

Supplementary Information for “Altered tRNA processing is linked to a distinct and unusual La protein in *Tetrahymena thermophila*”

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Figure S1. Multiple sequence alignments of La proteins from several eukaryotes demonstrate high conservation of the LaM and absence of the RRM1.

Figure S2. Mlp1 demonstrates preferential binding of certain tRNA isotypes and unprocessed pre-tRNAs.

Figure S3. Mlp1 binding to typical La protein target RNAs.

Figure S4. Mlp1 binding to uridylate RNA does not discriminate between the position of the uridylate.

Figure S5. tRNA mediated suppression protein expression in *Schizosaccharomyces pombe*.

Figure S6. Confirmation of the partial Mlp1 *Tetrahymena thermophila* knockout strain.

Figure S7. The effect of short 3'-trailer sequences on 5'-leader composition and Mlp1 binding affinity.

Figure S8. Primary sequence alignments of the 3'-exonuclease Rex1, 3'-endonuclease RNase Z and LSM2-8 complex from different eukaryotic species.

Table S1. Kd values from EMSAs determining binding of Mlp1 to different processed pre-tRNA intermediates and mature tRNA.

Table S2. Raw counts for 3'-U ending and 3'-CCA ending tRNAs from raw .fastq files from wild type (WT) input tRNA and Mlp1-immunoprecipitated tRNA (IP).

Table S3. Raw counts for 3'-U ending and 3'-CCA ending tRNAs from raw .fastq files from wild type (WT) and partial Mlp1 knockout (KO) strains.

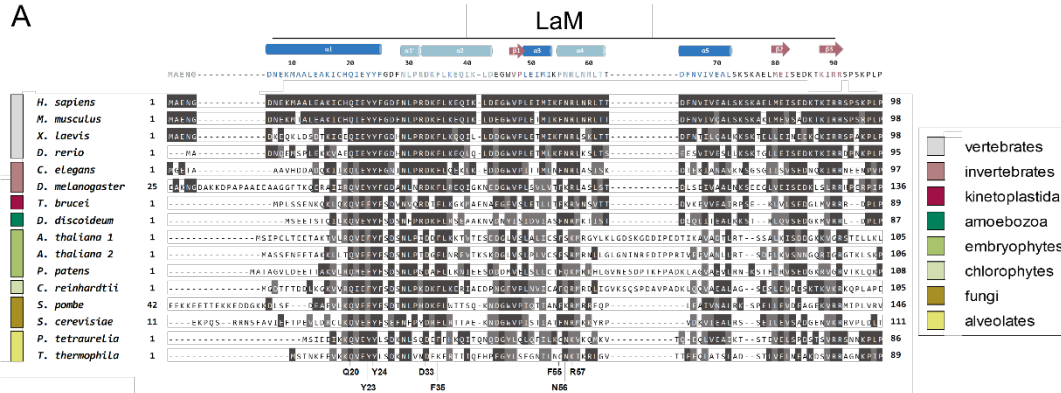
Table S4. Number of tRNA genes encoded in the genome of different eukaryotic species for each isotype (top) and isoacceptor (bottom).

Table S5. List of oligonucleotides used in this study.

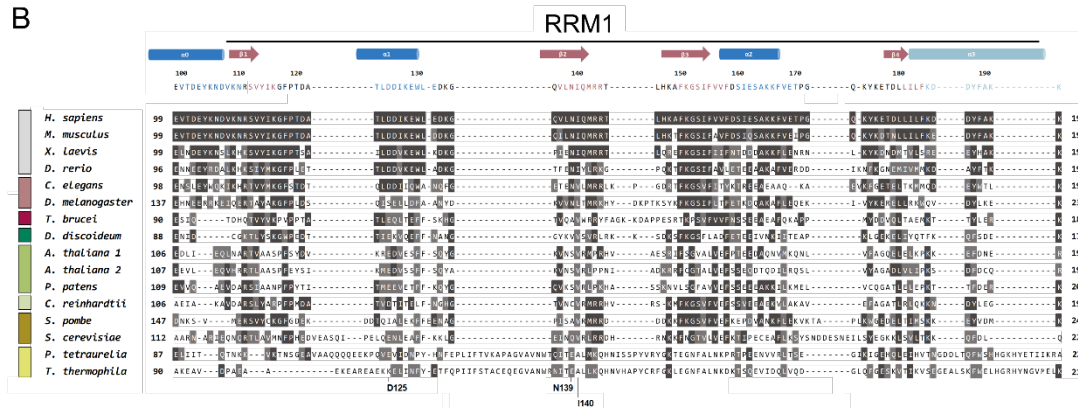
Table S6. NCBI accession numbers of primary amino acid sequences used for conservation analysis.

S1

A



B



■ identical residue ■ conserved residue □ non-conserved residue

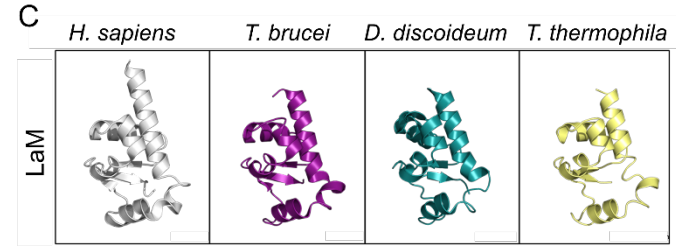
Figure S1. Multiple sequence alignments of La proteins from several eukaryotes demonstrate high conservation of the La Motif (LaM) and absence of the RNA recognition motif-1 (RRM1).

(A,B) Primary amino acid alignments of full RNA-binding domains LaM and RRM1 from different eukaryotic lineages. A dark grey background indicates identical residues, light grey conserved residues and white indicates no conservation. Most uridylylate binding residues are conserved in the LaM of alveolates, whereas residues in the RRM1 are more variable. Important uridylylate binding residues are shown at the bottom. The secondary structure motif of the high-resolution hLa protein structure is shown at the top with β -sheets shown in red and α -helices in blue (dark blue: typical α -helices found in the winged-helix fold and classic RRM, light blue: inserted α -helices found in La proteins specifically) (PDB: 2VOD).

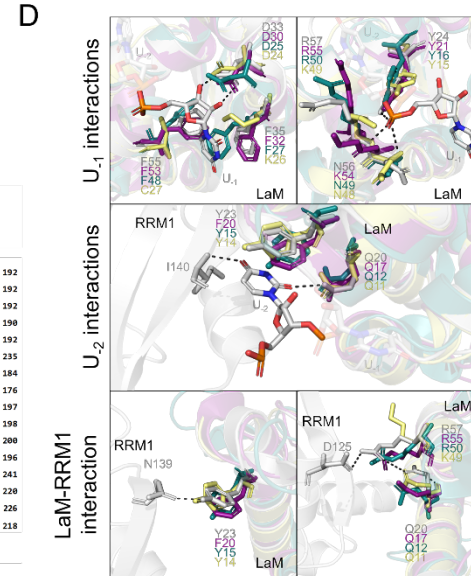
(C) High-resolution structures of the LaM in different species: *Homo sapiens* in white (PDB: 2VOD)¹⁴, *Trypanosoma brucei* in purple (PDB: 1S29)¹⁰, *Dictyostelium discoideum* in teal (PDB: 2M5W)⁵⁶ and the predicted tertiary structure for *Tetrahymena thermophila* in yellow⁴².

