

Assemblytics: a web analytics tool for the detection of assembly-based variants

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Table of Contents

Supplementary Note 1: Unique Anchor Filtering
Supplementary Figure 1: Schematic Diagram of Unique Anchor Filtering
Supplementary Note 2: Assemblytics Accuracy with Simulated Genomes
Supplementary Figure 2: Recall of Simulated Structural Variants
Supplementary Note 3: Assemblytics Web Interface
Supplementary Figure 3: Assemblytics Starting Page
Supplementary Figure 4: Assemblytics of MHAP <i>Homo sapiens</i> Assembly (brief)
Supplementary Note 4: Assemblytics Analysis of MHAP Genome Assemblies
Supplementary Figure 5: Assemblytics of MHAP <i>H. sapiens</i> Assembly (detailed)
Supplementary Figure 6: Assemblytics of MHAP <i>A. thaliana</i> Assembly
Supplementary Figure 7: Assemblytics of MHAP <i>D. melanogaster</i> Assembly
Supplementary Figure 8: Assemblytics of MHAP <i>S. cerevisiae</i> Assembly
Supplementary Figure 9: Assemblytics of MHAP <i>E. coli</i> Assembly

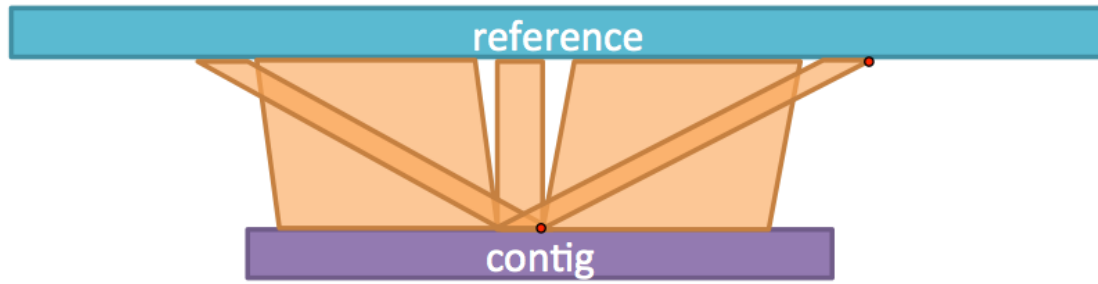
Supplementary Note 1: Unique Anchor Filtering

Assemblytics filters out repetitive sequences that have no unique anchoring alignments. Without such filtering, repetitive alignments could cause many false structural variations between each copy of the repeat. **Supplementary Figure 1 A-E** demonstrate a case where the repeat's "true alignment" could be chosen as the middle alignment based on the alignments of the neighboring sequences on the contig, but often there is no clear choice of consistent alignment (**Supplementary Figure 1 F and G**) and mutations could also be obscuring the mapping.

In an effort to conservatively call variants around repeats, Assemblytics does not attempt to choose the alignment for each repeat but rather filters out all alignments that do not contain at least a minimum amount (default: 10 kb) of uniquely anchoring sequence on the contig. The minimum anchoring sequence length can also be adjusted at runtime for different genome complexities and/or sequencing technologies. Note that this does not remove all sequences containing repeats but rather all sequences consist of only repeats without any unique sequence to anchor them.

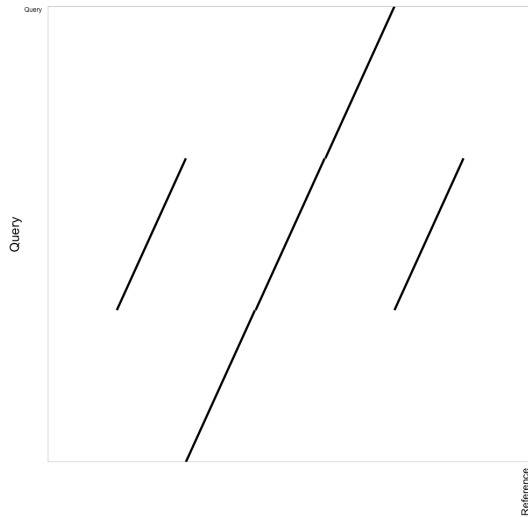
The sequences and delta files used in this figure are available at:
<http://qb.cshl.edu/assemblytics/reproducibility>

A.



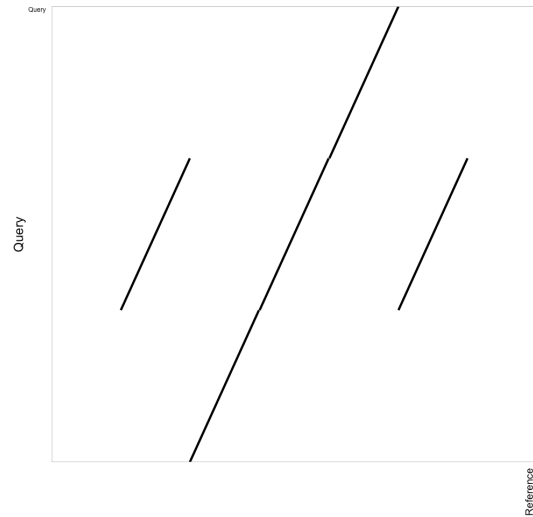
B.

All alignments



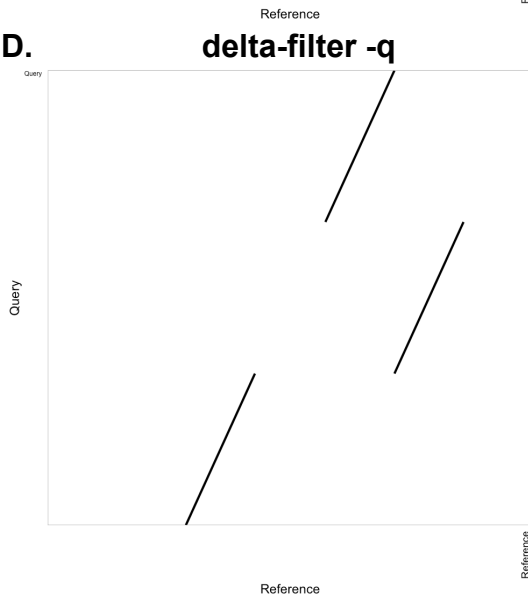
C.

delta-filter -r



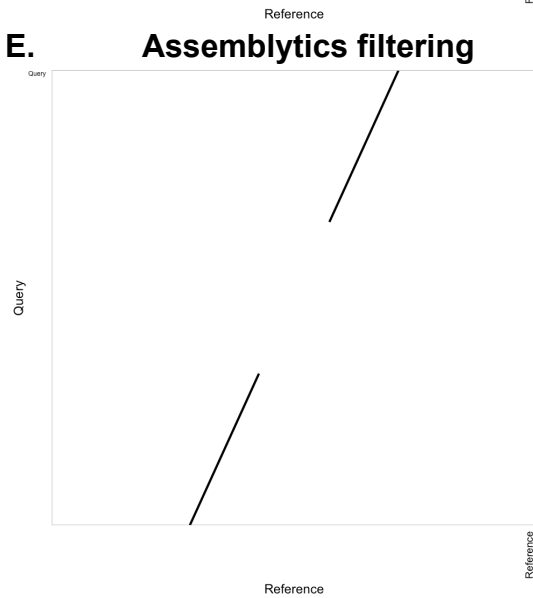
D.

delta-filter -q

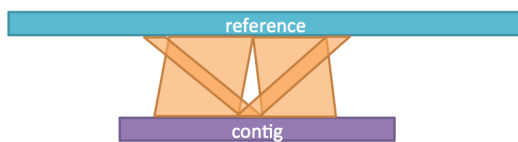


E.

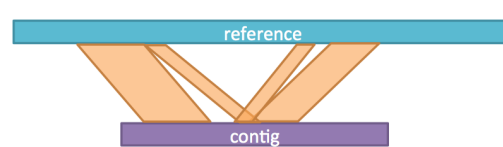
Assemblytics filtering



F.



G.



