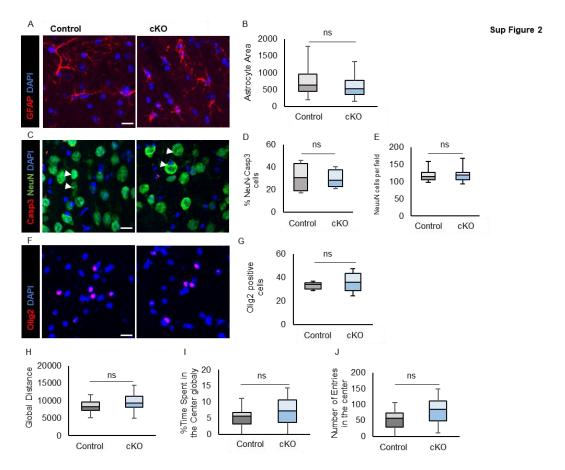
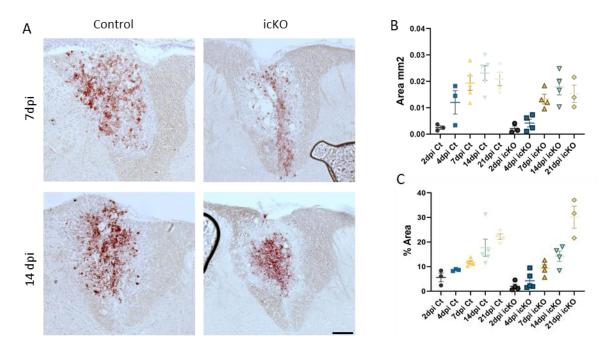


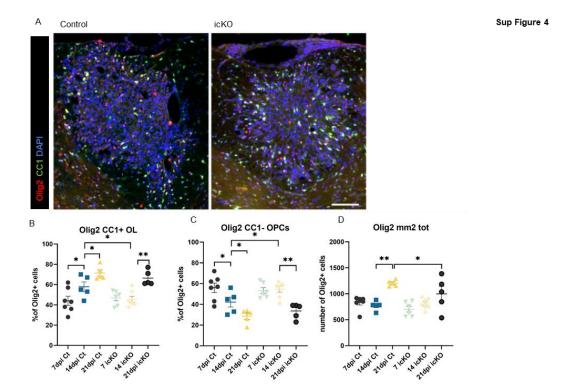
Supplementary figure 1. mGcm1 and mGcm2 expression. (A) Immunolabelling of CNS primary cultures for mGcm1 (grey), microglia (CD45, red) and DAPI (blue), N=3; scale bar 10 μm. (B) RNA ISH of brain sections on P14, 2 and 18month-old brain sections from control and cko animals: *Cx3cr1* (red), *mGcm2* (green) and DAPI (blue), N=3; scale bar: 10 μm. (C) RNAscope labelling on control and cKO brain sections at 18 months: *Cx3cr1* (red), *mGcm1* (green), DAPI (blue), N=3; scale bar: 10 μm. (D) Immunolabelling of resident macrophages from lung and adipose tissues at 24month-old animals: mGcm2 (grey), CD45 (red) and DAPI (blue). Scale bar: 100 μm. *Cx3cr1-Cre*^{+/-}; *mGcm2*^{flox/}(control) and *Cx3cr1-Cre*^{+/-}, *mGcm2*^{flox/flox}(cKO)



Supplementary figure 2. Characterisation of cell populations in the cortex of **24month-old animals**. (**A**) Immunolabelling of astrocytes from control and cKO animals for GFAP (red). (**B**) Quantification of the GFAP area. (**C**) Immunolabelling of neurons with the pan neuronal marker NeuN (green) and the cell death marker Caspase3 (red). (**D**) Quantification for double positive NeuN-Caspase3 cells to calculate the neuronal cell death and (**E**) the total neuronal number. (**F**) Immunolabelling for oligodendrocytes with Olig2 and (G) quantification of oligodendrocytes in control and cKO animals. (**H,J**) Open field test for control and cKO animals at 18 months. The animals were evaluated for the total distance covered (**H**), the percentage of time spent in the centre (**I**) and how many times they entered in the centre (**J**). p-value: *<0.05, **<0.01, ***<0.001, and ns for not significant. N=5-13; scale bar: 50 μm. Statistical significance was determined by one-way ANOVA followed 2-tailed, unpaired t-test. $Cx3cr1-Cre^{+/-}$; $mGcm2^{flox/}$ (control) and $Cx3cr1-Cre^{+/-}$, $mGcm2^{flox/}$ (cKO)

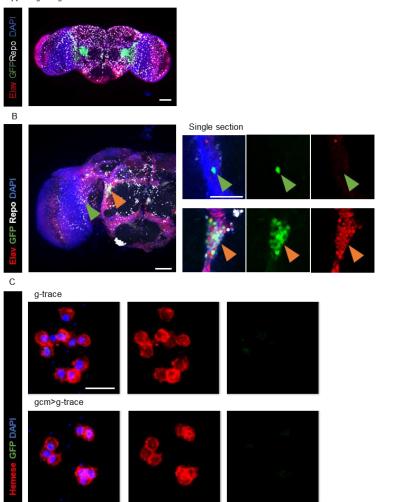


Supplementary Figure 3. The density of macrophages containing myelin debris is not altered in LPC demyelinated lesions of *mGcm2* icKO mice. (A) Oil-Red O staining of macrophages with myelin debris in control and *mGcm2* icKO LPC lesions, at 7 and 14 dpi. (B) Quantification of Oil-Red-O+ area in LPC lesions from 2 to 21 dpi, in control and *mGcm2* icKO animals. (C) Percentage of Oil-Red-O+ area in LPC lesions from 2 to 21 dpi in both experimental groups. Note that the clearance of myelin debris by macrophages in demyelinated lesions is not altered in *mGcm2* icKO with respect to control mice. Scale bar: (A), 100μm.



