PI3K block restores age-dependent neurovascular coupling defects associated with cerebral small vessel disease

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Abstract

Neurovascular coupling (NVC), a vital physiological process that rapidly and precisely directs localized blood flow to the most active regions of the brain, is accomplished in part by the vast network of cerebral capillaries acting as a sensory web capable of detecting increases in neuronal activity and orchestrating the dilation of upstream parenchymal arterioles. Here, we report a Col4a1 mutant mouse model of cerebral small vessel disease (cSVD) with age-dependent defects in capillary-to-arteriole dilation, functional hyperemia in the brain, and memory. The fundamental defect in aged mutant animals was the depletion of the minor membrane phospholipid phosphatidylinositol 4,5 bisphosphate (PIP2) in brain capillary endothelial cells, leading to the loss of inwardly rectifier K+ (Kir2.1) channel activity. Blocking phosphatidylinositol-3-kinase (PI3K), an enzyme that diminishes the bioavailability of PIP2 by converting it to phosphatidylinositol (3,4,5)-trisphosphate (PIP3), restored Kir2.1 channel activity, capillary-to-arteriole dilation, and functional hyperemia. In longitudinal studies, chronic PI3K inhibition also improved the memory function of aged Col4a1 mutant mice. Our data suggest that PI3K inhibition is a viable therapeutic strategy for treating defective NVC and cognitive impairment associated with cSVD.

One-sentence summary

PI3K inhibition rescues neurovascular coupling defects in cerebral small vessel disease.

Key words

COL4A1, extracellular matrix, cerebral small vessel disease, functional hyperemia, spatial working and recognition memory, PIP2, Kir2.1 channels, PI3K.
Introduction

A group of familial and idiopathic pathologies known as cerebral small vessel diseases (cSVDs) are a leading cause of vascular contributions to cognitive impairment and dementia (VCID) \(^1\), second only to Alzheimer’s disease as the most common cause of cognitive impairment in adults \(^2,3\). The global impact of cSVDs is massive and rapidly growing as the world’s population ages \(^4\), but little is currently known about the pathogenesis of the disease and no specific treatments exist. Brain atrophy, lacunes, enlarged perivascular spaces, intracerebral hemorrhages (ICH), white matter hyperintensities, and microinfarcts detected by magnetic resonance imaging (MRI) are clinical signs of irreversible brain damage brought on by cSVDs \(^5\). cSVDs also impair functional hyperemia, a crucial physiological process in which local blood flow is swiftly and precisely diverted to the most active brain regions \(^6-9\). Dysregulation of cerebral blood flow contributes to vascular dementia \(^1\), but it is not known if resolution of this impairment can delay or reverse cognitive decline. Autosomal dominant mutations in the genes encoding collagen type IV alpha 1 (COL4A1) and alpha 2 (COL4A2) cause an inherited form of cSVD as part of a multisystem disorder called Gould syndrome \(^10-13\). In this study, we used Col4a1 mutant mice to elucidate the molecular events that disrupt functional hyperemia in this type of cSVD and applied these findings to repair the deficit.

Functional hyperemia is accomplished by a collection of physiological processes called neurovascular coupling (NVC). During NVC, active neurons trigger the dilation of upstream pial arteries and parenchymal arterioles supplying the brain, boosting blood flow to fulfill regional metabolic demands \(^14\). The underlying mechanisms have been intensely studied for decades \(^15-17\), however much remains to be discovered. A new
paradigm has emerged envisioning the vast cerebral capillary network as a sensory
web capable of detecting increases in neuronal activity and orchestrating the dilation of
upstream parenchymal arterioles. In this arrangement, substances released from
nearby active neurons and/or astrocytic endfeet stimulate receptors on brain capillary
endothelial cells (ECs) to generate retrograde propagating vasodilator signals that travel
against the flow of blood and dilate upstream arterioles. Two types of ion channels
present on brain capillary ECs, inward-rectifying K+ (Kir2.1) channels, and transient
receptor potential ankyrin 1 (TRPA1) cation channels, are critical sensors of neuronal
activity and are necessary for NVC and functional hyperemia in the brain. Kir2.1
channels are activated by K+ ions released during neuronal activity and initiate rapid
retrograde propagating electrical signals that ultimately dilate upstream arterioles and
increase blood flow to meet neuronal demand. TRPA1 channels are activated by
reactive oxygen species metabolites and stimulate propagating intercellular Ca2+ waves
that dilate upstream arterioles. Capillary-to-arteriole dilation is impaired in the
CADASIL (Cerebral Autosomal Dominant Arteriopathy with Sub-cortical Infarcts and
Leukoencephalopathy) cSVD mouse model, but the impact of Col4a1 mutations on
this process, and NVC coupling, is unknown.

Here, we report that Kir2.1 channel activity in ECs of cerebral arteries and brain
capillaries from Col4a1 mutant mice is lost during aging. K+-induced capillary-to-
arteriole dilation was absent in vascular preparations from 12-month (M)-old mutant
animals, resulting in impaired functional hyperemia in the somatosensory cortex and
memory deficits. Reduced Kir2.1 channel activity was due to diminished
phosphatidylinositol 4,5 bisphosphate (PIP2), a minor membrane phospholipid that is an
essential co-factor for Kir2.1 activity. Kir2.1 channel activity and capillary-to-arteriole dilation was restored by blocking phosphoinositol 3-kinase (PI3K), an enzyme that converts PIP$_2$ to phosphatidylinositol (3,4,5)-trisphosphate (PIP$_3$). Chronic treatment of mutant mice with a PI3K antagonist restored Kir2.1 channel activity and capillary-to-arteriole dilation, improved functional hyperemia, and largely resolved memory deficits. Our findings reveal a novel age-dependent mechanism of impaired NVC associated with cSVD and identify PI3K as a potential therapeutic target for treating vascular cognitive impairment and dementia.

Results

Age-dependent loss of Kir2.1 channel activity and capillary-to-arteriole dilation in Col4a1$^{+/G394V}$ mice.

Mutant mice used for this study harbor a point mutation in one copy of the Col4a1 gene that results in the glycine (G) to valine (V) substitution at position 394 of the COL4A1 polypeptide (Col4a1$^{+/G394V}$). Wild type littermates were used as controls for all experiments. Both male and female mice were studied, and no sex-specific differences were observed. Age is the most critical risk factor for cSVD and vascular dementia. Therefore, we used Col4a1$^{+/G394V}$ and control mice at 3 and 12 months (M) of age, representing young adulthood and middle age, respectively, throughout this study.