(D) Magnified views of La-U₁ RNA, La-U₂ and LaM-RRM1 interactions. U₁ is the most 3'-terminal uridylylate possessing 2'-OH and 3'-OH ends. Most uridylylate binding residues located in the LaM are conserved in *Tetrahymena thermophila*, except for F35 and F55 (hLa numbering) which bind the most 3'-terminal U₁. Carbon atoms within the RNA (U₁ and U₂) from the *Homo sapiens* La structure are colored in white with other atoms colored by type: oxygen in red; nitrogen in blue and phosphorous in orange. Hydrogen bonds are shown as black dashed lines.

C



D



S2

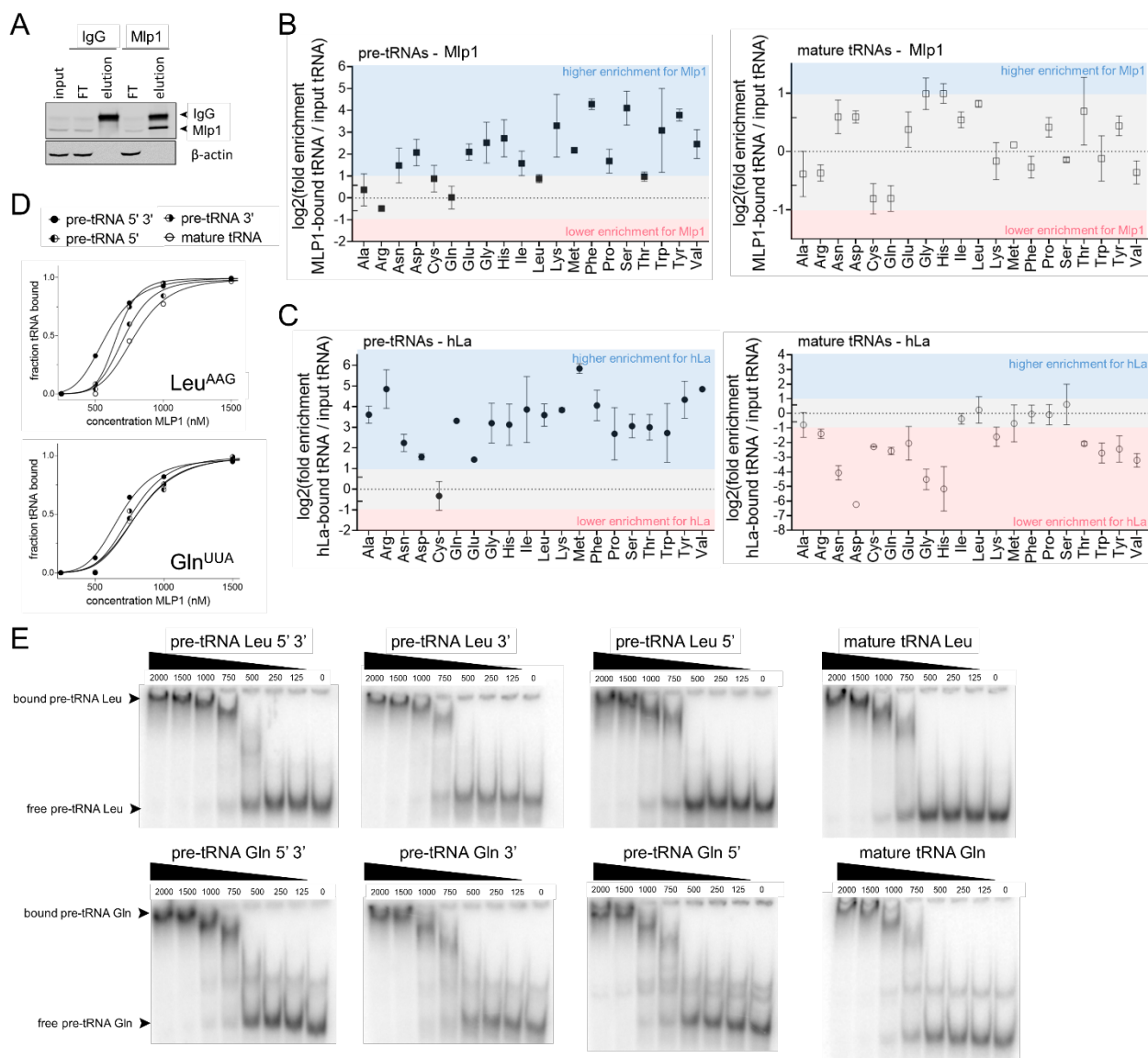
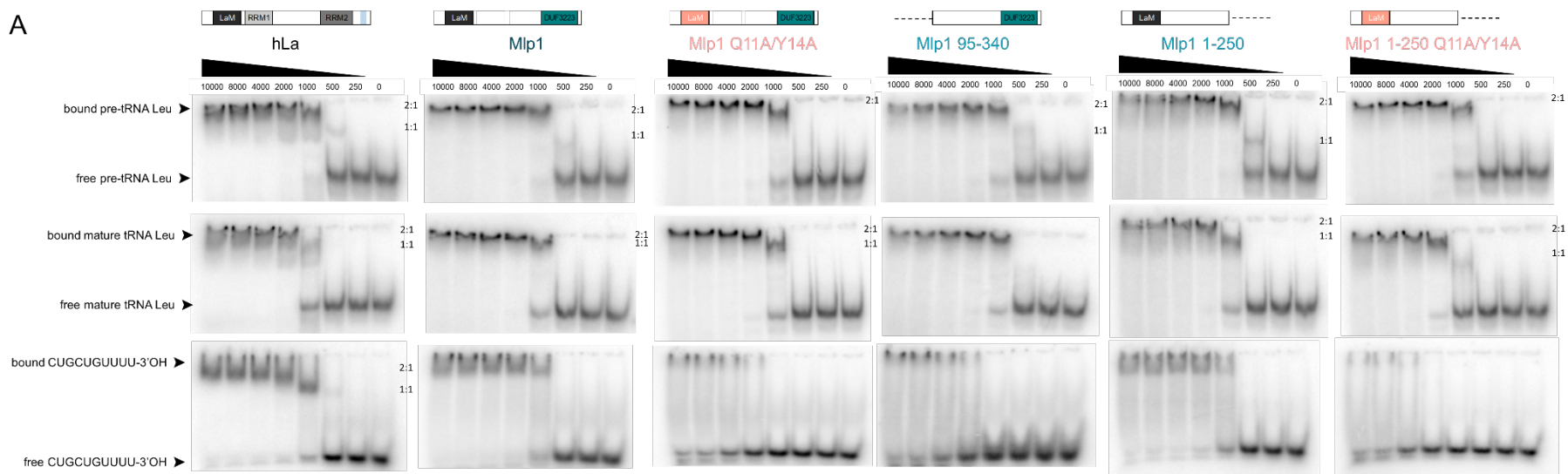


