

1 **Optimal cross selection for long-term genetic gain in a two-**  
2 **part genomic selection strategy**

3 **Supplementary Material S1:**  
4 **Additional Material and Methods**

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## 22 **Simulation setup**

23 We initiated simulation by defining a genome with 21 chromosome pairs. Each  
24 chromosome had genetic length of 1.43 Morgans and a physical length of  $8 \times 10^8$  base  
25 pairs. For each chromosome we generated whole chromosome sequences with the  
26 Markovian Coalescent Simulator (Chen et al. 2009). In the simulator we set: i)  
27 recombination rate to  $1.8 \times 10^{-9}$  per base pair (= 1.43 Morgans /  $8 \times 10^8$  base pairs), ii)  
28 mutation rate to  $2 \times 10^{-9}$  per base pair, and iii) effective population size to 50, with linear  
29 piecewise increases to 1,000 at 100 generations ago, 6,000 at 1000 generations ago,  
30 12,000 at 10,000 generations ago, and 32,000 at 100,000 generations ago. These values  
31 were chosen to roughly follow the evolution of effective population size in wheat  
32 (Thuillet et al. 2005; Peng et al. 2011). Finally, we randomly sampled the simulated  
33 chromosomes to establish 50 inbred founder genomes.

34 Out of all segregating variants in founders' sequences we randomly selected  
35 1,000 marker loci per chromosome and 1,000 causal loci per chromosome. Both types  
36 of loci were biallelic. In total there were 21,000 markers and 21,000 causal loci. Each  
37 causal locus was assigned an additive effect from a normal distribution with a mean of  
38 zero and a variance of one divided by the total number of causal loci. The sum of an  
39 individual's causal loci effects represents its genetic merit for a polygenic trait. To  
40 simulate individual's phenotype we added random error to its genetic merit. Errors were  
41 sampled from a normal distribution with mean zero and variance that varied with stages  
42 of a breeding program. Specifically, we varied error variance to obtain phenotypes with  
43 targeted narrow-sense heritability relative to genetic variance among the founders.

44

## 45 **Conventional program with phenotypic selection**

46 The conventional program (Conv) mimicked a wheat breeding program that uses  
47 doubled-haploid technology. In Fig. 1 this program is represented under the product  
48 development pane. We assumed the use of doubled-haploid technology to enable fair  
49 comparison (in terms of cycle time) with the two-part program. Selection was based on  
50 phenotypes, either directly on trial performance or indirectly on correlated traits. The  
51 key steps of this strategy were:

52 Year 1 Cross 50 parental lines to produce 100 bi-parental populations. The crosses  
53 are sampled without replacement from all possible parent combinations.

54 Year 1/2 Produce 100 doubled-haploid lines per bi-parental population

55 Year 3 Plant the 10,000 doubled-haploid lines in headrows. Visually select the best  
56 1,000 lines based on a phenotype with heritability 0.1 (i.e., visual  
57 selection).

58 Year 4 Evaluate the 1,000 lines in a preliminary trial. Select the best 100 lines  
59 based on a phenotype with heritability 0.2 (i.e., unreplicated trial). Advance  
60 the best 20 lines to next year's crossing block.

61 Year 5 Evaluate the 100 lines in an advanced trial. Select the best 10 lines based  
62 on a phenotype with heritability 0.4 (i.e., small multi-location replicated  
63 trial). The 10 lines go to elite trials and next year's crossing block.

64 Year 6/7 Evaluate the 10 lines in an elite trial. Select the best line based on a  
65 phenotype with heritability 0.6 (i.e., large multi-location replicated trial).

66 Year 8 Release variety.

67 We used the conventional program with phenotypic selection as a benchmark and  
68 designed other programs that had approximately equal costs. As in Gaynor et al. (2017)  
69 we assumed that the two dominating costs are creation of double-haploids and  
70 genotyping, which were respectively costed at \$35 and \$20. The conventional program

71 with phenotypic selection with 10,000 doubled-haploid lines had a per year cost  
 72 proportional to \$350,000 (Table S1.1).