Using whole-cell patch clamp electrophysiology, we first investigated how Kir2.1 currents in freshly isolated brain capillary ECs (Fig. 1A) were affected by Col4a1$^{G394V}$ mutation. Kir2.1 channels were activated by increasing the [K$^+$] of the bath solution to 60 mM, and currents were recorded as voltage ramps (-100 to +40 mV) were applied. The
selective Kir channel blocker BaCl$_2$ (10 µM) was used to isolate the current. Kir2.1 current densities in brain capillary ECs from young adult mice did not differ between mutant and control mice (Fig. 1B) but were significantly reduced in brain capillary ECs from 12 M-old Col4a1$^{+/G394V}$ mice compared with aged-matched control animals (Fig. 1C). We then utilized an innovative ex vivo cerebral microvascular preparation in which parenchymal arteriole segments with intact capillary branches were isolated from the brain, cannulated and pressurized$^{18,19}$ to determine if loss of Kir2.1 channel activity translated into impaired capillary-to-arteriole dilation. Capillaries were stimulated by locally applying pulses (7 seconds; 10 psi) of KCl (10 mM) dissolved in artificial cerebral spinal fluid (aCSF) using a micropipette attached to a picospritzer (Fig. 1D). Focal application of K$^+$ to the capillary beds of 3 M-old Col4a1$^{+/+}$ and Col4a1$^{+/-/G394V}$ mice produced robust, reversible, and reproducible dilations of the upstream arterioles (Fig. 1E and F). The addition of the nitric oxide donor sodium nitroprusside (SNP) to the tissue bath maximally dilated arterioles from both groups (Fig. 1G). Similar responses were observed when tissues from 12 M-old control mice were used (Fig. 1H and I). In contrast, focal application of K$^+$ onto capillaries from 12 M-old Col4a1$^{+/-/G394V}$ mice did not dilate upstream arterioles. However, SNP produced maximal dilation, demonstrating the viability of the preparation (Fig. 1J). These results show that the age-dependent impairment of Kir2.1 current activity in brain capillary ECs from Col4a1$^{+/-/G394V}$ mice eliminates capillary-to-arteriole dilation.

We also investigated the effects of the Col4a1 mutation on Kir2.1 channel activity in the ECs that line cerebral arteries (arteriolar ECs) (Fig. 1- supplement 1A) and found that these currents did not differ for 3 M-old mice but were blunted in 12 M-old
Col4a1+/G394V animals compared to age-matched controls (Fig. 1- supplement 1B and C). Prior studies show that cerebral arteries dilate when [K+] is raised from 3 mM to between 8 and 20 mM because increased Kir2.1 channel activity hyperpolarizes the membrane potential of vascular smooth muscle. Higher [K+] (greater than 30 mM) collapses the gradient, causing membrane depolarization and vasoconstriction. To determine the functional consequence of reduced Kir2.1 channel activity in arteriolar ECs from Col4a1 mutants, we used standard pressure myography techniques to investigate the effects of increasing [K+] on vasomotor responses of isolated cerebral arteries. We found that raising the external [K+] from 3 mM to a range of 8 to 20 mM dilated cerebral arteries from 12-M old control animals but had no effect on arteries from 12 M-old Col4a1+/G394V mice (Fig. 1 – supplement 1D and E). In contrast, vasoconstriction in response to higher [K+] (30 and 60 mM) did not differ between 12 M-old Col4a1+/G394V and control mice (Fig. 1 – supplement 1F). These results demonstrate that age-dependent loss of Kir2.1 currents impairs K+-induced vasodilation. In addition, our data show that fundamental voltage-dependent contractile mechanisms are not grossly affected.

Prior studies have demonstrated mild ICHs in the brains of 12 M-old Col4a1+/G394V mice. We further demonstrated this using in vivo susceptibility-weighted magnetic resonance imaging (SWI) to identify possible brain lesions. No hypointense lesions were detected on any SWI images from 12 M-old Col4a1+/+ and Col4a1+/G394V mice (Fig. 1 – supplement 2A), demonstrating that this mutation does not induce magnetic resonance (MR)-detectable hemorrhagic lesions. Volumetric analysis of the T2-weighted (T2W) images showed that the ventricle/brain ratio was not
significantly different between 12 M-old $\text{Col4a1}^{+/+}$ and $\text{Col4a1}^{+/G394V}$ mice (Fig. 1 – supplement 2B), further demonstrating that these mutant mice do not show obvious pathology in these imaging modalities.

**Age-dependent impairment of functional hyperemia in $\text{Col4a1}^{+/G394V}$ mice.**

The effects of the $\text{Col4a1}^{G394V}$ mutation on brain hemodynamics *in vivo* were investigated using the thinned-skull laser Doppler flowmetry method to measure blood flow changes in the somatosensory cortex in response to stimulating whiskers for 1, 2, or 5 seconds (s) (Fig. 2A and B) \(^{19,33,34}\). Stimulating contralateral whiskers for 1 s reproducibly increased blood flow in the somatosensory cortex of 3 M-old mice, with no differences in the amplitude, latency, or kinetics of the response between control and mutant mice (Fig. 2C to H). The magnitude of blood flow increases in response to 1 s stimulation were significantly blunted in 12 M-old $\text{Col4a1}^{+/G394V}$ mice compared to age-matched controls (Fig. 2I and J). In addition, the latency of the blood flow response was increased, and the rise and decay rates were diminished in 12 M-old mutant mice (Fig. 2K to N). Similar outcomes were observed for the 2 s (Fig. 2 – supplement 1), and 5 s (Fig. 2O to Z) stimulation protocols – no differences were detected in 3 M-old mice, but a blunted increases in blood flow, increased latency, and decreased rise rate and decay rate was detected for 12 M-old $\text{Col4a1}^{+/G394V}$ mice compared with controls. In addition, the duration of the blood flow increase was reduced in 12-M old mutant mice compared with controls in the 5 s stimulation protocol. Stimulation of ipsilateral whiskers failed to produce any change in the blood flow (Fig. 2 – supplement 2). These results demonstrate that loss of brain EC Kir2.1 channel activity and capillary-to-arteriole
dilation is associated with impaired functional hyperemia in 12 M-old Col4a1+/G394V mice in vivo.

**Age-dependent memory deficits in Col4a1+/G394V mice.**

Impaired NVC is associated with deficiencies in spatial working and recognition memory. Here, the spontaneous alternation behavioral assay was used to determine if Col4a1+/G394V mice develop memory deficits. Mice were freely allowed to explore all three arms of a Y-shaped maze for 10 minutes, and their consecutive entries into each of the arms were recorded and reported as % alternation (Fig. 3A). Spontaneous alternation, the maximum number of alternations, and the total distance moved did not differ between 3 M-old Col4a1+/G394V and control mice (Fig. 3B to D). However, spontaneous alternation was significantly diminished for 12 M-old Col4a1+/G394V mice compared with age-matched controls (Fig. 3E), suggesting deficits in spatial working memory. The maximum alternation and total distance moved did not differ between 12 M-old mutant and control mice, indicating that impaired mobility does not account for reduced alternation (Fig. 3F and G).