Supplementary Figure 1. Each repetitive element in a genome assembly can map ambiguously to multiple locations in the reference genome. *Delta-filter* is a component of MUMmer designed to filter repetitive alignments (Kurtz, *et al.* 2004; Phillippy, *et al.* 2008). It uses a longest-increasing subsequence (LIS) dynamic programming algorithm to identify subsets of long high identity alignments while penalizing overlapping alignments. In contrast, Assemblytics includes a filtering step to eliminate repeats without substantial unique anchoring sequence (by default 10kb). **A.** This simulated example demonstrates a case in which a 20 kb sequence in the contig (query) matches three locations in the reference exactly, except the addition of a single nucleotide (indicated by the red point) that matches better to the reference repeat on the right. **B.** Dot plot showing all alignments in the raw, unfiltered delta file output from *nucmer*. **C.** Dot plot showing results of applying *delta-filter -r* (same as unfiltered). **D.** Dot plot showing results of applying *delta-filter -q*. The 1 bp addition to a 20 kb repeat is enough to make *delta-filter -q* choose the third alignment. **E.** Dot plot showing results of the unique anchor filtering in Assemblytics. The repeat alignments are all removed since none of these include a unique anchor sequence of at least 10 kb that only aligns to a single position in the reference. Assemblytics tags any structural variants within this gap as repeat expansions for contractions, depending on whether the size of the gap between the two surrounding alignments is greater in the query or the reference, respectively. No variant is reported unless the size of the gap changes, so repeats themselves are not reported as structural variants, only changes in their size are reported as repeat expansions (increases in size) or repeat contractions (decreases in size). **F, G.** Diagrams showing additional ambiguous examples of repeats mapping to multiple locations.

Supplementary Note 2: Assemblytics Accuracy with Simulated Variants

To assess the accuracy of structural variation calling with Assemblytics, we embedded simulated insertions and deletions at known positions in the hg19 human reference genome. We then aligned the original reference genome sequence as well as the de novo assembly of the human genome assembled with MHAP presented by Berlin *et al* (2015) to this mutated reference in order to see if we could recover the simulated variants. The variants were created in the reference to allow an actual assembly to be used as the query without simulating reads, as the long read sequencers and long read assemblies have complex error models.

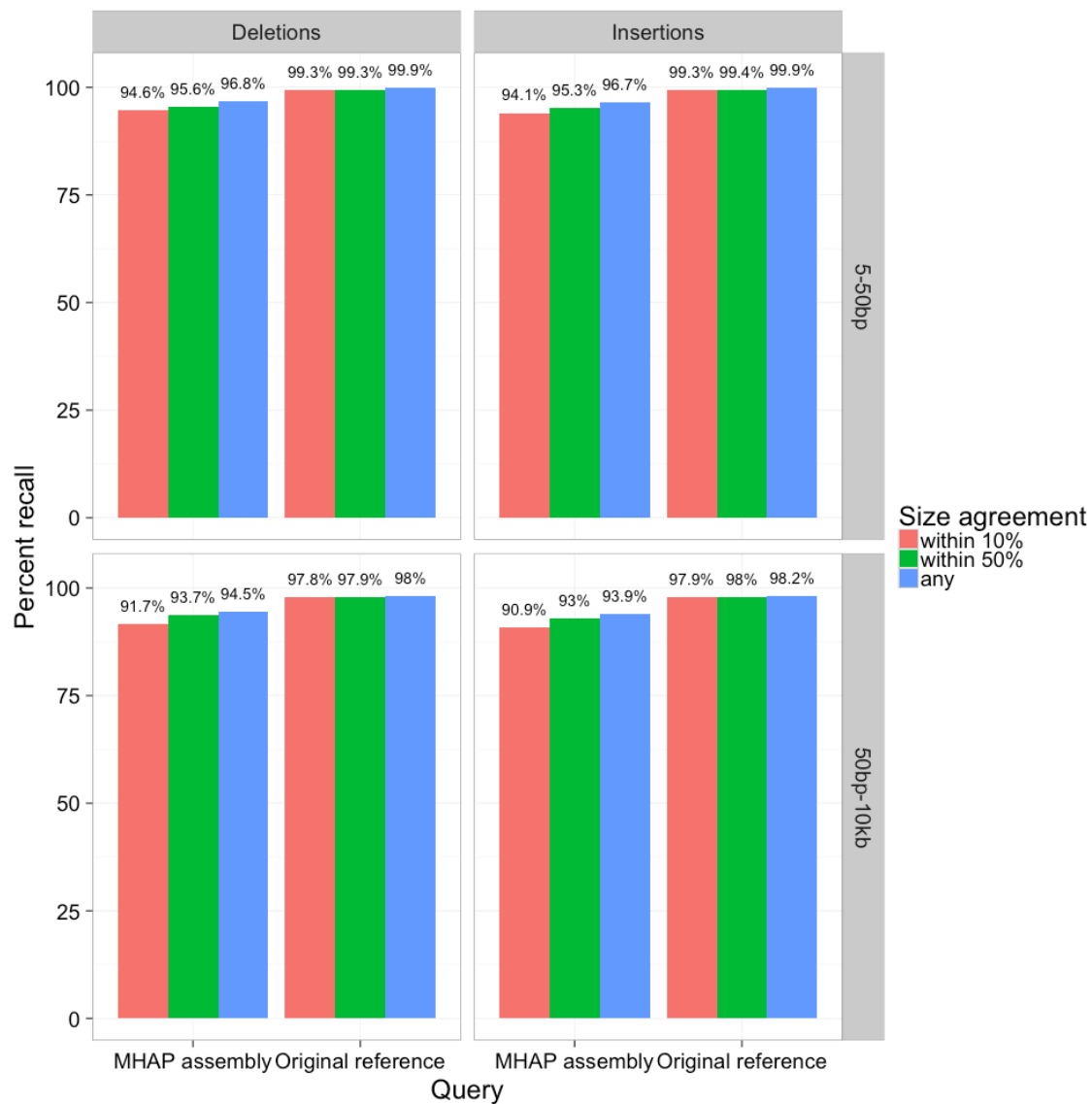
Variants were simulated at every 100 kbp, except where the 10kb sequences upstream and downstream contain a series of 100 or more Ns indicating problematic genomic regions with missing sequence; these are skipped. Variant sizes are chosen randomly from a uniform distribution in the range 50 bp to 10 kbp, and separately for the range of 5 bp to 50 bp to test whether Assemblytics performs differently on structural versus smaller variants. *Nucmer* was used for alignment and Assemblytics for variant-calling, and then *bedtools* was used to detect how many of the simulated variants were recovered. For insertions, the reference coordinates indicate only a single base-pair, so these were extended with a window of 10 bp to either side to allow for small arbitrary shifts in the local alignments.

The recall of the variants with the original reference and the MHAP assembly are presented in **Supplementary Figure 2** and shows that the vast majority of events can be detected. On inspection, variants that cannot be detected are those within long repetitive elements. The recall scores using the MHAP assembly are lower than for the original reference, as expected, since some sequences in the genome are hard to assemble and may not be represented in the MHAP assembly.

Since the MHAP assembly is from a different individual than the hg19 reference genome there are many real variants between these so the precision cannot be fully evaluated with this dataset. Instead we evaluated the precision of variants in the original reference with respect to the modified reference sequences. For deletions, 80 out of 28006 Assemblytics calls (0.286%) are false positives and among the insertions, 114 out of 28125 Assemblytics calls (0.405%) are false positives.

The simulated genomes fasta and delta files used in this figure are available at <http://qb.cshl.edu/assemblytics/reproducibility>

A.



Supplementary Figure 2. Simulation results for insertions and deletions in the size range 5 bp to 50 bp (top) and in the size range 50 bp to 10 kbp (bottom). Each of the four plots contains 28522 simulated variants, 1 at every 100 kbp simulated in the reference and using either the MHAP human assembly or the un-mutated hg19 reference.

Supplementary Note 3: Assemblytics Web Interface

A.

