dCypher assays demonstrated that the acetylation-mediated increase in H3K4 methylation does not involve stabilized interactions between MLL1C and the nucleosome (**Figure 2C**). In agreement with these *in vitro* findings, middle-down mass spectrometry showed that H3 hyperacetylation and H3K4 methylation co-occurred on the same histone tails in actively cycling cells; furthermore, upon butyrate treatment H3K4 methylation increased for the most highly acetylated proteoforms (**Figures 3 & S3**). The *in vivo* relationship required higher degrees of acetylation (preferring at least four acetyl groups per molecule: e.g., H3K9acK14acK18acK23acK27ac) for most effective conversion to the H3K4me3 state. Together with binding studies that identify the positive impact of *cis* H3 tail hyperacetylation on H3K4 reader engagement (**Figures 1 & S1**) (Marunde et al., 2022a; Morrison et al., 2018), our findings suggest a molecular switch that governs when and where the histone H3 N-terminus is available for H3K4-related transactions. Such a switch could be used to establish and heritably maintain the location and function of transcriptional promoters across the genome.

A continued observation from this study is that the binding preference of readers (in this case PHD-fingers that engage histone H3K4) are narrowed on nucleosomes relative to histone peptides (**Figure 1 & S1**) (Marunde et al., 2022a; Morgan et al., 2021). This difference in binding further highlights the importance of using nucleosomes as physiologically relevant substrates for *in vitro* interaction assays. We also add to the literature confirming importance of the physiological substrate for enzymatic studies (Strelow et al., 2016; Stützer et al., 2016). In this regard our

steady-state kinetic methylation assays with MLL1C revealed an intriguing (and previously undescribed) positive cooperativity with its preferred nucleosome substrates (**Figure 2B** versus **S2C**). It is important to consider the suggestion that allosteric factors could regulate interactions between MLL1C and the nucleosome, especially since previous kinetic analyses of this [enzyme:substrate] pair did not address such behavior (Park et al., 2019; Xue et al., 2019). However, positive cooperativity was not evident at the level of MLL1C - nucleosome binding, which was independent of substrate identity (*i.e.*, $\pm$ H3tri^{ac}) at ionic conditions similar to the catalytic assay (**Figure 2C**). Still to be determined are the signals that drive and regulate this cooperativity and its function in MLL1-catalyzed methylation, especially in a higher-order chromatin context.

Our study highlights a previously unrecognized regulatory mechanism for how writers might engage the histone H3 tail (specifically at H3K4) *in vivo*. Although this work focused on H3, it will be important to ascertain the consequence of acetylation (or indeed other acylation)-mediated changes in accessibility and the function of modifiers and readers of the other core histones (H2A, H2B, and H4). A more detailed analysis of how to regulate histone tail accessibility (in *cis* and *trans*) will likely add to the already complex language of the histone code.

Epigenetic studies have identified H3K4 methylation and H3 acetylation as active marks because of their co-occupancy on promoters and gene bodies of transcribed genes (Santos-Rosa et al., 2002; Strahl et al., 1999; Wozniak and Strahl, 2014). Our findings agree with these observations regarding H3K4 methylation, but it will be interesting to determine whether PTMs such as H3K9 methylation, normally (but not exclusively (Barral et al., 2022; Mauser et al., 2017)) a repressive mark, show similarly enhanced histone accessibility due to *cis* acetylation.

ACKNOWLEDGEMENTS

KJ is supported by a Postdoctoral Training Fellowship from the National Institutes of Health (NIH; T32CA217824) to the UNC Lineberger Cancer Center and a Postdoctoral Fellowship from the American Cancer Society (PF-20-149-01-DMC). This work was also supported by NIH grants to NLY (R01GM139295, P01AG066606, and R01CA193235), MSC (R01CA140522), to EpiCypher (R44GM117683, R44CA214076 and R44GM116584), and to BDS (R35GM126900). We thank colleagues for the generous supply of materials (see **Methods**) and members of the Cosgrove, Young, *EpiCypher* and Strahl labs for helpful discussions and suggestions.

AUTHOR CONTRIBUTIONS

KJ, MRM, JMB, MCK, and BDS conceptualized and designed the study. ZBG, SAH, MAC, and MJM assembled and validated PTM-defined dNucs and versaNucs. KK synthesized histone peptides used in versaNuc construction. MM, JB, SLG, KLR, IKP, NWH, AV, and ENW performed dCypher assays and analyzed data with input from KJ, MCK, and BDS. KEWN and MSC purified MLL1C for enzymatic analyses. KJ and SWC purified his-tagged MLL1C for binding assays. Methylation assays were performed and analyzed by KJ and SWC. MCF7 cells were cultured by KJ, and mass spectrometry was performed and analyzed by FJ, BCT, and NY. KJ and BDS wrote the first draft of the manuscript and all authors contributed to subsequent editing.

