Diversity of coelomycetous fungi in human infections: a 10-year experience of two European reference centres.

Dea Garcia-Hermoso[1], Nicomedes Valenzuela-Lopez[2,3], Olga Rivero-Menendez[4], Ana Alastruey-Izquierdo[4], Josep Guarro[2], José F. Cano-Lira[2#], Alberto M. Stchigel[2] and the French Mycoses Study Group

[1]Institut Pasteur, CNRS, National Reference Center for Invasive Mycoses and Antifungals (NRCMA), Molecular Mycology Unit, UMR2000, Paris, France.

[2] Mycology Unit, Medical School and IISPV, University Rovira i Virgili, C/ Sant Llorenç 21, 43201 Reus, Spain.

[3]Microbiology Unit, Medical Technology Department, Faculty of Health Science, University of Antofagasta, Chile.

[4]Mycology Reference Laboratory, Spanish National Center for Microbiology, Instituto de Salud Carlos III, Madrid, Spain

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[#]Corresponding author. E-mail: jose.cano@urv.cat Unitat de Micologia, Facultat de Medicina i Ciències de la Salut, IISPV, Universitat Rovira i Virgili, 21 Sant Llorenç St., 43201, Reus, Spain.

D.G-H. and N.V-L contributed equally to this article.

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ABSTRACT

The coelomycetous fungi are difficult to properly identify from their phenotypic characterization and their role as etiologic agents of human infections is not clear. We studied the species distribution of these fungi among clinical isolates that had been collected and stored over a ten-year period in two European reference laboratories (France and Spain). We identified phenotypically and molecularly 97 isolates by sequencing the D1-D2 fragment of the 28S nrRNA (LSU) gene. Species of the orders *Pleosporales* and *Glomerellales* were present in both collections, and *Botryosphaeriales* and *Diaporthales* only in the French one. The most prevalent species were *Medicopsis romeroi*, *Neocucurbitaria keratinophila*, *Neocucurbitaria unguis-hominis* and *Paraconiothyrium cyclothyrioides*, which had been recovered primarily from superficial tissues. The *Didymellaceae* was the most common family represented, with 27 isolates distributed into five genera.

Keywords: *Colletotrichum*, coelomycetes, coelomycetous fungi, *Didymella*, *Medicopsis*, mycosis, *Neocucurbitaria*, *Paraconiothyrium*, *Phoma*.

INTRODUCTION

Human infections by coelomycetous fungi are rare and poorly characterized due to the difficulty in identifying these fungi using only phenotypic tools. The coelomycetous fungi are characterized by the production of conidia into fruiting bodies (= conidiomata), and were originally included in the orders *Sphaeropsidales* and *Melanconiales* of the class Coelomycetes, taxa which today lack scientific validity due to the demonstrated polyphyletic character of this sort of fungus (1–3). They cause superficial or subcutaneous infections, mostly following a traumatic inoculation of contaminated plant material or soil particles during agricultural work in tropical and subtropical areas (4–6). The most common coelomycetous fungi involved in these infections are the etiologic agents of black-grain eumycetoma, such as *Biatriospora mackinnonii; Falciformispora* spp., *Medicopsis romeroi*, and *Pseudochaetosphaeronema larense*. Other common coelomycetous fungi include *Lasiodiplodia theobromae* and *Neoscytalidium dimidiatum* (synanamorph of *Hendersonula toruloidea*) (7–11), which typically cause onychomycosis, subcutaneous phaeohyphomycosis (12–15), and eumycetoma (16). In addition, many species of *Phoma* and *Pyrenochaeta* have been reported as occasional agents of localized and systemic infections in humans (9, 17–20). The taxonomy of several coelomycetous genera mentioned before have been revised recently but they still constitute a group of highly polyphyletic taxa that are usually difficult to identify phenotypically (2, 21–24).

In a recent study conducted in the USA, Valenzuela-Lopez *et al*. (6) identified 230 fungal strains by sequencing the D1-D2 domains of the 28S rRNA gene (LSU), from which 152 (66.1%) strains belonged to the order *Pleosporales*, the rest being distributed in several orders of the phylum Ascomycota. Most of these strains were recovered from superficial tissue. *Neoscytalidium dimidiatum, Paraconiothyrium*

66 *cyclothyrioides* and members of the family *Didymellaceae* were the most prevalent taxa.
67 In addition, those authors demonstrated the usefulness of the LSU as a good molecular
68 marker for a preliminary identification of coelomycetous fungi at genus level. In fact,
69 such locus is easily amplified and many sequences are available in the GenBank
70 database. However, the nucleotide sequences of more phylogenetically informative
71 genes need analysing in order to identify the fungi at species level. Genes such as the
72 RNA polymerase II subunit 2 (*rpb*2), translation elongation factor 1-alpha (*tef*1), beta-
73 tubulin (*tub*2) and the ribosomal internal transcribed spacer region (ITS), combined in a
74 multi-locus analysis, have all been recommended for this purpose (25)

75 Until now, the coelomycetous fungi involved in invasive fungal infections (IFIs)
76 are poorly known in Europe, probably due to the infrequency of these fungi and the
77 complexity of their identification in the absence of characteristic fruiting bodies when
78 grown on culture media used in the clinical lab. In a recent French study, eighteen
79 proven cases of cutaneous and subcutaneous primary infections by coelomycetous fungi
80 were reported and analysed in patients from tropical and subtropical regions (26).

81 For a better knowledge of the diversity of coelomycetous fungi involved in
82 human infections, we studied a large set of clinical isolates that had been identified in
83 two mycology reference centres in France and Spain, and determined their *in vitro*
84 antifungal susceptibility pattern.

85

RESULTS

Locations of infections

The majority of the isolates were recovered from superficial tissue, mainly skin (44%; 43/97), eyes (27%; 26/97), nails/hairs (18%; 17/97) and mouth/sinus (2%; 2/97). A few were recovered from deeper sites: bones (4%, 4/97), blood (2%, 2/97), cerebrospinal fluid (n=1), bone marrow (n=1) and lung (n=1) (Table 1 & 2).

