Modulation of dorsal premotor cortex disrupts neuroplasticity of primary

motor cortex in young and older adults

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Premotor effects on motor cortex plasticity in young and older adults
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Key points

- The influence of dorsal premotor cortex (PMd) on M1 plasticity is likely mediated by late I-wave interneuronal circuits within M1, but this communication may change with advancing age.
- We investigated the effects of intermittent theta burst stimulation (iTBS) to M1 when preceded by iTBS to PMd on transcranial magnetic stimulation (TMS) measures of M1 excitability in young and older adults.
- We found that PMd iTBS disrupts the plasticity induction of M1 iTBS in both young and older adults when measured with posterior-anterior (PA, early I-waves) current TMS, but not with anterior-posterior (AP, late I-waves) TMS measures of M1 excitability.
- The plasticity of early I-waves within M1 are specifically influenced by PMd iTBS in both young and older adults, suggesting that this communication is preserved with ageing.

1 Abstract

2	Previous research using transcranial magnetic stimulation (TMS) demonstrates that dorsal
3	premotor cortex (PMd) influences neuroplasticity within primary motor cortex (M1). This
4	communication is likely mediated by indirect (I) wave inputs within M1, the activity of
5	which are altered by ageing. However, it remains unclear if age-related changes in the I-wave
6	circuits modify the influence of PMd on M1 plasticity. The present study therefore
7	investigated the influence of PMd on the plasticity of early and late I-wave circuits within M1
8	of young and older adults. 15 young (mean \pm SD; 24.7 \pm 5.0 years) and 15 older adults (67.2
9	\pm 5.4 years) participated in two experimental sessions that examined the effects of
10	intermittent theta burst stimulation (iTBS) to M1 when preceded by iTBS (PMd iTBS-M1
11	iTBS) or sham stimulation (PMd sham-M1 iTBS) to PMd. Changes in M1 excitability post-
12	intervention were assessed with motor evoked potentials (MEP) recorded from right first
13	dorsal interosseous muscle, with posterior-to-anterior (PA) and anterior-to-posterior (AP)
14	current single-pulse TMS assessing corticospinal excitability (PA1mV; AP1mV; PA0.5mV, early I-
15	wave; AP _{0.5mV} , late I-wave). Although PA _{1mV} did not change post-intervention ($P = 0.628$),
16	PMd iTBS-M1 iTBS disrupted the expected facilitation of AP_{ImV} (to M1 iTBS in isolation) in
17	young and older adults ($P = 0.002$). Similarly, PMd iTBS-M1 iTBS disrupted PA _{0.5mV}
18	facilitation in young and older adults ($P = 0.030$), whereas AP _{0.5mV} facilitation was not
19	affected in either group ($P = 0.218$). This suggests that while PMd specifically influences the
20	plasticity of early I-wave circuits, this communication is preserved in older adults.

21 **Keywords:** Transcranial magnetic stimulation, ageing, dorsal premotor cortex,

22 neuroplasticity

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24 Introduction

25 One of the universal effects of ageing is widespread deficits in motor function. Although these deficits occur at all levels of the motor system, the structural, functional, and 26 biochemical changes within the brain are important (Seidler et al., 2010). In particular, 27 28 alterations to the ability of the brain's motor system to continuously modify its structure and 29 function are a critical factor. Termed neuroplasticity, this process is initially mediated by changes in the strength of synaptic communication with long-term potentiation (LTP) and 30 depression (LTD), and underpins the ability to learn new motor skills (Buonomano & 31 Merzenich, 1998; Sanes & Donoghue, 2000). While the capacity for neuroplastic change is 32 33 present across the lifespan, some studies using non-invasive brain stimulation (NIBS) show reduced plasticity in older adults (Müller-Dahlhaus et al., 2008; Fathi et al., 2010; Todd et 34 al., 2010; Freitas et al., 2011). This reduced plasticity may contribute to the motor deficits 35 36 that limit the ability of older adults to perform essential activities of daily life. However, the neurophysiological mechanisms underpinning these changes with advancing age remain 37 unclear. 38

Transcranial magnetic stimulation (TMS) is a type of NIBS that allows investigation of 39 specific neuronal networks within the motor system with high temporal resolution. 40 41 Application of TMS over primary motor cortex (M1) produces a complex series of descending volleys within corticospinal neurons that summate at the spinal cord, resulting in 42 a motor evoked potential (MEP) in targeted muscles (Di Lazzaro et al., 1998; Rossini et al., 43 2015). The first of these waves likely represent direct activation of corticospinal neurons, 44 45 whereas subsequent waves are thought to reflect the indirect activation of interneuronal inputs to the corticospinal neurons (Di Lazzaro et al., 2012; Ziemann, 2020). These responses 46 are referred to as indirect (I) waves and are named early (I_1) or late (I_2, I_3) based on the order 47

of their appearance, which occurs with a periodicity of ~ 1.5 ms (Di Lazzaro *et al.*, 2012; 48 Ziemann, 2020). Early and late I-waves can be preferentially recruited by applying low-49 intensity single-pulse TMS with different current directions (Sakai et al., 1997; Di Lazzaro et 50 al., 2001; Ni et al., 2010). For example, a posterior-to-anterior (PA) current (relative to the 51 central sulcus) preferentially recruits early I-waves, whereas an anterior-to-posterior (AP) 52 current preferentially recruits late I-waves (Sakai et al., 1997; Di Lazzaro et al., 2001; Ni et 53 54 al., 2010). Using these measures, previous work has shown that the ability to recruit late Iwaves predicts the response to plasticity-inducing TMS paradigms over M1 (Hamada et al., 55 56 2013; Wiethoff et al., 2014) and that the late I-waves are behaviourally relevant to the acquisition of fine motor skills (Hamada et al., 2014). 57 I-wave circuits are also involved in mediating the communication between other motor nodes 58 and M1 (Groppa et al., 2012; Volz et al., 2015; Spampinato et al., 2020; Opie et al., 2022; 59 Casarotto et al., 2023), which form a wider network that influences M1 plasticity and 60 61 learning (Huang et al., 2018; Liao et al., 2022). In particular, the dorsal premotor cortex (PMd) facilitates the planning, prediction, and correction of movements during motor 62 learning by updating the activity of M1 (Chouinard et al., 2005; Nowak et al., 2009; Parikh & 63 64 Santello, 2017). Previous studies have demonstrated that the application of repetitive TMS (rTMS) techniques (such as theta burst stimulation; TBS) over PMd is able to modify M1 65 excitability, plasticity, and motor skill acquisition (Huang et al., 2018; Meng et al., 2020). 66 Furthermore, while PMd influences both early and late I-wave excitability (Liao et al., 2023), 67 there is a stronger effect on the late I-waves (Volz et al., 2015; Aberra et al., 2020). Taken 68 69 together, it is likely that the influence of late I-waves on M1 plasticity reflects inputs from PMd. 70

Given the role of late I-wave circuits in mediating PMd-M1 communication, changes in late 71 I-wave activity may affect the influence of PMd on M1 plasticity. In particular, late I-wave 72 activity is known to be altered with advancing age (Opie et al., 2018). Age-related changes in 73 I-wave excitability have been investigated using the paired-pulse TMS protocol short 74 intracortical facilitation (SICF) (Opie et al., 2018), which revealed reduced I-wave 75 excitability and a specific delay in the temporal characteristics of the late I-waves in older 76 77 adults (Opie et al., 2018). Importantly, this delay influences NIBS-induced plasticity and is associated with specific aspects of motor behaviour in older adults (Opie et al., 2018; Opie et 78 79 al., 2020). In addition, it is also known that PMd-M1 effective connectivity (Ni et al., 2015) and direct PMd modulation of early I-waves within M1 is reduced in older adults (Liao et al., 80 2023). Consequently, it is possible that the influence of PMd on M1 plasticity is altered with 81 82 advancing age, but this remains to be tested.

