Using large soybean historical data to study genotype by environment variation and identify mega-environments with the integration of genetic and non-genetic factors

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1 Highlights

- A target population of environments can be split into mega-environments (MEs) according to phenotypic, geographic, and meteorological information.
- Reliable estimates of variance components are key to the identification of ME, which can be obtained by analyses of historical experimental data.
- From experimental soybean seed yields evaluated across 31 years of field trials, the phenotypic variance was mostly attributed to location and location by year effects. In terms of genotype-by-environment interactions (GEI), estimated variances of genotype by location interactions was more important than the genotype by year interactions.
- The GEI trend was successfully captured in terms of parametric probability distributions of variance components, that can be incorporated in simulation studies.

2 Abstract

Soybean (*Glycine max* (L.) Merr.) provides plant based protein for global food production and is extensively bred to create cultivars with greater productivity in distinct environments. Plant breeders evaluate new soybean genotypes using multi-environment trials (METs). Application of METs assume that trial sites provide representative environmental conditions that cultivars are likely to encounter when sold to farmers. Thus, it is important to understand the patterns of genotype by environment interactions (GEI) that occur in METs. In order to evaluate GEI for soybean seed yield and identify mega-environments, historical data were investigated with a retrospective analysis of 39,006 unique experimental soybean genotypes evaluated in preliminary and uniform trials conducted by public plant breeders from 1989-2019. Mega-environments (MEs) were identified using yield records of lines from the annual trials and geographic, soil, and meteorological records at the trial locations. Results indicate that yield variation was mostly explained by location and location by year interactions. The static portion of the GEI represented 26.30% of the total yield variance. Estimates of variance due to genotype by location were greater than estimates of variance due to genotype by year interaction effects. A trend analysis further indicated a two-fold increase in the genotypic variance. Furthermore, the heterogeneous estimates of genotypic, genotype by location, genotype by year, and genotype by location by year variances, were encapsulated by distinct probability distributions. The observed target population of environments (TPE) can be divided into at least two and at most three MEs, thereby suggesting improvements in the response to selection can be achieved when selecting directly for clustered (i.e. regions, ME) versus selecting across regions. Clusters obtained using phenotypic data, latitude, and soil variables plus elevation, were the most effective.

3 Keywords

Soybean; Genotype by Environment Interaction; Multi-environmental trials; Target Population of Environments; Mega-environments

4 Introduction

The terms genotype (G) and phenotype (P) were first coined by Johannsen (1911) after the rediscovery of Mendel's work. Since then, the understanding of the mapping function that links G to P has been an on-going research interest (Pigliucci 2001, p. 2). The mapping of G to P for most quantitatively expressed traits is further complicated by the differential response of genotype(s) to different environments, i.e. genotype by environment interactions (GEI), wherein phenotypic variation is shaped by G, Environment (E), and GEI (Tabery 2008; Sprague and Federer 1951). The GEI typically increases P variance and leads to a reduced estimates of heritability, complicating breeding decisions and lowering response to selection. Additionally, it leads to unpredictable adaptation of genotypic lines in targeted agro-ecological zones (Mackay *et al.* 2019) and influences plasticity response of varieties in variable environments (Cooper and DeLacy 1994; Haldane 1947). Hence, the GEI is of particular importance to breeders as they attempt to develop stable and responsive varieties (Comstock, R. E. and Moll 1962).

In order to reveal GEI patterns, plant breeders evaluate candidate genotypes in multi-environment trials (METs) (Oakey *et al.* 2016; Smith *et al.* 2001b). Sampled locations used in METs are assumed to represent the growing conditions that a candidate line is expected to encounter as a cultivar grown by farmers (Bustos-Korts *et al.* 2021). METs utilize locations that are sampled from a target population of environments (TPEs) which represent farm production environments. Hence, a TPE is composed of many environments (spatially across agro-ecological zones, and temporally over years) (Crespo-Herrera *et al.* 2021). The manifestation of GEI in a TPE has two components, the "static" environmental characteristics such as soil, longitude, latitude, and "non-static" seasonal characteristics such as weather and management practices (Cullis *et al.* 2000). If GEI is large and associated with consistent sub-groupings of environments within the TPE, greater gains from selection might be achieved by subdividing locations into Mega-Environments (MEs) (Crespo-Herrera *et al.* 2021; Yan 2016; Atlin *et al.* 2000a).

According to CIMMYT (1989) p. 58, "MEs are broad, not necessarily contiguous areas, defined by similar biotic and abiotic stresses, cropping system requirements …". Another definition is a group of environments that share the same winning genotypes (Kang 2020; Gauch and Zobel 1997), or that within ME there is minimal crossover interaction (COI) among the genotypes grown among environments (Smith *et al.* 2021). In a group of locations, if genotypes consistently perform the same relative to each other over a number of seasons, it is considered a ME (Singh *et al.* 2021, Chapter 4). One way of exploring GEI is to divide the TPE into MEs, and to select within ME (Yan 2016). Some studies have investigated strategies to subdivide the TPE in maize (Windhausen *et al.* 2012), barley (Atlin *et al.* 2000b), wheat (George and Lundy 2019; Bustos-Korts 2017), sorghum (da Silva *et al.* 2021), alfafa (Annicchiarico 2021), rice (Krishnamurthy *et al.* 2017), oat (Yan *et al.* 2010), and soybean (Zdziarski *et al.* 2019; Yan and Rajcan 2002).

There are several methods for dividing the TPE into MEs. For example, the genotype main effect plus GEI (GGE) Biplots (Yan *et al.* 2000) on soybean MET data was used by Zdziarski *et al.* (2019) to identify two MEs in Midwestern Brazil with contrasting altitudes, levels of fertilizer, and incidence of soybean cyst nematode profiles. da Silva *et al.* (2021) and Krishnamurthy *et al.* (2017) also took advantage of GGE Biplots to pinpoint MEs for pre-commercial sorghum hybrids in Brazil and rice genotypes in India, respectively. For wheat, Crespo-Herrera *et al.* (2021) defined three MEs in India with climate and soil data through principal component analysis, followed by a hierarchical clustering based on Euclidean distance with Ward's method. For

maize in Africa (CIMMYT's program), Windhausen *et al.* (2012) explored historical (2001-2009) METs data to determine MEs according to five subdivision systems (climate, altitude, geographic, country, and yield-level), and concluded there was enough genotype by subregion interaction relative to genotypic variance to justify the selection for the low and high-yielding sub-regions separately. Other methodologies such as the additive main effects and multiplicative interaction (AMMI) model (Bustos-Korts 2017; Gauch and Zobel 1997) and factor analytic (FA) models (Smith *et al.* 2021; Bustos-Korts 2017; Smith *et al.* 2015, 2001b; Piepho 1997) also have been used.

It should be noted that the terms subregion, region/regional, subdivision, clusters, zones, agro-climatic, ecogeographic and MEs are sometimes interchangeably used in the literature. For the METs data analysis, when MEs are ignored, the baseline model includes genotypes, locations, years (or the combination location-year, called environment), all two-way and three-way interactions (Malosetti *et al.* 2013). When MEs are included in the model, it is called a zone-based model; therefore, yielding zone-based predictions (Buntaran *et al.* 2019). One of the main advantages of modeling MEs in a mixed model framework is the ability to borrow information between zones from the genotype by ME interaction. This is particularly beneficial when fewer testing locations are available creating a sparse representation of genotypes in some locations (Piepho *et al.* 2016; Piepho and Möhring 2005).

The effectiveness of subdividing the TPE into MEs was assessed by Atlin *et al.* (2000a) based on the theory of correlated response to selection, first applied to the GEI problem by Falconer (1952). Effective selection occurs when subdivision increases response to selection, which might occur if the genotype by ME interaction variance, i.e. genotype by region (σ^2_{GR}), is large relative to the genotypic variance (σ^2_G). In terms of variance components, the GEI is composed of genotype by location (σ^2_{GL}), genotype by year (σ^2_{GY}), and genotype by location by year (σ^2_{GLY}) interaction variances. Both σ^2_{GY} and σ^2_{GLY} are non-static (unrepeatable) sources of variation. MEs can be identified with the static portion of the σ^2_{GL}, which is repeatable across years (Yan 2016). When MEs are identified and modelled, the σ^2_{GL} is partitioned into σ^2_{GR} and genotype by location within ME ($\sigma^2_{GL(R)}$). Furthermore, the σ^2_{GLY} is partitioned into a genotype by ME by year interaction (σ^2_{GRY}), and genotype by location within ME by year ($\sigma^2_{GL(R)Y}$) interaction (Atlin *et al.* 2000a). Consequently, the estimation of variance components provide important information for decision-making and accurate estimates are critical.

Variance components can be estimated with unbalanced historical data to provide information for designing novel breeding strategies and optimize resource allocation (Aguate *et al.* 2019). Efforts have been made to quantify component variability using historical METs data in wheat, maize, sunflower, sugar beet, potato, rye (Meyer *et al.* 2011; Laidig *et al.* 2008), among other commercial crops. However, proper modeling of historical data can be a significant challenge (Dias *et al.* 2020), and if not done properly can lead to erroneous interpretations. In terms of variance estimates, recent work from Aguate *et al.* (2019) and Hartung and Piepho (2021) considered both the imbalance of data (due to selection) and the properties of the residual maximum likelihood (REML) method (Patterson and Thompson 1971) to shed light onto the bias of the estimates obtained from METs using linear mixed models. Their results served as guideline to design the variance estimation portion of this work, which will be discussed later.

With the motivations of identifying and describing MEs for soybean in the primary production area of North America, we obtained historical soybean performance (seed yield) data from

Uniform Soybean Cooperative Tests (USDA 2021). We purposely chose this dataset because these trials have been used for decisions on variety release by public breeding organizations. Further, because flowering in soybean is extremely sensitive to daylength, soybean breeders first classify experimental genotypes into maturity groups (MGs) and subsequently restrict yield evaluations to appropriate maturity zones (MZ) that are defined by lattitude. Thus, the lattitude (MZ) of a location used for soybean field trials is an implemented element of MEs for soybean field trials. The dataset consisted of 39,006 unique experimental soybean line yield data from 63 locations between 1989 and 2019. Note that experimental lines were not evaluated at all locations within years and most were not evaluated in more than one year. The objectives of this study were to: (i) investigate if the observed TPE spanning 31 years of trial evaluations can be classified into MEs, and (ii) estimate probability density functions for the underlying trend of genotypic, genotype by location, genotype by year, genotype by location by year, and residual variance components. This modelling approach allowed us to fit parametric probability distributions to variance components in order to capture the GEI trend that can be used in future simulation studies, which will be needed for predicting plant breeding outcomes in changing climates. Currently, simulation studies rely on point estimates of variance components (Kleinknecht *et al.* 2016), or set heritability values (such as low or high) (Rutkoski 2019). By capturing the GEI trend using historical data, we generate reliable variance estimates that can be used to conduct more realistic simulation studies.

5 Data and Methods

5.1 Phenotypic data

Annual PDF reports from the Northern Region of the USDA Uniform Soybean Tests were obtained from `https://ars.usda.gov/mwa/lafayette/cppcru/ust`. The data retrieved from the published PDF files represent averages for seed yield for each genotype evaluated at each location-year combination (*i.e.,* the empirical best linear unbiased estimate, eBLUE), the CV%, and the number of replicates per trial. Seed yield was adjusted to 13% moisture and results were reported in bushels per acre (bu/ac). For more information about the trial field plot design and agronomic practices, please refer to the PDF files. Information from the PDF files were transcribed into CSV format files. The resulting files consist of eBLUE values for seed yield (bu/ac) of experimental genotypes and check varieties grown in field trials of soybean maturity groups (MGs) 00 through IV from 1941 to 2020. For our purposes we restricted our analyses to data belonging to MGs II and III from 1989 to 2019. Also, unusual data such as data from individual trials with estimates of reliability (i^2) less than 0.10, coefficients of variation (CV%) greater than 20%, and individual records within trials with estimated means less than 10 bu/ac were removed prior to further analyses. In addition, locations with less than three years of data were excluded from further analyses. The resulting data were comprised of 4,257 experimental genotypes evaluated at 63 locations, in 31 years, resulting in 591 location-year combinations (environments) with 39,006 yield values. However, because most experimental genotypes are only grown within appropriate MZs and are culled on an annual basis, only 0.47% of all potential combinations of experimental genotypes, locations and years exist in the data sets.

For subsequent data analyses, the trials were divided into Preliminary (PYTs) and Uniform

Regional (URTs) trials. Experimental genotypes were first evaluated in PTs, and if not culled, were subsequently evaluated in URTs. Because there are large numbers of experimental genotypes created by several public breeding programs within each MZ, the PTs are further split into two groups: PT-A and PT-B. In a given year, a PT was usually conducted at nine or more locations with two replicates of each experimental genotype evaluated at each location. Experimental genotypes retained for regional trials were evaluated at 15 locations representing a URT in the next year with three or four replicates per location. Some experimental genotypes might be evaluated in two subsequent years of URTs. Experimental genotypes with introgressed transgenic alleles were evaluated independently in trials referred to as PT/URT-RR or PT/URT-TM, depending on the transgenes. The field trials at each location utilized a randomized complete block field plot design. In addition to the experimental genotypes, entries in each field block included common check varieties ($\sim$ 3), but we noted that check varieties were seldom retained for more than four consecutive years.

5.2 Environmental data

In addition to phenotypic (PHE) data from yield trials, environmental data associated with trial locations were obtained. Elevation information was obtained from the "elevatr" package (Hollister *et al.* 2021). Soil characteristics at a depth of 5-15 cm were downloaded from Soilgrids (`https://soilgrids.org/`) with a modified R script available at `https://github.com/zecojls/downloadSoilGridsV2`, and further processed with the package "raster" (Hijmans 2021). The soil characteristics are referred to as soil variables (SV) and included: bulk density (SV1), cation exchange capacity (SV2), clay content (SV3), total nitrogen content (SV4), pH (SV5), sand content (SV6), silt content (SV7), and organic carbon content (SV8). Detailed information about SVs are available in the Soilgrids website. Latitudes for locations in the USA were downloaded from `https://simplemaps.com/data/us-cities`, and Canadian locations were obtained using Google Maps. Meteorological data, referred herein as MVs for each location were obtained from "NASA's Prediction of Worldwide Energy Resources" (NASA POWER, `https://power.larc.nasa.gov/`) with the package "nasapower" (Sparks 2018), and further processed with the "EnvRtype" package (Costa-Neto *et al.* 2021). In total, 19 MVs were retrieved on a daily basis (averages) from the average planting date until the average check variety maturity date (R8) for each environment (location by year combination). A summary of the environmental variables is provided in the appendix (Tables A1 and A2), and for more detailed information, please refer to the cited references.

5.3 Data analyses

A stage-wise approach to analyses composed of multiple models was followed (Piepho *et al.* 2012; Smith *et al.* 2001a; Frensham *et al.* 1997). The first-stage analyses were applied to individual trials within locations ($\mathbf{y_1}$), second-stage analyses were applied to all trials within locations ($\mathbf{y_2}$), and a third-stage analysis was conducted across locations and/or years ($\mathbf{y_3}$). All analyses were implemented using "Asreml-R" version 4 (Butler *et al.* 2017) in the R programming environment (R Core Team 2021). Variance components were estimated with REML followed by estimation/prediction of the fixed and random effects in Henderson's mixed models (Henderson

1950, 1963). When possible, computation time was sped-up with parallel processing by applying the "doParallel" and "foreach" packages (Microsoft Corporation and Weston 2020a,b).

5.3.1 First-stage analyses

The first-stage analyses were previously performed by the collaborators, i.e., public soybean breeders, before the data were submitted to the USDA for aggregating and reporting. Individual trials within locations were analyzed using a model in which genotypes and blocks were considered fixed effects, yielding eBLUE values, i.e., entry means for genotypes ($\mathbf{y_1}$). The eBLUE values were then analyzed with the following model to obtain an estimate of the genotypic variance (σ^2_G) that was subsequently used to estimate reliability (i^2) on an entry-mean basis:

$$\mathbf{y_1} = \boldsymbol{\mu} + \mathbf{Z}_g\mathbf{g} + \boldsymbol{\epsilon} \tag{1}$$

where $\mathbf{y_1}$ is the vector of entry means reported for each trial in the PDF files, $\boldsymbol{\mu}$ is the intercept, $\mathbf{Z}_g$ $(m \times m)$ is the incidence matrix of genotype effects, $\boldsymbol{g}$ $(m \times 1)$ is a vector of genotype random effects with $\boldsymbol{g} \sim \text{N}(\mathbf{0}, \sigma^2_G\mathbf{I})$, and $\boldsymbol{\epsilon}$ is a vector of residuals with $\epsilon \sim \text{N}(\mathbf{0}, \mathbf{\Sigma_1})$. The residual variance matrix $\mathbf{\Sigma_1}$ $(m \times m)$ is a diagonal matrix with elements equal to $\frac{1}{\text{SE}^2}$, where SE is the estimated standard error (Smith *et al.* 2001a; Frensham *et al.* 1997). The SE was estimated as $\frac{\sigma_\epsilon}{\sqrt{r}}$, where σ_ϵ is the residual standard deviation calculated from the reported CV%, and r is the reported number of replicates for each trial. Because phenotypic values from each replicate are not reported, we assumed all genotypes from a given trial had the same SE, i.e., equal replication. The i^2 was then estimated as $i^2 = \frac{\sigma^2_G}{\sigma^2_G + \frac{\sigma^2_\epsilon}{r}}$, where σ^2_G is the genotypic variance and σ^2_ϵ is the residual variance (Bernardo 2020, p. 173). Note σ^2_G could also be estimated from the variance of entry means (σ^2_F), where $\sigma^2_F = \frac{\sigma^2_\epsilon}{r} + \sigma^2_G$.

5.3.2 Second-stage analyses

Second-stage analyses utilized data from multiple trials with common entries among trials at the same location within a year (*e.g.* PT-A, PT-B, and UT), which were analyzed using:

$$\mathbf{y_2} = \boldsymbol{\mu} + \mathbf{X}_t\mathbf{t} + \mathbf{X}_g\mathbf{g} + \boldsymbol{\epsilon} \tag{2}$$

where $\mathbf{y_2}$ $(mj \times 1)$ is a vector of eBLUE values for m genotypes evaluated across j trials at location l, $\boldsymbol{\mu}$ is the intercept, $\mathbf{X}_t$ $(mj \times j)$ is the incidence matrix of fixed effects of trials, $\mathbf{t}$ $(j \times 1)$ is a vector of fixed effects of trials, $\mathbf{X}_g$ $(mj \times m)$ is the incidence matrix of fixed effects of genotypes, $\mathbf{g}$ $(m \times 1)$ is a vector of fixed effects of genotypes and $\boldsymbol{\epsilon}$ is a vector of residuals with $\epsilon \sim \text{N}(\mathbf{0}, \mathbf{\Sigma_1})$. The elements of the estimated residual variance matrix $\mathbf{\Sigma_1}$ $(mj \times mj)$ were obtained from the first stage analyses. Estimates of eBLUE values for genotypes and their SE from model 2 became input data ($\mathbf{y_3}$) for analyses across locations and years. Note the vector of observations $\mathbf{y_1}$ refers to the eBLUE values obtained from individuals trials in a single location, whereas $\mathbf{y_2}$ refers to multiple trials connected by checks or any common genotypes in a single location.