DECLARATIONS OF INTEREST

MSC owns stock/serves on the Consultant Advisory Board for Kathera Bioscience Inc. and holds US patents (8,133,690; 8,715,678; and 10,392,423) for compounds/methods for inhibiting SET1/MLL family complexes. EpiCypher is a commercial developer and supplier of reagents (e.g., PTM-defined semi-synthetic nucleosomes; dNucsTM and versaNucs[®]) and platforms (e.g., dCypher[®]) used in this study. MCK and BDS are board members of EpiCypher.

FIGURES AND TABLES

Figure 1. PHD finger reader domains show restricted binding on PTM-defined peptides vs. nucleosomes. *dCypher* assay alpha counts for interaction of GST-PHD queries (9.5 nM KDM7A (Uniprot #Q6ZMT4; residues 1-100); 2.4 nM DIDO1 (Uniprot #Q9BTC0; residues 250-340); 18 nM MLL5 (Uniprot #Q8IZD2; residues 100-180)) with PTM-defined peptides (**A**) vs. nucleosomes (**B**) (the potential targets). All error bars represent the range of two replicates. Key: H3.1N Δ2, H3.1N Δ32 and H4N Δ15 are nucleosomes assembled with histones lacking the indicated N-terminal residues of H3.1 or H4. All data was plotted using GraphPad Prism 9.0.

Figure 2. In the nucleosome context MLL1C methylation activity on H3K4 is significantly enhanced by co-incident H3tri^{ac}. **A)** Endpoint methylation assays of H3K4me0 (unmodified)-me1-me2-me3 nucleosomes and their cognate H3K9ac14ac18ac (H3tri^{ac}) partners (all 100 nM) with MLL1C (4nM). Reactions performed in triplicate with error bars as S.D. *p*-values were determined using a two-tailed t test: **** = <0.0001, *** = 0.0008, ** = 0.0038. **B)** MLL1 complex (4 nM) methylation activity on H3K4me0-me1-me2-me3 nucleosomes and their cognate H3tri^{ac} partners (all substrates: 1.5-fold serial dilution, 0-0.4 μ M). Reactions performed in triplicate with error bars as S.D. **C)** dCypher binding curves of 6HIS-tagged MLL1C (concentration as noted) with PTM-defined nucleosomes (20 nM). Error bars represent the range of two replicates. **D)** Graphical model for how H3 N-terminal tail acetylation may regulate H3K4 methylation by MLL1C.

271 Nucleosomal DNA is represented in black; each histone is as labeled, with the core histone N-
272 terminal tails colored to distinguish. MLL1C is in purple.

Figure 3. Middle-down MS analysis reveals a hierarchical dependence between H3K4 methylation and *cis* H3 acetylation in MCF-7 cells. A) Occupancy of H3K4 methyl states (me0-1-2-3) within each H3 acetyl state (0ac to 5ac) in asynchronous MCF-7 cells. B) Time course of H3K4me3 accumulation with respect to each H3 acetyl state after butyrate treatment. C) Occupancy of H3K4 methyl states within each H3 acetyl state after butyrate treatment (60 mins).

280 Asterisks represent p -values < 0.05 . All data were collected in biological triplicate with error bars
281 representing S.E.M.

TABLE 1: Steady-state Hill kinetic parameters

Substrate	$K_{0.5}$ (μM)	k_{cat} (min^{-1})	h (Hill coefficient)	R^2
H3 unmodified	0.13 ± 0.06	0.0018 ± 0.0004	1.213 ± 0.3430	0.8810
H3 K9/14/18Ac	0.12 ± 0.02	0.0304 ± 0.0036	1.741 ± 0.3528	0.9228
H3 K4me1	n.d. ^a	n.d. ^a	n.d. ^a	0.3186
H3 K4me1 K9/14/18Ac	0.14 ± 0.008	0.0092 ± 0.0004	2.032 ± 0.1333	0.9920
H3 K4me2	n.d. ^a	n.d. ^a	n.d. ^a	-0.1746
H3 K4me2 K9/14/18Ac	0.042 ± 0.005	0.0005 ± 0.00004	5.349 ± 2.682	0.8155
H3 K4me3	n.d. ^a	n.d. ^a	n.d. ^a	n.d. ^a
H3 K4me3 K9/14/18Ac	n.d. ^a	n.d. ^a	n.d. ^a	n.d. ^a

^a = methylation signal was indistinguishable from background: kinetic parameters could not be determined.

