

## **Genome-wide association analysis links multiple psychiatric liability genes to oscillatory brain activity.**

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Oscillations in neuronal activity are widely thought to play a crucial role in information processing and cortical communication<sup>1–8</sup>. Brain oscillations have been widely investigated as biomarkers of psychiatric disorders and variation in normal human behavior<sup>9,10</sup>, including intelligence<sup>11,12</sup>, schizophrenia<sup>13,14</sup>, attentional deficits<sup>15,16</sup>, and substance use<sup>17,18</sup>. Beyond the biomarker, oscillatory activity may indeed cause variation in behavior, as it has been shown that disrupting oscillatory activity by blocking GABAergic fast-spiking interneurons in the frontal cortex of mice impairs behavioral flexibility<sup>8</sup>, consistent with observed aberrant frontal cortical oscillatory responses during cognitive tasks requiring cognitive flexibility in humans<sup>19</sup>. Moreover, restoring oscillatory activity in *Dlx5/6+/-* transgenic mice with similar behavioral impairment—via optogenetic driving of GABA interneurons at gamma frequencies—restored normal behavioral flexibility, thus further supporting the causal involvement of oscillations in driving behavior<sup>8</sup>.

As one of the most heritable traits in humans<sup>20–25</sup>, studies have attempted to link oscillatory activity to the genetic liability psychiatric disorders via twin and family studies. However, studies linking specific genetic variants for oscillation strength to psychopathology remain scarce. Here, we performed a meta-analysis of genome-wide association studies (GWAS) between genetic variants and the oscillation strength in different frequency bands of electroencephalographic (EEG) recorded activity. In addition, we conducted gene-based analyses, compared these to known liability genes for psychiatric disorders, and examined gene-expression pathways.

The largest study of EEG power and peak frequency to date<sup>26</sup> did not yield genome-wide significant hits, but reported a significant contribution of common variants to heritability using random-effects modeling<sup>27</sup>. In other studies, the most consistent finding has been the involvement of GABA functioning in ~20 Hz beta oscillatory activity. Porjesz et al.<sup>28</sup> showed significant linkage between beta oscillations on chromosome 4 and the *GABRB1* microsatellite marker, which was overlying a cluster of GABA<sub>A</sub> receptor genes: *GABRG1*, *GABRA2*, *GABRA4* and *GABRB1*. Regional SNP association analysis subsequently pointed to SNPs intronic to *GABRA2* as accounting for the signal<sup>29</sup>. *GABRA2* was subsequently associated with both beta oscillations and alcohol use disorders<sup>29,30</sup>. A recent study conducted in 117 families of European ancestry from the Collaborative Study on the Genetics of Alcoholism associated several intergenic SNPs at 6q22 with >20 Hz fast beta oscillations<sup>31</sup>. One of the main aims of the current study is to investigate how genetic variation in *GABRA2* affects expression in brain tissues, extend to other oscillations frequencies, and whether liability genes for other psychopathological conditions influence brain function.

Since the effects of single genetic variants on brain activity are expected to be small, GWAS requires large sample sizes. Increases in sample size and statistical power may be obtained by meta-analysis of separate genome-wide association studies (GWAS). The current study describes the results from the ENIGMA-EEG workgroup of the ENIGMA consortium<sup>32,33</sup>. We developed EEG processing protocols to extract common measures for band power in the standard frequency bands delta, theta, alpha and beta power at the vertex (Cz) electrode, and occipital (O1, O2) alpha power and alpha peak frequency consistent with<sup>26</sup>. GWAS results of three population-based and two alcohol-dependence ascertained twin and family cohorts from the Netherlands, Australia, and US were combined in a meta-analysis for a total of 8425 individuals (see also<sup>34</sup>).

To gain further understanding of the results, extensive follow-up analyses were conducted. We performed enrichment analysis of expression Quantitative Trait Loci (eQTLs) in GTEx brain tissues, positional gene-based analysis, and SNP-based co-heritability analysis of our six EEG traits with related brain phenotypes and psychiatric traits. Follow-up analyses identified genetic variants previously implicated in schizophrenia, including eQTLs that influence gene expression in frontal cortical, anterior cingulate, and subcortical tissues. Hippocampal GABRA2 expression was linked to beta oscillations.

## RESULTS

### *Genome-wide association*

Supplementary figure S1 shows the Manhattan plots for the six EEG traits. Genome-wide significance was set at  $5 \cdot 10^{-8}$ . Two SNPs were genome-wide significant for Cz alpha power: (rs984924,  $p=4.7 \cdot 10^{-8}$  and rs10231372,  $p=2.9 \cdot 10^{-8}$ ); rs984924 on chromosome 4 is an intronic variant within protein kinase cGMP-dependent type II (PRKG2), and rs10231372 on chromosome 7 is an intronic variant within the long non-coding RNA gene LINC00996. Suggestive peaks ( $p < 5 \cdot 10^{-7}$ ) were found for Cz delta power on chromosome 5 (rs6867021,  $p=1.1 \cdot 10^{-7}$ ), chromosome 6 (rs17055223,  $p=3.1 \cdot 10^{-7}$ ), chromosome 2 (rs11677128,  $p=4.3 \cdot 10^{-7}$ ); Cz alpha power on chromosome 1 (rs10910665,  $p=1.8 \cdot 10^{-7}$ ) and on chromosome 13 (rs9514041,  $p=1.4 \cdot 10^{-7}$ ). Supplementary Table 1 shows the genome-wide significant SNPs, suggestive peaks, and FDR significant discoveries.

Q-Q plots for the meta-analysis are provided in Figure 1 (pink dots). Full genome median lambdas ranged from 1.02 to 1.06. Subsequent LD score-

intercepts—which can be used to parse inflation into true and spurious effects—were not significant ( $\text{abs}(z) < 1.50$ ); however, beta power did show significant inflation ( $z = 2.1$ ). Correction of p-values using the intercept had only minor effect and did not change any SNP or gene-based results. Overall, LD score intercept results indicated that there is no evidence of substantial inflation of statistics due to, for example, residual population stratification effects. *Positional gene-based analysis*  
We performed gene-based analysis using KGG Extended Simes test for each of the EEG traits, which combined the SNP p-values within genes plus flanking regions 50k basepair extensions in 5' and 3' UTR directions while taking into account the LD structure. Q-Q plots are included in Figure 1 (blue triangles). Plots showed inflation for gene p-values compared to SNP p-values. Figure 2 shows the gene-based Manhattan plots. FDR-corrected p-values showed significant genes for delta, theta, and alpha power at the vertex. Supplementary Table 2 shows the statistically significant gene discoveries (FDR  $q = 0.05$ ).

Consistent with the SNP-based findings, PRKG2 was significant for Cz alpha power ( $p = 0.019$ ). LOC101928942 ( $p = 0.019$ ) is an antisense noncoding RNA gene embedded in PRKG2. Both Cz and occipital alpha power showed a cluster of significant genes at 3p21 ranging from (hg19) basepair positions 52234203 to 52728499 (ALAS1 to GLT8D1). For Cz alpha power, 17 of these genes were significant discoveries at  $q = 0.05$ . For occipital alpha power, 11 genes reached significance of which 4 overlapped with Cz alpha. Figure S2 (top) shows the regional Cz alpha LocusZoom plot of the chromosome 3 region revealing high LD from about 52.2 to 52.8 Mb (hg19). Variants within the same region have been consistently associated with schizophrenia and bipolar disorder<sup>35,36</sup>. Figure S2 (bottom) shows the regional association plot for the second Psychiatric Genomics Consortium schizophrenia GWAS<sup>37</sup> for comparison. Top SNP in this region was rs7614727 ( $p = 2.0 \times 10^{-6}$ ), which is intronic to *WDR82*—previously associated with bipolar disorder and schizophrenia.