Phylogenetic analyses

The maximum-likelihood (ML) phylogenetic analysis of the LSU sequences (approximately 584 pb) demonstrated that the 97 isolates were distributed into four orders, but scattered into fourteen clades (Fig. 1). Most of the isolates (81%; 78/97) belonged to the order *Pleosporales*, which were distributed into nine clades corresponding to 23 species of twelve genera, followed by those of the *Botryosphaeriales* (8%; 8/97), the *Diaporthales* (6%; 6/97) and the *Glomerellales* (5%; 5/97).

The most common species identified was *Medicopsis romeroi* (11%; 11/97), followed by *Paraconiothyrium cyclothyrioides*, *Neocucurbitaria keratinophila* and *N. unguis-hominis* (8% each; 8/97). These species were mostly isolated from cutaneous lesions (Table 2).

Clade 1 of the *Pleosporales* corresponded to the family *Didymellaceae*, which included 27 isolates distributed into five genera, morphologically characterized by their production of pycnidial conidiomata and hyaline, aseptate conidia. The five genera were *Didymella*, *Epicoccum*, *Neoascochyta*, *Phoma* and *Xenodidymella*. *Didymella* was represented by 13 isolates, six of them clustering with the type strain of *D. gardeniae* (CBS 626.68), and the other seven clustered with a reference strain of *D. glomerata*

(CBS 528.66). The genus *Epicoccum* grouped five of the isolates, three of them clustering with a reference strain of *E. sorghinum* (CBS 179.80) and the other two with the type strain of the type species of the genus, *E. nigrum* (CBS 173.73). The genus *Phoma* was represented by seven clinical isolates and a reference strain of *Phoma herbarum* (CBS 615.75). Two additional isolates included in this clade (CNRMA 16.76 and CNM-CM 6201) grouped with the type strains of *Xenodidymella saxea* (CBS 419.92) and *Neoascochyta desmazieri* (CBS 297.69), respectively.

Clade 2 had two species of *Preussia*: CNM-CM 7335 grouped with a reference strain of *P. typharum* (CBS 107.69), while CNM-CM 7343 represented an unknown species forming a sister clade with the type strain of *P. terricola* (CBS 317.65).

Clade 3 grouped three isolates of *Paraphoma*, one of them (CNM-CM 8075) clustered with the type strain of *P. fimeti* (CBS 170.70), and the remaining two (CNRMA 15.665 and CNRMA 9.467) representing unidentified phoma-like species.

Clade 4 had two sister clades of the genus *Tintelnotia*, which produced pycnidia and hyaline, aseptate conidia. The isolate CNM-CM 7430 was identified as *T. destructans*. However, the other two isolates (CNM-CM 7080 and CNM-CM 7981) did not cluster with any known species of the genus and might represent new species.

Clade 5 had 20 isolates of *Neocucurbitaria*. *Neocucurbitaria keratinophila* and *N. unguis-hominis* were the most common species, both with eight isolates each. *Neocucurbitaria cava*, with a single isolate (CNRMA 15.708), was also included in this clade. Three Spanish isolates, CNM-CM 6489, CNM-CM 7025 and CNM-CM 7132 were identified as *Neocucurbitaria* sp. due to being phylogenetically different from the other isolates and, again, might be a new species of the genus. *Neocucurbitaria* spp.

produces pycnidia, ornamented or not, with bristle-like setose structures, and hyaline, aseptate conidia.

Clade 6 had eleven isolates of *Medicopsis romeroi* (syn. *Pyrenochaeta romeroi)*, which produces pycnidia and hyaline, aseptate conidia.

Clade 7 is represented by a single isolate (CNRMA 11.1115), phylogenetically distinct from the known pleosporalean fungi, possibly representing a novel taxon.

Clades 8 and 9 belonged to the family *Didymosphaeriaceae*. Clade 8 included a single isolate (CNM-CM 6000) phylogenetically related to a reference strain of *Paraphaeosphaeria michotii* (MFLUCC 13-0349). Clade 9 grouped ten isolates, two related to a reference strain of *Paraconiothyrium fuckelii* (CBS 797.95) and eight with the type strain of *Paraconiothyrium cyclothyrioides* (CBS 972.95). Members of the *Didymosphaeriaceae* form pycnidia and pale brown, 0-1 septate conidia.

The order *Botryosphaeriales* are present in Clades 10 to 12. Clade 10 had only one isolate (CNRMA 12.597) which clustered with a reference strain of *Neofusicoccum luteum* (CBS 110299); Clade 11 also had a single isolate (CNRMA 6.1007) that clustered with the type strain of *Diplodia seriata* (CBS 112555), and Clade 12 grouped six isolates, five of them clustering with the type strain of *Lasiodiplodia theobromae*, and CNRMA 15.383 identified as *Lasiodiplodia* sp. These fungi produce stromatic conidiomata and aseptate, hyaline to brown, thick-walled conidia.

Clade 13 included the type strain of *Diaporthe sclerotioides* (CBS 296.67) and six isolates corresponding to unidentified species of the genus *Diaporthe* (*Diaporthales*), none of them able to be morphologically distinguished since they produce pycnidia and small hyaline conidia.

Clade 14, corresponding to the *Glomerellales*, was used as outgroup. Five isolates nested in the *Colletotrichum* clade, two clustering with reference strains of *C. gigasporum* (CBS 159.75) and *C. gloeosporioides* (CBS 122687), respectively; and the other three, could not be identified. All the isolates showed the typical morphology of *Colletotrichum*, i.e., acervuli, conidia variable in shape, flattened with thickened tip branches (appressoria).

Antifungal susceptibility testing

The minimum inhibitory concentration (MIC) was determined for 46 of the isolates included here (16 from Spain and 30 from France) (Table 3, Table S1). Globally, the geometric mean (GM) and MIC_{50} values of itraconazole and caspofungin were the highest (Table 3). The MIC of amphotericin B (0.06-1 mg/L) was generally low among the *Pleosporales* with the exception of one isolate of *M. romeroi* and one of *D. gardeniae*, with MICs of 8 and 32 mg/L, respectively. The azole MIC ranged between 0.03 and 1 mg/L for isolates belonging to the genera *Paraconiothyrium*, *Paraphoma*, *Tintelnotia* and *Neocucurbitaria*, with the exception of two isolates of *N. unguis-hominis*, which showed higher values (16 mg/L). The terbinafine MIC was low except for *Diaporthe* spp. and a few isolates of *Colletotrichum* spp. and *M. romeroi*.

