Phylogenetically conservative trait 1 correlation: quantification and 2 interpretation 3 4 Version of 2022-12-27 5 6 7 Mark Westoby, School of Natural Sciences, Macquarie University, Sydney NSW 2109 8 Australia 9 10 Luke Yates, ARC Centre of Excellence for Plant Success in Nature and Agriculture, University of Tasmania, Sandy Bay TAS 7005 Australia 11 12 Barbara Holland, School of Natural Sciences, University of Tasmania, Sandy Bay TAS 13

- 14 7005 Australia
- 15
- 16 Ben Halliwell, ARC Centre of Excellence for Plant Success in Nature and Agriculture,
- 17 University of Tasmania, Sandy Bay TAS 7005 Australia
- 18
- 19 Corresponding author mark.westoby@mq.edu.au
- 20
- 21 Main text 4439 words, plus 1 Table, 4 Boxes, 2 Figures
- 22 Supplementary 1: Simulations
- 23 Supplementary 2: Mathematical aspects of phylogenetic mixed models
- 24 Supplementary 3: R code for the simulations reported in Figs 1 and 2
- 25
- 26 Short title: conservative trait correlation
- 27
- 28 Keywords: phylogenetic correction, trait correlation, evolutionary divergence, phylogenetic
- 29 niche conservatism, independence, partial correlation

30

31 Data statement: R code used for simulations is at Supplementary 3.

- 33 Author contributions: The manuscript developed from discussions between Westoby and
- 34 Halliwell. Westoby drafted successive versions, Halliwell, Yates and Holland contributed
- 35 expertise in multivariate phylogenetic mixed models, all authors critiqued drafts.

36 Abstract

- Correlation across species between two quantitative traits, or between a trait and a habitat
 property, can suggest that a trait value is effective in sustaining populations in some
 contexts but not others. It is widely held that such correlations should be controlled for
 phylogeny, via phylogenetically independent contrasts PICs or phylogenetic generalised
 least squares PGLS.
- 42 2. Two weaknesses of this idea are discussed. First, the phylogenetically conservative share
 43 of the correlation ought not to be excluded from consideration as potentially ecologically
 44 functional. Second, PGLS does not yield a complete or accurate breakdown of A-B
 45 covariation, because it corresponds to a generating model where B predicts variation in A
 46 but not the reverse.
- 3. Multi-response mixed models using phylogenetic covariance matrices can quantify 47 conservative trait correlation CTC, a share of covariation between traits A and B that is 48 phylogenetically conservative. Because the evidence is from correlative data, it is not 49 50 possible to split CTC into causation by phylogenetic history versus causation by continuing reciprocal selection between A and B. Moreover, it is quite likely biologically 51 52 that the two influences have acted in concert, through phylogenetic niche conservatism. 53 4. Synthesis: The CTC concept treats phylogenetic conservatism as a conjoint interpretation 54 alongside ongoing influence of other traits. CTC can be quantified via multi-response
- 55 phylogenetic mixed models.
- 56

57 Introduction

Ecological research often takes an interest in correlations across species between two traits, 58 or between a trait and a property of the species habitat. For example, seed size is correlated 59 (fairly loosely, $r^2 = 0.29$) with the size reached by species as adults (Falster, Moles, and 60 Westoby 2008). One motivation for investigating how closely traits are correlated is simply 61 to understand variation across the world's species, and to quantify how traits might be 62 clustered together into spectra of variation. For example, a unified size spectrum has been 63 64 suggested (e.g. Díaz et al. 2016) that embraces both seed size and adult size. Another 65 motivation is that an observed correlation might be consistent with some proposed mechanism connecting the two traits, or alternatively a lack of correlation might argue 66 67 against a mechanism. For example, it can be suggested that taller species typically suffer 68 more competitive mortality between seedling and reproductive stages, and this puts a stronger 69 selective premium on large seed size (Falster, Moles, and Westoby 2008). 70 71 A present-day correlation between seed size and potential plant size across species can be 72 interpreted as caused by trajectories of change through past evolution. Equally, the past 73 trajectories can be interpreted as movement toward evolutionary attractors, produced by an

ecological mechanism that exerts continuing selective pressure in the present day. Either of

those versions of causation are consistent with observed correlations between traits, or

- 76 between a trait and habitat.
- 77

78 It is important to be clear that correlations across species come from observational or survey 79 evidence. They can offer support for some proposed mechanism or argue against it, but they 80 can not significance-test them in the same sense as manipulative experimental treatments can. 81 In manipulative experiments, the treatment is cause and the outcome is effect, and other 82 factors are controlled or randomized so that each replicate yields an independent item of evidence for the link between cause and effect. Because the items of evidence are 83 84 independent, a P-value for the ensemble of events can be calculated with confidence. Whereas in survey evidence, some unmeasured or uncontrolled variable might be creating a 85 86 correlation between the two focal traits, or counteracting it.

87

Where investigators have thought about third and fourth variables as possible influences, and
have been able to obtain measurements for them, a more limited sort of independence can be
obtained by controlling or partialling for these third or fourth variables, or equivalently by

applying multiple regression. Residuals are obtained for the focal variables after regression
on the covariates, and correlations between the residuals are then inspected. But this is still a
very different sort of independence compared to the evidence that emerges from a
manipulative experiment. If an A-B correlation disappears after partialling for C, it still
remains a possibility that C was a secondary correlate and the true mechanism runs between
A and B. Plus there remain variables D, E, F and so forth that might have been the true cause
but were not measured or not even thought of.

99 Structured causal modeling SCM (Pearl 2009) or graphical causal modeling (Cronin and 100 Schoolmaster Jr. 2018) is a framework that purports to determine cause-and-effect 101 relationships from observational data (Arif and MacNeil 2022). However, the conditions for identifying causation unambiguously are stringent. The causal maps are required to be 102 directed acyclic graphs (DAGs), with no recursion to variables earlier in the causal chain. It 103 must be possible to list all competing causal hypotheses in order to compare them, and each 104 105 must correspond to a different chain of causation between variables. In our opinion (contra 106 Cronin and Schoolmaster Jr. 2018), these conditions are not ever met by the situations of 107 interest here, coordination across species among traits and habitat and their relationship to 108 phylogeny. Coordination between traits happens because the current value of each trait influences natural selection on the other (recursion). Traits also influence the habitats where 109 110 the clade is successful, and habitat in turn exerts natural selection on the traits (again recursion). A map leading from clade membership to trait values always has alternative 111 112 causal interpretations: (1) that traits are intrinsically slow to change so that clade signal 113 remains, and (2) niche conservatism, that there is continuing ecological selection from other 114 traits in combination with habitat.

115

It is widely held that correcting or controlling or accounting for phylogeny (methodology
summarized in Box 1) should be mandatory when ecologists consider present-day
functionality of traits in combination with each other or with different environments (e.g.
Losos 2011; Garamszegi 2014; Swenson 2020; Revell and Harmon 2022). Reviewers and
editors of ecology journals commonly require authors to control for phylogeny. Despite this
strong majority view insisting on the practice during review, experts have raised substantial
questions about what is achieved by controlling for phylogeny (Box 2).