Further significant findings were a cluster of three genes (METTL21C, TPP2, and CCDC168; FDR  $p = 0.033$  for all) for Cz alpha power; and at 2p15 for occipital alpha power.

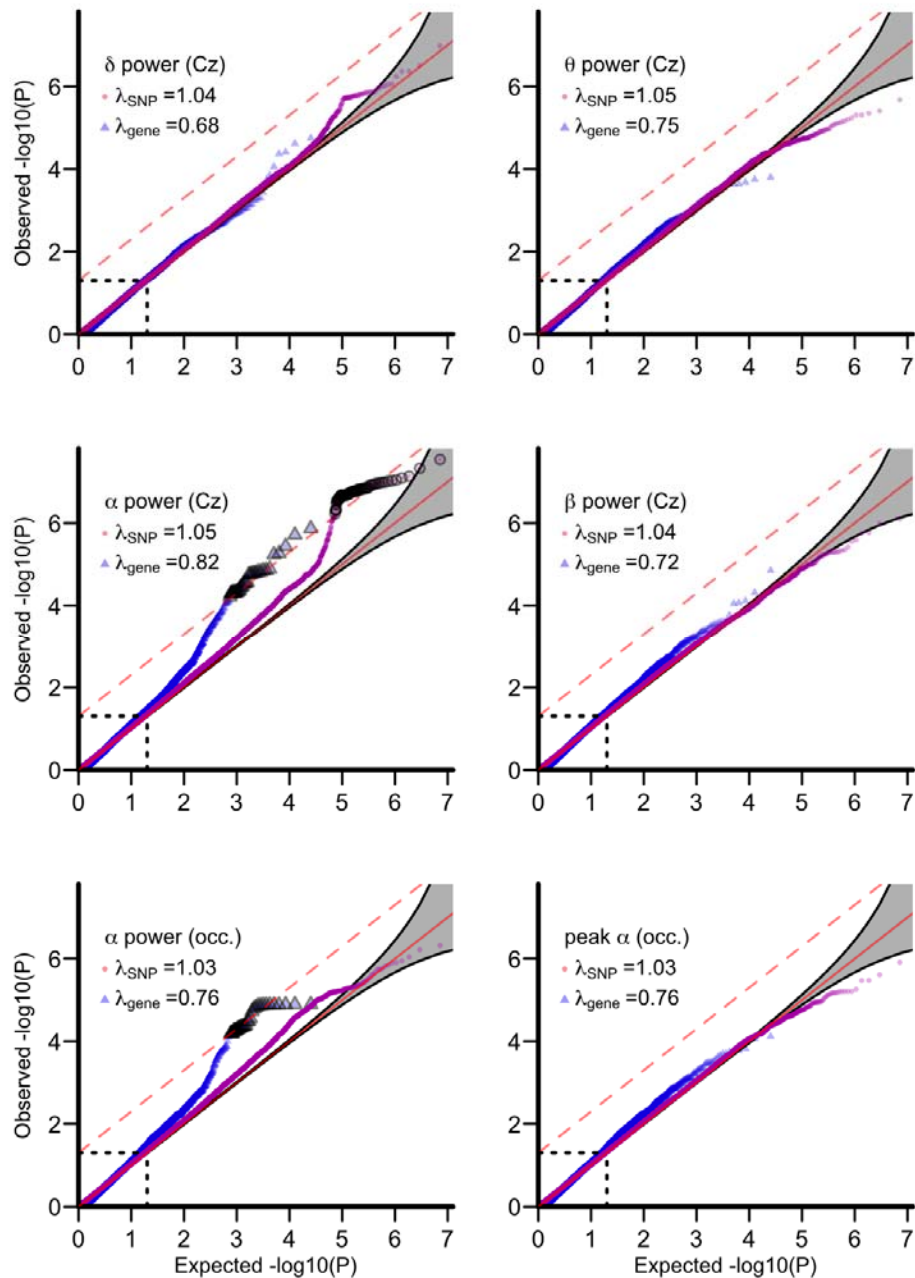


Figure 1. Quantile-quantile plots of observed versus expected  $-\log_{10}(p)$  for delta, theta, alpha, and beta EEG power at the vertex, occipital alpha power, and occipital alpha peak frequency. Red line is the expected null, grey area is the 95% confidence interval. Dashed red line is the Benjamini-Hochberg FDR  $q=0.05$  threshold. Pink dots are meta-analyzed SNP  $p$ -values. FDR-corrected significance is reached for alpha power at Cz. Blue triangles are KGG gene-based test  $p$ -values combining SNP effects within gene regions plus 50k base pairs 3' and 5' UTR. Many genes reach FDR significance for alpha oscillation power (Supplementary Table 2).

