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Amino acid and carbohydrate tradeoffs by honey bee nectar foragers and their implications for plant-pollinator interactions

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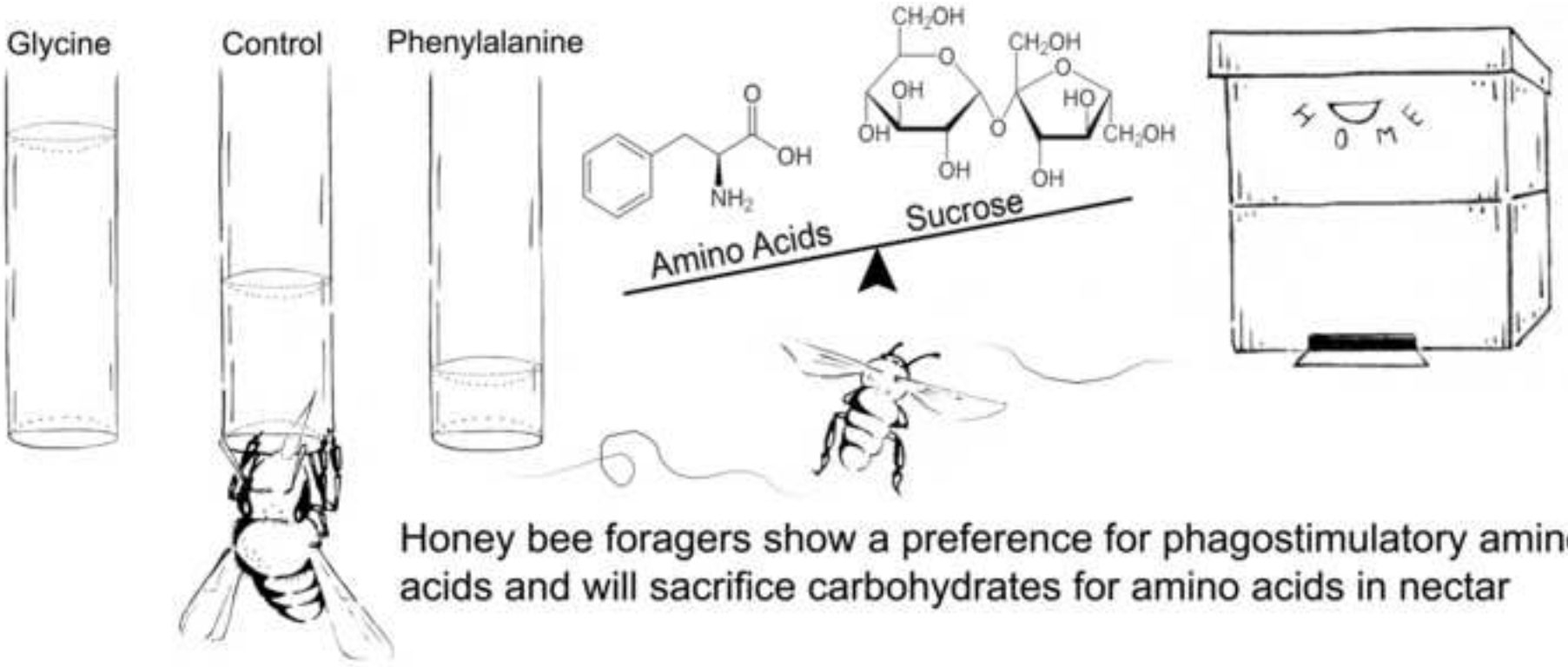
Keywords
pollination ecology / homing / essential amino acids / equivalence point / nutrient balance / pH deviation

Abstract
Honey bees are important pollinators, requiring floral pollen and nectar for nutrition. Nectar is rich in sugars, but contains additional nutrients, including amino acids (AAs). We tested the preferences of free-flying foragers between 20 AAs at 0.1% w/w in sucrose solutions in an artificial meadow. We found consistent preferences amongst AAs, with essential AAs preferred over nonessential AAs. The preference of foragers correlated negatively with AA induced deviations in pH values, as compared to the control. Next, we quantified tradeoffs between attractive and deterrent AAs at the expense of carbohydrates in nectar. Bees were attracted by phenylalanine, willing to give up 84 units sucrose for 1 unit AA. They were deterred by glycine, and adding 100 or more units of sucrose could resolve to offset 1 unit AA. In addition, we tested physiological effects of AA nutrition on forager homing performance. In a no-choice context, caged bees showed indifference to 0.1% proline, leucine, glycine or phenylanaline in sucrose solutions. Furthermore, flight tests gave no indication that AA nutrition affected flight capacity directly. In contrast, low carbohydrate nutrition reduced the performance of bees, with important methodological implications for homing studies that evaluate the effect of substances that may affect imbibition of sugar solution. In conclusion, low AA concentrations in nectar relative to pollen suggest a limited role in bee nutrition. Most of the 20 AAs evoked a neutral to a mild deterrent response in bees, thus it seems unlikely that bees respond to AAs in nectar as a cue to assess nutritional quality. Nonetheless, free choice behavior of foraging bees is influenced, for instance by phenylalanine and glycine. Thus, AAs in nectar may affect plant-pollinator interactions and thereby exhibit a selective pressure on the flora in the honey bee habitat.

*Graphical Abstract (for review)



Honey bee foragers show a preference for phagostimulatory amino acids and will sacrifice carbohydrates for amino acids in nectar

*Highlights (for review)
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HIGHLIGHTS

- Amino acids in artificial nectar elicit preferences from honey bee foragers

- Amino acid identity, pH, and essentiality explain preferences of bees

- A honey bee forager is willing to pay a premium of carbohydrates for amino acids

- Carbohydrate nutritional state affects flight performance of foraging bees

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1. Introduction

The honey bee, *Apis mellifera,* is a key contributor to the pollination of crops worldwide. The nutritional need of honey bee colonies for floral pollen and nectar is the fundamental driver for this valuable ecosystem service. Pollen contains nutrients for growth, such as essential amino acids (AAs) in proteins, certain lipids, and essential trace elements like minerals and vitamins (Free, 1987; Herbert and Shimanuki, 1978). Nectar is the main source of sugars and additionally it provides AAs, lipids, antioxidants, and potentially toxic secondary metabolites (Baker, 1977). The proportion of forager bees for pollen and nectar is determined by the nutritional state of the colony and the availability of floral resources in the environment (Calderone and Johnson, 2002; Camazine, 1993; Seeley, 2009).

It is still debated whether pollen-collecting insects, such as honey bees (Fewell and Winston, 1992; Saa-Otero et al., 2000) and bumblebees (Rasheed and Harder, 1997a, b), maximize foraging efficiency by preferentially switching to protein-rich pollen. Whereas Levin and Bohart (1955) found for five of six pollen types offered at feeding stations that the preference ranked according to crude protein content, Schmidt (1982) found no clear relation between preferences for eight pollens and their protein content. Similarly, choice experiments have shown that honey bees do not assess pollen protein content (Pernal and Currie, 2001, 2002). This is not surprising considering that protein is very rarely present in the pollenkitt, the oily layer coating the pollen grain, and free amino acids were not found in the pollenkitt of any of 69 plants studied (Dobson, 1988). It appears that cues other than protein content affect bee pollen preferences. Such cues may include odors, phagostimulants and phagodeterrents (Schmidt, 1982), grain size (Pernal and Currie, 2002), and pollen concentration; honey bees dance more rigorously to a feeder containing a higher pollen-to-cellulose ratio (Waddington, 2001; Waddington et al., 1998).

In contrast, far more is known on the ability of forager bees to evaluate the nutritional value of nectar. The main total dissolved solids (TDS) in nectar are sugars,

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and bees can readily determine sugar content. They can discriminate small differences in sugar concentration (Afik et al., 2006; Frisch, 1967; Shafir et al., 2008), and nectar volume and variability (Shafir et al., 2005; Waddington and Gottlieb, 1990). Honey bees are able to perceive and show preference for nectar mineral contents, secondary compounds and certain AAs (Afik et al., 2006; Carter et al., 2006; Cook et al., 2003; Gardener and Gillman, 2002; Kim and Smith, 2000; Singaravelan et al., 2005). The means of sensing nectar composition are receptor based, with different kinds of taste receptors on the proboscis and mouth parts (Goodman, 2003; Sanchez, 2011). It is unknown however, to what extent amino acid (AA) presence in nectar is valued, and further, if potential preferences are related to the nutritional state of the colony. Whereas pollen is the main source of AAs for bees, we cannot exclude the possibility that deficiencies in particular AAs in the colony may also modulate preference for nectars which contain those AAs.

A colony needs a minimum level of essential AAs to assimilate proteins for growth (De Groot, 1953). Hence, a preference for essential over non-essential AAs may be an adaptive strategy for bees. To date however, a general preference for essential AAs has not been found, though certain individual AAs are known to evoke phagostimulant or phagodeterrant effects on bees (Inouye and Waller, 1984). Honey bee studies have tested AAs singly, e.g. on consumptive responses (Inouye and Waller, 1984) or on olfactory perception (Linander et al., 2012). However, in the present study, we offered all AAs simultaneously in order to compare AA preferences in relation to one another. This may reveal insights in how honey bees in a floral habitat respond to a complexity of choice options. An AA-induced gustatory preference may influence plant pollinator interactions, which can have ecological and evolutionary consequences by affecting gene flow within and between plant populations (Gardener and Gillman, 2002; Gottsberger et al., 1984; Nepi et al., 2012). In addition to direct preference effects, AAs may induce more subtle plant pollinator interactions. AA levels in bees may stimulate or inhibit learning and memory (Chalisova et al., 2011), thereby influencing flower

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association by free flying foragers, as has been shown for nicotine in nectar (Wright et al., 2013).

Forager preferences for AAs may come with benefits. Proline for example, has been described as a flight muscle stimulant (Barker and Lehner, 1972; Carter et al., 2006; Micheu et al., 2000; Mollaei et al., 2013). At the same time, a preference may come at a cost when the perceived profitability exceeds the benefit. This would be the case if uptake due to phagostimulating AAs were in lieu of carbohydrate uptake.

If foragers alter their intake of carbohydrates, they might pay a physiological price, either in terms of fuel for flight, or their flight ability could change, for instance due to alterations in their carrying load. By means of homing experiments, we explored whether AAs in nectar elicit physiological effects on forager bees. In particular, we compared the indirect effect of phagostimulant and phagodeterent AAs on sucrose uptake, consequent nutritional state of the bee, and its effect on homing success and flight speed.

An additional aim of our study was to test the relative preferences of free flying nectar foragers between simultaneously presented AA-enriched sucrose feeders. Since AAs in artificial nectar may affect forager choice, we tested for a tradeoff between AAs and carbohydrate in collected nectar. We quantified evoked responses by comparing units of AAs to equivalent units of sugar, thereby calculating the energetic cost of choice.

2. Materials and Methods

The experiments were conducted at the Benjamin Triwaks Bee Research Center in Rehovot, Israel, with the local line based mostly on the Italian honey bee strain *Apis mellifera ligustica*.