5.3.3 Multi-location and multi-year analyses

For the third-stage of analyses the following "baseline" model was used to obtain estimates of variance components across multiple locations and years:

$$\mathbf{y_3} = \boldsymbol{\mu} + \mathbf{X}_l\mathbf{l} + \mathbf{Z}_g\mathbf{g} + \mathbf{Z}_{g.l}\mathbf{g.l} + \mathbf{Z}_{yr}\mathbf{yr} + \mathbf{Z}_{g.yr}\mathbf{g.yr} + \mathbf{Z}_{l.yr}\mathbf{l.yr} + \mathbf{Z}_{g.l.yr}\mathbf{g.l.yr} + \boldsymbol{\epsilon} \quad (3)$$

where $\mathbf{y_3}$ $(mjt \times 1)$ is a vector of eBLUE values for m genotypes evaluated across j locations and t years, $\boldsymbol{\mu}$ is the intercept, $\mathbf{l}$ $(j \times 1)$ is a vector of fixed effects of locations, $\mathbf{g}$ $(m \times 1)$ is a vector of random effects of genotypes with $g \sim \text{N}(\mathbf{0}, \sigma^2_G\mathbf{I})$, $\mathbf{g.l}$ $(mj \times 1)$ is a vector of random effects of genotype by location interactions with $g.l \sim \text{N}(\mathbf{0}, \sigma^2_{GL})$, $\mathbf{yr}$ $(t \times 1)$ is a vector of random effects of years with $yr \sim \text{N}(\mathbf{0}, \sigma^2_Y)$, $\mathbf{g.yr}$ $(mt \times 1)$ is a vector of random effects of genotype by year interaction with $g.yr \sim \text{N}(\mathbf{0}, \sigma^2_{GY})$, $\mathbf{l.yr}$ $(jt \times 1)$ is a vector of random effects of location by year interaction with $l.yr \sim \text{N}(\mathbf{0}, \sigma^2_{LY})$, $\mathbf{g.l.yr}$ $(mjt \times 1)$ is a vector of random effects of genotype by location by year interaction with $g.l.yr \sim \text{N}(\mathbf{0}, \sigma^2_{GLY})$, and $\boldsymbol{\epsilon}$ is a vector of residuals with $\epsilon \sim \text{N}(\mathbf{0}\ \boldsymbol{\Sigma_2})$. $\mathbf{X}_l$ $(mjt \times j)$, $\mathbf{Z}_g$ $(mjt \times m)$, $\mathbf{Z}_{g.l}$ $(mjt \times mj)$, $\mathbf{Z}_{yr}$ $(mjt \times t)$, $\mathbf{Z}_{g.yr}$ $(mjt \times mt)$, $\mathbf{Z}_{l.yr}$ $(mjt \times jt)$, and $\mathbf{Z}_{g.l.yr}$ $(mjt \times mjt)$ are incidence matrices for their respective effects. The elements of the residual variance matrix $\boldsymbol{\Sigma_2}$ $(mjt \times mjt)$ were obtained from model 2.

5.3.4 Probability distributions of estimated variance components

A modified jackknife resampling approach was used to obtain empirical probability distributions for the variance components σ^2_G, σ^2_{GL}, σ^2_{GY}, and σ^2_{GLY}. Following Aguate *et al.* (2019) and Hartung and Piepho (2021), the data were divided into four groups representing consecutive eras of soybean cultivar development: From 1989 to 1995, from 1996 to 2003, from 2004 to 2011, and from 2012 to 2019. For the first group (1989-1995), there were 181 environments; for 1996-2003, 194 environments; for 2004-2011, 100 environments; and for 2012-2019, 116 environments. The modified jackknife approach consisted of leaving-one-environment out (instead of one observation), and then estimating the variance components with a modified version of model 3, that considered locations as a random effect with variance σ^2_L. Estimates of variance components were then combined and evaluated for a best fit to probability distributions with the package "ForestFit" (Teimouri 2021). Given the lack of data from individual plots, trial-based estimates of σ^2_ϵ from the first-stage of analyses were used (i.e., no resampling). Distributional parameters were estimated via the expectation maximization (EM) algorithm (Dempster *et al.* 1977) using the log-likelihood functions of the Gamma, Log-Logistic, Log-Normal, Burr, and F univariate and multivariate distributions. In addition to a visual comparison of the modeled distributions relative to the empirical distributions, Akaike (AIC) and Bayesian (BIC) information criteria (Akaike 1974; Schwarz 1978) as well as the Kolmogorov-Smirnov (KS), Cramer-von Mises (CM), and Anderson-Darling (AD) goodness-of-fit statistics (Stephens 1986) were considered to select the best-fit distribution for each variance component. A classical penalized criteria based on the loglikehood (AIC, BIC) provided protection from overfitting.

5.3.5 Identification of mega-environments

Herein, "ME" and "cluster" are used interchangeably. We clustered 63 locations using six criteria: (i) phenotype, i.e., seed yield (PHE), (ii) eight soil variables (SVs) plus elevation (SoilE); (iii) lati-

tude, where locations were split into two groups (Lat2), (iv) latitude, where locations were split into three groups (Lat3); (v) 19 meteorological variables (MVs) with means across years (WA); and (vi) MVs with means nested within years (WW).

With the exception of Lat2 and Lat3, the optimal number of clusters was then defined based on the Silhouette and Elbow methods using the package "factoextra" (Kassambara and Mundt 2020), followed by a K-means clustering with the R base function *kmeans()* allowing for a maximum of 1,000 iterations and 100 multiple initial configurations of the K groups.

5.3.5.1 Clustering of PHE data

Several variance-covariance structures (VCOV) for the genotype by location (Σ_{gl}) and genotype by year (Σ_{gy}) interaction terms in model 3 were evaluated. The simplest model (M3-1) assumed independent years and locations with homogeneous variances. The next set of models allowed heterogeneous variances for locations (M3-2), years (M3-3), or both (M3-4). Specific pairwise covariances for both Σ_{gl} and Σ_{gy} were assessed with models M3-5, M3-6, ..., to M3-20. In all cases, the elements of the residual matrix was assumed to be known (Model 2). Results from 19 evaluated models are presented in Table 1. The VCOV models included identity (**I**), diagonal (**D**), and factor-analytic ($\mathbf{FA}_k$) of order k (Piepho 1997; Smith *et al.* 2001b, 2015). The best-fit model was selected according to the AIC selection criteria. For the FA models, the overall percentage of genetic variance accounted by each k factor, defined as $100[\text{tr}(\mathbf{\Lambda\Lambda}')/\text{tr}(\mathbf{\Lambda\Lambda}' + \mathbf{\Psi})]$, where "tr" is the trace of the matrix, $\mathbf{\Lambda}$ ($j \times k$) is the matrix of loadings, and $\mathbf{\Psi}$ ($j \times j$) is a diagonal matrix of specific variances associated with each location, was also considered. Models were selected based upon the AIC, the overall percentage of genotype by location [% Var(GL)] and genotype by year [% Var(GY)] variances explained by the FA models. With the best-fit FA model, locations were clustered based on the estimated Σ_{gl} loadings (Bustos-Korts 2017; Burgueño *et al.* 2008) after Varimax rotation. Genetic correlations between locations (**C**) were further estimated by $\mathbf{C} = \mathbf{DGD}$, where $\mathbf{G} = (\mathbf{\Lambda\Lambda}' + \mathbf{\Psi})$ is the estimator of genetic variances, and $\mathbf{D}$ is a diagonal matrix composed by the inverse of the square root of the diagonal values of $\mathbf{G}$ (Smith *et al.* 2015).

5.3.5.2 Clustering of SV data

First the SVs (including elevation) were centered and scaled to a unit variance. Subsequently, a principal component analysis (PCA) by non-linear iterative partial least squares (Wold 1966) was performed to reduce collinearity with the "pcaMethods" package (Stacklies *et al.* 2007). The number of principal components (PC) was selected with a 90% threshold of cumulative variance explained, followed by a Varimax rotation.

5.3.5.3 Clustering of MV data

Prior to conducting cluster analyses, a Critical Environmental Regressor through Informed Search (CERIS) procedure proposed by Li *et al.* (2018) was used to identify relevant MVs. The method consists of screening meteorological data in all environments to identify a period (window) of days after planting with the greatest Pearson correlation between the population means (i.e.,

environmental means) and the MVs. The idea is to identify periods of meteorological data that are most likely to affect stages of growth and development associated with the phenotypic results (yield, plant height, etc.). We further modified their approach to account for genotype by location deviations within years as follows:

$$\mathbf{y_3} = \boldsymbol{\mu} + \mathbf{X}_l\mathbf{l} + \mathbf{X}_g\mathbf{g} + \boldsymbol{\epsilon} \tag{4}$$

where $\mathbf{y_3}$ $(mj \times 1)$ is a vector eBLUE values of m genotypes in j locations, $\boldsymbol{\mu}$ is the intercept, $\mathbf{l}$ $(j \times 1)$ is a vector of fixed effects of locations, $\mathbf{g}$ $(m \times 1)$ is a vector of fixed effects of genotypes, and $\boldsymbol{\epsilon}$ is a vector of residuals with $\epsilon \sim \mathbf{N}(\mathbf{0}, \boldsymbol{\Sigma_2})$. $\mathbf{X}_l$ $(mj \times j)$ and $\mathbf{X}_g$ $(m \times m)$ are incidence matrices for their respective effects, and $\boldsymbol{\Sigma_2}$ $(mj \times mj)$ was previously defined. Model 4 was applied within years. The residuals represent the genotype by location deviations nested within years. Each location was then represented as the average of the residuals squared. The CERIS was computed for observed location-year combinations within (WW) and across years (WA), and the best window (i.e., highest correlation) for each of the MVs was selected for clustering. For WA, correlations were computed for each MV with the 591 observed environments. For example, if a given location was observed in five out of 31 years, five environmental means were computed with the same selected window, and the location represented as the mean of these five values. On the other hand, for WW, each observed year can have its own best window. A minimum window of seven days was considered in all cases. After identifying the most relevant window for each MVs, the resulting data were centered and scaled to unit variance. Subsequently clustering was conducted as described for the SVs. Note that for both WA and WW, the input data for the clustering analysis was a matrix of centered and scaled environmental means with dimension of 63 rows by 19 columns, which represent the number of locations and MVs, respectively.

5.3.5.4 Effectiveness of clustering

We used the ratio of correlated responses from selection across all environments relative to direct responses to selection within MEs (CR/DR) (Atlin *et al.* 2000a; Bustos-Korts 2017) as a metric to assess the relative effectiveness of clustering environments into MEs. As previously demonstrated, CR/DR can be determined using variance components obtained from linear models:

$$\begin{aligned}\mathbf{y_3} =& \mathbf{X}_r\mathbf{r} + \mathbf{Z}_{l(r)}\mathbf{l}_{(\mathbf{r})} + \mathbf{Z}_g\mathbf{g} + \mathbf{Z}_{g.l_r}\mathbf{g}.\mathbf{l}_{(\mathbf{r})} + \mathbf{Z}_y\mathbf{y} + \mathbf{Z}_{y.r}\mathbf{y}.\mathbf{r} \\ &+ \mathbf{Z}_{g.r}\mathbf{g}.\mathbf{r} + \mathbf{Z}_{l_r.y}\mathbf{l}_{(\mathbf{r})}.\mathbf{y} + \mathbf{Z}_{g.y}\mathbf{g}.\mathbf{y} + \mathbf{Z}_{g.y.r}\mathbf{g}.\mathbf{y}.\mathbf{r} + \mathbf{Z}_{g.y.l_r}\mathbf{g}.\mathbf{y}.\mathbf{l}_{(\mathbf{r})} + \boldsymbol{\epsilon}\end{aligned} \tag{5}$$

where $\mathbf{r}$ is a vector of fixed effects of clusters, and $\mathbf{l}_{(\mathbf{r})}$, $\mathbf{g}.\mathbf{l}_{(\mathbf{r})}$, $\mathbf{y}.\mathbf{r}$, $\mathbf{g}.\mathbf{r}$, $\mathbf{l}_\mathbf{r}.\mathbf{y}$, $\mathbf{g}.\mathbf{y}.\mathbf{r}$, and $\mathbf{g}.\mathbf{y}.\mathbf{l}_\mathbf{r}$, are random vectors with specific variances of locations within clusters, genotype by location within clusters interaction, year by clusters interaction, genotype by cluster interaction, locations nested in clusters by year interaction, and genotype by year by location within clusters, respectively. $\mathbf{X}_r$, and $\mathbf{Z}_{l_r}$ up to $\mathbf{Z}_{g.y.l_r}$, are incidence matrices for their respective effects and dimensions. The remaining model terms were previously defined.

Estimates of variance components from model 5 were used to obtain CR/DR:

$$CR/DR = \rho_g\sqrt{\frac{i_{\mathrm{L}}^2}{i_{\mathrm{SR}}^2}} \tag{6}$$

where ρ_g is the correlation between estimated genotypic effects in the non-clustered and clustered sets of environments and i^2_{L} and i^2_{SR} are the estimated reliabilities of genotype means in the non-clustered and clustered sets of environments, respectively. If $CR/DR < 1$, response to selection will be more effective if selections are made within clusters (Atlin *et al.* 2000a; Bustos-Korts 2017). Note that it is possible for $CR/DR > 1$, indicating that selection will be more effective if selection is based on eBLUE values obtained from non-clustered environments. As per (Atlin *et al.* 2000a) The terms in equation 6 are defined as follows:

$$\rho_g = \frac{\sigma^2_G}{\sqrt{\sigma^2_G(\sigma^2_G + \sigma^2_{GR})}} \tag{7}$$

$$i^2_{\text{L}} = \frac{\sigma^2_G}{\sigma^2_G + \frac{\sigma^2_{GR}}{nr} + \frac{\sigma^2_{GL(R)}}{nl \times nr} + \frac{\sigma^2_{GY}}{ny} + \frac{\sigma^2_{GRY}}{ny \times nr} + \frac{\sigma^2_{GL(R)Y}}{nl \times nr \times ny}} \tag{8}$$

$$i^2_{\text{SR}} = \frac{\sigma^2_G + \sigma^2_{GR}}{\sigma^2_G + \sigma^2_{GR} + \frac{\sigma^2_{GL(R)}}{nl} + \frac{\sigma^2_{GY}}{ny} + \frac{\sigma^2_{GRY}}{ny} + \frac{\sigma^2_{GL(R)Y}}{nl \times ny}} \tag{9}$$

where σ^2_G, σ^2_{GR}, $\sigma^2_{GL(R)}$, σ^2_{GY}, σ^2_{GRY}, and $\sigma^2_{GL(R)Y}$ are the genotypic, genotype by cluster, genotype by location nested in cluster, genotype by year, genotype by clusters by year, and genotype by location nested in cluster by year variance components, respectively. nr and ny are the harmonic means for the number of clusters and years in which genotypes were observed, respectively. nl is the median number of locations, obtained from harmonic means within clusters. The residual terms were omitted due to the lack of replicated data within environments.

The Jaccard similarity coefficient was used to compare the coincidence of locations within cluster across clustering-types. For the sake of simplicity, only the σ^2_{GR} was estimated for each clustering-type with the jackknife approach presented in section 5.3.4. Lastly, to evaluate if locations were simply allocated to clusters by chance, we randomly assigned locations into two, three, and four clusters, and assessed its CR/DR. This process was repeated 100 times.

6 Results

6.1 Single-trial and location analysis

The number of evaluated genotypes by year including checks ranged from 126 to 233, and the number of locations per year ranged from 10 to 32 (Table A3). Estimates of i^2 from single-trial analyses ranged from 0.10 to 0.99, with a median value of 0.55 (Figure 1). The majority of trials (844 out of 1423) had i^2 values greater than 0.50. The coefficient of variation (CV%) ranged from 1.30 to 19.9%, with a median value of 7.60% (Figure 1-A). Genotypic eBLUE values ranged from 10.16 to 112.40 bu/ac. Across years, there has been a positive trend for seed yield with an average increase of 0.49 bu/ac per year (Figure 1-B), although the relative contributions of genetic and non-genetic factors to this trend requires further analyses and is the subject of future research.

6.2 Variance components

Most of the estimated phenotypic variance for seed yield among annual soybean field trials has been due to location (σ^2_L) and location by year interaction (σ^2_{LY}) effects (Table 2). Among the estimated GEI variance components (σ^2_{GL} + σ^2_{GY} + σ^2_{GLY}), the static contribution (σ^2_{GL}) represented 26.30% (Table 2, model M3-1).

Estimated variance components reveal distinct multi-modal distributions over time (Figure 2). For example, the estimated genotypic variance (σ^2_G) more than doubled from $\sim$ 3.54 (bu/ac)^2 for the period 1989-1995 to $\sim$ 7.56 (bu/ac)^2 for the period 1996-2003. For the years 2004-2011, the estimated σ^2_G decreased to $\sim$ 6.67, but then increased to $\sim$ 7.49 for the most recent period of 2012 to 2019. While the smallest magnitudes of estimated GEI variance components were usually associated with the period from 1989 to 1995, subsequent changes across years were unique to each of the estimated variance component. A similar pattern was observed for empirical estimates of location, year, and location by year interaction variances (Figure A1).

Several multi-modal models were evaluated for goodness of fit for the empirical distributions of variance components (Table A4). The best-fit models for variance components across time consisted of: a mixture of five Log-Logistic distributions for the empirical distribution of σ^2_G, three Log-Logistic distributions for the empirical distribution of σ^2_{GL}, six Log-Logistic distributions for the empirical distribution of σ^2_{GY}, and five Gamma distributions for the empirical distribution of σ^2_{GLY} (Figure 3). For the empirical distribution of estimated residual variances (σ^2_ϵ), obtained directly rather then through jackknife resampling, the best-fit model was a univariate Log-Logistic distribution (Figure 4). Maximum Likelihood estimates for the distributional model parameters are reported for the selected models (Table 3).

The criteria for determining best-fit models included low AIC and BIC values, goodness of fit statistics (KS, CM, AD), as well as visual graphical alignment of the model with the plotted empirical distributions. In total, 102 models were assessed. Plots with empirical and fitted cumulative distribution functions are provided in Figures A2 and A3.

6.3 Clusters of meta-environments

The six clustering criteria revealed that the observed 63 environments can be divided into at least two and at most three MEs. Clusters based on PHE criteria had the best (lowest) relative effectiveness value, CR/DR $= 0.62$ (Table 4). This value was computed using results from analysis from model 5, and PHE determined by estimates of genotype by location interaction (Σ_{gl}) from model M3-18 (Table 1). The simplest, but unrealistic model (M3-1) assumed homogeneous and independent variances and had the greatest AIC value.