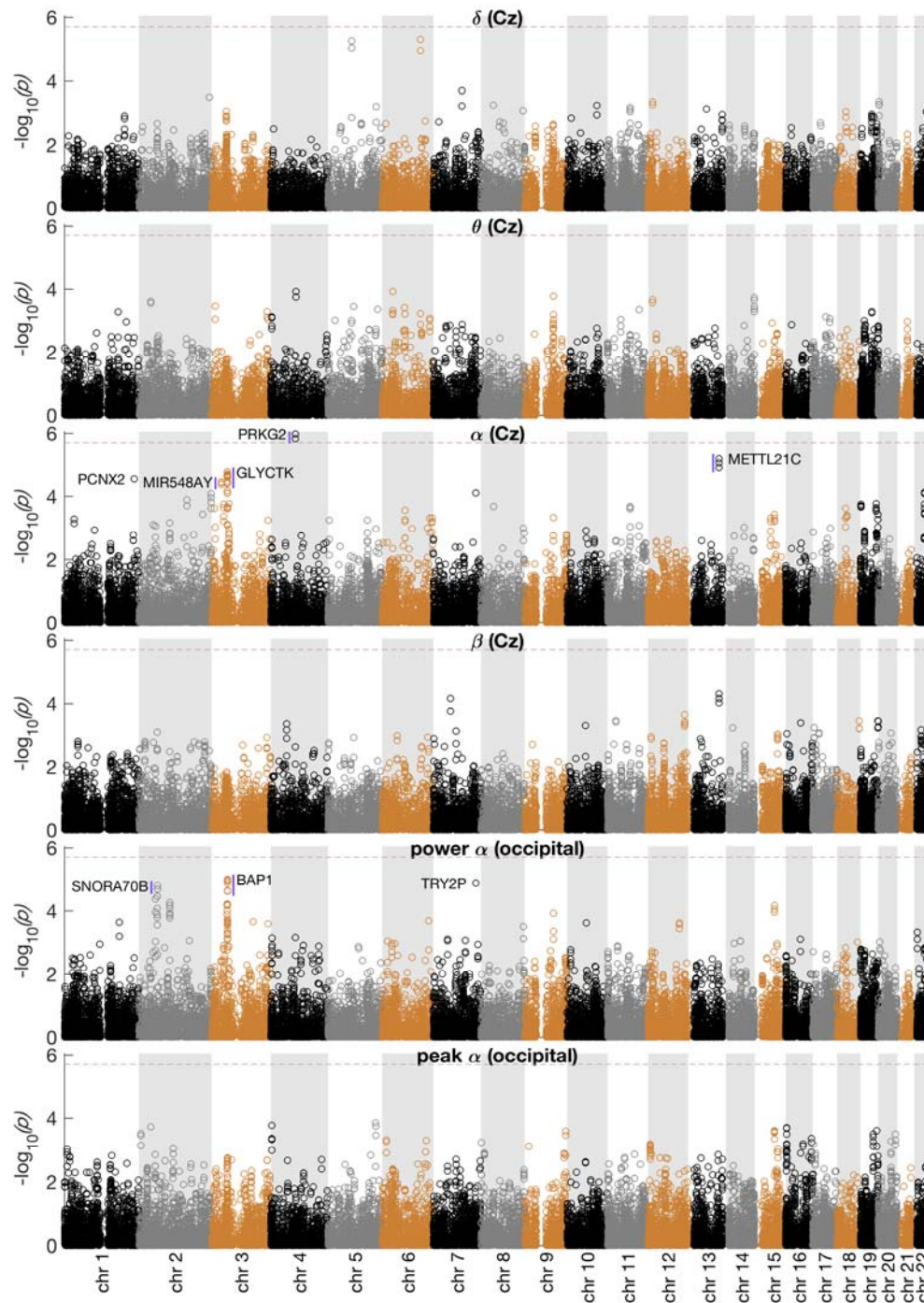


Figure 2. KGG gene-based test results Manhattan plots for the six EEG traits measured at the vertex electrode (Cz) and occipital (O1/O2). Dashed line is the threshold for genome-wide significance. Named genes are significant discoveries under FDR  $q=0.05$ . Only peak findings in the significant region marked with blue vertical lines are shown. A full listing of FDR significant genes is provided in Supplementary Table 2.

### eQTL Expression analysis

To investigate which genes are likely to mediate phenotypic variation in high LD regions and to elucidate mediating brain tissue expression pathways, we performed eQTL analysis. A substantial percentage of eQTLs affect the expression of genes at a distance, often including variants close to a different gene in a different LD region<sup>38</sup>. *Cis*-eQTLs are genetic variants within a 1Mb region of a gene that explain variability in the expression of the gene in a target tissue<sup>39–41</sup>. We selected eQTLs from eight brain tissues from the GTEx database<sup>41</sup>. Cz alpha power associated p-values resulted in inflated Q-Q plots for all tissues (Figure S3). Benjamini-Hochberg FDR significant effects at  $q=0.05$  were observed for the Frontal Cortex and Anterior Cingulate Gyrus, and the Hypothalamus. Occipital alpha power showed similar effects, but for different brain regions (Caudate, Nucleus Accumbens, Hippocampus) (Figure S4). The significant SNPs were frontal cortical tissue eQTLs for MTERF4, GNL3, and ITIH4; the latter two being schizophrenia/bipolar disorder liability genes at 3p21. Significant SNPs for occipital power were cortical-tissue eQTLs for genes IL1RL1, IL18R1, CLHC1, GLYCK, and ITIH4.

To test for overall significance of gene-expression enrichment in alpha oscillation power, we used the online tool FUMA<sup>42</sup>. We extracted the top 500 genes from the gene-based association (Cz and occipital alpha), which were matched against genes significantly up- or downregulated in each GTEx tissue compared to the average of other tissues (i.e., differentially expressed genes determined by a Bonferroni-corrected t-test). Significance of enrichment was determined by the

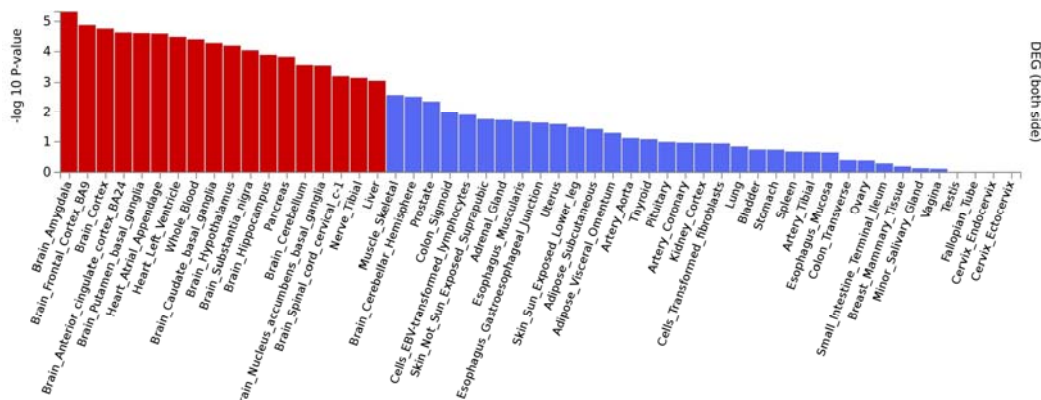
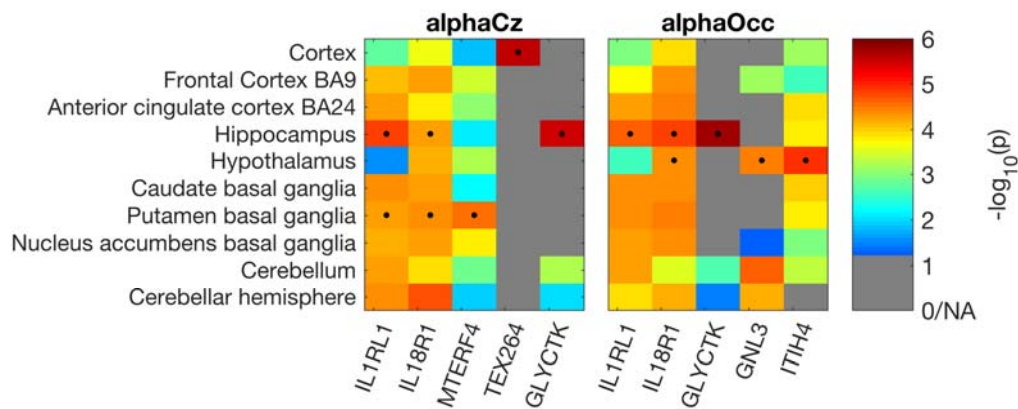


Figure 3. FUMA<sup>42</sup> enrichment analysis. The top 500 genes in the occipital alpha analysis are significantly enriched for genes that are up or down regulated in all cortical and subcortical brain tissues from the GTEx database, save cerebellar hemisphere. Other significantly enriched tissues are heart, whole blood, pancreas, tibial nerve, and liver.

hypergeometric test with Bonferroni correction. Figure 3 shows that brain derived tissues are almost invariably significant, and much more so than other tissues.