A second behavioral assay, the novel arm test, was also used to evaluate recognition memory function in mutant mice. This task is driven by the innate curiosity of mice to explore previously unvisited areas. Initially, mice were allowed to explore a Y-maze for 10 minutes with one arm of the maze blocked. After 24 hours, the mice were allowed to explore the maze for 10 minutes with all arms open (Fig. 3H). Mice with normal recognition memory spend more time exploring the novel arm than those explored on the previous day. The dwell time and the number of entries in the novel arm did not differ between 3 M-old Col4a1+/G394V and control mice (Fig. 3I to K). In contrast,
12 M-old Col4a1+/G394V mice spent significantly less time and had fewer entries into the novel arm compared with age-matched controls (Fig. 3L to N). These data provide further evidence of age-dependent memory deficits in Col4a1+/G394V mice.

**PIP2 depletion reduces Kir2.1 currents in 12 M-old Col4a1+/G394V mice.**

Kir2.1 channels require PIP2 for activity, and prior studies reported that pathogenic loss of PIP2 diminished Kir2.1 channel activity in brain capillary ECs from CADASIL cSVD mice and 5xFAD familial Alzheimer’s disease mice. Therefore, we investigated the possibility that PIP2 depletion also reduces Kir2.1 current density in ECs from 12 M-old Col4a1+/G394V mice. When exogenous diC8-PIP2 (10 µM) was included in the intracellular solution during whole-cell patch-clamp experiments, Kir2.1 current density in arteriolar ECs (Fig. 4A) and brain capillary EC (Fig. 4B) did not differ between 12 M-old control and age-matched Col4a1+/G394V mice, suggesting that PIP2 depletion is responsible for Kir2.1 current deficiencies in the mutant mice. The steady-state amount of PIP2 present in the inner leaflet of the plasma membrane is determined by the relative rates of PIP2 synthesis and degradation. PIP2 is synthesized by the sequential action of phosophatidylinositol 4-kinase (PI4K), an enzyme that converts phosphatidylinositol (PI) to phosphatidylinositol 4-phosphate (PIP), and phosphatidylinositol 4-phosphate 5-kinase (PIP5K), which converts PIP to PIP2. PI4K and PIP5K activity require high levels of ATP (i.e., the K_M of PI4K for ATP is ~0.4 to 1 mM) (Fig. 4 – supplement 1A). PIP2 deficiency in capillary ECs from CADASIL mice was attributed to reduced ATP levels and diminished synthesis. We measured ATP in isolated brain capillaries using a luciferase-based assay and found that ATP levels did not differ between 12 M-old Col4a1+/G394V and control mice (Fig. 4 – supplement 1B),
suggesting that PIP$_2$ synthesis is not impaired by lack of ATP in *Col4a1* mutants. Hydrolysis of PIP$_2$ by phospholipase C (PLC) to form inositol trisphosphate (IP$_3$) and diacylglycerol (DAG) diminishes steady-state PIP$_2$ levels (Fig. 4 – supplement 1C) $^{45}$. However, blocking PLC activity with U73122 (10 µM) did not restore Kir2.1 currents in capillary ECs from 12 M-old *Col4a1*+/*G394V* mice, suggesting that PIP$_2$ depletion does not result from elevated levels of PLC activity (Fig. 4 – supplement 1D). PIP$_2$ bioavailability is reduced when the enzyme PI3K phosphorylates it to PIP$_3$ (Fig. 4C). When PI3K was blocked using the selective inhibitor GSK1059615 (10 nM), Kir2.1 currents in capillary ECs from 12 M-old *Col4a1*+/*G394V* mice were restored to control levels (Fig. 4D), suggesting that PIP$_2$ depletion in mutant mice results from elevated PI3K activity. To determine if elevated PI3K activity is also responsible for cerebral microvascular dysfunction in mutant mice, capillary-to-arteriole dilation was assessed before and after treatment with GSK1059615 (10 nM, 30 min). This treatment had no effect on preparations from 12 M-old control animals (Fig. 4E and F). In contrast, PI3K block fully restored K$^+$-induced dilation of upstream arterioles from 12 M-old *Col4a1*+/*G394V* mice (Fig. 4G and H). In control studies, we found that the vehicle for GSK1059615 (0.01% v/v DMSO) did not affect capillary-to-arteriole dilation (Fig. 4 – supplement 2A to D) or maximal vasodilation in response to SNP (Fig. 4 – supplement 2E and F). These data suggest that elevated PI3K activity is responsible for the loss of capillary-mediated dilation in *Col4a1* mutant mice.
Chronic PI3K inhibition restores Kir2.1 currents, K⁺-induced dilation, functional hyperemia, and memory deficits in 12 M-old Col4a1+/G394V mice.

To determine if chronic inhibition of PI3K could resolve deficits in functional hyperemia and memory in mutant mice, 12 M-old Col4a1+/G394V and control mice were injected subcutaneously with GSK1059615 (10 mg/kg) or vehicle each day for 28 days (Fig. 5A). At the end of the treatment, we found that Kir2.1 currents in brain capillary ECs from Col4a1+/G394V mice injected with GSK1059615 were fully restored to control levels, whereas vehicle treatment had no effect (Fig. 5B). GSK1059615 treatment fully restored capillary-to-arteriole dilation in ex vivo microvascular preparations from Col4a1+/G394V mice (Fig. 5C and D). These data demonstrate that defects in brain capillary EC Kir2.1 channel function and associated microvascular vasomotor dysfunction can be restored by chronic PI3K blockade.

GSK1059615 treatment also increased the magnitude of blood flow increases in the somatosensory cortex induced whisker stimulation (1 s) in Col4a1+/G394V mice compared to vehicle-treated animals (Fig. 5E and F). The latency and rise rates of the blood flow response were also improved (Fig. 5G to J). Similar outcomes were observed for 2 s and 5 s stimulation protocols (Fig. 5 – supplement 1). In control studies, stimulation of ipsilateral whiskers failed to produce any change in the blood flow (Fig. 5 – supplement 2). These data demonstrate that chronic PI3K inhibition can restore functional hyperemia in Col4a1+/G394V mice.

In a longitudinal study, we found that GSK1059615 treatment significantly improved the Y-maze spontaneous alternation behavior of 12 M-old Col4a1+/G394V, whereas vehicle-treated mutant mice did not improve. The performance of
GSK1059615-treated Col4a1+/G394V mice did not differ from that of age-matched control animals (Fig. 5K). Interestingly, GSK1059615 treatment reduced the maximum number of alternations (Fig. 5L) and the total distance moved (Fig. 5M) for control and mutant mice. This unexpected side effect of prolonged GSK1059615 administration may be related to fatigue symptoms reported by cancer patients treated with PI3K inhibitors 46. Despite this potential limitation, our data identify PI3K inhibition as a novel therapeutic strategy for treating cognitive impairment associated with some forms of cSVD.

Discussion

Brain injuries and loss of fundamental blood flow control mechanisms due to cSVDs are significant causes of adult dementia. The brain pathology of Col4a1+/G394V cSVD mice used in this investigation was mild compared to other models, but NVC and functional hyperemia were significantly impaired. Thus, these animals allow the effects of impaired vascular control to be investigated independently of severe brain damage. Aged mutants performed worse than control mice in behavioral tests of working and recognition memory, suggesting that defects in cerebral blood flow control mechanisms are a primary cause of cognitive impairment in these animals. The fundamental defect leading to impaired NVC was the loss of Kir2.1 channel activity in brain capillary and arterial ECs due to PIP2 depletion. Accordingly, chronic PI3K blockade rescued Kir2.1 currents, NVC, functional hyperemia, and improved memory function in mutant mice, providing evidence that cognitive impairment associated with cSVD can be resolved by improving blood flow regulation in the brain.