For PHE, the optimal number of clusters was three, while for SoilE, WW, and WA it was two. For all clustering types, both Silhouette and Elbow criteria indicated similar results (Figure A4). The number of environments within clusters ranged from 7 to 54 (Table 4 and Figure 5). Calculated Jaccard distance metrics among the clusters created by different criteria showed that many clusters have common members. Out of the 75 pairwise similarity indexes, 68 presented non-zero values. The greatest similarity was between clusters 1 and 2 from PHE and Lat2, respectively (Table A5).

The best-fit model (M3-18) accounted for heterogeneous variances and pairwise genetic correlations with FA matrices of order $k = 5$ and $k = 2$ for Σ_{gl} and Σ_{gly}, accounting for 90.2% and

87.2% of the genetic variances, respectively (Table 1). Pairwise genetic correlations between the 63 observed locations show a higher average correlation within phenotypic clusters compared to across phenotypic clusters (Figure A5).

Prior to k-means clustering, a non-linear PCA was performed with centered and scaled environmental variables (elevation, SV, and MV). The non-linearity relationship for most variables was evident from scatterplots (Figures A6, A7, and A8). For SoilE, WA, and WW, the selected number of PC were five (93.06% of variance explained), four (93.42%), and six (91.12%), respectively (results not shown). Furthermore, for MV, just a small proportion of the data was used due to the CERIS biological filter. For WA, the CERIS revealed that four out of 19 variables had negative correlations between genotype by location deviations and the mean of the environmental variable for the selected window. The selected windows (i.e., with the highest Pearson correlation) were smaller than 20 days, and most of them started at the beginning of the cropping season (Table A1 and Figure A9). For WW and for the sake of simplicity, only the highest window for each MV are presented (Table A2), although locations were clustered as described in section 5.3.5.3.

The effectiveness of clustering was assessed using CR/DR, which compares the response to selection in the divided and undivided sets of environments. The metric, computed from estimated variance components from model 5 for each clustering type, presented ratios smaller than one in five out of the six cases. These results suggested an improvement in the response to selection when selecting directly within clusters (i.e., regions, ME) versus selecting across all locations for PHE, SoilE and Lat2, while a moderate response to selection was suggested for Lat3 and WA. For WW, no improvements were predicted. The relative effectiveness metric is affected by the correlation between genotypic effects in the undivided and divided sets of environments (ρ_g) and the reliability within subregion (i_{SR}). The PHE presented the lowest ρ_g value with an increase in the reliability (from i_L^2 to i_{SR}^2), followed by Lat2 and SoilE. For Lat3 and WA, although effective, the reliabilities remained constant. Regarding the differences within fitted clusters, for SoilE, with the exception of pH and bulk density, all variables presented a big contrast between the two clusters. For WA, these differences were more subtle. For example, for growing degree-days (GDD), clusters 1 and 2 presented a mean difference of 2.61 units. For temperature related variables (mean, maximum, minimum, and range), the difference ranged between 0.21 and 2.94 units. When locations were randomly assigned to two, three, or four clusters, the mean CR/DR values were always bigger than one, with large decreases in the reliability (Table 4).

REML estimates of variances decreased for σ^2_G when clusters were included in the complete dataset, with the exception of WW. The observed ratios σ^2_{GR}/σ^2_G and $\sigma^2_{GR}/\sigma^2_{GL}$ were greather than 0.50 for PHE, SoilE, and Lat2. In terms of the partitioning of σ^2_{GL}, the $\sigma^2_{GL(R)}$ portion was substantially reduced for PHE, Lat2, and Lat3. The σ^2_{GR} was better captured by PHE, SoilE, and Lat2, being ineffective for WW (estimate bounded at zero due to REML properties). On the other hand, for σ^2_{GLY}, just a small portion of the variation was captured by σ^2_{GRY} (Table 2). Both PHE and Lat2 clustering types were able to greatly capture σ^2_{GR} according to analysis for each group of years (Figure 6). Lastly, large reductions in the variation of years (σ^2_Y) were observed for SoilE and Lat2.

7 Discussion

Analyses of balanced data sets produced by METs using least square and mixed model estimators provide unbiased estimates of variance components. However, data generated by annual PYTs and URTs are unbalanced and sparse because every year most field plots (experimental units) are reserved for new experimental genotypes and most previously evaluated experimental genotypes are culled on an annual basis. For example, reports of soybean seed yield in MZ's II and III evaluated in PYTs and URTs conducted from 1989 and 2019 included less than one percent of all possible combinations genotypes, locations and years. As a consequence estimators of the variance components are biased (Rothschild *et al.* 1979). A question to consider is whether the biased estimators produce large bias in the estimates.

An additional challenge in using soybean seed yield data from PYTs and URTs is that the data consist of eBLUE values from individual trials within locations for each genotype. These values were transcribed from reports of individual trials formatted as PDF files. To the best of our knowledge, all genotypes were evaluated in replicated field trials organized as randomized complete block design (RCBD's) that were analyzed with a linear model consisting of fixed block and genotypic effects and random residual effects, where the residual effects were assumed to be estimated using equal numbers of replicates per genotype within a field trial. Implicitly this is equivalent to assuming there were no missing plots within any of the field trials, which is highly unlikely. Further, the estimates of σ^2_{GLY}, σ^2_{GL} and σ^2_{GY} are confounded and biased by σ^2_{ϵ}, i.e., plot to plot variability.

A consequence of using eBLUE values instead of individual plot data is that estimates of variance components needed to be obtained in multiple stages. If the reports of field trials had provided individual plot data it would have been possible to produce a variance–covariance matrix associated with adjusted means from each trial. Indeed, if individual plot data are provided, the field plot designs need not be restricted to RCBD's wherein the covariances among plots may be substantial and assumptions of independence among plots is inappropriate (Möhring and Piepho 2009). Under such field conditions use of spatial models and lattice designs where replicates are considered fixed and blocks as random effects (Möhring *et al.* 2015) can be utilized and the mixed model framework can provide appropriate weights for analyses of data combined across trials, locations and years. Toward this goal public soybean breeders are working with curators of "SoyBase" (Drs. Rex Nelson and David Grant, personal communication, 2020) to include results from individual plots in future reports of PYTs and URTs.

If data were obtained from only URTs they would provide very little information for estimating σ^2_{GLY} and σ^2_{GY} because most experimental genotypes are not grown for more than one year of URTs. Although check varieties were replicated across multiple years, the checks are a small sample of genotypes representing mostly commercial varieties. Thus, interpretation of variance components based on only check varieties is limited. In order to broaden the inferences about interactions involving genotypes and years in MZs II and III, we included eBLUE values from the PYTs, thus providing at least two years of data for a broader base of genotypes.

Even with inclusion of data from PYTs there is potential for bias in the estimates of σ^2_{GLY}, σ^2_{GL}, σ^2_{GY}, and σ^2_{G} due to the phenomenon of missing data between years. Many of the experimental genotypes evaluated in the PYTs will be culled before becoming entries in a subsequent year of URTs. As a consequence, the missing data are missing at random (MAR). If genotypes are MAR, selection is ignorable, and hence they will produce unbiased estimates of variance components

in likelihood-based analysis (Piepho and Möhring 2006; Rothschild *et al.* 1979). Note MAR does not state the missing genotypes are randomly eliminated (*i.e.,* no selection), but instead it depends on the observed data due to selection (Piepho and Möhring 2006). Also, URTs can have experimental genotypes that were not included in a previous year of PYTs, a condition known as missing completely at random (MCAR) (Little and Rubin 2020; Rubin 1976). Fortunately recent work from Hartung and Piepho (2021) demonstrated that both MAR and MCAR conditions for field trials conducted in sequential years result in minor bias for likelihood-based estimators of variance components (Piepho and Möhring 2006; Little and Rubin 2020; Hartung and Piepho 2021), a result previously noted for MAR by Piepho and Möhring (2006).

For the sake of interpreting variance components involving genotypes, we recognize that PYTs and URTs include genotypes from multiple breeding programs. Each breeding program operates independently with distinct breeding objectives and breeding strategies for distinct markets. For example, a couple of the breeding programs have objectives that include not only high seed yields, but also greater seed protein for food markets. Seed protein is negatively correlated with seed yield. Thus, the estimates of variance components involving genotypes from these trials is likely greater than it is for any individual breeding program.

Despite the various caveats mentioned above, the eBLUE values for genotypes from both PYTs and URTs provided estimates of variance components with similar relative magnitudes as other studies. For example, we determined that environmental, i.e., non-genetic sources of variability were the predominant source of variance for seed yield. Similar results were found in wheat in California (USA) (George and Lundy 2019), winter wheat field trials in Germany between 1983 to 2014 (Laidig *et al.* 2017a) and winter rye (Laidig *et al.* 2017b). Estimates of variance components (Table 2) revealed that the interactions of genotype with locations (σ^2_{GL}) was larger than genotype by year interactions (σ^2_{GY}). Also, the three-way interaction among genotype, location, and years (σ^2_{GLY}), was greater than either of the two way interactions involving genotypes. The same pattern was observed when years were combined into four 7 to 8 years periods. Friesen *et al.* (2016) found similar results for winter wheat evaluated in Canada from 2000 to 2009, where the reported σ^2_{GLY} represented 4.1% of the total variation, and both σ^2_{GL} and σ^2_{GY} together represented less than 2%. Similar trends were also observed for yield in wheat (George and Lundy 2019; Laidig *et al.* 2017a; Arief *et al.* 2015), rye (Laidig *et al.* 2017b), barley, maize, and sunflower (Laidig *et al.* 2008). While seed yields for multiple crops indicate $\sigma^2_{GL} > \sigma^2_{GY}$, it is not consistent for all traits. For example, Laidig *et al.* (2017b) evaluated the variation in crude protein content, amylogram viscosity and temperature in winter rye varieties, and reported that $\sigma^2_{GY} > \sigma^2_{GL}$, and that the year-to-year (σ^2_Y) variation was more important than the variation from location to location (σ^2_L). Authors attributed this to the rye seed susceptibility towards wetness, low temperature, and radiation during harvest time.

Aguate *et al.* (2019) simulated different METs with variable numbers of genotypes, locations and years to mimic wheat trials. They found that adding years is more beneficial than adding genotypes or locations for obtaining unbiased estimates of genotypic related variance components. Furthermore, even in highly imbalanced datasets, estimates from at least 8 years of trials produced less than 5% bias in the estimates, compared to biases of ∼18% for σ^2_G, >40% for σ^2_{GL}, and >15% for σ^2_{GY}, when only two years of METs were considered. Simulation results from Hartung and Piepho (2021) further demonstrated that non-significant bias can be achieved for estimates of σ^2_G, σ^2_{GL}, and σ^2_{GY}, with decreasing dropout rate and increasing number of years of testing. Given the dropout rate relies on the objectives and budget constraints of the breeding

program, in order to be confident with the REML estimates of variances obtained from METs, they recommended that at least seven to eight years of trials should be included. This result and the fact that soybean breeding cycles in public programs require seven to eight years, we created four subsets of data consisting of seven to eight years. The reader should keep in mind that the goal of minimizing bias in estimators is distinct from the magnitude and proportion of estimates of variance components. The former is a concern for algorithmic estimators while the latter is of concern for breeding decisions about whether to use more locations or more years in their crop species and for traits under selection.

Estimates of variance components were also used to fit parametric probability distributions and to quantify the effectiveness of dividing the sampled 63 environments/locations into MEs. For the distributions, a jackknife resampling approach was implemented and consisted of leaving-one-environment out and estimating the variance components in each group of years (1989-1995, 1996-2003, 2004-2011, 2012-2019). The selected distributions were Log-Logistic and Gamma (Tables 3 and A4). Especially due to the well-known properties of the analysis of variance (ANOVA) among the breeding community, there is a misconception regarding the distribution of estimates of variance components. If the underlying population are normally distributed, the mean squares are distributed as a chi-square (χ^2), whereas normality and independence are requirements to compute valid F tests from ANOVA (Rencher and Schaalje 2007, Chapter 5). The χ^2 distribution is further used for inferences about variance uncertainty, but as mentioned, it does assume that the random variable is normally distributed. The computed empirical distributions from jackknife can be of any form (distribution), likewise other approaches such hierarchical Bayesian models can be employed to obtain posterior distributions of variance estimates. But regardless of the type of inference, our motivation was essentially to capture the GEI trend. For example, the point estimate for the genotype by location variance in the whole period was $\hat{\sigma}^2_{GL} = 4.8$ (bu/ac)^2 (Table 2). This is a valid estimate, however, it does not allow a trend quantification. The GEI trend are crucial as they can be incorporated in simulation pipelines to depict genetic and non-genetic trends, which is current a topic under investigation.

Logically, plant breeders are motivated to ask: What is an environment? Costa-Neto and Fritsche-Neto (2021) defined environment as "... an emergent property derived from the balance of inputs and frequency across the plant's lifetime," and from an agronomic point of view, "... a certain time window between planting date and harvesting." Over 60 years ago, Comstock, R. E. and Moll (1962) described the differences between micro and macro-environment, and explained that GEI is the result of fluctuations in the macro-environment during a crop's lifetime. More recently, introduction of enviromics/envirotyping (Costa-Neto *et al.* 2021; Xu 2016; Cooper *et al.* 2014), coupled with high-throughput phenotyping/genotyping are being investigated as an approach to connect environment and biology for sustainable food production. While some agronomic research has focused on variation among agronomic systems linked with ideotype breeding, genotypic sources of variability continue to be the primary approach used for genetic improvement by plant breeders.

Results from the cluster analyses showed that all sampled environments can be effectively divided into three clusters using FA models. The obtained clusters can be considered disjoint subsets of environments with minimal genotypic crossover interaction (COI) (Burgueño *et al.* 2008; Cooper and DeLacy 1994). Burgueño *et al.* (2008) analysed a maize MET dataset from CIMMYT and identified five clusters of environments by fitting a FA(2) model. The authors computed the Euclidean distance between pairs of environments from the estimated loading matrix

($\mathbf{\Lambda}$, rotated with singular value decomposition), and environments were clustered based on complete linkage clustering strategy. Bustos-Korts (2017) analysed data embracing TPE in Denmark, Germany, The Netherlands, and the United Kingdom, with a FA(1) model. The results suggested improvements in the response to selection mostly for Denmark, where the CRS ratio was 0.93. More recently, Smith *et al.* (2021) proposed a new way to define groups of environments that exhibit minimal COI based on FA models. The idea is to take advantage of the traditional interpretation of factor and principal component analysis, and classify environments into clusters based on the sign (positive or negative) of the estimated and rotated factor loadings.

The identification of homogeneous environments was also accomplished by considering soil plus elevation (SoilE) and meteorological variables when CERIS was applied across environments (WA). The rational is that a portion of the GEI results from static, repeatable variation (Crespo-Herrera *et al.* 2021; Yan 2016). It is well-known that temperature is a key driving force in the rate of seasonal plant growth (Setiyono *et al.* 2007), which is why GDD is commonly a base unit in crop models (Holzworth *et al.* 2014). Photoperiod plays a significant role in soybean plant development, notably the change from vegetative to reproductive growth. Floral induction is essentially daylenght and temperature-independent (i.e. conversion of shoot apical and nodal meristems from a vegetative to floral mode). In soybeans, this induction occurs as soon as the first unifoliolate leaflets emerge and expand, becoming capable of measuring the night length (from dusk to dawn). Once floral induction occurs at a given apical or axillary node, the few-celled vegetative apical zone is transformed from a vegetative development pathway into a floral inflorescence development pathway. The development pathway is back under thermal control (Setiyono *et al.* 2007). Soybean is a quantitative long-night length sensing (not a short-day length sensing), and hence highly influenced by photoperiod and therefore by the latitude of the growing region/trial (Jackson 2009). This is a major reason that different soybean maturity groups are grown at different latitudes (Mourtzinis and Conley 2017). The estimated clusters (with the exception of the *ad-hoc* Lat2 and Lat3) follow a certain pattern in terms of latitude, which was also confirmed by the Jaccard similarity. In addition, the inclusion of the URT data in SoyBase would facilitate identifying critical crop growth periods in order to narrow down the amount of environmental data used by CERIS.

Herein, we identified MEs through reliable estimates of variance components. However, other environmental subsets would have been formed using different clustering strategies (Burgueño *et al.* 2008). Given this type of approach is an unsupervised learning (i.e., we do not know the truth about MEs), the objective is always to discover an interpretable grouping of members. We addressed interpretation using effectiveness of clustering (Atlin *et al.* 2000a). Other strategies for clustering can also be tested, for example, empirical knowledge of the TPE. But regardless of the definition/identification of MEs, breeders can take advantage of best linear unbiased prediction (BLUP) that borrows information (strength) between MEs from the genotype by ME interactions. This type of modelling can be highly beneficial for MEs that rely on a small number of locations (Buntaran *et al.* 2021; Piepho *et al.* 2016; Piepho and Möhring 2005). A natural continuation for this work would be to (i) evaluate the effectiveness of a combined cluster from soil, elevation, and meteorological variables filtered by CERIS; (ii) evaluate if BLUP based-models will improve the selection response upon regionalization in the Uniform Soybean Cooperative Tests; (iii) select an appropriate model that might account for heterogeneous covariances among MEs as well genetic relationships, because we confined attention to the compound symmetry model in order to facilitate comparison of the different clustering types; and (iv) leverage how far back

in the historical data should we go in order to take maximum advantage of the data in current models. It is also worth investigating if modelling maturity groups (specially when more data is considered) would enhance the ability of finding meaningful MEs using phenotypic models.

8 Conclusion

We dissected the sources of soybean seed yield variation using reports from Soybean Cooperative Tests for maturity groups II and III. We determined that sampled sets of environments can be split into mega-environments according to phenotypic, geographic, and meteorological information. Reasonable estimates of variance components are essential for analyses of data from historical field trials. Furthermore, it was possible to monitor trends in variance components involving genotypes in terms of parametric probability distributions. Historical field trials also evaluate traits like seed quality and size, iron deficiency chlorosis, green stem, seed oil, and protein content. The approach presented herein can be applied to variation of multiple economically important quantitative traits. Finally, in addition to the practical and theoretical results applied to soybean genetic improvement, the analysis performed in this study may be applied to quantitative traits evaluated in any crop using multi-environment trials.

9 Declaration of competing interest

The authors state there is no conflict of interest.

10 Author contributions

MDK and WDB conceived the research; MDK performed the statistical analyses and wrote the first drafts of the manuscript; KOGD provided insights into the methodology and helped in the interpretation of the results; AKS provided interpretation of the results and guidance in scientific writing; and WDB and AKS were responsible for acquiring funding to support the research. All authors critically revised drafts of the manuscript and approved the final version.