Correcting an A-B relationship for phylogeny uses the same logic as partialling it for a 124 continuous variable C. The commonest justification why phylogenetic correction should be 125 mandatory is to say that related species are not independent (Felsenstein 1985, and very often 126 repeated up to the present day, e.g. Symonds and Blomberg 2014). An A-B relationship 127 controlled for phylogeny is often interpreted as a corrected or improved version of the simple 128 129 cross-species relationship. This interpretation is not correct. Rather, phylogenetically controlled relationships measure different properties of the data, compared to relationships 130 across present-day species. They address a different question (see section below "What does 131

- 132 phylogenetic generalised least squares quantify?").
- 133

134 A statistical method corresponds to a generating model. Its equations, variables and

135 probability distributions express models for causation or for prediction. Only if the generating

136 model is well aligned with a biological hypothesis will a clear answer be delivered. The

137 statistical models fitted, the causal or predictive maps hypothesized, and the biological

- 138 questions of interest are all aspects of the same issue.
- 139

140 The PGLS and PIC methods mainly used for controlling for phylogeny correspond to 141 particular generating models. Our main aim here is to show that these generating models do not necessarily correspond to questions that ecologists want to ask. Further, the fact that they 142 143 are couched in terms of least squares regression of A against B does not adequately represent a generating process whereby A and B reciprocally influence each other. Another aim here is 144 145 to put forward multi-response or multivariate phylogenetic mixed models (MR-PMM). These 146 treat A and B as joint responses and partition the different correlations in a way that does not 147 treat phylogeny and present-day function as alternative interpretations. MR-PMM are not new (Lynch 1991; Housworth, Martins, and Lynch 2004), but have not come into common 148 149 use in ecology.

150

151 Quantifying conservative trait correlation via multi-trait response152 models

153 The most straightforward reason why controlling for phylogeny should not be interpreted as 154 automatically correcting or improving an A-B relationship, is that present-day influence from 155 B and phylogenetic conservatism overlap as explanations for variation in A. Controlling for

156 phylogeny is advocated with a view to discarding, or partialling out, A-B covariation that is

157 phylogenetically conservative from the A-B relationship. From the perspective of

understanding present-day ecological differences across species, this means that differences

- 159 between major clades are downweighted as contributors. To the extent differences between
- 160 major clades are important in present-day ecology, it risks throwing the baby (or large parts
- 161 of it) out with the bathwater (Hansen 2014; de Bello et al. 2015).
- 162

A constructive solution to this problem of interpretation lies in multi-response phylogenetic 163 mixed models (MR-PMM; Halliwell, Yates, and Holland 2022). These models decompose 164 165 trait-level covariance and variance into phylogenetic and independent components (details in Box 3 and Table 1). A component of A-B correlation that is also phylogenetically structured 166 can be identified and quantified. We refer to this quantity as the conservative trait correlation 167 CTC. In these MR-PMM, as in PGLS, a matrix of covariances expected from a phylogenetic 168 generating model appears as part of the residual structure on the right hand side. The key 169 170 difference from PGLS is that traits A and B are jointly modelled as response variables on the left hand side (hence the name multi-response), and both their phylogenetic and independent 171 172 correlations are parameters to be estimated. This makes it possible to decompose the A-B 173 correlation into a component that is also phylogenetically structured (conservative trait 174 correlation) and a component that is independent of phylogeny (Table 1). It has also the effect of treating the A-B relationship as a question of how they are coordinated rather than 175 176 as a question of how B predicts A, analogous to standardized major axis (SMA) relationships 177 rather than to ordinary least squares (OLS) regression (Warton et al. 2006). This will be 178 appropriate for most evolutionary questions, since selective influences between traits or 179 between a trait and a habitat property will be reciprocal.

180

181 From a statistical point of view, conservative trait correlation CTC is A-B covariation where 182 for each trait, phylogeny and the other trait jointly are associated. It is not possible to separate them. From the point of view of interpreting biological mechanism, it is quite likely that 183 phylogeny and each trait have acted in concert on the other trait, via phylogenetic niche 184 conservatism (next section). MR-PMM identifies phylogenetically-conservative covariation 185 186 between traits A and B (Table 1), and remains agnostic whether this covariation should be attributed to phylogenetic history or to continuing reciprocal selection between the traits, or 187 to the synergy between those two, known as niche conservatism. This is more constructive 188 than the PGLS partitioning, which is used with a view to separating phylogenetically-189

190 conservative covariation from the estimate of the A-B relationship (see also section on

191 PGLS).

192

193 Phylogenetic niche conservatism

194 Should the present-day pattern of trait-combinations across species be interpreted as caused by trajectories of change through past evolution? Or should the past trajectories be interpreted 195 196 as movement toward evolutionary attractors, which continue to be attractors in the present 197 day? A correlation between traits, or between a trait and habitat, can be interpreted in either 198 of these ways. Traits of ecological importance are expected often to evolve in a 199 phylogenetically conservative way. If a new ecological opportunity or niche arises, successful 200 occupants are most likely to be drawn from clades that already possess appropriate trait-201 combinations. Descendants from a clade are most likely to be successful in habitats or ways of life similar to those the clade is already adapted for. Through this phylogenetic niche 202 203 conservatism, large shares of present-day adaptation and phylogeny can often be bound together as a unified causal process. Differences between major clades are often important 204 205 contributors to the observed variation across ecological strategies. Phylogenetic history and present-day ecological competence are complementary explanations, not mutually exclusive 206 207 alternatives.

208

Consider the simulations described in Fig 1. In Fig 1a there is an overall correlation between
traits A and B, but the correlation is generated from the difference between two major clades,
and no correlation has been simulated within each clade. A similar pattern was shown in
Felsenstein's (1985) Fig 7. His interpretation was that "It can immediately be seen that the
apparent relationship is illusory". However, Price (1997) showed that a similar pattern
could in fact be produced by continuing selective forces. Fig 1b illustrates this, using a

- simulation driven by the same principles as Price.
- 216

In Fig 1b the broken line circumscribes an ecological attractor, a region of trait combinations in niche space that are ecologically competent. The shape of this region is of high interest for ecologists. Indeed, this is the motivation for looking at scattergrams of one trait vs another. It is supposed for both (a) and (b) of Fig 1 that orange and blue symbols represent sister clades that have diverged in trait space toward lower left and upper right. Panel (a) then assumes Brownian motion, while (b) assumes that new species can emerge only within the viable traitspace and tend to be drawn from the existing clade that is nearest in trait space. In both cases

- 224 each clade is phylogenetically conservative. In (a) the conservatism takes the form of
- sluggish Brownian motion. (If the Brownian motion is rapid, then the historical difference
- between the two major clades is quickly washed out.) In (b) conservatism arises from a
- 227 constrained range of ecological possibilities.
- 228
- The point is that the observed pattern across present-day species cannot help to decide which of these causative interpretations is more likely. Further, the process in Fig 1b is both phylogenetically conservative and also caused by ecological constraints continuing into the
- pryrogenetically conservative and also caused by ecological constraints continuing into the present day. Data analysis should not treat these as competing alternatives. Better for it to
- identify conservative trait correlation, the share of trait correlation that might be attributed
- either to phylogenetic history or to continuing functionality or to a combination of the two.
- 235