Missense mutations in COL4A1 and COL4A2 that alter G residues in collagen triple helices are the most common cause of Gould syndrome 47-49. Such mutations
prevent the proper assembly of collagen α1α1α2(IV) heterotrimers, potentially leading to intracellular retention, ER stress, and/or disrupting basement membranes to cause cSVD and other pathologies. The impact of specific disease-causing point mutations is highly variable and position dependent. Notably, spontaneous ICH is less severe for mutations nearer to the amino terminus, such as the Col4a1G394V mutation, and more severe for mutations closer to the carboxyl terminus. Consistent with this finding, Col4a1+/G394V mice did not show overt cerebral pathology in MRI scans. However, capillary-mediated NVC and functional hyperemia were impaired in an age-dependent manner. The age dependence of this pathology may be related to the unique properties of collagen. Collagens are extraordinarily durable - the in vivo half-life of collagen I, present in ligaments and tendons, is more than 100 years. The turnover rate of collagen α1α1α2(IV) is not precisely known, but an early study reported that the half-life of “vascular collagen” in rat aorta and mesenteric arteries was ~70 days. Further, collagen synthesis peaks during late embryonic development and growth as basement membranes form and then markedly decreases with age. Our data suggest that Col4a1+/G394V mice successfully develop and maintain functional basement membranes early in life when collagen production is maximal. We propose that diminishing collagen production and increasing degradation during aging result in a slow diminution of basement membranes over time, eventually leading to defects by middle age. It is conceivable that a similar process could contribute to idiopathic forms of age-related cSVD in humans.

Elevated TGF-β signaling has been recently described in Col4a1 mutant mice. Genetic knockdown of TGF-β ligand and blocking TGF-β signaling using neutralizing
antibodies or pharmacological inhibitors improved ocular and cerebrovascular pathogenesis associated with Col4a1 mutations\textsuperscript{32,56,57}. Activation of TGF-β receptors can stimulate PI3K activity through a non-canonical signaling pathway involving the TRAF6 ubiquitin ligase \textsuperscript{58,59}, suggesting that enhanced TGF-β signaling contributes to PIP\textsubscript{2} insufficiency in Col4a1\textsuperscript{+/G394V} mice. Further, disinhibition of TGF-β signaling causes CARASIL (cerebral autosomal recessive arteriopathy with subcortical infarcts and leukoencephalopathy), a rare genetic form of cSVD \textsuperscript{60}, suggesting that disturbances in the TGF-β pathway may be a generalized mechanism of cerebral vascular dysfunction.

Altered PIP\textsubscript{2} metabolism has emerged as a central pathogenic mechanism in multiple forms of cerebral microvascular disease. Prior reports show that PIP\textsubscript{2} insufficiency diminishes Kir2.1 channel activity in brain capillary ECs in mouse models of CADASIL cSVD \textsuperscript{22} and familial Alzheimer’s disease \textsuperscript{41}. The current data show that the same defect underlies impaired functional hyperemia and memory deficits in Col4a1\textsuperscript{+/G394V} mutant mice. Prior studies provide evidence that reduced levels of ATP in brain capillary ECs diminished the production of PIP\textsubscript{2} \textsuperscript{22}, whereas our findings indicate that increased activity of PI3K decreased PIP\textsubscript{2} bioavailability in Col4a1\textsuperscript{+/G394V} mice. We also show that PIP\textsubscript{2} levels and Kir2.1 channel activity are diminished in cerebral arteriolar ECs from 12 M-old Col4a1 mutant mice. In contrast, these cells are unaffected in CADASIL and 5xFAD animals, demonstrating cellular heterogeneity of PIP\textsubscript{2} depletion mechanisms among the different disease models. Despite these differences, we propose that repairing defective NVC by increasing PIP\textsubscript{2} levels in brain capillary ECs is a viable therapeutic strategy for multiple forms of cerebrovascular disease. We provide proof-of-concept by targeting the PI3K pathway to preserve PIP\textsubscript{2}, thereby restoring...
functional hyperemia and improving memory function in Col4a1 mutant mice. Several types of PI3K inhibitors are approved by the FDA for use against advanced cancers and could rapidly advance to clinical trials for the treatment of cSVDs.
Materials and methods

Chemical and reagents

Chemicals and other reagents were obtained from Sigma-Aldrich, Inc. (St. Louis, MO, USA) unless otherwise specified.

Animals

Young adult (3 M-old) and middle-aged (12 M-old) male and female littermate Col4a1+/+ and Col4a1+/G394V mice were used in this study. Animals were maintained in individually ventilated cages (<5 mice/cage) with ad libitum access to food and water in a room with controlled 12-hour light and dark cycles. All animal care procedures and experimental protocols involving animals complied with the NIH Guide for the Care and Use of Laboratory Animals and were approved by the Institutional Animal Care and Use Committees at the University of Nevada, Reno, and the University of California, San Francisco. Arteries for ex vivo experimentation were harvested from mice anesthetized with isoflurane (Baxter Healthcare, Deerfield, IL, USA) and euthanized by decapitation and exsanguination. Brains were isolated and placed in ice-cold Ca2+-free physiological saline solution (Mg-PSS; 134 mM NaCl, 5 mM KCl, 2 mM MgCl2, 10 mM HEPES, 10 mM glucose, 0.5% bovine serum albumin, pH 7.4 with NaOH).

In vivo magnetic resonance imaging

All in vivo magnetic resonance (MR) experiments were conducted on a 14.1 Tesla vertical MR system (Agilent Technologies, Palo-Alto, CA, USA) equipped with 100G/cm gradients and a single tuned millipede 1H proton coil (ØI = 40mm). For each imaging session, mice were anesthetized using isoflurane (1-1.5% in O2) and
positioned in a dedicated cradle maintaining constant anesthesia and placed in the MR
bore; respiration and temperature were continuously monitored during all acquisitions to
ensure animal well-being and data reproducibility. Susceptibility Weighted Imaging
(SWI) was performed to detect the potential presence of hemorrhagic lesions, using the
following parameters: gradient-echo scheme, field of view (FOV) = 20 x 20 mm², matrix
= 256 x 256, 16 slices, 0.4 mm slice thickness, 0.1 mm interslice gap, number of
averages = 16, echo time (TE)/ repetition time (TR) = 4.60 / 140 ms, flip angle = 10
degrees. T2-weighted (T2W) images were also acquired in a fast-spin-echo scheme to
calculate brain and ventricle volumes, using the same FOV geometry as SWI and the
following parameters: number of averages = 8, TE/TR = 21.38 / 2500 ms, flip angle = 90
degrees. For each animal, total brain and ventricles were manually delineated on the
T2W images, their volumes calculated, and the ventricle/brain ratio computed.