11 Acknowledgments

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12 Tables and Figures

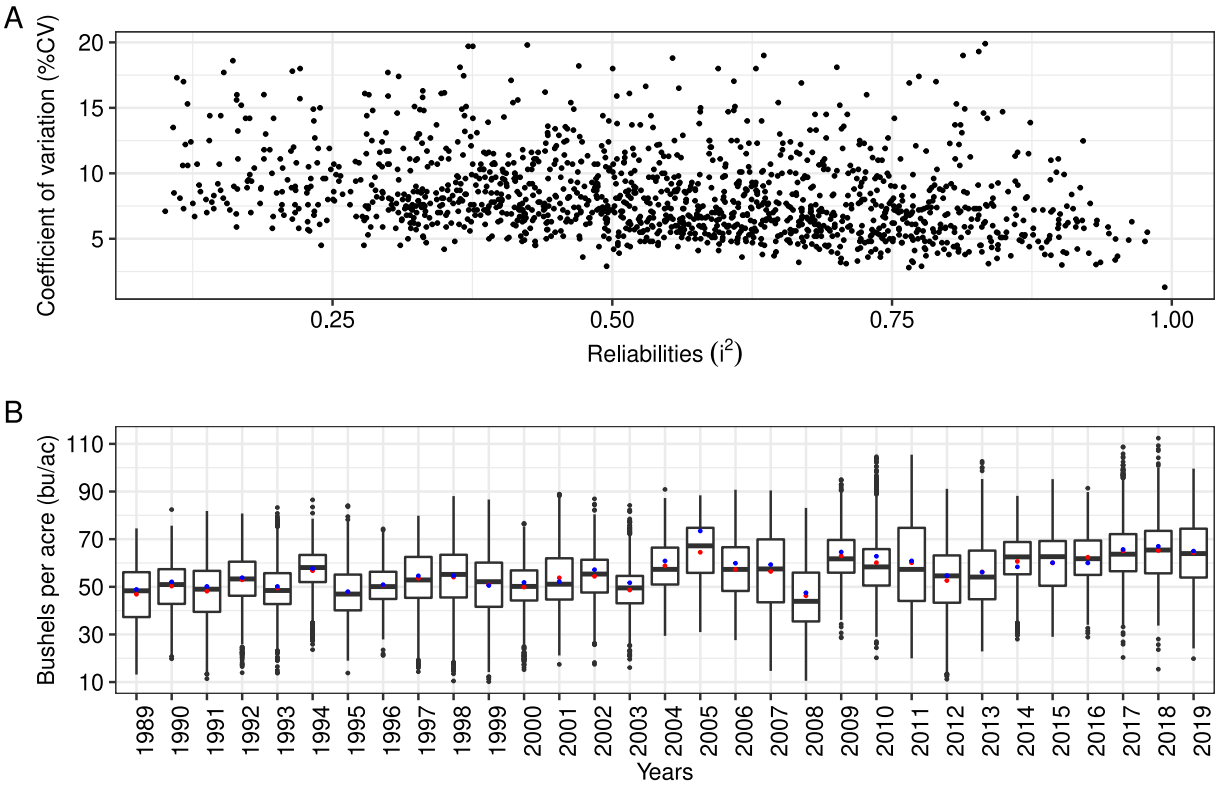

Figure 1: Estimates of reliabilities (i^2) and coefficient of variation (%CV) for 1423 soybean field trials conducted from 1989 to 2019 (A), and boxplots of empirical best linear unbiased estimates (eBLUEs) of seed yield plotted by year (B) from 1989 to 2019. Red dots in B depict the average yield of experimental cultivars excluding checks, whereas blue dots depict the average yield of the check varieties.

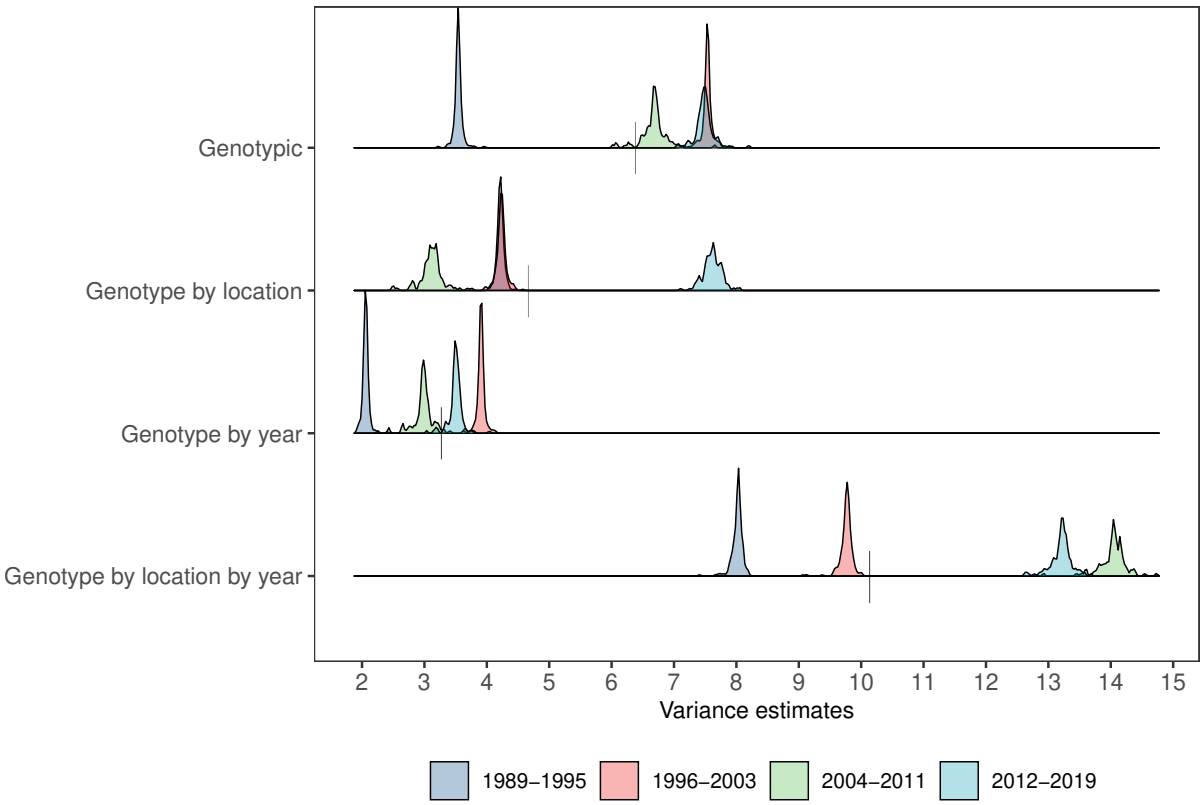

Figure 2: Empirical distributions of estimated variance components consisting of genotypic, genotype by location, genotype by year, and genotype by location by year variances for groups of years 1989-1995, 1996-2003, 2004-2011, and 2012-2019. Empirical estimates were obtained using a jackknife leave-one-location out method. Vertical bars on the x-axis represent point estimates across all years.

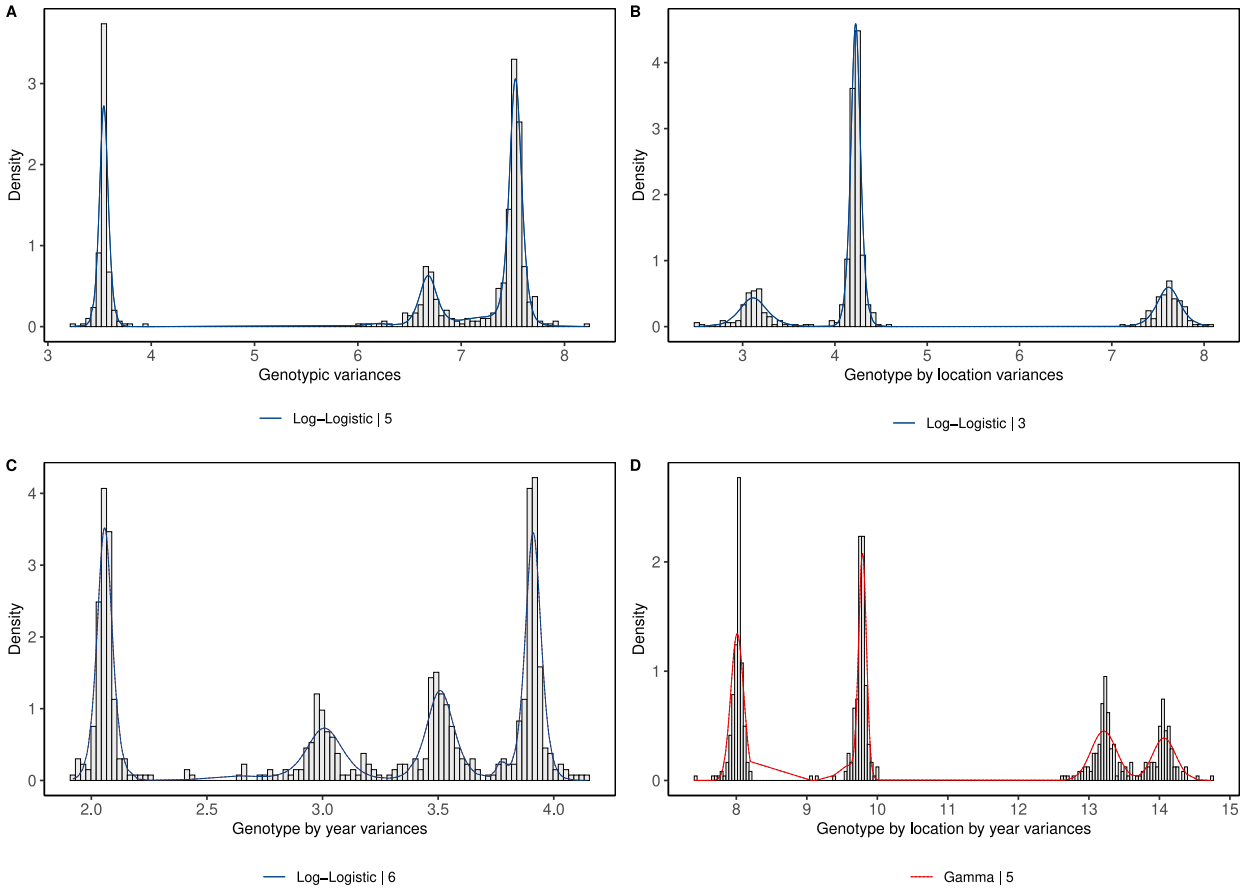

Figure 3: Empirical distributions and probability density function (PDF) of variance components for genotypic (A), genotype by location (B), genotype by year (C), and genotype by location by year (D). Empirical estimates were obtained using a jackknife leave-one-location out method. The best-fit models for PDF's are presented with different colors, and include the name of the distribution and its mixture number.

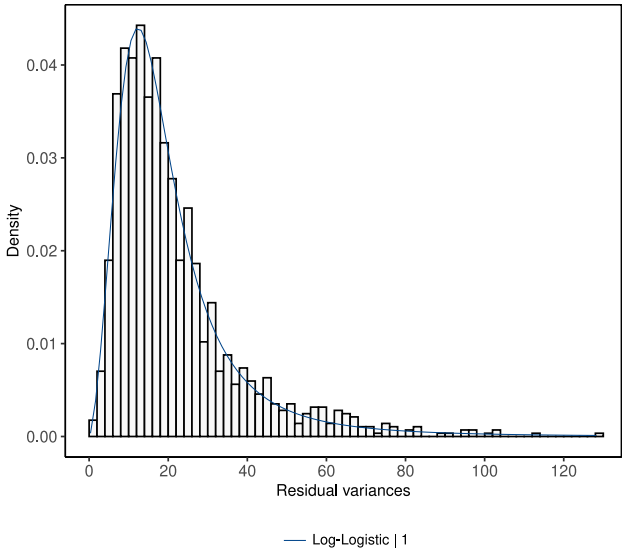

Figure 4: Empirical distribution and probability density function (PDF) for the unimodal Log-Logistic model of residual estimates from individual trials.

Table 1: Models and criteria used to evaluate the models for purposes of clustering locations into groups representing the most likely target population of environments. The best-fit model (M3-18) is highlighted in bold.

Model	Covariance structure[a]			Evaluation criteria		
	Σ_{gl}	Σ_{gy}	Σ_{ϵ}	AIC	% Var(GL)	% Var(GY)
M3-1	$\mathbf{I} \otimes \mathbf{I}$	$\mathbf{I} \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	175294	-	-
M3-2	$\mathbf{D} \otimes \mathbf{I}$	$\mathbf{I} \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	174256	-	-
M3-3	$\mathbf{I} \otimes \mathbf{I}$	$\mathbf{D} \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	174964	-	-
M3-4	$\mathbf{D} \otimes \mathbf{I}$	$\mathbf{D} \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	173951	-	-
M3-5	$\mathbf{FA}_1 \otimes \mathbf{I}$	$\mathbf{D} \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172845	48.5	-
M3-6	$\mathbf{FA}_2 \otimes \mathbf{I}$	$\mathbf{D} \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172560	67.3	-
M3-7	$\mathbf{FA}_3 \otimes \mathbf{I}$	$\mathbf{D} \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172359	78.2	-
M3-8	$\mathbf{FA}_4 \otimes \mathbf{I}$	$\mathbf{D} \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172303	87.6	-
M3-9	$\mathbf{FA}_1 \otimes \mathbf{I}$	$\mathbf{FA}_1 \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172754	48.0	63.4
M3-10	$\mathbf{FA}_2 \otimes \mathbf{I}$	$\mathbf{FA}_2 \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172472	66.2	82.5
M3-11	$\mathbf{FA}_3 \otimes \mathbf{I}$	$\mathbf{FA}_3 \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172288	77.0	96.4
M3-12[b]	$\mathbf{FA}_4 \otimes \mathbf{I}$	$\mathbf{FA}_4 \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	-	-	-
M3-13	$\mathbf{FA}_2 \otimes \mathbf{I}$	$\mathbf{FA}_1 \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172494	66.4	61.0
M3-14	$\mathbf{FA}_3 \otimes \mathbf{I}$	$\mathbf{FA}_1 \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172296	77.6	61.0
M3-15	$\mathbf{FA}_3 \otimes \mathbf{I}$	$\mathbf{FA}_2 \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172276	77.5	82.9
M3-16	$\mathbf{FA}_4 \otimes \mathbf{I}$	$\mathbf{FA}_2 \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172205	84.7	86.3
M3-17	$\mathbf{FA}_4 \otimes \mathbf{I}$	$\mathbf{FA}_3 \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172234	87.3	96.7
M3-18	$\mathbf{FA}_5 \otimes \mathbf{I}$	$\mathbf{FA}_2 \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	**171990**	**90.2**	**87.2**
M3-19	$\mathbf{FA}_5 \otimes \mathbf{I}$	$\mathbf{FA}_3 \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	172196	88.9	96.9
M3-20[b]	$\mathbf{FA}_6 \otimes \mathbf{I}$	$\mathbf{FA}_2 \otimes \mathbf{I}$	$\mathbf{\Sigma_2}$	-	-	-

[a] The evaluated variance-covariance structures were identity (**I**), diagonal (**D**), and factor analytic ($\mathbf{FA}_k$) from order k = 1, ..., 6. $\mathbf{\Sigma_2}$ is the residual variance matrix assumed to be known from single trial and location analysis.

[b] Singularity in the Average Information matrix.

Table 2: Point estimates and standard error of variance components for seed yield computed from Soybean Cooperative Tests (1989-2019) using Model 3-1 (baseline) and six clustering methods for clustering locations into mega-environments using Model 5.

Variance components[a]	Model 3-1	Clustering criteria[b]					
		PHE	SoilE	Lat2	Lat3	WA	WW
$\hat{\sigma}^2_G$	6.3 (0.3)	4.5 (0.4)	5.0 (0.5)	4.8 (0.4)	5.6 (0.4)	5.7 (0.4)	6.3 (0.4)
$\hat{\sigma}^2_L$	58.9 (13.1)[c]	56.4 (12.6)	58.6 (13.1)	59.5 (13.2)	48.9 (11.4)	59.1 (13.2)	60.5 (13.5)
$\hat{\sigma}^2_Y$	15.3 (5.4)	13.4 (6.6)	10.6 (7.2)	11.5 (6.3)	15.1 (6.1)	16.5 (6.5)	15.0 (6.4)
$\hat{\sigma}^2_{LY}$	81.4 (5.2)	74.6 (5.0)	79.9 (5.2)	76.5 (5.0)	77.7 (5.2)	80.0 (5.1)	79.9 (5.2)
$\hat{\sigma}^2_{GL}$	4.8 (0.3)	-	-	-	-	-	-
$\hat{\sigma}^2_{GY}$	3.2 (0.3)	2.8 (0.3)	2.9 (0.4)	3.2 (0.3)	3.1 (0.3)	2.8 (0.3)	3.0 (0.3)
$\hat{\sigma}^2_{GLY}$	10.1 (0.3)	-	-	-	-	-	-
$\hat{\sigma}^2_{RY}$	-	12.2 (5.7)	7.6 (6.5)	10.2 (5.2)	5.9 (3.9)	2.8 (3.7)	3.6 (4.1)
$\hat{\sigma}^2_{GR}$	-	4.1 (0.3)	2.9 (0.4)	2.5 (0.3)	1.5 (0.2)	1.1 (0.3)	0.0 (0.0)
$\hat{\sigma}^2_{GL(R)}$	-	2.5 (0.3)	4.6 (0.3)	3.6 (0.3)	3.8 (0.3)	4.4 (0.3)	4.8 (0.3)
$\hat{\sigma}^2_{GRY}$	-	0.9 (0.2)	0.5 (0.3)	0.1 (0.2)	0.1 (0.2)	0.7 (0.3)	0.5 (0.2)
$\hat{\sigma}^2_{GL(R)Y}$	-	9.7 (0.3)	9.9 (0.3)	10.3 (0.3)	10.2 (0.3)	9.9 (0.3)	9.9 (0.3)
Clusters	-	3	2	2	3	2	2

[a] Genotypic ($\hat{\sigma}^2_G$), location ($\hat{\sigma}^2_L$), year ($\hat{\sigma}^2_Y$), location by year ($\hat{\sigma}^2_{LY}$), genotype by location ($\hat{\sigma}^2_{GL}$), genotype by year ($\hat{\sigma}^2_{GY}$), genotype by location by year ($\hat{\sigma}^2_{GLY}$), cluster by year ($\hat{\sigma}^2_{RY}$), genotype by cluster ($\hat{\sigma}^2_{GR}$), genotype by location nested in cluster ($\hat{\sigma}^2_{GL(R)}$), genotype by cluster by year ($\hat{\sigma}^2_{GRY}$), and genotype by location nested in cluster by year ($\hat{\sigma}^2_{GL(R)Y}$).

[b] Phenotypic (PHE), soil and elevation (SoilE), latitude split into two groups (Lat2), latitude split into three groups (Lat3), weather from means across years (WA), and weather from means within years (WW).

[c] Locations were modelled as a random effect in model 3-1 (Table 1).

Table 3: Maximum Likelihood estimates of parameters for the best-fit univariate and multivariate probability distributions for empirical distributions obtained using jackknife resampling. Estimates of residual variance (σ^2_ϵ) were obtained from trials conducted from 1989 to 2019.