The purpose of the present study was, therefore, to investigate the influence of PMd on the 83 84 plasticity of early and late I-wave circuits in M1 of young and older adults. We applied intermittent TBS (iTBS) over PMd in young and older participants and assessed how this 85 influenced the neuroplastic response of M1 to iTBS. Different I-wave circuits were assessed 86 by varying the direction of current used to apply TMS over M1. Although we expected iTBS 87 over PMd to selectively modulate the plasticity of late I-wave circuits, we hypothesised that 88 the effect of PMd on M1 plasticity would be weaker in older adults, given the likely 89 alterations in late I-wave activity and PMd-M1 connectivity with advancing age. 90

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94 Methods

95 Sample Size and Participants

96	15 young (mean \pm standard deviation, 24.7 \pm 5.0 years; range , 19-36 years) and 15 older
97	adults (67.2 \pm 5.4 years; 61-78 years) were recruited for the study via advertisements placed
98	on notice boards within The University of Adelaide and the wider community, in addition to
99	social media platforms. Applicants for the study were excluded if they had a history of
100	psychiatric or neurological disease, current use of medication that affect the central nervous
101	system, pregnancy, metal implants, or left handedness, as assessed by a standard TMS
102	screening questionnaire (Rossi et al., 2011). The experiment was conducted in accordance
103	with the Declaration of Helsinki and was approved by The University of Adelaide Human
104	research Ethics Committee (H-026-2008). Subjects provided written, informed consent prior
105	to participation.

106 Experimental Arrangement

107 All participants attended two experimental sessions where iTBS or sham iTBS was applied to PMd, followed 30 minutes later by plasticity induction within M1 via iTBS (PMd iTBS-M1 108 iTBS, PMd sham-M1 iTBS). The same experimental protocol was used in both sessions Fig. 109 1), with the order of intervention randomised between participants, and a washout period of at 110 least 1 week was used between sessions. As diurnal variations in cortisol are known to 111 influence the neuroplastic response to TMS (Sale et al., 2008), all sessions were completed 112 between 11 am and 5 pm at approximately the same time of day for each participant. 113 During each experimental session, participants were seated in a comfortable chair with their 114 hands resting and relaxed. Surface electromyography (EMG) was recorded from the first 115 dorsal interosseous (FDI) of the right hand using two Ag-AgCl electrodes arranged in a belly-116

tendon montage on the skin above the muscle, with a third electrode attached above the 117 styloid process of the right ulnar used to ground the electrodes. EMG signals were amplified 118 (300x) and filtered (band-pass 20 Hz – 1 kHz) using a CED 1902 signal conditioner 119 (Cambridge Electronic Design, Cambridge, UK) before being digitised at 2 kHz using a CED 120 1401 analogue-to-digital converter. Signal noise associated with mains power was removed 121 using a Humbug mains noise eliminator (Quest Scientific, North Vancouver, Canada). EMG 122 123 signals were stored on a PC for offline analysis. Real-time EMG signals were displayed on an oscilloscope placed in front of the participant to facilitate muscle relaxation during the 124 125 experiment.

126 Experimental Procedures

127 Transcranial magnetic stimulation (TMS). A branding iron coil connected to two Magstim 200² magnetic stimulators (Magstim, Whitland, UK) via a BiStim unit was used to apply 128 TMS to left M1. The coil was held tangentially to the scalp at an angle of 45° to the sagittal 129 130 plane, inducing a posterior-to-anterior (PA) current relative to the central sulcus. The M1 131 hotspot was identified as the location producing the largest and most consistent MEPs within the relaxed FDI muscle of the right hand (Rossini et al., 2015). This location was marked on 132 the scalp for reference and continuously monitored throughout each experimental session. All 133 baseline, post-PMd iTBS, and post-M1 iTBS (5 minutes, 30 minutes) TMS was applied at a 134 rate of 0.2 Hz, with a 10% jitter between trials to avoid anticipation of the stimulus. 135

136 Resting motor threshold (RMT) was recorded as the lowest stimulus intensity producing an 137 MEP amplitude $\geq 50 \ \mu\text{V}$ in at least 5 out of 10 trials during relaxation of the right FDI. RMT 138 was assessed at the beginning of each experimental session and expressed as a percentage of 139 maximum stimulator output (% MSO) (Rossini *et al.*, 2015). Active motor threshold (AMT) 140 was then assessed, defined as the lowest % MSO producing an MEP amplitude $\geq 200 \ \mu\text{V}$ in at least 5 out of 10 trials during concurrent low-level activation (~10% voluntary activation) of the right FDI (Hamada *et al.*, 2013). These measures were then repeated using the AP current by rotating the coil 180°. Then, the stimulus intensities producing a standard MEP amplitude approximating 1 mV (MEP_{1mV}; PA_{1mV}, AP_{1mV}), in addition to an MEP amplitude approximating 0.5 mV (MEP_{0.5mV}; PA_{0.5mV}, AP_{0.5mV}), when averaged over 20 trials, were identified. The same intensities were then applied following PMd iTBS and following M1 iTBS to assess changes in corticospinal excitability.

148 *I-wave recruitment.* To investigate the ability to recruit I-waves, the onset latencies of PA

149 (early) and AP (late) MEPs were assessed relative to the MEP onset generated by direct

activation of corticospinal neurons using a lateral-to-medial (LM) current (Hamada *et al.*,

151 2013). A block of 15 MEP trials in the active FDI was recorded for 110% of AMT_{PA} and

AMT_{AP}, in addition to 150% AMT_{LM} (Hamada *et al.*, 2013). If 150% AMT_{LM} exceeded 100%

153 MSO, 100% MSO was used, or if 150% AMT_{LM} was below 50% MSO, 50% MSO was used

154 (Hamada *et al.*, 2013). The difference in mean onset latencies between PA and LM (PA-LM)

and AP and LM (AP-LM) were calculated as measures of early and late I-wave recruitment

efficiency, respectively (Hamada *et al.*, 2013). In an attempt to reduce the confounding

157 influence of muscle contraction on neuroplasticity induction (Huang *et al.*, 2008;

158 Thirugnanasambandam et al., 2011; Goldsworthy et al., 2015), these measures were recorded

at the start and at the end of the experimental session, at least 45 minutes apart from the

160 plasticity induction of PMd and M1.

161 *Theta burst stimulation (TBS).* Intermittent theta burst stimulation (iTBS) was delivered over

162 left PMd and left M1 using a Magstim Super-rapid stimulator (Magstim, Whitland, UK),

163 connected to an air-cooled figure-of-eight coil. The coil was held tangentially to the scalp, at

an angle of 45° to the sagittal plane, with the handle pointing backwards and laterally,

165	inducing a biphasic pulse with an initial PA current followed by an AP return current (Suppa
166	et al., 2008). In accordance with existing literature, iTBS consisted of bursts of three pulses
167	given at a frequency of 50 Hz. Each burst was repeated at 5 Hz for 2 s, and repeated every 8 s
168	for 20 cycles, totalling 600 pulses (Huang et al., 2005; Huang et al., 2008; Huang et al.,
169	2018; Meng et al., 2020). The location of left PMd was defined as 8% of the distance
170	between the nasion and inion (approximately $2.5 - 3$ cm) anterior to the M1 hotspot,
171	consistent with previous work (Münchau et al., 2002; Koch et al., 2007; Huang et al., 2018;
172	Meng et al., 2020). The location of both the M1 hotspot and left PMd site were logged
173	relative to the MNI-ICBM152 template using Brainsight neuronavigation (Rogue Research,
174	Montreal, Quebec, Canada). These locations were then used to guide the assessment of RMT
175	(RMT_{Rapid}) over M1 with the Magstim Super-rapid stimulator, in addition to the application
176	of iTBS over left PMd and M1 at 70% RMT _{Rapid} .
177	Sham iTBS to left PMd was delivered using a sham figure-of-eight coil (replicating the coil
178	click), with a bar electrode connected to a constant current stimulator (Digitimer,

179 Hertfordshire, UK) placed underneath the coil delivering electrical stimulation (1.5 mA) to

180 the scalp in order to mimic the pulse sensation. Following either intervention, participants

181 provided answers to a visual analogue scale (VAS) questionnaire indexing the degree of

discomfort, muscle activation, and localisation of scalp sensation during PMd iTBS.