Isolation of native brain capillary ECs

Individual brain capillary ECs were obtained as previously described. Brains
were denuded of surface vessels, and two 1 mm-thick were excised and homogenized
in artificial cerebrospinal fluid (aCSF; 124 mM NaCl, 3 mM KCl, 2 mM MgCl₂, 2 mM
CaCl₂, 1.25 mM Na₂HPO₄, 26 mM NaHCO₃, and 4 mM glucose) using a Dounce
homogenizer. The homogenate was filtered through a 70 µM filter, and capillary
networks captured on the filter were transferred to a new tube. Individual cells were
isolated by enzymatic digestion with 0.5 mg/ml neutral protease and 0.5 mg/ml elastase
(Worthington Biochemical Corporation) in endothelial cell (EC) isolation solution (55 mM
NaCl, 80 mM Na-glutamate, 6 mM KCl, 2 mM MgCl₂, 10 mM glucose, 0.1 mM CaCl₂, 10
mM HEPES, pH 7.3) for 45 min at 37°C. Following this, 0.5 mg/ml collagenase type I
(Worthington Biochemical Corporation) was added, and a second 2-min incubation at 37°C was performed. Digested networks were washed in ice-cold EC isolation solution, then triturated with a fire-polished glass Pasteur pipette to produce individual ECs. All cells used for this study were freshly dissociated on the day of experimentation.

Isolation of cerebral arteriolar ECs

Single arterial ECs were isolated as previously described. Cerebral arteries were dissected from mouse brains and washed in Mg-PSS. Arteries were transferred to EC isolation solution supplemented with 0.5 mg/mL neutral protease and 0.5 mg/mL elastase (Worthington Biochemical Corporation, Lakewood, NJ, USA), and incubated for 40 min at 37°C. Following this, 0.5 mg/mL collagenase type I (Worthington Biochemical Corporation) was added for an additional 2 min incubation at 37°C. A single-cell suspension was prepared by washing digested arteries three times with EC isolation solution to remove enzymes and triturating with a fire-polished glass pipette to dissociate cells. All cells used for this study were freshly dissociated on the day of experimentation.

Whole-cell patch-clamp electrophysiology

Enzymatically isolated native ECs were transferred to a recording chamber (Warner Instruments, Hamden, CT, USA) and allowed to adhere to glass coverslips for 15 minutes at room temperature. Pipettes were fabricated from borosilicate glass (1.5 mm outer diameter, 1.17 mm inner diameter; Sutter Instruments, Novato, CA, USA), fire-polished to yield a tip resistance of 3–6 MΩ. Currents were recorded at room temperature using an Axopatch 200B amplifier (Molecular Devices, San Jose, CA, USA)
equipped with an Axon CV 203BU headstage and Digidata 1440A digitizer (Molecular Devices) for all patch-clamp electrophysiology experiments. Currents were filtered at 1 kHz and digitized at 10 kHz. Kir2.1 currents were recorded using the conventional whole-cell configuration at a holding potential of -50 mV, with 400 ms ramps from -100 to +40 mV. The external bathing solution was composed of 134 mM NaCl, 6 mM KCl, 1 mM MgCl₂, 2 mM CaCl₂, 10 mM glucose, and 10 mM HEPES. The composition of the pipette solution was 10 mM NaCl, 30 mM KCl, 10 mM HEPES, 110 mM K⁺ aspartate, and 1 mM MgCl₂ (pH 7.2). Kir2.1 currents were activated by increasing extracellular [K⁺] concentration from 6 to 60 mM ([NaCl] adjusted to maintain isotonic solution) and blocked using BaCl₂ (10 μM). All recordings were performed at room temperature. Clampex and Clampfit software (pClamp version 10.2; Molecular Devices) were used for data acquisition and analysis, respectively.

**Pressure myography**

The current best practices guidelines for pressure myography experiments were followed³⁰. Pressure myograph experiments were performed on cerebral arteries and parenchymal arteriole-capillary preparations¹⁹. Surface cerebral arteries were carefully isolated and mounted between two glass cannulas (approximate outer diameter 40–50 μm) in a pressure myograph chamber (Living Systems Instrumentation, St Albans City, VT, USA) and secured by a nylon thread. Intraluminal pressure was controlled using a servo-controlled peristaltic pump (Living Systems Instrumentation), and preparations were visualized with an inverted microscope (Accu-Scope Inc., Commack, NY, USA) coupled to a USB camera (The...
Imaging Source LLC, Charlotte, NC, USA). Changes in luminal diameter were assessed using IonWizard software (version 7.2.7.138; IonOptix LLC, Westwood, MA, USA). Arteries were bathed in warmed (37°C), oxygenated (21% O2, 6% CO2, 73% N2) PSS (119 mM NaCl, 4.7 mM KCl, 21 mM NaHCO3, 1.17 mM MgSO4, 1.8 mM CaCl2, 1.18 mM KH2PO4, 5 mM glucose, 0.03 mM EDTA) at an intraluminal pressure of 5 mmHg. Following equilibration for 15 min, intraluminal pressure was increased to 110 mmHg, and vessels were stretched to their approximate \textit{in vivo} length, after which pressure was reduced back to 5 mmHg for an additional 15 min. Vessel viability was assessed for each preparation by evaluating vasoconstrictor responses to high extracellular [K+] PSS, made isotonic by adjusting the [NaCl] (60 mM KCl, 63.7 mM NaCl). Arteries that showed less than 10% constriction in response to elevated extracellular [K+] were excluded from further investigation. Changes in lumen diameter were recorded at different concentrations of extracellular [K+] (8 to 60 mM, [NaCl] adjusted to maintain isotonic solution). Arteries were pressurized to 20 mmHg and superfused with 10 nM endothelin-1 to induce vasoconstriction. Passive lumen diameter was determined by superfusing vessels Ca2+-free PSS supplemented with EGTA (2 mM) and the voltage-dependent Ca2+ channel blocker diltiazem (10 μM) to inhibit SMC contraction. Change in diameter was calculated at each [K+] concentration as the change in diameter (%) = (change in lumen diameter / passive diameter) × 100.

Parenchymal arterioles with intact capillary segments deriving from the middle cerebral artery were carefully dissected, cannulated, and secured onto a pressure myograph chamber using the same method described above. Preparations were bathed in a warmed, oxygenated aCSF solution at an intraluminal pressure of 5 mmHg.
Following equilibration for 15 min, intraluminal pressure was increased to 20 mmHg and superfused with 10 nM endothelin-1 to induce vasoconstriction. Localized application of drugs onto the capillary extremities was achieved by placing a micropipette attached to a Picospripter III (Parker Hannifin, Cleveland, OH, USA) adjacent to capillary segments. Capillaries were stimulated by locally applying a 7 s pulse of aCSF containing elevated $[K^+] (10 \text{ mM}, [\text{NaCl}]$ adjusted to maintain isotonic solution) onto capillary extremities. To determine preparation viability, SNP (10 µM) was superfused into the circulating aCSF. Changes in lumen diameter were calculated as vasodilation (%) = (change in lumen diameter / baseline diameter) $\times$ 100.

