

## 1 Title

2 Helminth ecological requirements shape the impact of climate change on the hazard of infection

## 3 Running title

4 Contrasting responses of helminths to climate

5

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13

## 14 Authorship statement

15 CV performed the literature analysis, modeling work, result visualization and wrote the first draft of the  
16 manuscript. LM, RC and MG guided the modeling development. IMC developed the original idea,  
17 advised on the analysis, results interpretation and contributed to the first draft writing. All authors  
18 contributed substantially to manuscript drafting and editing.

19

## 20 Data accessibility statement

21 The data that support the findings are fully available in Figshare repository (DOI:  
22 10.6084/m9.figshare.24065613).

## 23 Keywords

24 Trichostrongylidae; Free-living stages; Hatching; Development and mortality; Seasonality; Spatial  
25 distribution; Co-circulation

## 26 Abstract

27 Outbreaks and spread of infectious diseases are often associated with seasonality and changes caused  
28 by global warming. Free-living stages of soil-transmitted helminths are highly susceptible to  
29 environmental drivers, however, how multiple climatic variables affect helminth species, and the long-  
30 term consequences of these interactions, is poorly understood. We used experiments on nine  
31 trichostrongylid species to develop a temperature- and humidity-dependent model of infection hazard,  
32 which was then implemented at the European scale under climate change scenarios. Intestinal and  
33 stomach helminths exhibited contrasting climatic responses, with the former group strongly affected by  
34 temperature while the latter primarily impacted by humidity. These differences generated seasonal  
35 changes in the timing and intensity of the infection hazard and spatial heterogeneities within and  
36 between the two groups. A future range expansion of both groups toward northern latitudes is  
37 expected to create new opportunities for the co-circulation of the studied helminth species.

## 38 Introduction

39 The many forms of disruption associated with climate change, like warming temperature, extreme  
40 climatic events or shifts in climatic ranges, are expected to strongly affect ectotherm species by

41 selectively targeting components of their life cycle and dynamics (Deutsch et al., 2008; Paaijmans et al.,  
42 2013; Wagner et al., 2023). These climatic changes are also predicted to alter the severity and spread of  
43 many circulating infections, whose transmission depends on the survival and development of stages that  
44 live free in the environment, as is the case of soil-transmitted helminths with direct life cycle (Hoar et al.,  
45 2012; Molnár et al., 2013b, 2017; Rose et al., 2016; Smith, 1990), as well as of those parasites that  
46 require intermediate invertebrate hosts or vectors for maturation and transmission (Carraro et al., 2017;  
47 Kutz et al., 2002, 2005a; Molnár et al., 2013a; Schjetlein and Skorpning, 1995). Assessing the net effect of  
48 climate warming on parasite transmission has proved to be challenging because of the non-linear  
49 thermal responses that parasite traits often exhibit (Gehman et al., 2018; Molnár et al., 2017). Indeed,  
50 the trade-off of faster development but lower survival commonly observed for parasites (Hoar et al.,  
51 2012; Rose et al., 2016; Smith, 1990) and vectors (Mordecai et al., 2019, 2017) exposed to increasing  
52 temperatures suggests that transmission is far from being linearly related to temperature, but more  
53 likely the result of complex interactions with asymmetric humped-shaped thermal responses. Therefore,  
54 the assumption that warming will aggravate the prevalence and severity of current endemic infections  
55 cannot be fully generalized among parasite species, and needs to be carefully evaluated across a broad  
56 range of temporal and spatial settings (Lafferty and Mordecai, 2016).

57  
58 Seasonality is one of the strongest environmental forces that affects the fluctuation of parasite  
59 prevalence and abundance over time (Altizer et al., 2006; Dowell, 2001; Harvell et al., 2002).  
60 Temperature warming could increase or decrease the intensity of parasite transmission without  
61 affecting the seasonal trend, alternatively, it could impact both the magnitude and duration of  
62 transmission and thus its seasonal shape (Altizer et al., 2006). This latter scenario has been proposed for  
63 a few helminth species under climate change, whose current season of spring-to-fall transmission is  
64 expected to split into two separate periods, spring and fall, divided by the emergence of a summer  
65 minimum (Altizer et al., 2013). For example, a bimodal pattern has been described for the sheep  
66 helminth *Haemonchus contortus* in southern Europe (Rose et al., 2016) and for the reindeer nematode  
67 *Ostertagia gruehneri* in the arctic Canada (Hoar et al., 2012; Kutz et al., 2014; Molnár et al., 2013b) where  
68 temperatures have been found to exceed the parasite thermal optimum. In addition to seasonal  
69 changes, we should also expect the geographical expansion of parasites in those areas where climate  
70 will become more suitable for survival and persistence, but a range contraction where parasites have  
71 limited thermal tolerance or slow adaptation to rapid climate changes (Kafle et al., 2020; Kenyon et al.,  
72 2009; Kutz et al., 2013, 2009; Short et al., 2017). Importantly, given the strong heterogeneity of global  
73 warming expected across geographical areas in the coming decades, these spatial trends should be  
74 more apparent at the extremes of the parasite range of distribution, where life history traits are under  
75 stronger constraints.

76  
77 While studies on parasites and vectors suggest that climate warming can alter the seasonal profile of  
78 infectious disease transmission and the range of species distribution, when and where changes in these  
79 patterns arise, what climatic variables affect these trends, and whether the observed patterns will be  
80 maintained as climate becomes warmer needs careful investigation. The frequent assumption is that  
81 many of the relationships between parasite life-history strategies and climate can be generalizable  
82 across species. However, parasite species might have different thermal responses to the same  
83 demographic trait and it is important to examine how these responses change among species or groups  
84 of species that have distinct ecological requirements (Molnár et al., 2013a). Moreover, given that

85 natural and domestic animal populations are often infected by a community of parasite species,  
86 evaluating whether climate change will promote or prevent opportunities for co-circulation of multiple  
87 species and risk of co-infection remains an overlooked issue (Clerc et al., 2018; Graham et al., 2007;  
88 Johnson and Buller, 2011). In fact, although these patterns ultimately depend on the distribution and  
89 abundance of the host populations, climate modulates the dynamics of free-living stages, including the  
90 viability of infective stages and thus the hazard of infection over time and space (Dobson et al., 2015;  
91 Molnár et al., 2017; Rose et al., 2016).

92  
93 In this study, we focused on the most common soil-transmitted helminths of some of the most familiar  
94 domestic and wild herbivore species and examined the historical and future impact of climate on the  
95 hazard of infection and related life history traits (i.e. egg hatching and mortality, larval development and  
96 survival) of stages free-living in the environment. Our goal is to identify commonalities and  
97 dissimilarities in the temporal and spatial demography of helminth species with diverse life histories  
98 under the direct effect of temperature and relative humidity. Given our interest in the free-living stages,  
99 and to reduce the complexity of using several host species, we did not explicitly address the role of the  
100 host populations or additional anthropogenic factors, like anthelmintic treatment or livestock  
101 management, on the hazard of infection. We focused on disentangling the climatic dependence of rates  
102 that drive the dynamics of free-living stages, not of their absolute abundance, making our framework  
103 easy to compare with other host-helminth systems. We selected parasites from the Trichostrongylidae  
104 family, which includes the genera *Trichostrongylus*, *Haemonchus*, *Ostertagia*, *Cooperia*, and  
105 *Nematodirus*, and commonly infect a large number of herbivore species, such as sheep, goats, and cattle  
106 for livestock as well as rabbits and hares for wildlife (Anderson, 2000). These helminths have a direct life  
107 cycle where infection occurs by ingestion of infective stages available on the pasture; adults colonize the  
108 gastrointestinal tract of their hosts, either the intestine or the stomach, and shed eggs in the  
109 environment via host's feces. These helminths represent classical examples of climate dependency  
110 identified through laboratory studies (Dobson et al., 2015; Hoar et al., 2012; Kafle et al., 2020; Kutz et  
111 al., 2005b; Molnár et al., 2017, 2013b; Rose et al., 2016). In addition, the Trichostrongylidae family is  
112 among the ones that cause consistent economic costs to the livestock industry (Charlier et al., 2020),  
113 and a better understanding of the relationship between hazard of infection and climate could improve  
114 livestock management and animal health. The selected species also have many similarities with soil-  
115 transmitted helminths of humans, and insights from their thermal dynamics can be used for public  
116 health prevention in endemic areas under climate warming.