Variance[a]	Distribution	Number of distributions	Parameters[b]		
			$\hat{w}_i$	$\hat{\alpha}_i$	$\hat{\beta}_i$
σ^2_G	Log-Logistic	5	0.30	125.98	3.54
			0.14	119.87	6.68
			0.08	44.03	7.23
			0.01	88.03	6.18
			0.47	192.69	7.53
σ^2_{GL}	Log-Logistic	3	0.63	122.27	4.23
			0.17	32.00	3.12
			0.20	92.56	7.62
σ^2_{GY}	Log-Logistic	6	0.02	258.41	3.77
			0.31	172.61	3.91
			0.20	89.05	3.51
			0.30	94.97	2.06
			0.02	32.81	2.64
			0.15	57.79	3.01
σ^2_{GLY}	Gamma	5	0.05	4078.04	0.0024
			0.31	7755.00	0.0011
			0.20	5306.62	0.0025
			0.28	31922.50	0.0003
			0.16	7301.02	0.0020
σ^2_ϵ	Log-Logistic	1	1	2.56	17.03

[a] Genotypic (σ^2_G), genotype by location (σ^2_{GL}), genotype by year (σ^2_{GY}), and genotype by location by year (σ^2_{GLY}) variance components.

[b] Estimates of weight parameters (w_i) sums to one, and both Gamma and Log-Logistic distributions include a shape (α_i) and scale (β_i) parameter.

Table 4: The ratio of correlated responses from selection across all environments relative to direct responses to selection within mega-environments (CR/DR) for each clustering type. ρ_g is the correlation between estimated genotypic effects in the non-clustered and clustered sets of environments, i^2_{L} and i^2_{SR} are the reliabilities of genotype means in the non-clustered and clustered sets of environments, respectively.

Clustering type[a]	Number of		Estimates of			
	Clusters	Locations	$\hat{\rho}_g$	$\hat{i}^2_{\mathrm{L}}$	$\hat{i}^2_{\mathrm{SR}}$	$\hat{CR/DR}$
PHE	3	36 / 7 / 20	0.72	0.38	0.51	0.62
SoilE	2	9 / 54	0.88	0.39	0.45	0.81
Lat2	2	35 / 28	0.81	0.39	0.48	0.73
Lat3	3	16 / 36 / 11	0.89	0.44	0.44	0.89
WA	2	25 / 38	0.92	0.45	0.45	0.92
WW	2	19 / 44	1.00	0.51	0.43	1.08
	2	-	0.99	0.52	0.44	1.07
At random	3	-	0.99	0.51	0.39	1.14
	4	-	0.99	0.51	0.36	1.18

[a] Phenotypic (PHE), soil and elevation (SoilE), latitude split into two groups (Lat2), latitude split into three groups (Lat3), weather from means across years (WA), and weather from means within years (WW).

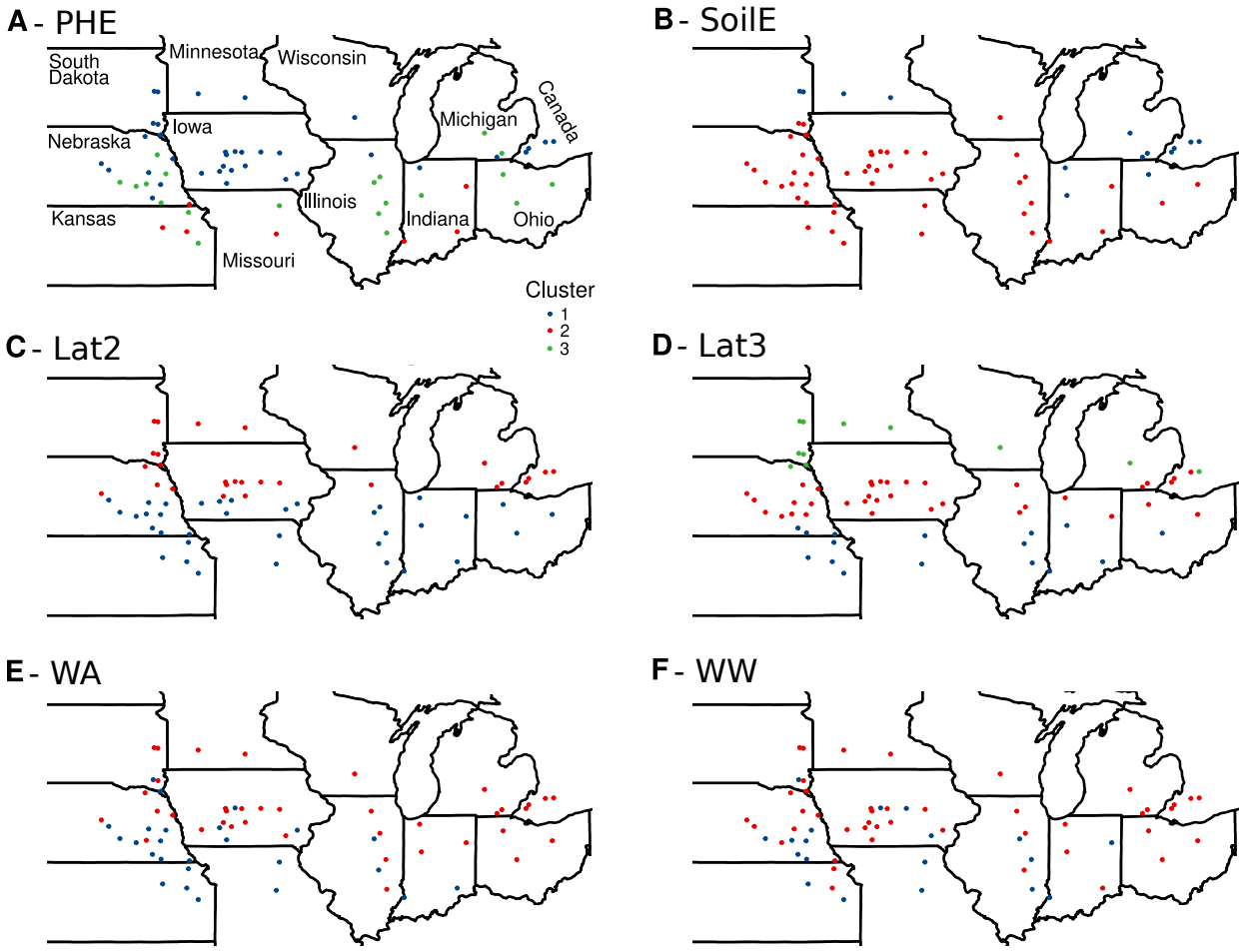

Figure 5: Geographic visualization of the target population of environments divided according to phenotypic (A), soil + elevation (B), latitude split into two groups (C), latitude split into three groups (D), weather across years (E), and weather within years (F) clustering types. In (A), the states' names are provided for geographic orientation.

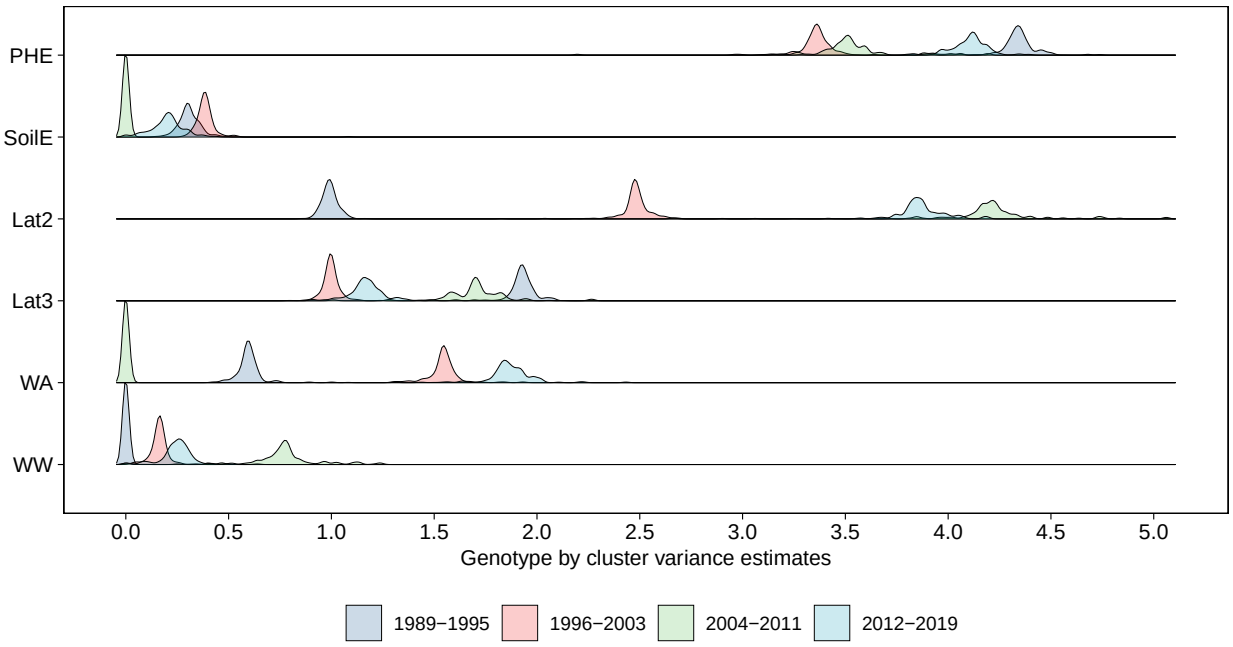

Figure 6: Jackknife estimates of genotype by cluster variances for the groups of years 1989-1995, 1996-2003, 2004-2011, and 2012-2019, for phenotypic (PHE), soil + elevation (SoilE), latitude split into two groups (Lat2), latitude split into three groups (Lat3), weather across years (WA), and weather within years (WW) clustering types.

References

Aguate, F., J. Crossa, and M. Balzarini, 2019 Effect of missing values on variance component estimates in multi-environment trials. Crop Science **59**: 508–517.

Akaike, H., 1974 A New Look at the Statistical Model Identification. IEEE Transactions on Automatic Control **19**: 716–723.

Annicchiarico, P., 2021 Breeding gain from exploitation of regional adaptation: An alfalfa case study. Crop Science **61**: 2254–2271.

Arief, V. N., I. H. DeLacy, J. Crossa, T. Payne, R. Singh, H. J. Braun, T. Tian, K. E. Basford, and M. J. Dieters, 2015 Evaluating Testing Strategies for Plant Breeding Field Trials: Redesigning a CIMMYT International Wheat Nursery. Crop Science **55**: 164–177.

Atlin, G. N., R. J. Baker, K. B. McRae, and X. Lu, 2000a Selection response in subdivided target regions. Crop Science **40**: 7–13.

Atlin, G. N., K. B. McRae, and X. Lu, 2000b Genotype x region interaction for two-row barley yield in Canada. Crop Science **40**: 1–6.

Bernardo, R., 2020 *Breeding for Quantitative Traits in Plants*. Stemma Press, Woodbury, MN, third edition.

Buntaran, H., J. Forkman, and H. P. Piepho, 2021 Projecting results of zoned multi-environment trials to new locations using environmental covariates with random coefficient models: accuracy and precision. Theoretical and Applied Genetics **134**: 1513–1530.

Buntaran, H., H. P. Piepho, J. Hagman, and J. Forkman, 2019 A cross-validation of statistical models for zoned-based prediction in cultivar testing. Crop Science **59**: 1544–1553.

Burgueño, J., J. Crossa, P. L. Cornelius, and R. C. Yang, 2008 Using factor analytic models for joining environments and genotypes without crossover genotype x environment interaction. Crop Science **48**: 1291–1305.

Bustos-Korts, D., 2017 *Modelling of genotype by environment interaction and prediction of complex traits across multiple environments as a synthesis of crop growth modelling, genetics and statistics*. Ph.D. thesis, Wageningen University.

Bustos-Korts, D., M. P. Boer, K. Chenu, B. Zheng, S. Chapman, and F. A. Van Eeuwijk, 2021 Genotype-specific P-spline response surfaces assist interpretation of regional wheat adaptation to climate change. In Silico Plants **3**: 1–23.

Butler, D. G., B. R. Cullis, A. R. Gilmour, B. G. Gogel, and R. Thompson, 2017 ASReml-R Reference Manual Version 4.

CIMMYT, 1989 *Toward the 21st century*. CIMMYT, Mexico, D.F.

Comstock, R. E. and H. Moll, 1962 Genotype-environment interactions. In *Statistical Genetics and Plant Breeding*, pp. 164–196, National Academy of Sciences-National Research Council, Washington, D.C.

Cooper, M. and I. H. DeLacy, 1994 Relationships among analytical methods used to study genotypic variation and genotype-by-environment interaction in plant breeding multi-environment experiments. Theoretical and Applied Genetics **88**: 561–572.

Cooper, M., C. D. Messina, D. Podlich, L. R. Totir, A. Baumgarten, N. J. Hausmann, D. Wright, and G. Graham, 2014 Predicting the future of plant breeding: complementing empirical evaluation with genetic prediction. Crop and Pasture Science **65**: 311.

741 Costa-Neto, G. and R. Fritsche-Neto, 2021 Enviromics: bridging different sources of data, building one framework.
742 Crop Breeding and Applied Biotechnology **21**: 393521–393533.

743 Costa-Neto, G., G. Galli, H. F. Carvalho, J. Crossa, and R. Fritsche-Neto, 2021 EnvRtype: a software to interplay
744 enviromics and quantitative genomics in agriculture. G3 (Bethesda, Md.) **11**.

745 Crespo-Herrera, L. A., J. Crossa, J. Huerta-Espino, S. Mondal, G. Velu, P. Juliana, M. Vargas, P. Pérez-
746 Rodríguez, A. K. Joshi, H. J. Braun, and R. P. Singh, 2021 Target Population of Environments for Wheat Breeding
747 in India: Definition, Prediction and Genetic Gains. Frontiers in Plant Science **12**: 1–15.

748 Cullis, B. R., A. Smith, C. Hunt, and A. Gilmour, 2000 An examination of the efficiency of Australian crop variety
749 evaluation programmes. Journal of Agricultural Science **135**: 213–222.

750 da Silva, K. J., P. E. Teodoro, M. J. da Silva, L. P. R. Teodoro, M. J. Cardoso, V. d. P. C. Godinho, J. H. Mota, G. A.
751 Simon, F. D. Tardin, A. R. da Silva, F. L. Guedes, and C. B. de Menezes, 2021 Identification of mega-environments
752 for grain sorghum in Brazil using GGE biplot methodology. Agronomy Journal **113**: 1–12.

753 Dempster, A. P., N. M. Laird, and D. B. Rubin, 1977 Maximum Likelihood from Incomplete Data Via the EM Algo-
754 rithm. Journal of the Royal Statistical Society: Series B (Methodological) **39**: 1–38.

755 Dias, K. O., H. P. Piepho, L. J. Guimarães, P. E. Guimarães, S. N. Parentoni, M. O. Pinto, R. W. Noda, J. V.
756 Magalhães, C. T. Guimarães, A. A. Garcia, and M. M. Pastina, 2020 Novel strategies for genomic prediction
757 of untested single-cross maize hybrids using unbalanced historical data. Theoretical and Applied Genetics **133**:
758 443–455.

759 Falconer, D. S., 1952 The Problem of Environment and Selection. The American Naturalist **86**: 293–298.

760 Frensham, A., B. Cullis, and A. Verbyla, 1997 Genotype by Environment Variance Heterogeneity in a Two-Stage
761 Analysis. Biometrics **53**: 1373–1383.

762 Friesen, L. F., A. L. Brûlé-Babel, G. H. Crow, and P. A. Rothenburger, 2016 Mixed model and stability analysis
763 of spring wheat genotype yield evaluation data from Manitoba, Canada. Canadian Journal of Plant Science **96**:
764 305–320.

765 Gauch, H. and R. Zobel, 1997 Identifying mega-environments and targeting genotypes. Crop Science **37**: 311–326.

766 George, N. and M. Lundy, 2019 Quantifying genotype × environment effects in long-term common wheat yield
767 trials from an agroecologically diverse production region. Crop Science **59**: 1960–1972.

768 Haldane, J., 1947 The interaction of nature and nurture. Annals of Human Genetics **17**: 197–205.

769 Hartung, J. and H. Piepho, 2021 Effect of missing values in multi-environmental trials on variance component
770 estimates. Crop Science pp. 1–11.

771 Henderson, C. R., 1950 Estimation of genetic parameters. Annals of Mathematical Statistics **21**: 309–310.

772 Henderson, C. R., 1963 Selection index and expected genetic advance. In *Statistical genetics and plant breeding*, p.
773 623, National Academy of Genetic Advance - National Research Council, Washington DC.

774 Hijmans, R. J., 2021 raster: Geographic Data Analysis and Modeling.

775 Hollister, J., T. Shah, A. L. Robitaille, M. W. Beck, and M. Johnson, 2021 elevatr: Access Elevation Data from
776 Various APIs.

777 Holzworth, D. P., N. I. Huth, P. G. deVoil, E. J. Zurcher, N. I. Herrmann, G. McLean, K. Chenu, E. J. van
778 Oosterom, V. Snow, C. Murphy, A. D. Moore, H. Brown, J. P. Whish, S. Verrall, J. Fainges, L. W. Bell, A. S.
779 Peake, P. L. Poulton, Z. Hochman, and P. J. Thorburn, 2014 APSIM – Evolution towards a new generation of
780 agricultural systems simulation. Environmental modelling & software : with environment data news. **62**: 327–350.

781 Jackson, S. D., 2009 Plant responses to photoperiod. New Phytologist **181**: 517–531.

782 Johannsen, W., 1911 The genotype conception of heredity. The American Naturalist **45**: 129–159.

783 Kang, M., 2020 Genotype-environment interaction and stability analyses: an update. In *Quantitative genetics, ge-*
784 *nomics and plant breeding*, edited by M. Kang, chapter 9, pp. 140–161, CABI, second edition.

785 Kassambara, A. and F. Mundt, 2020 factoextra: Extract and Visualize the Results of Multivariate Data Analyses.

786 Kleinknecht, K., J. Möhring, F. Laidig, U. Meyer, and H. P. Piepho, 2016 A simulation-based approach for evalu-
787 ating the efficiency of multienvironment trial designs. Crop Science **56**: 2237–2250.

788 Krishnamurthy, S. L., P. C. Sharma, D. K. Sharma, K. T. Ravikiran, Y. P. Singh, V. K. Mishra, D. Burman, B. Maji,
789 S. Mandal, S. K. Sarangi, R. K. Gautam, P. K. Singh, K. K. Manohara, B. C. Marandi, G. Padmavathi, P. B.
790 Vanve, K. D. Patil, S. Thirumeni, O. P. Verma, A. H. Khan, S. Tiwari, S. Geetha, M. Shakila, R. Gill, V. K.
791 Yadav, S. K. Roy, M. Prakash, J. Bonifacio, A. Ismail, G. B. Gregorio, and R. K. Singh, 2017 Identification of
792 mega-environments and rice genotypes for general and specific adaptation to saline and alkaline stresses in India.
793 Scientific Reports **7**: 1–14.

794 Laidig, F., T. Drobek, and U. Meyer, 2008 Genotypic and environmental variability of yield for cultivars from 30
795 different crops in German official variety trials. Plant Breeding **127**: 541–547.