#### *Imputed gene-expression association of alpha oscillations*

To further elucidate tissue-expression pathways of the chromosome 2 cytokine receptor genes and the schizophrenia liability genes on chromosome 3 affecting alpha oscillation power we applied MetaXcan using the all GTEx brain tissues<sup>40,43</sup>. MetaXcan may have increased power to detect significant gene / phenotype associations by combining genetic variants in a sparse elastic net prediction model. Four genes near the 3p21 region showed at least one FDR significant association (ITIH4, GNL3, GLYCTK, and TEX264). Figure 4 shows the expression profiles for chromosome 3 genes with at least one FDR significant effect. ITIH4 showed a more widespread association across tissues (significant in hypothalamus), whereas GLYCTK showed a rather specific hippocampal expression for both occipital and Cz alpha power. Immune genes IL1RL1 and IL18R1 on chromosome 2 also reached the threshold for significance in the significant association with cortical and subcortical expression with alpha oscillations (see Figure 4).



*Figure 4. MetaXcan gene-expression results for chromosomes 2 and 3 indicate which positional gene-based discoveries and which brain tissues may be involved in affecting alpha oscillation power. Uncorrected p-values are shown, FDR significant discoveries are marked with a dot. Out of the 24 significant genes in region 3p21, GLYCTK associated with alpha power via hippocampal expression, TEX264 via general cortical expression, GNL3 and ITIH4 via hypothalamic expression. Furthermore, IL18R1, IL1RL1, and MTERF4 on chromosome 2 were not discovered in the positional analysis. These genes affected alpha oscillations via the Putamen and Hippocampus, but IL1RL1 and IL18R1 expression in many other tissues just failed to reach the FDR threshold. Note that FDR correction was performed within tissue.*



### *GABA<sub>A</sub> receptor genes*

Planned comparisons for two GABA<sub>A</sub> receptor genes at 4p12 were made in relation to beta power. The results from the KGG GATES positional gene-based analysis showed  $p=0.024$  for GABRA2 and  $p=0.15$  for GABRB1. After removing the results from the COGA sample—on which the previous positive findings were based—the  $p$ -value increased to  $p=0.052$  for GABRA2. MetaXcan analysis showed that imputed expression was associated with beta power for hippocampal expression of GABRA2 ( $p=0.0024$ ) and not with other tissues ( $p>0.2$ ).

### *SNP-based heritability*

LD score regression for Hapmap 3 annotated SNPs ( $N_{\text{SNP}}>819\text{k}$ ) was carried out for SNP effects from the meta-analysis. Heritability estimates (Figure S5) were lower than estimates from twin and family studies of EEG power (ranging from 0.45 to 0.9; e.g., see <sup>21</sup>), but are in line with the GCTA random-effects modeling on common SNPs for the same EEG measures <sup>26</sup>.

### *Genetic correlation analysis*

Bivariate LD score regression was used to calculate genetic correlation  $r_G$  between traits. Figure S6 shows the genetic correlations between the EEG phenotypes. Strong and positive genetic correlations were observed among the EEG power phenotypes. Occipital and Cz alpha power correlated 1.0, and correlated highly with beta power ( $r_G=.67$  and  $r_G=0.71$  respectively). Slow oscillatory power (delta and theta) also correlated near 1.0. Only the theta with beta power genetic correlation was modest and not statistically significant. Negative correlations between peak alpha frequency and the slow oscillation power phenotypes were observed, and a nonsignificant positive correlation with beta power.

LD score regression based genetic correlations with a wide range of human phenotypes with published GWAS results are shown in Figure S7. Significant effects were observed but failed to survive multiple testing correction (FDR or Bonferroni), which may be expected in light of the still relatively low sample size of the current GWAS and the subsequently large confidence intervals. Top effects included  $r_G=-0.35$  (uncorrected  $p=0.0094$ ) between theta power and autism spectrum disorder, and  $r_G=0.55$  (uncorrected  $p=0.014$ ) between beta power and generalized epilepsy.

## DISCUSSION

We have presented results from the first international consortium for investigating the molecular genetic basis of brain functional activity as measured by resting EEG. The results revealed two genome-wide significant associations for Cz alpha power: on chromosome 4, a SNP intronic to PRKG2 (rs984924), and on chromosome 7, a SNP intronic to LINC00996 (rs10231372). FDR correction yielded 68 significant SNPs in the same PRKG2 and LINC00996 regions, plus intronic variants within PCNX2 and METTL21C. Gene-based analyses identified multiple genes significantly associated with Cz and occipital alpha power, including PRKG2, METTL21C, and several genes in a region on chromosome 3. PRKG2 influences anthropometric and blood pressure-related traits<sup>44,45</sup> and also affects multiple phenotypes in mice, including skeletal and adipose tissues. Humans with 4q21 microdeletion syndrome—which includes PRKG2 and flanking genes—show similar skeletal symptoms, including facial bone and growth retardations, but also neuropsychological symptoms, including speech and mental retardation<sup>46,47</sup>.

KGG gene-based analyses implicated a high-LD region on chromosome 3 that included many significant genes associated with Cz and occipital alpha power. Variants in this region have been associated with schizophrenia and bipolar disorder. Significant brain-tissue eQTLs pointed to ITIH4, GNL3, and GLYCTK as genes with altered expression. MetaXcan significantly associated widespread brain expression for ITIH4, with hypothalamic expression reaching significance. For GNL3, hypothalamic and cerebellar tissues significantly associated with alpha oscillations. Hippocampal GLYCTK expression associated with Cz and occipital alpha. MetaXcan further associated cortical TEX264 expression. By using expression analyses, we were able to strongly reduce the number of target genes in the chromosome 3 region from twenty-four to four, and localize their effects to hypothalamic and hippocampal expression as most strongly associated with alpha oscillations.

The association schizophrenia liability genes with oscillatory brain activity and the specific tissues with significantly altered expression highlights where oscillatory brain activity changes with increased disease risk. Altered expression of ITIH4 in the frontal cortex in the context of schizophrenia has recently been reported<sup>48</sup>, and is consistent with reduced alpha oscillatory activity in the frontal cortex observed in schizophrenia<sup>13,49–52</sup>. FDR significant SNPs were eQTLs for ITIH4, GNL3, and MTERF4 in the frontal cortex for Cz alpha oscillations. Our results indicate that schizophrenia liability gene ITIH4 affects oscillatory brain function, and adds GNL3 and MTERF4 as possible target genes. Brain eQTLs further pointed to cytokine receptor genes IL1RL1 and IL18R1, which are immune system genes linked to asthma, celiac disease, IBS, and atopic dermatitis<sup>53–56</sup>. MetaXcan imputed

expression analysis indicated that these genes are also brain expressed, and associated with alpha oscillation power for widespread cortical and subcortical tissues, reaching significance for the Hippocampus and Putamen. The association of IL18R1 expression with schizophrenia was reported recently<sup>57</sup>. Our results indicate that these immunological liability genes also affect oscillatory brain function by altering widespread expression in the brain.

Results confirmed GABA<sub>A</sub> signaling as being involved in fast oscillatory (beta) activity in the full meta-analysis ( $p=0.024$ ). Expression analysis significantly associated hippocampal GABRA2 expression to beta oscillations ( $p=0.0024$ ). This result fits well with observations that beta oscillations are influenced by GABA<sub>A</sub> receptor  $\alpha$  2 agonists such as benzodiazepines<sup>58–60</sup>, and the crucial role of GABA<sub>A</sub> interneurons for synchronized fast rhythms in the brain<sup>61</sup>. In our view, there is now strong evidence that GABA mediates the relation between resting-state EEG beta power and alcohol dependence<sup>17,29,62</sup>. The selective hippocampal expression association suggests that the genetic variants affecting beta oscillations also affect hippocampal GABA<sub>A</sub> receptor's sensitivity to interneuron inhibition.