117

## 118 **Materials and methods**

### 119 **Literature search and data analysis**

120 We performed a systematic review of published studies on the effect of temperature and relative  
121 humidity on free-living stages of soil-transmitted Trichostrongylidae of mammal herbivores following  
122 PRISMA (<https://prisma-statement.org/>). We limited our analysis to studies from the laboratory at  
123 controlled climatic regimes, which allowed us to include extreme conditions that might not be  
124 experienced in current natural settings but could represent future situations due to climate change. We  
125 screened an initial number of 244 publications and selected a total of 22 papers that met our criteria.

126 Nine species were identified from five genera (*Trichostrongylus*, *Haemonchus*, *Ostertagia*, *Cooperia*, and  
127 *Nematodirus*) (Sl.1, Table S1 and Figures S1,S2).

128 An initial analysis was undertaken to examine if the ecological and phylogenetic diversity of the  
129 considered species could explain differences in the traits of their stages free-living in the environment,  
130 specifically egg hatching and survival, 1<sup>st</sup> stage larval development, and 3<sup>rd</sup> stage larval survival (Figure  
131 1). ANOVA was used to test the contribution of parasite's phylogeny, host species, and site of infection,  
132 including their interactions with climatic variables, to variation in the observed traits (Sl.2, Table S2-S4).

### 133 Climate-driven model of free-living stages

134 The free-living component of the life cycle of soil-transmitted helminths can be described by the  
135 following set of ordinary differential equations, which captures the dynamics of eggs ( $E$ ), L1-stage larvae  
136 ( $L_1$ ), and L3-stage infective larvae ( $L_3$ ) in time with climate-dependent parameters, as follows (Figure 1):  
137

$$\begin{cases} \frac{dE}{dt} = \varphi - (h(T) + \mu_E(T))E \\ \frac{dL_1}{dt} = h(T)E - [d(T) + \mu_{L_1}(T)]L_1 \\ \frac{dL_3}{dt} = d(T)L_1 - [\mu_{L_3}(T, H) + \gamma Y]L_3 \end{cases} \quad (1)$$

138 where  $t$  represents time in days,  $\varphi$  is the daily rate of eggs shed in the environment,  $h$  is the egg  
139 hatching rate into L1,  $\mu_E$  is the egg mortality rate,  $d$  is the larval development rate from L1 to L3,  $\mu_{L_1}$  and  
140  $\mu_{L_3}$  are the larval mortality rate of L1 and L3, respectively, and  $\gamma Y$  represents the uptake of infective  
141 larvae by the hosts, with  $\gamma$  and  $Y$  being the grazing rate and the host abundance, respectively. The three  
142 stages are affected by mean air temperature ( $T$ , thereafter referred to as temperature, °C) and/or air  
143 relative humidity ( $H$ , thereafter referred to as humidity, %). Eggs and L1 live in the host's feces and we  
144 assumed that feces maintain the minimum humidity required for their viability; in contrast, L3 larvae  
145 emerge from the feces to live on the pasture and are under the direct influence of climatic conditions.  
146 For simplicity, we assumed that the mortality of L1 in the feces is negligible ( $\mu_{L_1} = 0$ ) and omitted to  
147 model the intermediate larval stage L2, which also lives in the feces and is morphologically similar to  
148 L1.  
149

150 We expressed the rate of egg hatching as a temperature-dependent process, described by the following  
151 Degree-Day ( $DD$ ) functions (Dobson et al., 2015; Hernandez et al., 2013; Hsu and Levine, 1977):  
152

$$\begin{aligned} DD(t) &= \begin{cases} T(t) - T_H & \text{if } T(t) > T_H \\ 0 & \text{if } T(t) \leq T_H \end{cases} \\ CDD(t) &= \sum_{\xi=t_s}^t DD(\xi) \\ CDD(t_H) &= H^* \end{aligned} \quad (2)$$

153 where  $t_s$  and  $t_H$  (days) are the egg shedding time and the hatching time, respectively,  $T_H$  (°C) is the  
154 baseline temperature for  $DD$  accumulation, and  $H^*$  is the Cumulative  $DD$  ( $CDD$ ) threshold needed by  
155 eggs to hatch. The thermal-dependent mortality of eggs was modeled as a sum of exponential functions  
156 of temperature described by the non-negative parameters  $b_{1E}$ ,  $\theta_{1E}$ ,  $b_{2E}$ , and  $\theta_{2E}$  (Smith, 1990) as:

157

$$\mu_E(T) = b_{1E}e^{-\beta_{1E}T} + b_{2E}e^{\beta_{2E}T} \quad (3)$$

158

159 The rate at which larvae L1 develop into infective L3 depends on temperature following a *DD* model  
 160 similarly to equation 2 for egg hatching (Dobson et al., 2015; Hernandez et al., 2013; Hsu and Levine,  
 161 1977):

$$DD_D(t) = \begin{cases} T(t) - T_{Di} & \text{if } T(t) > T_D \\ 0 & \text{if } T(t) \leq T_D \end{cases}$$

$$CDD_D(t) = \sum_{\xi=t_H}^t DD_D(\xi) \quad (4)$$

$$CDD_D(t_D) = D^*$$

162

163 with  $t_D$  representing the development time in days,  $T_D$  being the baseline temperature for degree day  
 164 accumulation ( $^{\circ}\text{C}$ ) and  $D^*$  being the *CDD* threshold needed by L1 to develop into L3 larvae.

165 We assumed that the L3 mortality rate depends both on the daily temperature and humidity, with a  
 166 functional relationship that was modeled as a sum of exponential functions of temperature (Smith,  
 167 1990) and as a linear function of humidity and as a linear function of humidity (Dagostin et al., 2023;  
 168 Mignatti et al., 2016) as:

$$\mu_{L3}(T, H) = \begin{cases} b_{1L}e^{-\beta_{1L}T} + b_{2L}e^{\beta_{2L}T} + c_L H & \text{if } \mu_{L3} > 0 \\ 0 & \text{if } \mu_{L3} \leq 0 \end{cases} \quad (5)$$

169 Where the non-negative parameters  $b_{1L}$ ,  $\beta_{1L}$ ,  $b_{2L}$  and  $\beta_{2L}$  depict the impact of temperature while  $c_L$   
 170 describes the effect of humidity.

171 We used the gathered experimental observations grouped in intestinal and stomach helminths (see SI.2  
 172 for details) and calibrated the climate-driven functions of egg hatching and mortality, L1 development  
 173 and L3 mortality (eqs. 2-5) minimizing the root mean square error (*ERR*) for each group independently  
 174 as:

$$ERR = \sqrt{\frac{\sum_{z=1}^n (\log(\bar{x}_z / \hat{x}_z))^2}{n}} \quad (6)$$

175 where  $\bar{x}_z$  and  $\hat{x}_z$  represent the observed and estimated, via the described model equations, free-living  
 176 rates, respectively, which were log-transformed to better compare rates of different order of  
 177 magnitude, while  $n$  is the sample size.

178 Finally, we evaluated the net effect of the different rates on the pool of infective L3 larvae in the  
 179 environment. Assuming that  $T$  and  $H$  vary on a longer time scale than  $E$ ,  $L_1$ ,  $L_3$ , we can calculate L3 at  
 180 quasi-equilibrium for any given  $T$  and  $H$  as:

$$r(T, H) = \bar{L}_3 = \frac{d(T)h(T)\varphi}{[d(T) + \mu_{L1}(T)][h(T) + \mu_E(T)][\mu_{L3}(T, H) + \gamma Y]} \quad (7)$$

181 This quantity represents a proxy for the climatic-dependent hazard of infection for any given host  
 182 exposed to these helminths. To simplify the analysis, the infection hazard was calculated per unit of egg  
 183 shedding rate ( $\varphi=1$ ) while host mortality induced by L3 uptake was considered to be negligible ( $\gamma Y = 0$ ).

184 To identify the parameters with the strongest effect on the hazard of infection and to evaluate the  
185 sensitivity to parameter uncertainty, we implemented the Latin Hypercube Sampling and Partial Rank  
186 Correlation (PRC) analysis (Blower and Dowlatabadi, 1994; Marino et al., 2008; McLeod et al., 2006),  
187 independently for stomach and intestinal helminths (SI.5).