183 Data Analysis

Visual inspection of EMG data was completed offline, with any trials obtained from the
resting muscle having EMG activity exceeding 25 µV in the 100 ms prior to stimulus
application excluded from analysis (approximately 6.8% removed). The amplitude of MEPs
obtained from resting muscle recordings was measured peak-to-peak and expressed in mV.
The MEP onset latencies obtained from active muscle recordings was assessed with a semi-

automated process using a custom script within the Signal program (v 6.02, Cambridge 189 Electronic Design) and expressed in ms. MEP latency was recorded as the period from 190 stimulus application to the resumption of voluntary EMG activity. This was defined as the 191 point at which post-stimulus EMG amplitude exceeded the mean EMG amplitude recorded 192 within the 100 ms pre-stimulus, plus 2 standard deviations. MEP onset latencies were 193 averaged over individual trials within each subject and coil orientation. Within each 194 195 participant, the mean LM MEP latencies were subtracted from the mean PA and AP MEP latencies to determine PA-LM and AP-LM MEP latency differences. Following TBS 196 197 interventions, changes in MEP latency differences were quantified by expressing the postintervention responses as a percentage of the baseline responses. Changes in MEP amplitude 198 due to PMd iTBS were quantified by expressing post-PMd iTBS responses as a percentage of 199 200 baseline MEP amplitude. For post-M1 iTBS, changes in MEP amplitude were quantified by expressing post-M1 iTBS responses as a percentage of post-PMd iTBS responses. 201

202 Statistical Analysis

203 Visual inspection and Kolmogorov-Smirnov tests of the data residuals revealed non-normal, positively-skewed distributions for all TMS data. Consequently, generalised linear mixed 204 models (GLMM's), which can account for non-normal distributions (Lo & Andrews, 2015; 205 Puri & Hinder, 2022), were used to perform all statistical analyses. Each model assessing 206 MEP amplitude included single trial data with repeated measures and was fitted with Gamma 207 208 distributions (Puri & Hinder, 2022), with all random subject effects included (intercepts and slopes) (Barr et al., 2013). Identity link functions were used for baseline MEP amplitude and 209 210 latency differences while log link functions were used for post-iTBS normalised MEP amplitude and latency differences (Lo & Andrews, 2015; Puri & Hinder, 2022). To optimise 211 model fit, we tested different covariance structures and the structure providing the best fit 212 213 (assess with the Bayesian Schwartz Criterion; BIC) within a model that was able to converge

was used in the final model. Two-factor GLMMs were used to compare effects of session
(PMd iTBS-M1 iTBS, PMd sham-M1 iTBS) and age (young, older) at baseline in four
separate models for PA_{0.5mV}, AP_{0.5mV}, PA_{1mV}, and AP_{1mV}. A three-factor model was used to
compare the effects of session, age, and orientation (PA, AP) on PA-LM and AP-LM latency
differences at baseline.

Changes in corticospinal excitability following PMd iTBS were investigated by assessing 219 effects of session and age in four separate models for baseline-normalised PA_{1mV}, AP_{1mV}, 220 PA_{0.5mV}, and AP_{0.5mV} MEP amplitude. Changes in corticospinal excitability following PMd 221 222 iTBS-M1 iTBS and PMd sham-M1 iTBS were investigated by assessing effects of session, time (5 minutes, 30 minutes) and age in four separate models for PA_{1mV}, AP_{1mV}, PA_{0.5mV}, and 223 AP_{0.5mV} MEP amplitudes normalised to the mean post-PMd iTBS MEP amplitudes. Changes 224 in I-wave recruitment following the intervention were investigated by assessing effects of 225 session, age, and coil orientation on baseline-normalised average PA-LM and AP-LM latency 226 227 differences. For all models, investigation of main effects and interactions were performed using custom contrasts with Bonferroni correction, and significance was set at P < 0.05. Data 228 for all models are presented as estimated marginal means (EMMs) and 95% confidence 229 intervals (95% CI), whereas pairwise comparisons are presented as the estimated mean 230 difference (EMD) and 95% CI for the estimate. 231

Furthermore, we used Spearman's rank-order correlation to assess the relationship between different variables. Specifically, baseline MEP latency differences were correlated with changes in corticospinal excitability immediately following PMd iTBS to investigate if the ability to recruit I-waves is related to changes in corticospinal excitability. Baseline MEP latency differences were also correlated with changes in corticospinal excitability and I-wave recruitment during the PMd sham-M1 iTBS session to investigate if the ability to recruit I-

waves is related to changes in corticospinal excitability and I-wave latency differences 238 following M1 iTBS. In addition, individual ages were tested against changes in corticospinal 239 excitability immediately following PMd iTBS (during PMd iTBS-M1 iTBS) and PMd sham-240 M1 iTBS to investigate if age is related to changes in corticospinal excitability. Changes in 241 corticospinal excitability following PMd iTBS were also correlated with changes in 242 corticospinal excitability following M1 iTBS (during PMd iTBS-M1 iTBS) to investigate if 243 244 direct PMd modulation of M1 excitability is related to changes in M1 plasticity. Correlations are presented as Spearman's p with false discovery rate-adjusted P-value of 0.05 following 245 246 the Benjamini-Hochberg procedure. Lastly, differences in the perception of discomfort, extent of FDI activation, and localisation of stimulus during PMd iTBS and PMd sham were 247 investigated by comparing VAS responses using paired t-tests with Bonferroni correction (P 248 < 0.0167), with data presented as mean \pm standard deviation. 249

250 **Results**

All participants completed both experimental sessions without adverse reactions. We were 251 252 unable to record PA_{ImV} in one older male participant, $AP_{0.5mV}$ in two older participants (1) female, 1 male), and AP_{1mV} in five participants (1 young female; 3 older females, 1 older 253 male) due to high thresholds of activation (mean $RMT_{PA} = 80.0\%$ MSO, mean $RMT_{AP} =$ 254 73.0% MSO). Baseline stimulation intensities are presented in Table 1. Stimulation 255 intensities for AP_{1mV} differed between sessions ($F_{1.46} = 4.17$, P = 0.047), with post-hoc 256 257 comparisons showing higher intensities for the iTBS session relative to sham session (EMD = 2.3% MSO [0.0, 4.6], P = 0.047). There were no other main effects or interactions for all 258 other baseline stimulation intensities (all P > 0.05). 259

260 Baseline MEP amplitudes for corticospinal excitability and MEP latency differences are

shown in Table 2. For PA_{1mV} MEP amplitude, there was an interaction between session and

age ($F_{1,1121} = 4.194$, P = 0.041), with *post-hoc* comparisons revealing larger MEP amplitude for young participants relative to older participants (EMD = 0.14 mV [0.02, 0.26], P =0.024). For baseline MEP latency differences, responses differed between coil orientations ($F_{1,112} = 165.20$, P < 0.0001), where PA-LM latencies were shorter than AP-LM latencies (EMD = 1.95 ms [1.65, 2.25], P < 0.0001), as expected. There were no main effects or interactions for all other baseline MEP amplitudes or MEP latency differences (all P > 0.05).

268 Changes in corticospinal excitability following PMd iTBS

- 269 The participants' perceptions of PMd iTBS and PMd sham are shown in Table 3. While there
- were no differences between sessions in the extent of discomfort ($t_{29} = 0.25$, P = 0.804) or
- FDI activation ($t_{29} = 0.10$, P = 0.918) experienced by the participants, the locality of

stimulation differed ($t_{29} = 3.98$, P = 0.004), with the sensation of iTBS perceived as more

273 widespread relative to electrical scalp stimulation in sham.

274 Changes in MEP_{1mV} and MEP_{0.5mV} measures of corticospinal excitability following PMd iTBS are shown in Figure 2. PA_{ImV} MEP amplitude did not differ between sessions ($F_{1,1114}$ = 275 0.90, P = 0.343; Fig. 2A) or age groups ($F_{1,1114} = 0.12$, P = 0.726), and there was no 276 interaction between factors ($F_{1,1114} = 2.41$, P = 0.121). AP_{*lmV*} MEP amplitude did not vary 277 between sessions ($F_{1.996} = 2.33$, P = 0.127; Fig. 2B) or age groups ($F_{1.996} = 1.31$, P = 0.252), 278 and there was no interaction between factors ($F_{1,996} = 0.51$, P = 0.476). In contrast, while 279 $PA_{0.5mV}$ MEP amplitude did not differ between age groups ($F_{1.1152} = 0.11$, P = 0.740), 280 responses varied between sessions ($F_{1,1152} = 4.23$, P = 0.040; Fig. 2C), with increased MEP 281 282 amplitude following iTBS relative to sham (EMD = 26.3% [0.7, 51.9], P = 0.044). There was no interaction between factors ($F_{1,1152} = 0.11$, P = 0.741). AP_{0.5mV} MEP amplitude did not 283 vary between sessions ($F_{1,1073} = 1.04$, P = 0.308; Fig. 2D) or age groups ($F_{1,1073} = 2.80$, P =284 285 0.095), and there was no interaction between factors ($F_{1,1073} = 1.03$, P = 0.310).