[Assemblytics](#) [Home](#) [Contact](#) [Cite](#) [Examples](#) [+](#)

Assemblytics

Analyze your assembly by comparing it to a reference genome

Instructions

Upload a delta file to analyze alignments of an assembly to another assembly or a reference genome

1. Download and install [MUMmer](#)
2. Align your assembly to a reference genome using *nucmer* (from MUMmer package)

```
$ nucmer -maxmatch -l 100 -c 500 REFERENCE.fa ASSEMBLY.fa -prefix OUT
```

Consult the [MUMmer manual](#) if you encounter problems

3. Delta-filter to reduce file size before upload (from MUMmer package): Here the 10000 should match the "Unique sequence length required" selected on the right. The minimum you can choose is 1000 which runs more slowly than 10000 especially on large genomes. Check the size of the final OUT.l10000.delta file. If the file size is larger than 500 MB, it might take a long time to run.

```
$ delta-filter -l 10000 OUT.delta > OUT.l10000.delta
```

4. Upload the output file OUT.l10000.delta ([view example](#)) to Assemblytics

Run Assemblytics

Drop delta file here to upload

Description

my favorite organism

Unique sequence length required

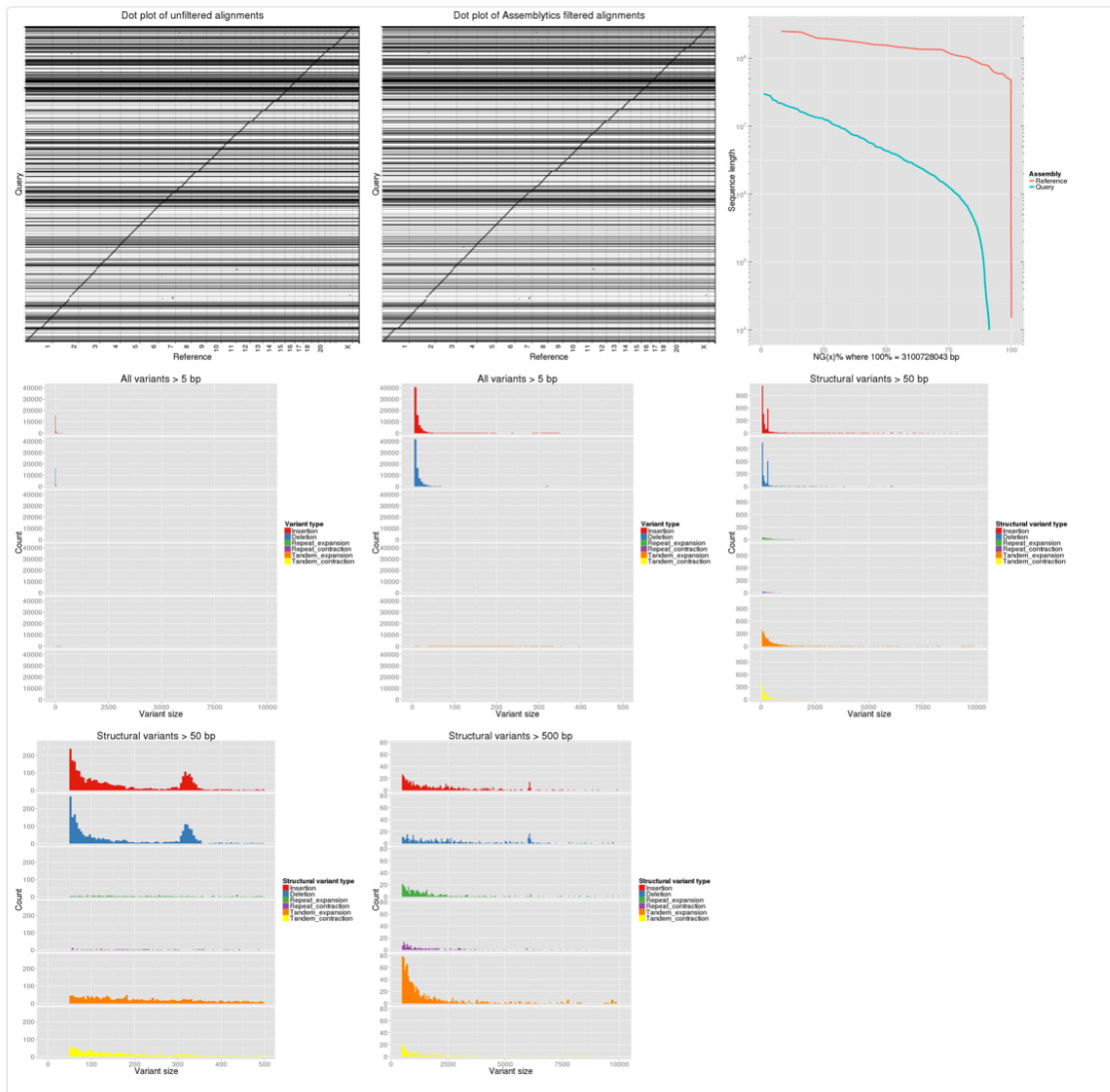
10000

Submit

Supplementary Figure 3. Front page of Assemblytics web application where users can upload a delta file from *Nucmer*, assign it a identifier, and choose the unique sequence length required to anchor alignments. The page includes instructions for running *nucmer*, and after users click the submit button any problems with the uploaded input is addressed before running Assemblytics. Examples are also available in the top navigation bar showing the output for five different organisms: Human, Drosophila, Arabidopsis, E. coli, and yeast, each assembled with MHAP and presented by Berlin et al (2015).

A.

Homo sapiens MHAP assembly



Assembly statistics

Reference:
 Number of sequences: 66
 Total sequence length: 3100728043
 Mean: 4.69807e+07
 Min: 15008
 Max: 249250621
 N50: 146364022

Query:
 Number of sequences: 4016
 Total sequence length: 2825853560
 Mean: 703649
 Min: 10024
 Max: 30140777
 N50: 5099387

B.

Variant summary statistics

Insertion		
	Count	Total bp
5-10 bp:	40664	263989
10-50 bp:	32707	567884
50-100 bp:	1133	76636
100-1,000 bp:	1732	495665
1,000-10,000 bp:	334	974016
Total:	76570	2378190
Deletion		
	Count	Total bp
5-10 bp:	41865	271523
10-50 bp:	32983	574504
50-100 bp:	1052	69054
100-1,000 bp:	1305	381744
1,000-10,000 bp:	295	1022768
Total:	77500	2319593
Tandem_expansion		
	Count	Total bp
5-10 bp:	85	600
10-50 bp:	195	5759
50-100 bp:	365	26726
100-1,000 bp:	2030	753960
1,000-10,000 bp:	493	1386016
Total:	3168	2173061
Tandem_contraction		
	Count	Total bp
5-10 bp:	76	520
10-50 bp:	203	6375
50-100 bp:	397	28357
100-1,000 bp:	884	247122
1,000-10,000 bp:	142	427004
Total:	1702	709378
Repeat_expansion		
	Count	Total bp
5-10 bp:	4	25
10-50 bp:	39	1047
50-100 bp:	41	3005
100-1,000 bp:	416	172969
1,000-10,000 bp:	219	580867
Total:	719	757913
Repeat_contraction		
	Count	Total bp
5-10 bp:	6	43
10-50 bp:	34	929
50-100 bp:	30	2158
100-1,000 bp:	226	88740
1,000-10,000 bp:	112	320425
Total:	408	412295
Total number of all variants: 160,067		
Total bases affected by all variants: 8,750,430 bp		
Total number of structural variants: 11,206		
Total bases affected by structural variants: 7,057,232 bp		

C.

Variant file preview

reference	ref_start	ref_stop	ID	size	strand	type	ref_gap_size	query_gap_size	query_coordinates
16	46397911	46397916	Assemblytics_v_1	5	+	Deletion	5	0	JSAF02000006.1:20231-20231++
16	46398849	46398854	Assemblytics_v_2	5	+	Deletion	5	0	JSAF02000006.1:21155-21155++
16	46398880	46398890	Assemblytics_v_3	10	+	Deletion	10	0	JSAF02000006.1:21181-21181++
16	46400004	46400004	Assemblytics_v_4	5	+	Insertion	0	5	JSAF02000006.1:22289-22294++
16	46400384	46400384	Assemblytics_v_5	6	+	Insertion	0	6	JSAF02000006.1:22674-22680++
16	46400845	46400851	Assemblytics_v_6	6	+	Deletion	6	0	JSAF02000006.1:23140-23140++
16	46402828	46402833	Assemblytics_v_7	5	+	Deletion	5	0	JSAF02000006.1:25109-25109++
16	46403725	46403735	Assemblytics_v_8	10	+	Deletion	10	0	JSAF02000006.1:25997-25997++
16	46404884	46404889	Assemblytics_v_9	5	+	Deletion	5	0	JSAF02000006.1:27144-27144++

Download all data

Download zip file of all results

Supplementary Figure 4. Results page of Assemblytics showing the results of aligning the *Homo sapiens* assembly from the MHAP study to the hg19 reference genome. **A.** Plots include two dot plots (before and after filtering), a query vs. reference length comparison showing cumulative sequence lengths, and five variant size distribution plots. Larger versions of these plots are shown in **Supplementary Figure 5**. Basic assembly statistics are also shown including N50. **B.** Variant summary statistics broken down by variant type and size class to show the exact counts and numbers of bases in the genome affected. **C.** A preview of the variants file in bed format, and the button where users can download all their data including the Assemblytics unique anchor filtered delta file, the full bed file of variants, and all the plots and summary tables shown on this page.