Twin and family studies have consistently indicated that EEG alpha power is one of the most heritable traits in humans at up to 96% for frontal alpha power in young adult samples<sup>20,21,24,63</sup>. The SNP heritability observed here using LD score regression was only able to retrieve a relatively small proportion of variance of the often highly heritable EEG traits. This discrepancy could be caused by a relative large contribution of rare SNPs that are poorly tagged by the common SNP arrays used here. The SNP-based genetic correlation analysis was more consistent with twin/family studies. Strong genetic correlations ( $>.70$ ) were observed among the slower (delta theta) and among faster oscillations (alpha beta). Results from twin studies generally ranged from 0.50 to 0.90, although the twin-based  $r_G$  between theta and delta oscillation power is generally not as strong as the SNP  $r_G$  observed here. This inconsistency between twin/family and SNP coheritability could perhaps be explained by the restricted scalp locations tested in the current analysis and/or the sample heterogeneity (e.g., COGA selected for alcohol use disorders).

Coheritability analysis showed a nominally significant genetic correlation ( $r_G=0.55$ ) between beta oscillations and generalized epilepsy. This is consistent with the putative role of fast beta/gamma oscillations in ictogenesis (in the present sample only nonaffected individuals were used). The largest epilepsy GWAS to date found suggestive evidence for the involvement of GABRA2<sup>64</sup>, which we found to be related to beta oscillation power. GABA is a main antiepileptic drug target, and is known to affect (motor) beta EEG via GABAergic modulation of pyramidal cells<sup>65–68</sup>. We

additionally observed a significant genetic correlation between autism and theta oscillations ( $r_G = -0.35$ ,  $p = 0.009$ ). Although deviant brain function in autism is most consistently found in the lower gamma band due to altered GABA inhibitory neuronal action<sup>69–72</sup>, cortical and hippocampal theta/lower alpha are known to show phase-amplitude coupling with fast oscillations (gamma<sup>73–75</sup>). The significant genetic correlation ( $r_G = 0.22$ ,  $p = 0.019$ ) between heart rate and alpha power is consistent with observations in concurrent EEG and ECG recordings<sup>76</sup>.

In sum, we have found evidence that (hippocampal) GABA receptor alpha 2 subunit is involved in altering beta power, possibly via hippocampal expression—consistent with its relation to epilepsy and alcohol dependence that are both well known for the involvement of GABA<sub>A</sub><sup>29,77</sup>. Schizophrenia liability genes on chromosome 2 and 3 affected alpha oscillation power. SNPs in the chromosome 3 region were eQTLs for ITIH4, GNL3, and GLYCTK. Significant eQTLs were tissue specific, including the frontal cortex, anterior cingulate cortex, hypothalamus, and hippocampus. Expression analysis further targeted immune system genes IL1RL1 and IL18R1 with altered expression in Putamen and Hippocampus.

GWAS is dependent on very large sample sizes as the effects of the tagging genetic variants (SNPs) are small, even for brain endophenotypes<sup>78,79</sup>. Our results suggest that EEG power GWAS may indeed be somewhat more powerful than of complex behavioral phenotypes, which generally require larger sample sizes. Current sample sizes may be adequate to reveal significant individual genetic associations, pathways for the expression of psychiatric liability in the brain, and prove hopeful for future GWAS of additional EEG parameters. For example, two recently published GWASs of bipolar EEG from families of African and European ancestry reported genome-wide signal at 3q26 and 6q22, respectively<sup>31,80</sup>. Bipolar EEG derivations show more localized activity than other EEG derivations and remove volume conduction effects, and have been particularly successful as a biomarker of alcohol dependence. Other EEG parameters of high interest are functional connectivity as biomarker for various neurodevelopmental and psychiatric disorders. The current results indicate that finding genetic variants or genes related to these EEG parameters is entirely feasible.

## METHODS

### *Subjects*

Resting state EEG and genome-wide genotyping were available for a total of 8,425 individuals from Australia, the Netherlands, and the USA. All subjects were part of

twin and family studies into the genetics of health and behavioral traits with additional psychophysiological assessments: the Minnesota Twin Family Study (MTFS), the Collaborative Study on the Genetics of Alcoholism (COGA), the Brisbane Adolescent Twin Study (BATS), and the Netherlands Twin Register (NTR). All sites excluded subjects with a history of neurological problems, including tumor and head trauma. Alcohol dependence ascertained samples (COGA EA and COGA case control) required alcohol testing before EEG recording. Full details and demographics for each cohort are given in the supplementary information.

#### *EEG recording and preprocessing*

For details on EEG assessment by the individual groups see supplementary material.

#### *EEG Power and Peak frequency analysis*

All groups analyzed the vertex recordings (Cz) and the average of Occipital leads (O1, O2) in standard frequency bands. Cleaned data were imported into MATLAB, epoched into 2s epochs, and power spectra calculated using FFT. Frequency bins were defined as delta (1-3.75Hz), theta (4-7.75 Hz), alpha (8-12.75Hz), and beta (15-25 Hz). Power was defined as the squared radius of the orthogonal sine and cosine amplitudes averaged over window size, and the mean value taken for the frequency band to obtain power density. Alpha peak frequency was determined using the power-weighted method in accordance with <sup>26</sup> between 7 and 14 Hz.

#### *QC and Genome-wide association analysis*

Groups used dosage based imputed SNP sets using CEU reference panels hg19/build 37 from 1000 genomes phase1 or phase3. Imputation followed ENIGMA imputation protocols <sup>32</sup> ([http://enigma.usc.edu/wp-content/uploads/2012/07/ENIGMA2\\_1KGP\\_cookbook\\_v3.pdf](http://enigma.usc.edu/wp-content/uploads/2012/07/ENIGMA2_1KGP_cookbook_v3.pdf)). Association analyses accounted for family relatedness. Sex, Age and age<sup>2</sup> were used as covariates plus Principal Components plus disease status (when applicable). See supplementary materials for more group specific methods and/or deviance from these standard analyses protocols, and covariate analysis results.

Pre-meta-analysis QC were performed using EasyQC <sup>81</sup>. We filtered on sample MAF (0.03 for the largest dataset, MTFS; 0.04 for the intermediate datasets, COGA case control and QIMR BATS; and 0.05 for NTR and COGA EA), as well as EUR 1000 Genomes reference set MAF (<0.03), N<200, HWE  $p < 10^{-7}$ , INFO<0.8, INFO>1.05, imputation  $R^2 < 0.4$ , invalid numbers (Inf, NA), 0.2 difference between sample and reference set allele frequencies. Further checks consisted of matching

alleles, duplicates, and strand flips. Meta-analyzed SNPs were filtered for combined  $N > 6000$ . The final datasets consisted of 4959085 to 4959521 SNPs depending on phenotype.

#### *SNP heritability*

LD score regression<sup>82</sup> uses the natural experiment present in the genome due to variable amounts of Linkage Disequilibrium (LD) between SNPs. Causal variants will cause straight slope decline in test statistics of nearby SNPs with decreasing levels of LD to the causal variant in the case of additive genetic variation. The slope of the regression line of the chi-square statistic against LD scores across the genome reflects the heritability of the trait<sup>82</sup>. SNP heritability estimation using LD score regression has the advantage of being insensitive to population stratification effects, as these will result in an upward shift across all LD score bins, thus affecting the intercept and not the slope of the LD-dependent regression.