796 Laidig, F., H. P. Piepho, D. Rentel, T. Drobek, U. Meyer, and A. Huesken, 2017a Breeding progress, environmental
797 variation and correlation of winter wheat yield and quality traits in German official variety trials and on-farm
798 during 1983–2014. Theoretical and Applied Genetics **130**: 223–245.

799 Laidig, F., H. P. Piepho, D. Rentel, T. Drobek, U. Meyer, and A. Huesken, 2017b Breeding progress, variation, and
800 correlation of grain and quality traits in winter rye hybrid and population varieties and national on-farm progress
801 in Germany over 26 years. Theoretical and Applied Genetics **130**: 981–998.

802 Li, X. X., T. Guo, Q. Mu, X. X. Li, and J. Yu, 2018 Genomic and environmental determinants and their interplay
803 underlying phenotypic plasticity. Proceedings of the National Academy of Sciences of the United States of America
804 **115**: 6679–6684.

805 Little, R. J. A. and D. B. Rubin, 2020 *Statistical analysis with missing data*. Wiley series in probability and statistics,
806 John Wiley & Sons, third edition.

807 Mackay, I., H. P. Piepho, and A. A. F. Garcia, 2019 Statistical Methods for Plant Breeding. In *Handbook of Statistical*
808 *Genomics*, edited by D. Balding, I. Moltke, and J. Marioni, chapter 17, pp. 501–530, John Wiley & Sons Ltd, Hoboken,
809 NJ, fourth edition.

810 Malosetti, M., J. M. Ribaut, and F. A. van Eeuwijk, 2013 The statistical analysis of multi-environment data: Mod-
811 eling genotype-by-environment interaction and its genetic basis. Frontiers in Physiology **4**: 1–17.

812 Meyer, U., F. Laidig, and T. Drobek, 2011 Optimization of number of trials in official VCU trial series of Germany.
813 Biuletyn Oceny Odmian **33**: 73–82.

814 Microsoft Corporation and S. Weston, 2020a doParallel: Foreach Parallel Adaptor for the 'parallel' Package.

815 Microsoft Corporation and S. Weston, 2020b foreach: Provides Foreach Looping Construct.

816 Möhring, J. and H. P. Piepho, 2009 Comparison of weighting in two-stage analysis of plant breeding trials. Crop
817 Science **49**: 1977–1988.

818 Möhring, J., E. Williams, and H. P. Piepho, 2015 Inter-block information: to recover or not to recover it? Theoretical
819 and Applied Genetics **128**: 1541–1554.

820 Mourtzinis, S. and S. P. Conley, 2017 Delineating soybean maturity groups across the United States. Agronomy
821 Journal **109**: 1397–1403.

822 Oakey, H., B. Cullis, R. Thompson, J. Comadran, C. Halpin, and R. Waugh, 2016 Genomic selection in multi-
823 environment crop trials. G3: Genes, Genomes, Genetics **6**: 1313–1326.

824 Patterson, H. D. and R. Thompson, 1971 Recovery of inter-block information when block sizes are unequal.
825 Biometrika **58**: 545–554.

826 Piepho, H.-P., 1997 Analyzing genotype-environment data by mixed models with multiplicative terms. Biometrics
827 **53**: 761–766.

828 Piepho, H. P. and J. Möhring, 2005 Best linear unbiased prediction of cultivar effects for subdivided target regions.
829 Crop Science **45**: 1151–1159.

830 Piepho, H. P. and J. Möhring, 2006 Selection in cultivar trials - Is it ignorable? Crop Science **46**: 192–201.

831 Piepho, H. P., J. Möhring, T. Schulz-Streeck, and J. O. Ogutu, 2012 A stage-wise approach for the analysis of
832 multi-environment trials. Biometrical Journal **54**: 844–860.

833 Piepho, H. P., M. F. Nazir, M. Qamar, A. U. R. Rattu, Riaz-Ud-Din, M. Hussain, G. Ahmad, Fazal-E-Subhan,
834 J. Ahmad, Abdullah, K. B. Laghari, I. A. Vistro, M. Sharif Kakar, M. A. Sial, and M. Imtiaz, 2016 Stability
835 analysis for a countrywide series of wheat trials in Pakistan. Crop Science **56**: 2465–2475.

836 Pigliucci, M., 2001 *Phenotypic plasticity: Beyond nature and nurture*. The Johns Hopkins University Press, Baltimore,
837 MD.

838 R Core Team, 2021 R: A Language and Environment for Statistical Computing.

839 Rencher, A. C. and G. B. Schaalje, 2007 *Linear Models in Statistics*. John Wiley & Sons, Inc., Hoboken, NJ, USA.

840 Rothschild, M., C. Henderson, and R. L. Quaas, 1979 Effects of selection on variances and covariances of simulated
841 first and second lactations. Journal of Dairy Science **62**: 996–1002.

842 Rubin, D., 1976 Inference and missing data. Biometrika **63**: 581–592.

843 Rutkoski, J. E., 2019 Estimation of realized rates of genetic gain and indicators for breeding program assessment.
844 Crop Science **59**: 981–993.

845 Schwarz, G., 1978 Estimating the dimension of a model. The Annals of Statistics **6**: 461–464.

846 Setiyono, T. D., A. Weiss, J. Specht, A. M. Bastidas, K. G. Cassman, and A. Dobermann, 2007 Understanding and
847 modeling the effect of temperature and daylength on soybean phenology under high-yield conditions. Field Crops
848 Research **100**: 257–271.

849 Singh, D. P., A. K. Singh, and A. Singh, 2021 *Plant Breeding and Cultivar Development*. Academic Press, first edition.

850 Smith, A., B. Cullis, and A. Gilmour, 2001a The analysis of crop variety evaluation data in Australia. Australian
851 and New Zealand Journal of Statistics **43**: 129–145.

852 Smith, A., B. R. Cullis, and R. Thompson, 2001b Analyzing variety by environment data using multiplicative mixed
853 models and adjustments for spatial field trend. Biometrics **57**: 1138–1147.

854 Smith, A., A. Norman, H. Kuchel, and B. Cullis, 2021 Plant Variety Selection Using Interaction Classes Derived
855 From Factor Analytic Linear Mixed Models: Models With Independent Variety Effects. Frontiers in Plant Science
856 **12**.

Smith, A. B., A. Ganesalingam, H. Kuchel, and B. R. Cullis, 2015 Factor analytic mixed models for the provision of grower information from national crop variety testing programs. Theoretical and Applied Genetics **128**: 55–72.

Sparks, A., 2018 nasapower: NASA-POWER Data from R.

Sprague, G. and W. Federer, 1951 A comparison of variance components in corn yield trials: II. error, year x variety, location x variety, and variety components. Agronomy Journal **43**: 535–541.

Stacklies, W., H. Redestig, M. Scholz, D. Walther, and J. Selbig, 2007 pcaMethods – a Bioconductor package providing PCA methods for incomplete data. Bioinformatics **23**: 1164–1167.

Stephens, M., 1986 Tests based on edf statistics. In *Goodness-of-fit techniques*, edited by R. D'Agostino and M. Stephens, pp. 97–194, Marcel Dekker, New York.

Tabery, J., 2008 R. A. Fisher, Lancelot Hogben, and the origin(s) of genotype-environment interaction. Journal of the History of Biology **41**: 717–761.

Teimouri, M., 2021 ForestFit: Statistical Modelling for Plant Size Distributions.

USDA, 2021 Uniform Soybean Tests, Northern Region.

Windhausen, V. S., S. Wagener, C. Magorokosho, D. Makumbi, B. Vivek, H. P. Piepho, A. E. Melchinger, and G. N. Atlin, 2012 Strategies to subdivide a target population of environments: Results from the CIMMYT-led maize hybrid testing programs in Africa. Crop Science **52**: 2143–2152.

Wold, H., 1966 Estimation of principal components and related models by iterative least squares. In *Multivariate Analysis*, edited by P. R. Krishnajah, pp. 391–420, Academic Press, New York.

Xu, Y., 2016 Envirotyping for deciphering environmental impacts on crop plants. Theoretical and Applied Genetics **129**: 653–673.

Yan, W., 2016 Analysis and handling of G x E in a practical breeding program. Crop Science **56**: 2106–2118.

Yan, W., J. Frégeau-Reid, D. Pageau, R. Martin, J. Mitchell-Fetch, M. Etienne, J. Rowsell, P. Scott, M. Price, B. de Haan, A. Cummiskey, J. Lajeunesse, J. Durand, and E. Sparry, 2010 Identifying essential test locations for oat breeding in Eastern Canada. Crop Science **50**: 504–515.

Yan, W., L. A. Hunt, Q. Sheng, and Z. Szlavnics, 2000 Cultivar Evaluation and Mega-Environment Investigation Based on the GGE Biplot. Crop Science **40**: 597–605.

Yan, W. and I. Rajcan, 2002 Biplot Analysis of Test Sites and Trait Relations of Soybean in Ontario. Crop Science **42**: 11–20.

Zdziarski, A. D., L. G. Woyann, A. S. Milioli, R. Zanella, L. V. Dallacorte, M. C. Panho, and G. Benin, 2019 Mega-environment identification for soybean (Glycine max) breeding and production in Brazilian Midwest region. Plant Breeding **138**: 336–347.

13 Appendix

Table A1: Meteorological variables and selected window across years (WA) with the highest Person correlation ($\hat{\rho}$).

Window			Parameter			$\hat{\rho}$
Beggining	End	Size	Name	Code	Description	
1	8	7	ALLSKY_SFC_LW_DWN	MV1	Downward thermal infrared (longwave) radiative flux	0.1719
9	16	7	ALLSKY_SFC_SW_DWN	MV2	Insolation incident on a horizontal surface	0.1956
9	16	7	ETP	MV3	Evapotranspiration	0.2007
3	10	7	FRUE	MV4	Effect of temperature on radiation use efficiency	0.2021
3	10	7	GDD	MV5	Growing degree-days	0.2012
9	16	7	n	MV6	Duration of sunshine hours	0.1904
52	71	19	PETP	MV7	Deficit of evapotranspiration	-0.1522
52	72	20	PRECTOT	MV8	Rainfall precipitation	-0.1267
3	10	7	PTR	MV9	Photothermal ratio (GDD / daylight in hours)	0.2037
3	10	7	PTT	MV10	Photothermal time (GDD $\times$ daylight in hours)	0.1981
1	8	7	RH2M	MV11	Relative humidity at 2 meters	0.0899
3	10	7	SPV	MV12	The slope of saturation vapor pressure curve	0.1995
3	11	8	T2M	MV13	Daily average temperature at 2 meters	0.2052
3	14	11	T2M_MAX	MV14	Daily minimum temperature at 2 meters	0.1849
1	8	7	T2M_MIN	MV15	Daily maximum temperature at 2 meters	0.2168
1	8	7	T2M_RANGE	MV16	Daily temperature range at 2 meters	-0.0943
1	8	7	T2MDEW	MV17	Dew point at 2 meters	0.2081
7	14	7	VPD	MV18	The deficit of vapor pressure	0.0909
69	77	8	WS2M	MV19	Wind speedy at 2 meters	-0.1397

Table A2: Meteorological variables and selected window within years (WW) with the highest Person correlation ($\hat{\rho}$).

Window				Parameter			$\hat{\rho}$
Beggining	End	Size	Year	Name	Code	Description	
40	47	7	2005	ALLSKY_SFC_LW_DWN	MV1	Downward thermal infrared (longwave) radiative flux	-0.8768
46	55	9	1990	ALLSKY_SFC_SW_DWN	MV2	Insolation incident on a horizontal surface	-0.8845
46	55	9	1990	ETP	MV3	Evapotranspiration	-0.8571
12	44	32	2005	FRUE	MV4	Effect of temperature on radiation use efficiency	-0.9041
12	44	32	2005	GDD	MV5	Growing degree-days	-0.9033
99	124	25	2014	n	MV6	Duration of sunshine hours	0.9126
104	120	16	2005	PETP	MV7	Deficit of evapotranspiration	0.9226
108	120	12	2005	PRECTOT	MV8	Rainfall precipitation	0.9198
17	44	27	2005	PTR	MV9	Photothermal ratio (GDD / daylight in hours)	-0.8852
11	44	33	2005	PTT	MV10	Photothermal time (GDD $\times$ daylight in hours)	-0.9178
31	38	7	2016	RH2M	MV11	Relative humidity at 2 meters	-0.8825
11	44	33	2005	SPV	MV12	The slope of saturation vapor pressure curve	-0.9004
12	44	32	2005	T2M	MV13	Daily average temperature at 2 meters	-0.8996
12	26	14	2005	T2M_MAX	MV14	Daily minimum temperature at 2 meters	-0.8835
108	124	16	2005	T2M_MIN	MV15	Daily maximum temperature at 2 meters	-0.8533
112	121	9	2016	T2M_RANGE	MV16	Daily temperature range at 2 meters	0.8354
5	14	9	2004	T2MDEW	MV17	Dew point at 2 meters	0.7601
22	36	14	2015	VPD	MV18	The deficit of vapor pressure	0.9195
66	80	14	1990	WS2M	MV19	Wind speedy at 2 meters	-0.8760

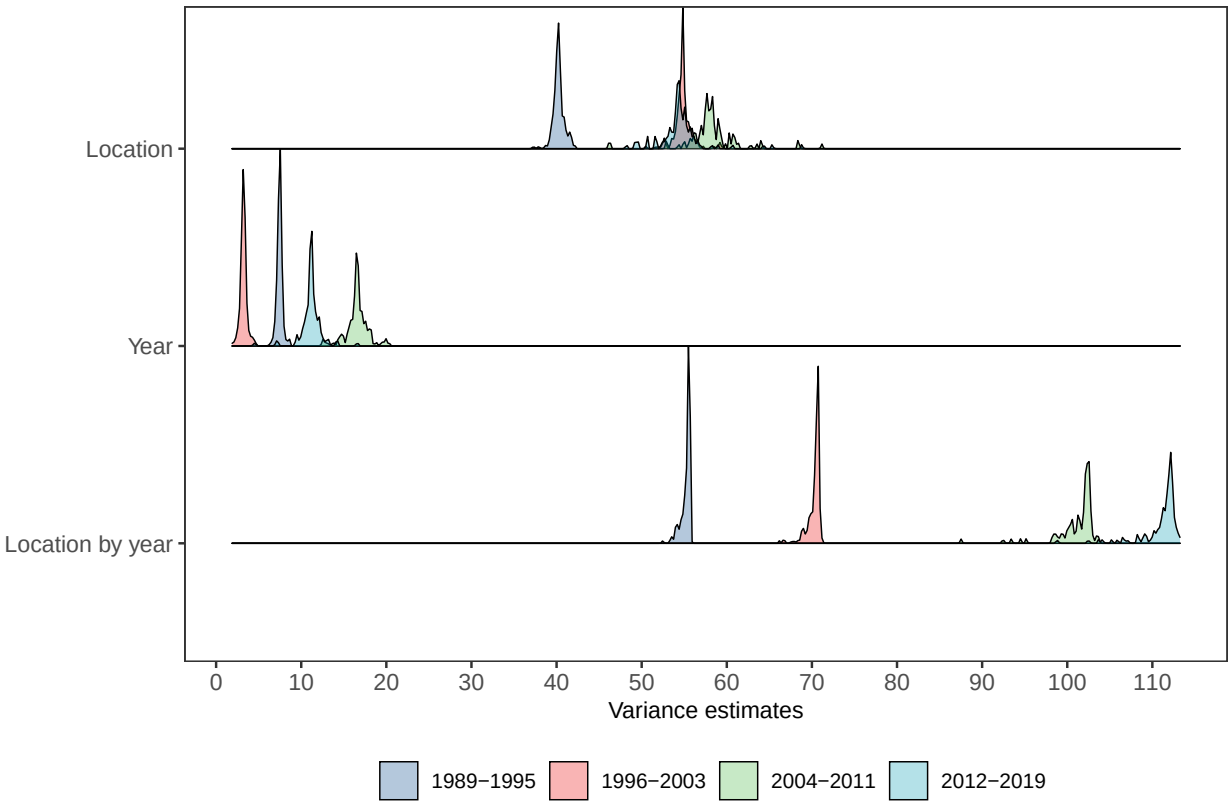

Figure A1: Jackknife estimates of location, year, and location by year variances for the groups of years 1989-1995, 1996-2003, 2004-2011, and 2012-2019.

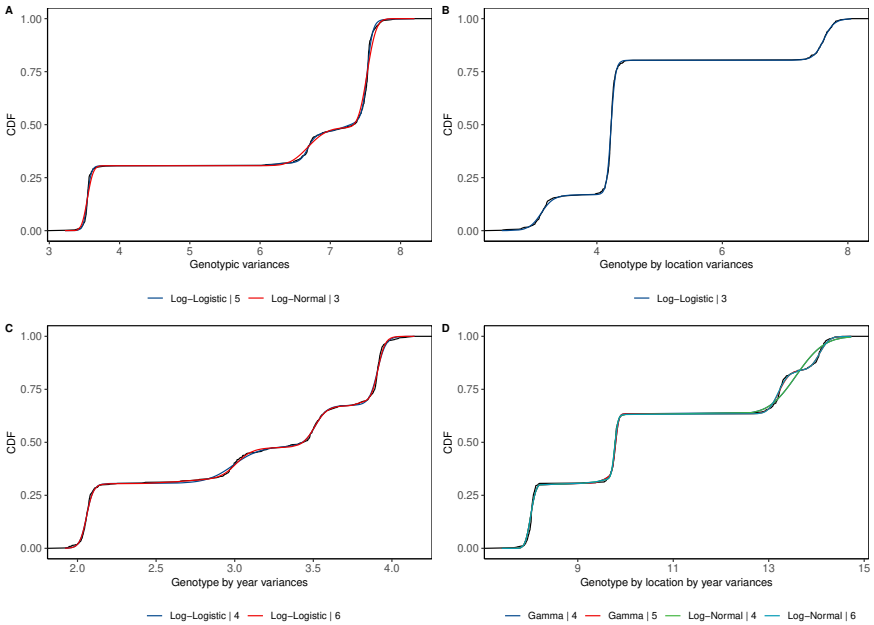

Figure A2: Cumulative distribution function (CDF) for the best-fit models according to the genotypic (A), genotype by location (B), genotype by year (C), and genotype by location by year (D) variances from jackknife. In each plot, the legends states for name of the distribution followed by its mixture number.

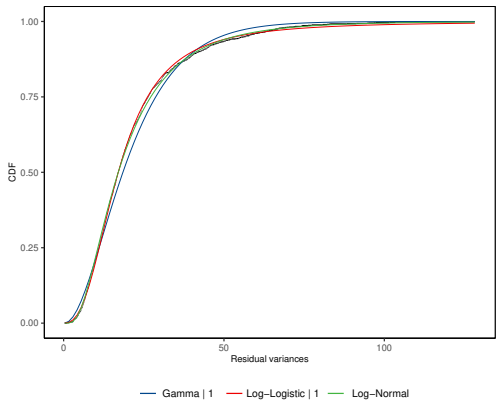

Figure A3: Cumulative distribution function (CDF) for the best-fit models according to the residual variances from individual trial level. The legends states for the name of the distribution followed by its mixture number.