METHODS

Expression and purification of GST-tagged PHD reader proteins

Recombinant expression constructs for GST-tagged PHD domains from KDM7A (Uniprot #Q6ZMT4; residues 1-100), DIDO1 (Uniprot #Q9BTC0; residues 250-340), and MLL5 (Uniprot #Q8IZD2; residues 100-180) were synthesized by BioMatik Corporation in a pGEX-4T-1 vector and provided by Dr. Mark T. Bedford (UT MD Anderson Cancer Center). These proteins were expressed and purified as described (Jain et al., 2020).

Expression, purification, and assembly of the MLL1 core complex (MLL1C)

Methods for the expression, purification, and assembly of MLL1 core complex (MLL1C) were adapted from published protocols (Usher et al., 2021). A polycistronic recombinant expression construct containing the MLL1 SET domain (Uniprot Q03164; residues 3745-3969), WDR5 (Uniprot P61964; residues 2-334), RbBP5 (Uniprot Q15291; residues 1-538) and Ash2L (Uniprot Q9UBL3-3; residues 1-534) (MWRA construct) in pST44 vector was a kind gift from Dr. Song Tan (Tan et al., 2005). WDR5 was cloned with an N-terminal hexahistidine (6HIS) tag and TEV protease site to enable purification via immobilized metal affinity chromatography (IMAC), and enzymatic cleavage for tag removal. Rosetta pLysS *E. coli* cells were transformed with the plasmid and grown on LB plates with 50 µg/mL carbenicillin. Single colonies were used to inoculate 50 mL starter cultures of Terrific Broth (TB) containing 50 µg/mL carbenicillin and grown at 37°C for 16 hours. This culture was transferred to 1L of TB + carbenicillin and grown to OD₆₀₀ ~0.6 (37°C, 200 RPM, ~ 4 hours). Recombinant protein expression was induced with 1 mM Isopropyl β-D-1-thiogalactopyranoside (*Sigma*; 16°C, 200 RPM, 20 hours). Cells were harvested by centrifugation (5000 RPM, 4°C) and flash frozen in liquid nitrogen.

Frozen cell pellets were resuspended in 50 mL of lysis buffer (50 mM Tris-HCl pH 7.5, 300 mM NaCl, 30 mM imidazole, 3 mM dithiothreitol, and 1 µM ZnCl₂) containing 0.5 mg/mL lysozyme

(*Sigma*), 250 U Pierce Universal Nuclease (*ThermoFisher*), and an EDTA-free protease inhibitor cocktail (*Roche*) and rotated at 4°C for 1 hour. The resultant mixture was then sonicated [five cycles of 30 sec on/30 sec off at 50% output] and centrifuged at 4°C, 15,000 RPM for 35 min. The clarified lysate was flowed over a 5 mL HisTrap nickel column (*Cytiva*) using an AKTA Pure FPLC (*Cytiva*) at 0.5 mL/min; all FPLC steps were conducted at 4°C. Unbound molecules were removed with 20 column volumes of wash buffer (WB: 50 mM Tris-HCl, pH 7.5, 300 mM NaCl, 30 mM imidazole, 3 mM DTT, and 1 μ M ZnCl₂ at 2 mL/min). The 6HIS-tagged MWRA was eluted in a 15-column volume linear gradient from WB to Elution Buffer (WB + 500 mM imidazole) at 2 mL/min. Fractions containing 6HIS-tagged MWRA were identified by SDS-PAGE, pooled, and supplemented with 6HIS-tagged TEV protease (purified as described: Nautiyal and Kuroda, 2018) at a 1:100 enzyme to substrate molar ratio to cleave the 6HIS-tag on WDR5. This mixture was dialyzed against three changes of WB (each 2L for at least four hours at 4°C) and 6HIS-TEV removed from cleaved MWRA via IMAC (Usher et al., 2021). MWRA flow-through protein solution was concentrated to ~15 mL using a 30 kDa MWCO centrifugal filter (*EMD Millipore*), ensuring not to concentrate to where solution became yellow/cloudy and viscous. MWRA complex was resolved over a HiLoad 16/60 Superdex 200 pg gel filtration (GF) column (*Cytiva*) pre-equilibrated in 20 mM Tris-HCl, pH 7.5, 300 mM NaCl, 1 mM TCEP, and 1 μ M ZnCl₂. Fractions containing stoichiometric MWRA sub-complex were identified by SDS-PAGE, pooled, and concentrated to ~15 mL.

HisDPY30 was expressed, purified, and cleaved to remove the 6HIS-tag as described (Patel et al., 2009; Usher et al., 2021). A two-fold molar excess of DPY30 was added to the MWRA sub-complex and incubated on ice for 1 hour. Following incubation, MLL1C was isolated by gel filtration as described for the MWRA complex, with fractions containing the stoichiometric complex identified by SDS-PAGE, pooled, concentrated to ~10 μ M, and flash frozen. For dCypher experiments with MLL1C, the 6HIS-tag was retained on DPY30.