DISCUSSION

The present study is the largest on this taxonomically complex group of fungi from clinical origin, with almost a hundred isolates morphologically and molecularly characterized from two southern European countries (France and Spain). Most of these coelomycetous fungi belonged to the order *Pleosporales* and were most commonly recovered from superficial infections. Similar results were observed in a previous work that focused on coelomycetous fungi collected at a North American reference centre (6). However, the diversity of the fungi identified in that study was higher, i.e. eleven orders were represented against four here.

In the present study, *Medicopsis romeroi* was the most frequently isolated species whereas the most common taxon in the American study was *Neoscytalidium dimidiatum*. Interestingly, while *M. romeroi* is usually reported as an etiologic agent of black grain eumycetoma (4, 11, 26–29), our isolates were mainly recovered from eye and non-mycetoma subcutaneous infections.

The second most frequently isolated species were *Paraconiothyrium cyclothyrioides*, *Neocucurbitaria unguis-hominis* and *N. keratinophila*. *Paraconiothyrium cyclothyrioides* is an emerging pathogen (6, 26, 30, 31) and was represented by eight isolates recovered from skin or superficial locations and mainly from tropical regions. *Neocucurbitaria unguis-hominis*, initially described as an agent of human onychomycosis (17), was equally distributed across both centres (n=8 isolates). Regarding *N. keratinophyla*, this species was reported for the first time from a corneal infection in Spain (18, 19). Interestingly, as well as being the first case reported for this species, all the isolates of *N. keratinophyla* were recovered in Spain from superficial tissue.

198 Other coelomycetous fungi we identified in the present work were *Didymella*
199 *glomerata* and *Phoma herbarum*. Although *Phoma* spp. are commonly reported as a
200 coelomycete involved in human infections (9, 20, 32–39), recent extensive changes in
201 taxonomy and nomenclature have spread all but one of the species into different genera
202 of the *Didymellaceae*, *Phoma herbarum* remaining as the unique species of the genus
203 (22–24). Interestingly, *Didymella gardeniae* was commonly found in our study (five
204 isolates from Spain and one from France).

205 Recently, Ahmed *et al*. (40) proposed *Tintelnotia destructans*, a new phoma-like
206 fungus belonging to the *Phaeosphaeriaceae* able to cause eye and nail infections. They
207 reported the successful use of terbinafine against a case of keratitis by this species. Two
208 of the Spanish isolates recovered from superficial specimens (one cutaneous exudate
209 and one nail sample) were molecularly related to the above-mentioned species but
210 phylogenetically different and might represent a new taxon.

211 *Lasiodiplodia theobromae* (order *Botryosphaeriales*) is the only species of this
212 genus involved in human opportunistic infections (41–46). Valenzuela-Lopez *et al*. (6)
213 found a higher species diversity in the North American study than we report here, since
214 five of the French isolates were identified as *L. theobromae*. The other three isolates of
215 the *Botryosphaeriales* we found were related, one to a different species of *Lasiodiplodia*
216 and the other two to other genera, specifically *Neofusicoccum* and *Diplodia*.

217 Four species of the genus *Diaporthe* (formerly *Phomopsis*; order *Diaporthales*),
218 i.e. *D. bougainvilleicola*, *D. longicolla*, *D. phaseolorum* and *D. phoenicicola*, are
219 considered opportunistic pathogens that cause mycoses that range from superficial to
220 deep infections (47–51). Six isolates from France were phylogenetically placed into the
221 latter genus. However, our results are only preliminary since only one phylogenetic

222 marker was analysed. Similar was observed in several polyphyletic genera of the
223 coelomycetes (52, 53).

224 We also report the finding of five clinical isolates of *Colletotrichum*. Two of the
225 isolates corresponded to *C. gigasporum* (formerly *C. crassipes*) and *C. gloeosporioides*,
226 taxa that have previously been reported as agents of keratitis, endophthalmitis and
227 phaeohyphomicotic cyst; the other three isolates could not be identified at species level.
228 This genus encompasses numerous plant pathogens that are found worldwide, although
229 mainly in tropical and subtropical regions (54). The taxonomy of *Colletotrichum* is
230 complicated and the genus is organized in species-complexes (55–59). Species such as
231 *C. coccodes*, *C. crassipes*, *C. dematium*, *C. gloeosporioides*, *C. graminicola* and *C.*
232 *truncatum* cause superficial and deep infections (endophthalmitis, keratitis,
233 subcutaneous cyst or more rarely arthritis) (60–65). Further studies, including different
234 phylogenetic markers, are needed to delimit the different species and clarify their
235 pathogenic role.

236 The antifungal susceptibility of coelomycetous fungi involved in human
237 infections is poorly known, mainly because they do not easily sporulate. In spite of the
238 limited number of isolates tested here, amphotericin B seemed the most active drug *in*
239 *vitro* together with terbinafine, in agreement with Valenzuela-Lopez *et al*. (6). Until
240 more *in vitro* data is available, the antifungal treatment of the infection by this sort of
241 fungus remains purely empirical. In a recent study, Guégan *et al*. (26) recommended
242 extensive surgical resection of affected tissues as a first-line treatment for solitary
243 subcutaneous lesions by coelomycetous fungi, followed by an antifungal therapy
244 (posaconazole or voriconazole) in the case of relapse or amphotericin B in refractory
245 cases.

Since our study is based on isolates from the two reference centres, we cannot comment on the incidence of infections due to coelomycetes nor compare their epidemiology between France and Spain. However, we still provide a good picture of the great diversity of coelomycetous fungi in the clinical context, and the basis for future studies on this interesting but neglected group of fungi.