2.1. Preference between nonessential and essential amino acids

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In March, October and November 2013, bees were trained to forage on ten 200 ml feeders filled with honey-tainted sucrose solutions (30% w/w), hanging at fences in the vicinity of two of our apiaries. At any time, 20 to 30 colonies of free-flying bees were present. Once foragers were visiting the fence sites, a few dozen 2 ml glass tubes with one open end were filled with 20% sucrose solutions and hung on 2.5 x 2 m fence sections to train the bees to visit the tubes. The feeding tubes together on the fence formed an artificial meadow to conduct experiments. In order to unravel the complexity of nectar foragers choice patterns, this novel test approach allows testing a wide range of treatments.

To test honey bee forager preferences among AA-enriched artificial nectars, a series of choice experiments was started by hanging new tubes with AA solutions in the fence sections. Each AA treatment was presented in 1.8 ml 20% sucrose solutions, enriched with an AA at 0.1% (w/w). Abbreviations for amino acid names used throughout the manuscript are listed in Supplement 2. Depending on molecular weights of each AA, treatment solutions were in range of 5.3 – 14.3 mM (Supplement 2). The solutions were always offered in presence of a control (20% sucrose solution). Considering a crop volume of maximally 70 µL, it takes at least 25 bee visits to empty a tube with a volume of 1.8 ml of liquid.

In three experimental designs, we tested 8 nonessential AAs (72 test tubes offered to bees in 8 replicate fence sections), 10 essential AAs (88 test tubes in 8 replicate sections), and 20 AAs together (542 feeding tubes in 25 replicate sections). The tubes were hung according a Latin square design, so that each treatment appeared in a different spatial position in every replicate fence section. The experiment with all the AAs was replicated at March 13 and 15, and November 16, at two apiaries. Each apiary contained 20-30 hives, which were exchanged from time to time. Additional details of the experimental designs are given in the supplementary material (Supplement 1: S1-a, S1-b and S1-c). The pH values of the solutions were measured for each batch of test solutions (Supplement 2).

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154 The fluid levels in the test tubes were marked at the meniscus at the start and
155 end of the experiment. As soon as the first tubes in a section were nearly or completely
156 empty, all tubes in that section were turned upwards, so that the content was no longer
157 available to the bees. We then measured the column height of the collected solution in
158 each glass tube to the nearest mm. For every AA treatment, proportional consumption
159 values were calculated on the total solution collected per replicate fence section.
160
161 **2.2. Tradeoff between carbohydrates and amino acids**
162
163 To quantify the tradeoff between sucrose and amino acids, glass tubes were hung at 12
164 replicate fence sections, offering 12 AA treatments to free flying foragers. The test was
165 conducted as a 12x12 Latin square design (Supplement 1: S1-d). Treatments were
166 control solutions of 15, 18, 21, and 24% sucrose (w/w), solutions of 15, 18, 21, and 24%
167 sucrose with 0.1% phenylalanine (Phe), and solutions of 15, 18, 21, and 24% sucrose
168 with 0.1% glycine (Gly). Based on the findings from the AA-choice experiment (see 2.1.),
169 phenylalanine was chosen to represent a phagostimulant, and glycine to represent a
170 phagodeterrent. The phenylalanine and glycine treatments had pH values similar to the
171 control ($pH_{Control}$ = 6.15, pH_{Phe} = 6.23, pH_{Gly} = 6.28). Data were collected on the
172 proportional consumption of treatment solutions per fence section. The data were then
173 used to extrapolate sucrose tradeoff levels where a 0.1% AA solution would be equally
174 collected compared with a sucrose control.
175 We tested these extrapolated values in two additional experiments. In one
176 experiment, there were three treatments: a 20% sucrose control, a lower sucrose
177 concentration with 0.1% phenylalanine, and a higher sucrose concentration with 0.1%
178 glycine. Throughout 9 fence sections, in which each solution was represented twice, 18
179 replicates of each solution were presented to free-flying bees (Supplement S4). The
180 results confirmed the equivalence sucrose concentration for phenylalanine, but
181 suggested a higher sucrose equivalence concentration for glycine. In a follow-up

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182 experiment, we presented four treatments: a 20% sucrose control, and 20%, 30% and
183 40% sucrose solutions each enriched with 0.1% glycine (Supplement S4).
184
185 **2.3. The effect of amino acid and carbohydrate consumption on homing success**
186
187 To study AA and sucrose nutrition effects on foragers, three homing tests were
188 performed (H1, H2 and H3). All test individuals were homecoming foragers that were
189 collected by closing the entrance to a test hive. Captured bees were placed on ice for
190 several minutes until they stopped moving, marked on their thorax with a color to
191 indicate the treatment they would receive, and placed in groups of 15 or 20 bees in a
192 clear plastic jar with a mesh opening on the side for ventilation. Over the three homing
193 experiments, a total of five hives were tested, with a total of 2079 bees placed in 130
194 clear plastic jars (details in Supplement 3).
195 Every jar received two glass tubes through holes in the lid, each containing 1.8
196 ml of test solution. The jars were then placed in an empty hivebox, on top of a
197 populated hivebox. The two boxes were separated by means of a queen excluder
198 covered with a 1 mm^2 mesh. This allowed the bees in the jars above to feel the warmth
199 and scent of the colony below. The levels of test solution were marked on the tubes as
200 soon as they were inserted into the jars. The levels were marked twice a day, in parallel
201 to noting the number of dead bees per jar, to determine how many individuals
202 contributed to consuming the treatment solutions over time, and to monitor for
203 treatment-related mortality effects.
204 On the day of the homing experiment, bees were released forenoon
205 approximately 600 m from the hive. Bees were released onto an arena. During the thirty
206 minutes after release, moribund and dead individuals were collected and counted. The
207 “dead” bees included those that died during the pre-phase of the experiment, whereas
208 “moribund” bees were those that were not able to fly away from the arena within thirty
209 minutes. Experimenters back at the test hives were notified when the bees were
210 released. The hive entrances were closed and every homecoming marked bee was

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captured, in order to prevent recounts, over a three hour time period. The return time and color for every test bee was recorded. After each experiment, the test hives were opened to verify the absence of marked bees.

During the first homing experiment (H1), the treatments were 40% sucrose (1), and 40% sucrose enriched by 0.1% phenylalanine (2), 0.1% leucine (3), 0.1% proline (4), or 0.1% glycine (5). Alike the fence experiments, these particular amino acids were chosen to represent phagostimulants, phagodeterrents, nonessential, and essential amino acids as well as being pH neutral in comparison to the control. Fifteen bees were kept in each plastic jar (n_{jars}=70), for a total of 1050 bees from two colonies (Supplement 3). To test for possible effect of foraging task specialization, foragers returning with pollen loads (n_P=300) were collected from one hive and foragers returning without pollen loads (n_N=300) from another. We presume that most bees returning without pollen loads were nectar foragers, and refer to them as such, though we did not examine their crop content (few may be water collectors or returning with empty crops). We conducted a replicate trial at a different date, in which the type of forager collected from each hive was switched (n_P=225, and n_N=225). The total imbibition time before release in the homing experiment was 36 h.

The second homing experiment (H2) was performed with three carbohydrate treatments at different concentrations: 8, 16, and 32% sucrose (w/w). Test bees were caught over two experimental days, again distinguishing pollen foragers from non-pollen foragers on two trial dates (Supplement 3). Twenty bees were kept per jar (n_{jars}=28), with a total of 549 marked bees from two colonies. Time until release was shortened to 16 h, to avoid mortality of bees fed the low concentration of sucrose solution.

A third homing experiment (H3) was completed with four treatments: a control solution of 20% sucrose (1), or 20% sucrose enriched with 0.1% phenylalanine (2), 0.1% glycine (3), or 0.1% proline (4). With a total of 480 marked bees, fifteen individuals were kept per jar (n_{jars}=32), and incubated for 20 h on top of two colonies. Each treatment was replicated eight times, including a duplication by separately testing pollen and non-

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pollen foragers, and the use of two colonies, others than used for H1 and H2 (Supplement 3). In H3 we tested a 20% sucrose solution, in contrast to 40% sucrose in H1. First, because amino acids taste might be masked at 40% sucrose, and second, to test within a concentration range where the carbohydrate nutrition can induce a limitation effect (see H2; survival or flight performance).

2.4. Data analysis

Statistics were performed with the JMP Pro software, version 10, SAS Institute Inc. Analyses were performed with general linear models, with a visual inspection of the standardized residuals to evaluate the assumptions of appropriate normality of the response variable, and normally distributed errors with homogeneous variance. In addition, parametric survival analyses (Weibull) were used to investigate flight times in the homing tests.

Forager preferences for AAs were analyzed according to the proportional consumption data by means of an Anova in which AA identity was entered as a factor with 9, 11 or 21 levels (including the controls), for nonessential, essential and all individual AAs, respectively. A post hoc Dunnett test was used to compare choice for each AA solution with the control. A potential seasonal effect on preference was also considered by testing the interaction between AAs as a factor, and the two test months March or October as a factor; this interaction was removed from the model when it was not significant. Choice among AA solutions was analyzed a second time, on AA essentiality (a 2 leveled factor; 10 essential versus 10 non-essential AAs per fence section) together with the deviation in pH value from the control (a continuous variable). The control values were excluded in this second analysis as the pH deviation was calculated relative to the control, and since the control solutions did not contain an essential or non-essential AA. Due to the double analyses on the source data, we applied a Bonferroni-correction and set the threshold for significance at $\alpha=0.025$.

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For the assessment of the tradeoff between AAs and carbohydrates, regressions were performed on the proportions of collected treatment solutions per fence section, considering a sucrose concentration gradient (a continuous variable of 15-24% sucrose solutions) and including the AA treatments (a 3 leveled factor: phenylalanine, glycine, control), with a post hoc Dunnett test to indicate differences between the two AAs and the control. The regression line equations allowed the extrapolation of a sucrose concentration equivalence point, which gave estimated equal consumptions of phenylalanine and glycine treatments, as compared to a control solution of 20% sucrose.

Hereafter, the phenylalanine and glycine treatment solutions, as previously extrapolated, were experimentally tested in a new artificial meadow set-up (Supplement 4), and the proportions of foraged treatment solutions per fence section were compared by means of an Anova with a Post Hoc Dunnett test, comparing the 0.1% phenylalanine and glycine solutions to the control.

For the homing experiments H1, H2 and H3, we used a factorial design, considering four factors: treatment, forager type, trial date and colony (Supplement 3). The two tested continuous response variables were the amount of test solution imbibed per bee, and the mortality during the imbibition period. The analyses were performed on mean values per replicate jar, applying post hoc Tukey tests to differentiate between levels. The return rates in homing flight tests of bees were compared by means of Chi-square tests on the numbers of returning bees versus non-returning bees, with one test for each of the factors mentioned above. The flight times in experiments were compared using the bee return data, employing survival analyses, once for every factor. P-value significance was considered after Bonferroni corrections for multiple comparisons.