PTM-defined nucleosomes

All nucleosomes were from the dNucTM or versaNuc[®] portfolios (*EpiCypher*). PTMs were confirmed by mass-spectrometry and immunoblotting (if an antibody was available) (Goswami et al., 2021; Marunde et al., 2022b; Weinberg et al., 2019).

dCypher assays

dCypher binding assays with PTM-defined nucleosomes were performed under standard conditions that titrate query (e.g., epitope-tagged reader domain(s)) to a fixed concentration of target (e.g., biotinylated PTM-defined nucleosome) with the appropriate Alpha Donor and Acceptor beads (*Perkin Elmer*) (Jain et al., 2020; Marunde et al., 2022b; Weinberg et al., 2019). Binding curves [query:target] were generated using a non-linear 4PL curve fit in Prism 9.0 (*GraphPad*). For each query, the relative EC₅₀ (EC₅₀^{rel}) and hillslope values were derived from the best binding target. EC₅₀^{rel} is the half maximal signal for the specified target. Where necessary, we excluded query concentration values determined to be beyond a query's hook point (signal inhibition due to query exceeding bead saturation). The EC₈₀^{rel} was selected as the optimal probing concentration for discovery screens because of the robust signal-to-background and to provide the best opportunity to bind targets without saturating the primary target signal. To compute EC₈₀^{rel} values, we used the formula $EC_F^{rel} = (F / (100 - F))^{1/H} \times EC_{50}^{rel}$; F = 80 and H = hillslope.

Briefly, 5 µL of GST-tagged reader domain was incubated with 5 µL of 10 nM biotinylated nucleosomes (e.g., *EpiCypher* #16-9001) for 30 minutes at room temperature in 20 mM HEPES pH 7.5, 250 mM NaCl, 0.01% BSA, 0.01% NP-40, 1 mM DTT in a 384-well plate. A mix of 10 µL of 2.5 µg/mL glutathione acceptor beads (*PerkinElmer*, AL109M) and 5 µg/mL streptavidin donor beads (*PerkinElmer*, 6760002) was prepared in 20 mM HEPES pH 7.5, 250 mM NaCl, 0.01% BSA, 0.01% NP-40 and added to each well. The plate was incubated at room temperature in

subdued lighting for 60 minutes, and AlphaLISA signal was measured on a PerkinElmer 2104 EnVision (680 nm laser excitation, 570 nm emission filter $\pm$ 50 nm bandwidth). Each binding interaction was performed in duplicate in a 20 μ L mix in 384 well plates.

MLL1C binding assays (**Figure 2C**) were performed as described above except using Nickel-chelate acceptor beads (10 μ g/mL; *Perkin Elmer* AL108M), streptavidin donor beads (20 μ g/mL; *Perkin Elmer*) and modified assay buffer (20 mM Tris pH 7.5 + 50 mM NaCl, 0.01% BSA, 0.01% NP-40 and 1 mM DTT); [NaCl] was optimized via a titration assay and 50 mM chosen for subsequent analyses.

In vitro methylation assays

Methylation assays (Shinsky et al., 2015) were performed at 15°C for three hours using purified MLL1C enzyme and nucleosome substrate in a reaction volume of 20 μ L [in 50 mM HEPES, pH 8.0, 1 mM DTT, 1 μ M ZnCl₂; 10 μ M of 9:1 S-adenosyl-L-methionine (SAM) *p*-toluenesulfonate salt (Sigma) to S-adenosyl-L-[methyl-³H]-methionine ([methyl-³H]-SAM) (*PerkinElmer*)]. Concentrations of MLL1C and NaCl were optimized by performing 2D-titration methylation assays at [0, 4 and 40 nM MLL1C] and [0, 50 and 300 mM NaCl] with 2 μ g of chicken oligo-nucleosome substrate (Morris et al., 2007). It is notable that, across a range of concentrations, MLL1C stability decreases as temperature increases and methyltransferase activity has been reported to be enhanced in sub-physiological NaCl concentrations (Namitz et al., 2019; Shinsky et al., 2015). For endpoint methylation assays, 4 nM MLL1C and 100 nM nucleosome substrates were tested as above. For steady-state kinetics, 4 nM MLL1C was incubated with a nucleosome substrate titration (0, 23.4, 35.1, 52.7, 79, 119, 178, 267 and 400 nM) and reactions quenched with 5 μ L of 5X SDS loading dye. For steady-state kinetic assays, 0.5 μ g bovine serum albumin was added to each reaction after quenching to act as a loading guide. To analyze methylation, quenched reactions were resolved by 15% Tris-Glycine SDS-PAGE. Gels were stained with Coomassie dye and bands containing mononucleosomes were excised (with serum albumin as a supporting lane

marker) and incubated in a solution of 50% Solvable (*PerkinElmer*) and 50% water at 50°C for three hours. Mixture and gel slices were then combined with 10 mL of Hionic-Fluor scintillation fluid (*PerkinElmer*), dark-adapted overnight, and radioactivity measured on a Liquid Scintillation Counter (*Beckman Coulter*).