The question how much to interpret functional traits in terms of past history versus in terms

- of present-day competence itself has a history (brief summary in Box 4). To some extent it
- reflects tension between the outlooks of evolutionists and ecologists.
- 239

240 When ecological selection has favoured high trait A in conjunction with high trait B through the length of phylogenetic history, as well as in the present day (as in Fig 1c), then ordinary 241 242 regression across species and PGLS will yield similar results (Fig 2, simulation S1), because the trait correlation pattern across phylogenetic divergences is similar to the pattern across 243 244 present-day species. This is a very common case in real datasets (Ackerly 1999; Carvalho, 245 Diniz-Filho, and Bini 2006). Nevertheless, this similarity should not be the basis for choosing 246 OLS in preference to PGLS or vice versa. These two analyses, and also MR-PMM, are 247 different in what features of the data they model. Analyses should be chosen to match the 248 assumptions of the generating model and the question being addressed, even though OLS and PGLS quite often yield similar slopes. 249

250

Controlling for phylogeny does not confer strong-sense independence The most common justification why controlling for phylogeny should be mandatory is to say that related species are not independent (Felsenstein 1985, and very often repeated up to the present day). This justification implies that independence is restored by controlling for phylogeny. But that implication is only correct in a very limited sense.

256

Independence is used with two meanings. The looser meaning is simply "uncorrelated". Seed
size can be said to lack independence from plant asymptotic height. Independence can be said
to be restored by replacing the absolute seed size with residuals around a regression of seed
size on plant height. However, this is a conditional independence, from a fitted function of
plant height only, not from all possible confounding variables.

262

The tighter meaning of independence is about formally significance-testing a hypothesized 263 264 causal mechanism. If causal events are independent, their probabilities can be multiplied to yield the probability of the ensemble of events. When phylogenetic correction is said to be 265 266 obligatory because cross-species correlations lack independence, the suggestion is that after correction, independence will be ensured in this tighter sense, and a reliable significance test 267 will ensue. But significance tests from survey or correlative data should not be interpreted as 268 clean tests of causation anyhow. An A-B correlation may be more or less consistent with a 269 270 proposed causation, but it does not provide significance-tested proof. Correcting for 271 phylogeny using structured residuals addresses one sort of cross-correlation, but not all. 272 There may always be further variables unmeasured or not even thought of that are correlated 273 with both A and B (Price 1997). And further, in a network of interconnected variables, 274 correlations do not by themselves resolve the pathways along which causation runs. 275 Criteria need to be invoked from outside the correlative data. Some causal pathways might 276

seem more plausible than others in light of known mechanisms, or parsimony can be invoked
in choosing among statistical models, or combinations of plausibility with parsimony (Yates,
Richards, and Brook 2021).

280

281 Independence in this sense of independent events showing causality can really only be assured in manipulative experiments. Treatments come before outcomes in an experiment's 282 283 timeline, so it is clear which is cause and which is effect. And factors other than treatments 284 are randomized or physically controlled, so each replicate becomes definitively an 285 independent instance of a treatment giving rise or failing to give rise to an outcome. In 286 situations such as social science surveys or comparisons across present-day species, there is 287 no way to assure independence in this rigorous sense (Hernán, Hsu, and Healy 2019). Significance tests should not be taken too literally when analysing survey data, but r^2 and 288

similar indices that quantify the strength of correlations are useful descriptors. Cross-species
relationships are correlations not causation, and remain so after adjusting for phylogeny.

To invoke independence in the context of estimating a P-value you need to specify what process is being tested for. Then the question is whether two or more events or links between variables are independent <u>as evidence for that process</u>. Independence is a property of the hypothesis as well as of the data structure. To say that past divergences are independent cases for a link between traits A and B while present-day species are not, is just another way of asserting that past divergences are a legitimate causative explanation while continuing present-day selection is not (Box 4).

299

300 Arguments over the primacy of causal processes cannot be resolved from data that are cross-

301 correlated. We should look to statistical methodologies that offer the most informative

302 decomposition of trait variance, without preferencing one causality over another. Causality

303 can then be interpreted in light of knowledge about physiological mechanisms, or

304 manipulative experiments demonstrating how particular trait values confer advantage

- 305 depending on other traits or on habitat.
- 306

307 What does phylogenetic generalised least squares quantify?

Saying that PGLS accounts for phylogeny does not tell us what it actually is. What is
measured by the PGLS slope and confidence intervals, and how should it be interpreted?

311 For phylogenetically independent contrasts PICs and for PGLS with λ set at 1, the slope

312 coefficient with associated confidence interval describes divergences in A as predicted from

313 divergences in B, taken across the population of all past divergences inferred at all the nodes

in the tree. The significance test (whether the confidence intervals on the slope span zero)

315 assesses consistency, the question whether trait divergences were correlated across most or

all of the nodes.

317

318 The interpretation of PGLS with λ estimated lies somewhere between the extreme cases of λ

319 = 0 (OLS) and $\lambda = 1$ (original PGLS). Fitted λ with a value less than 1 but still not zero can be

- interpreted as a rescaling of branch lengths in the phylogeny (Symonds and Blomberg 2014).
- 321 Low lambda elongates the terminal branches, reducing the expected similarity between

related species. With fitted λ the PGLS slope likely still reflects an ensemble of slopes across 322 all divergences, but with divergences calculated on a tree with branch lengths modified by λ . 323 324 PGLS assumes that the true generating process is consistent with 'B predicts A' and that B 325 has no phylogenetic signal. These assumptions are usually not true. Unless predictor B actually is free of phylogenetic signal, the slope is confounded with the phylogenetic 326 327 component of the residual variance (Supplementary 2, see also simulations in next section). This confounding will commonly have the result that some but not all of the phylogenetic 328 signal in the A-B relationship remains in the slope estimate. Analogous issues occur in spatial 329 330 statistics where environmental predictors with spatial signal are confounded with the spatial 331 component of the residual variance (Marques, Kneib, and Klein 2022; Warton 2022).

332

The focus of PICs and PGLS on describing correlation in past divergences accords with the 333 insistence of evolutionists that adaptation is defined as the selective circumstances when a 334 335 trait or trait value first came about (Box 4). PGLS does not directly measure the relationship between traits in the present day, unless $\lambda = 0$ when it becomes an ordinary least squares 336 337 regression. It is the nature of most trees that there are many nodes near the tips and rather few 338 deep in the tree. As a result, deep nodes have only a minor influence on the PGLS-estimated 339 relationship between divergence in A and divergence in B. But the consequences of a single deep divergence for the pattern across present-day species can sometimes be very substantial 340 341 (Fig 1). Consider for example the divergence between angiosperms and gymnosperms. As well as qualitative differences such as tracheids vs vessels for water transport, these two 342 343 major clades of seed plants have widely different strategies with regard to quantitative traits 344 such as vein density in the leaves, seed size and leaf mass per area (e.g. Ackerly and Reich 345 1999; Brodribb et al. 2005; Díaz et al. 2016). This only counts as one divergence among many in a PGLS, but it has large consequences in terms of ecological strategies sustained in 346 347 the present day.

