Video 1: Muscle contraction of the triceps in response to acetylcholine was observed as early as E13.5. The skin was removed from $ScxGFP^+$ (green) E12.5 – E14.5 forelimbs and exposed to 20 µm acetylcholine. Movement was observed at E13.5 and later in control limbs but not in *mdg* limbs. Video is shown at 1× and 4× speed, forward (\bowtie) and reverse (\bowtie). Scale bars: 500 µm. Representative videos from n = 3 biological replicates.

Video 2: The ectopic lateral triceps in $Prrx1^{CreTg/+}$; $Tbx3^{fl/fl}$ limbs contracted in response to acetylcholine at E18.5. The skin was removed from $ScxGFP^+$ (green) E18.5 forelimbs and exposed to 10 µm acetylcholine. Contraction was observed in the control and $Prrx1Cre^{Tg/+}$; $Tbx3^{fl/fl}$ ectopic lateral triceps (arrows). No movement was detected in *mdg* or $Prrx1Cre^{Tg/+}$; $Tbx3^{fl/fl}$; *mdg* limbs. Video is shown at 1× and 4× speed, forward ()) and reverse (). The end of the video shows control and *mdg* limbs in the same dish, exposed to the same spike of acetylcholine. Closed arrowheads: lateral triceps; open arrowheads: long triceps. Scale bars: 500 µm. Representative videos from n = 3 biological replicates.

Video 3: EMILIN1+ fibers were absent from the belly of the ectopic lateral triceps muscle in <u>Prrx1Cre^{Tg/+};Tbx3^{fl/fl};mdg limbs</u>. E14.5 decellularized forelimbs were stained for EMILIN1 (green), which marked fibers that were found within the lateral triceps of control, *mdg* (tendon: closed arrowhead) and Prrx1Cre^{Tg/+};Tbx3^{fl/fl} (tendon: closed arrow) limbs. However, there were no fibers in the Prrx1Cre^{Tg/+};Tbx3^{fl/fl};mdg ectopic lateral triceps. Red: WGA; blue: THBS4. EMILIN1+ fibers were present in the long triceps (tendon: open arrowhead) of all genotypes. 10× decell, $z = 114 \mu m$. Scale bars: 100 μm . Representative videos from n = 3 biological replicates.

Table S1: Raw data supporting the E18.5 *mdg* versus control triceps muscle protein comparisons and methods details for primers, antibodies dilutions, decell protocol, and confocal settings.

<u>**Table S2**</u>: Raw data supporting the E14.5 $Prrx1Cre^{Tg/+}$; $Tbx3^{fl/fl}$; mdg versus control limb protein comparisons.

<u>**Table S3**</u>: Raw data supporting the E18.5 $Prrx1Cre^{Tg/+}$; $Tbx3^{fl/fl}$; mdg versus control triceps protein comparisons.

<u>**Table S4:**</u> Raw data supporting the P21 $Prrx1Cre^{Tg/+}$; $Tbx3^{fl/fl}$; mdg versus control triceps protein comparisons and slow verse fast muscle classification support.



Figure S1: Muscle contraction was required for MTJ maturation in the wrist, expansion of COL22A1 at the joint surface, and nerve patterning. (A, A', C, C') COL22A1 (red) was expressed on joint surfaces (*) within the elbow and wrist in both control and *mdg* limbs at E14.5 (WGA, blue). (B, B', D, D') COL22A1 was expanded around the humerus and ulna (B) and wrist bones (D) joint surfaces (*) in E18.5 controls, but not in the *mdg* limbs. Ossification centers (O) were still present. (E, E', G-G') COL22A1 and EMILIN1 (red) were observed in the E14.5 extensor carpi ulnaris (ECU) in both control and mdg limbs. THBS4 (green) was not MTJ-specific at E14.5. (F, F', H, H', I, I') COL22A1 (red) and THBS4 (green) were reduced at the MTJ of mdg E18.5 ECU when compared to controls. (J, K) EMILIN1⁺ tendons (red) were slightly reduced in thickness in the mdg triceps. (L, M) LAM⁺ (red) blood vessel patterning was similar in control and *mdg* triceps. (N-Q) The distribution of neurofilament⁺ nerves (green or depth projection) was expanded in LAMA2⁺ (blue) mdg triceps muscles (dotted line) compared to controls. Tendons labeled by ScxGFP (red). A-D': $10 \times$ decell, 3D rendered, $z = 342 \mu m$ (A-B'), $z = 496.2 \mu m$ (C, C', D, D'); E-F', H-M: $63 \times$ decell, 3D rendered, $z = 65 \mu m$ (E-F', H-I'), $z = 59 \mu m$ (G,G'), z = 102 μ m (J, K), 80 μ m (L,M); N-Q: 10× wholemount, z-projection, z =1130 μ m. Scale bars: 100 μ m. Representative images from n = 3 biological replicates.