3. Results

3.1. Preferences between amino acids

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When tubes with 20% (w/w) sucrose solution enriched with individual AAs were offered in an artificial meadow, nectar foragers showed significant choice differences amongst the 8 nonessential AA enrichments with a control ($F_{8,63}$=8.16, P<0.001), amongst the 10 essential AA enrichments with a control ($F_{10,77}$=3.97, P<0.001), and also between all 20 AA enrichments with a control ($F_{20,472}$=8.94, P<0.001) (Fig. 1A). A consistency in the order of AA choice is indicated as the consumed proportions of the 10 essential AAs, when tested alone, strongly correlated to those when all 20 AAs were tested together (regression on proportion results of AAs between tests; r^2=0.74, $F_{1,9}$=25.5, P<0.001). Similarly, the results on the 8 non-essential AAs alone strongly correlated to those while testing all AAs together (r^2=0.82, $F_{1,7}$=31.2, P<0.001). The choice among AAs was consistent between the test months March and October (interaction AA and Months: $F_{40,434}$=1.22, P=0.24). A post hoc comparison indicated that glutamate, aspartate, histidine, cysteine and glycine were significantly disliked compared to the control (Fig. 1). Foragers preferred phenylalanine the most, although not significantly more than the control.

A significant preference was found for essential AA's over non-essential AA's ($F_{1,465}$=12.17, P<0.001), and at the same time, the AA induced deviations in pH (absolute value relative to the control) were a significant cause for dislike ($F_{1,465}$=28.54, P<0.001) (Fig. 1B). Overall, the acidity of the test solutions ranged between pH=2.26 (0.1% cysteine) and pH=9.33 (0.1% arginine) (Supplement 2).

3.2. Tradeoff between carbohydrates and amino acids

When the range of 15% to 24% sucrose solutions were offered to foragers in an artificial meadow (Fig. 2), they showed preferences for higher sucrose concentrations ($F_{1,140}$= 8.05, P=0.005), and simultaneously phenylalanine was a significant phagostimulant, and glycine a phago-deterrent ($F_{2,140}$=30.9, P<0.001; with posthoc significances for both glycine < control and phenylalanine > control). In Figure 2, the control regression line [y

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= 0.019x + 0.175] at concentration 20% indicates a consumed proportion of 0.549. At this level, the 0.1% phenylalanine regression line [y = 0.012x + 0.425] is at a concentration of 10.6% sucrose, hence at +Δ9.4% sucrose equivalence. At the same level, the 0.1% glycine regression line [y = 0.026x - 0.273] is at 31.4% sucrose, hence a -Δ11.4% sucrose equivalence. The equivalence extrapolation for phenylalanine reveals an expected AA to carbohydrate ratio of 1/94 parts w/w, and a ratio of 1/114 parts w/w for glycine.

In two additional experiments we compensated AA presence with sucrose in order to offset the attraction and deterrence of phenylalanine and glycine, respectively (Supplement 4). In the first experiment, foragers collected dissimilar proportions of test solutions ($F_{2,51}$=10.1, P<0.001, power=0.98 at α=0.05). An 11.6% sucrose solution with 0.1% phenylalanine was collected similar to the 20% sucrose control (P=0.29), whereas a 31.7% sucrose solution with 0.1% phenylalanine was collected significantly less (P<0.001). We therefore conducted a follow-up experiment to test sucrose compensation for the deterrent effect of glycine. Solutions were again dissimilarly collected ($F_{3,60}$=25.6, P<0.001, power=1.00 at α=0.05), with glycine reconfirmed to be unattractive to foragers (20% sucrose + 0.1% glycine) as compared to the control (P=0.026). However, in this experiment the glycine effect was offset by +Δ10% sucrose (P=0.08), whereas glycine +Δ20% sucrose was significantly more collected than the control (P<0.001). Thus, in line with our extrapolations, we could confirm the AA to carbohydrate ratio (1 part phenylalanine ≈ 84 parts sucrose, and 100 parts sucrose ≤ 1 part glycine < 200 parts sucrose).

3.3. The effect of amino acids and carbohydrates on homing success

The results of the homing test are summarized in Tables 1, 2 and 3. The addition of 0.1% AAs in sucrose solutions did not evoke effects on treatment solution imbibition (Fig. 3), mortality and return rates (Fig. 4), or flight times (Fig. 5), either at 40% (H1) or 20% (H3)

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354 sucrose solutions. Interactions between AA treatment and other factors were not
355 indicated.

356 Forager type effects were found, with nectar foragers imbibing less sucrose
357 solutions, and having a lower mortality in H1, though this finding was not present in H2
358 and H3. Exclusively in H2, pollen foragers returned a notable 10 minutes earlier than
359 nectar foragers (P: 25 min < N: 35 min; though insignificant after Bonferroni correction).
360 Colony effects were found throughout the homing experiments, on sucrose uptake and
361 bee mortality (between colonies A&B and D&E), return rates (between A&B and B&C),
362 and flight times (between B&C).

363 The test on the effect of carbohydrate nutrition on forager homing performance
364 (H2) showed that a relatively low volume per bee was consumed at the lowest
365 carbohydrate concentration. A significant increase in sucrose uptake in µL and mg was
366 observed over the sucrose gradient (P=0.01 and P<0.001, respectively, Fig. 3B). A high
367 sucrose uptake in mg (Fig 3.) correlated with a reduction in mortality (Linear regression;
368 P=0.01, R^2=0.22), with 66, 17, and 1 deaths at 8%, 16% and 32% sucrose, respectively
369 (Fig. 4-H2). The return rates of bees were not different for the three carbohydrate
370 treatments. However, the median flight time of 14.5 minutes at the highest sucrose
371 concentration was significantly shorter than the 35 and 36.5 minutes median it took
372 those that received the lower sucrose concentrations (Table 2, Fig. 5B).

373

374 **4. Discussion**

375

376 As main results, we found consistent preferences of free-flying honey bee foragers for
377 certain AAs over others, and for essential over non-essential AAs (Fig. 1). Furthermore,
378 we found that bees were willing to pay a considerable carbohydrate premium for their
379 AA preferences (Fig. 2). Bees were significantly attracted by 0.1% enriched
380 phenylalanine solutions, as compared to the controls, and willing to give up 84 units
381 sucrose for 1 unit AA. The bees were significantly deterred by glycine, but adding a 100
382 or more units of sucrose could resolve to offset the effect of 1 unit AA.

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Roubik et al. (1995a) found the stingless bee *Melipona fuliginosa* to avoid glutamate, glycine, serine, alanine, and arginine at concentrations of 35 to 80 mM AA in 50% sucrose solutions (≈ 0.6% w/w on average), and *M. fuliginosa* valued the deterrence equal to control solutions of 20 to 40% sucrose (–Δ10% to –Δ30% sucrose). It reveals a ratio in range of 1/17 to 1/50 parts AA to carbohydrate to offset deterrence. This is less expensive than for honey bees, considering their glycine deterrence compensation with 100 or more parts sucrose.

The premiums in sucrose that bees are willing to pay for AA can be biologically significant; mortality of bees fed an 8% sucrose solution was greater than those fed 16 or 32% sucrose (Fig. 4), and it took those that survived twice as long to home back to the hive (Fig. 5B). For a honey bee colony, the caloric uptake of carbohydrates is lowered when foragers systematically prefer low sucrose nectars with phagostimulant AAs, or evade high sucrose nectars with deterrent AAs. From the plant perspective, plants can substitute expensive carbohydrates in their nectar with minute concentrations of phagostimulating AAs, or modulate pollinator visits by adding phagodeterrent AAs.

An artificial meadow assay allowed us to test the preference of free flying foragers between 20 AAs that were simultaneously present, and we found a preference for essential over non-essential AAs. Insect research has shown that an animal's sensitivity to taste can change according to the need for nutrients due to the under-representation of nutrients evoking over-sensitivity on the receptor level (Abisgold and Simpson, 1987; Simmonds et al., 1992; Simpson et al., 1991). Perhaps, the current finding that essential AAs are more preferred than nonessentials, might relate to the yet unknown ability of forager bees to respond to specific nutritional shortcomings within their colony. We found a consistent AA preference profile between colonies at the beginning of spring growth (March) and autumn (October), but we did not specifically manipulate or assess colony needs in this study. When testing honey bees in a no-choice assay with single AAs, Inouye and Waller (1984) did not find preference effects due to essentiality of AAs. Similarly, in dual choice preference tests with *Drosophila,* Toshima

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411 and Tanimura (2012) found AA preferences unrelated to the classification of essential or
412 non-essential, nor to other chemical properties.

413 The natural pH of floral nectar is reported to range between 4.2 and 8.5 (Baker,
414 1977). The pH of the dissolved purified AAs at 0.1% (w/w) solutions were between 2.3
415 and 9.3, which lies beyond the natural range in nectar (Supplement 2). We found that
416 0.1% AA could affect nectar pH, and that foragers were sensitive to the pH (Fig. 1B). The
417 statistical model showed independent effects on bee preferences of whether AAs were
418 essential or not and of their pH. Furthermore, the pH of glycine and phenylalanine
419 enriched solutions were similar, yet the former was consistently disliked by the bees
420 whereas the latter was consistently preferred (Fig. 2). Thus, pH affects forager choice
421 but only partially explains preferences between AAs. The pH of the resulting solution
422 should always be considered in taste or preference studies that dissolve purified AAs in
423 solution.

424 In our study, bees could use both olfaction and gustation to discriminate
425 between AA solutions. Linander et al. (2012) showed that bees could discriminate
426 between the odor of single dissolved AAs and the control solvent, specifically aspartate,
427 cysteine, proline, tryptophan, and tyrosine. A conversion by molecular weight indicates
428 our AA concentrations in range of 5.3 – 14.3 mM (supplement 2). This is one order of
429 magnitude smaller than the test concentration of 100 mM as used by Linander et al.
430 (2012). Nonetheless, the discrimination between cysteine and aspartate from the
431 control in our experiments (Fig. 1) could thus have been facilitated by olfactory cues. We
432 found that free-flying foragers could also discriminate phenylalanine and glycine from
433 the control (Fig. 2). This discrimination may have relied on gustation. Although bees are
434 able to discriminate proline, tryptophan and tyrosine from the control by scent at
435 100mM (Linander et al., 2012), we did not find preference effects for these AAs at the
436 tested concentration of 0.1% (<10mM).

437 Honey bees show a significant phagostimulatory effect of phenylalanine and a
438 phagodeterrent effect of glycine (Fig. 2), which is consistent with the findings of Inouye
439 and Waller (1984). However, considering the results in Figure 1, the choice among AAs

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for which bees did not show extreme preferences was not always consistent with other studies. For example, proline at concentrations similar to those that we studied, has generally been reported to be attractive to bees (Bertazzini et al., 2010; Carter et al., 2006; Inouye and Waller, 1984). Similarly, bees in our study favored alanine, but not significantly more than the control; Bertazzini et al. (2010) and Inouye and Waller (1984) found alanine to be significantly preferred to the control. Different preferences of bees between studies may be due to methodology (e.g., choice vs no-choice, free-flying vs caged), or differences in sensitivity thresholds of different honey bee strains. Such inconsistencies are more likely to show for AAs that evoke less pronounced preferences; AAs at the extreme ends seem to be valued more consistently.