348

349 The relationships quantified by OLS and by PGLS are different, OLS a pattern across 350 present-day species, PGLS a pattern across past evolutionary divergences (at least with $\lambda =$ 351 1). Biologically, these are naturally complementary questions, but they are different, and one 352 should not be seen as replacing the other. MR-PMM quantifies both types of relationship 353 within a single analysis, but not in quite the same way, since it models phylogenetic signal in 354 both traits and formulates the relationship as A-B coordination rather than as predicting A 355 from B.

356

Simulations to illustrate how MR-PMM compares to PGLS 357 358 To illustrate issues around niche conservatism and the decomposition provided by MR-PMM, 359 we simulated datasets of two traits A and B. For each simulation, A and B were given the 360 same independent or residual covariance, but different phylogenetic covariances (Supplementary 1 for details). We simulated 400 replicate datasets by generating random 361 362 pure birth trees of 200 taxa. The two traits A and B were simulated from the full crosscovariance structure of the MR-PMM, comprising phylogeny-independent covariances 363 364 crossed with phylogenetic covariances, rather than generating A as a scalar multiple of B, as is assumed in PGLS (e.g. Revell 2010). This structure allows the independent and 365 366 phylogenetic variance in A and B to be defined separately and explicitly. 367 MR-PMM successfully recovers the trait-level covariances of each generating model (Fig 2), 368 as expected since it corresponds to the generating model. Comparing these covariance 369 370 estimates with PGLS is more complicated since PGLS is a single-response model and does 371 not report covariances directly, only the β slope coefficient. To facilitate comparisons, the independent and phylogenetic variances of each trait were set to one which places the slope 372 coefficients on the same scale as the correlation coefficients (see Supplementary 1 for 373 374 details). This choice of scale means that the β_{OLS} estimate is approximately equal to the mean 375 of the simulation values for independent and phylogenetic correlation components, $\beta_{\lambda=1}$ is

approximately equal to independent component, and $\beta_{\lambda=MLE}$ attains an intermediate value depending on the estimated λ .

378

These simulations illustrate the following. First, when covariances between traits are similar 379 380 on the phylogenetic and independent level (Fig 2, S1), then $\beta_{\lambda=1}$, $\beta_{\lambda=MLE}$ and β_{OLS} are also similar. Biologically, this is a common outcome (Price 1997; Ackerly 1999; Carvalho, Diniz-381 Filho, and Bini 2006). Second, $\beta_{\lambda=1}$ has the effect of disregarding covariance associated with 382 phylogenetic history during calculation of the β coefficient. Our central point in this paper is 383 that phylogenetically-associated covariance should not be automatically set aside, because 384 385 niche conservatism is both phylogenetic and also represents selective attractors that continue into the present day (Figure 1). Third, in the extreme case where the correlation between a 386 pair of traits occurs exclusively on the phylogenetic level, $\beta_{\lambda=1}$ is likely to report no 387 388 relationship (Figure 2, S4). To the extent that differences between major clades are important in present-day ecology, this result represents a false negative. Fourth, optimising λ does not

resolve this problem, rather it represents a compromise between the assumptions of PGLS

and OLS. Finally, because $\beta_{\lambda=1}$, $\beta_{\lambda=MLE}$ and β_{OLS} are all products of single-response models,

they represent single-number summaries of the different components of A-B covariance

393 present in the data (Supplementary Information 2). This means they are poor approximations

of the true generating model when phylogenetic and residual covariances differ in sign (S3)

395 or even magnitude (S2).

396

397 Conclusion

Both evolutionary and ecological questions about traits are important, but they are not the same. For ecologists interested in the present-day relationship of traits to habitat or each other, phylogenetic correction has been justified largely from the perspective that trait correlation across species might be misleading. This formulation is missing the point from the outset. Correlative data are undoubtedly capable of being misleading, and need to be approached with that mindset. But it is wrong to think that controlling or accounting for phylogeny obviates the problem.

405

Phylogenetically independent contrasts PICs ask about the history of divergences at nodes.
The divergences, rather than the present-day species, are the population of interest. The
question whether divergences in A have been consistently associated with divergences in B is
a natural one for evolutionists to ask. It is complementary to the ecological question about
trait-combinations that are competent in the present day, but it is not the same question.

411

412 Phylogenetic generalized least squares PGLS is currently widely recommended and used.

413 When used with $\lambda = 1$, it is mathematically equivalent to PICs. However, the actual historical

414 divergences are not inspected or graphed as they are for PICs. The slope estimate with $\lambda = 1$

415 describes the power of divergence in B to predict divergence in A, across the ensemble of

416 divergences or nodes. As was the case for PICs, this slope estimate answers an interesting

417 question, but not the question how traits are related across present-day species.

418

419 When PGLS is used with λ fitted to the data, λ will usually lie intermediate between 0 and 1,

420 since for most traits there is some phylogenetic signal but not perfect correlation with a

421 phylogenetic generating model. The strength of the residual phylogenetic influence is then

422 measured via λ. The estimated slope is intermediate between the slope across divergences423 and the slope across present-day species.

424

425 Multi-response phylogenetic mixed models open a path to interpreting covariance structure

- 426 better in two ways, we believe. First, their generating model deals in A-B covariation, which
- 427 reflects the nature of reciprocal influences between traits and habitat more satisfactorily than
- 428 regression-style models predicting A from B. Second, they quantify the variance and
- 429 covariance components more comprehensively. In particular, they quantify conservative trait
- 430 correlation CTC, and remain agnostic about whether it is caused by history, by continuing
- 431 evolutionary attractors, or by both. Historical and present-day accounts of causation are, in
- 432 fact, complementary. Over evolutionary time, new ecological opportunities will very often
- 433 have been taken up by speciation from clades that already possess a configuration of traits
- 434 close to what will be most successful.

435 Acknowledgments

- 436 Ian Wright kindly put the authors in touch with each other. Warm thanks also to Daniel
- 437 Falster and David Warton for much discussion of this topic over the years. Halliwell and
- 438 Yates were partly funded by The Australian Research Council Centre of Excellence for Plant
- 439 Success in Nature and Agriculture (CE200100015).
- 440

441 Literature cited

- 442 Ackerly, D. D. 1999. "Comparative Plant Ecology and the Role of Phylogenetic
 443 Information." In *Physiological Plant Ecology*, edited by M.C. Press, J.D Scholes, and
 444 M.G. Barker, 391–413. Blackwell Science.
- Ackerly, D. D., and P.B. Reich. 1999. "Convergence and Correlations among Leaf Size and
 Function in Seed Plants: A Comparative Test Using Independent Contrasts." *American Journal of Botany* 86 (9): 1272–81.
- Arif, S., and M. A. MacNeil. 2022. "Applying the Structural Causal Model Framework for
 Observational Causal Inference in Ecology." *Ecological Monographs* n/a (n/a):
 e1554. https://doi.org/10.1002/ecm.1554.
- Bello, Francesco de, Matty P. Berg, André T. C. Dias, Jose Alexandre F. Diniz-Filho, Lars
 Götzenberger, Joaquín Hortal, Richard J. Ladle, and Jan Lepš. 2015. "On the Need for
 Phylogenetic 'Corrections' in Functional Trait-Based Approaches." *Folia Geobotanica* 50 (4): 349–57. https://doi.org/10.1007/s12224-015-9228-6.
- Blomberg, S. P., J. G. Lefevre, J. A. Wells, and M. Waterhouse. 2012. "Independent
 Contrasts and PGLS Regression Estimators Are Equivalent." *Systematic Biology* 61
 (January): 382–91. https://doi.org/10.1093/sysbio/syr118.