Figure S2: Development of the MTJ, tendon, and cartilage at E13.5-E14.5 in *Prrx1Cre^{Tg/+};Tbx3^{fl/fl}* limbs. (A-D') At E13.5, COL22A1 (green or depth projection) was deposited at the MTJ of the lateral and long triceps. No discernable COL22A1 was observed at the ectopic insertion. (E-F) *Scx*GFP (red) and SOX9 (green) did not co-localize at the ectopic insertion at E13.5, but did co-localize at the enthesis of the normal insertion. MY32 = muscle (blue). (G-I') *Scx*GFP⁺ (red) cells were inconsistently observed at the TNC⁺ (green) ectopic insertion at E14.5, and was not observed in the same area within control limbs. MY32 = muscle (blue). A-D': 25× decell, 3D rendering, $z = 50 \ \mu m$ (A-B), $z = 56 \ \mu m$ (C-D); E, F: 10× wholemount, z-projection, $z = 458 \ \mu m$ (E, F); G-H': 10× wholemount, z-projection, $z = 381.6 \ \mu m$. Scale bars: 100 $\ \mu m$. Representative images from n = 3 biological replicates.



Figure S3: Development of the nerve and cartilage from E13.5 – E18.5 in *Prrx1Cre^{Tg/+};Tbx3^{fl/fl}* limbs. (A) Volcano plot of matrisome components identified in control and *Prrx1Cre^{Tg/+};Tbx3^{fl/fl}* E18.5 triceps forelimbs (**Table S3**). Grey lines indicate \geq 2-fold change and p < 0.05 (two-tailed t-test). Average of n = 3 biological replicates. (**B-G**) Neurofilament⁺ (depth projection) nerves (arrowheads) were not observed at E13.5 in either the *Prrx1Cre^{Tg/+};Tbx3^{fl/fl}* or control muscles. At E14.5 and E18.5, neurofilament⁺ nerves were observed within the lateral triceps (dotted line), and were oriented perpendicular to the direction of muscle fibers, even in the mispatterned *Prrx1Cre^{Tg/+};Tbx3^{fl/fl}* limbs. (**H-I**") While overlap between the TNC⁺ (red) tendon and COL2A1⁺ (green) cartilage was observed in the normal enthesis at E18.5 (*), COL2A1 did not co-localize with the TNC⁺ ectopic insertion (arrows). LAMA2 = muscle (blue). B-G: 10× wholemoutt (B, C), SeeDB-cleared (D-G), z-projection, z = 280 µm (B, C), z = 427 µm (D, E), z = 802 µm (F, G); H-I": 10× cryosections (H, I); 63×, cryosections (H', H'', I', I''). Scale bars: 100 µm. Representative images from n = 3 biological replicates.



Figure S4: There was increased fat, POSTN, and TNC accumulation in the *Prrx1^{CreTg/+}; Tbx3^{fl/fl}* ectopic triceps. (A-B') The *Prrx1Cre^{Tg/+};Tbx3^{fl/fl}* lateral triceps had increased fat (*) compared to the controls. (C-C'''') In some P21 limbs, it appeared an ectopic origin was also present, connected to the elbow by a THBS4⁺/*Scx*GFP⁺ tendon (*). Both the long triceps insertion and ectopic triceps origin muscle fibers terminated in a mature MTJ. (D-K') POSTN and TNC were enriched near the *Scx*GFP⁺ ectopic insertion. COL5 was found thorough the interstitial spaces surrounding the muscle. C: 10× wholemount, z-projection, $z = 1687 \mu m I$; $z = 1160 \mu m (D, E)$, C'-C''' 63× wholemount, z-projection, $z = 48 \mu m (C'-C''')$, $z = 51 \mu m (D', D'', E', E'')$, F-K: 10× cryosections, F'-K': 10× cryosections. Scale bars: 1 mm (A-B', C-K); 100 $\mu m (C'-C''', D', D'', E', E'', F'-K')$. Representative images from n = 3 biological replicates.