MATERIAL AND METHODS

Fungal isolates

We studied 97 isolates of coelomycetous fungi recovered from clinical specimens, 51 of which were provided by the French National Reference Centre for Invasive Mycoses and Antifungals (NRCMA) at the *Institut Pasteur*, Paris (CNRMA isolates, n=51). The NRCMA offers expertise on difficult-to-identify fungi and the epidemiological surveillance of all cases of IFIs, which are notified on a voluntary basis either through active or passive surveillance programmes. The Spanish National Centre of Microbiology at the *Instituto de Salud Carlos III*, Madrid provided 46 isolates (CNM-CM isolates, n=46). This mycology reference laboratory receives isolates from the National Health System on a voluntary basis, the main aim of which is to support it by identifying and profiling the antifungal susceptibility of fungal isolates. The isolates were collected between 2005 and 2015. Table 1 gives information about the country of isolation and the location of the infection in the body.

Morphological and physiological characterization

For morphology studies, the isolates were cultured on oatmeal agar (OA; 30 g of filtered oat flakes, 15 g of agar-agar, 1 L tap water) and malt extract agar (MEA; 40 g of malt extract, 15 g of agar-agar, 1 L distilled water) at 20 ± 1ºC for 14 days in darkness.

The morphological features of the vegetative and reproductive structures were studied using an Olympus CH2 bright-field microscope (Olympus Corporation, Tokyo, Japan) in wet mounts (on water and lactic acid) and slide cultures (by growing the isolates on OA and MEA) of the fungal isolates, following Valenzuela-Lopez *et al*. (6). Colour standards by Kornerup & Wanscher (66) were used in colony description. Photomicrographs were taken with an Axio-Imager M1 microscope (Zeiss, Oberkochen, Germany).

DNA extraction, amplification and sequencing

Total genomic DNA was extracted from colonies grown on potato dextrose agar (PDA; 4 g of potato infusion, 20 g dextrose, 15 g of agar-agar, 1 L tap water) after seven days of incubation at 20 ± 1ºC, using the FastDNA kit protocol (Bio101, Vista, CA), with a FastPrep FP120 instrument (Thermo Savant, Holbrook, NY) following the manufacturer's protocol. DNA was quantified using the Nanodrop 2000 (Thermo Scientific, Madrid, Spain). LSU was amplified with the primer pair LR0R and LR5 (67). The amplicons were sequenced in both directions with the same primer pair used for amplification at Macrogen Europe (Macrogen Inc., Amsterdam, The Netherlands). The consensus sequences were obtained using the SeqMan software version 7.0.0 (DNAStar Lasergene, Madison, WI, USA).

Molecular identification and phylogenetic analysis

Preliminary molecular identification of the isolates was made using LSU nucleotide sequences in $BLAST_N$ searches. Twenty-eight LSU sequences of type or reference strains deposited in the GenBank database by the Westerdijk Fungal Biodiversity Institute (CBS) and the Mae Fah Luang University (MFLUCC) culture collections were used for identification and phylogenetic purposes. DNA sequences

294 generated in this study were deposited in GenBank (accession numbers are given in
295 Table 1).

296 For the phylogenetic study, sequences were aligned using the ClustalW
297 application (68) of the MEGA 6.06 (69) computer program, and manually adjusted
298 using the same software platform. Phylogenetic reconstructions were made by
299 maximum-likelihood (ML) and Bayesian inference (BI) with MEGA 6.06 and MrBayes
300 3.2.4 (70), respectively. The best substitution model for the gene matrix (TN93+G) was
301 estimated using MEGA 6.06. For ML analyses, nearest-neighbour interchange was used
302 as the heuristic method for tree inference. Support for internal branches was assessed by
303 1,000 ML bootstrapped pseudoreplicates. Bootstrap support (BS) of ≥70 was considered
304 significant. For BI analyses, Markov chain Monte Carlo (MCMC) sampling was carried
305 out with four million generations, with samples taken every 1,000 generations. The 50%
306 majority rule consensus trees and posterior probability values (PP) were calculated after
307 removing the first 25% of the resulting trees for burn-in. A PP value of ≥0.95 was
308 considered significant. Reference strains of *Colletotrichum gigasporum* (CBS 159.75),
309 *C. gloeosporioides* (CBS 122687) and *C. hippeastri* (CBS 241.78) were used as
310 outgroup.

311 **Antifungal susceptibility testing**

312 The *in vitro* susceptibility testing in both reference centres (n= 46 isolates)
313 followed the European Committee on Antimicrobial Susceptibility Testing (EUCAST)
314 procedure (71, 72). The antifungals used were amphotericin B (Sigma-Aldrich Química,
315 Madrid, Spain), itraconazole (Sigma-Aldrich Química, Madrid, Spain), posaconazole
316 (Schering-Plough Research Institute, Kenilworth, N.J.), voriconazole (Pfizer S.A.,
317 Madrid, Spain), caspofungin (Merck & Co., Inc., Rahway, N.J.), micafungin (Astellas

318 Pharma Inc, Tokyo, Japan) and terbinafine (Novartis, Basel, Switzerland). For the
319 NCRMA, all antifungal drugs were obtained from ALSACHIM, Strasbourg, France.

320 The isolates were cultured on potato carrot agar (PCA; 20 g each of filtered
321 potatoes and carrots, 20 g of agar, 1 L of distilled water) or OA for seven to 30 days at
322 25°C and 30°C to obtain sporulation. Conidia were then collected in sterile water
323 containing 0.01% (v/v) Tween 80 (Sigma-Aldrich, St. Louis, MO, USA), and the
324 suspension was adjusted to 2–5 x 10^5 conidia/mL. The minimal effective concentration
325 (MEC) was determined for each echinocandin and the minimal inhibitory concentration
326 (MIC) for the other drugs (90% inhibition for amphotericin B and 80% for the azoles)
327 after 24 h and 48 h of incubation at 35°C. *Aspergillus flavus* ATCC 204304 and
328 *Aspergillus fumigatus* ATCC 204305 were used as quality control strains in all tests
329 carried out. Susceptibility profiles were determined for 46 isolates since non-sporulating
330 isolates were excluded at the NRCMA.

331

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560

561 **FIGURE LEGEND**

562 **FIG 1** Maximum likelihood tree obtained from the D1-D2 of LSU (584 bp) sequences
563 of the 125 strains, where 28 belong to type or reference strains. The branch lengths are
564 proportional to phylogenetic distance. Bayesian posterior probability scores ≥ 0.95 and
565 Bootstrap support values ≥ 70% are indicated on the nodes. Some branches were
566 shortened to fit them to the page, these are indicated by two diagonal lines with the
567 number of times a branch was shortened. The species of the genus *Colletotrichum* were
568 used to root the tree. Superscript ^{T} indicated the type strains.