## 188 **Historical and future hazard to helminth infection in Europe**

189 The developed climate-dependent model was then used to examine the climatic responses of eggs, L1,  
190 and L3 rates and the resulting hazard of infection intra-annually and throughout Europe, for stomach  
191 and intestinal helminths. Using daily mean temperature and relative humidity estimates from the  
192 EUROCORDEX (European Coordinated Regional Climate Downscaling Experiment) dataset at a spatial  
193 resolution of  $\sim 12.5 \times 12.5$  km<sup>2</sup> (<https://www.euro-cordex.net/>), we considered a total of 44,331 spatial  
194 cells, reasonably assuming spatial-independence with no movements of eggs or larvae between cells.

195 We approached this analysis in two phases. First, using a daily time step for each cell of the spatial  
196 domain, we assessed the infection hazard and related free-living stage rates, both for stomach and  
197 intestinal helminths, in the selected historical period of 20-years (1981-2000) to establish a  
198 representative baseline scenario. Second, we evaluated changes in the infection hazard of the two  
199 groups in a future 20-year period (2071-2090), under the Representative Concentration Pathway (RCP)  
200 8.5 of “business as usual” high emission scenario (IPCC, 2013). Moreover, we chose the RCP 8.5  
201 projections as representative of extreme, albeit not completely unrealistic scenarios, and we expect that  
202 more moderate scenarios would lie in-between the historical and the RCP 8.5 trajectories.

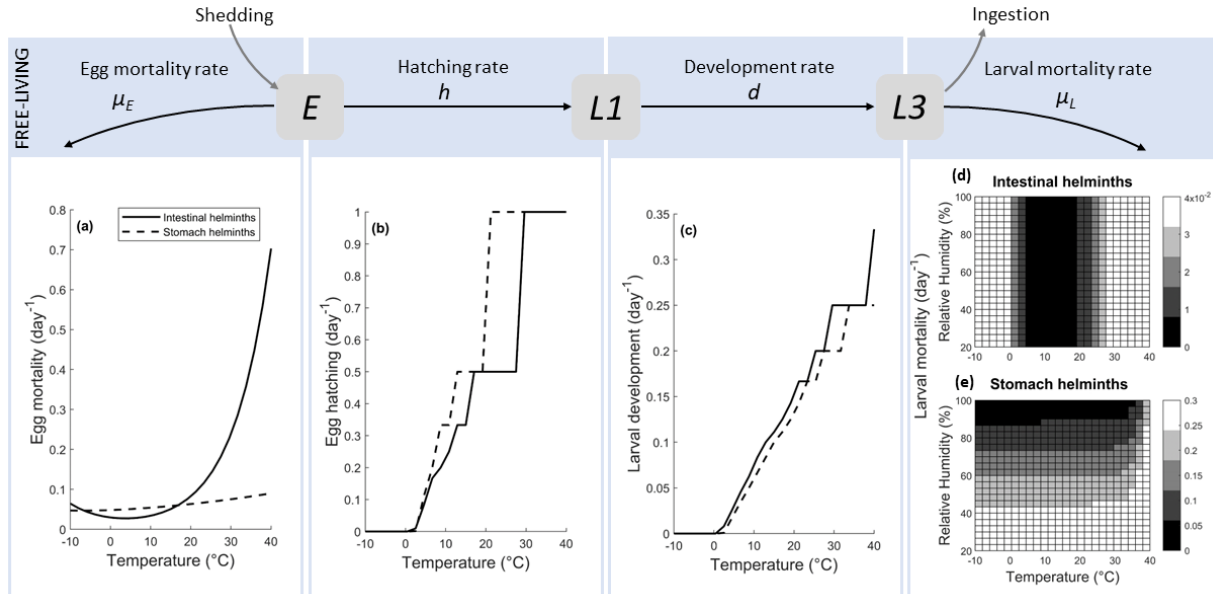
203 To account for the climatic variability of the European territory, we divided Europe in three main  
204 latitudinal areas, which correspond to the main European climatic zones, namely Mediterranean,  
205 temperate, and Nordic climates for southern, central and northern Europe, respectively (SI.3, Figure S3).

## 206 **Results**

### 207 **Climatic responses of helminth free-living stages and hazard**

208 The observed rates of egg hatching and mortality, larval L1 development, and L3 mortality were  
209 significantly different when the site of infection of the nine Trichostrongylidae species was considered  
210 ( $p < 0.05$ ), while neither the phylogenetic distance between species nor their host species had a  
211 significant effect (SI.2, Tables S2-S4). Therefore, we clustered the gathered data in two groups, intestinal  
212 (total observations=128) and stomach (total observations=162) helminths, and all the subsequent  
213 analyses were carried out independently for each group (SI.4, Table 1, Figure S4,S5). Overall, we found  
214 contrasting thermal responses between the two groups. Intestinal helminths showed to have lower  
215 baseline temperatures ( $T_H$  and  $T_D$ ) and higher degree day ( $DD$ ) cumulated thresholds ( $H^*$  and  $D^*$ ) both for  
216 egg hatching, but not for L1 development, compared to stomach helminths (Figure 1b,c and Table 1).  
217 Moreover, the egg mortality rate was more clearly affected by temperature for intestinal than stomach  
218 helminths (Figure 1a). Stronger differences were found for L3 mortality, for intestinal helminths the  
219 mortality rate was well described by a skewed U-shaped temperature function with a minimum at  
220 10.5°C, while for stomach helminths it was primarily driven by a negative linear relationship with  
221 humidity, and an increase in mortality above the temperature extreme of 30°C (Figure 1d,e).

222



223  
224 **Figure 1.** Life cycle of free-living helminths (upper blue panel) and climate dependencies of demographic rates for intestinal  
225 (bold lines a-c and d) and stomach (dashed lines a-c and e) helminths from laboratory data.  
226

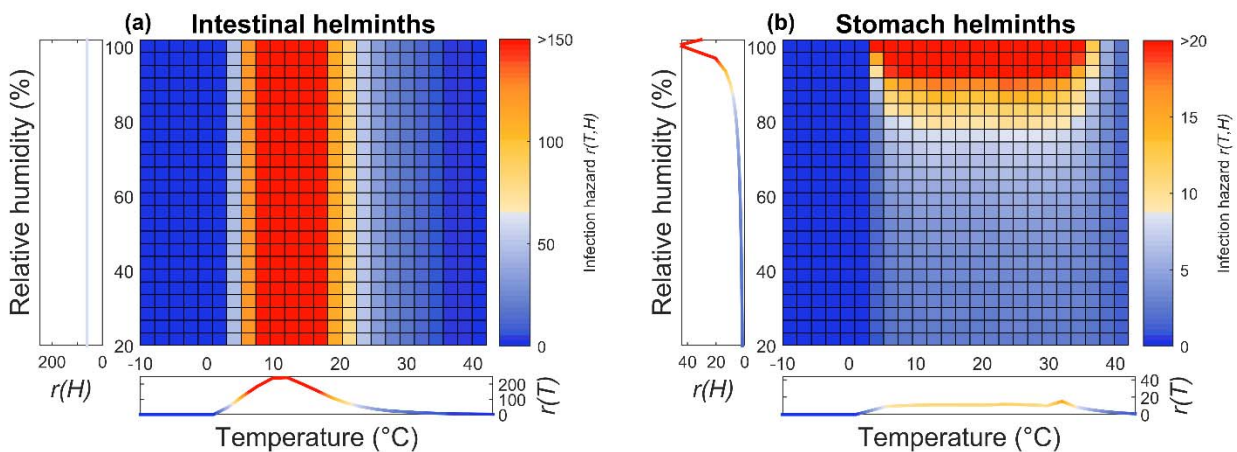
227 **Table 1.** Calibrated parameters ( $b_{1E}$ ,  $\theta_{1E}$ ,  $b_{2E}$ ,  $\theta_{2E}$ ,  $T_H$ ,  $H^*$ ,  $T_D$ ,  $D^*$ ,  $b_{1L}$ ,  $\theta_{1L}$ ,  $b_{2L}$ ,  $\theta_{2L}$  and  $c_L$ ) for the rates of egg mortality and  
228 hatching, larval L1 development and L3 mortality as functions of temperature and relative humidity for intestinal and stomach  
229 helminths from laboratory data;  $n$  indicates the sample size and  $ERR$  represents the Root Mean Square Error described in eq. 6.  
230