Of course, preference and deterrence effects are also dependent on AA concentrations. However, AAs in nectar vary greatly within plants and between species within several orders of magnitude (Baker, 1977; Nepi et al., 2012; Weiner et al., 2010). Therefore, we chose to standardize our experimental setup, and test all AAs at the same concentration of 0.1% (w/w), which lies within the higher boundary of the natural range. It is for instance comparable to *Lantana camara* nectar, with 16 mM total AA (an approximate 0.1% w/w); a plant visited by bees and many butterflies and described to have a relative high amino acid concentration (Alm et al., 1990).

In addition to honey bees, AA preferences are reported for tropical stingless bees (Gardener et al., 2003; Roubik et al., 1995b), solitary bees (Weiner et al., 2010), butterflies (Erhardt and Rusterholz, 1998), ants (Bluthgen and Fiedler, 2004a, b; Wada et al., 2001), flies (Potter and Bertin, 1988), and nectivorous bats (Rodriguez-Pena et al., 2013). Differences in taste perception between diverse pollinators likely affect nectar perception and may be a factor in how plants bias pollinator visits, thereby affecting gene flow within and amongst plant populations (Nepi et al., 2012).

AA concentrations in nectar are low relative to pollen, thereby suggesting a limited role in bee nutrition. For pollen, it has been described that plants visited by oligolectic insects have a lower pollen quality, for instance in the ratio of essential to non-essential AAs, as compared to plants visited only by non-oligolectic insects (Weiner

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et al., 2010). The noted pollen quality differentiation was however not reflected in the free amino acids content of the pollens. Thus, since free amino acids are the most likely cue for a potential taste-based evaluation of AA nutrition, it is not likely that pollinators respond to nutritional quality based on AA taste alone. Also, most of the 20 AAs tested in the current study evoked neutral to mild deterrent responses in honey bees, thus it seems unlikely that bees respond to AAs in nectar as a cue to assess nutritional quality.

A substantial question within the current study addressed if AAs in nectar may affect the physiology and performance of bees. As mentioned, proline may be directly used as a minor source of flight fuel (Barker and Lehner, 1972; Carter et al., 2006; Micheu et al., 2000; Mollaei et al., 2013). However, the study by Mollaei et al. (2013) showed no notable honey bee flight muscle stimulation at 0.1% proline treatment. In our homing studies proline at 0.1% did also not evoke an effect, neither on the return rate, nor the median flight time of bees (Table 1 and 3).

In contrast to the consistent preferences exhibited by free-flying bees, caged foragers in a no-choice context imbibed equally from sucrose solution or sucrose solution enriched with AAs (Fig. 3). An dietary enrichment with either Phenylalanine, Glycine, Leucine or Proline did not affect their homing performance (Fig. 5A) or other physiological measures of performance (Table 1 and 3). However, physiology and performance were directly influenced by carbohydrate nutrition (H2): foragers fed with lower sucrose concentrations took longer to come back to the hive (Fig. 4). This can have important methodological implications for homing studies that evaluate the effect of substances that may affect the imbibition of sugar solution.

Forager task specialization affects various physiological and behavioral traits (Nelson et al., 2007; Page, 2013; Robinson and Vargo, 1997). In one of our homing tests (H1) pollen foragers imbibed more test solution than non-pollen foragers (Table 1). Homing ability is an effective assay in eco-toxicological tests, and the amount of test solution imbibed would affect exposure to toxicants. Our study suggests that controlling for task specialization could reduce experimental variability.

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We have presented a novel and useful bioassay by means of an artificial meadow set-up. We illustrate high test sensitivity, in which steps of only Δ3% sucrose concentration show a clear gradient effect, in addition to detecting an AA treatment effect (Fig. 2). This simple setup could be useful for tackling additional questions about bee perception of nectar components. Specifically for AAs, we tested them singly, but Alm et al. (1990) found that honey bees consumed more and longer from nectar sources which were enriched with a mixture of AAs, compared to nectar without AAs. This supports the hypothesis that the AAs in nectar contribute to pollinator attraction. Higher stimulatory effects of mixtures of AAs over single AAs have been frequently reported also for ants (Lanza, 1988; Lanza and Krauss, 1984). The artificial meadow set-up would be useful for further research into the complexity of AA attraction of mixtures compared to single AAs.

In conclusion, we found three variables that explain observed AA preferences of honey bee nectar foragers: AA identity, pH, and essentiality. We identified sucrose equivalents that foragers are willing to exchange for phagostimulant or phagodeterrent AAs. Flight performance of foraging bees was affected by nutritional state: foragers were directly affected by carbohydrate levels, but not directly by either proline, leucine, phenylalanine or glycine, nor indirectly by their excitatory or deterrent tastes. Both pH and AAs were detected by the bees in artificial nectar at natural concentrations. Thus, such preferences by pollinators could affect floral evolution, shaping the biodiversity in natural ecosystems, and could affect pollination services in agricultural landscapes.

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References

Abisgold, J.D., Simpson, S.J., 1987. The Physiology of Compensation by Locusts for Changes in Dietary-Protein. Journal of Experimental Biology 129, 329-346.

Afik, O., Dag, A., Kerem, Z., Shafir, S., 2006. Analyses of avocado (Persea americana) nectar properties and their perception by honey bees (Apis mellifera). Journal of Chemical Ecology 32, 1949-1963.

Alm, J., Ohmeiss, T.E., Lanza, J., Vriesenga, L., 1990. Preference of Cabbage White Butterflies and Honey-Bees for Nectar That Contains Amino-Acids. Oecologia 84, 53-57.

Baker, H.G., 1977. Non-Sugar Chemical Constituents of Nectar. Apidologie 8, 349-356.

Barker, R.J., Lehner, Y., 1972. Free Amino-Acids in Thoraces of Flown Honey Bees, Apis-Mellifera L (Hymenoptera-Apidae). Comparative Biochemistry and Physiology 43, 163-&.

Bertazzini, M., Medrzycki, P., Bortolotti, L., Maistrello, L., Forlani, G., 2010. Amino acid content and nectar choice by forager honeybees (Apis mellifera L.). Amino Acids 39, 315-318.

Bluthgen, N., Fiedler, K., 2004a. Competition for composition: Lessons from nectar-feeding ant communities. Ecology 85, 1479-1485.

Bluthgen, N., Fiedler, K., 2004b. Preferences for sugars and amino acids and their conditionality in a diverse nectar-feeding ant community. Journal of Animal Ecology 73, 155-166.

Calderone, N.W., Johnson, B.R., 2002. The within-nest behaviour of honeybee pollen foragers in colonies with a high or low need for pollen. Animal Behaviour 63, 749-758.

Camazine, S., 1993. The Regulation of Pollen Foraging by Honey Bees: How Foragers Assess the Colony's Need for Pollen. Behavioral Ecology and Sociobiology 32, 265-272.

Carter, C., Shafir, S., Yehonatan, L., Palmer, R.G., Thornburg, R., 2006. A novel role for proline in plant floral nectars. Naturwissenschaften 93, 72-79.

Chalisova, N.I., Kamyshev, N.G., Lopatina, N.G., Kontsevaya, E.A., Urtieva, S.A., Urtieva, T.A., 2011. Effect of Encoded Amino Acids on Associative Learning of Honeybee Apis mellifera. Journal of Evolutionary Biochemistry and Physiology 47, 607-610.

Cook, S.M., Awmack, C.S., Murray, D.A., Williams, I.H., 2003. Are honey bees' foraging preferences affected by pollen amino acid composition? Ecological Entomology 28, 622-627.

De Groot, A.P., 1953. Protein and amino acid requirements of the honeybee (Apis mellifica L.), Physiol. Comp. Oecologia, pp. 197-285.

Dobson, H.E.M., 1988. Survey of Pollen and Pollenkitt Lipids - Chemical Cues to Flower Visitors. American Journal of Botany 75, 170-182.

Erhardt, A., Rusterholz, H.P., 1998. Do Peacock butterflies (Inachis io L.) detect and prefer nectar amino acids and other nitrogenous compounds? Oecologia 117, 536-542.

Fewell, J.H., Winston, M.L., 1992. Colony State and Regulation of Pollen Foraging in the Honey-Bee, Apis-Mellifera L. Behavioral Ecology and Sociobiology 30, 387-393.

Free, J.B., 1987. The Biology of the Honey-Bee - Winston,Ml. Science 238, 1591-1592.

Frisch, K.v., 1967. Dance language and orientation of bees. Cambridge, Mass., Belknap Press of Harvard University Press.

Gardener, M.C., Gillman, M.P., 2002. The taste of nectar - a neglected area of pollination ecology. Oikos 98, 552-557.

Gardener, M.C., Rowe, R.J., Gillman, M.P., 2003. Tropical Bees (Trigona hockingsi) Show No Preference for Nectar with Amino Acids. Biotropica 35, 119-125.

Goodman, L., 2003. Form and function in the honey bee. . Cardiff: The Westdale Press.

