

Supplementary Table 1. Gene Trap Mutations in Rat Strains Screened for Reproduction Phenotypes

Gene symbol	Intron	Native Protein	Trapped Protein	Product (blue font = translated epitope from gene trap construct)
<i>Abca13</i>	49 of 62	5041 aa	4416 aa	exon 49 to bgeo splice (aa4377) VWVYNQKGFHSLPSYLNHLNLLWQNLPAHAADWRQYARG
<i>Alk3</i>	1 of 10	532 aa	25 aa	exon 1 to bgeo splice MTQLTYIRLLGACLFHSHVQARG
<i>Atg13</i>	17 of 17	516 aa	515 aa	exons 1-17 to bgeo splice METDLSQQDRKDLDKFIKFFALKTVQVIVQARLGEKICTRSSSSPTGSDWFNLAIKDIPVTHEAKKALSGQLPAVGRSM CVEISLKTSEGDMSMELEIWCLEMNEKCDKEIKVSYTVNRLSLLSLAIAITRVTPAYRLSRKQGHVILYRIYFGEVQLNG LGEGFQIVRVGTVGTVPVGTLLTSCAYRINLAFMSTRQFERTPPIMGIIDHFVDRPYPPSSPMHPCNRYRTAEDAGVAYPS VEDSQEVCCTSFSTSPSQSSRLSYQPAVLGLGSADLAYPVVFTAGLNTTAAHQLMVPGKEGGVPLAPNHPAHGAQ ADQERLIVHIMPSDGTHCAATPSSSEDETETVSNSSSEGRASPHDILETIFVRKVGAFVNKPINQVTLTSLDIPFAMFAPKNL ELEDADPMVNPDPSETTSPHGLHSEGGSSGSSGNAHDDFVMIDKPAFSKDDILPMDLGTFYREFQNPQLSSLSI DFGAOSMAEDLL LAVEDKLF AVFPV GIDGIDKLDL
<i>Birc</i>	1 of 14	619 aa	40 aa	exon 1 to bgeo splice MDPAEAVLQEKALKFML LA VEDKLF AVFPV GIDGIDKLDL
<i>Dlg1</i>	6 of 28	911 aa	218 aa	exons 1-6 to bgeo splice

				<p>MPVRRKQDTRALHLLLEEYRSKLSQTEDRQLRSSIERVISIFQSNLFAQALIDIQEFYEVTLLDNPCKVDHSKQCEPVPQGNP WESGSLSSAAVTSSELPGGGLSPVVEKYRYQDEEVLPSERISPOYPLAVEDKLFVFPVIGIDGIDKLDL</p>
<i>Exoc6b</i>	1 of 21	810 aa	1355 aa	<p>exon 1 to bgeo splice</p> <p>MERVKMAEESELETAAEHERILREIESTDTACIGPTLRSVYDGEEHGRFMEKLETRIRNHDRREIEKMCNIFHY QGFVDSITELLKVRGEAQKLNQVTDNPKLQHEGKELVIAMEELKQCRLQQRNISATVDKMLCLPYLE MYSKLRDQMKTKRHYPAKLTLEHLEHTYLPQVSHYRFCKVMVDNIPKLRREEIKDVSMSDLKDFLESIRKH SDKIGETAMKQAQQQRNLDNIVLQQPRLGSKRKSKKDVYTFIDTEVESTSPKSEQDSGLDVEDEEDDEE VPGAQDLVDFSPVYRQLHYSVLGARETFENYRKRQRKQARLVLQPPSNMHETLDGYYRKYFNQIVGFF VVEDHILHTTQGLVNRAYIDELWEMALSKTIAALRTHSSYCSDPNLVLDKNLIVLFADTLQVYGFVNLQLF DMLLEIRDQYSETLLKRWAGVFRNILDSDNYSPIPVTSSETYKKVVGQFPFQDTELEKQPPFKKPFSEFV PKVYNQIKEFIYAQLKFSEDLHLSSTEVDDMIRKSTNLLTRLSNSLQNVIKRKNIGLTELVOIINTTHEKS CKYLEEFITNITNVLPEYVHTTKLYGTTTFKDPARHAAEEIYTNLNQKIDQFLQADYDWMITGELDNKASDY LVDLIAFLRSTFAVFTHLPGKVAQTAGMSACKHLATSLMQLLLEAEVRQLTLGALQQFNLDVRECEQFAR SGPVPGFQEDTLQAFIDLRLQLDLFIQWDWSTYLADYGGPNCKYLRVNPVTALTLEKMKDTSRKNMIF AQFRKNERDKQKLIDTVAKQLRGLISSHHS</p>
<i>Gsg1l</i>	3 of 6	230 aa	84 aa	<p>exons 1-3 to bgeo splice</p> <p>MDKGOQEAQPGGGGEEKCRSFIDLAPASEKGVLWLSVSEVLYILLVGFSLMCELLHSSVIDGLKLNFAAVFTVL SARG</p>
<i>Grik3</i>	In exon 11 of 16	910 aa	557 aa	<p>Exons 1- truncated 11 to bgeo genomic</p>