- Brodribb, T. J., N. M. Holbrook, M. A. Zwieniecki, and B. Palma. 2005. "Leaf Hydraulic
 Capacity in Ferns, Conifers and Angiosperms: Impacts on Photosynthetic Maxima." *New Phytologist* 165 (3): 839–46.
- Brooks, D.R., and D.A. McLennan. 1991. *Phylogeny, Ecology, and Behavior*. Chicago: U
 Chicago Press. https://press.uchicago.edu/ucp/books/book/chicago/P/bo3626827.html.
- 463 Carvalho, P., J. A. F. Diniz-Filho, and L. M. Bini. 2006. "Factors Influencing Changes in
 464 Trait Correlations across Species after Using Phylogenetic Independent Contrasts."
 465 *Evolutionary Ecology* 20 (6): 591–602. https://doi.org/10.1007/s10682-006-9119-7.
- 466 Cronin, J. P., and D. R. Schoolmaster Jr. 2018. "A Causal Partition of Trait Correlations:
 467 Using Graphical Models to Derive Statistical Models from Theoretical Language."
 468 *Ecosphere* 9 (9): e02422. https://doi.org/10.1002/ecs2.2422.
- 469 Díaz, S., J. Kattge, J. H. C. Cornelissen, I. J. Wright, Sandra Lavorel, S. Dray, B. Reu, et al.
 470 2016. "The Global Spectrum of Plant Form and Function." *Nature* 529 (7585): 167–
 471 71. https://doi.org/10.1038/nature16489.
- Falster, D. S., A. T. Moles, and M. Westoby. 2008. "A General Model for the Scaling of
 Offspring Size and Adult Size." *American Naturalist* 172 (3): 299–317.
 https://doi.org/10.1086/589889.
- Felsenstein, J. 1985. "Phylogenies and the Comparative Method." *American Naturalist* 125:
 1–15.
- 477 Garamszegi, L.Z., ed. 2014. Modern Phylogenetic Comparative Methods and Their
 478 Application in Evolutionary Biology: Concepts and Practice. Berlin: Springer-Verlag.
- 479 Gould, S. J., and E. S. Vrba. 1982. "Exaptation—a Missing Term in the Science of Form."
 480 *Paleobiology* 8 (1): 4–15. https://doi.org/10.1017/S0094837300004310.
- 481 Grafen, A. 1989. "The Phylogenetic Regression." *Phil. Trans. Roy. Soc. Lond. B* 326: 119–
 482 57.
- Halliwell, B, L. Yates, and B. Holland. 2022. "Multi-Response Phylogenetic Mixed Models:
 Concepts and Application." *BioRxiv*.
- 485 https://biorxiv.org/cgi/content/short/2022.12.13.520338v1.
- Hansen, T.F. 2014. "Use and Misuse of Comparative Methods in the Study of Adaptation."
 In *Modern Phylogenetic Comparative Methods and Their Application in Evolutionary Biology*, 351–79. Berlin: Springer-Verlag. DOI: 10.1007/978-3-662-43550-2 14.
- Harvey, P.H., and M.D. Pagel. 1991. *The Comparative Method in Evolutionary Biology*.
 Edited by R.M. May and P.H. Harvey. Oxford Series in Ecology and Evolution.
 Oxford: Oxford University Press.
- Hernán, M. A., J. Hsu, and B. Healy. 2019. "A Second Chance to Get Causal Inference
 Right: A Classification of Data Science Tasks." *CHANCE* 32 (1): 42–49.
 https://doi.org/10.1080/09332480.2019.1579578.
- Housworth, E. A., E. P. Martins, and M. Lynch. 2004. "The Phylogenetic Mixed Model." *The American Naturalist* 163 (1): 84–96. https://doi.org/10.1086/380570.
- Huey, R. B., T. Garland, and M. Turelli. 2019. "Revisiting a Key Innovation in Evolutionary
 Biology: Felsenstein's 'Phylogenies and the Comparative Method." *The American Naturalist* 193 (6): 755–72. https://doi.org/10.1086/703055.
- Losos, J. B. 2011. "Seeing the Forest for the Trees: The Limitations of Phylogenies in
 Comparative Biology." *The American Naturalist* 177 (6): 709–27.
 https://doi.org/10.1086/660020.
- Lynch, M. 1991. "Methods for the Analysis of Comparative Data in Evolutionary Biology."
 Evolution 45 (5): 1065–80. https://doi.org/10.2307/2409716.
- Marques, I., T. Kneib, and N. Klein. 2022. "Mitigating Spatial Confounding by Explicitly
 Correlating Gaussian Random Fields." *Environmetrics* 33 (5): e2727.
 https://doi.org/10.1002/env.2727.

- Martins, E. P., and T. F. Hansen. 1997. "Phylogenies and the Comparative Method: A
 General Approach to Incorporating Phylogenetic Information into the Analysis of
 Interspecific Data." *The American Naturalist* 149 (4): 646–67.
 https://doi.org/10.1086/286013.
- Pagel, M. 1999. "Inferring the Historical Patterns of Biological Evolution." *Nature* 401 (6756): 877–84. https://doi.org/10.1038/44766.
- Paradis, E. 2014. "An Introduction to the Phylogenetic Comparative Method." In L. Z. *Garamszegi (Ed.), Modern Phylogenetic Comparative Methods and Their Application in Evolutionary Biology*, 3–18. Berlin: Springer-Verlag.
- 517 Pearl, J. 2009. *Causality: Models, Reasoning and Inference*. 2nd ed. Cambridge University
 518 Press.
- 519 Price, T. 1997. "Correlated Evolution and Independent Contrasts." *Phil. Trans. R. Soc. Lond.*520 *B* 352: 519–29.
- Revell, L. J. 2010. "Phylogenetic Signal and Linear Regression on Species Data." *Methods in Ecology and Evolution* 1 (4): 319–29. https://doi.org/10.1111/j.2041-210X.2010.00044.x.
- Revell, L. J., and L. J. Harmon. 2022. *Phylogenetic Comparative Methods in R.* Princeton,
 N.J.: Princeton U.P.
- Swenson, N. G. 2020. *Phylogenetic Ecology: A History, Critique and Remodeling*. Chicago:
 University of Chicago Press.
- 528 Symonds, M.R.E., and S. P. Blomberg. 2014. "A Primer on Phylogenetic Generalised Least
 529 Squares." In *Modern Phylogenetic Comparative Methods and Their Application in*530 *Evolutionary Biology*, edited by L Z Garamszegi, 105–30. Springer Berlin
 531 Heidelberg. https://doi.org/10.1007/978-3-662-43550-2 5.
- 532 Uyeda, J. C, R. Zenil-Ferguson, and M. W. Pennell. 2018. "Rethinking Phylogenetic
 533 Comparative Methods." *Syst Biol* 67 (6): 1091–1109.
- Warton, D. I. 2022. *Eco-Stats: Data Analysis in Ecology*. Cham, Switzerland: Springer
 Nature Switzerland AG.
- Warton, D. I., I. J. Wright, D. S. Falster, and M. Westoby. 2006. "Bivariate Line-Fitting
 Methods for Allometry." *Biological Reviews* 81 (2): 259–91.
- Yates, L. A., S. A. Richards, and B. W. Brook. 2021. "Parsimonious Model Selection Using Information Theory: A Modified Selection Rule." *Ecology* 102 (10): e03475.
 https://doi.org/10.1002/ecy.3475.
- 541