	Parameter	Unit	Intestinal helminths	Stomach helminths
<b>Egg mortality rate</b> ( $n=60$ ) $ERR_{intestinal}=0.570$ $ERR_{stomach}=1.095$	$b_{1E}$	day <sup>-1</sup>	$2.05 \times 10^{-2}$	$5.09 \times 10^{-3}$
	$\theta_{1E}$	°C <sup>-1</sup>	0.110	$7.90 \times 10^{-2}$
	$b_{2E}$	day <sup>-1</sup>	$9.16 \times 10^{-3}$	$4.31 \times 10^{-2}$
	$\theta_{2E}$	°C <sup>-1</sup>	0.109	$1.84 \times 10^{-2}$
<b>Egg hatching rate</b> ( $n=92$ ) $ERR_{intestinal}=0.384$ $ERR_{stomach}=0.418$	$T_H$	°C	2.25	2.48
	$H^*$	DD	26.2	18.2
<b>Larval L1 development rate</b> ( $n=52$ ) $ERR_{intestinal}=0.635$ $ERR_{stomach}=0.291$	$T_D$	°C	1.73	2.41
	$D^*$	DD	110	122
<b>Larval L3 mortality rate</b> ( $n=86$ ) $ERR_{intestinal}=0.917$ $ERR_{stomach}=1.005$	$b_{1L}$	day <sup>-1</sup>	$2.11 \times 10^{-2}$	0.418
	$\theta_{1L}$	°C <sup>-1</sup>	0.253	$7.92 \times 10^{-5}$
	$b_{2L}$	day <sup>-1</sup>	$3.74 \times 10^{-4}$	$2.30 \times 10^{-9}$
	$\theta_{2L}$	°C <sup>-1</sup>	0.166	0.418
	$c_L$	day <sup>-1</sup>	0	$-4.13 \times 10^{-3}$

231  
232 We then examined the infection hazard for each helminth group under a range of temperature and  
233 humidity conditions, and differences between these two groups were evident (Figure 2). For intestinal  
234 helminths, the hazard was strongly affected by temperature peaking at the optimal temperature of  
235 10°C, which corresponds to the highest L3 survival, and dropping to zero below 2°C and above 35°C

236 because of a decrease in egg hatching and development or high L3 mortality, respectively. Humidity  
237 appeared to have no clear effect on the hazard of intestinal helminths (Figure 2a). In contrast, humidity  
238 substantially impacted the hazard of stomach helminths in that the greatest hazard was found at above  
239 80% humidity within the 5°C-35°C temperature range (Figure 2b). The partial rank correlation  
240 coefficients from the performed sensitivity analysis reinforce our findings by showing that the relative  
241 contribution of the different parameters to the infection hazard consistently differs between the two  
242 helminth groups (Figure S6).

243  
244 These findings highlight the contrasting climatic performance of intestine and stomach helminths when  
245 exposed to the same temperature and humidity conditions. Moreover, these differences show that the  
246 net climatic effect on the infection hazard is determined by the complex balance between different  
247 demographic traits and their thermal dependence, namely, how fast eggs can hatch and for how long  
248 eggs and infective L3 can remain viable in the environment.  
249



250  
251 **Figure 2.** Response of the infection hazard  $r(T,H)$  of (a) intestinal and (b) stomach helminths under different temperatures (x-  
252 axis) and relative humidities (y-axis), estimated from laboratory data. Profiles on the x- and y-axis represent the average  
253 infection hazard by temperature  $r(T)$ , and by humidity  $r(H)$ , respectively.

254

## 255 Historical and future seasonality of helminth hazard

256 The relationships between climatic variables and demography of free-living helminths, previously  
257 identified and trained on laboratory data, were then used to investigate how the infection hazard is  
258 affected by seasonality in the three European zones for the historical (1981-2000) period and future  
259 (2071-2090) projections under the RCP 8.5 climate change scenario (Figures 3, S7, SI.6).

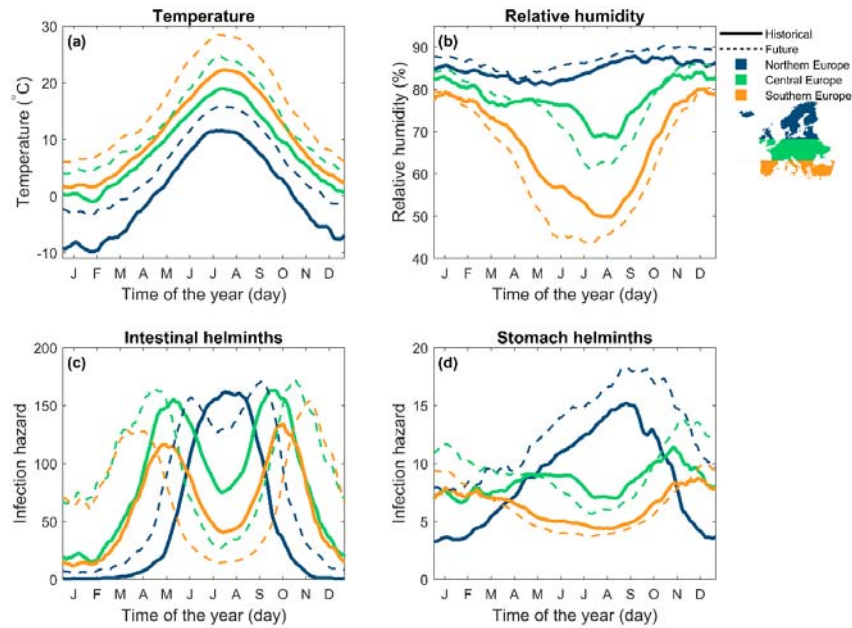
260 During the selected historical period, temperature exhibited the classical annual unimodal profile with a  
261 peak in summer in all the three European zones and variation in the absolute values according to their  
262 latitudinal gradient (Figure 3a). The seasonal trend of humidity was more variable among the three  
263 zones: humidity was above 80% and relatively constant in northern Europe but exhibited a summer  
264 minimum more pronounced in southern than central Europe (Figure 3b). Our model simulations indicate  
265 that climate seasonality led to contrasting patterns in the hazard of infection between the two helminth  
266 groups and among the geographical zones within each group (Figure 3). The hazard of intestinal



267 helminths exhibited a unimodal trend peaking in summer for northern Europe and a biannual pattern  
268 with peaks in spring and fall for central and southern Europe (Figure 3c). The striking difference between  
269 latitudinal zones appears to be caused by the summer mismatch between the rates of  
270 hatching/development and mortality, being the latter greater for eggs and larvae at low latitudes  
271 compared to high latitudes (Figure S7). The hazard of stomach helminth infection depicted a different  
272 scenario characterized by a late-summer peak in northern Europe, probably facilitated by a generally  
273 low L3 mortality coupled with fast egg hatching and L1 development in summer (Figure 3d, S7). In the  
274 remaining European zones, the hazard followed the humidity profile, except for winter months when  
275 cold temperatures are a restraining factor for hatching and development (Figure 3d). At lower European  
276 latitudes, the two helminth groups showed the most compelling seasonal differences, suggesting strong  
277 differences in the fulfillment of their ecological requirements.

278 Projections at the end of the XXI century under the RCP 8.5 scenario indicated that climate change is  
279 expected to cause an increase of temperatures, which will maintain the same seasonal trends for the  
280 three European zones (Figure 3a). Humidity is also projected to change and exacerbate the preexisting  
281 latitudinal differences by causing a more humid northern Europe and drier central and southern zones,  
282 particularly in summer months (Figure 3b). Projections suggested that the climatic responses of free-  
283 living stages will reflect these seasonal changes by showing an increase in the dissimilarities between  
284 the two helminth groups, already highlighted during the historical period. The bimodal seasonality noted  
285 for intestinal helminths in the historical period is expected to become more prominent with the shift  
286 and increase of the two peaks towards early spring and late fall. We also expect the emergence of a  
287 similar bimodal trend in northern Europe and a general increase of the hazard of infection in the winter  
288 months in three European zones (Figure 3c). The intensification of this bimodality is mainly caused by a  
289 lower egg and larval survival during the summer months (Figure S7). The hazard of stomach helminth  
290 infection will drastically increase throughout the year in northern Europe. At lower latitudes, the  
291 coupling of drier and warmer conditions from spring to fall is expected to negatively affect the hazard,  
292 while warmer and slightly wetter winters will favor infections (Figure 3).