Table A3: Number of genotypes (NG) and locations (NL) within each year, the number of common genotypes between years (upper diagonal), and the number of common locations between years (lower diagonal).

Years	89	90	91	92	93	94	95	96	97	98	99	00	01	02	03	04	05	06	07	08	09	10	11	12	13	14	15	16	17	18	19	NG
89		7	18	9	5	4	2																									192
90	12		28	14	5	4	1																									144
91	22	13		45	13	5	4																									180
92	20	12	21		48	20	8	2	1	1	1																					194
93	16	11	21	24		40	15	2	1	1	1																					179
94	14	11	18	20	25		45	7	1	1	1																					178
95	15	10	18	21	24	21		42	8	4	1																					193
96	14	9	16	20	21	18	21		32	10	4	2	2	2	2	1	1	1	1	1												187
97	16	10	19	21	24	21	21	22		45	17	5	3	2	2	1	1	1	1	1												184
98	17	12	19	22	23	20	20	20	25		43	14	5	3	3	2	1	1	1	1												181
99	15	10	17	17	19	18	18	17	21	25		37	9	5	3	3	2	1	1	1												183
00	11	7	13	15	15	14	12	14	16	17	21		32	10	7	2	1	1	1	1												173
01	7	6	9	10	11	11	9	10	12	14	17	18		44	12	3	1	1	1	1												151
02	11	7	13	15	15	13	13	14	16	17	19	20	18		24	4	2	1	1	1												180
03	5	3	7	6	8	6	7	7	8	8	11	12	11	11		19	4	2	1	1												126
04	6	3	7	8	9	8	8	9	10	9	11	12	12	14	9		46	17	6	3	1	1	1	1	1	1	1	1				171
05	4	1	4	6	5	5	5	5	6	6	8	9	7	10	5	9		42	18	9	5	2	2	3	3	2	3	1				164
06	6	2	6	7	7	6	6	7	8	8	9	9	8	11	6	11	8		50	16	6	2	3	4	4	3	4	2	1	1	1	185
07	4	2	4	5	6	4	4	5	6	6	7	7	7	9	6	10	7	9		49	15	2	3	4	4	3	4	2	1	1	1	229
08	6	2	6	8	7	5	6	7	7	7	8	9	7	11	6	9	8	9	8		41	10	6	4	4	3	4	2	1	1		217
09	8	4	8	10	9	7	8	9	9	9	11	11	8	12	6	10	8	10	8	11		36	13	8	6	4	5	3	1	1		231
10	5	3	5	6	5	4	5	5	5	5	8	8	4	8	4	6	7	6	6	8	11		21	7	6	6	4	3	2	2	1	137
11	4	3	4	6	5	5	4	5	5	5	7	8	5	8	2	6	6	7	5	7	11	9		27	13	6	5	4	3	3	2	139
12	4	2	4	6	5	4	4	5	5	6	8	8	6	8	3	7	6	8	6	7	10	7	11		43	10	8	8	5	5	4	179
13	7	3	6	8	7	5	6	7	7	7	7	7	4	8	3	7	6	8	6	7	11	9	11	13		34	16	8	8	6	4	161
14	6	3	6	6	5	4	5	5	5	6	7	7	3	7	3	5	5	5	4	5	8	7	6	8	9		41	17	6	5	3	140
15	4	3	4	5	4	4	4	4	4	4	5	6	2	5	2	4	4	4	3	4	7	7	6	8	9	9		48	19	10	7	154
16	5	3	5	6	4	4	4	4	4	5	6	6	3	6	2	4	4	5	2	4	6	4	5	7	7	9	7		56	26	13	169
17	9	6	8	11	8	7	8	8	8	11	12	10	8	11	6	5	4	5	3	6	9	7	6	8	8	8	9	8		60	28	175
18	6	4	6	7	7	5	6	6	6	6	8	8	6	8	7	5	3	3	3	5	7	6	4	5	7	5	7	5	14		63	183
19	5	3	3	6	3	3	3	3	3	4	4	5	3	5	2	2	4	2	2	3	4	5	3	4	6	5	7	4	11	13		233
NL	24	15	27	30	32	27	26	23	27	31	30	25	21	23	14	15	12	12	10	11	15	12	13	16	17	12	11	11	20	16	13	

Table A4: Goodness-of-fit (GOF) statistics and selection criteria for the fit of univariate and multivariate probability distributions for genotypic, genotype by location, genotype by year, and genotype by location by year variance components estimated from the jackknife analysis, and residuals variances from trial-level. The best-fit model is highlighted in bold. KS, CM, AD, AIC, and BIC stand for Kolmogorov-Smirnov, Cramer-von Mises, Anderson-Darling, Akaike's and Bayesian Information Criterion, respectively.

Variance	Distribution	Mixture of	GOF statistics			GOF criteria		
			KS	CM	AD	AIC	BIC	Log-likelihood
Genotypic	Gamma	2	0.19	4.39	25.23	672.09	694.00	-331.05
	Log-Logistic	2	0.15	3.32	19.97	598.57	620.48	-294.29
	Log-Normal	2	0.19	4.48	25.66	680.36	702.27	-335.18
	Burr	2	0.50	40.79	193.24	3978.69	4000.60	-1984.35
	F	2	0.67	77.08	356.80	4338.62	4360.52	-2164.31
	Gamma	3	0.07	0.81	5.56	182.11	217.16	-83.06
	Log-Logistic	3	0.15	3.26	19.45	554.84	589.90	-269.42
	Log-Normal	3	0.08	0.66	4.90	170.74	205.79	-77.37
	Burr	3	0.52	44.39	207.91	3978.03	4013.09	-1981.02
	F	3	0.67	77.08	356.80	4338.62	4360.52	-2164.31
	Gamma	4	0.09	1.03	6.20	177.38	225.58	-77.69
	Log-Logistic	4	0.15	3.26	19.45	565.62	613.82	-271.81
	Log-Normal	4	0.08	0.52	3.61	74.67	122.87	-26.33
	Burr	4	0.52	44.39	207.91	3984.04	4032.24	-1981.02
	F	4	0.67	75.30	347.50	4351.57	4399.77	-2164.79
	Gamma	5	0.10	0.74	5.85	155.12	216.47	-63.56
	Log-Logistic	**5**	0.05	0.21	1.67	-9.63	51.71	18.82
	Log-Normal	5	0.06	0.36	2.53	89.40	150.75	-30.70
	Burr	5	0.53	45.19	211.25	3989.54	4050.89	-1980.77
	F	5	0.67	75.93	350.80	4357.11	4418.45	-2164.55
	Gamma	6	0.07	0.46	3.15	94.30	168.79	-30.15
	Log-Logistic	6	0.05	0.20	1.89	32.97	107.47	0.51
	Log-Normal	6	0.06	0.37	2.68	103.17	177.66	-34.59
	Burr	6	0.52	43.68	205.00	3996.91	4071.40	-1981.45
	F	6	0.66	74.44	343.11	4364.38	4438.87	-2165.19
Genotype by Location	Gamma	2	0.30	10.51	48.78	1164.59	1186.50	-577.29
	Log-Logistic	2	0.24	8.35	49.77	1139.99	1161.90	-565.00
	Log-Normal	2	0.30	10.80	50.82	1194.97	1216.87	-592.48
	Burr	2	0.50	42.63	197.90	3466.48	3488.39	-1728.24
	F	2	0.63	67.82	305.85	3830.34	3852.24	-1910.17
	Gamma	3	0.30	10.52	48.87	1185.88	1220.94	-584.94
	Log-Logistic	**3**	0.04	0.14	0.72	-80.42	-45.36	48.21
	Log-Normal	3	0.06	0.49	2.38	-5.10	29.96	10.55
	Burr	3	0.50	42.78	198.50	3477.58	3512.63	-1730.79
	F	3	0.63	67.82	305.85	3830.34	3852.24	-1910.17
	Gamma	4	0.30	10.51	48.83	1185.69	1233.89	-581.85
	Log-Logistic	4	0.04	0.13	0.62	-80.72	-32.52	51.36
	Log-Normal	4	0.06	0.48	2.20	-10.68	37.52	16.34
	Burr	4	0.50	41.96	195.32	3482.35	3530.55	-1730.18
	F	4	0.65	73.19	331.38	3854.78	3902.98	-1916.39
	Gamma	5	0.02	0.07	0.56	-34.66	26.69	31.33
	Log-Logistic	5	0.03	0.10	0.59	-73.99	-12.64	50.99
	Log-Normal	5	0.02	0.06	0.49	-66.28	-4.93	47.14

Table A4 continued from previous page

Variance	Distribution	Mixture of	GOF statistics			GOF criteria		
			KS	CM	AD	AIC	BIC	Log-likelihood
	Burr	5	0.49	41.22	192.48	3488.02	3549.36	-1730.01
	F	5	0.65	72.56	328.33	3860.42	3921.76	-1916.21
	Gamma	6	0.02	0.06	0.40	-30.91	43.58	32.46
	Log-Logistic	6	0.03	0.09	0.46	-74.96	-0.47	54.48
	Log-Normal	6	0.03	0.04	0.25	-71.57	2.92	52.78
	Burr	6	0.49	41.61	194.00	3494.23	3568.72	-1730.11
	F	6	0.65	72.49	328.02	3866.55	3941.05	-1916.28
	Gamma	2	0.13	2.33	17.77	805.22	827.13	-397.61
	Log-Logistic	2	0.12	1.39	10.73	522.98	544.89	-256.49
	Log-Normal	2	0.13	1.80	12.89	546.28	568.19	-268.14
	Burr	2	0.46	34.26	165.40	2641.18	2663.09	-1315.59
	F	2	0.67	80.39	375.82	3042.72	3064.63	-1516.36
	Gamma	3	0.12	1.48	10.61	438.20	473.26	-211.10
	Log-Logistic	3	0.08	0.29	1.48	69.69	104.75	-26.85
	Log-Normal	3	0.13	1.36	10.01	402.06	437.11	-193.03
	Burr	3	0.49	38.99	184.26	2636.35	2671.40	-1310.17
	F	3	0.67	80.39	375.82	3042.72	3064.63	-1516.36
	Gamma	4	0.12	1.34	9.44	391.85	440.05	-184.92
Genotype	Log-Logistic	4	0.04	0.11	1.14	-21.98	26.22	21.99
by	Log-Normal	4	0.05	0.26	2.48	69.53	117.73	-23.77
Year	Burr	4	0.48	36.59	174.58	2642.16	2690.36	-1310.08
	F	4	0.65	73.44	338.37	3062.06	3110.26	-1520.03
	Gamma	5	0.07	0.45	4.18	142.71	204.05	-57.35
	Log-Logistic	5	0.03	0.09	1.07	-25.40	35.94	26.70
	Log-Normal	5	0.05	0.27	2.52	73.76	135.10	-22.88
	Burr	5	0.46	33.87	163.84	2652.24	2713.58	-1312.12
	F	5	0.65	73.45	338.42	3068.16	3129.51	-1520.08
	Gamma	6	0.05	0.29	2.90	78.16	152.65	-22.08
	Log-Logistic	**6**	0.03	0.08	1.04	-35.86	38.63	34.93
	Log-Normal	6	0.04	0.13	1.11	2.00	76.49	16.00
	Burr	6	0.45	32.15	157.07	2645.95	2720.44	-1305.98
	F	6	0.65	73.41	338.24	3073.76	3148.25	-1519.88
	Gamma	2	0.18	3.72	23.32	2027.94	2049.84	-1008.97
	Log-Logistic	2	0.17	3.40	21.40	2104.98	2126.88	-1047.49
	Log-Normal	2	0.18	3.73	23.33	2028.46	2050.36	-1009.23
	Burr	2	0.57	46.63	216.45	4962.90	4984.81	-2476.45
	F	2	0.68	68.15	308.55	5298.60	5320.51	-2644.30
	Gamma	3	0.06	0.52	3.94	905.88	940.93	-444.94
	Log-Logistic	3	0.17	3.22	19.52	1938.73	1973.78	-961.36
	Log-Normal	3	0.06	0.52	3.96	909.36	944.42	-446.68
	Burr	3	0.60	51.37	235.46	4969.12	5004.18	-2476.56
Genotype	F	3	0.68	68.15	308.57	5304.63	5339.68	-2644.31
by	Gamma	4	0.06	0.41	3.50	830.66	878.86	-404.33
Location	Log-Normal	4	0.06	0.36	3.30	810.14	858.34	-394.07
by	Burr	4	0.60	51.37	235.46	4975.11	5023.31	-2476.55
Year	F	4	0.69	71.27	323.14	5313.82	5362.02	-2645.91
	Gamma	**5**	0.05	0.21	1.74	709.98	771.33	-340.99
	Log-Normal	5	0.06	0.22	1.15	715.12	776.47	-343.56
	Burr	5	0.60	51.37	235.46	4981.10	5042.45	-2476.55

Table A4 continued from previous page

Variance	Distribution	Mixture of	GOF statistics			GOF criteria		
			KS	CM	AD	AIC	BIC	Log-likelihood
	F	5	0.70	75.75	344.66	5317.16	5378.51	-2644.58
	Gamma	6	0.06	0.32	2.58	781.22	855.71	-373.61
	Log-Normal	6	0.05	0.16	1.53	691.83	766.32	-328.91
	Burr	6	0.58	48.52	223.96	4989.49	5063.98	-2477.75
	F	6	0.68	68.92	312.13	5322.81	5397.30	-2644.40
Residual (Trial-level)	Gamma	1	0.06	1.60	9.57	11139.50	11150.02	-5567.80
	Log-Logistic	**1**	0.02	0.07	0.72	11063.82	11074.34	-5529.90
	Log-Normal	1	0.02	0.09	0.67	11066.60	11077.12	-5531.10
	Burr	1	0.41	79.00	377.80	13896.60	13907.11	-6946.30
	F	1	0.54	131.37	594.40	14677.45	14687.97	-7336.73

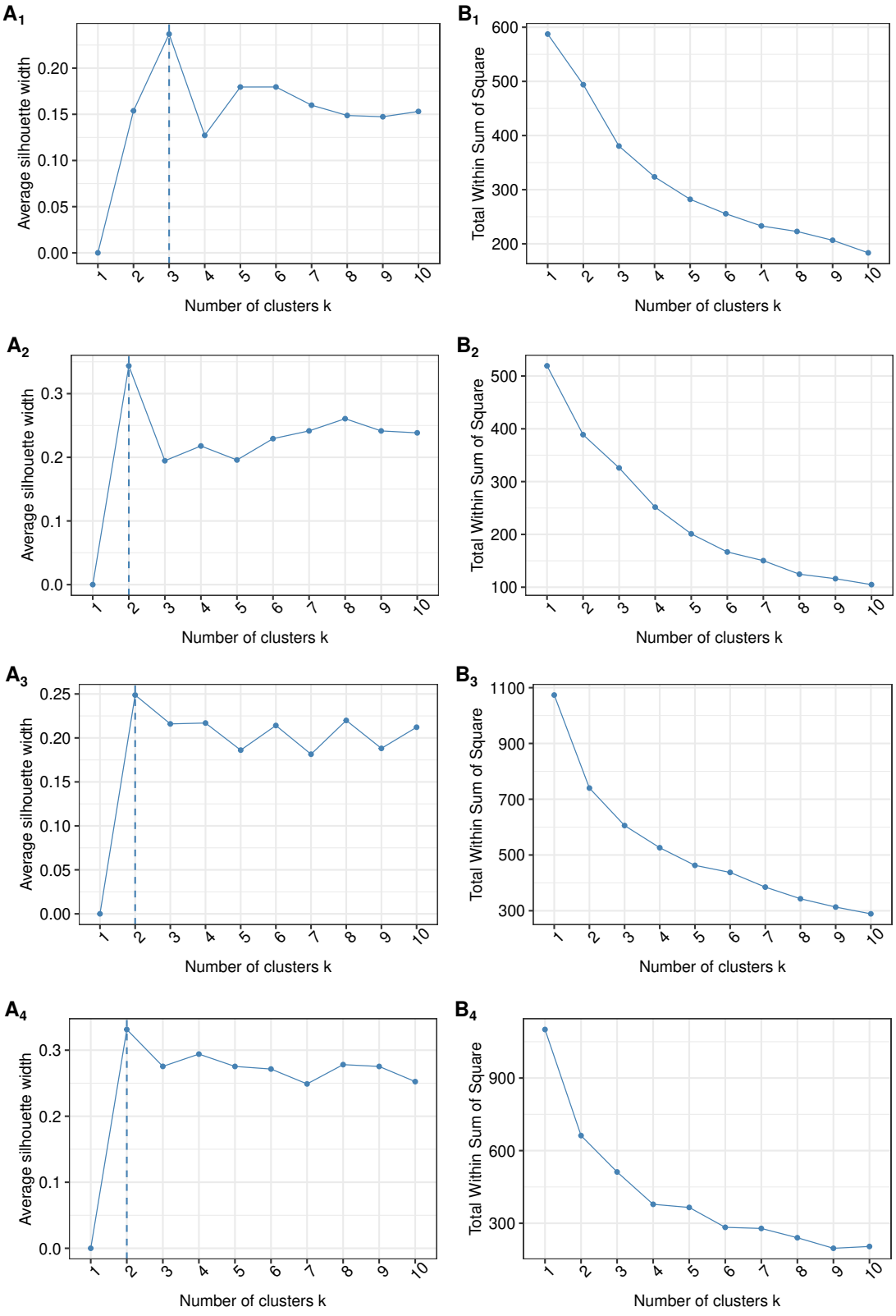

Figure A4: Graphical display of the optimal number of clusters based on the Silhouette (A) and Elbow (B) methods for the phenotypic clustering type (PHE, A_1 and B_1), soil + elevation variables (SoilE, A_2 and B_2), weather within year variables (WW, A_3 and B_3), and weather across year variables (WA, A_4 and B_4).