Supplementary Note 4: Assemblytics Analysis of MHAP Genome Assemblies

The five assemblies presented in the MHAP paper by Berlin et al (2015) were aligned using *nucmer* against the following reference genomes:

Human: hg19 version used in 1000 Genomes project.

Arabidopsis: TAIR10

Drosophila:

ftp://ftp.flybase.net/genomes/Drosophila_melanogaster/dmel_r6.09_FB2016_01/fasta/dmel-all-chromosome-r6.09.fasta.gz

Saccharomyces:

http://downloads.yeastgenome.org/sequence/S288C_reference/genome_releases/S288C_reference_genome_Current_Release.tgz

E. coli: http://www.ncbi.nlm.nih.gov/nuccore/NC_000913

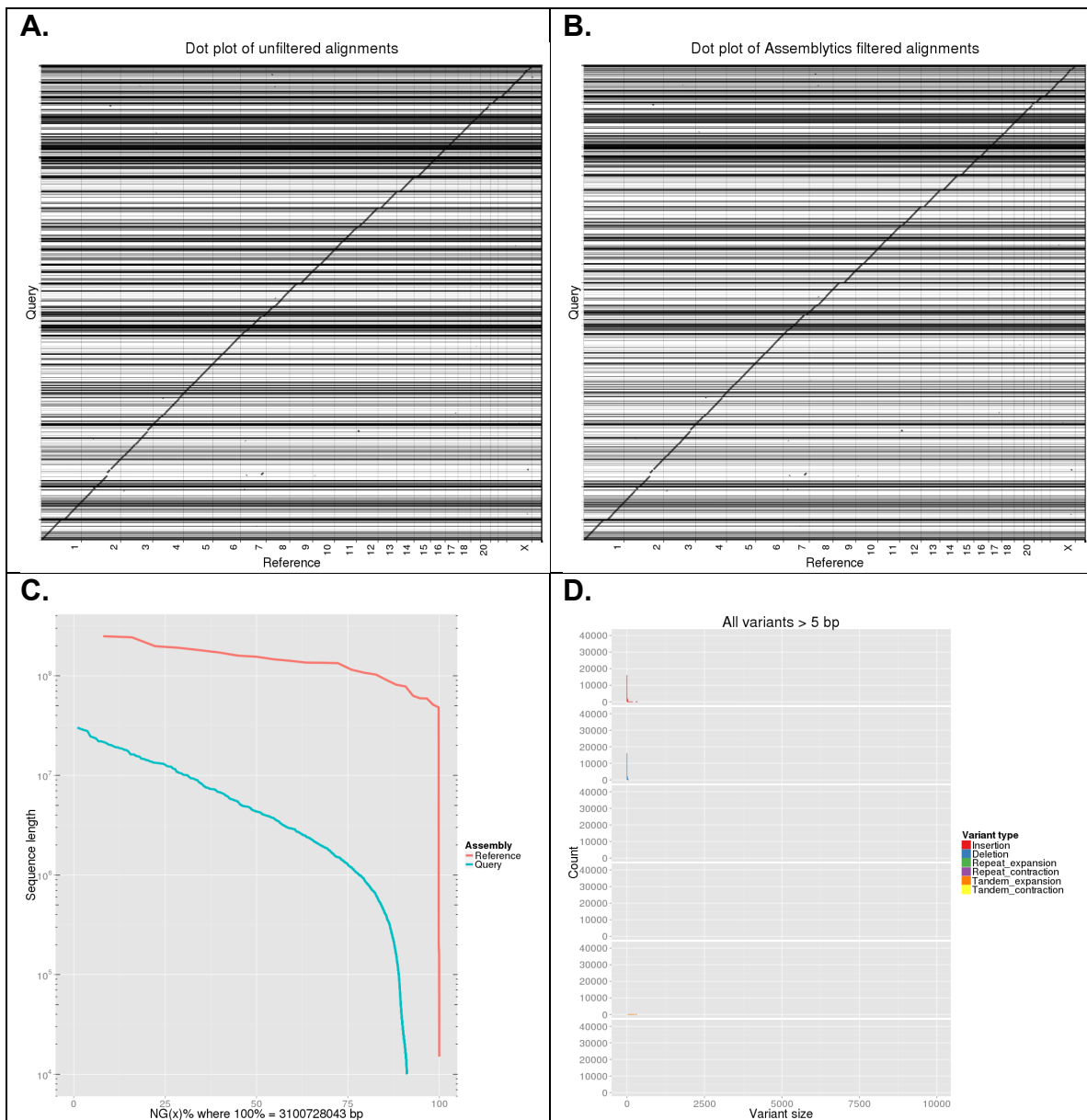
The summary statistics of the variants are presented in **Supplementary Table 1**. The full Assemblytics reports are presented in **Supplementary Figures 5-9** and are on the Assemblytics website.

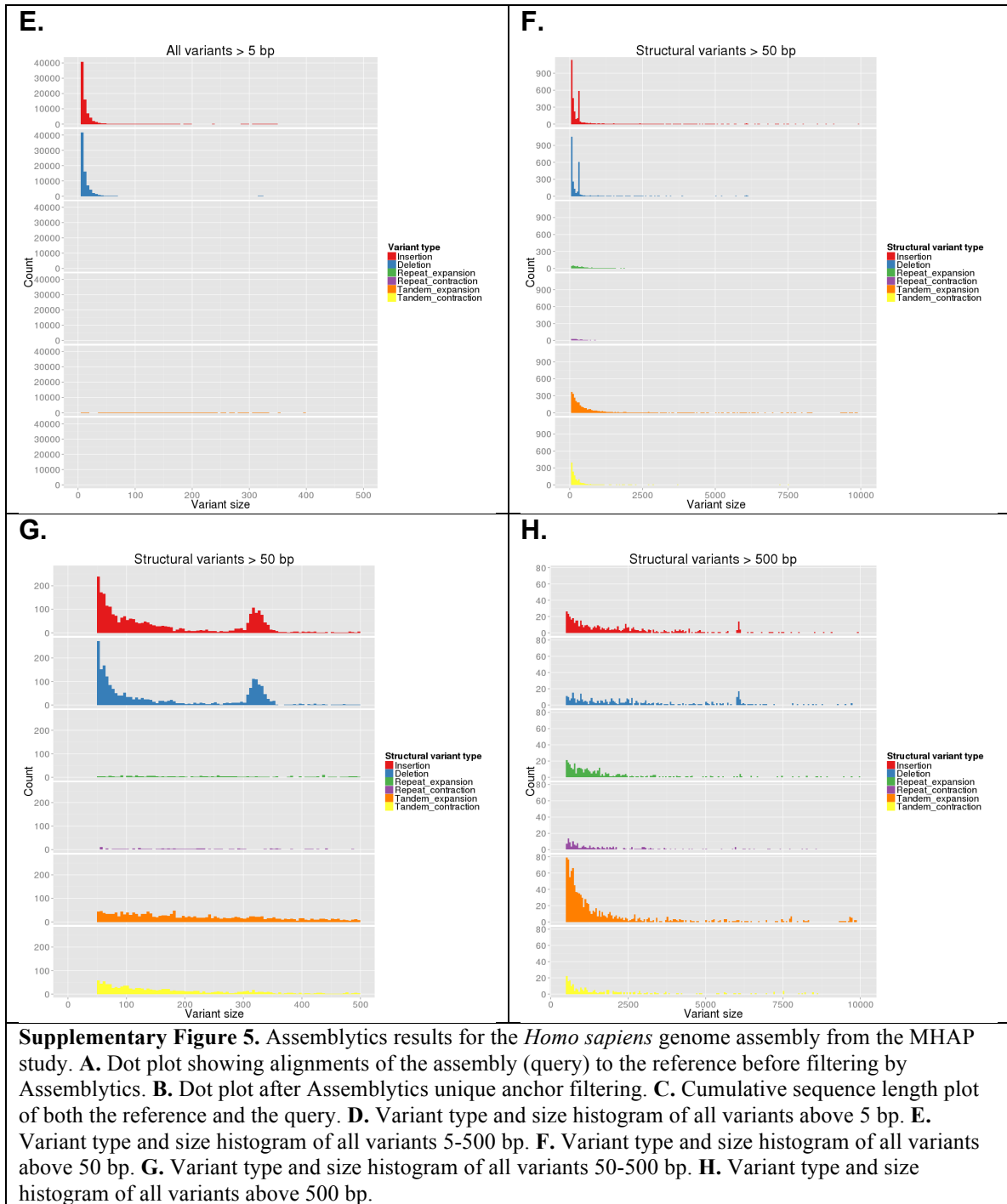
All delta files from this section are available at:

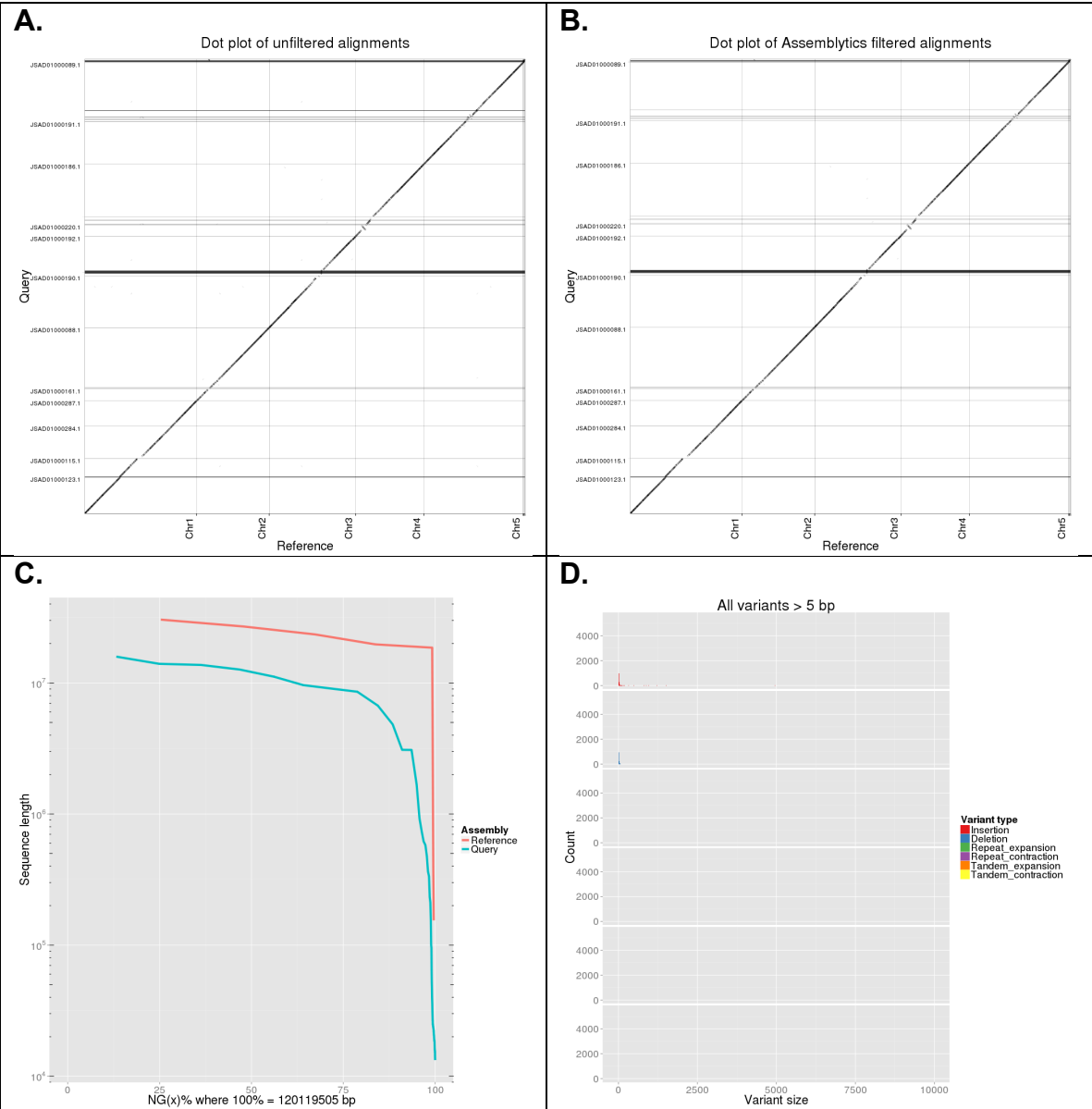
<http://qb.cshl.edu/assemblytics/reproducibility>

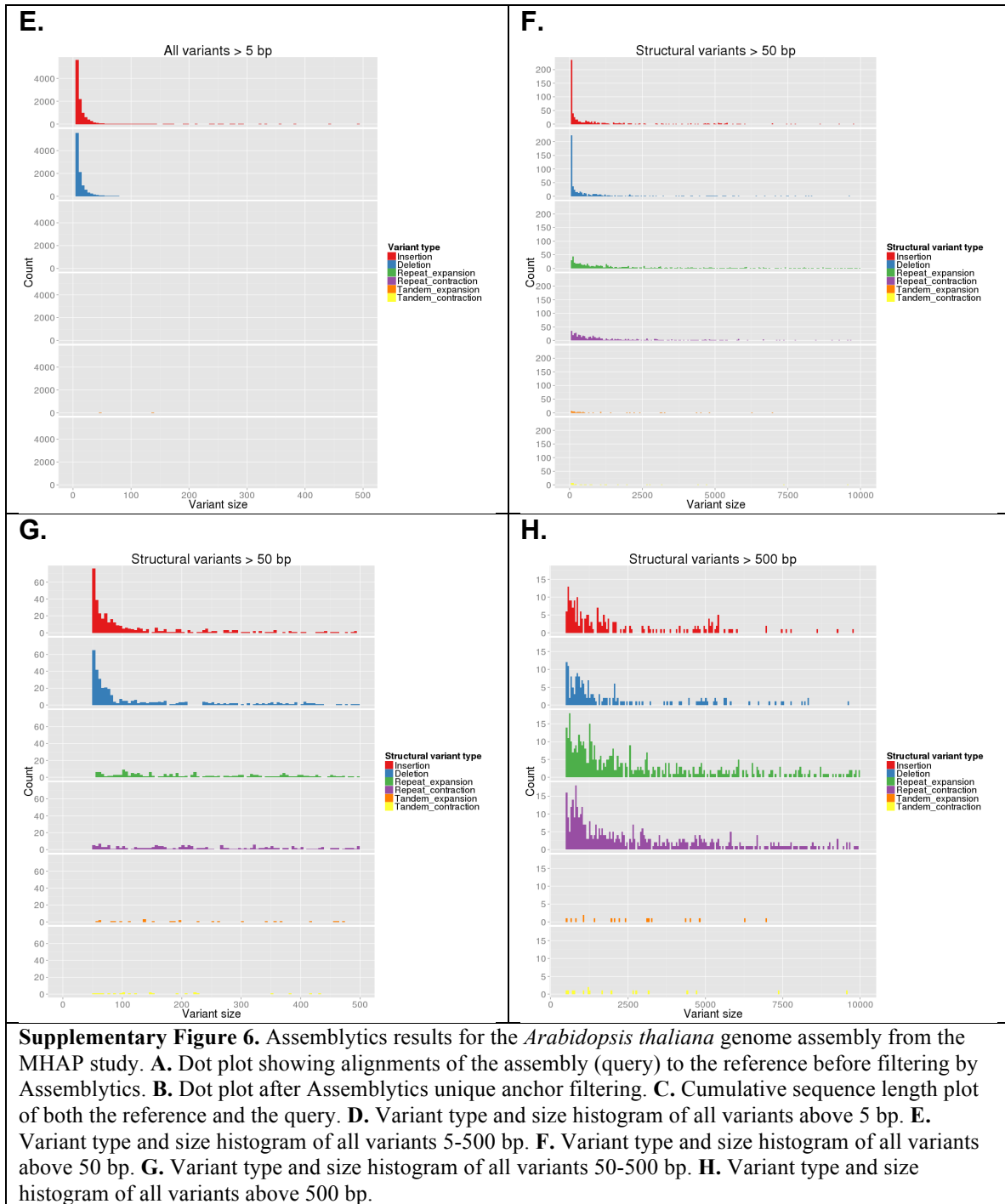
Species	Reference genome size	Structural variants (>50bp)	Total variants (>5 bp)
Homo sapiens	3,100,728,043	11,206	160,067
Drosophila melanogaster	139,047,227	204	1,189
Arabidopsis thaliana	119,667,750	2,501	23,035
Saccharomyces cerevisiae	12,157,105	42	166
Escherichia coli	4,641,652	4	4
Supplementary Table 1. Each assembly from the MHAP study was aligned to its respective reference genome using <i>nucmer</i> , and structural variants were called using Assemblytics. The count of total variants includes the structural variants.			