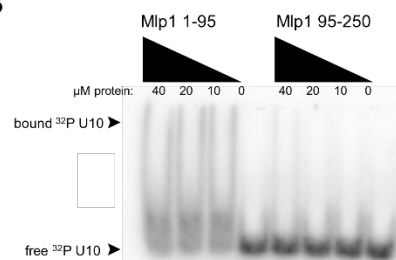
Figure S2. Mlp1 demonstrates preferential binding of certain tRNA isotypes and unprocessed pre-tRNAs. (A) Western blot confirming Mlp1-specific immunoprecipitation from *Tetrahymena thermophila* using an affinity purified rabbit anti-Mlp1 antibody and a rabbit isotype immunoglobulin G (IgG) control. The Mlp1-specific antibody was used for both ribonucleoprotein-immunoprecipitation (RNP-IP) and western blotting. The heavy chain (50 kDa) of the antibodies was detected with the secondary anti-rabbit antibody and shown as IgG. Loading control: β -actin. (B-C) Next generation sequencing data of tRNAs split by tRNA isotypes for Mlp1 (n=3) (B) and hLa from Gogakos *et al.* (n=2) (C). Fold enrichment is shown as the log₂ transformed ratio between Mlp1- or hLa-immunoprecipitated tRNAs and input tRNAs. Both Mlp1 and hLa have a higher binding affinity for pre-tRNAs compared to mature tRNAs. (D-E) Binding curves from EMSAs (D) and corresponding native gels (E) comparing binding affinity of Mlp1 between ³²P-labeled pre-tRNA containing 5'- and 3'-extensions or 5'- or 3'-extensions and mature tRNA for Leu^{AAG} and Gln^{UUA}. The highest binding affinity is found for 5'- and 3'-end containing pre-tRNAs. See Table S1 for K_d quantifications.

S3

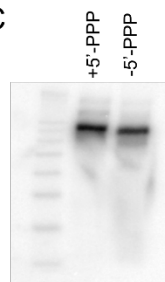
A



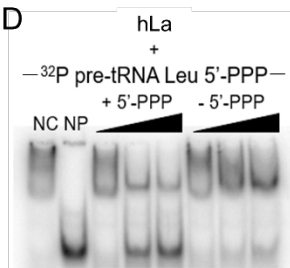
B



C



D



E

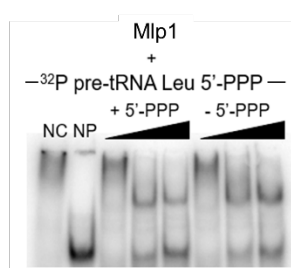


Figure S3. Mlp1 binding to typical La protein target RNAs.

(A) Native gels comparing binding to ^{32}P -labeled pre-tRNAs, mature tRNAs and a 3'-trailer sequence CUGCUGUUUU-3'OH. Unbound RNA is found at the bottom of the gel, while protein-RNA complexes shift upwards. The numbering on the side of the gels indicate binding events. A single protein bound to the RNA is indicated at 1:1 and two proteins bound to the same RNA is denoted at 2:1.

(B) Native gels comparing binding to ^{32}P -labeled U10 RNA for mutants Mlp1 1-95 (LaM only) and Mlp1 95-250 (middle domain only).

(C) Denaturing gel of *in vitro* transcribed ^{32}P -labeled 5'-leader containing, 3'-trailer processed pre-tRNAs produced side-by-side as unlabelled-competitor pre-tRNAs (lacking ^{32}P -labeled labeling) used in (E,F) to confirm successful removal of the 5'-triphosphate group.

(D,E) Native gels comparing unlabelled-competitors 5'-triphosphate containing pre-tRNA (+5'PPP) and dephosphorylated pre-tRNA (-5'PPP) for ^{32}P -labeled +5'PPP binding on hLa (D) or Mlp1 (E). Competition is seen as a decrease in protein-RNA complex and increase in unbound RNA. NP: no protein lane, NC: no unlabeled competitor lane

S4

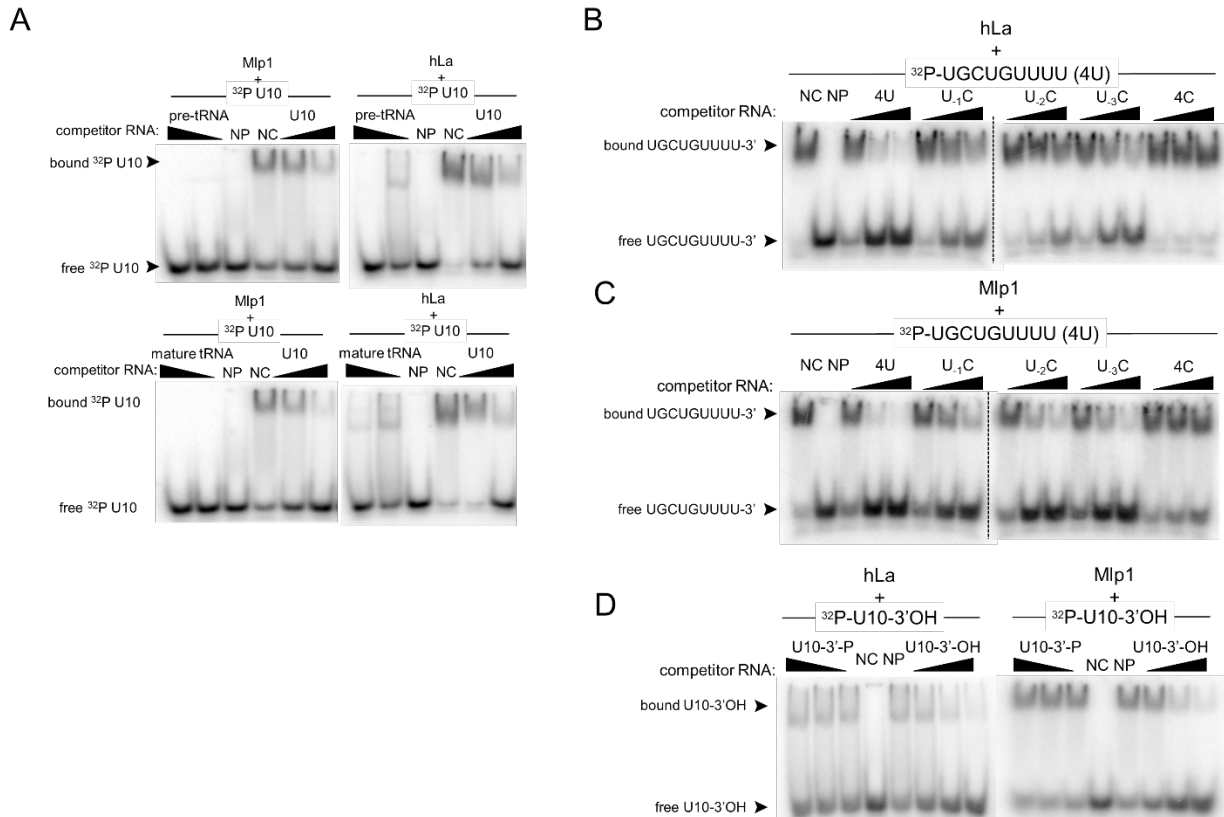


Figure S4. Mlp1 binding to uridylate RNA does not discriminate between the position of the uridylate.

(A) Native gels comparing unlabelled-competitors U10 (positive control), pre-tRNA and mature tRNA for ³²P-labeled U10 binding. Competition is seen as a decrease in protein-RNA complex and increase in unbound RNA. NP: no protein lane, NC: no competitor lane.

(B-C) Native gels comparing unlabelled-competitors CUGCUGUUUU (4U), CUGCUGUUUC (U₁C), CUGCUGUUCU (U₂C), CUGCUGUCUU (U₃C) and CUGCUGCCCC (4C) for ³²P-labeled 4U binding on hLa (B) or Mlp1 (C). Competition is seen as a decrease in protein-RNA complex and increase in unbound RNA. NP: no protein lane, NC: no competitor lane.