569

570

TABLE 1 Taxonomical identification of the isolates studied, origin and GenBank accession numbers. New sequences generated are indicate in bold.

Order	Species	Strain no. [a]	Origin	Country	GenBank accesion no. [b]
Botryosphaeriales	*Diplodia seriata*	CBS 112555^{T}	*Vitis vinifera* dead plant	Portugal	KF766327
		CNRMA 6.1007	bone	France	**LT965964**
	Lasiodiplodia sp.	CNRMA 15.383	eye	France (West Indies, Guadeloupe)	**LT965965**
	Lasiodiplodia theobromae	CBS 164.96^{T}	fruit along coral reef coast	Papua New Guinea	NG_042460
		CNRMA 10.1369	skin	France (West Indies, Martinique)	**LT965966**
		CNRMA 10.813	eye	France (West Indies, Martinique)	**LT965967**
		CNRMA 11.360	eye	France (West Indies, Martinique)	**LT965968**
		CNRMA 13.891	skin	France	**LT965969**
		CNRMA 14.708	eye	France (West Indies, Guadeloupe)	**LT965970**
	Neofusicoccum luteum	CBS 110299	*Vitis vinifera* cane	Portugal	AY928043
		CNRMA 12.597	eye	France	**LT965971**
Diaporthales	*Diaporthe sclerotioides*	CBS 296.67^{T}	*Cucumis sativus* root	The Netherlands	AF439628
	Diaporthe sp.	CBS 477	*Cucumis sativus*	USA	AF439631
		CNRMA 8.522	eye	France	**LT965972**
		CNRMA 9.205	eye	France (West Indies, Guadeloupe)	**LT965973**
		CNRMA 11.385	eye	France (West Indies, Martinique)	**LT965974**
		CNRMA 12.311	blood	France	**LT965975**
		CNRMA 13.515	skin	France	**LT965976**
		CNRMA 14.198	skin	France	**LT965977**

Glomerellales	*Colletotrichum gigasporum*	CBS 159.75	air and stored grains	India	DQ286206
		CNRMA 16.553	skin	France (West Indies, Guadeloupe)	**LT965978**
	Colletotrichum gloeosporioides	CBS 122687	*Leucospermum* sp. leaf litter	South Africa	EU552111
		CNRMA 15.504	eye	France (West Indies, Martinique)	**LT965979**
	Colletotrichum hippeastri	CBS 241.78	*Hippeastrum* sp.	The Netherlands	DQ286167
	Colletotrichum sp.	CNM-CM4760	corneal swab	Spain	**LT965980**
		CNM-CM 6116	conjuntival	Spain	**LT965981**
		CNM-CM 7345	humor acuosus	Spain	**LT965982**
Pleosporales	*Didymella gardeniae*	CBS 626.68^{T}	*Gardenia jasminoides* leaf	India	GQ387595
		CNM-CM 3697	nail	Spain	**LT965983**
		CNM-CM 3895	nail	Spain	**LT965984**
		CNM-CM 5036	scales	Spain	**LT965985**
		CNM-CM 5814	conjunctival exudate	Spain	**LT965986**
		CNM-CM 7499	conjunctival exudate	Spain	**LT965987**
		CNRMA 11.794	skin	France	**LT965988**
	Didymella glomerata	CBS 528.66	*Chrysanthemum* sp. cutting	The Netherlands	EU754184
		CNM-CM 3356	toenail	Spain	**LT965989**
		CNM-CM 3546	nail	Spain	**LT965990**
		CNM-CM 4675	nail	Spain	**LT965991**
		CNM-CM 7099	cutaneous exudate	Spain	**LT965992**
		CNRMA 9.1046	skin	France	**LT965993**

	CNRMA 10.867	skin	France	**LT965994**
	CNRMA 15.6	mouth/sinus	France	**LT965995**
Epicoccum nigrum	CBS 173.73^{T}	*Dactylis glomerata* seed	USA	GU237975
	CNM-CM 5281	skin	Spain	**LT965996**
	CNM-CM 5724	vitreous humor	Spain	**LT965997**
Epicoccum sorghinum	CBS 179.80	*Sorghum vulgare*	Puerto Rico	GU237978
	CNRMA 7.167	bone	France (New Caledonia)	**LT965998**
	CNRMA 10.947	skin	France (New Caledonia)	**LT965999**
	CNRMA 10.948	skin	France (New Caledonia)	**LT966000**
Medicopsis romeroi	CBS 252.60^{T}	maduromycosis	Venezuela	EU754207
	CNM-CM 3387	knee ulcer	Spain	**LT966001**
	CNM-CM 7645	cutaneous exudate	Spain	**LT966002**
	CNRMA 4.200	eye	France	**LT966003**
	CNRMA 5.321	skin	France	**LT966005**
	CNRMA 7.1225	skin	France	**LT966007**
	CNRMA 8.1363	skin	France	**LT966008**
	CNRMA 11.680	skin	France	**LT966010**
	CNRMA 11.949	bone	France	**LT966011**
	CNRMA 14.407	skin	France	**LT966013**
	CNRMA 15.461	bone	France	**LT966014**
	CNRMA 15.7	skin	France	**LT966015**
Neoascochyta desmazieri	CBS 297.69^{T}	*Lolium perenne*	Germany	KT389726
	CNM-CM 6201	nail	Spain	**LT966016**