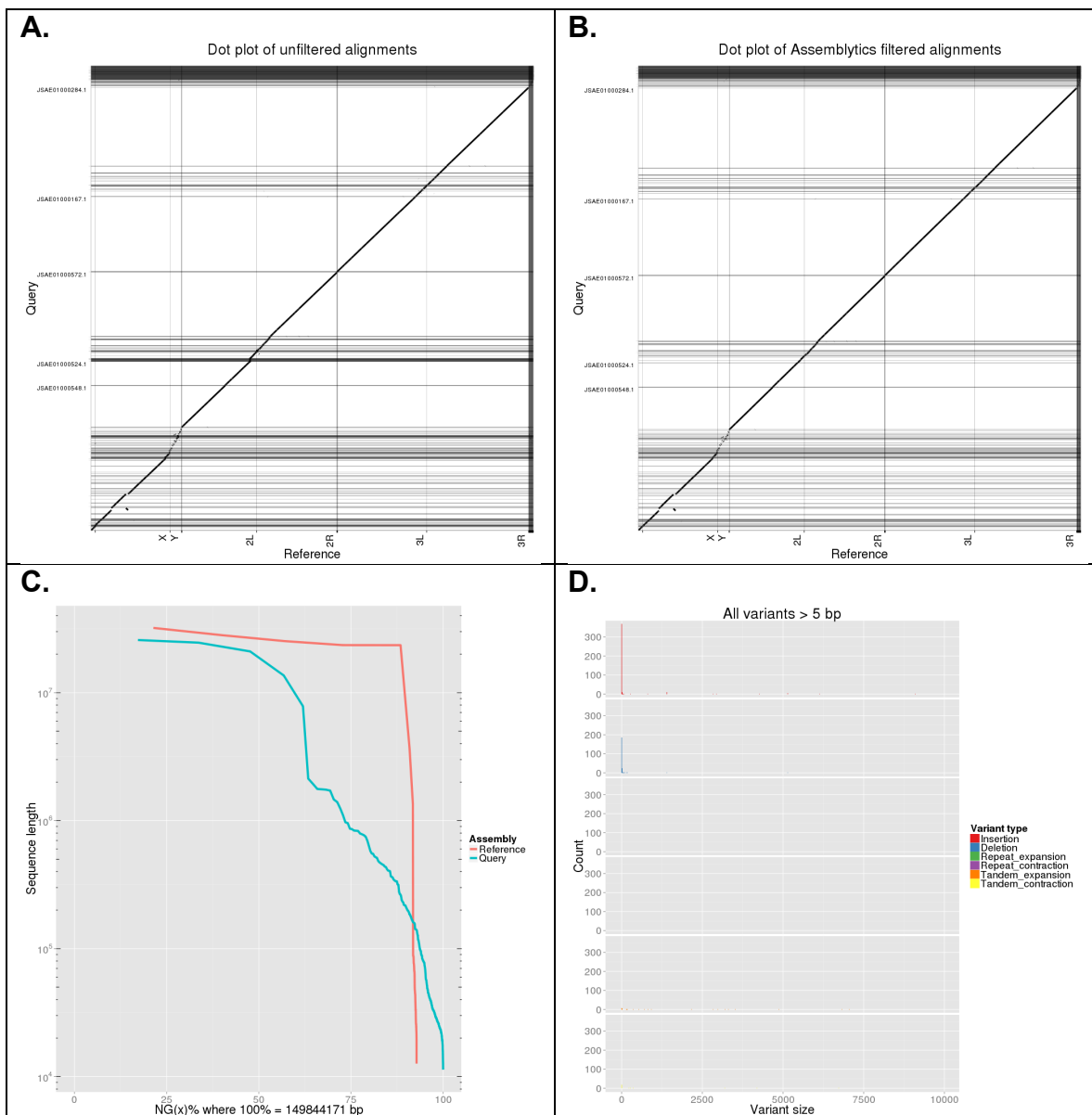
In the human assembly there is a noticeable peak of insertions and deletions around 320 bp. Extracting all deletions in the size range 300-350 bp from the variants output bed file, we see that 579 out of 604 (96%) of these intersect with known Alu elements in the RepeatMasker database of repeats for the hg19 reference genome using bedtools. This is consistent with a high level of Alu elements being deleted while there is a smaller background level of other variants that happen to fall within this size range. Since insertions of Alu elements would not be already marked in the RepeatMasker database for the reference genome, we instead extracted the sequences from the fasta file for all 588 insertions between 300 and 350 bp in size and used BLAST to align them to the Human ALU repeat elements database, where 548 (93%) of these aligned. Thus we conclude that the majority of the enrichment of insertions and deletions around 320 bp are due to jumping Alu elements.