(D) Native gels comparing a 3'-phosphorylated substrate (U10-3'-P) and normal 3'-OH containing substrate (U10-3'-OH) for binding of ³²P-labeled U10-3'-OH. Competition is seen as a decrease in protein-RNA complex and increase in unbound RNA. No competition was observed with the phosphorylated target for either Mlp1 or hLa. NP: no protein lane, NC: no competitor lane.

S5

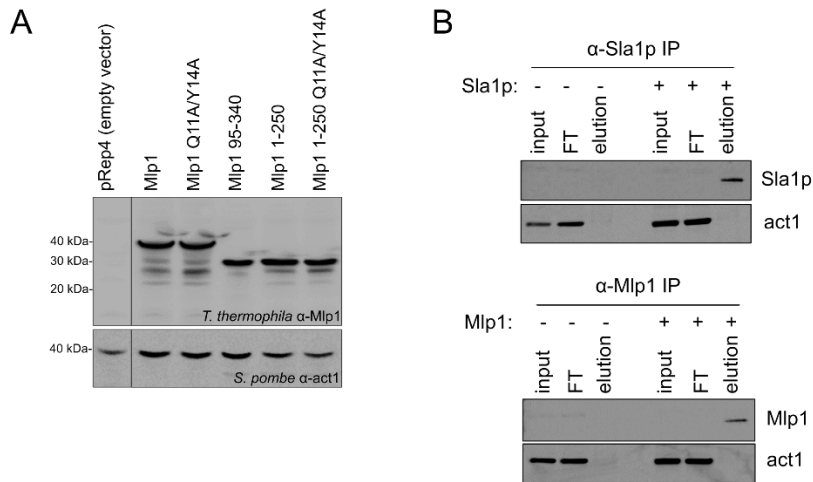


Figure S5. tRNA mediated suppression protein expression in *Schizosaccharomyces pombe*.

(A) Western blot confirming Mlp1 protein expression levels in *Schizosaccharomyces pombe* following pRep4 plasmid transformation in the tRNA-mediated suppression assay Figure 4A. Loading control: act1.

(B) Western blot confirming immunoprecipitation (IP) of Sla1p and Mlp1 from Sla1p- and Mlp1-transformed *Schizosaccharomyces pombe* ySH9 tRNA suppressor strains. pRep4 empty vector transformed strains were used as a control for background immunoprecipitation using Sla1p and Mlp1 antibodies. Loading control: act1. FT: flow through.

S6

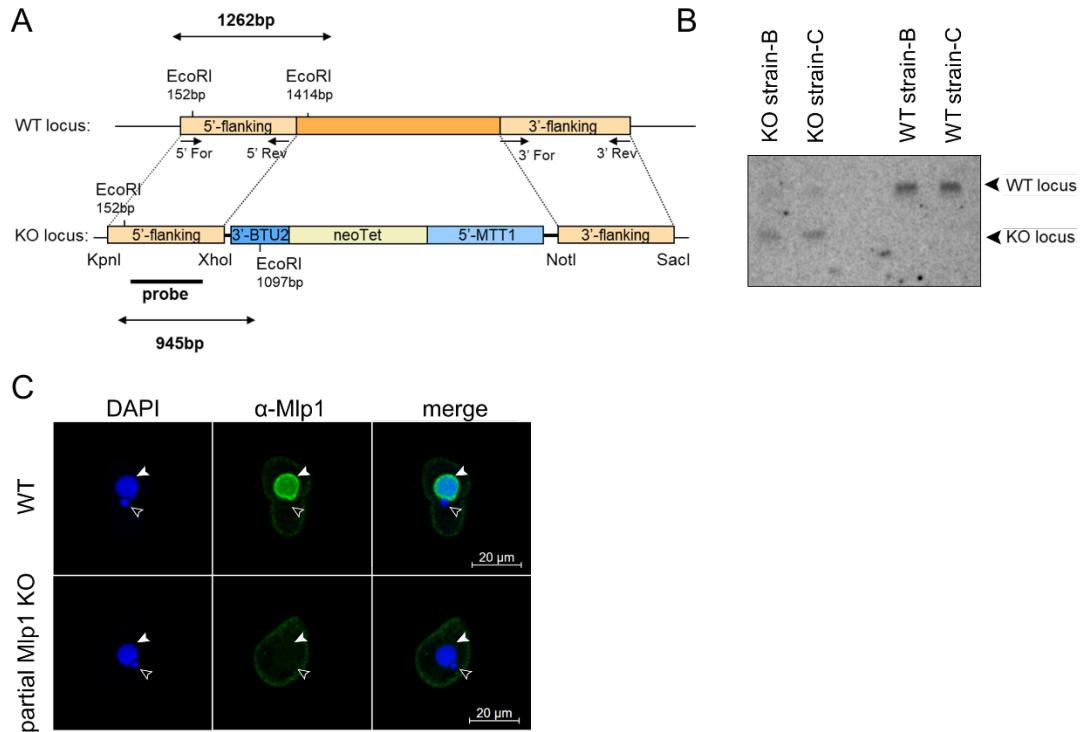


Figure S6. Confirmation of the partial Mlp1 *Tetrahymena thermophila* knockout strain.

(A) Schematic overview of the *Tetrahymena thermophila* wild type (WT) locus encoding endogenous Mlp1 and knockout (KO) locus following homologous recombination between 5'- and 3'-flanking regions of the genome with transformation plasmid encoding the neomycin (neoTet) selection marker. Restriction enzymes used for cloning 5'- and 3'-flanking regions for homologous recombination are shown below the KO locus. Genomic DNA digest before southern blotting were performed with *EcoRI* restriction enzyme generating a 1262 bp fragment for the WT locus and a 945 bp fragment for the KO locus.

(B) Southern blot of genomic DNA digested with restriction enzyme *EcoRI* probed with ^{32}P -labeled PCR-generated probe reveals almost complete KO of Mlp1 (see schematic in A).

(C) Indirect immunofluorescent staining of *Tetrahymena thermophila* WT and partial Mlp1 KO strains using the polyclonal rabbit anti-Mlp1 antibody. Nuclei were stained using DAPI. Full white arrows are denoting the transcriptionally active macronucleus and empty white arrows show the transcriptionally inactive micronucleus.