286 Changes in corticospinal excitability and I-wave recruitment following M1 iTBS.

287 *Corticospinal excitability*

Changes in MEP_{1mV} measures of corticospinal excitability following PMd iTBS-M1 iTBS 288 and PMd sham-M1 iTBS are presented in Figure 3. PA_{1mV} MEP amplitudes (Fig. 3A) did not 289 vary between sessions ($F_{1,2234} = 2.20, P = 0.138$), time points ($F_{1,2234} = 0.15, P = 0.696$), or 290 age groups ($F_{1,2234} = 1.17$, P = 0.279), and there were no interactions between factors (all P >291 0.05). AP_{1mV} MEP amplitudes also did not differ between sessions ($F_{1,1922} = 1.16$, P = 0.281), 292 293 time points ($F_{1,1922} = 1.15$, P = 0.283), or age groups ($F_{1,1922} = 1.70$, P = 0.193), but there was an interaction between session and time ($F_{1,1922} = 10.02$, P = 0.002; Fig 3B). Post-hoc 294 comparisons showed that MEP amplitudes following PMd sham-M1 iTBS was increased at 5 295 296 minutes compared to PMd iTBS-M1 iTBS (EMD = 30.4% [7.5, 53.3], P = 0.009), and compared to 30 minutes (EMD = 30.4% [7.4, 53.3], P = 0.009). There were no other 297 interactions (all P > 0.05). 298

299 Changes in MEP_{0.5mV} measures of corticospinal excitability are presented in Figure 4. While 300 PA_{0.5mV} MEP amplitudes did not differ between time points ($F_{1,2311} = 0.03$, P = 0.874) or age

301 groups ($F_{1,2311} = 0.17$, P = 0.678), responses varied between sessions ($F_{1,2311} = 17.4$, P < 10.17

0.05), with increased MEP amplitudes following PMd sham-M1 iTBS (EMD = 34.3% [17.5,

51.0], P < 0.05). Furthermore, there was an interaction between session, time, and age ($F_{1,2311}$

= 4.71, P = 0.030; Fig. 4A). *Post-hoc* analysis revealed increased MEP amplitudes at 30

minutes for young adults (EMD = 50.7% [20.1, 81.3], P = 0.001), and at 5 (EMD = 43.0%

306 [13.9, 72.0], P = 0.004) and 30 minutes (EMD = 32.0% [3.6, 60.4], P = 0.027) for older

adults following PMd sham-M1 iTBS compared to PMd iTBS-M1 iTBS. For AP_{0.5mV}, MEP

amplitude did not vary between sessions ($F_{1,2141} = 0.13$, P = 0.723) or age groups ($F_{1,2141} =$

309 3.12, P = 0.077) (Fig. 4B). However, responses differed between time points ($F_{1,2141} = 5.91$,

- P = 0.015; Fig. 4C), with *post-hoc* analysis revealing that MEP amplitude was increased at 5
- minutes relative to 30 minutes post-M1 iTBS (EMD = 22.0% [3.9, 40.0], P = 0.017). There
- 312 were no interactions between factors (all P > 0.05).
- 313 *I-wave recruitment*
- There was no difference between sessions ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$, P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), P = 0.399), coil orientations ($F_{1,112} = 0.72$), $F_{1,$
- 315 0.09, P = 0.766), or age groups ($F_{1,112} = 0.38$, P = 0.538), and there were no interactions
- 316 between factors (all P > 0.05).

317 *Correlation analyses*

Baseline PA-LM and AP-LM latencies were not related to changes in single-pulse measures

of corticospinal excitability (PA_{1mV}, AP_{1mV}, PA_{0.5mV}, AP_{0.5mV}) following PMd iTBS (all P >

320 0.05). Baseline PA-LM and AP-LM latencies were not related to changes in single-pulse

321 measures of corticospinal excitability or changes in PA-LM and AP-LM latencies following

- 322 PMd sham-M1 iTBS (all P > 0.05). Furthermore, individual age did not predict changes in
- 323 single-pulse measures of corticospinal excitability following PMd iTBS or corticospinal
- excitability and MEP latency differences following PMd sham-M1 iTBS (all P > 0.05). In
- 325 contrast, while changes in AP_{ImV} MEP amplitude following PMd iTBS were not related to
- changes in AP_{1mV} responses following M1 iTBS ($\rho = -0.361$, P = 0.076; Fig. 5B), changes in

327 PA_{1mV}, PA_{0.5mV}, and AP_{0.5mV} MEP amplitude following PMd iTBS were negatively correlated

- 328 with changes in PA_{1mV} ($\rho = 0.-577$, P = 0.001; Fig. 5A), PA_{0.5mV} ($\rho = -0.616$, P = 0.0003; Fig.
- 329 5C), and AP_{0.5mV} (ρ = -0.551, *P* = 0.002; Fig. 5D) responses following M1 iTBS,
- 330 respectively.

331

332 **Discussion**

333	In the present study, we investigated the influence of PMd on the plasticity of early and late I-
334	wave-generating circuits in M1 of young and older adults. This was achieved by applying
335	PMd iTBS as a priming intervention to modify the neuroplastic response of M1 to subsequent
336	iTBS (PMd iTBS-M1 iTBS, PMd sham-M1 iTBS). We measured changes in corticospinal
337	excitability (PA1mV, AP1mV, PA0.5mV, AP0.5mV) and I-wave recruitment (PA-LM latency, AP-
338	LM latency) following the intervention. The findings show that PMd iTBS specifically
339	modulated the excitability of the early I-wave circuits in both young and older adults.
340	Moreover, PMd iTBS disrupted the neuroplastic response of the early I-wave circuits to M1
341	iTBS in both young and older adults, whereas the neuroplastic response of the late I-wave
342	circuits was unaffected in both age groups.

343 PMd influence on corticospinal excitability in young and older adults

Previous work has reported that application of iTBS to PMd facilitates PA_{1mV} measures of 344 345 M1 corticospinal excitability in young adults by ~30%, which is thought to stem from the induction of LTP-like effects within PMd, resulting in increased excitability within M1 346 (Meng *et al.*, 2020). Furthermore, we have demonstrated previously that this effect on PA_{ImV} 347 is preserved with ageing and extends to AP_{ImV} measures of corticospinal excitability (Liao et 348 al., 2023). The absence of any changes in PA_{1mV} or AP_{1mV} within the present study is 349 therefore inconsistent with these previous findings. However, inter- and intraindividual 350 variability in the changes in M1 excitability following TBS is well-documented (Hamada et 351 al., 2013; Corp et al., 2020; Guerra et al., 2020). In particular, there is some variability in the 352 time course of facilitation following PMd iTBS. For example, one study reported that the 353 facilitation of MEP amplitude only occurred at 15 minutes (Meng et al., 2020), whereas we 354 previously demonstrated facilitation of MEP amplitude that persisted from 5 to 40 minutes 355

following PMd iTBS (Liao *et al.*, 2023). Consequently, our decision to record MEPs at 5
minutes post-PMd iTBS may have limited the ability to detect changes in corticospinal
excitability due to the priming intervention.

Although PA_{1mV} and AP_{1mV} MEP amplitude were not modulated following PMd iTBS, 359 360 $PA_{0.5mV}$ was facilitated (by ~30%) for both young and older adults. The conventional interpretation of how TMS intensity and current direction influence I-wave recruitment 361 suggests that low-intensity PA TMS preferentially recruits early I-waves, whereas low-362 intensity AP TMS preferentially recruits late I-waves (Hamada et al., 2013), with either 363 current direction able to recruit both I-waves as the stimulation intensity is increased (Di 364 Lazzaro et al., 2001; Di Lazzaro et al., 2003). We therefore applied single-pulse TMS at 365 relatively lower intensities compared to MEP_{1mV} (PA_{0.5mV}, AP_{0.5mV}), where PA_{0.5mV} is likely 366 more selective for activation of the early I-waves, while $AP_{0.5mV}$ is likely more selective for 367 the late I-waves (Opie *et al.*, 2022). Given that we previously reported potentiation of both 368 369 PA_{0.5mV} and AP_{0.5mV} (by ~50-100%) following PMd iTBS (Liao et al., 2023), the increase in 370 PA_{0.5mV} within the present study suggests that the effect of PMd iTBS on early I-wave excitability may be immediate and more consistent. Importantly, previous work has shown 371 that PMd iTBS applied as it was in the current study is unlikely to have activated M1 directly. 372 Specifically, Huang and colleagues (2009) assessed the intensity required to activate M1 373 when TMS was applied over PMd, and showed that 80% of this (matching the level applied 374 during iTBS) applied to M1 does not influence M1 excitability (Huang et al., 2009). Given 375 that we located PMd using similar methods (Huang et al., 2009; Huang et al., 2018; Meng et 376 377 al., 2020), it is therefore unlikely that PMd iTBS activated M1 directly in the present study. Despite the present findings demonstrating that PMd iTBS increased early I-wave 378