We used LD score regression to estimate the SNP based heritability of the six phenotypes following the recommendations in<sup>82</sup>, including pruning for Hapmap 3 SNPs. Next, the LD-score regression intercept was used to assess quality of the GWAS and removal of stratification effects by the population Principal Components (see for example<sup>83</sup>). Finally, we used bivariate LD-score regression to estimate genetic correlation  $r_G$ , between the EEG phenotypes and GWASs available in LD Hub (<http://ldsc.broadinstitute.org>)<sup>84,85</sup>. This includes GWASs on schizophrenia, bipolar disorder, subjective wellbeing, and neuroticism, and ENIGMA subcortical volumes, intracranial volume, and brain volume (See Supplementary Methods for a full reference list). We extended these with generalized epilepsy and educational attainment<sup>64,86</sup>.

## References

- 1 Başar E. A review of alpha activity in integrative brain function: Fundamental physiology, sensory coding, cognition and pathology. *Int J Psychophysiol* 2012; **86**: 1–24.
- 2 Buzsáki G. *Rhythms of the brain*. Oxford University Press US, 2006.
- 3 Jokisch D, Jensen O. Modulation of Gamma and Alpha Activity during a Working Memory Task Engaging the Dorsal or Ventral Stream. *J Neurosci* 2007; **27**: 3244–3251.
- 4 Pandey AK, Kamarajan C, Manz N, Chorlian DB, Stimus A, Porjesz B. Delta, theta, and alpha event-related oscillations in alcoholics during Go/NoGo task: Neurocognitive deficits in execution, inhibition, and attention processing. *Prog Neuropsychopharmacol Biol Psychiatry* 2016; **65**: 158–171.
- 5 Uhlhaas PJ, Singer W. Abnormal neural oscillations and synchrony in schizophrenia. *Nat Rev Neurosci* 2010; **11**: 100–113.
- 6 Marín O. Interneuron dysfunction in psychiatric disorders. *Nat Rev Neurosci* 2012; **13**: 107.
- 7 Cardin JA, Carlén M, Meletis K, Knoblich U, Zhang F, Deisseroth K *et al*. Driving fast-spiking cells induces gamma rhythm and controls sensory responses. *Nature* 2009; **459**: 663–667.
- 8 Cho KKA, Hoch R, Lee AT, Patel T, Rubenstein JLR, Sohal VS. Gamma Rhythms Link Prefrontal Interneuron Dysfunction with Cognitive Inflexibility in Dlx5/6+/- Mice. *Neuron* 2015; **85**: 1332–1343.
- 9 de Geus E. From genotype to EEG endophenotype: a route for post-genomic understanding of complex psychiatric disease? *Genome Med* 2010; **2**: 1–4.
- 10 Gottesman II, Gould TD. The endophenotype concept in psychiatry: etymology and strategic intentions. *Am J Psychiatry* 2003; **160**: 636–645.
- 11 Doppelmayr M, Klimesch W, Stadler W, Pöllhuber D, Heine C. EEG alpha power and intelligence. *Intelligence* 2002; **30**: 289–302.
- 12 Thatcher RW, North D, Biver C. Intelligence and EEG current density using low-resolution electromagnetic tomography (LORETA). *Hum Brain Mapp* 2007; **28**: 118–133.
- 13 Sponheim SR, Clementz BA, Iacono WG, Beiser M. Resting EEG in first-episode and chronic schizophrenia. *Psychophysiology* 1994; **31**: 37–43.
- 14 Boutros NN, Arfken C, Galderisi S, Warrick J, Pratt G, Iacono W. The status of spectral EEG abnormality as a diagnostic test for schizophrenia. *Schizophr Res* 2008; **99**: 225–237.
- 15 Clarke AR, Barry RJ, McCarthy R, Selikowitz M. EEG analysis in Attention-Deficit/Hyperactivity Disorder: a comparative study of two subtypes. *Psychiatry Res* 1998; **81**: 19–29.

- 16 Snyder SM, Hall JR. A meta-analysis of quantitative EEG power associated with attention-deficit hyperactivity disorder. *J Clin Neurophysiol Off Publ Am Electroencephalogr Soc* 2006; **23**: 440–455.
- 17 Rangaswamy M, Porjesz B, Chorlian DB, Wang K, Jones KA, Bauer LO *et al*. Beta power in the EEG of alcoholics. *Biol Psychiatry* 2002; **52**: 831–842.
- 18 Struve FA, Straumanis JJ, Patrick G, Price L. Topographic Mapping of Quantitative EEG Variables in Chronic Heavy Marijuana Users: Empirical Findings with Psychiatric Patients. *Clin Electroencephalogr* 1989; **20**: 6–23.
- 19 Minzenberg MJ, Firl AJ, Yoon JH, Gomes GC, Reinking C, Carter CS. Gamma Oscillatory Power is Impaired During Cognitive Control Independent of Medication Status in First-Episode Schizophrenia. *Neuropsychopharmacology* 2010; **35**: 2590–2599.
- 20 Anokhin AP, Van Baal G, Van Beijsterveldt C, De Geus E, Grant J, Boomsma D. Genetic correlation between the P300 event-related brain potential and the EEG power spectrum. *Behav Genet* 2001; **31**: 545–554.
- 21 Smit D, Posthuma D, Boomsma D, Geus EJC. Heritability of background EEG across the power spectrum. *Psychophysiology* 2005; **42**: 691–697.
- 22 Tang Y, Chorlian DB, Rangaswamy M, Porjesz B, Bauer L, Kuperman S *et al*. Genetic influences on bipolar EEG power spectra. *Int J Psychophysiol* 2007; **65**: 2–9.
- 23 Van Beijsterveldt C, Molenaar P, De Geus E, Boomsma D. Heritability of human brain functioning as assessed by electroencephalography. *Am J Hum Genet* 1996; **58**: 562.
- 24 Zietsch BP, Hansen JL, Hansell NK, Geffen GM, Martin NG, Wright MJ. Common and specific genetic influences on EEG power bands delta, theta, alpha, and beta. *Biol Psychol* 2007; **75**: 154–164.
- 25 Smit CM, Wright MJ, Hansell NK, Geffen GM, Martin NG. Genetic variation of individual alpha frequency (IAF) and alpha power in a large adolescent twin sample. *Int J Psychophysiol* 2006; **61**: 235–243.
- 26 Malone SM, Burwell SJ, Vaidyanathan U, Miller MB, McGue M, Iacono WG. Heritability and molecular-genetic basis of resting EEG activity: A genome-wide association study. *Psychophysiology* 2014; **51**: 1225–1245.
- 27 Yang J, Benyamin B, McEvoy BP, Gordon S, Henders AK, Nyholt DR *et al*. Common SNPs explain a large proportion of the heritability for human height. *Nat Genet* 2010; **42**: 565–569.
- 28 Porjesz B, Almasy L, Edenberg HJ, Wang K, Chorlian DB, Foroud T *et al*. Linkage disequilibrium between the beta frequency of the human EEG and a GABAA receptor gene locus. *Proc Natl Acad Sci* 2002; **99**: 3729–3733.
- 29 Edenberg HJ, Dick DM, Xuei X, Tian H, Almasy L, Bauer LO *et al*. Variations in GABRA2, Encoding the  $\alpha 2$  Subunit of the GABAA Receptor, Are Associated with Alcohol Dependence and with Brain Oscillations. *Am J Hum Genet* 2004; **74**: 705–714.