S7

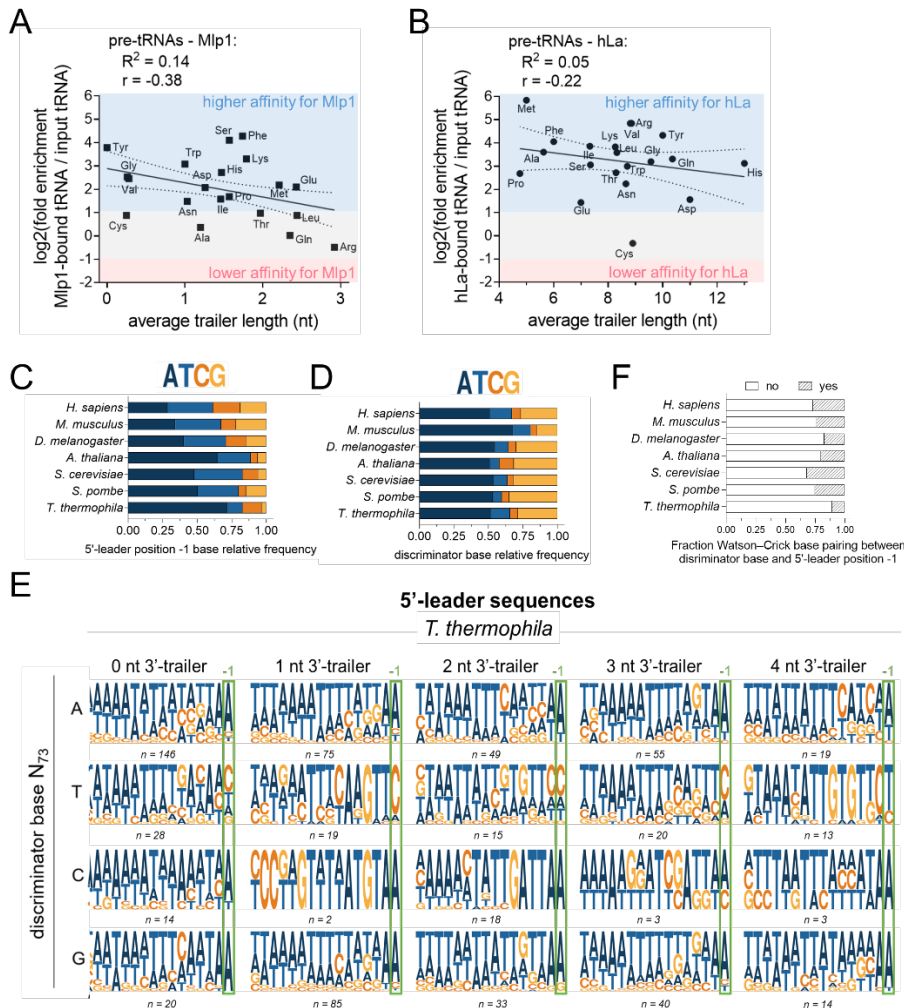


Figure S7. The effect of short 3'-trailer sequences on 5'-leader composition and Mlp1 binding affinity.

(A,B) Next generation sequencing data of Mlp1-bound pre-tRNAs (n=3) (A) and hLa-bound pre-tRNAs (n=2) (B) split by tRNA isotypes compared against 3'-trailer lengths.

(C) Distribution of the nucleotide at the most 3'-terminal position in the 5'-leader of pre-tRNAs from different eukaryotes. *Tetrahymena thermophila* has a high frequency of adenosines in this position.

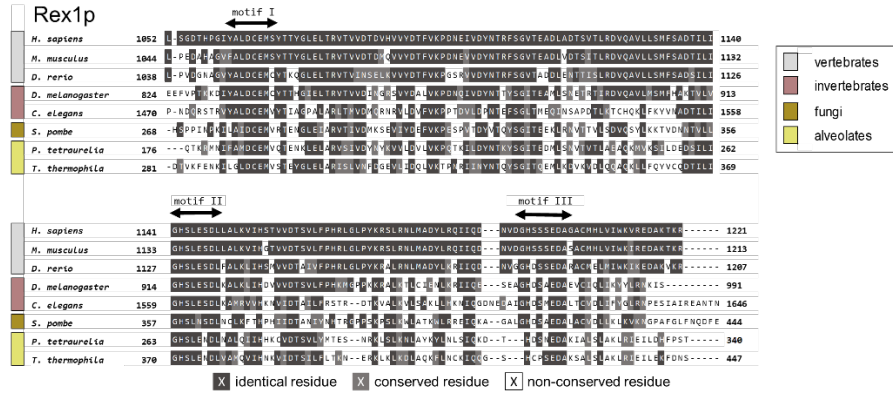
(D) Distribution of the nucleotide found as the discriminator base (N₇₃) in different eukaryotes. Distribution is equal between different species.

(E) Logo analysis of *Tetrahymena thermophila* 5'-leader sequences split by the discriminator base identity and 3'-trailer length. The number of analyzed pre-tRNAs is shown underneath each logo.

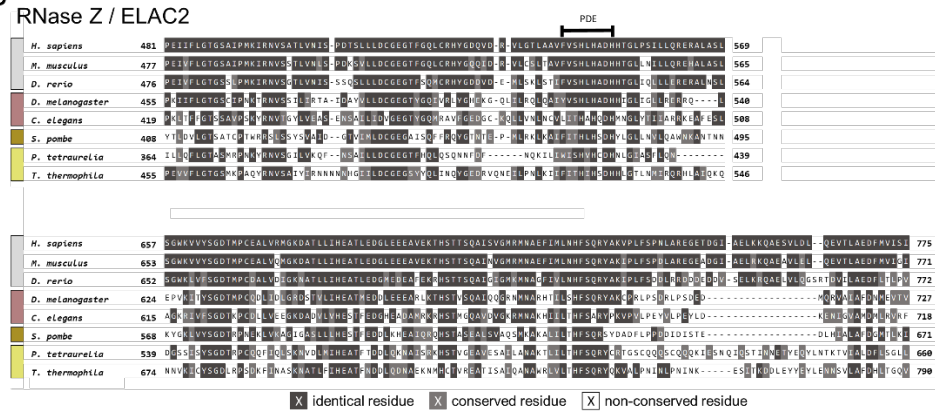
(F) Distribution of pre-tRNAs containing a perfect Watson-Crick base pairing between the discriminator base (N₇₃) preceding the 3'-trailer and most 3'-terminal 5'-leader nucleotide (N₋₁).