Assessment of functional hyperemia in the brain using laser Doppler flowmetry

Functional hyperemia in the brain was assessed as previously described 19. Mice were anesthetized with isoflurane (5% induction, 2% maintenance), the head was immobilized in a stereotaxic frame, and the skull was exposed. The skull of the right hemisphere was carefully thinned using a drill to visualize the surface vasculature of the somatosensory cortex. Isoflurane anesthesia was replaced with combined $\alpha$-chloralose (50 mg/kg, i.p.) and urethane (750 mg/kg, i.p.) to eliminate confounding vasodilatory effects of isoflurane. Changes in perfusion were assessed via a laser-Doppler flowmetry probe (PeriFlux System PF5000, Perimed AB, Jakobsberg, Sweden) positioned directly above the somatosensory cortex. The contralateral whiskers were stimulated for either 1, 2, or 5 s, and changes in perfusion were recorded. Contralateral whiskers were stimulated three times at 2 min intervals. Ipsilateral whiskers were also stimulated as a control for potential vibration artifacts. Changes in perfusion were calculated as \%Δ Blood flow = (perfusion during stimulus / baseline perfusion) $\times$ 100. Kinetics of the
response, including latency, duration, rise rate, and decay rate, were also obtained and analyzed.

**Y-Maze behavioral assay**

Memory function was assessed using the Y-maze (Maze Engineers, Skokie, IL, USA) using two different configurations.

The spontaneous alternation behavioral assay was used to assess short-term spatial working memory. Mice were placed into one of the three arms of the maze (start arm) and were allowed to explore all three arms for 10 minutes. Session videos were recorded and analyzed using Ethovision XT software (version 16.0.1536, Noldus Information Technology, Leesburg, VA, USA). Spontaneous alternation was evaluated by scoring the order of entries into each arm during the 10-minute recording period. Spontaneous alternation was calculated as alternation index (%) = (number of spontaneous alternations/max alternation) \times 100, where the spontaneous alternation is defined as the number of consecutive entries into each arm of the maze in any order without any repeats, and the max alternation is the total number of alternations possible (max alternation = total number of arm entries – 2).

The novel arm configuration was used to evaluate longer-term spatial reference memory. This test was performed across 2 days. On day one, mice were placed into one of the three arms of the maze (start arm) and allowed to explore only two arms for 10 minutes (training trial). On day two, the test trial was conducted with the closed arm opened, which served as the novel arm. Mice were returned to the maze via the same start arm and were allowed to explore all three arms for 10 minutes. Session videos were recorded and analyzed using Ethovision XT software (version 16.0.1536, Noldus Information Technology, Leesburg, VA, USA). Spontaneous alternation was evaluated by scoring the order of entries into each arm during the 10-minute recording period. Spontaneous alternation was calculated as alternation index (%) = (number of spontaneous alternations/max alternation) \times 100, where the spontaneous alternation is defined as the number of consecutive entries into each arm of the maze in any order without any repeats, and the max alternation is the total number of alternations possible (max alternation = total number of arm entries – 2).
were recorded and analyzed using Ethovision XT software, and the time spent in the novel arm and entries into the novel arm were measured and analyzed.

**Measurement of [ATP] in brain capillary ECs**

The intracellular [ATP] was determined by lysing brain capillary ECs and quantifying luminescence produced by luciferase-induced conversion of ATP to light using the CellTiter-Glo Assay 3D (Promega, Madison, WI, USA). Approximately ~500 ECs were added to each reaction. To further ensure equal input of ECs, the CellTiter-Glo 3D Assay was multiplexed with CellTox Green Assay (Promega), a fluorescent dye that selectively and quantitatively binds double-stranded DNA. Dye fluorescence is directly proportional to DNA concentration and the number of cells in each assay. ATP concentration (luminescence) was normalized to the number of cells (fluorescence) to determine intracellular ATP per cell. Luminescence and fluorescence (485_{Ex}/538_{Em}) were measured using a FlexStation 3 (Molecular Devices). A serial tenfold dilution of ATP (1 nM to 1 uM; 80 µl contains 8^{-14} to 8^{-11} moles of ATP, respectively) was measured to calibrate the linear working range of ATP detection. Data were expressed as the ratio of ATP luminescence/DNA fluorescence.

**Statistical analysis**

All summary data are presented as means ± SEM. Statistical analyses and graphical presentations were performed using GraphPad Prism software (version 9.4.1, GraphPad Software, Inc, USA). The value of n refers to the number of cells for patch-clamp electrophysiology experiments and measurement of [ATP], vessel preparations for myography experiments, and animals used for MRI acquisition, the Y-maze.
behavioral assays and functional hyperemia assessment. Statistical analyses were performed using Student's paired or unpaired two-tailed t-test, or repeated measures or non-repeated measures two-way analysis of variance (ANOVA) with a Šidák correction for multiple comparisons. A value of p<0.05 was considered statistically significant.
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Author contributions

S.E. and D.B.G. initiated and supervised the project. S.E. designed the experiments. X.G. and M.M.C. performed in vivo MRI experiments. S.A., E.Y., and A.S.S. performed patch-clamp electrophysiology experiments. E.Y. and P.T. performed myography experiments. P.T. conducted in vivo functional hyperemia experiments and performed Y-maze behavioral tests. E.Y. conducted luciferase assay for the measurement of the cellular level of ATP. P.T., E.Y., S.A., and S.E. analyzed the data. P.T. and S.E. wrote the manuscript and prepared the figures. P.T., E.Y., C.L.D., D.B.G., and S.E. revised the manuscript.

Competing interests

The authors declare that they have no competing interests.
Data and materials availability

All data needed to evaluate the conclusions are present in the paper or the Supplementary Materials.
Figure legends

**Fig. 1. Age-dependent loss of Kir2.1 channel activity and capillary-to-arteriole dilation in Col4a1+/G394V mice.** (A) Illustration of the brain capillary EC isolation procedure. Scale bar = 10 µm. (B) Representative I-V traces and summary data showing Kir2.1 current densities in freshly isolated capillary ECs from 3 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 4–5 cells from 4 to 5 animals per group, ns = not significant, unpaired t-test). (C) Representative I-V traces and summary data showing Kir2.1 current densities in freshly isolated capillary ECs from 12 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 7–8 cells from 4 animals per group; *p<0.05, unpaired t-test). (D) Illustration of the microvascular preparation. Parenchymal arterioles with intact capillaries were carefully dissected and cannulated onto a pressure myograph chamber, and compounds of interest were focally applied to capillary extremities. Scale bar = 50 µm. (E and F) Representative traces (E) and summary data (F) showing K⁺ (10 mM, blue box)-induced dilation of upstream arterioles in preparations from 3 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 6 preparations from 3 animals per group, ns = not significant, unpaired t-test). (G) The dilation produced by superfusing SNP (10 µM) in preparations from 3 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 6 preparations from 3 animals per group, ns = not significant, unpaired t-test). (H and I) Representative traces (H) and summary data (I) showing K⁺ (10 mM, blue box)-induced dilation of upstream arterioles in preparations from 12 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 11 preparations from 6 to 7 animals per group, *p<0.05, unpaired t-test). (J) The dilation produced by superfusing SNP (10 µM) in preparations from 12 M-old Col4a1+/+ and
Col4a1+/G394V mice (n = 6–8 preparations from 3 to 5 animals per group, ns = not significant, unpaired t-test).