379 excitability, this effect was not different between young and older adults, suggesting that the

influence of PMd on early I-wave excitability may be preserved with ageing. This contrasts 380 with our previous work, which specifically demonstrated weakened direct PMd modulation 381 of early I-waves in older adults (Liao et al., 2023). Given that both studies employed the 382 same methods to assess changes in M1 excitability following PMd iTBS, participant factors 383 such as genetics, pharmacology, aerobic exercise, and diet that are known to influence 384 cortical plasticity (Ridding & Ziemann, 2010; Phillips, 2017) may have confounded the 385 386 present findings. As the contributions of participant characteristics on PMd-M1 communication were not examined in the present study, and the small sample sizes were not 387 388 powered for such subanalyses, it will be important to characterise their involvement in future studies. 389

PMd influence on M1 plasticity in young and older adults

Previous work in young participants demonstrated that applying continuous TBS (cTBS) to 391 PMd disrupts the neuroplastic response of M1 to both iTBS and cTBS, assessed using PA_{ImV} 392 MEPs (Huang et al., 2018). This demonstrated that LTP- and LTD-like effects within M1 can 393 be modulated by PMd cTBS, which was thought to arise from heterosynaptic metaplastic 394 395 effects, where the modulation of local synaptic plasticity within PMd affected subsequent changes in remote synapses (that were not initially activated) within M1 (Huang et al., 2018). 396 In the present study, we demonstrated that applying iTBS to PMd also disrupts the LTP-like 397 398 effects of M1 iTBS for AP_{1mV} measures of corticospinal excitability. However, given that iTBS produces LTP-like effects while cTBS produces LTD-like effects, this disruption of 399 AP_{1mV} facilitation may stem from a different mechanism more consistent with homeostatic 400 401 metaplasticity (Müller et al., 2007; Todd et al., 2009; Murakami et al., 2012). Importantly, 402 this response did not differ between young and older adults, suggesting that the influence of PMd on the plasticity of AP circuits within M1 is maintained with age. 403

Furthermore, PMd iTBS disrupted the effects of M1 iTBS on PA_{0.5mV} (early), but not AP_{0.5mV} 404 (late) circuits. This suggests that the influence of PMd on M1 plasticity is specific to the early 405 I-waves. Although this may appear counterintuitive to our AP_{ImV} findings, this can be 406 explained by the possibility that the higher stimulus intensity required to record AP_{1mV} 407 resulted in mixed recruitment of early and late I-waves, but that changes in AP_{ImV} were 408 driven specifically by the early I-waves (Di Lazzaro et al., 2001; Di Lazzaro et al., 2003; 409 410 Liao et al., 2022). This is complemented by the correlation analysis results demonstrating that larger facilitation of PA_{0.5mV} post-PMd iTBS is correlated with smaller facilitation of PA_{0.5mV} 411 412 post-M1 iTBS, suggesting that this homeostatic metaplastic effect is likely related to the early I-wave circuits. While a similar correlation was also shown for PA_{1mV} and AP_{0.5mV}, PMd 413 iTBS-M1 iTBS did not disrupt the potentiation of these measures when compared to PMd 414 sham-M1 iTBS session. It is possible that the higher stimulus intensities required for PA_{1mV} 415 and AP_{0.5mV} (relative to PA_{0.5mV}) may have also resulted in mixed recruitment of early and 416 late I-waves (Liao et al., 2022). In particular, given that there is growing evidence to suggest 417 that PA and AP TMS can activate distinct populations of early and late I-waves (i.e., PA- and 418 AP-sensitive early and late I-waves) (Spampinato et al., 2020; Opie & Semmler, 2021), 419 PA_{lmV} and $AP_{0.5mV}$ may have recruited other I-wave circuits that were less sensitive to the 420 modulatory effects of iTBS. However, this will need to be clarified in future research using 421 techniques that are more selective to these different I-waves, such as modifying the TMS 422 423 pulse width (Hannah & Rothwell, 2017). Despite this, we provide new evidence that PMd iTBS specifically modulates M1 plasticity of early I-wave circuits recruited by AP 424 stimulation. 425

While M1 iTBS in isolation (PMd sham-M1 iTBS) potentiated PA_{0.5mV} responses (compared
with PMd iTBS-M1 iTBS) in both age groups, the timing of this response varied between
groups. Whereas differences between sham and real PMd iTBS sessions were immediate for

older adults, they were only apparent after 30 minutes in young adults. Given that M1 iTBS 429 has not been shown to differentially modulate corticospinal excitability in young and older 430 adults (Di Lazzaro et al., 2008; Young-Bernier et al., 2014; Dickins et al., 2015; Opie et al., 431 2017), this outcome seems unlikely to reflect effects of age within M1. An alternative 432 explanation could be that the modulatory effects of PMd iTBS differed between groups, with 433 younger adults having a stronger response that was more resistant to the subsequent effects of 434 435 M1 iTBS. This is supported by the amplitude of PA_{0.5mV} being reduced 5 minutes after M1 iTBS in older, but not young adults in the session involving real PMd iTBS (Fig. 4A). 436 437 Although speculative, this outcome would be consistent with our previous finding that the influence of PMd iTBS on PA_{0.5mV} is reduced in older adults (Liao et al., 2023). However, 438 this speculation will require additional studies that more effectively characterise the time 439 440 course of facilitation in young and older adults. For example, previous work investigating the effects of PMd cTBS on M1 neuroplastic response to iTBS or cTBS monitored changes in 441 corticospinal excitability for two hours following PMd cTBS (during which excitability 442 returned to baseline levels) before applying subsequent M1 iTBS or cTBS (Huang *et al.*, 443 2018). 444

445 PMd and M1 influence on I-wave recruitment in young and older adults

446 The ability to recruit both early and late I-waves can be investigated by comparing the

447 latencies evoked by PA and AP TMS to the latencies of direct corticospinal activation (PA-

LM, early; AP-LM, late) (Hamada *et al.*, 2013). The prototypical values for these measures

449 reveal shorter PA-LM latencies (~1.5 ms) compared to AP-LM latencies (~3 ms), providing

- 450 an index of early and late I-wave recruitment, respectively (Hamada *et al.*, 2013).
- 451 Importantly, previous studies have shown that the ability to recruit late I-waves with AP TMS
- 452 predicts the neuroplastic response of M1 to iTBS (Hamada *et al.*, 2013; Volz *et al.*, 2019),

with AP inputs thought to originate from PMd (Volz et al., 2015; Aberra et al., 2020). It has 453 also been demonstrated that AP-LM latencies can be shortened using M1 iTBS, which was 454 suggested to reflect the direct modulation of the late I-wave circuitry (Volz et al., 2019). 455 Although we also assessed changes in PA-LM and AP-LM latencies following PMd sham-456 M1 iTBS in the present study, the intervention failed to modulate the I-wave latencies. It is 457 possible that changes in AP-LM latencies occur immediately following iTBS, as the MEP 458 459 latency measures were recorded at least 45 minutes either side of PMd and M1 iTBS in order to avoid complications involving the effects of muscle activation on neuroplasticity responses 460 461 (Huang et al., 2008; Thirugnanasambandam et al., 2011; Goldsworthy et al., 2015). Consequently, the effects of M1 iTBS on I-wave latencies will have to be clarified in future 462 studies. 463 Importantly, baseline I-wave recruitment was not correlated with changes in corticospinal 464 excitability following M1 iTBS in isolation, in contrast to previous findings (Hamada et al., 465 466 2013; Volz et al., 2019). While the difference between the present study and previous studies is that we included older participants, correlation analyses did not reveal any relationship 467 between age and changes in corticospinal excitability following M1 iTBS. The variability in 468 the present findings may therefore involve contributions from other factors. For example, 469 recent work assessing variability of M1 iTBS has suggested that the ability of iTBS to engage 470 neural oscillations in the β range (13-30 Hz) may be an important predictor of the 471 neuroplastic response to iTBS (Leodori et al., 2021). Enhancing premotor-M1 472 communication using cortico-cortical paired associated stimulation (ccPAS) has been 473 474 recently shown to improve the synchronisation of neural oscillations (which is thought to mediate neuronal communication and plasticity) in the β range (Trajkovic *et al.*, 2023). 475 476 Further investigation involving these measures may therefore better characterise the

477 variability of iTBS, and may also have applications in understanding PMd-M1

478 communication.