542

544 Boxes and Tables

545 Box 1: Phylogenetic correction in brief

Consider a dataframe giving traits or habitat properties (columns) across a number of present-546 day species or other entities (rows). Also, the species in the data table are connected by a tree 547 structure representing their phylogeny, as best it is known. Phylogenetic correction of 548 549 correlations between columns in such a dataframe has two elements. There is a statistical 550 procedure, and then an interpretive step whereby the phylogenetically -adjusted relationship between two traits or between a trait and a habitat is seen as corrected, compared to the raw 551 552 correlations across present day species. The implication is that the phylogenetically adjusted 553 relationship is more reliable, or more enlightening, or that the model is more complete. 554 Statistical method and interpretation are linked. What generating process is being assumed by the statistical model, and hence what question exactly does a given statistical method ask? 555 556

557 One version of the statistical procedure is to transform a set of present-day species into a set 558 of evolutionary divergences or phylogenetically independent contrasts PICs (Felsenstein 559 1985). At each node in the tree, an evolutionary divergence or PIC is inferred for each trait. 560 These divergences, rather than present-day species, then become the objects under study, and 561 the cases or items of evidence in a statistical procedure. The question is whether divergences 562 in trait A tend to be correlated in size and direction with divergences in trait B. (For a 563 polytomy, there is a regression between trait A and trait B across the set of descendant species or nodes. Indeed for a dichotomy, the divergences can also be thought of as a two-564 point regression.) The effect has something in common with a pairing design in social 565 science, where individuals are matched for (say) gender or age or income, then differences 566 ("contrasts") are calculated across the pair for other variables, and the analysis proceeds 567 568 using those contrasts as items of evidence, rather than the individuals themselves. 569

Currently the method most often used is phylogenetic generalised least squares PGLS
(Grafen 1989; Martins and Hansen 1997). This is a regression model for relationships
between traits across species. The expected residual covariances between each pair of species
are modelled in such a way that higher covariance is expected when the species have
diverged more recently on the phylogenetic tree. If two species are outliers in the same
direction and also have a relatively recent common ancestor, then some covariance between

them is seen as expected, and the influence of those residuals on the position of the fitted line
is downweighted accordingly. In other words, the idea that traits are for unspecified reasons
slow to change through evolutionary time (phylogenetic inertia) is part of the causation being
modeled.

580

The phylogenetically expected covariances in PGLS scale with the combined branch lengths 581 582 shared between species, reflecting a Brownian-motion or diffusion model for trait change. 583 Often a parameter λ (Pagel 1999) is fitted by maximum likelihood as part of the model. This is a multiplication factor in the range 0 to 1 for the off-diagonal elements of the 584 585 phylogenetically expected residual covariance matrix. If λ is near zero, this effectively makes 586 the terminal branches of the tree very long, with little covariance expected even between sister species. (In phylogenetic mixed models PMM discussed in Box 3, an equivalent scaling 587 is estimated for each response trait (Halliwell et al. 2022)). In addition to the basic Brownian-588 589 motion model, a variety of more complex models have been developed (overview in 590 Garamszegi 2014), that fit parameters for rates of trait change that vary through time or in 591 response to other variables.

592

593 PGLS with λ fixed to 1 is mathematically equivalent to PICs, which iteratively calculate 594 divergences or contrasts at each node through the phylogenetic tree and treat those as a 595 population of events (Blomberg et al. 2012; Symonds and Blomberg 2014). Under these 596 circumstances the regression slope and confidence intervals reported by PGLS are 597 summarizing the population of regression slopes across all the divergences or nodes in the 598 phylogenetic tree. PGLS with $\lambda = 0$ yields the ordinary least squares regression slope across present-day species. With intermediate λ , the slope will lie somewhere in between those two 599 600 meanings. Mathematical treatment is provided as Supplementary 2.

- 601
- 602

Box 2. Selected quotes that illustrate uncertainty among expertsabout what is achieved when controlling for phylogeny

- 606 The majority or standard view is expressed by Garamszegi (2014) in the preface to an edited
- 607 book: "Statistically, the effect of phylogeny can be regarded as a confounding factor that
- 608 violates assumptions about non-independence of the unit of analysis, and that potentially
- 609 *introduces spurious correlations across traits.* "Similarly Huey et al. (2019): "Independent
- 610 contrasts enabled comparative biologists to avoid the statistical dilemma of nonindependence
- 611 of species values, arising from shared ancestry ... Felsenstein (1985) rapidly and radically
- 612 *changed both evolutionary and organismal biology ... No one would consider ignoring*
- 613 *phylogeny when analyzing data involving multiple species* ... "
- 614

605

615 As against that majority view, the following quotes make the point that adaptation to niche

- and phylogenetic history should not be treated as competing alternatives. Housworth et al
- 617 (2004) wrote "the heritable component contains not only genetic changes but also nongenetic
- 618 *contributions to the phenotype, such as environmental or cultural contributions, that are*
- 619 *described by the phylogenetic relationship among the taxa.*". Hansen (2014) wrote *"if related*
- 620 species tend to occur in similar environments (i.e., having similar values of their predictor
- 621 *variables), then we still expect a phylogenetic signal in the response variable. Correcting for*
- 622 *phylogeny in this situation is throwing the baby out with the bathwater ... [perhaps] the*
- 623 application of phylogenetic comparative methods has done more harm than good in the study
- 624 *of adaptation.*" De Bello et al (2015) wrote *"Phylogenetic relatedness between species*
- 625 should not be considered a bias to be corrected, but rather an evolutionary signal that allows
- 626 *results to be interpreted at different evolutionary scales.*"
- 627

Any given model reflects a hypothesis about processes generating the observed data (Uyeda,
Zenil-Ferguson, and Pennell 2018): *"[phylogenetic comparative models] PCMs are powerful tools for drawing inferences from interspecific data but they necessarily imply some types of causal structures and negate others. It is too much to ask of our methods to decide what questions we ought to ask."*

- 634 And causation can not be decisively inferred from survey data: *"the validity of causal"*
- 635 inferences depends on structural knowledge, which is usually incomplete, to supplement

- *the information in the data. As a consequence, no algorithm can quantify the accuracy of*
- *causal inferences from observational data*" (Hernán, Hsu, and Healy 2019).