293 In summary, climate warming is expected to increase the intensity and timing of the hazard of infection  
294 of both helminth groups on the pasture in the fall-winter months but will have an opposite effect in the  
295 summer months; this change will also be associated with a temporal shift and increase of the peak.



296  
297 **Figure 3.** Average historical (1981-2000, bold lines) and future (2071-2090 under RCP 8.5 scenario, dashed lines) seasonality of  
298 (a) temperature, (b) relative humidity, and (c,d) infection hazard of (c) intestinal and (d) stomach helminths in northern (blue),  
299 central (green), and southern (orange) Europe, plotted with two weeks moving average for visual representation.

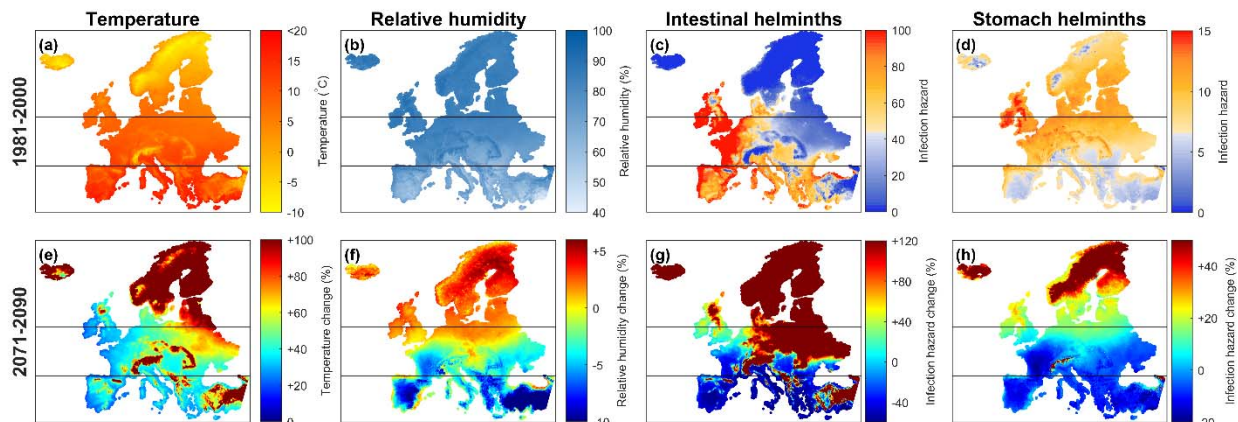
### 300 **Historical and future spatial distribution of helminth hazard**

301 We examined the spatial distribution of the helminth hazard during the selected historical period (1981-  
302 2000) and the expected percentage changes in the future (2071-2090) under the RCP 8.5 climate change  
303 scenario. (Figure 4, S8, S9, Sl.7).

304 The historical spatial distribution of the average annual temperature and humidity confirms the well-  
305 characterized latitudinal trends (Figure 4a,b). These geographical differences were found to have  
306 contrasting effects on the spatial distribution of the demographic rates withintestinal helminths  
307 exhibiting a minimum hazard of infection in the north-eastern part of the continent, which on the  
308 contrary, represented an infection hotspot with high hazard for stomach helminths (Figures 4c,d, S8).

309 The projected changes of temperature and humidity at the end of the XXI century will not occur  
310 uniformly throughout Europe (Figure 4e,f). Southern and northern Europe are expected to experience a  
311 drastic warming, up to +6 °C, with a decrease of humidity in the former (-10%) and an increase in the  
312 latter (+5%) zone. In central Europe, temperature is projected to increase by +4 °C, but humidity should  
313 remain mostly unchanged. Consequently, Scandinavian countries are expected to experience a greater  
314 hazard of infection for both helminth groups (increase of 100% for intestinal helminths and 55% for  
315 stomach helminths) due to a fast egg hatching and L1 development paired with high egg and L3 survival  
316 (Figures 4g,h, S9). On the contrary, the substantial warmer temperatures and drier conditions in the  
317 Mediterranean countries are projected to reduce the hazard of both helminth groups (decrease of 60%  
318 for intestinal helminths and 20% for stomach helminths) due to a drastic increase of egg and L3  
319 mortality and (Figures 4g,h, S9). Additionally, our projections highlight dissimilarities between the two  
320 groups for high altitude areas, such as Pyrenees, Alps, Pontic mountains, where we expect a higher  
321 increase in the hazard of intestinal helminth infections compared to stomach helminths (Figure 4g,h).

322 Overall, in addition to identifying a spatial shift towards northern latitudes of the hazard of infection and  
323 associated decrease at lower latitude for both helminths, our maps confirm the strong regional variation  
324 in the climate-hazard relationship.



325 **Figure 4.** Average historical (1981-2000) (a) temperature, (b) relative humidity, hazard of infection of (c) intestinal and (d)  
326 stomach helminths and future (2071-2090) expected changes (e-h) under the RCP 8.5 climate change scenario. Horizontal lines  
327 separate northern, central and southern zones.  
328  
329

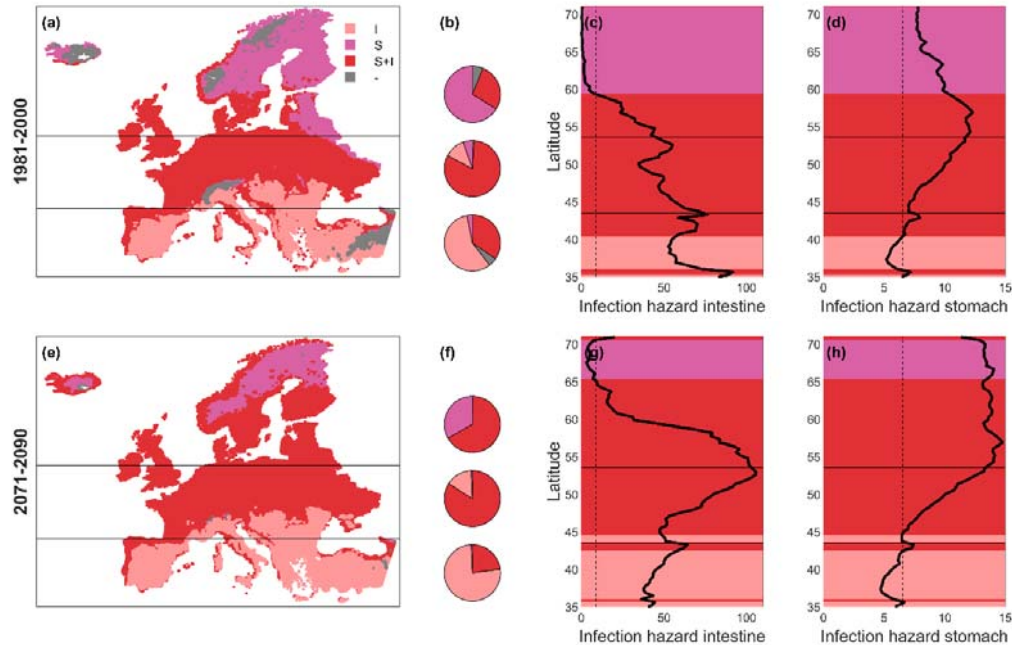
### 330 **Spatiotemporal changes of helminth co-occurrence**

331 The co-occurrence of intestinal and stomach helminths was evaluated to examine the degree of spatial  
332 overlapping and to determine whether areas of co-circulation may expand in the future (SI.8, Figure  
333 S10).

334 In central Europe, the co-occurrence of intestinal and stomach helminths (82% of spatial cells) is  
335 facilitated by climatic conditions favorable for both groups, particularly intermediate temperatures and  
336 high humidity for intestinal and stomach helminths, respectively. Southern Europe generates climatic  
337 areas where single circulation of intestinal species predominates (57%), whereas single circulation of  
338 stomach species appears more common in northern Europe (65%) (Figure 5a,b). Unfavorable areas to  
339 both helminth groups are only represented by 4% of the entire European territory and are mainly  
340 located in the Alps, Scandinavian mountains, and Iceland. When we examined the average infection  
341 hazard across the latitudinal gradient, results confirmed that the co-circulation of both groups in the  
342 environment predominates, except for very high latitudes where single infection by stomach helminths  
343 appears to prevail (Figure 5c,d).