- 30 Lydall GJ, Saini J, Ruparelia K, Montagnese S, McQuillin A, Guerrini I *et al.* Genetic association study of GABRA2 single nucleotide polymorphisms and electroencephalography in alcohol dependence. *Neurosci Lett* 2011; **500**: 162–166.
- 31 Meyers JL, Zhang J, Wang JC, Su J, Kuo SI, Kapoor M *et al.* An endophenotype approach to the genetics of alcohol dependence: a genome wide association study of fast beta EEG in families of African ancestry. *Mol Psychiatry* 2017. doi:10.1038/mp.2016.239.
- 32 Thompson PM, Stein JL, Medland SE, Hibar DP, Vasquez AA, Renteria ME *et al.* The ENIGMA Consortium: large-scale collaborative analyses of neuroimaging and genetic data. *Brain Imaging Behav* 2014; **8**: 153–182.
- 33 Thompson PM, Andreassen OA, Arias-Vasquez A, Bearden CE, Boedhoe PS, Brouwer RM *et al.* ENIGMA and the individual: Predicting factors that affect the brain in 35 countries worldwide. *NeuroImage* 2017; **145, Part B**: 389–408.
- 34 Thompson PM, Andreassen OA, Arias-Vasquez A, Bearden CE, Boedhoe PS, Brouwer RM *et al.* ENIGMA and the individual: Predicting factors that affect the brain in 35 countries worldwide. *NeuroImage* doi:10.1016/j.neuroimage.2015.11.057.
- 35 Ripke S, O’Dushlaine C, Chambert K, Moran JL, Kähler AK, Akterin S *et al.* Genome-wide association analysis identifies 13 new risk loci for schizophrenia. *Nat Genet* 2013; **45**: 1150–1159.
- 36 Sklar P, Ripke S, Scott LJ, Andreassen OA, Cichon S, Craddock N *et al.* Large-scale genome-wide association analysis of bipolar disorder identifies a new susceptibility locus near ODZ4. *Nat Genet* 2011; **43**: 977–983.
- 37 Ripke S, Neale BM, Corvin A, Walters JT, Farh K-H, Holmans PA *et al.* Biological Insights From 108 Schizophrenia-Associated Genetic Loci. *Nature* 2014; **511**: 421–427.
- 38 Ramasamy A, Trabzuni D, Guelfi S, Varghese V, Smith C, Walker R *et al.* Genetic variability in the regulation of gene expression in ten regions of the human brain. *Nat Neurosci* 2014; **17**: 1418–1428.
- 39 Gamazon ER, Badner JA, Cheng L, Zhang C, Zhang D, Cox NJ *et al.* Enrichment of cis-regulatory gene expression SNPs and methylation quantitative trait loci among bipolar disorder susceptibility variants. *Mol Psychiatry* 2013; **18**: 340–346.
- 40 Gamazon ER, Wheeler HE, Shah KP, Mozaffari SV, Aquino-Michaels K, Carroll RJ *et al.* A gene-based association method for mapping traits using reference transcriptome data. *Nat Genet* 2015; **47**: 1091–1098.
- 41 Lonsdale J, Thomas J, Salvatore M, Phillips R, Lo E, Shad S *et al.* The Genotype-Tissue Expression (GTEx) project. *Nat Genet* 2013; **45**: 580–585.
- 42 Watanabe K, Taskesen E, Bochoven A van, Posthuma D. FUMA: Functional mapping and annotation of genetic associations. *bioRxiv* 2017; : 110023.
- 43 Barbeira A, Shah KP, Torres JM, Wheeler HE, Torstenson ES, Edwards T *et al.* MetaXcan: Summary Statistics Based Gene-Level Association Method Infers Accurate PrediXcan Results. *bioRxiv* 2016; : 45260.

- 44 Sung YJ, de las Fuentes L, Schwander KL, Simino J, Rao DC. Gene–Smoking Interactions Identify Several Novel Blood Pressure Loci in the Framingham Heart Study. *Am J Hypertens* 2015; **28**: 343–354.
- 45 Wood AR, Esko T, Yang J, Vedantam S, Pers TH, Gustafsson S *et al.* Defining the role of common variation in the genomic and biological architecture of adult human height. *Nat Genet* 2014; **46**: 1173–1186.
- 46 Bonnet C, Andrieux J, Béri-Dexheimer M, Leheup B, Boute O, Manouvrier S *et al.* Microdeletion at chromosome 4q21 defines a new emerging syndrome with marked growth restriction, mental retardation and absent or severely delayed speech. *J Med Genet* 2010; **47**: 377–384.
- 47 Dukes-Rimsky L, Guzauskas GF, Holden KR, Griggs R, Ladd S, del Carmen Montoya M *et al.* Microdeletion at 4q21.3 is associated with intellectual disability, dysmorphic facies, hypotonia, and short stature. *Am J Med Genet A* 2011; **155**: 2146–2153.
- 48 Ohi K, Shimada T, Nitta Y, Kihara H, Okubo H, Uehara T *et al.* Schizophrenia risk variants in ITIH4 and CALN1 regulate gene expression in the dorsolateral prefrontal cortex. *Psychiatr Genet* 2016; **26**: 142–143.
- 49 Iacono WG. Bilateral electrodermal habituation-dishabituation and resting EEG in remitted schizophrenics. *J Nerv Ment Dis* 1982; **170**: 91–101.
- 50 Itil TM, Saletu B, Davis S, Allen M. Stability studies in schizophrenics and normals using computer-analyzed EEG. *Biol Psychiatry* 1974; **8**: 321–335.
- 51 Klimesch W, Sauseng P, Hanslmayr S. EEG alpha oscillations: The inhibition–timing hypothesis. *Brain Res Rev* 2007; **53**: 63–88.
- 52 Sponheim SR, Clementz BA, Iacono WG, Beiser M. Clinical and biological concomitants of resting state EEG power abnormalities in schizophrenia. *Biol Psychiatry* 2000; **48**: 1088–1097.
- 53 Barreto-Luis A, Pino-Yanes M, Corrales A, Campo P, Callero A, Acosta-Herrera M *et al.* Genome-wide association study in Spanish identifies ADAM metallopeptidase with thrombospondin type 1 motif, 9 (ADAMTS9), as a novel asthma susceptibility gene. *J Allergy Clin Immunol* 2016; **137**: 964–966.
- 54 Dubois PCA, Trynka G, Franke L, Hunt KA, Romanos J, Curtotti A *et al.* Multiple common variants for celiac disease influencing immune gene expression. *Nat Genet* 2010; **42**: 295–302.
- 55 Liu JZ, van Sommeren S, Huang H, Ng SC, Alberts R, Takahashi A *et al.* Association analyses identify 38 susceptibility loci for inflammatory bowel disease and highlight shared genetic risk across populations. *Nat Genet* 2015; **47**: 979–986.
- 56 Paternoster L, Standl M, Waage J, Baurecht H, Hotze M, Strachan DP *et al.* Multi-ethnic genome-wide association study of 21,000 cases and 95,000 controls identifies new risk loci for atopic dermatitis. *Nat Genet* 2015; **47**: 1449–1456.
- 57 Xu Y, Yue W, Shugart YY, Yuan J, Wang G, Wang HZ *et al.* Potential involvement of the interleukin-18 pathway in schizophrenia. *J Psychiatr Res* 2016; **74**: 10–16.
- 58 Manmaru S, Matsuura M. Quantification of benzodiazepine-induced topographic EEG changes by a computerized wave form recognition method: application of a