Neocucurbitaria cava	CBS 257.68^{T}	wheat-field soil	Germany	EU754199
	CNRMA 15.708	mouth/sinus	France	**LT966017**
Neocucurbitaria keratinophila	CBS 121759^{T}	corneal scrapings	Spain	LT623215
	CNM-CM 5882	cutaneous exudate	Spain	**LT966018**
	CNM-CM 6401	fingernail	Spain	**LT966019**
	CNM-CM 6455	cutaneous exudate	Spain	**LT966020**
	CNM-CM 7013	cutaneous exudate	Spain	**LT966021**
	CNM-CM 7457	cutaneous exudate	Spain	**LT966022**
	CNM-CM 7731	cutaneous exudate	Spain	**LT966023**
	CNM-CM 8010	conjunctival exudate	Spain	**LT966024**
	CNM-CM 8674	toenail	Spain	**LT966025**
Neocucurbitaria unguis-hominis	CBS 112.79	airborn	Wales	GQ387622
	CNM-CM 7037	nail	Spain	**LT966026**
	CNM-CM 7089	cutaneous lession	Spain	**LT966027**
	CNM-CM 8717	urine	Spain	**LT966028**
	CNM-CM 8743	toenail	Spain	**LT966029**
	CNRMA 4.1112	eye	France	**LT966030**
	CNRMA 6.243	eye	France	**LT966031**
	CNRMA 16.153	eye	France	**LT966032**
	CNRMA 16.19	lung	France	**LT966033**
Neocucurbitaria sp.	CNM-CM 6489	wound exudate	Spain	**LT966034**
	CNM-CM 7025	hair	Spain	**LT966035**
	CNM-CM 7132	toenail	Spain	**LT966036**

Paraconiothyrium cyclothyrioides	CBS 972.95^{T}	soil	Papua New Guinea	JX496232
	CNM-CM 6313	conjunctival exudate	Spain	**LT966037**
	CNM-CM 6513	nail	Spain	**LT966038**
	CNM-CM 4767	abscess	Spain	**LT966039**
	CNRMA 11.383	skin	France (West Indies, Martinique)	**LT966041**
	CNRMA 11.855	skin	France	**LT966042**
	CNRMA 13.245	skin	France	**LT966043**
	CNRMA 16.374	skin	France (West Indies, Guadeloupe)	**LT966044**
	CNRMA 16.556	skin	France (West Indies, Guadeloupe)	**LT966045**
Paraconiothyrium fuckelii	CBS 797.95	*Rubus* sp. dead stem	Denmark	JX496226
	CNRMA 3.240	eye	France	**LT966046**
	CNRMA 4.493	eye	France	**LT966047**
Paraphaeosphaeria michotii	MFLUCC 13-0349	*Poaceae* dead leaves	Italy	KJ939282
	CNM-CM 6000	skin	Spain	**LT966048**
Paraphoma fimeti	CBS 170.70^{T}	*Apium graveolens* seeds	The Netherlands	GQ387584
	CNM-CM 8075	wound exudate	Spain	**LT966049**
Paraphoma sp.	CNRMA 9.467	skin	France	**LT966050**
	CNRMA 15.665	skin	France	**LT966051**
Phaeosphaeriopsis obtusispora	CBS 246.64	*Aloe arborescens* dead leaf	Portugal	JX681119
Phoma herbarum	CBS 615.75	*Rosa multiflora* dead stem	The Netherlands	EU754186

	CNM-CM 2132	right toe	Spain	**LT966052**
	CNM-CM 3526	bone marrow	Spain	**LT966053**
	CNM-CM 3597	blood culture	Spain	**LT966054**
	CNM-CM 8031	nail	Spain	**LT966055**
	CNRMA 9.1095	skin	France	**LT966056**
	CNRMA 11.1097	eye	France	**LT966057**
	CNRMA 12.1227	eye	France	**LT966058**
pleosporelean fungus	CNRMA 11.1115	skin	France	**LT966059**
Preussia sp.	CNM-CM 7343	nail	Spain	**LT966060**
Preussia terricola	CBS 317.65^{T}	*Musa sapientum* rhizosphere	Honduras	GQ203725
Preussia typharum	CBS 107.69	Dung of deer	Japan	GQ203726
	CNM-CM 7335	nail	Spain	**LT966061**
Pseudophaeosphaeria rubi	MFLUCC 14-0259	*Rubus idaeus* dead branch	Italy	KX765299
Tintelnotia destructans	CBS 127737^{T}	anterior eye chamber cornea	Germany	KY090664
	CNM-CM 7430	Unknown	Spain	**LT966062**
Tintelnotia sp.	CNM-CM 7080	nail	Spain	**LT966063**
	CNM-CM 7981	cutaneous exudate	Spain	**LT966064**
Xenodidymella saxea	CBS 419.92^{T}	Corroded mediterranean marble	Unknown	GU238141

	CNRMA 16.76	Cerebrospinal fluid	France	**LT966065**

[a] **CBS**: Strains from Westerdijk Fungal Biodiversity Institute, Utrecht, The Netherlands; **CNM-CM**: Isolates from the National Centre for Microbiology, Instituto Carlos III, Madrid, Spain; **CNRMA**: Isolates from the National Reference Center for Invasive Mycoses and Antifungals; Institut Pasteur, Paris, France; **MFLUCC**: Strains from Mae Fah Luang University Culture Collection, Chiang Rai, Thailand. Type strains are indicated by a superscript [T]