479	In conclusion, the application of iTBS over PMd potentiated corticospinal excitability and
480	disrupted the effects of subsequent M1 iTBS. Specifically, our results show that PMd may
481	more consistently influence the excitability of early I-waves in young and older adults.
482	Importantly, we provide new evidence that PMd disrupts M1 plasticity of early I-wave
483	circuits in both age groups. It will therefore be useful in future studies to investigate how
484	PMd modulation of M1 plasticity influences different feature of motor skill learning in young
485	and older adults.

486 Data Availability Statement

487 Data from this study will be made available to qualified investigators upon reasonable request488 to the corresponding author.

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497 **References**

498 499 500	Aberra AS, Wang B, Grill WM & Peterchev AV. (2020). Simulation of transcranial magnetic stimulation in head model with morphologically-realistic cortical neurons. <i>Brain Stimul</i> 13 , 175-189.
501 502 503	Barr DJ, Levy R, Scheepers C & Tily HJ. (2013). Random effects structure for confirmatory hypothesis testing: Keep it maximal. <i>J Mem Lang</i> 68, 255-278.
504 505 506	Buonomano DV & Merzenich MM. (1998). Cortical plasticity: from synapses to maps. <i>Annu Rev</i> Neurosci 21, 149-186.
507 508 509 510	Casarotto A, Dolfini E, Cardellicchio P, Fadiga L, D'Ausilio A & Koch G. (2023). Mechanisms of Hebbian-like plasticity in the ventral premotor – primary motor network. <i>J Physiol</i> 601, 211- 226.
511 512 513	Chouinard PA, Leonard G & Paus T. (2005). Role of the primary motor and dorsal premotor cortices in the anticipation of forces during object lifting. <i>J Neurosci</i> 25 , 2277-2284.
514 515 516 517 518	Corp DT, Bereznicki HGK, Clark GM, Youssef GJ, Fried PJ, Jannati A, Davies CB, Gomes-Osman J, Stamm J, Chung SW, Bowe SJ, Rogasch NC, Fitzgerald PB, Koch G, Di Lazzaro V, Pascual- Leone A & Enticott PG. (2020). Large-scale analysis of interindividual variability in theta-burst stimulation data: Results from the 'Big TMS Data Collaboration'. <i>Brain Stimul</i> 13 , 1476-1488.
519 520 521 522	Di Lazzaro V, Oliviero A, Pilato F, Mazzone P, Insola A, Ranieri F & Tonali PA. (2003). Corticospinal volleys evoked by transcranial stimulation of the brain in conscious humans. <i>Neurol Res</i> 25, 143-150.
523 524 525 526	Di Lazzaro V, Oliviero A, Saturno E, Pilato F, Insola A, Mazzone P, Profice P, Tonali P & Rothwell J. (2001). The effect on corticospinal volleys of reversing the direction of current induced in the motor cortex by transcranial magnetic stimulation. <i>Exp Brain Res</i> 138 , 268-273.
527 528 529 530	Di Lazzaro V, Pilato F, Dileone M, Profice P, Oliviero A, Mazzone P, Insola A, Ranieri F, Meglio M, Tonali PA & Rothwell JC. (2008). The physiological basis of the effects of intermittent theta burst stimulation of the human motor cortex. <i>J Physiol</i> 586, 3871-3879.
531 532 533	Di Lazzaro V, Profice P, Ranieri F, Capone F, Dileone M, Oliviero A & Pilato F. (2012). I-wave origin and modulation. <i>Brain Stimul 5,</i> 512-525.
534 535 536 537	Di Lazzaro V, Restuccia D, Oliviero A, Profice P, Ferrara L, Insola A, Mazzone P, Tonali P & Rothwell JC. (1998). Effects of voluntary contraction on descending volleys evoked by transcranial stimulation in conscious humans. <i>J Physiol</i> 508 (Pt 2), 625-633.
538	

539 540	Dickins DS, Sale MV & Kamke MR. (2015). Plasticity Induced by Intermittent Theta Burst Stimulation in Bilateral Motor Cortices Is Not Altered in Older Adults. <i>Neural plasticity</i> 2015 , 323409.
541 542 543 544	Fathi D, Ueki Y, Mima T, Koganemaru S, Nagamine T, Tawfik A & Fukuyama H. (2010). Effects of aging on the human motor cortical plasticity studied by paired associative stimulation. <i>Clin Neurophysiol</i> 121 , 90-93.
545 546 547 548	Freitas C, Perez J, Knobel M, Tormos JM, Oberman L, Eldaief M, Bashir S, Vernet M, Peña-Gómez C & Pascual-Leone A. (2011). Changes in cortical plasticity across the lifespan. <i>Front Aging Neurosci</i> 3 , 5-5.
549 550 551 552	Goldsworthy MR, Müller-Dahlhaus F, Ridding MC & Ziemann U. (2015). Resistant Against De- depression: LTD-Like Plasticity in the Human Motor Cortex Induced by Spaced cTBS. <i>Cereb</i> <i>Cortex</i> 25, 1724-1734.
553 554 555 556 557	Groppa S, Schlaak BH, Münchau A, Werner-Petroll N, Dünnweber J, Bäumer T, van Nuenen BFL & Siebner HR. (2012). The human dorsal premotor cortex facilitates the excitability of ipsilateral primary motor cortex via a short latency cortico-cortical route. <i>Hum Brain Mapp</i> 33 , 419-430.
558 559 560	Guerra A, López-Alonso V, Cheeran B & Suppa A. (2020). Variability in non-invasive brain stimulation studies: Reasons and results. <i>Neurosci Lett</i> 719 , 133330.
561 562 563 564	Hamada M, Galea JM, Di Lazzaro V, Mazzone P, Ziemann U & Rothwell JC. (2014). Two distinct interneuron circuits in human motor cortex are linked to different subsets of physiological and behavioral plasticity. <i>J Neurosci</i> 34, 12837-12849.
565 566 567	Hamada M, Murase N, Hasan A, Balaratnam M & Rothwell JC. (2013). The Role of Interneuron Networks in Driving Human Motor Cortical Plasticity. <i>Cereb Cortex</i> 23, 1593-1605.
568 569 570 571	Hannah R & Rothwell JC. (2017). Pulse Duration as Well as Current Direction Determines the Specificity of Transcranial Magnetic Stimulation of Motor Cortex during Contraction. <i>Brain Stimul</i> 10, 106-115.
572 573 574	Huang Y-Z, Chen R-S, Fong P-Y, Rothwell JC, Chuang W-L, Weng Y-H, Lin W-Y & Lu C-S. (2018). Inter- cortical modulation from premotor to motor plasticity. <i>J Physiol</i> 596, 4207-4217.
575 576 577	Huang Y-Z, Edwards MJ, Rounis E, Bhatia KP & Rothwell JC. (2005). Theta Burst Stimulation of the Human Motor Cortex. <i>Neuron</i> 45, 201-206.
578 579 580	Huang Y-Z, Rothwell JC, Edwards MJ & Chen R-S. (2008). Effect of Physiological Activity on an NMDA- Dependent Form of Cortical Plasticity in Human. <i>Cereb Cortex</i> 18, 563-570.
195	