				<p>MTAPWRRRLRSLWVEYWAGFLVCAFWIPDSRGM/PHVIRIGIFEYADGPNACQVMNAEEHAFRSANIINRNTLLPNT TLTYDIQRHFDSEATKKACDQLALGVVAIFGSPSQSCTNAVOSIGNALEVPHIQLRWKHHPLDNKDTFYVNLYPDYA SLSHAILDLVOSLKWRSATVYVDSTGLRQLQELIMAPSRYNRLKIRQLPIDSDSRLLKEMKRGREFRIIFDCSHITMAA QILKQAMAMGMNTEYYHFITLIDLVALDLEPYRSGVNLTFRILNVDNPHVSAIVEKWSMERLQAAAPRAESGLLDG VMMTDAALLYDAVHIVSVCYQARAPQMTVNSLQCHRHKAWRFGGRFMINFIKEAQWEGLTGRIVFNKTSGLRTDFDL IISLKEDGLEKYGVWSPADGLNITEVAKGRGNVTDLSLNRSLIVTTLLEPFVIMFRKSDRTLTYGNDREFEGYCIDLKEIAH ILGFSYEIRLVEDGKYGAQDDKGQWNGMVKELIDHKADLAVAPLTIHVREKAIDFSKPFMTLGVSLQLKSEVYHILSW SH</p>
<i>FstI5</i>	3 of 15	847 aa	56 aa	<p>1-3 to bgeo splice</p> <p>MFKCWSAVLLIGFIFLASEGRPTKESGVGLKYYQPLTRLRHKQEKSOESSRIKARG</p>
<i>Pqm3</i>	5 of 18	833 aa	278 aa	<p>exons 1-5 to bgeo splic</p> <p>MNSGGGLPPPSAAASPSSSSLAANAVALAVAAASSGVGVPGGPAAGVKLKCYCRYAKDKTCFYGECCQFLHEDPASGA APGLGLHSNSVPLALAAASGAGFPGLPGGGAGPPAGPKPELGVPGAATAGGGLDGPRAVAPGMDGGALTDTSLT DSYFSTSHIGVNGFGSPVETKYPLMQRMITSSSSPILLNDSAKPYTGHDPLTSSASSLFNDFGALNISQRRKTPNP TASEFI PKGGSTRLSNV/SQSNMSAFSQVFSHPMSMGSPATAGLAPARG</p>
<i>PcIb</i>	3 of 24	4880- 5085 aa	1075 aa	<p>exons 1-3 to bgeo splice</p>