344 Future projections indicate an expansion of the distribution of small intestine helminths in the  
345 Mediterranean regions (+19%) and co-circulation of both groups in northern Europe (+39%, Figure 6e,f).  
346 Unfavorable areas to helminth infection are expected to decrease in the future (from 4% to less than  
347 1%). Overall, our predictions suggest that for mid-high latitudes the probability of co-occurrence of  
348 stomach and intestinal helminths in the environment is expected to be higher given that the relative  
349 hazard of infection of both groups will substantially increase.

350



351  
352 **Figure 5.** Historical (1981-2000) (a-d) and future (2071-2090) (e-h) single occurrence of intestinal helminths (I, pink), single  
353 occurrence of stomach helminths (S, purple), co-occurrence of both helminth groups (S+I, red), and no occurrence of both  
354 groups (-, grey) in Europe. Spatial distribution (a,e), composition of spatial cells (b,f) in northern Europe (top pie), central  
355 Europe (mid pie), southern Europe (bottom pie) based to their occurrence type, and average infection hazard of intestinal (c,g)  
356 and stomach (d,h) helminths. Black bold lines (c,d,g,h) represent the average infection hazard across the European latitudes,  
357 the vertical dashed black line is the threshold  $\bar{r}$  and the color in the background represents the average occurrence type at each  
358 latitude.  
359

## 360 Discussion

361 We developed a climate-driven mechanistic model to examine the demographic responses of  
362 trichostrongylid helminths, aggregated in intestinal and stomach groups, and their hazard of infection.  
363 This framework was then applied at the European continental scale to evaluate changes in the seasonal  
364 and spatial distribution of the two groups during historical and projected climatic scenarios. Our  
365 predictions suggest that under the high emission scenario RCP 8.5 climate will severely modify the  
366 seasonal transmission and the spatial distribution of the two groups according to their distinct ecological  
367 requirements and climatic tolerance along the latitudinal and altitudinal gradients. A drastic increase of  
368 the infection hazard at mid-high latitudes will likely increase the co-circulation of stomach and intestine  
369 helminth species with critical consequences for animal health in the future.

370 We found clear differences in the response of helminths to temperature and humidity based on their  
371 site of infection in the mammal host. Multiple ecological factors could contribute to explaining this  
372 contrasting trend. One possibility could be related to the intrinsic differences in the life history  
373 properties between the two groups. For example, the body size of adult female is bigger in stomach  
374 than intestine helminths, irrespective of their mammal host, a pattern that is partially reflected in the  
375 volume of their eggs (Table S1). Individual body size is a fundamental characteristic that influences the  
376 thermal sensitivity of an organism, especially for ectotherms, where larger sizes are commonly  
377 associated with higher temperature tolerance and fitness (Kingsolver and Huey, 2008). It is indeed  
378 possible that the bigger stomach helminths produce eggs, and likely infective larvae, of better quality

379 and larger size that can thrive under high temperatures. Differences in the immune response between  
380 the two groups, and likely affecting female fecundity including egg quantity and quality, could also  
381 contribute to the observed patterns. Previous studies have suggested that host immunity can constrain  
382 the fitness, in terms of body size and/or egg abundance in uterus, of adult helminth females and  
383 consequently the hatching potential of their eggs (Hein et al., 2010; Lambert et al., 2015; Viney and  
384 Cable, 2011; Wakelin, 1987). For example, a negative relation between the host immune response and  
385 both female length and number of eggs in uterus was found for the stomach helminth *O. circumcincta* in  
386 sheep (Stear et al., 1995) and the intestinal *T. retortaeformis* in rabbits (Cattadori et al., 2005, 2019,  
387 2014; Chylinski et al., 2009). Given the complex relation between host immunity and helminth fitness,  
388 we cannot exclude that the immune response could affect the way helminths respond to different  
389 climatic conditions.

390

391 Our model projections suggest that there are significant differences in the seasonal and spatial trends of  
392 the hazard of infection both between and within helminth groups throughout Europe. The emergence of  
393 a bimodal seasonal hazard with a minimum in summer for intestinal helminths is determined by the  
394 overshoot of the thermal optimum during extreme hot periods, a profile described for the historical  
395 time in central-southern Europe and likely to be exacerbated in future warmer conditions. Importantly,  
396 a similar bimodal pattern is expected for the northern regions in the long-term as they are projected to  
397 experience extreme hot temperatures. These findings are in agreement with climate change predictions  
398 from theoretical models (Altizer et al., 2013; Molnár et al., 2013a), and frameworks applied to  
399 trichostrongylid infections under different climate change scenarios (Rose et al., 2016, 2015). For  
400 stomach helminths, whose spatiotemporal dynamics are mainly affected by humidity, central-northern  
401 Europe could be potentially expose to a higher infection hazard due to relative warm and humid  
402 summers within the tolerance range of these helminth species. Our simulations indicate that stomach  
403 helminths are distributed across Europe following an increasing south-north gradient, which is the  
404 opposite for the intestinal helminths. Noteworthy, the infection hazard of both helminth groups is  
405 expected to expand towards the northern latitudes and contract in the southern territories. These  
406 climate-driven spatial shifts are in agreement with projected distributions of helminths of wild (e.g. Dall  
407 sheep, muskoxen) and farm (e.g. sheep) animals in northern Europe and Canada, where a more  
408 permissive climate is already contributing to expand the spatial ranges of the local parasite species  
409 (Jenkins et al., 2006; Kutz et al., 2009, 2005b; Rose et al., 2016). Similarly, the warming of the arctic  
410 tundra has been shown to likely allow for the completion of the parasite life cycles facilitating the  
411 establishment and expansion towards northern areas (Kafle et al., 2020; Kutz et al., 2013). Anticipating  
412 the spatiotemporal shifts of the hazard of helminth infection is particularly important from an applied  
413 perspective. At present, helminth treatment represents 20% of the total economic burden to the  
414 livestock industry in Europe (Charlier et al., 2020) and an alarming positive trend is expected to continue  
415 in the future due to the spread of anthelmintic resistance (Geerts and Gryseels, 2001; Rose Vineer et  
416 al., 2020) and/or the lack of regular prophylactic. Redesigning the timing and the area of interest of  
417 these treatment strategies could reduce the spread of anthelmintic resistance and contribute to a  
418 more effective control of helminth infections.

419 By investigating the response of closely related species exposed to the same climatic drivers, our work  
420 provides a more comprehensive understanding of the climate change impact on the co-circulation and  
421 potential coinfection of multiple helminth species. Model projections identify the European mid-high

422 latitudes as future critical areas for the co-circulation of intestinal and stomach helminths, with an  
423 expected increase of the hazard by +100% and +30%, respectively. Moreover, future climate changes  
424 might increase the likelihood of synchronous seasonal peaks of infection hazard to both groups. This  
425 spatial and seasonal overlapping could facilitate coinfection events, given the exposure to a competent  
426 host, and increase disease severity and transmission with consequences for species co-endemicity and  
427 infection hotspots. Recently, we showed that while under the same climatic conditions coinfecting  
428 rabbits shed more *T. retortaeformis* and *G. strigosum* eggs that survive for longer in the environment,  
429 compared to single-infected hosts (Dagostin et al., 2023), supporting the important role that co-infected  
430 individuals could play in disease transmission under climate change.

431 Our results bring a new perspective to the study of climate and infectious diseases by addressing the  
432 role of humidity as climatic driver of helminth transmission. Indeed, despite the well-recognized  
433 humidity requirements for helminth life cycles (Beveridge et al., 1989; Pandey et al., 1993; Prasad, 1959)  
434 this variable is usually neglected (but see (Dagostin et al., 2023; Maya et al., 2010)) in favor of  
435 temperature, which is more frequently recorded in the field and easy to manipulate in the laboratory.  
436 We show that humidity is a key environmental variable for understanding the climate-driven dynamics  
437 of stomach helminth free-living stages and important for generating heterogeneities in the demographic  
438 responses of both intestinal and stomach group. Moreover, the inclusion of humidity provides a more  
439 accurate understanding of the relationship between climate and hazard of infection, including a more  
440 rigorous projection under future climate change.

441 Ultimately, our work shows that the ecological requirements of different soil-transmitted helminth  
442 species modulate whether and how climate change will modify the magnitude, seasonality and  
443 distribution of the infection hazard and the co-circulation of multiple helminth species over time and  
444 space.