Fig. 2. Age-dependent impairment of functional hyperemia in Col4a1+/G394V mice.
(A) Illustration demonstrating the functional hyperemia assessment procedure in the mouse somatosensory cortex. (B) Illustration demonstrating the parameters that were analyzed. (C and D) Representative traces (C) and summary data (D) showing the increase in blood flow following 1 s contralateral whisker stimulation (WS) in 3 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 6 animals per group, ns = not significant, unpaired t-test). (E to H) Latency (E), duration (F), rise rate (G), and decay rate (H) were also analyzed (n= 6 animals per group, ns = not significant, unpaired t-test). (I and J) Representative traces (I) and summary data (J) showing the increase in blood flow following 1 s contralateral WS in 12 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 6 animals per group, *p<0.05, unpaired t-test). (K to N) Latency (K), duration (L), rise rate (M), and decay rate (N) were also analyzed (n = 6 animals per group, *p<0.05, ns = not significant, unpaired t-test). (O and P) Representative traces (O) and summary data (P) showing the increase in blood flow following 5 s contralateral WS in 3 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 6 animals per group, ns = not significant, unpaired t-test). (Q to T) Latency (Q), duration (R), rise rate (S), and decay rate (T) were also analyzed (n= 6 animals per group, ns = not significant, unpaired t-test). (U and V) Representative traces (U) and summary data (V) showing the increase in blood flow following 5 s contralateral WS in 12 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 6 animals per group,
*p<0.05, unpaired t-test). (W to Z) Latency (W), duration (X), rise rate (Y), and decay rate (Z) were also analyzed (n = 6 animals per group, *p<0.05, unpaired t-test).

**Fig. 3. Age-dependent memory deficits in Col4a1+/G394V mice.** (A) Illustration of the Y-maze spontaneous alternation behavior assay showing examples of a spontaneous (top) and nonspontaneous (bottom) alternation. (B) Summary data showing alternation index, an indicator spatial working memory, in 3 M-old *Col4a1*+/+ and *Col4a1*/+/G394V mice (n = 10 animals per group, ns = not significant, unpaired t-test). (C and D) Summary data showing max alternation (C) and distance moved (D), indicative of exploratory activity, in 3 M-old *Col4a1*+/+ and *Col4a1*/+/G394V mice (n = 10 animals per group, ns = not significant, unpaired t-test). (E) Summary data showing the alternation index of 12 M-old *Col4a1*+/+ and *Col4a1*/+/G394V mice (n = 10 animals per group, *p<0.05, unpaired t-test). (F and G) Summary data showing max alternation (F) and distance moved (G) in 12 M-old *Col4a1*+/+ and *Col4a1*/+/G394V mice (n = 10 animals per group, ns = not significant, unpaired t-test). (H) Illustration demonstrating typical and impaired Y-maze novel arm behavior. (I) Representative heatmaps showing the time (s) 3 M-old *Col4a1*+/+ and *Col4a1*/+/G394V mice spent in areas of the Y-maze during the novel arm test. (J and K) Summary data showing the time spent (J) and entries (K) into the novel arm (n = 8–9 animals per group, ns = not significant, unpaired t-test). (L) Representative heatmaps showing the time (s) 12 M-old *Col4a1*+/+ and *Col4a1*/+/G394V mice spent in areas of the Y-maze during the novel arm test. (M and N) Summary data showing the time spent (M) and entries (N) into the novel arm (n = 10 animals per group, *p<0.05, unpaired t-test).

**Fig. 4. PIP2 depletion reduces Kir2.1 currents in 12 M-old Col4a1+/G394V mice.** (A) Representative I-V traces and summary data showing Kir2.1 current densities in freshly
isolated cerebral artery ECs from 12 M-old $Col4a1^{+/+}$ and $Col4a1^{+/G394V}$ mice with internal solution supplemented with diC8-PIP2 (10 µM) ($n = 6$ cells from 4 animals per group, ns = not significant, unpaired t-test). (B) Representative I-V traces and summary data showing Kir2.1 current densities in freshly isolated brain capillary ECs from 12 M-old $Col4a1^{+/+}$ and $Col4a1^{+/G394V}$ mice with internal solution supplemented with diC8-PIP2 (10 µM) ($n = 5–6$ cells from 4 to 5 animals per group, ns = not significant, unpaired t-test). (C) PIP2 depletion pathway. (D) Representative I-V traces and summary data showing Kir2.1 current densities in freshly isolated brain capillary ECs from 12 M-old $Col4a1^{+/+}$ and $Col4a1^{+/G394V}$ mice treated with the PI3K blocker GSK1059615 (10 nM) ($n = 4–8$ cells from 3 to 6 animals per group, ns = not significant, unpaired t-test). (E and F) Representative trace (E) and summary data (F) showing K$^+$ (10 mM, blue box)-induced dilation of upstream arterioles in preparations from 12 M-old $Col4a1^{+/+}$ mice before and after superfusing the PI3K blocker GSK1059615 (10 nM, 30 min) ($n = 6$ preparations from 5 animals per group, ns = not significant, paired t-test). (G and H) Representative trace (G) and summary data (H) showing K$^+$ (10 mM, blue box)-induced dilation of upstream arterioles in preparations from 12 M-old $Col4a1^{+/G394V}$ mice before and after superfusing the PI3K blocker GSK1059615 (10 nM, 30 min) ($n = 6$ preparations from 5 animals per group, *$p<0.05$, paired t-test).