582 583 584	Huang Y-Z, Rothwell JC, Lu C-S, Wang J, Weng Y-H, Lai S-C, Chuang W-L, Hung J & Chen R-S. (2009). The effect of continuous theta burst stimulation over premotor cortex on circuits in primary motor cortex and spinal cord. <i>Clin Neurophysiol</i> 120 , 796-801.
585 586 587 588	Koch G, Franca M, Mochizuki H, Marconi B, Caltagirone C & Rothwell JC. (2007). Interactions between pairs of transcranial magnetic stimuli over the human left dorsal premotor cortex differ from those seen in primary motor cortex. <i>J Physiol</i> 578, 551-562.
589 590 591 592	Leodori G, Fabbrini A, De Bartolo MI, Costanzo M, Asci F, Palma V, Belvisi D, Conte A & Berardelli A. (2021). Cortical mechanisms underlying variability in intermittent theta-burst stimulation- induced plasticity: A TMS-EEG study. <i>Clin Neurophysiol</i> 132, 2519-2531.
593 594 595 596	Liao W-Y, Opie GM, Ziemann U & Semmler JG. (2023). Modulation of dorsal premotor cortex differentially influences I-wave excitability in primary motor cortex of young and older adults. <i>J Physiol</i> .
597 598 599	Liao W-Y, Sasaki R, Semmler JG & Opie GM. (2022). Cerebellar transcranial direct current stimulation disrupts neuroplasticity of intracortical motor circuits. <i>PLoS One</i> 17, e0271311.
600 601 602	Lo S & Andrews S. (2015). To transform or not to transform: using generalized linear mixed models to analyse reaction time data. <i>Front Psychol</i> 6, 1171-1171.
603 604 605 606	Meng H-J, Cao N, Zhang J & Pi Y-L. (2020). Intermittent theta burst stimulation facilitates functional connectivity from the dorsal premotor cortex to primary motor cortex. <i>PeerJ</i> 8, e9253-e9253.
607 608 609 610	Müller-Dahlhaus JFM, Orekhov Y, Liu Y & Ziemann U. (2008). Interindividual variability and age- dependency of motor cortical plasticity induced by paired associative stimulation. <i>Exp Brain</i> <i>Res</i> 187, 467-475.
611 612 613 614	Müller JF, Orekhov Y, Liu Y & Ziemann U. (2007). Homeostatic plasticity in human motor cortex demonstrated by two consecutive sessions of paired associative stimulation. <i>The European journal of neuroscience</i> 25, 3461-3468.
615 616 617 618	Münchau A, Bloem BR, Irlbacher K, Trimble MR & Rothwell JC. (2002). Functional connectivity of human premotor and motor cortex explored with repetitive transcranial magnetic stimulation. <i>J Neurosci</i> 22, 554-561.
619 620 621 622	Murakami T, Müller-Dahlhaus F, Lu MK & Ziemann U. (2012). Homeostatic metaplasticity of corticospinal excitatory and intracortical inhibitory neural circuits in human motor cortex. <i>J Physiol</i> 590, 5765-5781.
623 624 625 626	Ni Z, Charab S, Gunraj C, Nelson AJ, Udupa K, Yeh IJ & Chen R. (2010). Transcranial Magnetic Stimulation in Different Current Directions Activates Separate Cortical Circuits. <i>J</i> <i>Neurophysiol</i> 105, 749-756.

627 628 629	Ni Z, Isayama R, Castillo G, Gunraj C, Saha U & Chen R. (2015). Reduced dorsal premotor cortex and primary motor cortex connectivity in older adults. <i>Neurobiol Aging</i> 36, 301-303.
630 631 632 633	Nowak DA, Berner J, Herrnberger B, Kammer T, Grön G & Schönfeldt-Lecuona C. (2009). Continuous theta-burst stimulation over the dorsal premotor cortex interferes with associative learning during object lifting. <i>Cortex</i> 45, 473-482.
634 635 636	Opie GM, Cirillo J & Semmler JG. (2018). Age-related changes in late I-waves influence motor cortex plasticity induction in older adults. <i>J Physiol</i> 596, 2597-2609.
637 638 639 640	Opie GM, Hand BJ & Semmler JG. (2020). Age-related changes in late synaptic inputs to corticospinal neurons and their functional significance: A paired-pulse TMS study. <i>Brain Stimul</i> 13, 239-246.
641 642 643	Opie GM, Liao W-Y & Semmler JG. (2022). Interactions Between Cerebellum and the Intracortical Excitatory Circuits of Motor Cortex: a Mini-Review. <i>Cerebellum</i> 21 , 159-166.
644 645 646 647	Opie GM & Semmler JG. (2021). Preferential Activation of Unique Motor Cortical Networks With Transcranial Magnetic Stimulation: A Review of the Physiological, Functional, and Clinical Evidence. <i>Neuromodulation</i> 24, 813-828.
648 649 650	Opie GM, Vosnakis E, Ridding MC, Ziemann U & Semmler JG. (2017). Priming theta burst stimulation enhances motor cortex plasticity in young but not old adults. <i>Brain Stimul</i> 10, 298-304.
651 652 653	Parikh PJ & Santello M. (2017). Role of human premotor dorsal region in learning a conditional visuomotor task. <i>J Neurophysiol</i> 117, 445-456.
654 655 656 657	Phillips C. (2017). Lifestyle Modulators of Neuroplasticity: How Physical Activity, Mental Engagement, and Diet Promote Cognitive Health during Aging. <i>Neural plasticity</i> 2017, 3589271.
658 659 660	Puri R & Hinder MR. (2022). Response bias reveals the role of interhemispheric inhibitory networks in movement preparation and execution. <i>Neuropsychologia</i> 165 , 108120.
661 662 663	Ridding MC & Ziemann U. (2010). Determinants of the induction of cortical plasticity by non-invasive brain stimulation in healthy subjects. <i>J Physiol</i> 588, 2291-2304.
664 665 666	Rossi S, Hallett M, Rossini PM & Pascual-Leone A. (2011). Screening questionnaire before TMS: An update. <i>Clin Neurophysiol</i> 122, 1686.
667 668 669 670	Rossini PM, Burke D, Chen R, Cohen LG, Daskalakis Z, Di Iorio R, Di Lazzaro V, Ferreri F, Fitzgerald PB, George MS, Hallett M, Lefaucheur JP, Langguth B, Matsumoto H, Miniussi C, Nitsche MA, Pascual-Leone A, Paulus W, Rossi S, Rothwell JC, Siebner HR, Ugawa Y, Walsh V & Ziemann

671 672 673 674	U. (2015). Non-invasive electrical and magnetic stimulation of the brain, spinal cord, roots and peripheral nerves: Basic principles and procedures for routine clinical and research application. An updated report from an I.F.C.N. Committee. <i>Clin Neurophysiol</i> 126 , 1071-1107.
675 676 677 678	Sakai K, Ugawa Y, Terao Y, Hanajima R, Furubayashi T & Kanazawa I. (1997). Preferential activation of different I waves by transcranial magnetic stimulation with a figure-of-eight-shaped coil. <i>Exp Brain Res</i> 113 , 24-32.
679 680 681	Sale MV, Ridding MC & Nordstrom MA. (2008). Cortisol inhibits neuroplasticity induction in human motor cortex. <i>J Neurosci</i> 28, 8285-8293.
682 683 684	Sanes JN & Donoghue JP. (2000). Plasticity and primary motor cortex. <i>Annu Rev Neurosci</i> 23, 393- 415.
685 686 687 688	Seidler RD, Bernard JA, Burutolu TB, Fling BW, Gordon MT, Gwin JT, Kwak Y & Lipps DB. (2010). Motor control and aging: links to age-related brain structural, functional, and biochemical effects. <i>Neurosci Biobehav Rev</i> 34 , 721-733.
689 690 691	Spampinato DA, Celnik PA & Rothwell JC. (2020). Cerebellar-Motor Cortex Connectivity: One or Two Different Networks? <i>J Neurosci</i> 40 , 4230-4239.
692 693 694 695	Suppa A, Bologna M, Gilio F, Lorenzano C, Rothwell JC & Berardelli A. (2008). Preconditioning Repetitive Transcranial Magnetic Stimulation of Premotor Cortex Can Reduce But Not Enhance Short-Term Facilitation of Primary Motor Cortex. <i>J Neurophysiol</i> 99, 564-570.
696 697 698 699 700	Thirugnanasambandam N, Sparing R, Dafotakis M, Meister IG, Paulus W, Nitsche MA & Fink GR. (2011). Isometric contraction interferes with transcranial direct current stimulation (tDCS) induced plasticity – evidence of state-dependent neuromodulation in human motor cortex. <i>Restor Neurol Neurosci</i> 29 , 311-320.
701 702 703	Todd G, Flavel SC & Ridding MC. (2009). Priming theta-burst repetitive transcranial magnetic stimulation with low- and high-frequency stimulation. <i>Exp Brain Res</i> 195 , 307-315.
704 705 706	Todd G, Kimber TE, Ridding MC & Semmler JG. (2010). Reduced motor cortex plasticity following inhibitory rTMS in older adults. <i>Clin Neurophysiol</i> 121, 441-447.
707 708 709 710	Trajkovic J, Romei V, Rushworth MFS & Sel A. (2023). Strengthening connectivity between premotor and motor cortex increases inter-areal communication in the human brain. <i>bioRxiv</i> , 2023.2002.2015.528606.
711 712 713 714	Volz LJ, Hamada M, Michely J, Pool E-M, Nettekoven C, Rothwell JC & Grefkes Hermann C. (2019). Modulation of I-wave generating pathways by theta-burst stimulation: a model of plasticity induction. J Physiol 597 , 5963-5971.