S8

A



B



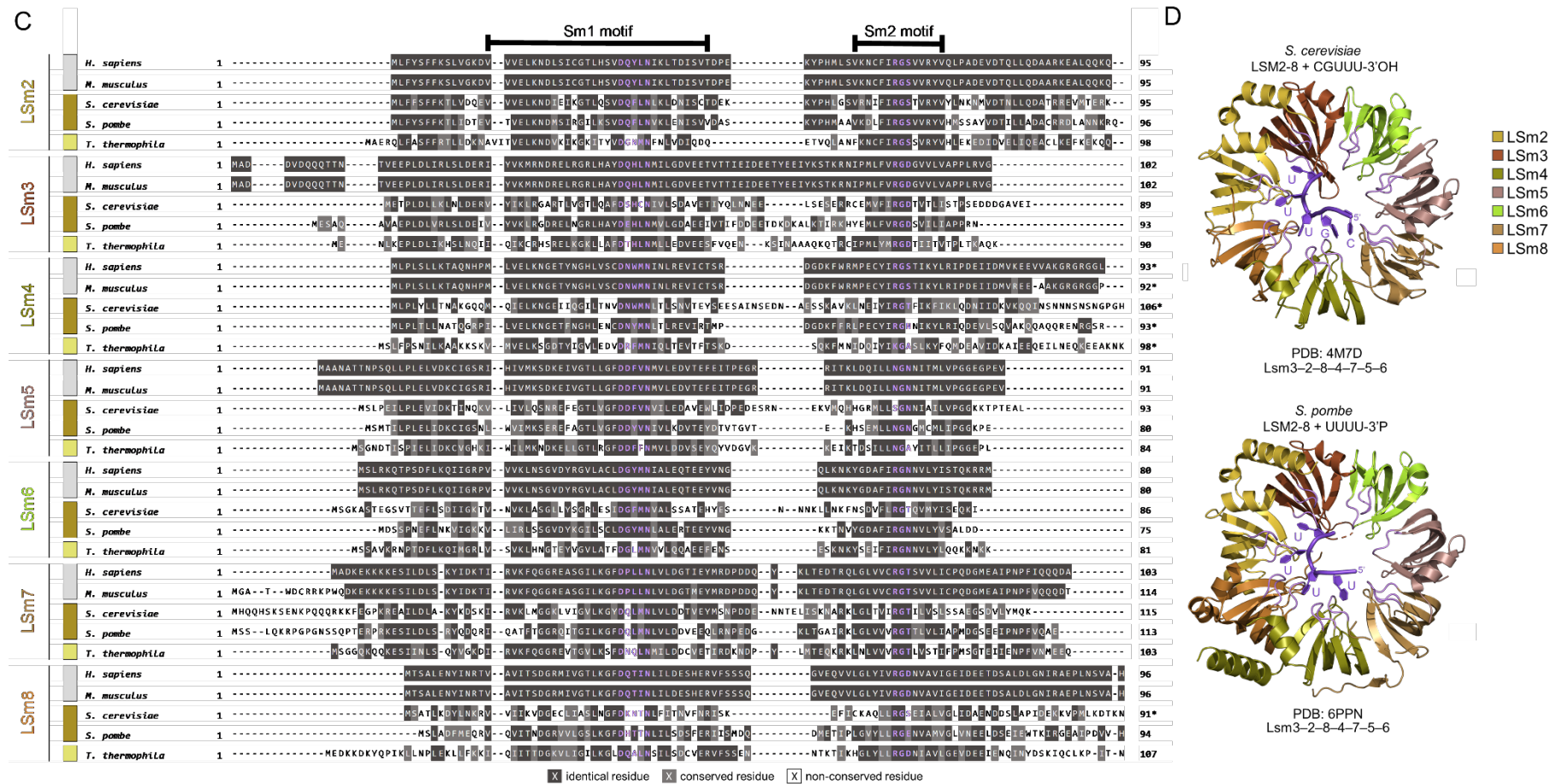


Figure S8. Primary sequence alignments of the 3'-exonuclease Rex1p, 3'-endonuclease RNase Z / ELAC2 and Lsm2-8 complex from different eukaryotic species.

(A-C) Primary amino acid alignments from different eukaryotic lineages revealing conservation of the 3'-exonuclease Rex1p (A), 3'-endonuclease RNase Z/ELAC2 (B) and Lsm2-8 complex (C). Regions important for Rex1p function are highlighted at motif I, motif II and motif III. The RNase Z catalytic domain is shown as PDE. Lsm2-8 complex conserved RNA interacting residues are highlighted in purple and conserved Sm1 and Sm2 motifs are shown. A dark grey background indicates identical residues, light grey conserved residues and white indicates no conservation.

(D) High-resolution structures of the Lsm2-8 complex in *Saccharomyces cerevisiae* (PDB: 4M7D) and *Schizosaccharomyces pombe* (PDB: 6PPN) in complex with 3'-uridylyate RNA shown in dark purple. The conserved RNA interacting residues, highlighted in purple C, are also shown in light purple and are found in the loops.

Table S1. K_d values from EMSAs determining binding of Mlp1 to different processed pre-tRNA intermediates and mature tRNA.

	Leu AAG K_d (nM)	Gln UUA K_d (nM)
pre-tRNA 5' 3'	586 ± 15	687 ± 44
pre-tRNA 3'	658 ± 36	740 ± 40
pre-tRNA 5'	714 ± 21	804 ± 16
mature tRNA	831 ± 33	801 ± 62

Table S4. Number of tRNA genes encoded in the genome of different eukaryotic species for each isotype (top) and isoacceptor (bottom). The number of tRNA genes was obtained by counting the number of entries for each tRNA isotype and anticodon from the Genomic tRNA Database (GtRNAdb) and from the UCSC Genome Browser for *Tetrahymena thermophila* in Additional Table 3. NNN anticodons were excluded from the analysis.

tRNA isotype	<i>T. thermophila</i>	<i>S. pombe</i>	<i>S. cerevisiae</i>	<i>A. thaliana</i>	<i>D. melanogaster</i>	<i>M. musculus</i>	<i>H. sapiens</i>
Ala	44	12	16	33	17	46	45
Arg	36	13	19	37	26	25	31
Asn	31	6	10	16	10	7	36
Asp	31	8	16	27	14	22	20
Cys	16	3	4	16	7	62	32
Gln	52	6	10	17	12	19	31
Glu	44	10	16	27	21	22	28
Gly	42	12	21	41	20	32	36
His	17	4	7	10	6	11	10
Ile	41	9	15	22	12	18	24
iMet	0	4	5	2	6	10	10
Leu	55	13	21	42	22	29	38
Lys	29	12	21	32	19	41	40
Met	33	3	5	23	6	9	10
Phe	38	5	10	16	8	7	17
Pro	28	9	12	67	17	20	25
Ser	46	13	17	64	20	22	28
Sup	0	0	0	1	0	0	1
Thr	29	10	16	23	18	1	22
Trp	21	3	6	14	8	0	7
Tyr	19	4	8	76	10	0	16
Val	32	12	18	30	15	0	34
Sum	684	171	273	636	294	403	541

tRNA isotype	anticodon	<i>T. thermophila</i>	<i>S. pombe</i>	<i>S. cerevisiae</i>	<i>A. thaliana</i>	<i>D. melanogaster</i>	<i>M. musculus</i>	<i>H. sapiens</i>
Ala		44	12	16	33	17	46	45
	AGC	31	9	11	16	12	22	30
	CGC	2	1		7	3	10	5
	GGC						3	
	TGC	11	2	5	10	2	11	10
Arg		36	13	19	37	26	25	31
	ACG	8	8	6	9	10	6	7
	CCG	1	1	1	4		3	4
	CCT	2	1	1	8	3	5	8
	TCG	1	1		6	10	5	6
	TCT	24	2	11	10	3	6	6
Asn		31	6	10	16	10	7	36
	ATT							2
	GTT	31	6	10	16	10	13	35
Asp		31	8	16	27	14	22	20
	GTC	31	8	16	27	14	16	19
Cys		16	3	4	16	7	62	32
	ACA						1	1
	GCA	16	3	4	16	7	61	31
Gln		52	6	10	17	12	19	31
	CTA *	9						
	CTG		2	1	9	8	12	22
	TTA *	30						
	TTG	13	4	9	8	4	7	9
Glu		44	10	16	27	21	22	28
	CTC	7	6	2	13	15	14	12
	TTC	37	4	14	14	6	8	16
Gly		42	12	21	41	20	32	36
	ACC						2	
	CCC	1	1	2	5		7	10
	GCC	29	8	16	23	14	15	15
	TCC	12	3	3	13	6	8	11
His		17	4	7	10	6	11	10
	ATG							1
	GTG	17	4	7	10	6	10	10
Ile		41	9	15	22	12	18	24
	AAT	29	8	13	17	10	12	16
	GAT						1	3
	TAT	12	1	2	5	2	5	5
iMet			4	5	2	6	10	10
	CAT		4	5	2	6	10	10
Leu		55	13	21	42	22	29	38
	AAG	19	5		12	4	5	13
	CAA	11	4	10	10	4	4	7
	CAG	2	1		3	8	10	9
	GAG			1	1			
	TAA	19	2	7	6	4	7	5
	TAG	4	1	3	10	2	3	4
Lys		29	12	21	32	19	41	40
	CTT		9	14	18	13	26	21
	TTT	29	3	7	14	6	15	19
Met		33	3	5	23	6	9	10
	CAT	33	3	5	23	6	9	10
Phe		38	5	10	16	8	7	17
	AAA	1						
	GAA	37	5	10	16	8	7	17
Pro		28	9	12	67	17	20	25
	AGG	20	6	2	16	7	8	11
	CGG	1	1		5	5	3	4
	GGG						1	1
	TGG	7	2	10	45	5	8	9
Ser		46	13	17	64	20	22	28
	ACT							1
	AGA	20	7	11	37	8	9	11
	CGA	2	1	1	4	4	3	4
	GCT	15	3	2	13	6	9	8
	GGA				1			
	TGA	9	2	3	9	2	1	4
Sup					1			1
	TTA				1			1
Thr		29	10	16	23	18	1	22
	AGT	22	7	11	10	9	1	10
	CGT	1	1	1	5	3		6
	TGT	6	2	4	8	6		6
Trp		21	3	6	14	8		7
	CCA	21	3	6	14	8		7
Tyr		19	4	8	76	10		16
	ATA							1
	GTA	19	4	8	76	10		15
Val		32	12	18	30	15		34
	AAC	21	9	14	15	6		11
	CAC	5	1	2	8	7		18
	TAC	6	2	2	7	2		5
Sum		684	171	273	636	294	403	541