Box 3: Multi-response phylogenetic mixed models MR-PMM asapplied to dissecting covariance between two traits across species

In multi-response phylogenetic mixed models, two or more traits appear as responses on the 642 643 left hand side of the model equation. Terms on the right hand side include a matrix of covariances expected from a model of trait change through the phylogeny, as well as trait-644 645 level intercepts and possibly fixed or random variables. With respect to a single-response model, changing the status of trait B from a predictor for A to a joint response variable with 646 647 phylogenetically structured residuals allows phylogenetically conservative A-B correlation (conservative trait correlation CTC) to be quantitatively identified. The multi-response 648 649 approach treats the A-B relationship as a question of trait coordination rather than a question of predicting A from B. For allometric relationships, this joint view yields a consistent 650 estimate of trait coordination, via a decomposition of their residual covariation, unlike the 651 652 predictive view where the slope estimates depend on whether A is predicted from B or vice 653 versa (Warton et al. 2006). Indeed, for data generated from a MR-PMM, -slope estimates for 654 B from a misspecified single-response model such as PGLS confound various components of the generating model (explained further in Supplementary 2). 655

656

641

MR-PMMs offer a sufficiently complex and more biologically appropriate model structure than their single-response analogues. These models simultaneously account for phylogenetic signal in all included traits and permit a decomposition of the estimated trait correlation according to dependence on phylogeny. For two species traits A and B, a multi-response mixed model with phylogenetic covariances modelled as a random effect takes the form 662

$$\binom{A}{B} = \binom{\mu_A + b_A + e_A}{\mu_B + b_B + e_B}$$

664

663

The μ 's are vectors of fixed effects, which can be any linear predictive equation. When the interest is only in the relationship between traits A and B, i.e. there are no predictors in the model, these fixed effects would contain only an_intercept for each trait.

The phylogenetic random effects b_A, b_B and the phylogeny-independent effects e_A, e_B are
drawn from multivariate normal distributions

$$(b_A, b_B) \sim MVN(0, \Sigma^{phy} \otimes C)$$

$$(e_A, e_B) \sim MVN(0, \Sigma^{ind} \otimes I)$$

674

For two response traits, A and B, and n species in the phylogeny, the covariance matrices for 675 the random effects and independent errors are of dimensions 2n x 2n. The covariance of the 676 phylogenetic random effects $\Sigma^{phy} \otimes C$ is the Kronecker product of a 2 x 2 trait-level 677 correlation matrix, Σ^{phy} , with C, the n x n matrix of expected error covariances given a 678 model of trait evolution applied to a phylogenetic tree. For the simplest case of Brownian 679 motion, C is the phylogenetic relatedness matrix. The covariance structure of the residuals or 680 phylogeny-independent elements $\Sigma^{ind} \otimes I$, is- the Kronecker product of a -2 x 2 trait-level 681 correlation matrix Σ^{ind} , with I, an n x n identity matrix (1 for diagonal elements and 0 for 682 off-diagonal elements). For the two-trait PMM, we estimate two phylogenetic variances for A 683 and B (Σ_{AA}^{phy} and Σ_{BB}^{phy}) and the phylogenetic covariance between A and B (Σ_{AB}^{phy}). The same 684 is true for independent (co)variances in the elements of Σ^{ind} . When scaled by the relevant 685 variance components, these covariances yield phylogenetic and residual correlations. Thus, 686 687 when appropriately parameterized, the MR-PMM estimates each element listed in Table 1. 688

MR-PMM looks at the correlation between A and B rather than at predicting one from the
other. Given a fitted MR-PMM, further derived quantities such as (standardized) major axes
relating A to B (Warton et al. 2006) can be constructed from either point estimates or
posterior distributions of the variance and covariance parameters associated with the two
traits.

694

In principle, models with this layout can have any number of species traits or habitat
properties on the left hand side, and also other predictors included in the fixed-effect terms on
the right hand side. More complex models require more replication to yield reliable estimates
(Housworth, Martins, and Lynch 2004). For simplicity, we have confined this explanation to
the correlation between two Gaussian traits, but response variables are not required to be
Gaussian distributed. See Halliwell, Yates and Holland (2022) for details including worked
examples in two popular R packages, 'MCMCglmm' and 'brms'.

704 Box 4: Past and present-day causation

The question whether adaptation should be interpreted as a past versus a present-day process has long been debated. Palaeobiologists and evolutionists have insisted that adaptation should refer only to the selective circumstance that initially gave rise to a trait. For example, Gould and Vrba (1982) coined "exaptation" for functionality that came about subsequent to a trait's origin, in order to reserve adaptation for functionality at the time of origin. (For a quantitative trait such as adult body size, they must have meant "origin" to refer to the time the trait arrived at a particular value.)

712

713 This defining of terms by evolutionists has mostly stuck over the ensuing 40 years. For 714 example, Paradis (2014) wrote: "we can define the phylogenetic comparative method as the 715 analytical study of species, populations, and individuals in a historical framework with the 716 aim to elucidate the mechanisms at the origin of the diversity of life." Losos's (2011) 717 presidential address to American Society of Naturalists discussed traits and phylogenies. 718 Summarizing the history of ideas, he wrote "the key turning point was the publication of 719 Felsenstein's (1985) article in the American Naturalist, which presented the issue of shared 720 ancestry as a difficulty in comparative analysis and the independent contrasts method as the 721 solution publication of books by Brooks and McLennan (1991) and Harvey and Pagel (1991) completed the revolution. Since that time, there has been a continuous, unabated rise 722 in the development and use of phylogenetic comparative methods. Comparative studies now 723 724 are essentially unpublishable unless analyzed in a phylogenetic context ...". Losos 2011 also 725 wrote in a footnote: "many reviewers ... have been concerned that this article will give 726 license to ecologists and other ne'er-do-wells to ignore phylogenetic approaches entirely. So, 727 just to be clear, I will say it again: phylogenetics is an important approach for studying historical events This article should not be read as license to ignore phylogenetic 728 729 information in comparative studies!"

730

Phylogenies are indeed essential for studying the history of divergences. But what has
happened here is that comparative studies have been defined as being about history, in the
same way as adaptation earlier was defined as being about history. On the other hand,
ecologists have a continuing interest in the question what traits or trait-combinations make
species successful in what situations in the present day. Losos intended, no doubt, to express
collegiality from evolutionists toward ecologists when he breezily called them ne'er-do-

wells. But the collegiality did not extend to permitting ecologists to consider adaptation andcomparative studies as questions about the present day.