## 445 References

- 446 Altizer, S., Dobson, A., Hosseini, P., Hudson, P., Pascual, M., Rohani, P., 2006. Seasonality and the  
447 dynamics of infectious diseases. *Ecol. Lett.* 9, 467–484.
- 448 Altizer, S., Ostfeld, R.S., Johnson, P.T., Kutz, S., Harvell, C.D., 2013. Climate change and infectious  
449 diseases: from evidence to a predictive framework. *science* 341, 514–519.
- 450 Anderson, R.C., 2000. Nematode parasites of vertebrates: their development and transmission. Cabi.
- 451 Beveridge, I., Pullman, A.L., Martin, R.R., Barelds, A., 1989. Effects of temperature and relative humidity  
452 on development and survival of the free-living stages of *Trichostrongylus colubriformis*, *T.*  
453 *rugatus* and *T. vitrinus*. *Vet. Parasitol.* 33, 143–153.
- 454 Blower, S.M., Dowlatabadi, H., 1994. Sensitivity and uncertainty analysis of complex models of disease  
455 transmission: an HIV model, as an example. *Int. Stat. Rev. Int. Stat.* 229–243.
- 456 Carraro, L., Bertuzzo, E., Mari, L., Fontes, I., Hartikainen, H., Strepparava, N., Schmidt-Posthaus, H.,  
457 Wahli, T., Jokela, J., Gatto, M., 2017. Integrated field, laboratory, and theoretical study of PKD  
458 spread in a Swiss prealpine river. *Proc. Natl. Acad. Sci.* 114, 11992–11997.
- 459 Cattadori, I. m, Boag, B., Bjørnstad, O. n, Cornell, S. j, Hudson, P. j, 2005. Peak shift and epidemiology in a  
460 seasonal host–nematode system. *Proc. R. Soc. B Biol. Sci.* 272, 1163–1169.  
461 <https://doi.org/10.1098/rspb.2004.3050>
- 462 Cattadori, I.M., Pathak, A.K., Ferrari, M.J., 2019. External disturbances impact helminth–host  
463 interactions by affecting dynamics of infection, parasite traits, and host immune responses. *Ecol.*  
464 *Evol.* 9, 13495–13505. <https://doi.org/10.1002/ece3.5805>

- 465 Cattadori, I.M., Wagner, B.R., Wodzinski, L.A., Pathak, A.K., Poole, A., Boag, B., 2014. Infections do not  
466 predict shedding in co-infections with two helminths from a natural system. *Ecology* 95, 1684–  
467 1692.
- 468 Charlier, J., Rinaldi, L., Musella, V., Ploeger, H.W., Chartier, C., Vineer, H.R., Hinney, B., von Samson-  
469 Himmelstjerna, G., Băcescu, B., Mickiewicz, M., 2020. Initial assessment of the economic burden  
470 of major parasitic helminth infections to the ruminant livestock industry in Europe. *Prev. Vet.  
471 Med.* 182, 105103.
- 472 Chylinski, C., Boag, B., Stear, M.J., Cattadori, I.M., 2009. Effects of host characteristics and parasite  
473 intensity on growth and fecundity of *Trichostrongylus retortaeformis* infections in rabbits.  
474 *Parasitology* 136, 117–123.
- 475 Clerc, M., Devevey, G., Fenton, A., Pedersen, A.B., 2018. Antibodies and coinfection drive variation in  
476 nematode burdens in wild mice. *Int. J. Parasitol.* 48, 785–792.  
477 <https://doi.org/10.1016/j.ijpara.2018.04.003>
- 478 Dagostin, F., Vanalli, C., Boag, B., Casagrandi, R., Gatto, M., Mari, L., Cattadori, I.M., 2023. The enemy of  
479 my enemy is my friend: Immune-mediated facilitation contributes to fitness of co-infecting  
480 helminths. *J. Anim. Ecol.* 92, 477–491. <https://doi.org/10.1111/1365-2656.13863>
- 481 Deutsch, C.A., Tewksbury, J.J., Huey, R.B., Sheldon, K.S., Ghalambor, C.K., Haak, D.C., Martin, P.R., 2008.  
482 Impacts of climate warming on terrestrial ectotherms across latitude. *Proc. Natl. Acad. Sci.* 105,  
483 6668–6672.
- 484 Dobson, A., Molnár, P.K., Kutz, S., 2015. Climate change and Arctic parasites. *Trends Parasitol.* 31, 181–  
485 188.
- 486 Dowell, S.F., 2001. Seasonal variation in host susceptibility and cycles of certain infectious diseases.  
487 *Emerg. Infect. Dis.* 7, 369.
- 488 Geerts, S., Gryseels, B., 2001. Anthelmintic resistance in human helminths: a review. *Trop. Med. Int.  
489 Health* 6, 915–921.
- 490 Gehman, A.-L.M., Hall, R.J., Byers, J.E., 2018. Host and parasite thermal ecology jointly determine the  
491 effect of climate warming on epidemic dynamics. *Proc. Natl. Acad. Sci.* 115, 744–749.
- 492 Graham, A.L., Cattadori, I.M., Lloyd-Smith, J.O., Ferrari, M.J., Bjørnstad, O.N., 2007. Transmission  
493 consequences of coinfection: cytokines writ large? *Trends Parasitol.* 23, 284–291.
- 494 Harvell, C.D., Mitchell, C.E., Ward, J.R., Altizer, S., Dobson, A.P., Ostfeld, R.S., Samuel, M.D., 2002.  
495 Climate warming and disease risks for terrestrial and marine biota. *Science* 296, 2158–2162.
- 496 Hein, W.R., Pernthaner, A., Piedrafita, D., Meeusen, E.N., 2010. Immune mechanisms of resistance to  
497 gastrointestinal nematode infections in sheep. *Parasite Immunol.* 32, 541–548.
- 498 Hernandez, A.D., Poole, A., Cattadori, I.M., 2013. Climate changes influence free-living stages of soil-  
499 transmitted parasites of European rabbits. *Glob. Change Biol.* 19, 1028–1042.
- 500 Hoar, B.M., Ruckstuhl, K., Kutz, S., 2012. Development and availability of the free-living stages of  
501 *Ostertagia gruehneri*, an abomasal parasite of barren-ground caribou (*Rangifer tarandus*  
502 *groenlandicus*), on the Canadian tundra. *Parasitology* 139, 1093–1100.
- 503 Hsu, C.K., Levine, N.D., 1977. Degree-day concept in development of infective larvae of *Haemonchus*  
504 *contortus* and *Trichostrongylus colubriformis* under constant and cyclic conditions [Nematodes,  
505 sheep]. *Am. J. Vet. Res.*
- 506 IPCC, 2013. Climate Change 2013: The physical Science basis. Contribution of Working Group I to the  
507 Fifth Assessment Report of the Intergovernmental Panel On Climate Change. U. K. Cambridge  
508 University Press Cambridge & N. Y. (USA), Eds.; tech. rep.
- 509 Jenkins, E.J., Veitch, A.M., Kutz, S.J., Hoberg, E.P., Polley, L., 2006. Climate change and the epidemiology  
510 of protostrongylid nematodes in northern ecosystems: *Parelaphostrongylus odocoilei* and  
511 *Protostrongylus stilesi* in Dall's sheep (*Ovis d. dalli*). *Parasitology* 132, 387–401.

