Supplementary information

Materials and Methods Table S1-S7, Figure legends S1-S7

Materials and Methods

 Table S1: Introduction of point mutations in EGFPMBP-BRCA2, 2xMBP-BRCA2₁₋₂₅₀

Mutation	Oligo name	Sequence (5'-3')
S193A	Fw : oAC543	CCC ACC CTT AGT TCT GCT GTG CTC ATA GTC
	Rv : oAC544	GAC TAT GAG CAC AGC AGA ACT AAG GGT GGG
M192T	Fw : oAC283	GTGGATCCTGATACGTCTTGGTCAAGTTC
	Rv : oAC284	GA ACT TGA CCA AGA CGT ATC AGG ATC CAC
S196N	Fw : oAC026	CCTGATATGTCTTGGTCAAATTCTTTAGCTACACCACC
	Rv : oAC027	GGTGGTGTAGCTAAAGAATTTGACCAAGACATATCAGG
S206C	Fw : oAC028	CCACCCACCCTTAGTTGTACTGTGCTCATAGTCAG
	Rv : oAC029	CTGACTATGAGCACAGTACAACTAAGGGTGGGTGG
T200K	Fw : oAC285	CAA GTT CTT TAG CTA AAC CAC CCA CCC TTA G
	Rv : oAC286	CTA AGG GTG GGT GGT TTA GCT AAA GAA CTT G
T207A	Fw : oAC545	GGA TCC TGA TAT GGC TTG GTC AAG TTC TTT AGC
	Rv : oAC546	GCT AAA GAA CTT GAC CAA GCC ATA TCA GGA TCC

and GST-BRCA2₁₉₀₋₂₈₃ constructs

Table S2: Sequencing primers

Construct	Oligo name	Binding	Sequence (5'-3')
		site	
GFPMBP-	Rv : oAC131	aa 273	TTAGTTCGACTTATCCAATGTGGTCTTT
BRCA2, GST-		BRCA2	
BRCA2 ₁₉₀₋₂₈₃			
2xMBP-	Fw : oAC149	aa 1-6	TTATTTGCTAGCCCTATTGGATCCAAAGAG
BRCA2 ₁₋₂₅₀		BRCA2	

Construct	Oligo name	Sequence (5'-3')
Amplifying	Fw: oAC130	TTAGGATCCATGTCTTGGTCAAGTTCT
BRCA2 ₁₉₂₋₂₈₃	Rv: oAC131	TTAGTTCGACTTATCCAATGTGGTCTTT
L191D in NTshort	Fw: oAC593	CAG GGG CCC GAT ATG TCT TGG TCA AGT TCT
	Rv: oAC594	AGA ACT TGA CCA AGA CAT ATC GGG CCC CTG

 Table S3: Primers used to subclone BRCA2₁₉₂₋₂₈₃ into the pGEX-6P-1 vector

Table S4: Primers used to subclone PLK1 cDNA into pFastBac HT

Primer name	Sequence (5'-3')
GA_pFBtev_R	GCCCTGAAAATACAGGTTTTCGGTCGTTGGGAT
GA_pFB_UTR_F	TTGTCGAGAAGTACTAGAGGATCATAATCA
GA_hPLK_F	ATCCCAACGACCGAAAACCTGTATTTTCAGGGCATGAGTGCTGCAG
	TGACTGCA
GA_hPLK_R	TGATTATGATCCTCTAGTACTTCTCGACAATTAGGAGGCCTTGAGAC
	GGTT

Table S5: Primers used to subclone PLK1_{PBD} (aa 326-603) into pT7-His6-SUMO

Primer name	Sequence (5'-3')
GA_PLKPDBwt_F	ATTGAGGCTCACCGCGAACAGATTGGTGGCTCGATTGCTCCCA
	GCAGCCT
GA_PLKPDBwt_R	TTCCTTTCGGGCTTTGTTAGCAGCCGGTCATTAGGAGGCCTTGA
	GACGGT

Table S6: Peptide sequences for Isothermal Titration Calorimetry (ITC)

Peptide	Sequence
pS197	DMSWSS{pS}LAT
T207	WSSSLATPPTLSSTVLI
pT207	WSSSLATPPTLSS{pT}VLI
T207A	WSSSLATPPTLSSAVLI
СрТ207	WSSSLATPPTLSC{pT}VLI

Table S7: Primers for amplifying BRCA2 (aa 1-267) from genomic DNA

Primer name	Sequence (5'-3')
Fw : OAC035	GGTCGTCAGACTGTCGATGAAGCC
Rv : OAC056	CAAAGAGAAGCTGCAAGTCATGGATTTGAAAAAACATCAGGG

Figure Legends

Figure S1. Related to Figure 1. **PLK1 phosphorylation kinetics of BRCA2**₁₉₀₋₂₈₃ **and conservation of PLK1 phosphosites**

(a) Phosphorylation kinetics resulting from a duplicate experiment performed with a different PLK1 kinase aliquot on ¹⁵N labelled BRCA2₁₉₀₋₂₈₃ (283K, 600 MHz). Error bars correspond to SD (n=2). (b) Alignment of the region 190-283 of BRCA2 in 30 different species. Amino acids conserved in more than 66 % of the species are highlighted with coloured background. The percentage of identity to human BRCA2 sequence is added after the species name. Arrows show the amino acids identified as phosphorylated by PLK1 in the NMR experiments. (c) Bar graph showing the evolutionary conservation of the amino acids phosphorylated by PLK1 detected by NMR.