Supplementary Figure 4. Loss of mGcm2 function in microglia delays oligodendrocyte differentiation in demyelinated lesions. (A) Immunolabelling with Olig2 (red) and CC1 (green) in LPC lesions of the spinal cord at 14 dpi, in control and mGcm2 icKO mice. Nuclei are counterstained with DAPI. (B,C) Graphs indicating the percentage of Olig2/CC1-double positive differentiated oligodendrocytes and Olig2-positive/CC1-negative OPCs in demyelinated lesions from 7 to 21 dpi, in control and mGcm2 icKO mice. (D) Quantification of the number of total Olig2-positive oligodendroglia in LPC lesions at 7, 14 and 21 dpi, in control and mGcm2 icKO animals. Two-way ANOVA followed by Tukey's multiple comparison tests were used for statistical analysis. *p< 0.05, **p< 0.01. Scale bar: (A), 100 μ m.

A gcm>g-trace Sup.Figure 5



Supplementary Figure 5. An inducible system tracing *gcm* expression. (A) Immunolabelling of adult *Drosophila* brain with *gcm* tracing (*gcm>g-trace*=GFP) in green, Repo (white), Elav (white) and DAPI (blue). (B) Immunolabelling of adult *Drosophila* brain with *gcm* tracing (*gcm>g-trace*) in green, Repo (grey), Elav (red) and DAPI (blue), Scale bar: 20 μm. (C) Immunolabelling of hemocytes from gcm>g-trace strain. The pan-hemocyte marker Hemese (red), g-trace (green), DAPI (blue). Animals were raised at 18°C and shifted at 29°C at second instar larval. scale bar: 20 μm.

Host	Target	Dilution	Reference		
mouse	anti-Arginase1	1/200	sc-271430		
rat	anti-CD11b	1/400	MCA-74G		
rat	anti-CD45	1/50	MCD-4500		
rat	anti-CD68	1/400	MCA-1957		
rat	anti-F4/80	1/100	MCA-497		
rabbit	anti-Gcm1	1/50	B. Nait-Oumesmar		
rabbit	anti-Gcm2	1/200	ab201170		
rabbit	anti-Iba1	1/300	1919741		
mouse	anti-iNOS	1/100	ab210823		
mouse	anti-GFAP	1/100	PCN		
mouse	anti-NeuN	1/100	ab279296		
rabbit	anti-Casp3	1/200	mAb #9664		
mouse	anti-MOG	1/20	clone C18C5, C. Linnington		
rabbit	anti-Olig2	1/400	AB9610		
mouse	anti-APC	1/100	OP80		
mouse	anti-HLA	1/100	CR3/43		
mouse	anti-Repo	1/20	8D12 (DSHB)		
rat	anti-Elav	1/100	7E8A10 (DSHB)		
mouse	anti-Hemese	1/40	I. Ando		
mouse	anti-L4	1/50	I. Ando		
chicken	anti-GFP	1/500	703095155		
donkey	anti-mouse-Cy3	1/500	715-165-151		
donkey	anti-rabbit-Cy3	1/500	711-165-152		
goat	anti-rat-Cy3	1/500	112-165-167		
goat	anti-mouse-Alexa647	1/500	115-605-166		
goat	anti-rabbit-Cy5	1/500	111-175-144		
goat	anti-rat-Cy5	1/500	112-175-144		
donkey	anti-chicken-FITC	1/500	A11039		
goat	anti-rat-Alexa568	1/1000	A11077		
donkey	anti-rat-Alexa488	1/1000	A21208		
donkey	anti-rat-Alexa647	1/1000	AB150155		
donkey	anti-rabbit-Alexa568	1/1000	A10042		
goat	anti-mouse-Alexa488	1/1000	1070-02		
donkey	anti-mouse-Alexa488	1/1000	A21202		
goat	anti-mouse-FITC	1/100	A21141		

Supplementary Table 1. Primary and secondary antibodies

MS cases	Age	Sex	PMD	Disease	Disease course	Lesions analyzed			
				duration		Active	Chronic	Chronic	Shadow
							active	inactive	plaques
MS94CLA6	42	F	11	6	PP	1			
MS100CLC2	46	M	7	8	SP			1	
MS121CLC5	49	F	24	14	PR	1		1	
MS106L6C8	39	F	18	21	PP		1		
Control									
C26CLA5	79	F	18		Cardiac failure				

Supplementary Table 2. MS and control cases used to study hGCM2 expression

	Target	Taqman probes
	Hprt	HPRT TaqMan® Gene Expression Assays (Mm03024075_m1)
	mGcm2	GCM2 TaqMan® Gene Expression Assays (Mm00492312_m1)
M1	iNOS	NOS2 TaqMan® Gene Expression Assays (Mm00440502_m1)
M1	TLR2	TLR2 TaqMan® Gene Expression Assays (Mm00442346_m1)
M2a	Arg-1	ARG1 TaqMan® Gene Expression Assays (Mm00475988_m1)
M2b	Il-4ra	IL-4a TaqMan® Gene Expression Assays (Mm01275139_m1)
M2c	CD163	Cd163 TaqMan® Gene Expression Assays (Mm00474091_m1)

Supplementary Table 3. Taqman probes used for qRT-PCR analysis of LPC lesions