Middle-down mass spectrometry of MCF-7 cells ± KDAC inhibition

MCF-7 breast cancer cells (ATCC HTB-22) were grown in MEM (*Gibco*) supplemented with 10% fetal bovine serum (*VWR*), 100 I.U. penicillin, 100 µg/mL streptomycin (*Corning*), 0.01 mg/mL human recombinant insulin (*Gibco*), and 5 µg/mL plasmocin (*Invivogen*) at 37°C and 5% CO₂.

For mass spectrometric (MS) analysis ± KDAC inhibition, cells were cultured in 150 mm dishes to ~80% confluence and treated with 5 mM sodium butyrate (or equivalent volume of water) in triplicate for 0, 10, 20, 30, 60, 120 mins. Cells were washed with cold PBS (11.9 mM phosphates, 137 mM NaCl, 2.7 mM KCl) to remove residual sodium butyrate, harvested by scraping, and flash frozen in liquid nitrogen. Histones were acid extracted after nuclei isolation as described (Holt et al., 2021). Isolated histones were resuspended in 85 µL 5% acetonitrile, 0.2% trifluoroacetic acid (TFA) and resolved by offline high-performance liquid chromatography (HPLC) as described (Holt et al., 2021). Reverse Phase HPLC fractionation was performed with a Thermo U3000 HPLC system (*ThermoFisher*) with a 150 × 2.1-mm Vydac 218TP 3 µm C18 column (*HiChrom* # 218TP3215), at a flowrate of 0.2 mL/min using a linear gradient from 25% B to 60% B in 60 min. The composition of buffers used were A: 5% acetonitrile and 0.2% TFA and B: 95% acetonitrile and 0.188% TFA. After chromatographic separation and fraction collection, histone H3.1 was selected, diluted in 100 mM ammonium acetate (pH = 4) and digested with Glu-C protease (*Roche*) at 10:1 protein:enzyme for 1 h at room temperature prior to mass spectrometric analysis.

The digested samples were diluted to 2 µg/µL. Online HPLC was performed on a Thermo U3000 RSLC nano Pro-flow system using a C3 column (Zorbax 300SB-C3, 5 µm; *Agilent*). Samples were maintained at 4 °C and 1 µL injected for each analysis using a microliter pickup. A linear 70-minute gradient of 4-15% B was used (Buffer A: 2% acetonitrile, 0.1% formic acid and Buffer B: 98% acetonitrile and 0.1% formic acid) with a flow rate of 0.2 µL/min. The column eluant was introduced into an Orbitrap Fusion Lumos mass spectrometer (*ThermoFisher*) by nano-electrospray ionization. A static spray voltage of 1900 V and an ion transfer tube temperature of 320 °C were set for the source.

A Fusion Lumos mass spectrometer was used to generate MS the data. The 9th charge state of histone H3 was targeted for analysis. MS1 analysis was acquired in the orbitrap with a normal mass range and a 60k resolution setting in positive ion mode. An Automatic Gain Control (AGC) target of 5.0E5 with a 200 ms maximum injection time, three microscans, and a scan range of 585-640 *m/z* were used to identify desired ions for fragmentation. MS2 acquisition was performed in both orbitrap and ion trap mode. Both modes used electron transfer dissociation (ETD), a reaction time of 18 ms, and an injection time of 200 ms. A normal scan range was used for the orbitrap mode with a resolution setting of 30k and an AGC target of 5.0E5, with two microscans. A narrow scan range of 470-530 *m/z*, targeting ions indicative of K4 modification states, was used for the ion trap mode MS2 with an ACG target of 3.0E4, quadrupole isolation, maximum injection time of 100 ms, and eight microscans.

These MS methods were used with two technical replicates per biological replicate (n=3). An MS ion trap mode with a targeted mass list was used to increase sensitivity to identify known low abundance K4me3-containing proteoforms. The ion trap MS2 spectra were averaged for each H3 acetyl state based on known retention times, and the intensities of ions indicative of the K4 methylation states were manually recorded. Retention times for each acetyl state were approximated as 0ac 35-40 min, 1ac 45-55 min, 2ac 55-60 min, 3ac 62-68 min, 4ac 69-73 min, and 5ac 74-78 min. Precursor mass was used as an additional confirmation and filter of correct

439 acetyl state. For scans yielding low signal and high noise (*i.e.*, 5ac at 0 min butyrate treatment),
 440 data were manually curated before averaging. Within acetyl states, the relative proportion of
 441 fragment ions for unmodified, mono-, di-, and trimethylation of the H3K4 ion at respective *m/z* of
 442 474, 488, 502, and 516 were recorded per MS run. Values were averaged across replicates of
 443 the same conditions and normalized to one hundred percent. A two-tailed t-test was used for
 444 significance. Raw MS data will be available in a public repository.