[b] LSU, large subunit ribosomal DNA sequences

TABLE 2 Localization of infections due to coelomycetous fungi isolates

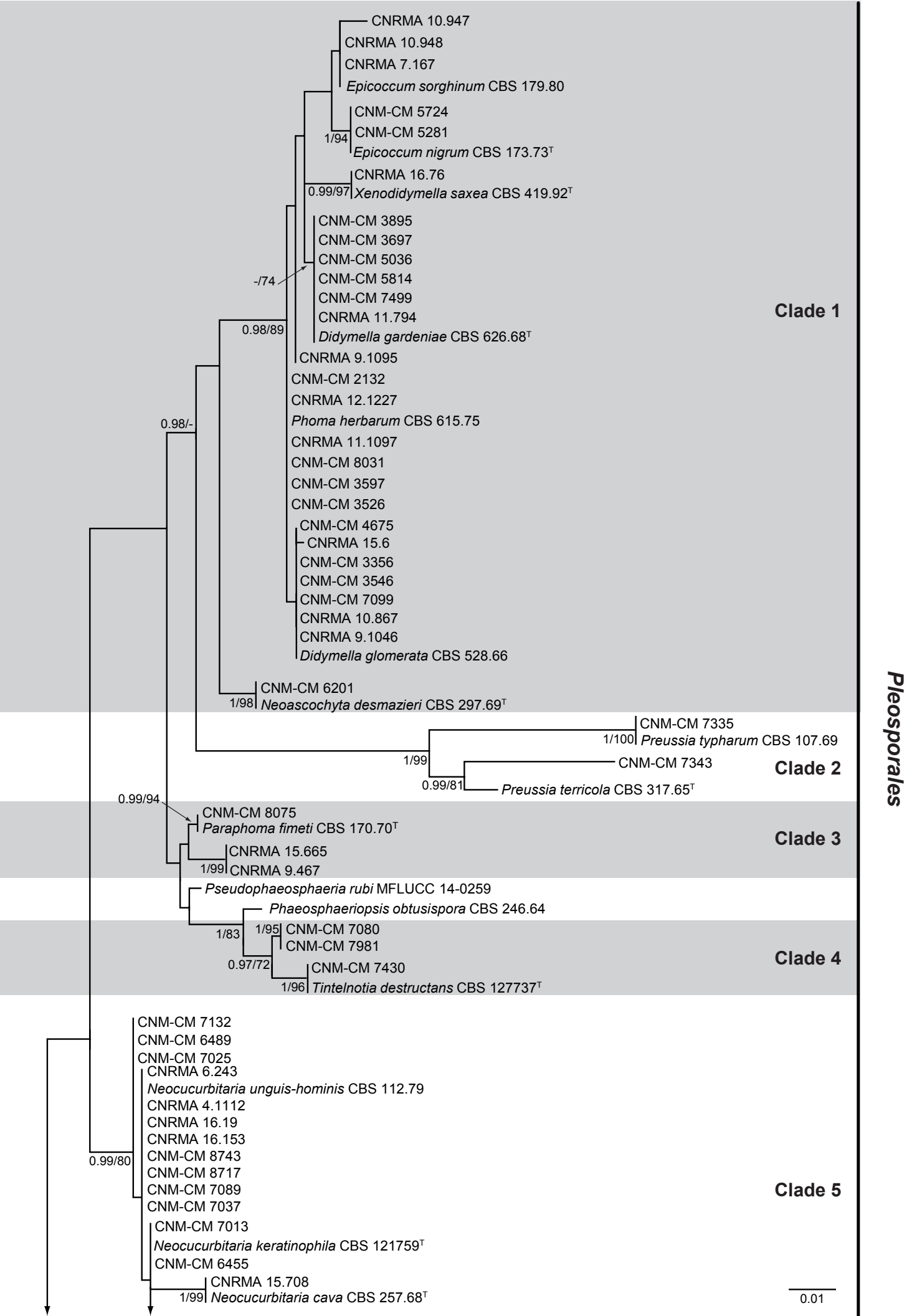
	no. of isolates obtained from:		
Orders	**Superficial infection**	**Deep infection**	**Total no. of isolates**
Botryosphaeriales	7	1	8
Diaporthales	5	1	6
Glomerellales	5		5
Pleosporales	71	7	78
Total no. of isolates (%)	**88 (91)**	**9 (9)**	**97 (100)**

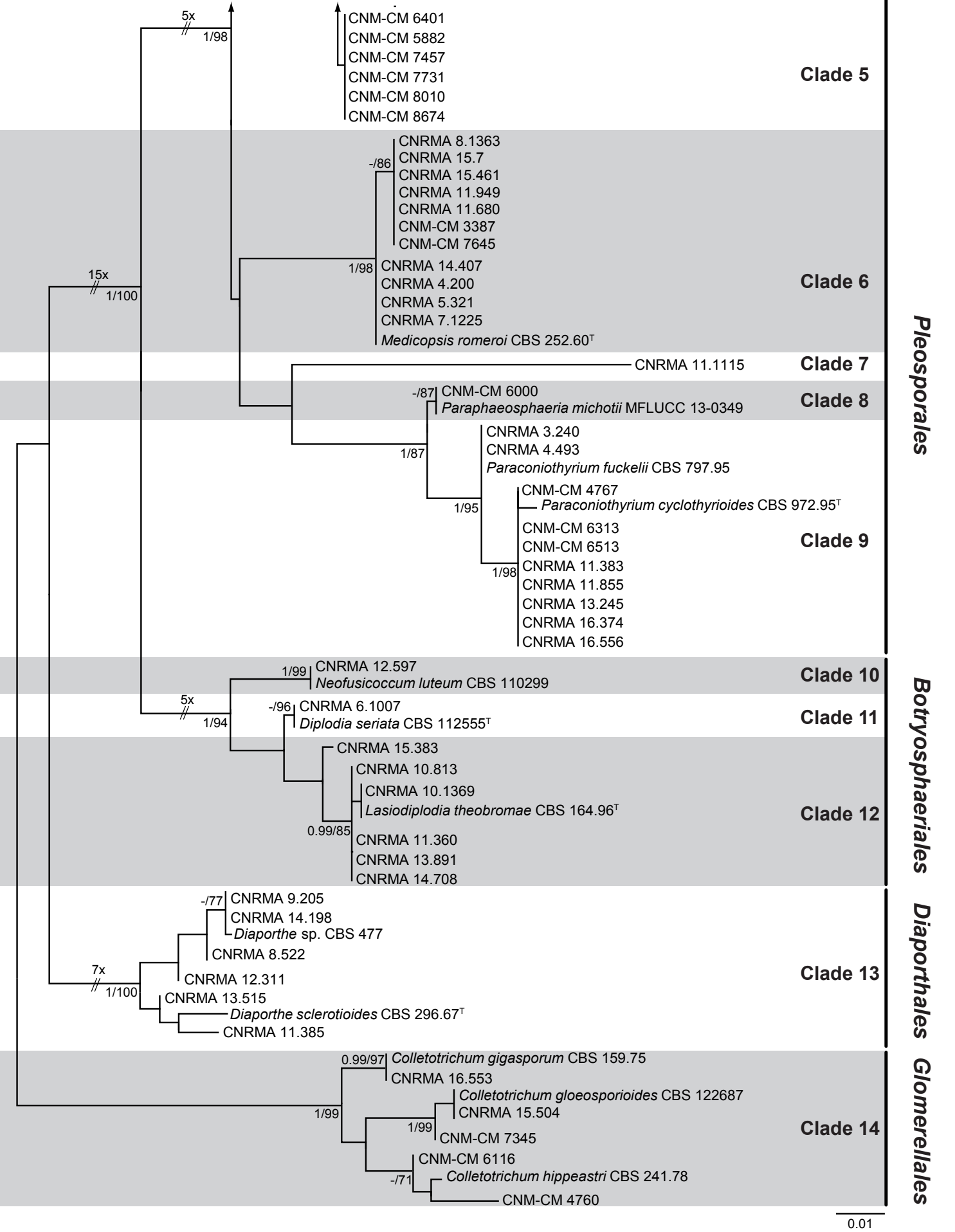
TABLE 3 Overall i*n vitro* antifungal activity against the 46 coelomycetous isolates as determined by EUCAST[a] methodology