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572 Gottsberger, G., Schrauwen, J., Linskens, H.F., 1984. Amino-Acids and Sugars in Nectar, and Their
573 Putative Evolutionary Significance. Plant Systematics and Evolution 145, 55-77.
574 Herbert, E.W., Shimanuki, H., 1978. Chemical Composition and Nutritive-Value of Bee-Collected
575 and Bee-Stored Pollen. Apidologie 9, 33-40.
576 Inouye, D.W., Waller, G.D., 1984. Responses of Honey Bees (Apis-Mellifera) to Amino-Acid
577 Solutions Mimicking Floral Nectars. Ecology 65, 618-625.
578 Kim, Y.S., Smith, B.H., 2000. Effect of an amino acid on feeding preferences and learning
579 behavior in the honey bee, Apis mellifera. Journal of Insect Physiology 46, 793-801.
580 Lanza, J., 1988. Ant Preferences for Passiflora Nectar Mimics That Contain Amino-Acids.
581 Biotropica 20, 341-344.
582 Lanza, J., Krauss, B.R., 1984. Detection of Amino-Acids in Artificial Nectars by 2 Tropical Ants,
583 Leptothorax and Monomorium. Oecologia 63, 423-425.
584 Levin, M.D., Bohart, G.E., 1955. Selection of pollens by honey bees. Am Bee Journal 95, 392-393,
585 402.
586 Linander, N., de Ibarra, N.H., Laska, M., 2012. Olfactory Detectability of L-Amino Acids in the
587 European Honeybee (Apis mellifera). Chemical Senses 37, 631-638.
588 Micheu, S., Crailsheim, K., Leonhard, B., 2000. Importance of proline and other amino acids
589 during honeybee flight (Apis mellifera carnica POLLMANN). Amino Acids 18, 157-175.
590 Mollaei, M., Hoseini, S.A., Karimi, M., Hekmat, Z., 2013. Short communication. Impact of the
591 amino acid proline on the cold hardiness of honey bee, Apis mellifera L. Spanish Journal of
592 Agricultural Research 11, 714-717.
593 Nelson, C.M., Ihle, K.E., Fondrk, M.K., Page, R.E., Amdam, G.V., 2007. The gene vitellogenin has
594 multiple coordinating effects on social organization. Plos Biology 5, 673-677.
595 Nepi, M., Soligo, C., Nocentini, D., Abate, M., Guarnieri, M., Cai, G., Bini, L., Puglia, M., Bianchi,
596 L., Pacini, E., 2012. Amino acids and protein profile in floral nectar: Much more than a simple
597 reward. Flora 207, 475-481.
598 Page, R.E., 2013. The Spirit of the Hive: The Mechanisms of Social Evolution. Harvard University
599 Press.
600 Pernal, S.F., Currie, R.W., 2001. The influence of pollen quality on foraging behavior in
601 honeybees (Apis mellifera L.). Behavioral Ecology and Sociobiology 51, 53-68.
602 Pernal, S.F., Currie, R.W., 2002. Discrimination and preferences for pollen-based cues by
603 foraging honeybees, Apis mellifera L. Animal Behaviour 63, 369-390.
604 Potter, C.F., Bertin, R.I., 1988. Amino-Acids in Artificial Nectar - Feeding Preferences of the Flesh
605 Fly Sarcophaga Bullata. American Midland Naturalist 120, 156-162.
606 Rasheed, S.A., Harder, L.D., 1997a. Economic motivation for plant species preferences of pollen-
607 collecting bumble bees. Ecological Entomology 22, 209-219.
608 Rasheed, S.A., Harder, L.D., 1997b. Foraging currencies for non-energetic resources: pollen
609 collection by bumblebees. Animal Behaviour 54, 911-926.
610 Robinson, G.E., Vargo, E.L., 1997. Juvenile hormone in adult eusocial hymenoptera:
611 Gonadotropin and behavioral pacemaker. Archives of Insect Biochemistry and Physiology 35,
612 559-583.
613 Rodriguez-Pena, N., Stoner, K.E., Ayala-Berdon, J., Flores-Ortiz, C.M., Duran, A., Schondube, J.E.,
614 2013. Nitrogen and amino acids in nectar modify food selection of nectarivorous bats. Journal of
615 Animal Ecology 82, 1106-1115.
616 Roubik, D., W., Yanega, D., Aluja, Buchmann, S., L., Inouye, D., W., 1995a. On optimal nectar
617 foraging by some tropical bees (Hymenoptera: Apidae). Apidologie 26, 197-211.
618 Roubik, D.W., Yanega, D., Aluja, M., Buchmann, S.L., Inouye, D.W., 1995b. On Optimal Nectar
619 Foraging by Some Tropical Bees (Hymenoptera, Apidae). Apidologie 26, 197-211.

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620 Saa-Otero, M.P., Diaz-Losada, E., Fernandez-Gomez, E., 2000. Analysis of fatty acids, proteins
621 and ethereal extract in honeybee pollen - Considerations of their floral origin. Grana 39, 175-
622 181.
623 Sanchez, M.G.D., 2011. Taste Perception in Honey Bees. Chemical Senses 36, 675-692.
624 Schmidt, J.O., 1982. Pollen Foraging Preferences of Honey Bees (Hymenoptera, Apidae).
625 Southwestern Entomologist 7, 255-259.
626 Seeley, 2009. The Wisdom of the Hive: the social physiology of honey bee colonies.
627 Shafir, S., Menda, G., Smith, B.H., 2005. Caste-specific differences in risk sensitivity in
628 honeybees, *Apis mellifera*. Animal Behaviour 69, 859-868.
629 Shafir, S., Reich, T., Tsur, E., Erev, I., Lotem, A., 2008. Perceptual accuracy and conflicting effects
630 of certainty on risk-taking behaviour. Nature 453, 917-920.
631 Simmonds, M.S.J., Simpson, S.J., Blaney, W.M., 1992. Dietary Selection Behavior in Spodoptera-
632 Littoralis - the Effects of Conditioning Diet and Conditioning Period on Neural Responsiveness
633 and Selection Behavior. Journal of Experimental Biology 162, 73-90.
634 Simpson, S.J., James, S., Simmonds, M.S.J., Blaney, W.M., 1991. Variation in Chemosensitivity
635 and the Control of Dietary Selection Behavior in the Locust. Appetite 17, 141-154.
636 Singaravelan, N., Nee'man, G., Inbar, M., Izhaki, I., 2005. Feeding responses of free-flying
637 honeybees to secondary compounds mimicking floral nectars. Journal of Chemical Ecology 31,
638 2791-2804.
639 Toshima, N., Tanimura, T., 2012. Taste preference for amino acids is dependent on internal
640 nutritional state in Drosophila melanogaster. The Journal of Experimental Biology 215, 2827-
641 2832.
642 Wada, A., Isobe, Y., Yamaguchi, S., Yamaoka, R., Ozaki, M., 2001. Taste-enhancing effects of
643 glycine on the sweetness of glucose: a gustatory aspect of symbiosis between the ant,
644 Camponotus japonicus, and the larvae of the lycaenid butterfly, Niphanda fusca. Chemical
645 Senses 26, 983-992.
646 Waddington, K.D., 2001. Subjective evaluation and choice behavior by nectar- and pollen-
647 collecting bees, in: Chittka, L., Thompson, J.D. (Eds.), Cognitive ecology of pollination: animal
648 behavior and floral evolution. Cambridge University Press, Cambridge, pp. 41-60.
649 Waddington, K.D., Gottlieb, N., 1990. Actual vs perceived profitability: a study of floral choice of
650 honey bees. Journal of Insect Behavior 3, 429-441.
651 Waddington, K.D., Nelson, C.M., Page, R.E., 1998. Effects of pollen quality and genotype on the
652 dance of foraging honey bees. Animal Behaviour 56, 35-39.
653 Weiner, C.N., Hilpert, A., Werner, M., Linsenmair, K.E., Bluthgen, N., 2010. Pollen amino acids
654 and flower specialisation in solitary bees. Apidologie 41, 476-487.
655 Wright, G.A., Baker, D.D., Palmer, M.J., Stabler, D., Mustard, J.A., Power, E.F., Borland, A.M.,
656 Stevenson, P.C., 2013. Caffeine in Floral Nectar Enhances a Pollinator's Memory of Reward.
657 Science 339, 1202-1204.

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TABLES

Table 1: H1 homing test results. Treatments were 40% sucrose control, and 40% sucrose enriched with 0.1% AA (glycine, leucine, phenylalanine, or proline). The imbibition test concerned 1050 bees, 70 jars, with 36 hours before testing (see Suppl. 3).

Endpoints H1:	**Uptake** (µl bee^{-1} jar^{-1})	**Mortality** (mean % jar^{-1})	**Return rate** (% bees)	**Flight time** (minutes)
AA treatment	$F_{4,62}$=1.98, *P*=0.11	$F_{4,62}$=1.37, *P*=0.26	X^2=1.35, *Df*=3, *P*=0.85	X^2=5.38, *Df*=4, *P*=0.25
P & N foragers	$F_{1,62}$=89.1, *P*<0.001*	$F_{1,62}$=12.6, *P*<0.001*	X^2=1.61, *Df*=1, *P*=0.20	X^2=4.56, *Df*=1, *P*=0.033 (ns)
Colony A & B	$F_{1,62}$=10.4, *P*=0.002*	$F_{1,62}$=22.8, *P*<0.001*	X^2=39.4, *Df*=1, *P*<0.001*	X^2=0.18, *Df*=1, *P*=0.66
Trial Sep & Oct	$F_{1,62}$=45.1, *P*<0.001*	$F_{1,62}$=27.5, *P*<0.001*	X^2=58.9, *Df*=1, *P*<0.001*	X^2=23.2, *Df*=1, *P*<0.001*
Overall mean	123 µl	157/1050 bees	330/807 bees	median 32 min

* Significant differences between levels had following effect sizes: Solution uptake per forager type (P: 141 µl > N: 107 µl), per colony (A: 119 µl < B: 130 µl), and trial (Sept: 112 µl < Oct: 136 µl). Mortality per forager type (P: 22% > N: 10%), per colony (A: 8% < B: 24%), and trial (Sept: 8% > Oct: 25%). Return rates differed per colony (A: 217 non-homers vs 224 homers; B: 206 non-homers vs 106 homers), and trial (Sept: 369 non-homers vs 169 homers; Oct: 109 non-homers vs 161 homers). Median flight time differed per trial (Sept: 18 min < Oct: 40 min), but not for forager type (P: 25 min < N: 35 min; Bonferroni corrected). No interaction between AA treatment and another factor was indicated.

Table 2: H2 homing test results. Treatments were three sucrose solution concentrations: 8, 16, and 32%. Beside pollen and nectar foragers, effects by two colonies were considered, as tested on individual trial dates. The imbibition test concerned 549 bees, 28 jars, with 16 hours before testing (see Suppl. 3).

Endpoints H2	**Uptake** (µl bee^{-1} jar^{-1})	**Mortality** (mean % jar^{-1})	**Return rate** (% bees)	**Flight time** (minutes)
Carbohydrate	$F_{2,23}$=5.59, *P*=0.011*	$F_{2,23}$=64.0, *P*<0.001*	X^2=2.88, *Df*=2, *P*=0.24	X^2=39.7, *Df*=2, *P*<0.001*
P & N foragers	$F_{1,23}$=0.13, *P*=0.72	$F_{1,23}$=0.01, *P*=0.93	X^2=0.17, *Df*=1, *P*=0.68	X^2=4.38, *Df*=1, *P*=0.036 (ns)
Colony B & C	$F_{1,23}$=2.12, *P*=0.16	$F_{1,23}$=2.70, *P*=0.11	X^2=7.00, *Df*=1, *P*=0.008*	X^2=18.8, *Df*=1, *P*<0.001*
Overall mean	50.1 µl	93/548 bees	384/460 bees	median 27 min

* Significant differences between imbibed solutions by bees (42 µl [8] < 58 µl [16 and 32]), mortality rates (24.7% [8] > 8.8% [16] > 1.6% [32]), and also in flight time (36.5 min [8] and 35 min [16] > 14.5 min [32]). Further a colony effect was indicated for return rates (B: 50 non-homers vs 189 homers; C: 26 non-homers vs 195 homers), and median flight times per colony (B: 33 min; C: 19min). No interaction between AA treatment and another factor was indicated.

Table 3: H3 homing test results. Treatments were 20% sucrose control, and 20% sucrose enriched with 0.1% AA (glycine, phenylalanine, or proline). The imbibition test concerned 480 bees, 24 jars, with 20 hours before testing. See Suppl. 3.