REFERENCES

1. Barral, A., Pozo, G., Ducrot, L., Papadopoulos, G.L., Sauzet, S., Oldfield, A.J., Cavalli, G., and Déjardin, J. (2022). SETDB1/NSD-dependent H3K9me3/H3K36me3 dual heterochromatin maintains gene expression profiles by bookmarking poised enhancers. *Molecular Cell* 82, 816-832.e12.
2. Garcia, B.A., Pesavento, J.J., Mizzen, C.A., and Kelleher, N.L. (2007). Pervasive combinatorial modification of histone H3 in human cells. *Nature Methods* 4, 487–489.
3. Ghoneim, M., Fuchs, H.A., and Musselman, C.A. (2021). Histone Tail Conformations: A Fuzzy Affair with DNA. *Trends in Biochemical Sciences* 46, 564–578.
4. Goswami, J., MacArthur, T., Bailey, K., Spears, G., Kozar, R.A., Auton, M., Dong, J.F., Key, N.S., Heller, S., Loomis, E., et al. (2021). Neutrophil Extracellular Trap Formation and Syndecan-1 Shedding Are Increased After Trauma. *Shock (Augusta, Ga.)* 56, 433–439.
5. Greer, E.L., Beese-Sims, S.E., Brookes, E., Spadafora, R., Zhu, Y., Rothbart, S.B., Aristizábal-Corrales, D., Chen, S., Badeaux, A.I., Jin, Q., et al. (2014). A Histone Methylation Network Regulates Transgenerational Epigenetic Memory in *C. elegans*. *Cell Reports* 7, 113–126.
6. Holt, M. v., Wang, T., and Young, N.L. (2019). High-Throughput Quantitative Top-Down Proteomics: Histone H4. *Journal of the American Society for Mass Spectrometry* 30, 2548–2560.
7. Holt, M. v, Wang, T., and Young, N. (2021). Expeditious Extraction of Histones from Limited Cells or Tissue Samples and Quantitative Top-Down Proteomic Analysis. *Current Protocols* 1.
8. Hörmanseder, E., Simeone, A., Allen, G.E., Bradshaw, C.R., Figlmüller, M., Gurdon, J., and Jullien, J. (2017). H3K4 Methylation-Dependent Memory of Somatic Cell Identity

- Inhibits Reprogramming and Development of Nuclear Transfer Embryos. *Cell Stem Cell* 21, 135-143.e6.
9. Jain, K., Fraser, C.S., Marunde, M.R., Parker, M.M., Sagum, C., Burg, J.M., Hall, N., Popova, I.K., Rodriguez, K.L., Vaidya, A., et al. (2020). Characterization of the plant homeodomain (PHD) reader family for their histone tail interactions. *Epigenetics & Chromatin* 2020 13:1 13, 1–11.
10. Jenuwein, T., and Allis, C.D. (2001). Translating the Histone Code. *Science* 293, 1074–1080.
11. Lee, Y.T., Ayoub, A., Park, S.H., Sha, L., Xu, J., Mao, F., Zheng, W., Zhang, Y., Cho, U.S., and Dou, Y. (2021). Mechanism for DPY30 and ASH2L intrinsically disordered regions to modulate the MLL/SET1 activity on chromatin. *Nature Communications* 12.
12. Marunde, M.R., Fuchs, H.A., Burg, J.M., Popova, I.K., Vaidya, A., Hall, N.W., Meiners, M.J., Watson, R., Howard, S.A., Novitzky, K., et al. (2022a). Nucleosome conformation dictates the histone code. *BioRxiv* 2022.02.21.481373.
13. Marunde, M.R., Popova, I.K., Weinzapfel, E.N., and Keogh, M.-C. (2022b). The dCypher Approach to Interrogate Chromatin Reader Activity Against Posttranslational Modification-Defined Histone Peptides and Nucleosomes. 231–255.
14. Mauser, R., Kungulovski, G., Keup, C., Reinhardt, R., and Jeltsch, A. (2017). Application of dual reading domains as novel reagents in chromatin biology reveals a new H3K9me3 and H3K36me2/3 bivalent chromatin state. *Epigenetics and Chromatin* 10, 1–19.
15. Morgan, M.A.J., Popova, I.K., Vaidya, A., Burg, J.M., Marunde, M.R., Rendleman, E.J., Dumar, Z.J., Watson, R., Meiners, M.J., Howard, S.A., et al. (2021). A trivalent nucleosome interaction by PHIP/BRWD2 is disrupted in neurodevelopmental disorders and cancer. *Genes & Development*.
16. Morris, S.A., Rao, B., Garcia, B.A., Hake, S.B., Diaz, R.L., Shabanowitz, J., Hunt, D.F., Allis, C.D., Lieb, J.D., and Strahl, B.D. (2007). Identification of histone H3 lysine 36