- 512 Johnson, P.T.J., Buller, I.D., 2011. Parasite competition hidden by correlated coinfection: using surveys  
513 and experiments to understand parasite interactions. *Ecology* 92, 535–541.  
514 <https://doi.org/10.1890/10-0570.1>
- 515 Kafle, P., Peller, P., Massolo, A., Hoberg, E., Leclerc, L.-M., Tomaselli, M., Kutz, S., 2020. Range expansion  
516 of muskox lungworms track rapid arctic warming: implications for geographic colonization under  
517 climate forcing. *Sci. Rep.* 10, 17323. <https://doi.org/10.1038/s41598-020-74358-5>
- 518 Kenyon, F., Sargison, N.D., Skuce, P.J., Jackson, F., 2009. Sheep helminth parasitic disease in south  
519 eastern Scotland arising as a possible consequence of climate change. *Vet. Parasitol.* 163, 293–  
520 297.
- 521 Kingsolver, J.G., Huey, R.B., 2008. Size, temperature, and fitness: three rules. *Evol. Ecol. Res.* 10, 251–  
522 268.
- 523 Kutz, S.J., Checkley, S., Verocai, G.G., Dumond, M., Hoberg, E.P., Peacock, R., Wu, J.P., Orsel, K., Seegers,  
524 K., Warren, A.L., 2013. Invasion, establishment, and range expansion of two parasitic nematodes  
525 in the Canadian Arctic. *Glob. Change Biol.* 19, 3254–3262.
- 526 Kutz, S.J., Hoberg, E.P., Molnár, P.K., Dobson, A., Verocai, G.G., 2014. A walk on the tundra: host–  
527 parasite interactions in an extreme environment. *Int. J. Parasitol. Parasites Wildl.* 3, 198–208.
- 528 Kutz, S.J., Hoberg, E.P., Nishi, J., Polley, L., 2002. Development of the muskox lungworm,  
529 *Umingmakstrongylus pallikuukensis* (Protostrongylidae), in gastropods in the Arctic. *Can. J. Zool.*  
530 80, 1977–1985.
- 531 Kutz, S.J., Hoberg, E.P., Polley, L., Jenkins, E.J., 2005a. Global warming is changing the dynamics of Arctic  
532 host–parasite systems. *Proc. R. Soc. B Biol. Sci.* 272, 2571–2576.
- 533 Kutz, S.J., Hoberg, E.P., Polley, L., Jenkins, E.J., 2005b. Global warming is changing the dynamics of Arctic  
534 host–parasite systems. *Proc. R. Soc. B Biol. Sci.* 272, 2571–2576.
- 535 Kutz, S.J., Jenkins, E.J., Veitch, A.M., Ducrocq, J., Polley, L., Elkin, B., Lair, S., 2009. The Arctic as a model  
536 for anticipating, preventing, and mitigating climate change impacts on host–parasite  
537 interactions. *Vet. Parasitol.* 163, 217–228.
- 538 Lafferty, K.D., Mordecai, E.A., 2016. The rise and fall of infectious disease in a warmer world.  
539 F1000Research 5.
- 540 Lambert, K.A., Pathak, A.K., Cattadori, I.M., 2015. Does host immunity influence helminth egg  
541 hatchability in the environment? *J. Helminthol.* 89, 446–452.
- 542 Marino, S., Hogue, I.B., Ray, C.J., Kirschner, D.E., 2008. A methodology for performing global uncertainty  
543 and sensitivity analysis in systems biology. *J. Theor. Biol.* 254, 178–196.
- 544 Maya, C., Ortiz, M., Jiménez, B., 2010. Viability of *Ascaris* and other helminth genera non larval eggs in  
545 different conditions of temperature, lime (pH) and humidity. *Water Sci. Technol.* 62, 2616–2624.
- 546 McLeod, R.G., Brewster, J.F., Gumel, A.B., Slonowsky, A., 2006. Sensitivity and uncertainty analyses for a  
547 SARS model with time-varying inputs and outputs. *Math. Biosci. Eng.* 3, 527.
- 548 Mignatti, A., Boag, B., Cattadori, I.M., 2016. Host immunity shapes the impact of climate changes on the  
549 dynamics of parasite infections. *Proc. Natl. Acad. Sci.* 113, 2970–2975.  
550 <https://doi.org/10.1073/pnas.1501193113>
- 551 Molnár, P.K., Dobson, A.P., Kutz, S.J., 2013a. Gimme shelter—the relative sensitivity of parasitic  
552 nematodes with direct and indirect life cycles to climate change. *Glob. Change Biol.* 19, 3291–  
553 3305.
- 554 Molnár, P.K., Kutz, S.J., Hoar, B.M., Dobson, A.P., 2013b. Metabolic approaches to understanding climate  
555 change impacts on seasonal host-macroparasite dynamics. *Ecol. Lett.* 16, 9–21.
- 556 Molnár, P.K., Sckrabulis, J.P., Altman, K.A., Raffel, T.R., 2017. Thermal performance curves and the  
557 metabolic theory of ecology—a practical guide to models and experiments for parasitologists. *J.*  
558 *Parasitol.* 103, 423–439.



- 559 Mordecai, E.A., Caldwell, J.M., Grossman, M.K., Lippi, C.A., Johnson, L.R., Neira, M., Rohr, J.R., Ryan, S.J.,  
560 Savage, V., Shocket, M.S., 2019. Thermal biology of mosquito-borne disease. *Ecol. Lett.* 22,  
561 1690–1708.
- 562 Mordecai, E.A., Cohen, J.M., Evans, M.V., Gudapati, P., Johnson, L.R., Lippi, C.A., Miazgowicz, K.,  
563 Murdock, C.C., Rohr, J.R., Ryan, S.J., 2017. Detecting the impact of temperature on transmission  
564 of Zika, dengue, and chikungunya using mechanistic models. *PLoS Negl. Trop. Dis.* 11, e0005568.
- 565 Paaijmans, K.P., Heinig, R.L., Seliga, R.A., Blanford, J.I., Blanford, S., Murdock, C.C., Thomas, M.B., 2013.  
566 Temperature variation makes ectotherms more sensitive to climate change. *Glob. Change Biol.*  
567 19, 2373–2380.
- 568 Pandey, V.S., Chaer, A., Dakkak, A., 1993. Effect of temperature and relative humidity on survival of eggs  
569 and infective larvae of *Ostertagia circumcincta*. *Vet. Parasitol.* 49, 219–227.
- 570 Prasad, D., 1959. The effects of temperature and humidity on the free-living stages of *Trichostrongylus*  
571 *retortaeformis*. *Can. J. Zool.* 37, 305–316.
- 572 Rose, H., Caminade, C., Bolajoko, M.B., Phelan, P., van Dijk, J., Baylis, M., Williams, D., Morgan, E.R.,  
573 2016. Climate-driven changes to the spatio-temporal distribution of the parasitic nematode,  
574 *Haemonchus contortus*, in sheep in Europe. *Glob. Change Biol.* 22, 1271–1285.
- 575 Rose, H., Wang, T., van Dijk, J., Morgan, E.R., 2015. GLOWORM-FL: a simulation model of the effects of  
576 climate and climate change on the free-living stages of gastro-intestinal nematode parasites of  
577 ruminants. *Ecol. Model.* 297, 232–245.
- 578 Rose Vineer, H., Morgan, E.R., Hertzberg, H., Bartley, D.J., Bosco, A., Charlier, J., Chartier, C., Claerebout,  
579 E., De Waal, T., Hendrickx, G., 2020. Increasing importance of anthelmintic resistance in  
580 European livestock: creation and meta-analysis of an open database. *Parasite* 27, 69.
- 581 Schjetlein, J., Skorpning, A., 1995. The temperature threshold for development of *Elaphostrongylus*  
582 *rangiferi* in the intermediate host: an adaptation to winter survival? *Parasitology* 111, 103–110.
- 583 Short, E.E., Caminade, C., Thomas, B.N., 2017. Climate change contribution to the emergence or re-  
584 emergence of parasitic diseases. *Infect. Dis. Res. Treat.* 10, 1178633617732296.
- 585 Smith, G., 1990. The population biology of the free-living phase of *Haemonchus contortus*. *Parasitology*  
586 101, 309–316.
- 587 Stear, M.J., Bishop, S.C., Doligalska, M., Duncan, J.L., Holmes, P.H., Irvine, J., McCRIE, L., McKELLAR,  
588 Q.A., Sinski, E., Murray, M., 1995. Regulation of egg production, worm burden, worm length and  
589 worm fecundity by host responses in sheep infected with *Ostertagia circumcincta*. *Parasite*  
590 *Immunol.* 17, 643–652. <https://doi.org/10.1111/j.1365-3024.1995.tb01010.x>
- 591 Viney, M., Cable, J., 2011. Macroparasite life histories. *Curr. Biol.* 21, R767–R774.
- 592 Wagner, T., Schliep, E.M., North, J.S., Kundel, H., Custer, C.A., Ruzich, J.K., Hansen, G.J., 2023. Predicting  
593 climate change impacts on poikilotherms using physiologically guided species abundance  
594 models. *Proc. Natl. Acad. Sci.* 120, e2214199120.
- 595 Wakelin, D., 1987. The role of the immune response in helminth population regulation. *Int. J. Parasitol.*  
596 17, 549–557.
- 597