Endpoints H3	**Uptake** (µl bee^{-1} jar^{-1})	**Mortality** (mean % jar^{-1})	**Return rate** (% bees)	**Flight time** (minutes)
AA treatment	$F_{3,26}$=1.27, *P*=0.31	$F_{3,26}$=0.33, *P*=0.80	X^2=1.18, *Df*=3, *P*=0.76	X^2=3.15, *Df*=3, *P*=0.37
P & N foragers	$F_{1,26}$=0.85, *P*=0.36	$F_{1,26}$=1.14, *P*=0.29	X^2=0.77, *Df*=1, *P*=0.37	X^2=0.13, *Df*=1, *P*=0.72
Colony D & E	$F_{1,26}$=10.7, *P*=0.003*	$F_{1,26}$=9.81, *P*=0.004*	X^2=1.18, *Df*=1, *P*=0.28	X^2=2.86, *Df*=1, *P*=0.09
Overall mean	92.5 µl	152/480 bees	200/272 bees	median 30 min

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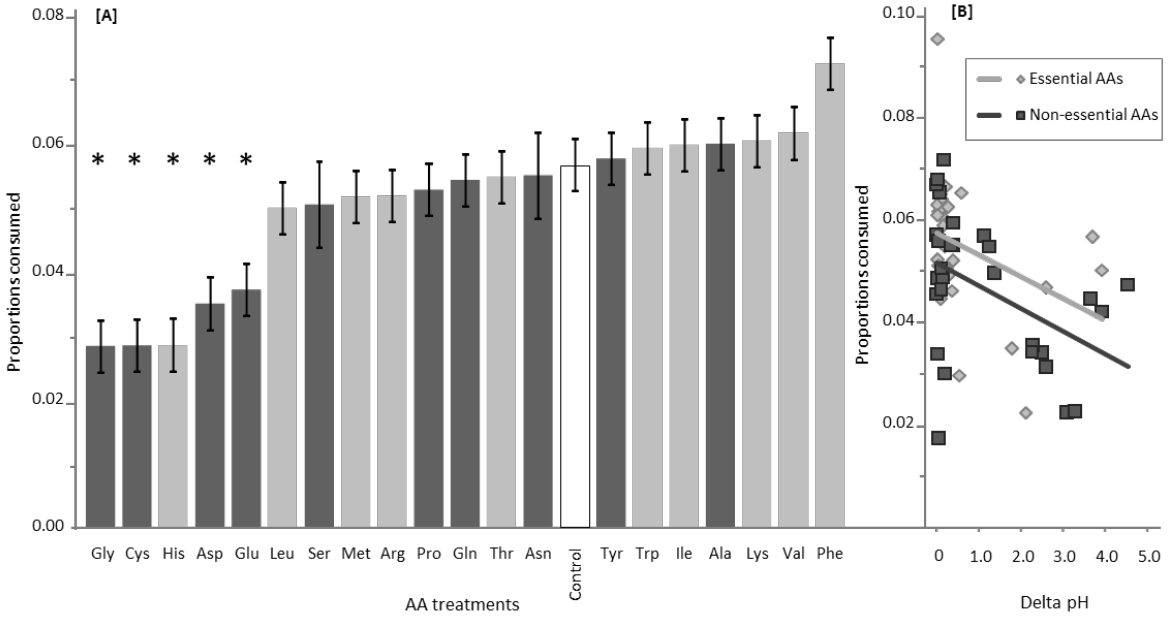
682 * Significant differences between imbibed solutions by bees per colony (D: 87.0 µl; E: 98.1 µl), and in mortality rates
683 per colony (D: 23.1%; E: 38.5%). No interaction between AA treatment and another factor was indicated.

684 **FIGURES**

685

686

687



688

689 **Fig. 1.** Amino acid preferences amongst free-flying foragers in an artificial meadow, according to collected
690 proportions of AA solutions, tested simultaneously against a control. In pane A, as from the left, AAs are
691 placed from least to most favored. The bars show mean values of collected solution amount per fence
692 section (mean ± SE, n sections; see Supplement 1). The data represent both non-essential (dark grey) and
693 essential (light grey) AAs. Asterisks (*) indicate AAs that were significantly different as compared to the
694 control (white bar). Pane B shows the result of a regression between deviation in pH values (as compared
695 to the control), and the factor essentiality. Shown are trend lines through pH deviation and mean
696 proportional consumption data (for each AA, per trial): both pH deviation and essentiality were
697 explanatory significant. The abbreviations for amino acids are listed in Supplement 2.

698

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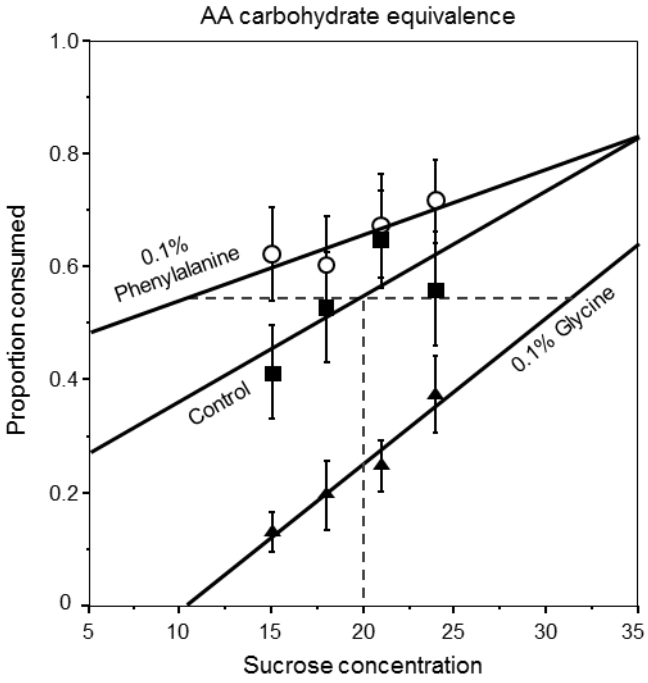

Fig. 2. Tradeoff between AAs and carbohydrates. Bees collected sucrose solutions over a gradient of 15%, 18%, 21% and 24% sucrose, with and without AA's. The points with error bars indicate consumed treatment solutions (mean ± SE), squares for the control, open circles for phenylalanine, and triangles for glycine. The control regression line [y = 0.019x + 0.175] at concentration 20% indicates a consumed proportion of 0.549. The horizontal dotted line indicates the extrapolated equivalence at which 0.1% AA solutions are expected to be consumed equally to the control. At this level, the 0.1% phenylalanine regression line [y = 0.012x + 0.425] is at 10.6% sucrose concentration, hence at +Δ9.4% sucrose equivalence. At the same level, the 0.1% glycine regression line [y = 0.026x - 0.273] is at 31.4% sucrose, hence at -Δ11.4% sucrose equivalence.

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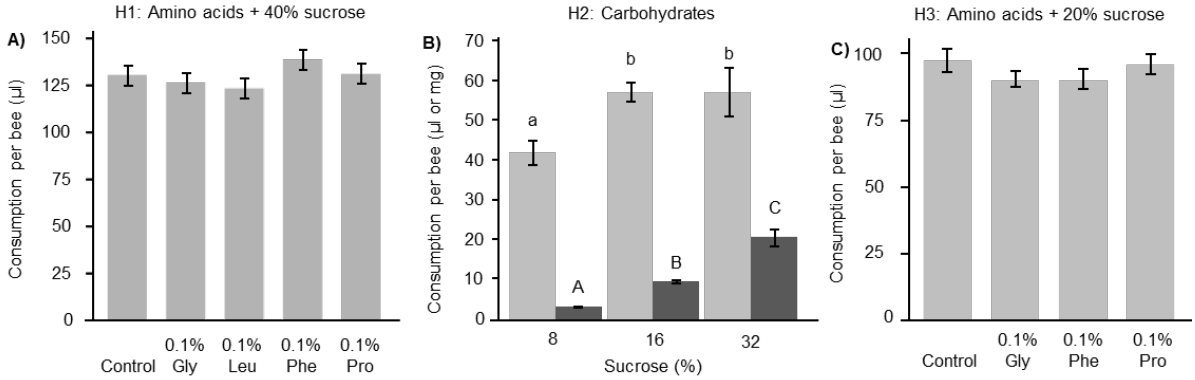

711

712 **Fig. 3.** Dietary uptake (mean ± SE) of sucrose solution preceding three homing experiments, to
713 determine whether AA and carbohydrate nutrition influence flight performance of foragers. In
714 the first experiment (pane A: H1), forager bees were kept in jars for 36 h, whereas in the
715 following experiments (pane B and C: H2 and H3) bees were kept in jars for 16 h and 20 h,
716 respectively. Effect of AAs in 40% (H1) and 20% (H3) sucrose solution consumption are shown
717 (µl bee^{-1} jar^{-1}). The effect of a concentration gradient of 8%, 16%, and 32% sucrose (H2) on
718 consumption is measured in µl solution (light bars), and given additionally as absolute amount
719 of sucrose in mg (dark bars). Different letters represent significantly different values ($P<0.05$;
720 Tukey). Notable is the indifference in imbibition of phenylalanine and glycine solutions.

721

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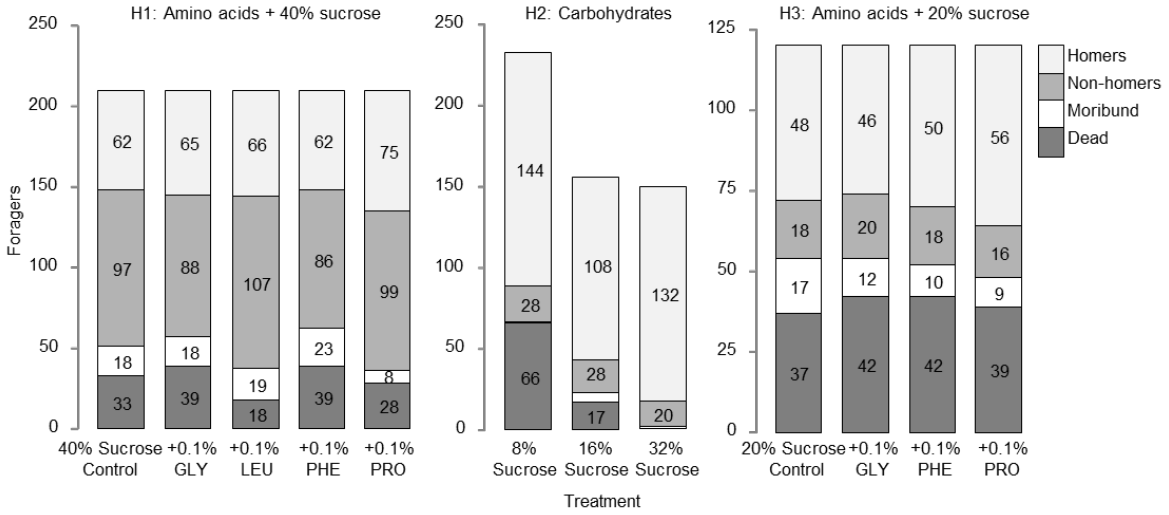
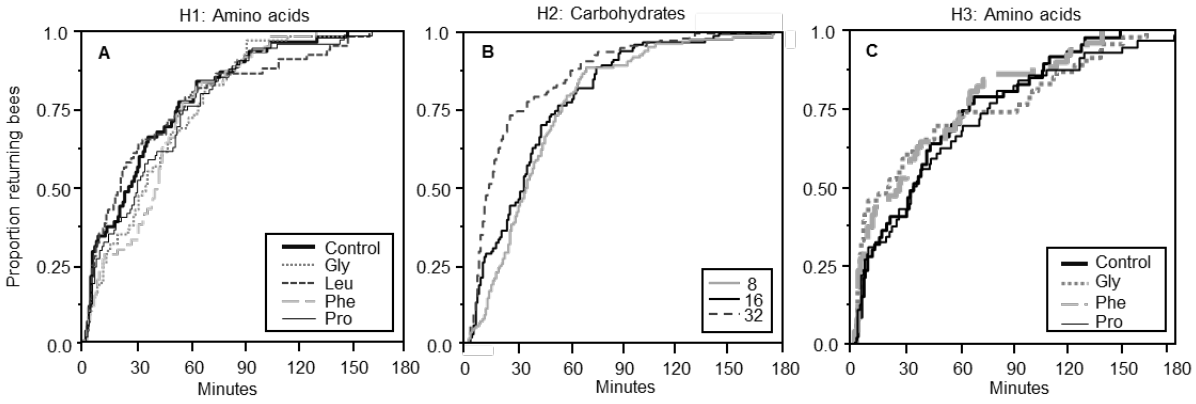