- acetylation as a highly conserved histone modification. *Journal of Biological Chemistry* 282, 7632–7640.
17. Morrison, E.A., Bowerman, S., Sylvers, K.L., Wereszczynski, J., and Musselman, C.A. (2018). The conformation of the histone H3 tail inhibits association of the BPTF PHD finger with the nucleosome. *ELife* 7.
18. Muramoto, T., Müller, I., Thomas, G., Melvin, A., and Chubb, J.R. (2010). Methylation of H3K4 Is Required for Inheritance of Active Transcriptional States. *Current Biology* 20, 397–406.
19. Musselman, C.A., Lalonde, M.-E., Côté, J., and Kutateladze, T.G. (2012). Perceiving the epigenetic landscape through histone readers. *Nature Structural & Molecular Biology* 2012 19:12 19, 1218–1227.
20. Namitz, K.E.W., Tan, S., and Cosgrove, M.S. (2019). Hierarchical assembly of the MLL1 core complex within a biomolecular condensate regulates H3K4 methylation. *BioRxiv* 870667.
21. Nautiyal, K., and Kuroda, Y. (2018). A SEP tag enhances the expression, solubility and yield of recombinant TEV protease without altering its activity. *New Biotechnology* 42, 77–84.
22. Ng, H.H., Robert, F., Young, R.A., and Struhl, K. (2003). Targeted Recruitment of Set1 Histone Methylase by Elongating Pol II Provides a Localized Mark and Memory of Recent Transcriptional Activity. *Molecular Cell* 11, 709–719.
23. Park, S.H., Ayoub, A., Lee, Y.T., Xu, J., Kim, H., Zheng, W., Zhang, B., Sha, L., An, S., Zhang, Y., et al. (2019). Cryo-EM structure of the human MLL1 core complex bound to the nucleosome. *Nature Communications* 10.
24. Patel, A., Dharmarajan, V., Vought, V.E., and Cosgrove, M.S. (2009). On the mechanism of multiple lysine methylation by the human mixed lineage leukemia protein-1 (MLL1) core complex. *Journal of Biological Chemistry* 284, 24242–24256.

25. Patel, A., Vought, V.E., Dharmarajan, V., and Cosgrove, M.S. (2011). A Novel Non-SET Domain Multi-subunit Methyltransferase Required for Sequential Nucleosomal Histone H3 Methylation by the Mixed Lineage Leukemia Protein-1 (MLL1) Core Complex *. *Journal of Biological Chemistry* 286, 3359–3369.
26. Peach, S.E., Rudomin, E.L., Udeshi, N.D., Carr, S.A., and Jaffe, J.D. (2012). Quantitative Assessment of Chromatin Immunoprecipitation Grade Antibodies Directed against Histone Modifications Reveals Patterns of Co-occurring Marks on Histone Protein Molecules. *Molecular & Cellular Proteomics* 11, 128–137.
27. Rao, R.C., and Dou, Y. (2015). Hijacked in cancer: The KMT2 (MLL) family of methyltransferases. *Nature Reviews Cancer* 15, 334–346.
28. Rossetto, D., Avvakumov, N., and Côté, J. (2012). Histone phosphorylation. *Epigenetics* 7, 1098–1108.
29. Santos-Rosa, H., Schneider, R., Bannister, A.J., Sherriff, J., Bernstein, B.E., Emre, N.C.T., Schreiber, S.L., Mellor, J., and Kouzarides, T. (2002). Active genes are tri-methylated at K4 of histone H3. *Nature* 2002 419:6905 419, 407–411.
30. Sha, L., Ayoub, A., Cho, U.S., and Dou, Y. (2020). Insights on the regulation of the MLL/SET1 family histone methyltransferases. *Biochimica et Biophysica Acta - Gene Regulatory Mechanisms* 1863.
31. Shinsky, S.A., Monteith, K.E., Viggiano, S., and Cosgrove, M.S. (2015). Biochemical reconstitution and phylogenetic comparison of human SET1 family core complexes involved in histone methylation. *Journal of Biological Chemistry* 290, 6361–6375.
32. Simon, M.D., and Shokat, K.M. (2012). A Method to Site-Specifically Incorporate Methyl-Lysine Analogues into Recombinant Proteins. *Methods in Enzymology* 512, 57–69.
33. Smith, L.M., and Kelleher, N.L. (2013). Proteoform: a single term describing protein complexity. *Nature Methods* 2013 10:3 10, 186–187.