Antifungal agent	MIC/MEC values (mg/L)[b]				
	range	median	GM	MIC_{50}	MIC_{90}
Amphotericin B	0.03 -16	0.5	0.41	0.25	1
Itraconazole	0.014 -16	2	1.72	0.5	16
Voriconazole	0.03 -16	0.5	0.70	0.6	4
Posaconazole	0.014 -16	0.5	0.58	0.25	8
Caspofungin	0.125- 16	2	2.17	1	8
Micafungin	0.015- 16	0.5	0.53	0.125	8
Terbinafin	0.014- 16	0.25	0.39	0.25	2

[a]EUCAST, European Committee on Antimicrobial Susceptibility Testing procedure (71);

[b]MIC, minimum inhibitory concentration; MEC, minimal effective concentration; MIC_{50} and MIC_{90}, MIC encompassing 50 and 90% of isolates tested, respectively.


CNRMA 10.947
CNRMA 10.948
CNRMA 7.167
Epicoccum sorghinum CBS 179.80
CNM-CM 5724
CNM-CM 5281
Epicoccum nigrum CBS 173.73T
CNRMA 16.76
Xenodidymella saxea CBS 419.92T
CNM-CM 3895
CNM-CM 3697
CNM-CM 5036
CNM-CM 5814
CNM-CM 7499
CNRMA 11.794
Didymella gardeniae CBS 626.68T
CNRMA 9.1095
CNM-CM 2132
CNRMA 12.1227
Phoma herbarum CBS 615.75
CNRMA 11.1097
CNM-CM 8031
CNM-CM 3597
CNM-CM 3526
CNM-CM 4675
CNRMA 15.6
CNM-CM 3356
CNM-CM 3546
CNM-CM 7099
CNRMA 10.867
CNRMA 9.1046
Didymella glomerata CBS 528.66
CNM-CM 6201
Neoascochyta desmazieri CBS 297.69T
CNM-CM 7335
Preussia typharum CBS 107.69
CNM-CM 7343
Preussia terricola CBS 317.65T
CNM-CM 8075
Paraphoma fimeti CBS 170.70T
CNRMA 15.665
CNRMA 9.467
Pseudophaeosphaeria rubi MFLUCC 14-0259
Phaeosphaeriopsis obtusispora CBS 246.64
CNM-CM 7080
CNM-CM 7981
CNM-CM 7430
Tintelnotia destructans CBS 127737T
CNM-CM 7132
CNM-CM 6489
CNM-CM 7025
CNRMA 6.243
Neocucurbitaria unguis-hominis CBS 112.79
CNRMA 4.1112
CNRMA 16.19
CNRMA 16.153
CNM-CM 8743
CNM-CM 8717
CNM-CM 7089
CNM-CM 7037
CNM-CM 7013
Neocucurbitaria keratinophila CBS 121759T
CNM-CM 6455
CNRMA 15.708
Neocucurbitaria cava CBS 257.68T
1/94
0.99/97
-/74
0.98/89
0.98/-
1/98
1/100
1/99
0.99/81
0.99/94
1/99
1/83
1/95
0.97/72
1/96
0.99/80
1/99
Clade 1
Clade 2
Clade 3
Clade 4
Clade 5
Pleosporales
0.01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1/98
CNM-CM 6401
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CNM-CM 8010
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CNRMA 8.1363
-/86
CNRMA 15.7
CNRMA 15.461
CNRMA 11.949
CNRMA 11.680
CNM-CM 3387
CNM-CM 7645
1/98
CNRMA 14.407
CNRMA 4.200
CNRMA 5.321
CNRMA 7.1225
Medicopsis romeroi CBS 252.60T
Clade 6
15x
1/100
CNRMA 11.1115
Clade 7
-/87
CNM-CM 6000
Paraphaeosphaeria michotii MFLUCC 13-0349
Clade 8
CNRMA 3.240
CNRMA 4.493
1/87
Paraconiothyrium fuckelii CBS 797.95
CNM-CM 4767
1/95
Paraconiothyrium cyclothyrioides CBS 972.95T
CNM-CM 6313
CNM-CM 6513
CNRMA 11.383
1/98
CNRMA 11.855
CNRMA 13.245
CNRMA 16.374
CNRMA 16.556
Clade 9
Pleosporales
1/99
CNRMA 12.597
Neofusicoccum luteum CBS 110299
Clade 10
5x
-/96
CNRMA 6.1007
1/94
Diplodia seriata CBS 112555T
Clade 11
CNRMA 15.383
CNRMA 10.813
CNRMA 10.1369
Lasiodiplodia theobromae CBS 164.96T
0.99/85
CNRMA 11.360
CNRMA 13.891
CNRMA 14.708
Clade 12
Botryosphaeriales
-/77
CNRMA 9.205
CNRMA 14.198
Diaporthe sp. CBS 477
CNRMA 8.522
7x
CNRMA 12.311
1/100
CNRMA 13.515
Diaporthe sclerotioides CBS 296.67T
CNRMA 11.385
Clade 13
Diaporthales
0.99/97
Colletotrichum gigasporum CBS 159.75
CNRMA 16.553
Colletotrichum gloeosporioides CBS 122687
1/99
CNRMA 15.504
1/99
CNM-CM 7345
CNM-CM 6116
-/71
Colletotrichum hippeastri CBS 241.78
CNM-CM 4760
Clade 14
Glomerellales
0.01