- principal component analysis. *Electroencephalogr Clin Neurophysiol* 1989; **72**: 126–132.
- 59 Montagu JD. Effects of diazepam on the EEG in man. *Eur J Pharmacol* 1972; **17**: 167–170.
- 60 van Lier H, Drinkenburg WHIM, van Eeten YJW, Coenen AML. Effects of diazepam and zolpidem on EEG beta frequencies are behavior-specific in rats. *Neuropharmacology* 2004; **47**: 163–174.
- 61 Buzsáki G, Chrobak JJ. Temporal structure in spatially organized neuronal ensembles: a role for interneuronal networks. *Curr Opin Neurobiol* 1995; **5**: 504–510.
- 62 Dick DM, Bierut L, Hinrichs A, Fox L, Bucholz KK, Kramer J *et al*. The Role of GABRA2 in Risk for Conduct Disorder and Alcohol and Drug Dependence across Developmental Stages. *Behav Genet* 2006; **36**: 577–590.
- 63 van Beijsterveldt CEM, van Baal GCM. Twin and family studies of the human electroencephalogram: a review and a meta-analysis. *Biol Psychol* 2002; **61**: 111–138.
- 64 ILAE Consortium. Genetic determinants of common epilepsies: a meta-analysis of genome-wide association studies. *Lancet Neurol* 2014; **13**: 893–903.
- 65 Gaetz W, Edgar JC, Wang DJ, Roberts TPL. Relating MEG measured motor cortical oscillations to resting  $\gamma$ -Aminobutyric acid (GABA) concentration. *NeuroImage* 2011; **55**: 616–621.
- 66 Rowland LM, Edden RAE, Kontson K, Zhu H, Barker PB, Hong LE. GABA Predicts Inhibition of Frequency-Specific Oscillations in Schizophrenia. *J Neuropsychiatry Clin Neurosci* 2013; **25**: 83–87.
- 67 Hall SD, Barnes GR, Furlong PL, Seri S, Hillebrand A. Neuronal Network Pharmacodynamics of GABAergic Modulation in the Human Cortex Determined Using Pharmaco-Magnetoencephalography. *Hum Brain Mapp* 2010; **31**: 581–594.
- 68 Yamawaki N, Stanford IM, Hall SD, Woodhall GL. Pharmacologically induced and stimulus evoked rhythmic neuronal oscillatory activity in the primary motor cortex in vitro. *Neuroscience* 2008; **151**: 386–395.
- 69 Blatt GJ, Fatemi SH. Alterations in GABAergic biomarkers in the autism brain: research findings and clinical implications. *Anat Rec Hoboken NJ* 2007 2011; **294**: 1646–1652.
- 70 Gandal MJ, Edgar JC, Ehrlichman RS, Mehta M, Roberts TPL, Siegel SJ. Validating  $\gamma$  oscillations and delayed auditory responses as translational biomarkers of autism. *Biol Psychiatry* 2010; **68**: 1100–1106.
- 71 Nelson SB, Valakh V. Excitatory/Inhibitory Balance and Circuit Homeostasis in Autism Spectrum Disorders. *Neuron* 2015; **87**: 684–698.
- 72 Rojas DC, Wilson LB. Gamma-band abnormalities as markers of autism spectrum disorders. *Biomark Med* 2014; **8**: 353–368.
- 73 Canolty RT, Edwards E, Dalal SS, Soltani M, Nagarajan SS, Kirsch HE *et al*. High gamma power is phase-locked to theta oscillations in human neocortex. *Science* 2006; **313**: 1626–1628.

- 74 Khan S, Gramfort A, Shetty NR, Kitzbichler MG, Ganesan S, Moran JM *et al.* Local and long-range functional connectivity is reduced in concert in autism spectrum disorders. *Proc Natl Acad Sci* 2013; **110**: 3107–3112.
- 75 Lisman JE, Jensen O. The Theta-Gamma Neural Code. *Neuron* 2013; **77**: 1002–1016.
- 76 de Munck JC, Gonçalves SI, Faes TJC, Kuijter JPA, Pouwels PJW, Heethaar RM *et al.* A study of the brain's resting state based on alpha band power, heart rate and fMRI. *NeuroImage* 2008; **42**: 112–121.
- 77 Jurkiewicz MT, Gaetz WC, Bostan AC, Cheyne D. Post-movement beta rebound is generated in motor cortex: Evidence from neuromagnetic recordings. *NeuroImage* 2006; **32**: 1281–1289.
- 78 Stein JL, Medland SE, Vasquez AA, Hibar DP, Senstad RE, Winkler AM *et al.* Identification of common variants associated with human hippocampal and intracranial volumes. *Nat Genet* 2012; **44**: 552–561.
- 79 Hibar DP, Stein JL, Renteria ME, Arias-Vasquez A, Desrivieres S, Jahanshad N *et al.* Common genetic variants influence human subcortical brain structures. *Nature* 2015; **520**: 224–229.
- 80 Meyers JL, Zhang J, Manz N, Rangaswamy M, Kamarajan C, Wetherill L *et al.* A genome wide association study of fast beta EEG in families of European ancestry. *Int J Psychophysiol* 2017; **115**: 74–85.
- 81 Winkler TW, Day FR, Croteau-Chonka DC, Wood AR, Locke AE, Mägi R *et al.* Quality control and conduct of genome-wide association meta-analyses. *Nat Protoc* 2014; **9**: 1192–1212.
- 82 Bulik-Sullivan BK, Loh P-R, Finucane HK, Ripke S, Yang J, Schizophrenia Working Group of the Psychiatric Genomics Consortium *et al.* LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat Genet* 2015; **47**: 291–295.
- 83 Okbay A, Baselmans BML, De Neve J-E, Turley P, Nivard MG, Fontana MA *et al.* Genetic variants associated with subjective well-being, depressive symptoms, and neuroticism identified through genome-wide analyses. *Nat Genet* 2016; **48**: 624–633.
- 84 Bulik-Sullivan B, Finucane HK, Anttila V, Gusev A, Day FR, Loh P-R *et al.* An atlas of genetic correlations across human diseases and traits. *Nat Genet* 2015; **47**: 1236–1241.
- 85 Zheng J, Erzurumluoglu AM, Elsworth BL, Kemp JP, Howe L, Haycock PC *et al.* LD Hub: a centralized database and web interface to perform LD score regression that maximizes the potential of summary level GWAS data for SNP heritability and genetic correlation analysis. *Bioinformatics* 2017; **33**: 272–279.
- 86 Okbay A, Beauchamp JP, Fontana MA, Lee JJ, Pers TH, Rietveld CA *et al.* Genome-wide association study identifies 74 loci associated with educational attainment. *Nature* 2016; **533**: 539–542.

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