34. Strahl, B.D., and Allis, C.D. (2000). The language of covalent histone modifications. *Nature* *403*, 41–45.
35. Strahl, B.D., Ohba, R., Cook, R.G., and Allis, C.D. (1999). Methylation of histone H3 at lysine 4 is highly conserved and correlates with transcriptionally active nuclei in *Tetrahymena*. *Proceedings of the National Academy of Sciences* *96*, 14967–14972.
36. Strelow, J.M., Xiao, M., Cavitt, R.N., Fite, N.C., Margolis, B.J., and Park, K.J. (2016). The Use of Nucleosome Substrates Improves Binding of SAM Analogs to SETD8. *Journal of Biomolecular Screening* *21*, 786–794.
37. Stützer, A., Liokatis, S., Kiesel, A., Schwarzer, D., Sprangers, R., Söding, J., Selenko, P., and Fischle, W. (2016). Modulations of DNA Contacts by Linker Histones and Post-translational Modifications Determine the Mobility and Modifiability of Nucleosomal H3 Tails. *Molecular Cell* *61*, 247–259.
38. Su, Z., and Denu, J.M. (2016). Reading the Combinatorial Histone Language. *ACS Chemical Biology* *11*, 564–574.
39. Tan, S., Kern, R.C., and Selleck, W. (2005). The pST44 polycistronic expression system for producing protein complexes in *Escherichia coli*. *Protein Expression and Purification* *40*, 385–395.
40. Taverna, S.D., Ueberheide, B.M., Liu, Y., Tackett, A.J., Dias, R.L., Shabanowitz, J., Chait, B.T., Hunt, D.F., and Allis, C.D. (2007). Long-distance combinatorial linkage between methylation and acetylation on histone H3 N termini. *Proceedings of the National Academy of Sciences of the United States of America* *104*, 2086–2091.
41. Taylor, B.C., and Young, N.L. (2021). Combinations of histone post-translational modifications. *Biochemical Journal* *478*, 511–532.
42. Usher, E.T., Namitz, K.E.W., Cosgrove, M.S., and Showalter, S.A. (2021). Probing multiple enzymatic methylation events in real time with NMR spectroscopy. *Biophysical Journal* *120*, 4710–4721.

43. Vaughan, R.M., Kupai, A., and Rothbart, S.B. (2021). Chromatin Regulation through Ubiquitin and Ubiquitin-like Histone Modifications. *Trends in Biochemical Sciences* 46, 258–269.
44. Weinberg, D.N., Papillon-Cavanagh, S., Chen, H., Yue, Y., Chen, X., Rajagopalan, K.N., Horth, C., McGuire, J.T., Xu, X., Nikbakht, H., et al. (2019). The histone mark H3K36me2 recruits DNMT3A and shapes the intergenic DNA methylation landscape. *Nature* 573, 281–286.
45. Weinberg, D.N., Rosenbaum, P., Chen, X., Barrows, D., Horth, C., Marunde, M.R., Popova, I.K., Gillespie, Z.B., Keogh, M.C., Lu, C., et al. (2021). Two competing mechanisms of DNMT3A recruitment regulate the dynamics of de novo DNA methylation at PRC1-targeted CpG islands. *Nature Genetics* 53, 794–800.
46. Weiss, J.N. (1997). The Hill equation revisited: uses and misuses. *The FASEB Journal : Official Publication of the Federation of American Societies for Experimental Biology* 11, 835–841.
47. Wozniak, G., and Strahl, B. (2014). Hitting the “mark”: interpreting lysine methylation in the context of active transcription. *Biochimica et Biophysica Acta* 1839, 1353–1361.
48. Xue, H., Yao, T., Cao, M., Zhu, G., Li, Y., Yuan, G., Chen, Y., Lei, M., and Huang, J. (2019). Structural basis of nucleosome recognition and modification by MLL methyltransferases. *Nature* 573, 445–449.
49. Young, N.L., DiMaggio, P.A., Plazas-Mayorca, M.D., Baliban, R.C., Floudas, C.A., and Garcia, B.A. (2009). High throughput characterization of combinatorial histone codes. *Molecular and Cellular Proteomics* 8, 2266–2284.
50. Young, N.L., DiMaggio, P.A., and Garcia, B.A. (2010). The significance, development and progress of high-throughput combinatorial histone code analysis. *Cellular and Molecular Life Sciences* 2010 67:23 67, 3983–4000.