739

740 The essential point for ecologists is that patterns such as in Fig 1 where a trait is correlated

both with another trait and with phylogenetic history, called here conservative trait

correlation CTC, can potentially arise from a deep historical divergence followed by limited

subsequent change, or from continuing selection in the present day. The observed pattern

does not give a basis for preferring one explanation to the other, and moreover the two need

not be mutually exclusive. For ecologists aiming to describe trait combinations that confer

746 present-day competence, it is not sensible to remove the conservative trait correlation from

consideration. That is why we recommend instead the partitioning of variation provided by

748 MR-PMM (Table 1).

750 Table 1. Where variation is attributed by the multi-response phylogenetic mixed

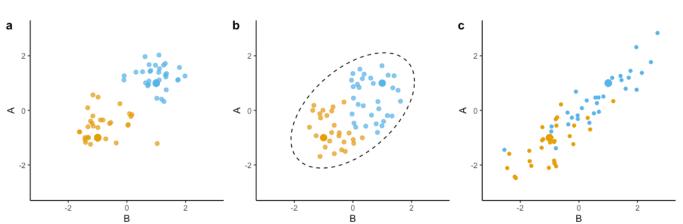
751 model (MR-PMM) described here. Key parameters estimated are four standard

752 deviations σ_A^{phy} , σ_B^{phy} , σ_A^{ind} , and σ_B^{ind} , and two correlations ρ_{AB}^{phy} and ρ_A^{ind}	752	ns ρ_{AB}^{pny} and ρ_{AB}^{ing}	correlations	and two	and $\sigma_{\rm P}^{ind}$.	σ^{ind} .	$\sigma_{\rm p}^{phy}$.	deviations σ_{\star}^{pny}	752
--	-----	--	--------------	---------	------------------------------	------------------	--------------------------	-----------------------------------	-----

deviations σ_A^{phy} , σ_B^{phy} , σ_A^{thu} , and σ_B^{thu} , and two correl	ations $ ho_{AB}^{\mu\nu}$ and $ ho_{AB}^{\mu\mu}$.
Components of variation	expression
total variation in A	$\sigma_A^2 = \left(\sigma_A^{phy} ight)^2 + \left(\sigma_A^{ind} ight)^2$
phylogenetic signal in A	$\lambda_A = rac{\left(\sigma_A^{phy} ight)^2}{\sigma_A^2}$
total variation in B	$\sigma_B^2 = \left(\sigma_B^{phy} ight)^2 + \left(\sigma_A^{ind} ight)^2$
phylogenetic signal in B	$\lambda_B = rac{\left(\sigma_B^{phy} ight)^2}{\sigma_B^2}$
A-B correlation associated with phylogeny (called here conservative trait correlation CTC)	$ ho_{AB}^{phy} = rac{\Sigma_{AB}^{phy}}{\sqrt{\Sigma_{AA}^{phy}\Sigma_{BB}^{phy}}}$
A-B correlation independent from phylogeny	$ ho_{AB}^{ind} = rac{\Sigma_{AB}^{ind}}{\sqrt{\Sigma_{AA}^{ind}\Sigma_{BB}^{ind}}}$
total A-B correlation	$ \rho_{AB} = $
(expression in terms of lambda as given by Housworth, Martins, and Lynch 2004)	$rac{\Sigma^{phy}_{AB}+\Sigma^{ind}_{AB}}{\sigma_A\sigma_B}= ho^{phy}_{AB}\sqrt{\lambda_A\lambda_B}+$
	$ ho_{AB}^{ind}\sqrt{(1-\lambda_A)(1-\lambda_B)}$

753

bioRxiv preprint doi: https://doi.org/10.1101/2023.02.05.527214; this version posted February 5, 2023. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.



755

Figure 1. Data simulated under different evolutionary models, beginning from two clades 756 757 (orange and blue) separated in a space described by two traits A and B. Large solid points represent the most recent common ancestor for the orange and blue clades in each simulation. 758 759 Species then radiate, and traits diversify, within each clade. In simulation (a), radiation of 760 each major clade proceeds by Brownian motion. The overall correlation between traits has been produced entirely by the starting points of the two major clades. In simulation (b), the 761 radiations are positioned at random within a region of trait space (broken line) whereby only 762 trait combinations within the line are competent to support viable populations. New viable 763 species are more likely to arise from clades that have existing species nearer to them in trait 764 space. The observations in present-day species are not distinguishable between simulations 765 (a) and (b), illustrating how historical vs present-day determination of an overall correlation 766 can often not be distinguished by analysis of present-day data. A version of this comparison 767 was first given by Price (1997). In simulation (c), data are produced from a MR-PMM. 768 Positive correlations between A and B are operating on both the phylogenetic and 769 independent level ($\rho_{AB}^{phy} = \rho_{AB}^{ind} = 0.7$), resulting in a tight overall relationship between A 770 771 and B. Importantly, correlations within each of the major clades and also between them, are 772 both important drivers of the present-day spectrum of variation.

bioRxiv preprint doi: https://doi.org/10.1101/2023.02.05.527214; this version posted February 5, 2023. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. All rights reserved. No reuse allowed without permission.

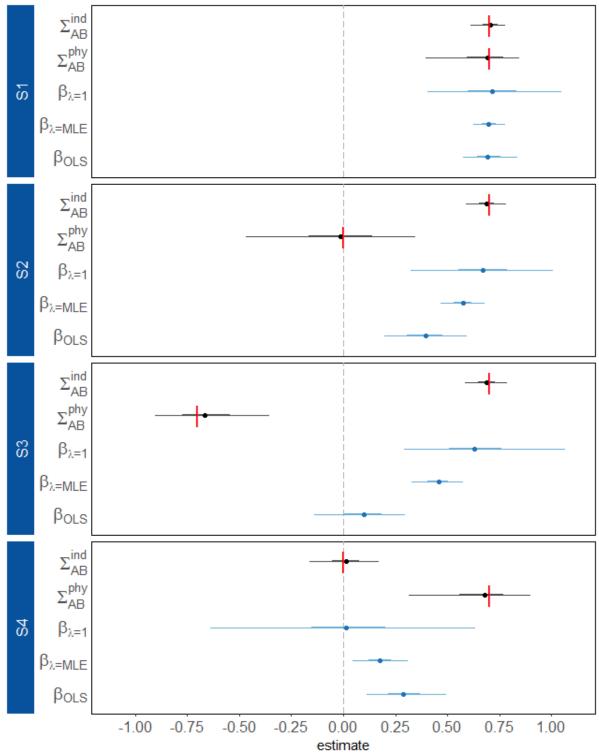




Figure 2. Parameter estimates for phylogenetic and independent covariances (black) from MR-PMM, and beta coefficients (blue) from PGLS ($\beta_{\lambda=1}$), PGLS with lambda optimised ($\beta_{\lambda=MLE}$), and OLS (β_{OLS}) fit to simulated datasets (S1-4). Points represent the median posterior estimate across 400 model fits, with heavy and light wicks showing the 50% and 90% sample quantiles, respectively. For each simulation, $\Sigma_{AA}^{ind} = \Sigma_{BB}^{ind} = \Sigma_{AA}^{phy} = \Sigma_{BB}^{phy} = 1$. True values for Σ_{AB}^{ind} and Σ_{AB}^{phy} used to generate the data for each simulation are indicated by