Figure S2. Related to Figure 2. PLK1 phosphorylation kinetics of WT vs mutated forms of BRCA2₁₉₀₋₂₈₃

Phosphorylation kinetics resulting from a duplicate experiment performed with a different PLK1 kinase aliquot on ¹⁵N labelled BRCA2₁₉₀₋₂₈₃ (283K, 600 MHz). Comparisons of the WT kinetics with the phosphorylation kinetics of **(a)** T207A and **(b)** T200K are displayed.

Figure S3. Related to Figure 4. **Isothermal Titration Calorimetry (ITC)** thermogram showing binding of PLK1_{PBD} to the fragment BRCA2₁₉₀₋₂₈₃ or a 10 aa BRCA2 peptide containing pS197

Thermogram showing the binding affinity of $PLK1_{PBD}$ to (a) phosphorylated or the (b) non-phosphorylated BRCA2₁₉₀₋₂₈₃ fragment. (c) Thermogram showing the binding affinity of $PLK1_{PBD}$ to a 10 aa BRCA2 peptide comprising pS197.

Figure S4. Related to Figures 4-8. **BRCA2 protein levels of BRCA2**^{-/-} **stable** clones expressing the variants as compared to the WT expressing cells utilized in this study and interaction of BRCA2 in these clones with PLK1.

(a) Protein levels of EGFP-MBP-BRCA2 WT or the VUS (S206C and T207A) in total protein extracts from respective BRCA2^{-/-} stable clones analyzed by western blot using anti-BRCA2 (OP95) and anti-GFP antibodies. (b) GFP-trap pull-down of BRCA2 from mitotic cell extracts of stable clones expressing WT BRCA2, or the variants S206C or T207A. The interaction with PLK1 was revealed by anti-PLK1 antibody and BRCA2 was detected with a BRCA2 specific antibody.

4

Figure S5. Effect of PLK1 inhibitors and phosphatase treatment on pT680-BUBR1.

(a) Protein levels of pT680-BUBR1 in BRCA2 WT stable clone after treatment with PLK1 inhibitors. After 14h culture with media containing nocodazole (100 ng/µl), PLK1 inhibitors (Bi2536 (50 nM) or BTO (50 µM)) were added to the media and the cells were cultured for additional 2h before harvesting and extraction of proteins. The levels of pT680-BUBR1 in total protein lysate was analyzed by western blot. (b) Phosphatase (Fast AP phosphatase) treatment of total protein lysate extracted from nocodazole (100 ng/µl) treated DLD1 BRCA2 WT stable clone, levels of pT680-BUBR1 was analyzed by western blot. For (a, b), stain-free images of the 4-15% SDS-PAGE gel before transfer were used as loading control (cropped images are shown).

Figure S6. Related to Figure 5 and 8. Cell cycle profile of the BRCA2^{-/-} stable clones in nocodazole versus untreated conditions and plate efficiency of unchallenged cells. PLK1 co-immunoprecipitates with BRCA2 in cells exposed or not to IR.

(a, b) Cell cycle distribution of DLD1 BRCA2^{-/-} stable clones expressing EGFP-MBP-BRCA2 WT or the VUS treated with nocodazole (b) or DMSO (c) as in Figure 5. Statistical significance of differences was calculated with two-way ANOVA test with Tukey's multiple comparisons test (the asterisks show differences compared to WT; ns p > 0.05, * $p \le 0.05$, ** $p \le 0.01$, *** $p \le 0.001$, **** $p \le 0.0001$). (c) Crystal violet stained colonies of unchallenged cells from Figure 8a. (d) Co-immunoprecipitation of endogenous PLK1 after exposure to IR (6 Gy) in nocodazole treated U2OS cells expressing exogenous 2xMBP-BRCA2₁₋₂₅₀ (WT). StainFree image of the 7.5% SDS- PAGE (7.5%) gel before transfer was used as loading control (cropped image is shown).

Figure S7. Purification of PLK1 and PLK1_{PBD} protein

(a) SDS-PAGE showing purified PLK1. Human PLK1 was expressed and purified from sf9 insect cells using Ni-NTA column followed by a second purification step with a cationic exchange Capto S column. The purified protein was loaded on a 4-20% SDS-PAGE (3 μ g) and the detected by Coomassie staining. (b) SDS-PAGE showing purified PLK1_{PBD}. 6His-Sumo-PLK1_{PBD} was expressed and purified from bacteria using a His-TRAP column, the His-tag was cleaved with 6xHis-SUMO Protease and the cleaved PLK1_{PBD} was further purified using Ni-NTA agarose resin. The purified protein was loaded on a 4-20% SDS-PAGE (1.4 μ g) and detected by Coomassie staining. (c) *In vitro* kinase assay with the purified PLK1 (0.1 μ g) from (a) or PLK1 purchased from Abcam, 0.1 μ g PLK1 was used in the kinase reaction with either RAD51 (25 ng) or purified 2xMBP-BRCA2₁₋₂₅₀ WT (0.5 μ g) as substrate in the presence of γ^{32} P-ATP. The samples were resolved by 7.5 % SDS-PAGE and ³²P-labeled products were detected by autoradiography.