* Anticodons encoding stop codons in eukaryotes except in *T. thermophila*

Table S5. List of oligonucleotides used in this study.

gBlock Gene Fragments used as DNA template for cloning MLP1	
gBlock MLP1 CDS codon optimized	ATGTCAATCAATAAAGAAGAGGTAATAAAGCAAGTCAACTACTCTGTGCGACAAGAAGTGGTAAATGATGAGAAATTCGCACTATCATCAAGAGCACCCAGAGGGTATTTATCCTTCGGCAATATCTTAACTGTAATAAGATTAAAGCGCTGGGGTGTGACGACATTCGAACAACGGCAACGTCCTTGTCTGATAGTACCTTAGTTGAAATTAATGAGGCAAGGACTCTGTGCGTCTGTGCGGAAATAAACCCATTCCAGCGAAGGAAGCAGTTGACCCCGCTGAAGCTGCAGAAAAAGAAGCCGGTGAGGC CGAAAAAAGGAACTGATCAACTTTTATGAACTTTTCAACCGATTATCTTCTCAACCGCTGCGAACAAGAGGGGGTAGCGAACTGGCGCAACATCACAGAGCCCTTGCTGAAGCAACATAACGTTTCATGCTCCGTATTGCCGCTTTGGCAAGTTGGAGGGTAACTTTGCATTGAATAAAGACAAAACTTCGCAAGAGGTTATCGACCAGCTTGCAAGACGGGCTTCAATTTGGAGAGTCTAAGGTGACTATCAAAGTGAAGTGAAGGCGAAGCCCTGTCAAAGTTTTGGGAGCTGCATGGACGCCACTACAACGGTGTGATGGAATTGAAAAAGAAGGAGTCAATCAGACAGTTAAAGCTAAGAAAGATAAAAAGGAGAAAAACAAAAACGTAATTGCAATTCGGGGGAGAAAAATATACAGATGATTGACTATAAAAATTTGTTAAGGGAATCTTAGGGCGTACGGCGAATGGACAAAAGATCATTAGTCCCTTATCAGGATGTTGAGAGCCTCTTGAGTATCAAAATAACAAAGAAGCCAAATTAAGGATTTAGATCACTTACGGTAGACGTCATCCTGAGCATAAAGATACAGTTGTTCTTGTGCTCAATCGGACGGAAACGAAGGAAATTTCTCAGCGTTAAGTGCATCTCAAACTTTGAGGAAAACTTAAACTTTGA

Oligonucleotides used for cloning MLP1 and MLP1-mutants in pET28a (bacterial expression - protein purification) and pRep4 (yeast expression - tRNA mediated suppression assay). Restriction enzyme cleavage sites are underlined. 6X His tag is shown in bold and blue. Point mutations are shown in bold and pink.			
MLP1 Sall 6X His For	pRep4	5'	GCGCGGTCGACATG <u>CACCATCACCATCACCAT</u> TCAATCAATAAAGAAGAGGTAATAAAGC
MLP1 Q11A/Y14A Sall 6X His For	pRep4	5'	GCGCGGTCGACATG <u>CACCATCACCATCACCAT</u> TCAATCAATAAAGAAGAGGTAATAAAG GCA GTCGAA GCCT ATCTGTGCGACAAGAAGTGGTAAATG
MLP1 95 Sall 6X His For	pRep4	5'	GCGCGGTCGACATG <u>CACCATCACCATCACCAT</u> GACCCCGCTGAAGCTGCAGAAAAAGAAGCGCGTG
MLP1 NheI For	pET28a	5'	GCGCGGCTAGCATGTCAATCAATAAAGAAGAGGTAATAAAGCAAGTC
MLP1 Q11A/Y14A NheI For	pET28a	5'	GCGCGGCTAGCATGTCAATCAATAAAGAAGAGGTAATAAAG GCA GTCGAA GCCT ATCTGTGCGACAAGAAGTGGTAAATG
MLP1 95 NheI For	pET28a	5'	GCGCGGCTAGCATGGACCCCGCTGAAGCTGCAGAAAAAGAAGCGCGTG
MLP1 BamHI Rev	pET28a/pRep4	5'	GCGCGGGATCCTCAAAGTTAAGTTTTCTCAAAGTTTGG
MLP1 95 BamHI Rev	pET28a/pRep4	5'	GCGCGGGATCCTCAGTCAACTGCTTCTTCGCTGGAATGGGTTTTATTTCCG
MLP1 250 BamHI Rev	pET28a/pRep4	5'	GCGCGGGATCCTCAATCTGTATATTTTTCTCCCGAATTCGAATTC

Oligonucleotides used to generate T7 DNA template for <i>in vitro</i> transcription of tRNAs for electromobility shift assays (EMSAs). T7 promotor sequence is shown in bold and blue.	
Leu AAG DNA template	5' GATGAAGTGCCGAGCGGTCTAAGCGGTAGATTAAGGCTCTATTCGAAAGGGCGCGAGTTCGAATCTCGCCTTCATCA
Leu AAG T7 promoter + 5'-leader For	5' GCTAATACGACTCACTAT <u>Agagcga</u> GATGAAGTGCCGAGCGGTC
Leu AAG T7 promoter + mature For	5' GCTAATACGACTCACTAT AGATGAAGTGCCGAGCGGTC
Leu AAG 3'-trailer + (U) ₅ Rev	5' aaaaatgTGATGAAGGCGAGATTCGAAC
Leu AAG mature Rev	5' TGATGAAGGCGAGATTCGAAC
Gln TTA DNA template	5' GGTTCATAGTATAGTGGTTAGTACTGGGACTTTAAATCCCTTGACCTGGGTTGCAATCCAGTGGGACCT
Gln TTA T7 promoter + 5'-leader For	5' GCTAATACGACTCACTAT <u>Agcatgtcc</u> GGTTCATAGTATAGTGGTTAGTAC
Gln TTA T7 promoter + mature For	5' GCTAATACGACTCACTAT AGGTTCCATAGTATAGTGGTTAGTAC
Gln TTA 3'-trailer + (U) ₅ Rev	5' aaaaagcAGGTCCCACTGGGATTCGAACCC
Gln TTA mature Rev	5' AGGTCCCACTGGGATTCGAACCC