**Fig. 5.** Chronic PI3K inhibition restores Kir2.1 currents, K$^+$-induced dilation, functional hyperemia, and memory deficits in 12 M-old Col4a1$^{+/G394V}$ mice. (A) Illustration showing GSK1059615 treatment plan. (B) Representative I-V traces and summary data showing Kir2.1 current densities in freshly isolated capillary ECs from 12 M-old $Col4a1^{+/+}$ and $Col4a1^{+/G394V}$ mice treated with vehicle (saline) or GSK1059615 (10 nM, 30 min) ($n = 6$ preparations from 5 animals per group, ns = not significant, paired t-test).
mg/kg) for 28 days, s.c. (n = 5–8 cells from 3 to 4 animals per group, *p<0.05, ns = not significant, non-repeated measures two-way ANOVA). (C and D) Representative traces (C) and summary data (D) showing K⁺ (10 mM, blue box)-induced dilation of upstream arterioles in preparations from 12 M-old Col4a1+/+ and Col4a1+/G394V mice treated with vehicle (saline) or GSK1059615 (10 mg/kg) for 28 days, s.c. (n = 6–7 preparations from 3 animals per group, *p<0.05, ns = not significant, non-repeated measures two-way ANOVA). (E and F) Representative traces (E) and summary data (F) showing the increase in blood flow following 1 s contralateral whisker stimulation (WS) in 12 M-old Col4a1+/+ and Col4a1+/G394V mice treated with vehicle (saline) or GSK1059615 (10 mg/kg) for 28 days, s.c. (n = 5 animals per group, *p<0.05, ns = not significant, non-repeated measures two-way ANOVA). (G to J) Latency (G), duration (H), rise rate (I), and decay rate (J) were also analyzed (n= 5 animals per group, *p<0.05, ns = not significant, non-repeated measures two-way ANOVA). (K) Summary data showing alternation index, an indicator spatial working memory, of 12 M-old Col4a1+/+ and Col4a1+/G394V mice before and after treatment with vehicle (saline) or GSK1059615 (10 mg/kg) for 28 days, s.c. (n = 8–9 animals per group, *p<0.05, ns = not significant, repeated measures two-way ANOVA). (L and M) Summary data showing max alternation (L) and distance moved (M), indicative of exploratory activity, in 12 M-old Col4a1+/+ and Col4a1+/G394V mice before and after treatment with vehicle (saline) or GSK1059615 (10 mg/kg) for 28 days, s.c. (n = 8–9 animals per group, *p<0.05, ns = not significant, repeated measures two-way ANOVA).
Supplemental figure legends

Fig. 1 – supplement 1. Kir2.1 currents in cerebral artery ECs. (A) Illustration of the cerebral artery EC isolation procedure. Scale bar = 10 μm. (B) Representative I-V traces and summary data showing Kir2.1 current densities in freshly isolated cerebral artery ECs from 3 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 5–6 cells from 5 animals per group, ns = not significant, unpaired t-test). (C) Representative I-V traces and summary data showing Kir2.1 current densities in freshly isolated cerebral artery ECs from 12 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 6–10 cells from 3 to 4 animals per group; *p<0.05, unpaired t-test). (D and E) Representative traces (D) and summary data (E) demonstrating the vasodilator response to raising extracellular [K+] in cerebral arteries from 12 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 9–13 preparations from 6 to 10 animals per group, *p<0.05, non-repeated measures two-way ANOVA). (F) Summary data showing the contractile response to 30 and 60 mM [K+] in cerebral arteries from 12 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 7–12 preparations from 5 to 10 animals per group, ns = not significant, unpaired t-test).

Fig. 1 – supplement 2. High field magnetic resonance imaging. (A) Representative susceptibility weighted (SWI) and T2 weighted (T2W) MRI images from 12 M-old Col4a1+/+ and Col4a1+/G394V mice. (B) Quantification of ventricle/brain ratio for 12 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 7 animals per group, ns = not significant, unpaired t-test).

Fig. 2 – supplement 1. Functional hyperemic response following 2 s whisker stimulation. (A and B) Representative traces (A) and summary data (B) showing the increase in blood flow following 2 s contralateral whisker stimulation (WS) in 3 M-old
Col4a1+/+ and Col4a1+/G394V mice (n = 6 animals per group, ns = not significant, unpaired t-test). (C to F) Latency (C), duration (D), rise rate (E) and decay rate (F) were also analyzed (n= 6 animals per group, ns = not significant, unpaired t-test). (G and H) Representative traces (G) and summary data (H) showing the increase in blood flow following 2 s contralateral WS in 12 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 6 animals per group, *p<0.05, unpaired t-test). (I to L) Latency (I), duration (J), rise rate (K) and decay rate (L) were also analyzed (n = 6 animals per group, *p<0.05, ns = not significant, unpaired t-test).

**Fig. 2 – supplement 2. Ipsilateral whisker stimulation.** (A to C) Summary data showing no change in blood flow following ipsilateral whisker stimulation for 1 s (A), 2 s (B), and 5 s (C) in 3 M- and 12 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 6 animals per group, ns = not significant, unpaired t-test).

**Fig. 4 – supplement 1. ATP levels in brain capillary ECs.** (A) PIP2 synthesis pathway. (B) Summary data showing normalized [ATP] in brain capillary ECs from 12 M-old Col4a1+/+ and Col4a1+/G394V mice. (n = 9 sets of ~500 cells from three mice per group, ns = not significant, unpaired t-test). (C) PIP2 depletion pathway. (D) Representative I-V traces and summary data showing Kir2.1 current densities in freshly isolated brain capillary ECs treated with the PLC inhibitor U73122 (10 µM) from 12 M-old Col4a1+/+ and Col4a1+/G394V mice (n = 4 cells from 3 animals per group, *p<0.05, unpaired t-test).

**Fig. 4 – supplement 2. GSK1059615 vehicle and SNP response.** (A and B) Representative trace (A) and summary data (B) showing K+ (10 mM, blue box)-induced dilation of upstream arterioles in preparations before and after superfusing the vehicle.
Fig. 5 – supplement 1. **Functional hyperemic response following 2 s and 5 s whisker stimulation in GSK1059615-treated animals.** (A and B) Representative traces (A) and summary data (B) showing the increase in blood flow following 2 s contralateral whisker stimulation (WS) in 12 M-old Col4a1+/+ and Col4a1+/G394V mice treated with vehicle (saline) or GSK1059615 (10 mg/kg) for 28 days, s.c. (n = 5 animals per group, *p<0.05, ns = not significant, non-repeated measures two-way ANOVA). (C to F) Latency (C), duration (D), rise rate (E) and decay rate (F) were also analyzed (n= 5 animals per group, *p<0.05, ns = not significant, non-repeated measures two-way ANOVA). (G and H) Representative traces (G) and summary data (H) showing the increase in blood flow following 5 s contralateral WS in 12 M-old Col4a1+/+ and Col4a1+/G394V mice treated with vehicle (saline) or GSK1059615 (10 mg/kg) for 28 days, s.c. (n = 5 animals per group, *p<0.05, ns = not significant, non-repeated measures two-way ANOVA). (I to L) Latency (I), duration (J), rise rate (K) and decay rate (L) were also analyzed (n= 5 animals per group, *p<0.05, ns = not significant, non-repeated measures two-way ANOVA).
analyzed (n= 5 animals per group, *p<0.05, ns = not significant, non-repeated measures two-way ANOVA).

**Fig. 5 – supplement 2. Ipsilateral whisker stimulation in GSK1059615 treated animals.** (A to C) Summary data showing no change in blood flow following ipsilateral whisker stimulation for 1 s (A), 2 s (B), and 5 s (C) in 12 M-old Col4a1+/+ and Col4a1+/G394V mice treated with vehicle (saline) or GSK1059615 (10 mg/kg) for 28 days, s.c. (n = 5 animals per group, ns = not significant, non-repeated measures two-way ANOVA).
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