73 Table S1.1: Breeding program characteristics (number of crosses, number of doubled-  
 74 haploid lines per cross, total number of doubled-haploid lines, and total cost)

Program	#Crosses	#Lines / cross	#Lines	Cost (\$)
Conv	100	100	10,000	350,000
ConvP	100	95	9,500	352,500
ConvH	100	64	6,400	352,000
TwoPart	/	/	6,100	348,300
product development	100	41	4,100	225,500
population improvement	64	31.2	2,000	122,800

75 <sup>1</sup>Conv – conventional program with phenotypic selection; ConvP/H – conventional program  
 76 with genomic selection at the preliminary trial stage / headrow stage; TwoPart – two-part  
 77 program with genomic selection

78

79 **Conventional program with genomic selection**

80 The conventional program with genomic selection followed closely the conventional  
81 program with phenotypic selection. The difference was high-density genotyping and  
82 genomic selection of lines for trials and next year's crossing block to reduce cycle time  
83 (Fig. 1). We performed genomic selection either at the preliminary trial stage (ConvP)  
84 or at the headrow stage (ConvH). This reduced cycle time for one year with genomic  
85 selection at the preliminary trial stage or for two years with genomic selection at the  
86 headrow stage. Genomic selection increases the total costs in comparison to phenotypic  
87 selection due to high-density genotyping. We equalized the costs by decreasing the  
88 number of doubled-haploid lines per bi-parental population (Table S1.1); to 95 with  
89 genomic selection at the preliminary trial stage (9,500 headrows in total, 1,000 of them  
90 genotyped at the preliminary trial stage) and to 64 with genomic selection at the  
91 headrow stage (6,400 headrows in total, all of them genotyped). The large difference in  
92 number of doubled-haploid lines per bi-parental population was needed due to  
93 genotyping 1,000 lines at the preliminary trial stage and 6,400 lines at the headrow  
94 stage.

95

## 96 **Two-part program with genomic selection**

97 The two-part program (TwoPart) differed from the conventional program in explicit  
98 separation of product development and population improvement (Fig. 1). The  
99 population improvement component is based in a greenhouse that enables several  
100 cycles of recurrent genomic selection per year, while the product development  
101 component is the same as the conventional program with minor modifications  
102 (Table S1.1). We initialized the population improvement component in the last year of  
103 burn-in with a half-diallel cross among the existing parents and another round of  
104 random crossing to avoid large founder effects and to increase number of  
105 recombinations. After the initialization the two components were ready for the year 21.  
106 We have run the two-part program under different scenarios: i) truncation selection  
107 with two numbers of parents or optimal cross selection in the population improvement  
108 component, ii) 1, 2, 3, 4, 5, or 6 cycles of recurrent genomic selection per year, and iii)  
109 constrained or unconstrained costs incurred by high-density genotyping.

110 In the following we first describe: i) a cycle of population improvement with truncation  
111 selection, ii) a cycle of product development, iii) interactions between the two  
112 components, and iv) how we equalized the costs relative to the conventional program.  
113 Then we describe modifications with optimal cross selection, more than one recurrent  
114 selection cycle per year, and lifting the cost constraints.

## 115 **A cycle of population improvement with truncation selection**

116 We assumed that each cross produced 14 seeds out of which 10 were designated for  
117 selection candidates and 4 were designated for production of doubled-haploid lines  
118 (passed to the product development component). With 64 crosses there were  
119 640 selection candidates (Table S1.1 and Table 1). We ranked the candidates based on  
120 genomic prediction and selected the best 32 or 128 as parents of the next cycle  
121 (Table 1), which we respectively denote as TwoPartTS and TwoPartTS+ scenarios; TS  
122 stands for truncation selection and + for a larger number of parents. These two scenarios  
123 respectively correspond to 5% and 20% selected individuals (selection intensity of 2.06  
124 and 1.40). We use these scenarios to demonstrate the effect of selection and drift caused  
125 by the different number of parents. The selected individuals were randomly split into  
126 male and female pools to model potential flowering time differences. Assuming that

127 one wheat plant has four tillers and that we need to produce 64 crosses, we crossed  
128 16 males with 16 females (four crosses per plant) when 32 parents were used or  
129 64 males with 64 females (one cross per plant) when 128 parents were used.