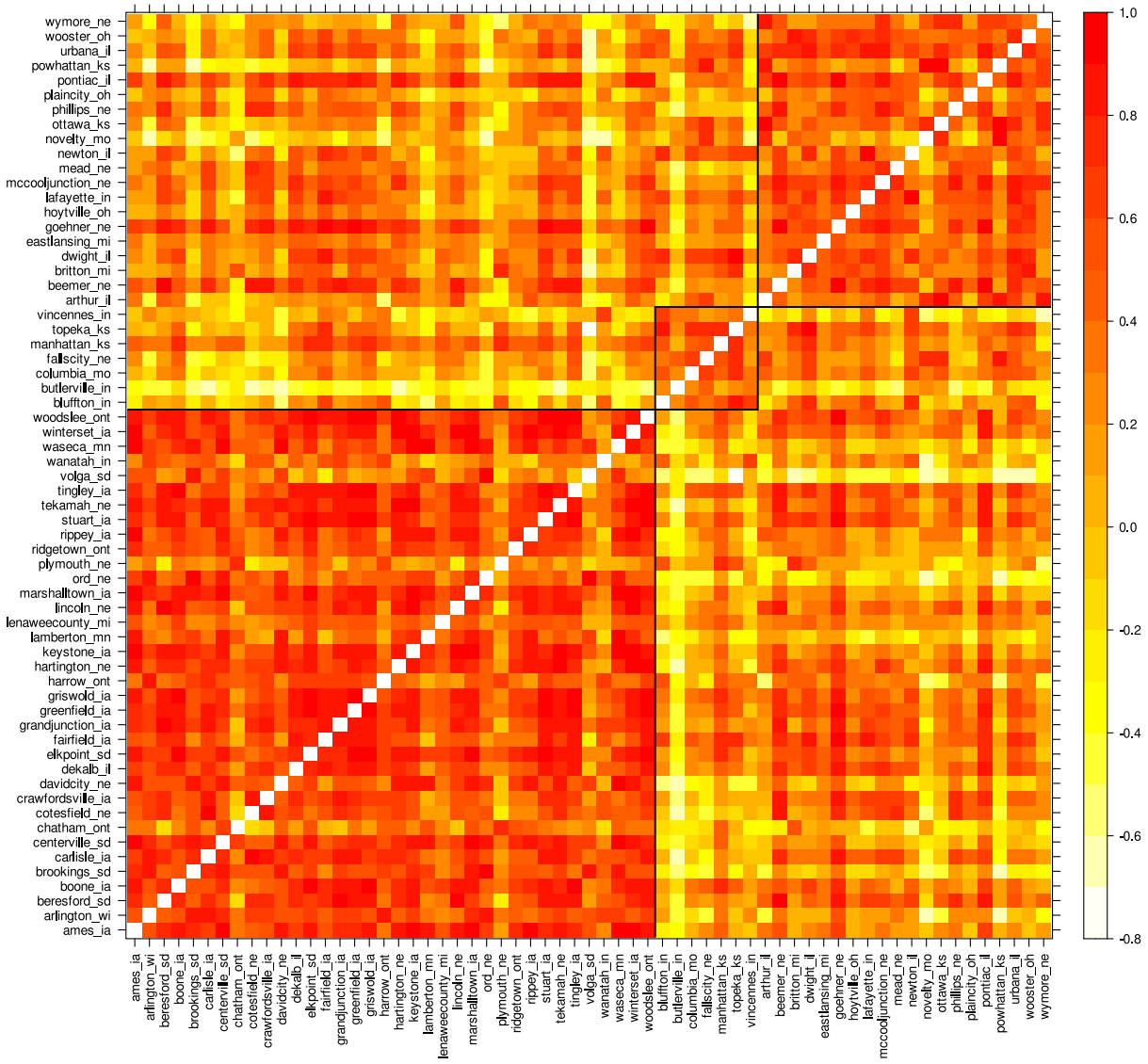

Figure A5: Heatmap of genetic correlations between the 63 observed locations from 1989 to 2019, estimated from the factor analytic (FA) model M3-18. Fitted k-means clustering are ordered, from 1 to 3.

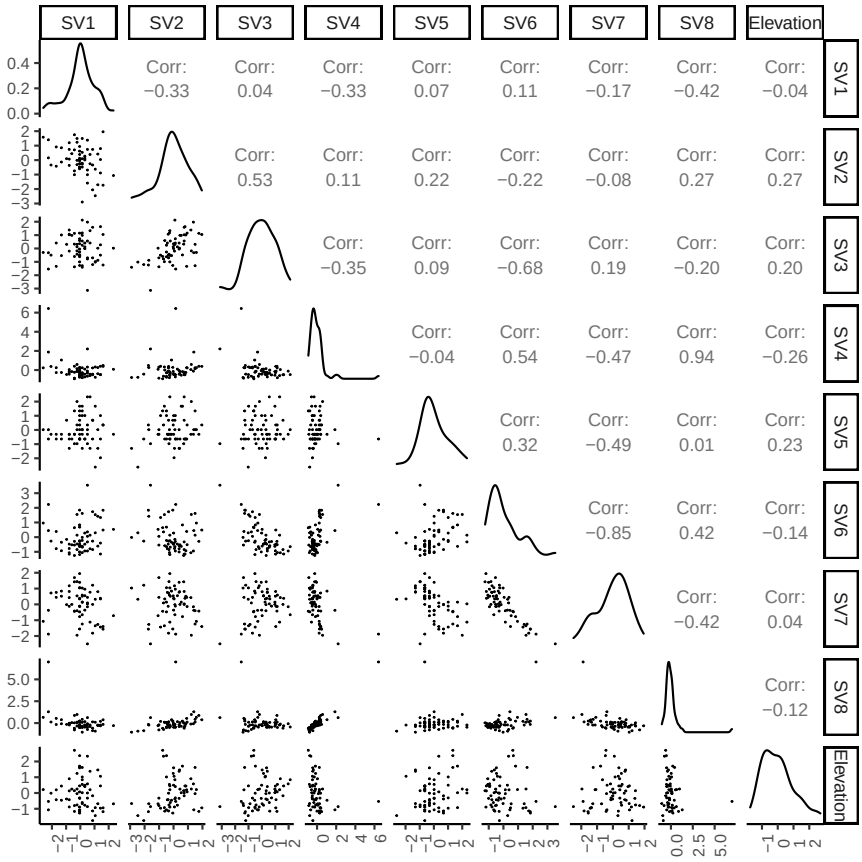

Figure A6: Scatterplots of the scaled and centered soil variables and elevation. Pearson correlation is displayed on the right. Variable distribution is available on the diagonal. The labels SV1, SV2, to SV8 are the soil variables described in section 5.3.5.

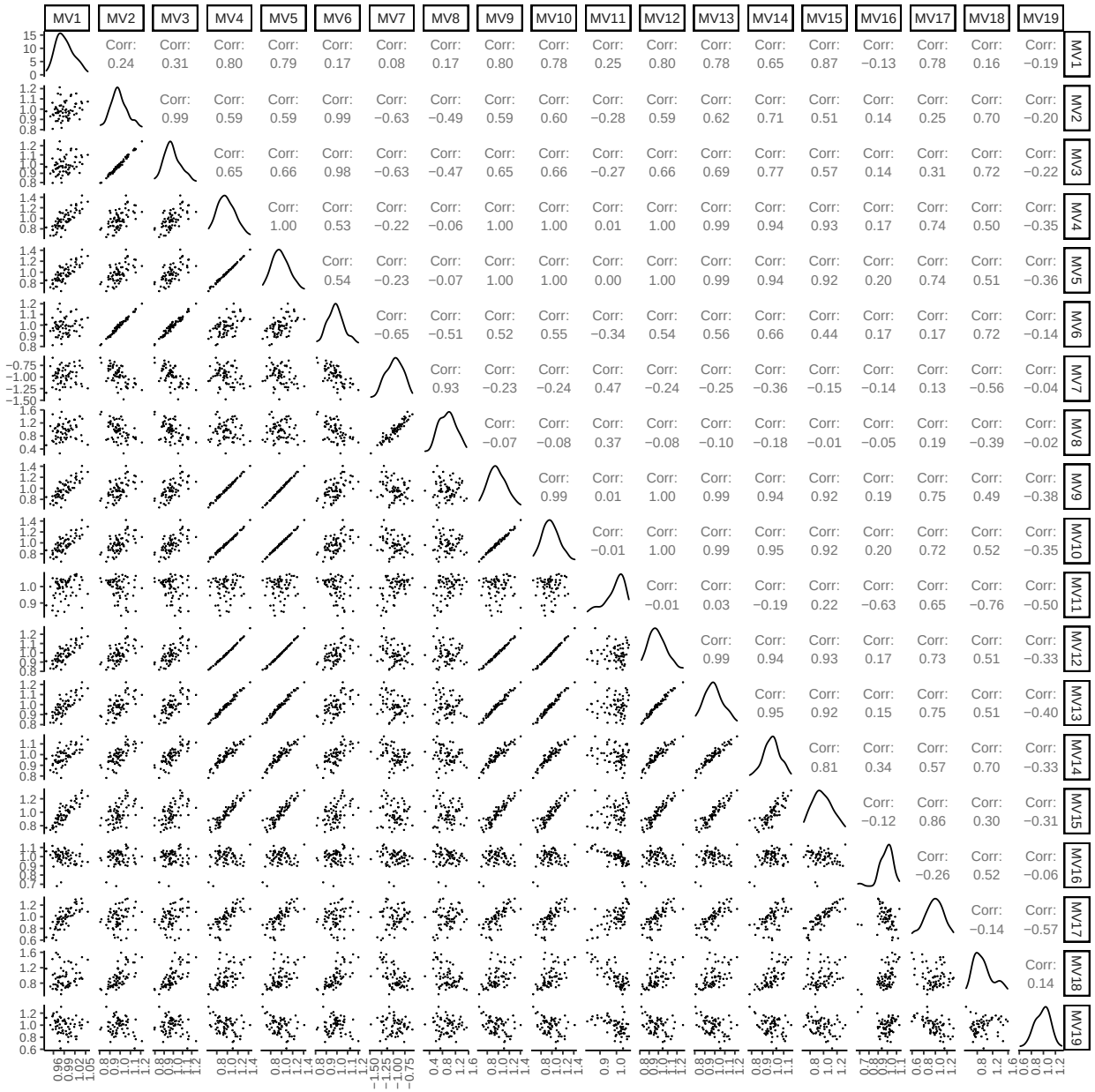

Figure A7: Scatterplots of the scaled and centered weather variables computed across years. Pearson correlation is displayed on the right. Variable distribution is available on the diagonal.The labels MV1, MV2, ..., to MV19, are the weather variables described in Table A1.

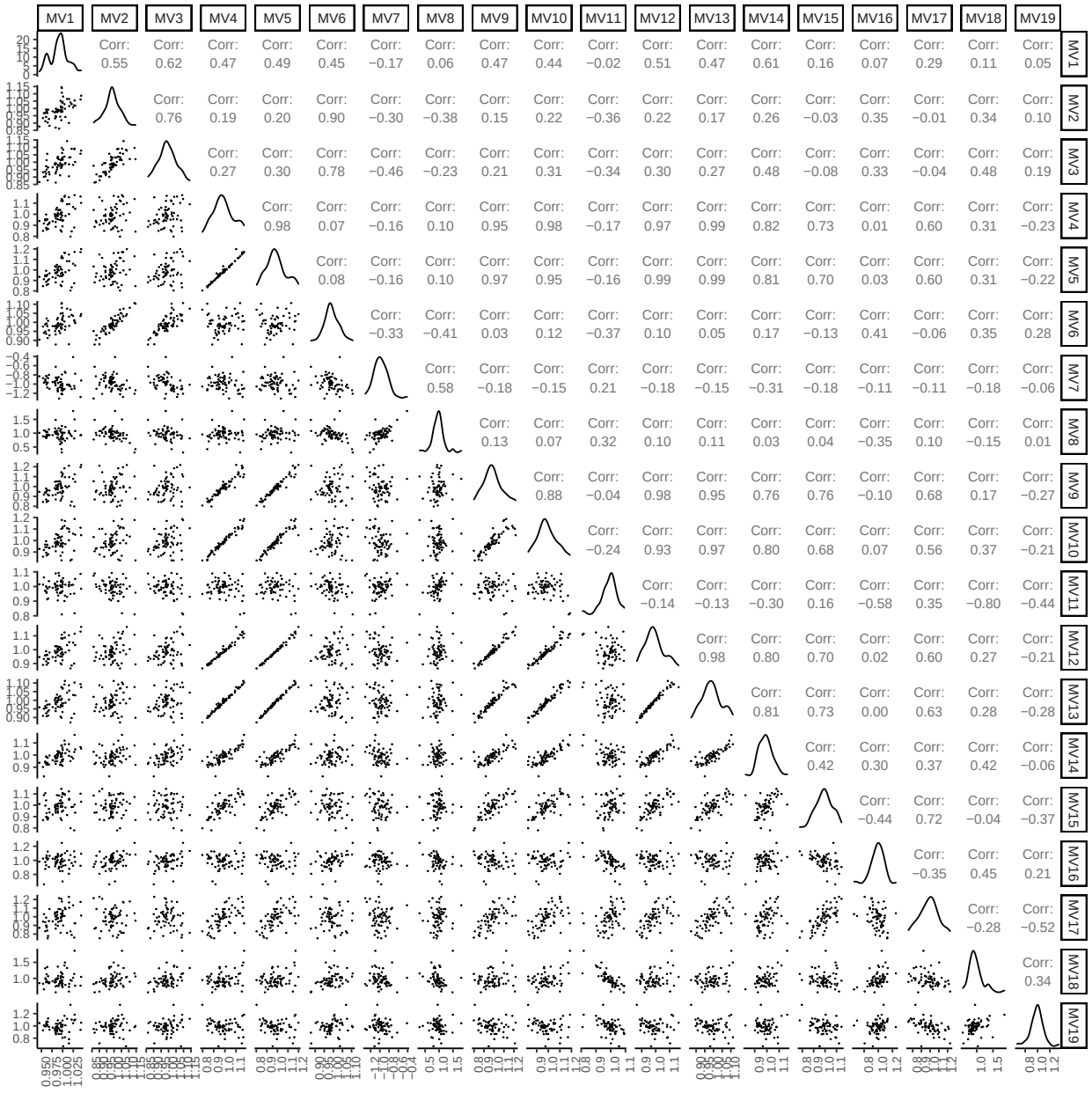

Figure A8: Scatterplots of the scaled and centered weather variables computed within years. Pearson correlation is displayed on the right. Variable distribution is available on the diagonal. The labels MV1, MV2, ..., to MV19, are the weather variables described in Table A2.

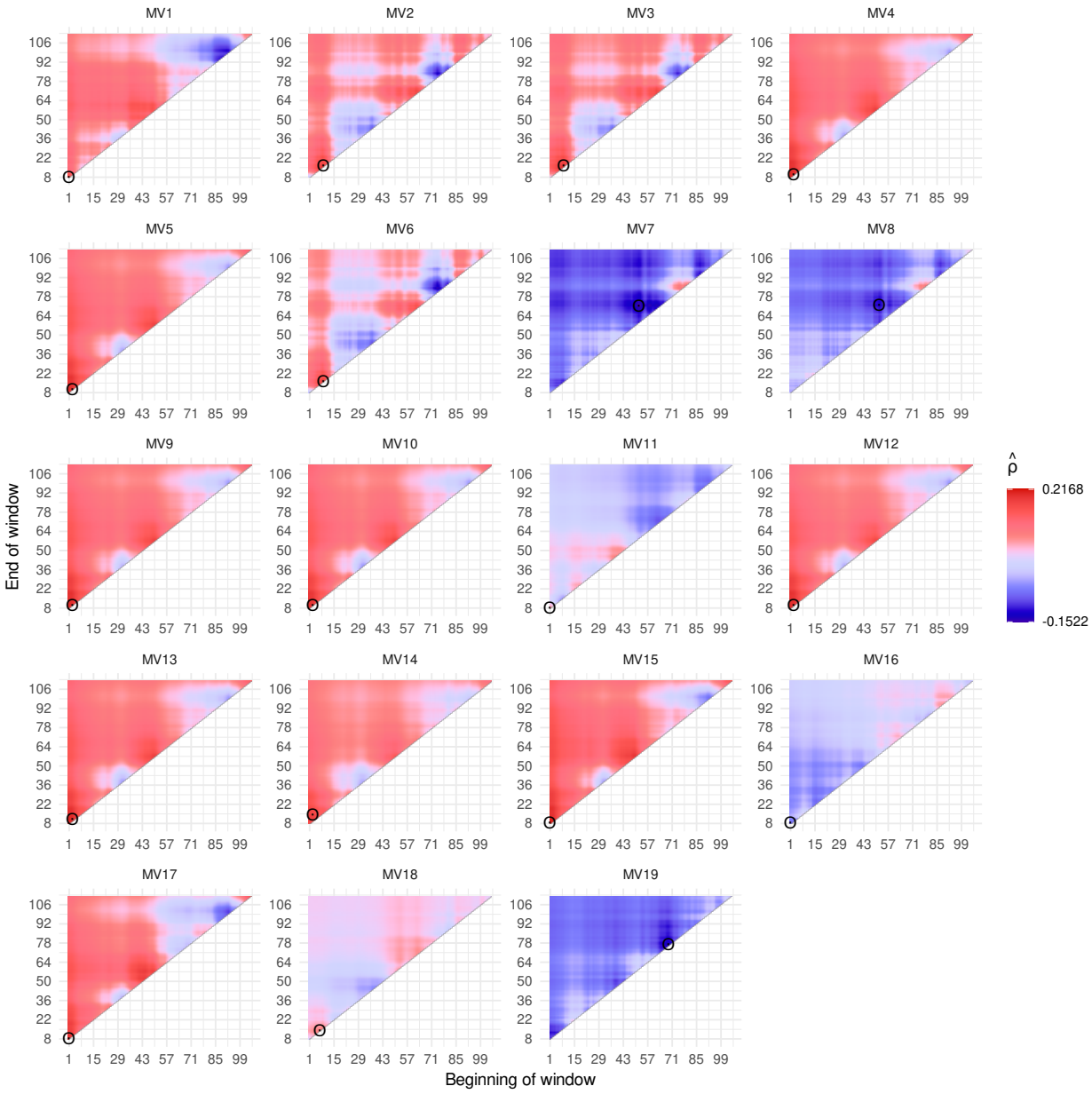

Figure A9: Exhaustive search from the critical environmental window computed from the Pearson correlation between the weather variables across years and the genotype by location deviations of each environment (location-year combination). The dots depict the highest window correlation. The labels MV1, MV2, to MV19 are the weather variables described in Table A1.

Table A5: Jaccard similarity matrix between clustering-types (phenotypic - PHE, soil and elevation - SoilE, latitude split into 2 clusters - Lat2, latitude split into 3 clusters - Lat3, weather across years - WA, and weather within years - WW).

Clusters		PHE			SoilE		Lat2		Lat3			WA		WW	
		1	2	3	1	2	1	2	1	2	3	1	2	1	2
PHE	1	1	-	-	0.25	0.44	0.18	0.64	0.02	0.53	0.27	0.20	0.54	0.12	0.60
	2		1	-	0.00	0.14	0.20	0.00	0.35	0.02	0.00	0.23	0.02	0.18	0.06
	3			1	0.13	0.30	0.45	0.07	0.33	0.22	0.03	0.25	0.23	0.30	0.21
SoilE	1				1	-	0.07	0.35	0.03	0.16	0.32	0.00	0.37	0.00	0.32
	2					1	0.62	0.28	0.30	0.52	0.09	0.51	0.38	0.39	0.48
Lat2	1						1	-	0.46	0.37	0.00	0.58	0.22	0.42	0.32
	2							1	0.00	0.36	0.39	0.06	0.61	0.07	0.53
Lat3	1								1	-	-	0.41	0.08	0.35	0.13
	2									1	-	0.22	0.51	0.20	0.51
	3										1	0.06	0.23	0.03	0.22
WA	1											1	-	0.42	0.21
	2												1	0.12	0.64