Oligonucleotides used for cloning MLP1 partial knockout strain		
MLP1 5'-flanking KpnI For	pNeo4	5' CGCGGGTACCGGTATGTTGTTTGGATCTTATTAGAATCAC
MLP1 5'-flanking XhoI Rev	pNeo4	5' CGCGCTCGAGTTATTCAAAGTAGTAACTCCGAGTCTCTT
MLP1 3'-flanking NotI For	pNeo4	5' CGCGCCGCCGCTCACGAAATGGTAAATAAATAAATACGATT
MLP1 3'-flanking SacI Rev	pNeo4	5' ATTAGAGCTCAGTATATTAAGCAATCAATAAATAAATAA

DNA probes used for northern blotting. Anticodon sequence shown in bold and 5'-leader, 3'-trailer and intron sequences in lowercase if applicable.

<i>T. thermophila</i> U5 snRNA	5' CACATTGAAAAACCCAGCTCACGG
<i>T. thermophila</i> 5.8S rRNA	5' CTGCAATTCGCATTGCGT
<i>T. thermophila</i> Tyr GTA pre-tRNA 3'-trailer	5' aaaaaaaaaTCCGAAGCTCCCGGG
<i>T. thermophila</i> Tyr GTA mature tRNA	5' GACCACCGGATTACAGTCCG
<i>T. thermophila</i> Ile TAT pre-tRNA intron	5' GACacacttttataaggttggtcagcaaa TATAATC
<i>T. thermophila</i> Ile TAT/Leu TAA pre-tRNA 3'-trailer	5' aaaaaaaaaagTAGCTCAGAGAGGGTTC
<i>T. thermophila</i> Ile TAT/Leu TAA pre-tRNA 5'-leader	5' ACCACTGAGCCtttcaaag
<i>T. thermophila</i> Leu TAA pre-tRNA intron	5' GACactcttttctaaatgtaggagatgaacG
<i>T. thermophila</i> Val CAC pre-tRNA intron	5' TTAAGGTTtattagacct CGTGATA
<i>T. thermophila</i> Val CAC pre-tRNA 3'-trailer	5' aaaaaaGTCGACACTAGGGTTTGAA
<i>T. thermophila</i> Val CAC pre-tRNA 5'-leader	5' ACTACTTCACCATGCCGACtatagcag
<i>T. thermophila</i> Arg TCT mature tRNA	5' CAACCAAGGTGGGACTCGAACCAC
<i>S. pombe</i> U5 snRNA	5' CTGGTAAAAGGCAAGAAACAGATACG
<i>S. pombe</i> suppressor Ser TCA pre-tRNA intron	5' gaatacagga TTGA GTCT
<i>S. pombe</i> suppressor Ser TCA pre-tRNA intron (unlabelled probe)	5' gaatacagga TTCA GTCT
<i>S. pombe</i> suppressor Ser TCA mature tRNA	5' TGA AGTCTAACTCCTT
<i>S. pombe</i> suppressor Ser TCA mature tRNA (unlabelled probe)	5' TCA AGTCTAACTCCTT
<i>S. pombe</i> Lys CTT pre-tRNA intron	5' CTTCTGATaccattcg TAAG AGTC
<i>S. pombe</i> Lys CTT pre-tRNA 3'-trailer	5' aaattaaccTCCCAAG
<i>S. pombe</i> Lys CTT pre-tRNA 5'-leader	5' ATTGAGCCACTCGGGAcgctt
<i>S. pombe</i> Lys CTT mature tRNA	5' CTCCAAGGCGAGACTCGAACTCGCAA

DNA probes used for pre-tRNA intron-specific PCR amplification for Sanger sequencing RIP-3'-RACE

Lys CTT pre-tRNA intron	5' TCTGACTCTTATGATGGTAATCAGA
Tyr GTA pre-tRNA intron	5' CCGGCTGTAATTATAAAGATACC

RNA for electromobility shift assays (EMSAs)

U10	5' rUrUrUrUrUrUrUrUrUrU
U10-3'-P	5' rUrUrUrUrUrUrUrUrUrU/3Phos/
4U (wild type)	5' rCrUrGrCrUrGrUrUrUrU
U ₁ C	5' rCrUrGrCrUrGrUrUrUrC
U ₂ C	5' rCrUrGrCrUrGrUrUrCrU
U ₃ C	5' rCrUrGrCrUrGrUrCrUrU
4C	5' rCrUrGrCrUrGrCrCrCrC

Table S6. NCBI accession numbers of primary amino acid sequences used for conservation analysis

	La	Rex1p	RNase Z / ELAC2	LSm2	LSm3	LSm4	LSm5	LSm6	LSm7	LSm8	
Vertebrates	<i>Homo sapiens</i>	NP_003133.1	NP_065746.3	NP_060597.4	NP_067000.1	NP_055278.1	NP_036453.1	NP_036454.1	NP_009011.1	NP_057283.1	NP_057284.1
	<i>Mus musculus</i>	NP_033304.1	NP_080128.2	NP_001349912.1	NP_001103571.1	NP_080585.1	NP_056631.2	NP_079796.1	NP_001177933.1	NP_001346078.1	NP_598700.1
	<i>Xenopus laevis</i>	NP_001081021.1									
	<i>Danio rerio</i>	NP_955841	NP_001119888.1	NP_001243133.1							
Invertebrates	<i>Caenorhabditis elegans</i>	NP_491411.1	NP_498135.2	NP_001023110.1							
	<i>Drosophila melanogaster</i>	NP_477014.1	NP_001034073.1	NP_724916.1							
Amoebozoa	<i>Dictyostelium discoideum</i>	XP_640625.1									
Kinetoplastide	<i>Trypanosoma brucei</i>	XP_822491.1									
Embryophytes	<i>Arabidopsis thaliana</i>	NP_567904.1									
		NP_178106.2									
	<i>Physcomitrella patens</i>	XP_024359573.1									
Chlorophytes	<i>Chlamydomonas reinhardtii</i>	XP_042914771.1									
Fungi	<i>Schizosaccharomyces pombe</i>	NP_593315.1	NP_594627.2	NP_595514.1	NP_588459.1	NP_595747.1	NP_001342832.1	NP_596373.1	NP_594380.1	NP_588340.1	NP_588509.1
	<i>Saccharomyces cerevisiae</i>	NP_010232.1			NP_009527.1	NP_013543.3	NP_011037.3	NP_011073.1	NP_010666.2	NP_014252.2	NP_012556.2
Alveolates	<i>Paramecium tetraurelia</i>	XP_001439258.1	XP_001435945.1	XP_001346883.1							
	<i>Tetrahymena thermophila</i>	XP_001019287.2	XP_001033374.2	XP_001031902.2	XP_012654399	XP_012656392.1	XP_001008519.2	XP_012655775.1	XP_012653392.1	XP_001031297.1	XP_001470817.1

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