722

723 **Fig 4.** Fates of all the 2079 bees in homing experiments. Those that died during the feeding
724 phase were classified as dead. Moribund are those that were alive at release but did not take
725 off. Non-homers took off but did not return to the hive. Homers are those that returned to the
726 hive. In H2 where a sucrose gradient was tested, we began with more bees in the 8% sucrose
727 treatment, anticipating higher mortality.

728



729

730 **Fig 5.** Flight time analyses of cumulative forager homecoming rates. A) Experiment H1, testing
731 the effect of 0.1% AA nutrition in presence of 40% sucrose. B) Experiment H2, testing the effect
732 of carbohydrate nutrition (a gradient of 8, 16 and 32% sucrose solution). C) Experiment H3,
733 testing the effect of 0.1% AA nutrition in presence of 20% sucrose.

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734

735 **Supplement 1**

736

737 S1: EXPERIMENTAL DESIGNS OF FENCE TESTS: AA PREFERENCE AND EQUIVALENCE

738 The experimental designs of the preference tests of AA on free flying foragers.
739 Treatment solutions were offered to the bees at fence sites close to apiaries. The design
740 follows a Latin square considering that a treatment position in a replicate section is
741 represented only once, so that potential spatial effects are equally balanced within the
742 experimental setups. Three geographically different fence sites were used, on two of
743 our apiaries. Each apiary contained 20-30 hives which were exchanged from time to
744 time. The tests were performed over several months.

745

746 **S1-a Test on essential AAs**. March 08, 2013, apiary 1, fence site 1

747 10 essential AA + 1 control: 8 replicate linear sections

Ile	A	SECTION 1	B	K	H	G	I	J	E	F	C	D	A
Trp	B	SECTION 2	D	B	I	C	F	G	H	K	J	A	E
Met	C	SECTION 3	F	E	B	J	C	K	D	G	A	H	I
Arg	D	SECTION 4	H	C	F	B	K	D	G	A	I	E	J
His	E	SECTION 5	J	K	D	H	B	E	A	I	C	G	F
Lys	F	SECTION 6	E	G	I	J	A	C	B	F	H	D	K
Phe	G	SECTION 7	I	F	E	A	G	H	J	B	D	K	C
Val	H	SECTION 8	G	A	J	K	D	I	C	E	F	B	H
Thr	I												
Leu	J												
Con	K												

748

749 **S1-b Test on nonessential AAs**. March 10, 2013, apiary 1, fence site 1

750 8 non-essential AA + 1 control: 8 replicate linear sections

Ala	A	SECTION 1	B		H	G	I		E	F	C	D	A
Asp	B	SECTION 2	D	B	I	C	F	G	H			A	E
Cys	C	SECTION 3	F	E	B		C		D	G	A	H	I
Glu	D	SECTION 4	H	C	F	B		D	G	A	I	E	
Gln	E	SECTION 5			D	H	B	E	A	I	C	G	F
Gly	F	SECTION 6	E	G	I		A	C	B	F	H	D	
Pro	G	SECTION 7	I	F	E	A	G	H		B	D		C
Tyr	H	SECTION 8	G	A			D	I	C	E	F	B	H
CON	I												

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S1-c **Test on all AAs** (Fig 1): 20 AA + 1 control25 replicate block sections:

10 sections (black) March 13, 2013, apiary 1, fence sites 1

6 sections (red) March 15, 2013, apiary 1, fence sites 1

9 sections (green) October 16, 2013, apiary 2, fence sites 2

Glu	1		16	5	10		4		6	11	15	2	1	17		9	18	14	7	12	3	8	13	2		19
Leu	2	6	11		15	4	9	1	17			3	18	12	7	14	13	19	2		8	10	5	16		
Arg	3	17		1		9	14	18	12	7	3	8	13		2	19	5		16		10	15		11	6	4
Ala	4	12	7	18	3	14	19	13		2	8	10	5		16			4	11	6	15		1		17	9
Control	5		2	13	8	19		5		16	10	55		6	11	4	1	9		17		3	18	7	12	14
Asp	6	14	12	7	18	3	8	2	19		13	5	16			10	11	15	6	4		1		17	9	
Gln	7	9	17		1		3	7	14	12	18	13	2	19		8	16	10			5		11	6	4	15
Met	8	4	6	11		15			9	17	1	18	7	14	12	3	2	8		19	13	5	16			10
Lys	9			16	5	10	15	11	4	6		1		9	17		7	3	12	14	18	13	2		19	8
Ile	10	19		2	13	8	10	16			5		11	4	6	15			17	9	1	18	7	12	14	3
Trp	11	3	14	12	7	18	13		8	19	2	16		10		5	6		4	15	11		17	9		1
Pro	12	8	19		2	13	5		10		16	11	6	15	4		17	1	9			7	12	14	3	18
Cys	13	10			16	5		6	15	4	11		17		9	1	12	18	14	3	7	2		19	8	13
His	14	15	4	6	11		1	17		9		7	12	3	14	18		13	19	8	2	16			10	5
Tyr	15		9	17		1	18	12	3	14	7	2		8	19	13		5		10	16	11	6	4	15	
Gly	16	1		9	17		7	14	18	3	12		19	13	8	2	21	16	10	5		6	4	15		11
Val	17	18	3	14	12	7	2	19	13	8			21	5	10	16	4	11	15		6	17	9	20	1	
Thr	18	13	8	19		2	16	21	5	10		6	4		15	11	9		20	1	17	12	14	3	18	7
Phe	19	5	10			16	11	4		15	6	17	9	1	20		14	7	3	18	12		19	8	13	2
Asn	20		15	4	6	11		9	1	20	17	12	14	18	3	7	19	2	8	13			21	10	5	16
Ser	21	16	5	10	21		6	15	11		4	9	20		1	17	3	12	18	7	14	19	8	13	2	
		2	13	8	19			10	16	5	21	4	15	11		6	20	17	1		9	14	3	18	7	12
		7	18	3	14	12		8	2	13	19	21	10	16	5		15	6		11	4	9	20	1		17
			1	20	9	17	12	3	7	18	14	19	8	2	13		10		5	16	21	4	15		11	6
		11		15	4	6	17	20		1	9	14	3	7	18	12	8		13	2	19	21	10	5	16	

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S1-d **Test on AA equivalence** (Fig. 2): Tradeoff between carbohydrates and AAs

12 replicate block sections, apiary 1, fence site 3

2 sections (black) November 5, 2013

4 sections (red) November 7, 2013

6 sections (green) November 20, 2013

1	15% Sucrose	4	11	8	9	3	6	1	5	7	10	2	12
2	18% Sucrose	5	1	9	4	10	8	3	12	2	7	6	11
3	21% Sucrose	7	10	3	1	12	2	8	11	6	5	9	4
4	24% Sucrose	2	6	12	5	11	7	10	9	4	1	8	3
5	15% Sucrose + 0.1% GLY	9	5	11	8	1	10	7	6	12	3	4	2
6	18% Sucrose + 0.1% GLY	1	7	6	3	5	11	2	4	9	8	12	10
7	21% Sucrose + 0.1% GLY	3	4	2	6	9	12	5	10	8	11	1	7
8	24% Sucrose + 0.1% GLY	8	12	10	2	7	4	11	1	3	6	5	9
9	15% Sucrose + 0.1% PHE	11	8	1	10	2	9	4	7	5	12	3	6
10	18% Sucrose + 0.1% PHE	10	3	4	12	8	1	6	2	11	9	7	5
11	21% Sucrose + 0.1% PHE	12	2	7	11	6	5	9	3	1	4	10	8
12	24% Sucrose + 0.1% PHE	6	9	5	7	4	3	12	8	10	2	11	1

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766

767 **Supplement 2**

768

769 S2: TABLE AMINO ACID INFORMATION (Fig. 1; Supplement S1-c)

770 Background information on amino acids for tests on preferences by nectar foragers

771

Amino acids		L/D Isomere	Molecular weight	mM (0.1% w/w)	Proportion	SD	(n)	pH	(n)	Delta pH
Arginine	Arg	L-ARGININE base	174.2	6.17	5.19%	2.0%	(25)	9.46	(3)	3.53
Histidine	His	L-HISTIDINE base	155.16	6.93	2.86%	2.3%	(25)	7.39	(3)	1.63
Isoleucine	Ile	L-ISOLEUCINE	131.18	8.20	5.98%	2.0%	(25)	6.20	(3)	0.42
Leucine	Leu	L-LEUCINE	131.18	8.20	5.00%	1.9%	(25)	5.92	(3)	0.17
Lysine	Lys	L-LYSINE HCL	182.65	5.89	6.04%	2.2%	(25)	5.74	(3)	0.18
Methionine	Met	L-METHIONINE	149.21	7.21	5.18%	2.0%	(25)	5.79	(3)	0.17
Phenylalanine	Phe	L-PHENYLALANINE	165.19	6.51	7.26%	2.6%	(25)	5.90	(3)	0.03
Threonine	Thr	L-THREONINE	119.12	9.03	5.49%	1.9%	(25)	5.94	(3)	0.11
Tryptophan	Trp	L-TRYPTOPHAN	204.25	5.26	5.94%	2.3%	(25)	5.83	(3)	0.12
Valine	Val	L-VALINE	117.15	9.18	6.17%	2.1%	(25)	6.01	(3)	0.10
Alanine	Ala	L-ALANINE	89.1	12.07	6.00%	1.8%	(25)	5.99	(3)	0.09
Asparagine	Asn	L-ASPARAGINE	132.12	8.14	5.51%	2.8%	(9)	5.20	(1)	0.35
Aspartate	Asp	L-ASPARTIC ACID	133.1	8.08	3.50%	1.5%	(25)	2.89	(3)	2.89
Cysteine	Cys	L-CYSTEINE HCL	175.63	6.12	2.85%	1.9%	(25)	2.26	(3)	3.52
Glutamine	Gln	L-GLUTAMINE	146.15	7.36	5.44%	1.6%	(25)	4.65	(3)	1.23
Glutamate	Glu	L-GLUTAMIC ACID	147.13	7.31	3.72%	1.8%	(25)	3.16	(3)	2.62
Glycine	Gly	GLYCINE	75.07	14.32	2.84%	1.9%	(25)	6.00	(3)	0.11
Proline	Pro	L-PROLINE	115.13	9.34	5.28%	1.8%	(25)	5.80	(3)	0.12
Serine	Ser	L-SERINE	105.09	10.23	5.06%	2.8%	(9)	5.44	(1)	0.11
Tyrosine	Tyr	L-TYROSINE	181.19	5.93	5.77%	2.1%	(25)	6.06	(3)	0.24
Control	Con				5.68%	2.2%	(25)	5.91	(3)	0.00
		[1]		[2]	[3]			[4]		[5]

772

773 [1] The amino acids were ordered from Sigma-Aldrich (Israel) and PureBulk (USA).