715 716 717	Volz LJ, Hamada M, Rothwell JC & Grefkes C. (2015). What Makes the Muscle Twitch: Motor System Connectivity and TMS-Induced Activity. <i>Cereb Cortex</i> 25 , 2346-2353.
718 719 720	Wiethoff S, Hamada M & Rothwell JC. (2014). Variability in Response to Transcranial Direct Current Stimulation of the Motor Cortex. <i>Brain Stimul</i> 7, 468-475.
721 722 723 724	Young-Bernier M, Tanguay AN, Davidson PS & Tremblay F. (2014). Short-latency afferent inhibition is a poor predictor of individual susceptibility to rTMS-induced plasticity in the motor cortex of young and older adults. <i>Front Aging Neurosci</i> 6 , 182.
725 726 727	Ziemann U. (2020). I-waves in motor cortex revisited. <i>Exp Brain Res</i> 238, 1601-1610.
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Figure Legends

Figure 1. (A) Subject sample and experimental setup. (B) Experimental procedure. PA, posterior-to-anterior; AP, anterior-to-posterior; LM, lateral-to-medial; RMT, resting motor threshold; AMT, active motor threshold; MEP_{*ImV*}, standard MEP of ~ 1mV at baseline; MEP_{0.5mV}, MEP of ~ 0.5 mV at baseline; PMd, dorsal premotor cortex; iTBS, intermittent theta burst stimulation.

Figure 2. Changes in PA_{1mV} (A, blue), AP_{1mV} (B, orange), $PA_{0.5mV}$ (C, blue), and $AP_{0.5mV}$ (D, orange) measures of corticospinal excitability following PMd iTBS (darker hue) and sham (lighter hue) stimulation in all participants. Data show EMM (95% CI) with individual subject means. **P* < 0.05.

Figure 3. Changes in PA_{ImV} (A, blue) and AP_{ImV} (B, orange) measures of corticospinal excitability following PMd iTBS-M1 iTBS (darker hue) and PMd sham-M1 iTBS (lighter hue) in young (no stripes) and older adults (stripes) at 5 and 30 minutes. Data show EMM (95% CI) with individual subject means. **P* < 0.05. #*P* < 0.05 compared to 5 minutes in same session.

Figure 4. Changes in PA_{0.5mV} (A, blue) and AP_{0.5mV} (B, orange) measures of corticospinal excitability following PMd iTBS-M1 iTBS (darker hue) and PMd sham-M1 iTBS (lighter hue) in young (no stripes) and older adults (stripes) at 5 and 30 minutes. (C) Changes in AP_{0.5mV} following M1 iTBS in all participants (dark orange) at 5 and 30 minutes. Data show EMM (95% CI) with individual subject means. *P < 0.05.

Figure 5. Correlation of ranked changes in post-PMd iTBS measures of corticospinal excitability (PA_{1mV}, A; AP_{1mV}, B; PA_{0.5mV}, C; AP_{0.5mV}, D) with ranked changes in post-M1 iTBS measures of corticospinal excitability.

Table 1. Baseline TMS intensities between sessions for young and older adults.					
Measure	Young		Older		
	PMd iTBS-M1 iTBS	PMd sham-M1 iTBS	PMd iTBS-M1 iTBS	PMd sham-M1 iTBS	
PA					
RMT _{PA} (% MSO)	47.3 [42.3, 52.3]	47.8 [42.8, 52.8]	50.4 [45.4, 55.4]	51.3 [46.3, 56.3]	
AMT _{PA} (% MSO)	39.9 [36.4, 43.4]	39.3 [35.8, 42.8]	42.6 [39.1, 46.1]	43.2 [39.7, 46.7]	
1mV _{PA} (% MSO)	56.5 [49.9, 63.1]	57.4 [50.8, 64.0]	65.4 [58.5, 72.2]	63.3 [56.4, 70.1]	
0.5mV _{PA} (% MSO)	53.0 [46.1, 59.9]	53.8 [46.9, 60.7]	61.5 [54.5, 68.4]	61.3 [54.3, 68.2]	
AP					
RMT _{AP} (% MSO)	61.3 [55.6, 67.0]	62.3 [56.6, 68.0]	66.3 [60.4, 72.2]	65.3 [59.4, 71.2]	
AMT _{AP} (% MSO)	54.2 [49.0, 59.4]	54.1 [49.0, 59.3]	59.1 [54.0, 64.3]	57.2 [52.0, 62.4]	
1mV _{AP} (% MSO)	73.9 [65.6, 82.1]	73.8 [65.6, 82.0] ^a	83.8 [74.6, 93.1]	79.3 [70.0, 88.5] ^a	
0.5mV _{AP} (% MSO)	70.9 [63.5, 78.3]	71.3 [63.9, 78.7]	79.4 [71.4, 89.4]	76.4 [68.4, 84.4]	
LM					
AMT _{LM} (% MSO)	45.3 [40.5, 50.2]	45.3 [40.4, 50.1]	49.9 [45.1, 54.8]	48.8 [43.9, 53.7]	
TBS					
RMT _{Rapid} (% MSO)	55.7 [50.8, 60.6]	57.8 [52.9, 62.7]	57.7 [52.8, 62.6]	58.1 [53.2, 63.0]	
	1 1 30 .005	1, 'TDC '			

 Table 1. Baseline TMS intensities between sessions for young and older adults.

Data show EMM [95% CI; lower, upper]. ${}^{a}P < 0.05$ compared to iTBS session.

Table 2. Baseline responses of corticospinal excitability and I-wave recruitment between sessions.						
Measure	Young		Older			
	PMd iTBS-M1 iTBS	PMd sham-M1 iTBS	PMd iTBS-M1 iTBS	PMd sham-M1 iTBS		
PA						
PA-LM latency (ms)	1.39 [0.89, 1.88]	1.54 [1.04, 2.03]	1.97 [1.47, 2.46]	2.02 [1.52, 2.51]		
$1 m V_{PA} (m V)$	1.03 [0.94, 1.12]	0.93 [0.85, 1.01]	0.89 [0.81, 0.97] ^a	0.96 [0.87, 1.04]		
$0.5 m V_{PA} (mV)$	0.53 [0.46, 0.60]	0.49 [0.42, 0.56]	0.50 [0.43, 0.57]	0.51 [0.44, 0.58]		
AP						
AP-LM latency (ms)	3.49 [3.00, 3.98] ^b	3.54 [3.05, 4.03] ^b	3.69 [3.20, 4.18] ^b	3.99 [3.50, 4.48] ^b		
$1 m V_{AP} (m V)$	0.97 [0.87, 1.06]	0.88 [0.79, 0.97]	1.02 [0.91, 1.14]	0.99 [0.88, 1.10]		
$0.5 m V_{AP} (m V)$	0.47 [0.41, 0.53]	0.44 [0.39, 0.50]	0.45 [0.40, 0.51]	0.45 [0.39, 0.50]		

Data show EMM [95% CI; lower, upper].^aP < 0.05 compared to young. ^bP < 0.05 compared to PA of same measure.

Table 3. Comparison of VAS responses (mean ± STD) between sessions.					
Question	PMd iTBS-M1 iTBS	PMd sham-M1 iTBS			
How uncomfortable were the TMS pulses (0,					
not uncomfortable at all; 10, highly	2.67 ± 2.60	2.5 ± 2.79			
uncomfortable)?					
If there were any twitches in the right hand,					
how strong were they (0, no twitches; 10, very	0.63 ± 1.40	0.60 ± 1.13			
strong cramp)?					
How localised were the sensations from TMS	2.03 ± 2.47	$0.50 \pm 1.04*$			
pulses (0, highly localised; 10, widespread)?	2.05 ± 2.47	0.50 ± 1.04			

Data show mean \pm standard deviation.**P* < 0.0167 compared to iTBS.



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Figure 5. Correlation of ranked changes in post-PMd iTBS measures of corticospinal excitability (PA_{1mV}, A; AP_{1mV}, B; PA_{0.5mV}, C; AP_{0.5mV}, D) with ranked changes in post-M1 iTBS measures of corticospinal excitability.