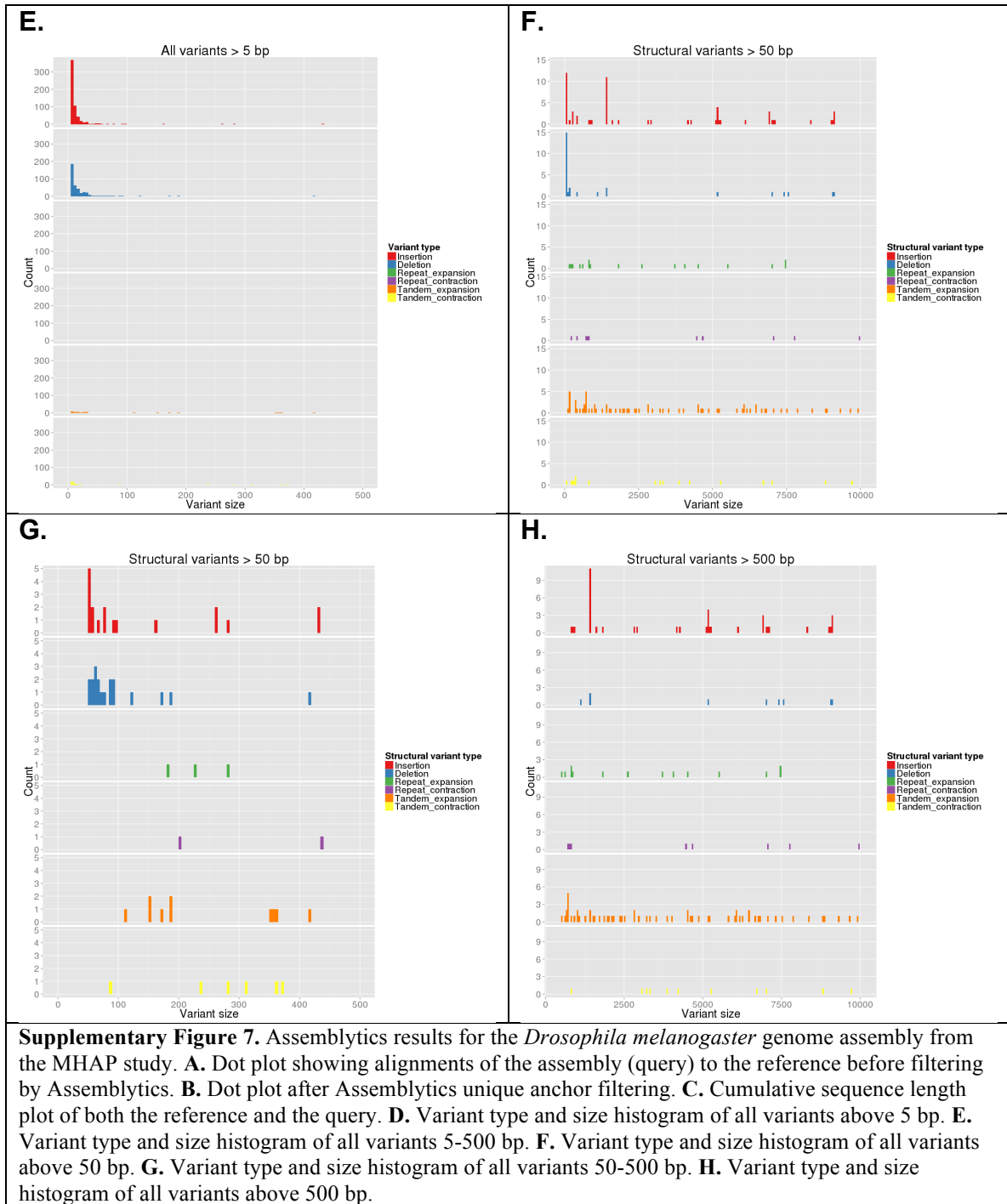


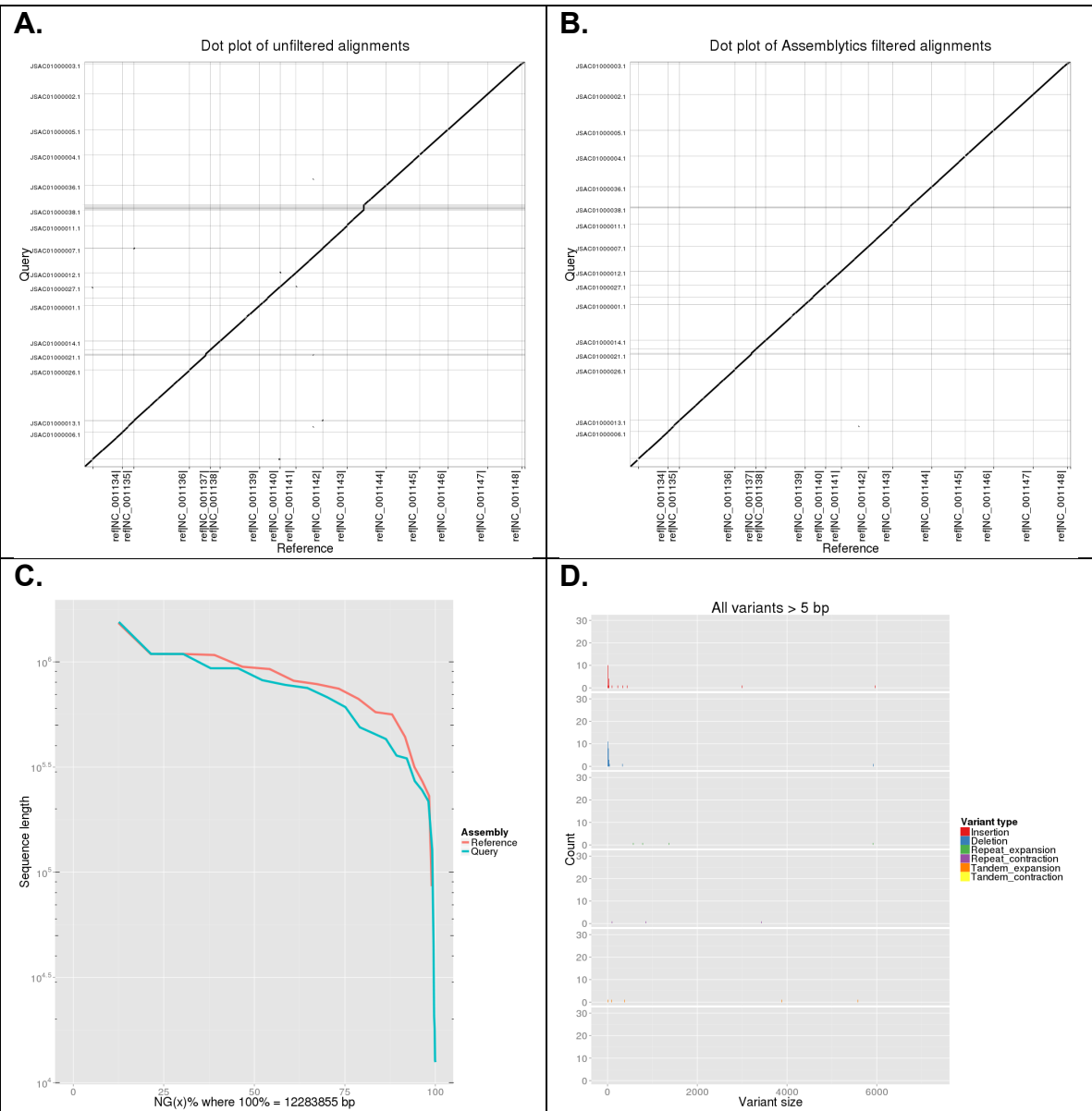


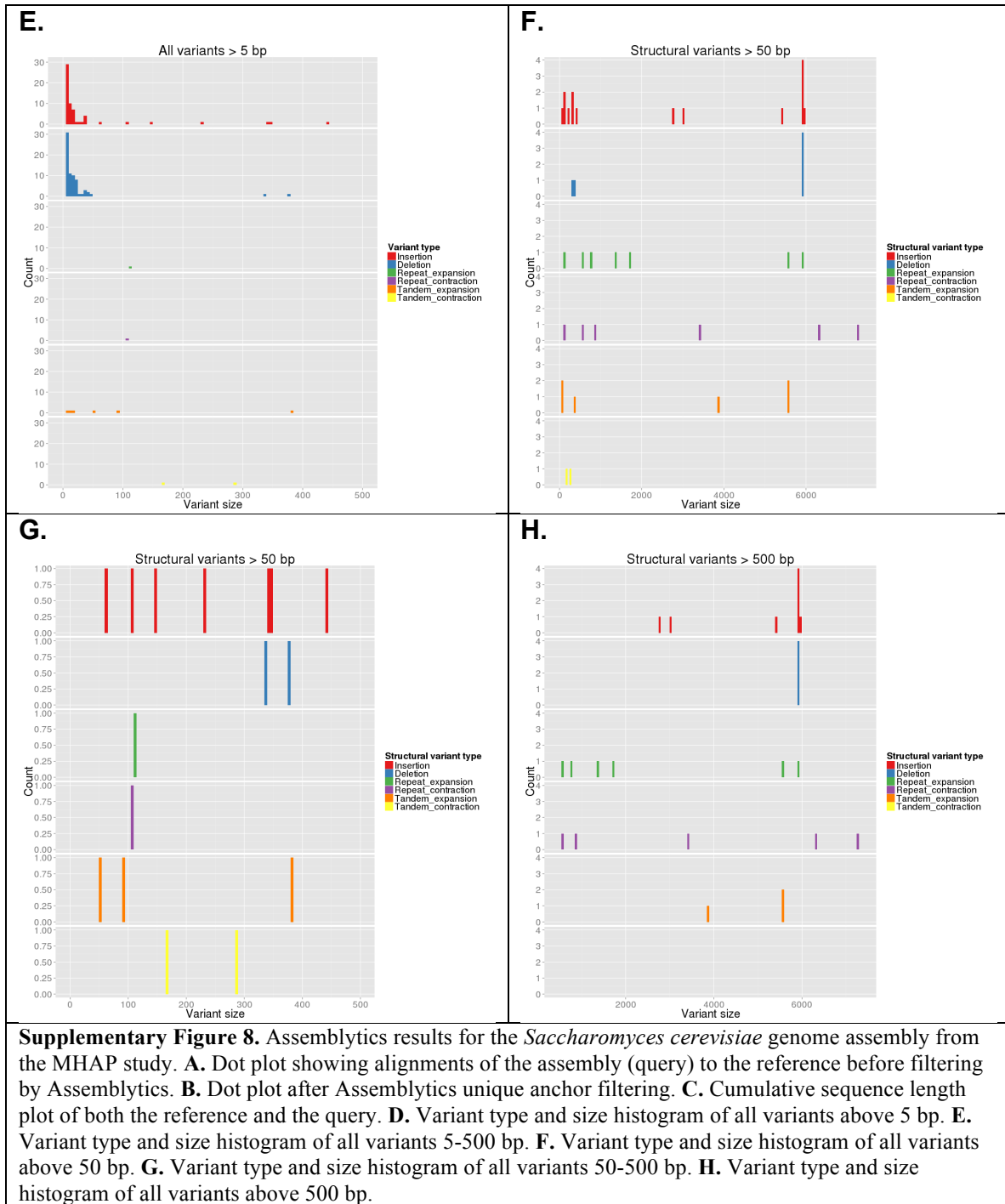


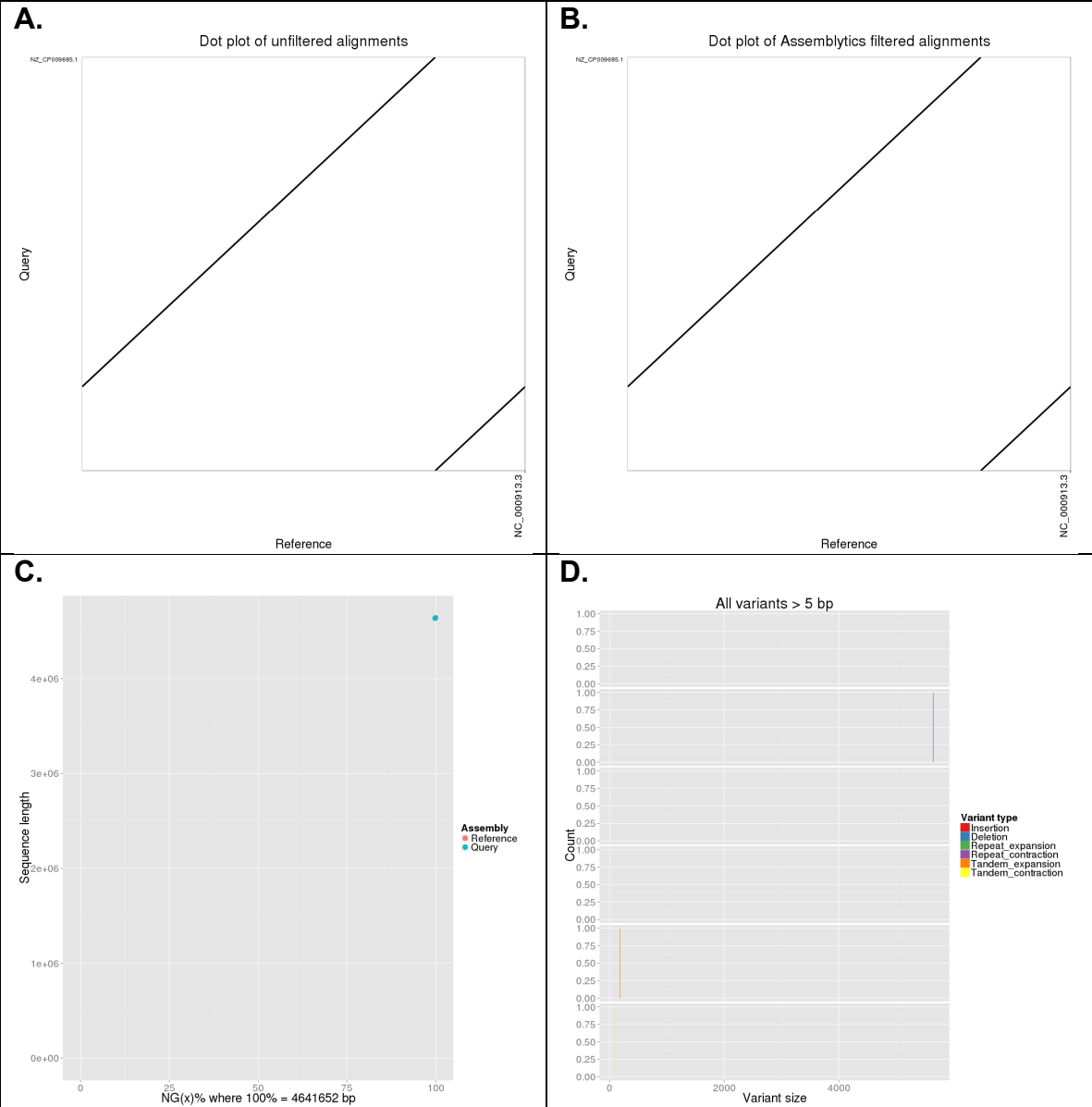


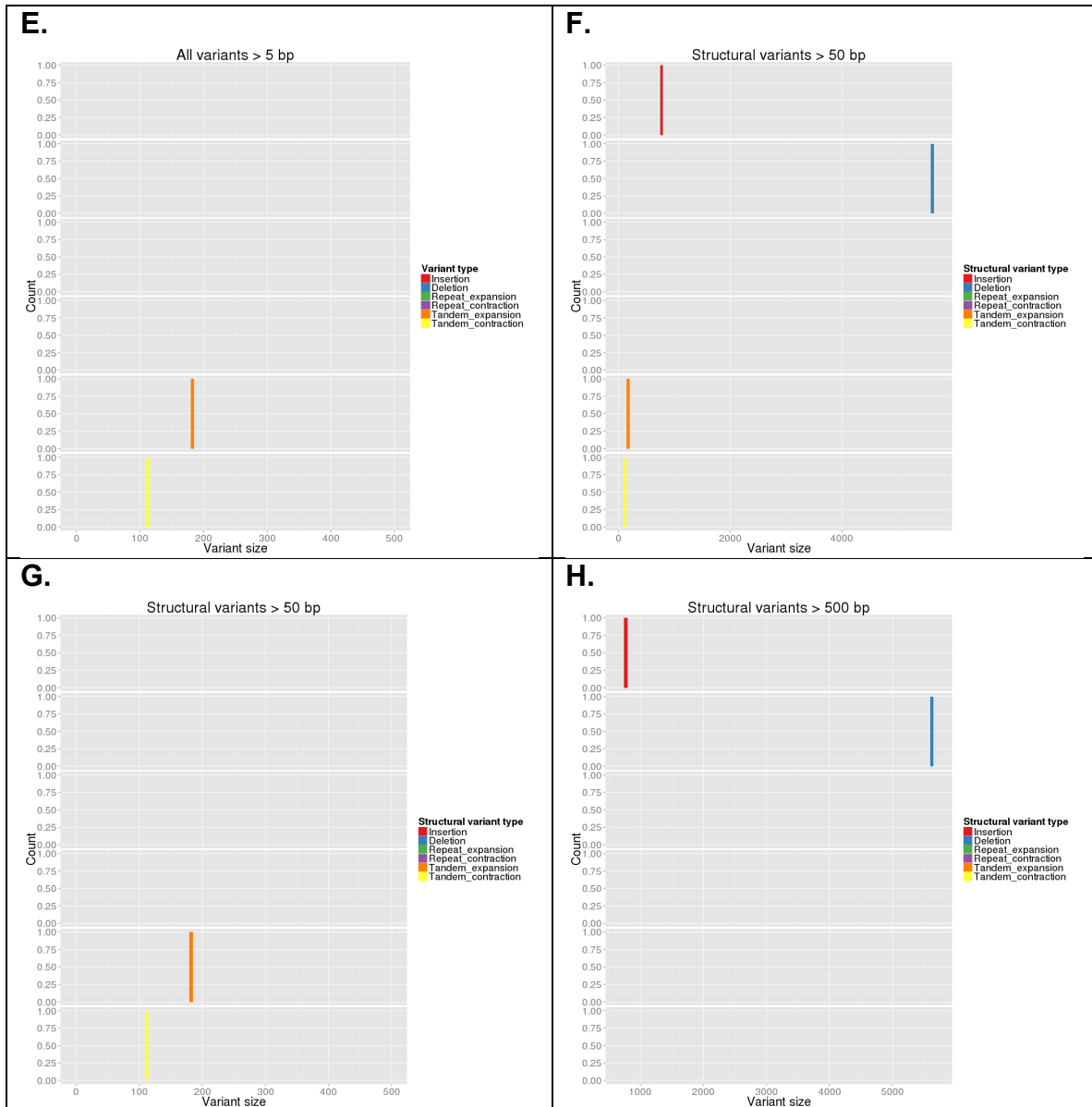












Supplementary Figure 9. Assemblytics results for the *Escherichia coli* genome assembly from the MHAP study. **A.** Dot plot showing alignments of the assembly (query) to the reference before filtering by Assemblytics. **B.** Dot plot after Assemblytics unique anchor filtering. **C.** Cumulative sequence length plot of both the reference and the query. **D.** Variant type and size histogram of all variants above 5 bp. **E.** Variant type and size histogram of all variants 5-500 bp. **F.** Variant type and size histogram of all variants above 50 bp. **G.** Variant type and size histogram of all variants 50-500 bp. **H.** Variant type and size histogram of all variants above 500 bp.