774 Certificates of analysis (COA) per AA indicated purity of a mean 99.7 ± 0.7% SD.

775 [2] 0.1% equals 1.0 g AA / 1.0 kg solution (w/w), thus 1.0 g / 0.930 L (w/v), which equals MW / 0.930 moles per L.

776 This conversion of 0.1% AA (w/w) to molarity is based on the volume of 1.0 Kg of a 20% sucrose solution: 930 ml.

777 Equally: our tested concentration was at 108 mg / 100 ml, allowing comparisons, e.g. to Inouye and Waller (1984).

778 [3] The preferences for 20 AAs as indicating by free flying bees visiting replicate fence sections which all contained

779 each AA once. A unique spatial pattern for each section was considered (Supplement S1-c). Proportions collected

780 solutions were calculated per replicate fence section.

781 [4] The reported standard deviation of pH measurements was between separate batches of measurements.

782 Measurements were conducted with a Microprocessor pH Meter, "pH 211", Hanna Instruments. A mean pH is given

783 of n trials, at which all pH values were an average of a threefold measurement.

784 [5] pH deviation data were derived of all AAs in experiment S1-c, in comparison to controls of the same date.

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Supplement 3

S3: TABLE HOMING EXPERIMENTS

Experimental designs of three homing experiments, as described in the materials and methods section. The tests aimed to monitor AA nutrition effects on forager flight (H1 and H3) considering AA-induced alterations in carbohydrate uptake. Separately, carbohydrate uptake effects were tested in the second experiment (H2).

H1: AAs in 40% Sucrose	DATE	HIVE	Pol/Nec	COLOR	JARS	BEES
Control	September 24, 2013	B	N	WHITE	4	60
Control	October 2, 2013	A	N	BLUE	3	45
Control	September 24, 2013	A	P	WHITE	4	60
Control	October 2, 2013	B	P	BLUE	3	45
0.1% Gly	September 24, 2013	B	N	YELLOW	4	60
0.1% Gly	October 2, 2013	A	N	GREEN	3	45
0.1% Gly	September 24, 2013	A	P	YELLOW	4	60
0.1% Gly	October 2, 2013	B	P	GREEN	3	45
0.1% Leu	September 24, 2013	B	N	BLUE	4	60
0.1% Leu	October 2, 2013	A	N	RED	3	45
0.1% Leu	September 24, 2013	A	P	BLUE	4	60
0.1% Leu	October 2, 2013	B	P	RED	3	45
0.1% Phe	September 24, 2013	B	N	RED	4	60
0.1% Phe	October 2, 2013	A	N	YELLOW	3	45
0.1% Phe	September 24, 2013	A	P	RED	4	60
0.1% Phe	October 2, 2013	B	P	YELLOW	3	45
0.1% Pro	September 24, 2013	B	N	GREEN	4	60
0.1% Pro	October 2, 2013	A	N	WHITE	3	45
0.1% Pro	September 24, 2013	A	P	GREEN	4	60
0.1% Pro	October 2, 2013	B	P	WHITE	3	45
5 treatments	**2**	**2**	**2**	**5**	**70**	**1050**

H2: Carbohydrates	DATE	HIVE	Pol/Nec	COLOR	JARS	BEES
8% sucrose	November 5, 2013	B	N	BLUE	3	60
8% sucrose	November 12, 2013	C	N	BLUE	3	60
8% sucrose	November 5, 2013	B	P	YELLOW	3	54
8% sucrose	November 12, 2013	C	P	YELLOW	3	63
16% sucrose	November 5, 2013	B	N	WHITE	2	40
16% sucrose	November 12, 2013	C	N	WHITE	2	40
16% sucrose	November 5, 2013	B	P	GREEN	2	36
16% sucrose	November 12, 2013	C	P	GREEN	2	40
32% sucrose	November 5, 2013	B	N	AZURE	2	40
32% sucrose	November 12, 2013	C	N	AZURE	2	40
32% sucrose	November 5, 2013	B	P	RED	2	36
32% sucrose	November 12, 2013	C	P	RED	2	40
3 treatments	**2**	**2**	**2**	**6**	**28**	**549**

H3: AAs in 20% Sucrose	DATE	HIVE	Pol/Nec	COLOR	JARS	BEES
Control	April 10, 2014	D	N	DARK GREEN	2	30
Control	April 10, 2014	E	N	DARK GREEN	2	30
Control	April 10, 2014	D	P	LIGHT GREEN	2	30
Control	April 10, 2014	E	P	LIGHT GREEN	2	30
0.1% Gly	April 10, 2014	D	N	YELLOW	2	30
0.1% Gly	April 10, 2014	E	N	YELLOW	2	30
0.1% Gly	April 10, 2014	D	P	WHITE	2	30
0.1% Gly	April 10, 2014	E	P	WHITE	2	30
0.1% Phe	April 10, 2014	D	N	DARK BLUE	2	30
0.1% Phe	April 10, 2014	E	N	DARK BLUE	2	30
0.1% Phe	April 10, 2014	D	P	LIGHT BLUE	2	30
0.1% Phe	April 10, 2014	E	P	LIGHT BLUE	2	30
0.1% Pro	April 10, 2014	D	N	ORANGE	2	30
0.1% Pro	April 10, 2014	E	N	ORANGE	2	30
0.1% Pro	April 10, 2014	D	P	RED	2	30
0.1% Pro	April 10, 2014	E	P	RED	2	30
6 treatments	**1**	**2**	**2**	**8**	**32**	**480**
[1]	[2]	[3]	[4]	[5]	[6]	[7]

[1] Treatments were offered to bees in jars, either with a 36 hr. (H1), a 16 hr. (H2) or 20 hr. (H3) imbibition period.

[2] Homing bees at H1 and H3 were monitored by 4 experimenters (2 hives per date), and H2 with 2 experimenters (1 hive per date).

[3] Five different hives: One story 10-frame hive, with 4-6 frames brood, with an empty hive box on top to contain the jars with bees.

[4] Pollen and nectar collectors, caught as incoming foragers at a closed hive entrance, were contained and tested separately.

[5] Thorax markers: Solvent based “Edding 750 paint marker, Japan" (H1 & H2) / water based "uni POSCA, Japan" (H3).

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[6] A total 130 jars with 15 bees (H1 and H3) or 20 bees (H2), for consumption data per alive bee, and mortality over time.
[7] A total of 2079 bees entered the setup of the three homing experiments.

Supplement 4

S4: EQUIVALENCE EXPERIMENTS

For the AAs phenylalanine and glycine, an extrapolation assessed the equivalence of 0.1% AA to % sucrose (S1-d). The extrapolated points were beyond the tested gradient, thus additional experiments were performed to verify the expected equivalence: 20% sucrose ≈ 10.6% sucrose + 0.1 % phenylalanine ≈ 31.4% sucrose + 0.1% glycine

Design A: 3 treatments abc, 9 replicate sections, 54 test tubes, November 25, 2013, Apiary 1, fence site 3

1		2		3		4		5		6		7		8		9	
a	c	b	b	a	c	b	a	c	c	b	a	a	c	a	c	b	b
a	b	c	a	c	b	b	a	a	b	c	c	c	b	a	b	c	a
c	b	a	c	b	a	c	c	b	a	a	b	b	a	c	b	a	c

a (n=18) 11.6% sucrose[1] + 0.1% phenylalanine pH=8.05[2]
b (n=18) 20.0% sucrose[1] pH=8.11[2]
c (n=18) 31.7% sucrose[1] + 0.1% glycine[4] pH=8.09[2]

Design B: 4 treatments ABCD, 25 replicate sections, 100 test tubes, April 29, 2014, Apiary 1, fence site 3

1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
A	D	B	C	B	D	C	A	D	A	C	B	D	B	C	A
C	B	D	A	C	A	B	D	B	C	A	D	C	A	D	B
D	A	C	B	A	C	D	B	A	D	B	C	A	C	B	D
B	C	A	D	D	B	A	C	C	B	D	A	B	D	A	C

A (n=16) 20.0% sucrose[1] pH=6.55[3]
B (n=16) 20.0% sucrose[1] + 0.1% glycine[5] pH=6.91[3]
C (n=16) 30.0% sucrose[1] + 0.1% glycine[5] pH=6.72[3]
D (n=16) 40.0% sucrose[1] + 0.1% glycine[5] pH=6.64[3]

[1] Refractometer / brix values. [2] pH values are the mean of 2 measurements. [3] pH values are the mean of 3 measurements. [4] Glycine "Purebulk", manufacture date January 17 2013, ordered February 2013, tested November 2013. [5] Glycine "Sigma", manufacture date unknown, ordered April 2014, tested April 2014.

	Effect Tests	Significance	Dunnett	Least Square Mean
Equivalence check A	$F_{2,51 = 10.1}$	$P < 0.001$		Proportion
20% control				0.437
11.6% + phenylalanine			$P = 0.29$	0.372
31.7% + glycine			$P < 0.001$ *	0.231
Equivalence check B	$F_{3,60 = 25.6}$	$P < 0.001$		Proportion
20% control				0.205
20% + glycine			$P = 0.026$ *	0.115
30% + glycine			$P = 0.083$	0.279
40% + glycine			$P < 0.001$ *	0.400

Results A: [a=b] 1 part phenylalanine ≈ 84 parts sucrose, and [b>c] 1 part glycine > 117 parts sucrose.
Results B: [A<B] 1 part glycine ≈ deterrent, and [B=C] 1 part glycine ≈ 100 parts sucrose (Δ10% sucrose).

Test strengths	α	σ	δ	Sample Size	Power
Equivalence check A	0.05	0.141	0.086	54	0.98
Equivalence check B	0.05	0.095	0.104	64	1.00

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830 Conclusion: 1 part phenylalanine ≈ 84 parts sucrose. 1 part glycine ≥ 100 parts, but <200 parts sucrose.