### 130 **A cycle of product development**

131 The product development component was the same as the conventional program with  
132 genomic selection at the headrow stage. The difference was only that both components  
133 produced doubled-haploid lines that were evaluated jointly at the headrow stage of the  
134 product development component (Fig. 1). This represents a likely application of the  
135 two-part program, where a breeder assigns a part of their resources for rapid population  
136 improvement and maintains the conventional strategy with specific lines to design  
137 specific crosses that improve/combine various properties of these specific lines.

### 138 **Interactions between the two components**

139 There were three interactions between the population improvement and product  
140 development components. First, doubled-haploid lines from the population  
141 improvement component were evaluated alongside the product development  
142 component lines at the headrow stage. Main results are shown for this joint group of  
143 individuals to enable comparison with the conventional programs. Second, genomic  
144 predictions in the population improvement component were based on training data  
145 collected in product development trials. Third, at the beginning of each year current  
146 crossing block lines of the product development component were considered as  
147 selection candidates of the population improvement component – in addition to outbred  
148 individuals in the population improvement component. We did this to potentially  
149 include lines with high genetic merit or increase genetic diversity.

### 150 **Costs**

151 The two-part program increased costs due to additional genotyping in the population  
152 improvement component. We equalized the total cost by decreasing the number of  
153 produced doubled-haploid lines (Table S1.1). The number of lines with the two-part  
154 program involved lines from the two components.

155 **Optimal cross selection**

156 Application of the optimal cross selection in the population improvement component  
157 changed selection of parents and their crossing. The method produced a crossing plan  
158 that determined which individuals were selected and how they should be mated to  
159 maximise genetic gain at a predefined loss of diversity. Practically this meant that  
160 between 32 and 128 individuals contributed to 64 crosses, i.e., a selection candidate  
161 could contribute to 0, 1, 2, 3, or 4 crosses. We ran optimal cross selection with a range  
162 of penalties on loss of diversity – operationalized with penalty degrees ( $1^\circ$ ,  $5^\circ$ ,  $10^\circ$ , ...,  
163  $85^\circ$ ; described in the optimal cross selection subsection in the manuscript). We did not  
164 use optimal cross selection in the product development component, because we  
165 assumed that a breeder would design crosses with specific criteria not controlled by  
166 optimal cross selection. However, at the beginning of each year we considered using  
167 the latest crossing block lines from the product development part into the optimal cross  
168 selection for the population improvement component.

169 **Number of recurrent selection cycles per year**

170 We have evaluated the effect of 1, 2, 3, 4, 5, or 6 cycles of recurrent selection per year  
171 assuming that this is possible with the intensive use of greenhouses. Increasing number  
172 of cycles per year increases per year genotyping costs. To account for this increase, we  
173 have scaled numbers of parents, crosses, and selection candidates per cycle such that  
174 the total number of selection candidates per year was approximately constant (640;  
175 Table 1).

176 **Lifting cost constraints**

177 Increasing the number of recurrent selection cycles per year increases the number of  
178 selection candidates per year and through that genotyping costs. In previous two-part  
179 scenarios we have avoided increasing costs by reducing the number of crosses per cycle  
180 (Table 1). We have run all these scenario also without cost constraints, i.e., keeping the  
181 number of crosses per cycle constant (64) irrespective of the number of cycles per year  
182 (Table 1).



183 **References**

- 184 Chen GK, Marjoram P, Wall JD (2009) Fast and flexible simulation of DNA sequence  
185 data. *Genome Res* 19:136–142 . doi: 10.1101/gr.083634.108
- 186 Gaynor RC, Gorjanc G, Bentley AR, et al (2017) A Two-Part Strategy for Using  
187 Genomic Selection to Develop Inbred Lines. *Crop Sci* 57:2372–2386 . doi:  
188 10.2135/cropsci2016.09.0742
- 189 Peng JH, Sun D, Nevo E (2011) Domestication evolution, genetics and genomics in  
190 wheat. *Mol Breed* 28:281–301 . doi: 10.1007/s11032-011-9608-4
- 191 Thuillet A-C, Bataillon T, Poirier S, et al (2005) Estimation of Long-Term Effective  
192 Population Sizes Through the History of Durum Wheat Using Microsatellite  
193 Data. *Genetics* 169:1589–1599 . doi: 10.1534/genetics.104.029553