Figure S5: The ectopic lateral triceps muscle, MTJ, and tendon were not observed in E18.5 *Prrx1Cre^{Tg/+};Tbx3^{fl/fl};mdg* limbs, but blood vessel structure was maintained. (A-D) LAMA2⁺ (blue) lateral triceps muscle and THBS4⁺ (green)/*Scx*GFP⁺ (green) tendon formed in the proper location in control and *mdg* limbs. The ectopic lateral triceps muscle was present in *Prrx1Cre^{Tg/+};Tbx3^{fl/fl}* but not *Prrx1Cre^{Tg/+};Tbx3^{fl/fl}*;mdg limbs. (E-H) COL22A1⁺ (depth projection) was present in the lateral and long triceps and ectopic insertion. In contrast, COL22A1 was weakly found at the MTJ of the long triceps tendon and not found at the location of the ectopic insertion in *Prrx1Cre^{Tg/+};Tbx3^{fl/fl};mdg* limbs. * Indicates joint surface. (I-L') CD31⁺ (red) blood vessels were present in the location of the lateral triceps in *mdg* and *Prrx1Cre^{Tg/+};Tbx3^{fl/fl};mdg* limbs. TNC = green; MY32 = blue. A-D, I-L': 10× wholemount, z-projection, z = 387 µm (A, C, D), z = 501 µm (B), z = 497 µm (I), z = 1130 µm (J), z = 1009 µm (K), z = 602 µm (L); E-H: 10× decell, 3D rendering, z = 570 µm (E-H). Scale bars: 100 µm. Representative images from n = 3 biological replicates.



Figure S6: No lateral triceps or COL22A1⁺ MTJ was present in the E16.5 *Prrx1Cre^{Tg/+};Tbx3^{fl/fl};mdg* lateral triceps ectopic insertion. (A-H) Lateral triceps muscle (MY32, LAMA2 = blue) and tendon (TNC, THBS4 = green; *Scx*GFP⁺ = red) formed in the proper location in the control and *mdg* limbs. The lateral triceps was ectopically located in *Prrx1Cre^{Tg/+};Tbx3^{fl/fl}* limbs, but was not present in *Prrx1Cre^{Tg/+};Tbx3^{fl/fl};mdg* limbs. (I-L) COL22A1⁺ MTJs were present in the long and lateral triceps in both control and *Prrx1Cre^{Tg/+};Tbx3^{fl/fl}* limbs at E16.5. COL22A1 was reduced in the lateral and long triceps *mdg* MTJs. In *Prrx1Cre^{Tg/+};Tbx3^{fl/fl};mdg* limbs, COL22A1 was lost at the site of the ectopic insertion, but was present in the long triceps. * Indicates joint surface. A-N: 10× wholemount, z-projection, $z = 513 \mu m$ (A-D), $z = 547 \mu m$ (E-G); H-L: 10× decell, 3D rendering, $z = 399 \mu m$. Scale bars: 100 µm. Representative images from n = 3 biological replicates.



 Figure S7: The MTJ was not present in the E14.5 $Prrx1Cre^{Tg/+}$; $Tbx3^{fl/fl}$; mdg ectopic lateral triceps. (A-H) COL22A1 (green) was expressed in MTJs on the ulnar side of the limb, such as the extensor carpi ulnaris and the bicep brachial (u = ulna, r = radius, h = humerus). (I-L) EMILIN1 (green) and WGA (red) was found in the muscle belly fibers of the lateral triceps (dotted line) in control, $Prrx1Cre^{Tg/+}$; $Tbx3^{fl/fl}$ and mdg, but not $Prrx1Cre^{Tg/+}$; $Tbx3^{fl/fl}$; mdg limbs. (M-P) EMILIN1 was present in control and $Prrx1Cre^{Tg/+}$; $Tbx3^{fl/fl}$ tendons, whereas it was reduced in mdg and absent in the ectopic insertion in the $Prrx1Cre^{Tg/+}$; $Tbx3^{fl/fl}$; mdg limbs at E14.5. A-H, M-P: 25× decell, 3D rendering, $z = 50 \ \mu m$ (A-D), $z = 44 \ \mu m$ (E-H), $z = 97 \ \mu m$ (M-P), I-L: 10× decell, z-projection, $z = 114 \ \mu m$. Scale bars: 100 μm . Representative images from n = 3 biological replicates.