				<p>MGN EASLEGEGLPEGLAAAAGAGSGSALHPGIPAGMEADLSQLSEERRQIAAVMISRAOGLPKGSVPAAAEPSMI HRKQELDSSQAPQQPGKPPDPGRPTQPGLSKSRITTDTFRSEQKLPGRSPSTLSLKSRSRTDFKEEVKSSMMMPGFFSDV NPLSAVSSVWNKFNPFDLISDSEASQEEITTKQKVVOKEQKSEGMAKPPLQQPSPKPIPKQQGQVKEVIOQDSSPKSV SSQQAEEKVKPQAPGTGKPSQQSPAQTVAQQASPGKVAQQPQGSAKATVQQPGRPAKSPAQPAQTGKSPAQPAKTP GQQAAGLEKTSSSQQPGPKSLAQTPEGHGFPLGPKVSPAQQPGTAKHPAQQPGPQTAAKVPGPTKTPAQQSGPGKTP AQQPGPTKPSPPQQPIPAKPPQPPVATKTOPQQSAPAKPQQPAPAKPQQPTPAKPPQPPPTPAKPPQPPPT ATKPPQPPPTATKPHHQQGLAKPSAQQPTKISISQTVTGRPLQPPPTSAAQTPAQGLSKTICPLCNTTELLHIPKANF NTCTECQSTVCSLGFNPNPHLTEIKEWLCNCOMQRALGGDLAAAISSPQPTPKAATAPTATASKSPVPSQQASPKK EPPSKQDSPKALESKKPEPKKPPPEPKKPPPLVKQPTLHGPTPATAPQLPVAEALPEPAPPEKPSGRLPEQAKAP VGDVERKQPKMTETRADIOSSSTTKPDLSSQVQSAQVKTASPLKTDSAKPSQSFPPTGEKTPPLDSKAMPPRPA SDSKI ISQPGPGESEKDPKHIDPIQKKDEPKKAQPKGSPKPETKPVPKGSPTPSGTRPTAGQAAPPSSQQPPKQEQSRRFSLNLG GITDAPKSQPTTPOETVTGKLFGGASIFSOASNLSTAGQQGPHPQTGPAAPSKQAPTPSOSPAAQGPAKSTGQLPPA PAKATAVKKEAKAAAAENLESKPEQAPTAKKTEKDKKPPPAKVGGKPPSEPEKAVPAHKPKDKTTKPKPACPLCRTEINLG SQEPPNFNCTECKNQVCNLCGFNPTPHLTELAVEDKLFVAVFPVGDIDKLDL</p>
<i>Sicta3</i>	2 of 9	543 aa	63 aa	<p>exons 1-2 to bgeo splice</p> <p>MTKSNNGEPRMGSRMERFQQGVRRKRTLAKKKVQNTTKEDVKSYLFRNAPVLLTVSAVIVARG</p>
<i>Spacc6</i>	2 of 8	324 aa	100 aa	<p>exons 1-2 to bgeo splice</p> <p>MGLVALVGSFVLLLLLIFRASTWACLFCFTTHEERLSLCRMVGSSEDSKIRKCRDALTDAFEGFSDMIENYDEKSHLHDEF TQMTVFLOEVAAVQGESRG</p>
<i>Trmx4</i>	5 of 7	336 aa	193 aa	<p>exons 1-5 to bgeo splice</p> <p>MTGGFCVPVFLAAWLAAAAAEEGLEQAALPAEESRVQPMTASNWTLVMEGEWMLKFFYAPWCPSCQQQTDSEWET FAKNGETLQISVGKVDVQIEPGLSGRFFVTTLPAFFHAKDGIFRRYRGVIEDLQNVILEKKWQSV EPLTGWKSPASLT MSGMAGLFSISGIW LAVEDKLFVAVFPVGDIDKLDL</p>
<i>Ubez2k</i>	1 of 6	200 aa	45 aa	<p>exon 1 to bgeo splice</p>

				MANIAVQRIKREFKEVLKSEE LAVEDKLFVFPVGGIDGIDKLDL
<i>Ube2q2</i>	1 of 14	345 aa	87 aa	exon 1 to bgeo splice MSVSGLKAELKFLASIFDKNHLLFRIVSWKLDLHCQFLVPPPPGSSHSPPPLTLHCNITLAVEDKLFVFPVGGIDGID KLDDL
<i>Zmynd8</i>	3 of 21	1208 aa	82 aa	exons 1-3 to bgeo splice MDISTRSKDPGSTERTAQKRKVPSPPHSSNGHSPQDSSTSPIKKKKKPGLLNSSKDQLAVEDKLFVFPVGGIDGIDKLDD L

Supplementary Table 2. Reproduction Phenotypes of Rats with Gene Trap Mutations

Mutated Gene	Intronic Insertion	Breeders	Breeding Pairs	Litter Number	Mean Litter Size (±S.E.M.)	Genotypes Ratio (%WT:Het:Homo)	Sex Ratio (Female:Male)	Reproduction Phenotype	
Wildtype	None	WT x WT	3	10	11.8±1.27	100:0:0	53:47	Normal	
<i>Abca13</i>	Intron 49 of 62	Het x Het	2	6	7.33±0.80	p=0.01*	25:61:14	55:45	Subfertile
		F Homo x WT	2	6	9.50±1.19		0:100:0	52:48	Normal
		M Homo x WT	2	6	12.8±1.08		0:100:0	39:61	Normal
<i>Alk3</i>	Intron 1 of 10	Het x Het	2	6	5.83±0.60	p=0.001*	49:51:0	43:57	Embryonic Lethal
		F Homo x WT	0	0	-		-	-	-
		M Homo x WT	0	0	-		-	-	
<i>Alg13</i>	Intron 16 of 16	Het x Het	2	6	8.33±1.08	p=0.06*	30:42:28	42:58	Subfertile
		F Homo x WT	2	0	0		-	-	Adult Lethal
		M Homo x WT	2	0	0		-	-	Adult Lethal
<i>Birc</i>	Intron 1 of 14	Het x Het	2	6	11.6±1.50		28:43:29	47:53	Normal
		F Homo x WT	2	6	9.00±1.88		0:100:0	54:46	Normal
		M Homo x WT	2	0	0		-	-	Gametogenesis Defect
<i>Dlg1^t</i>	Intron 6 of 28	Het x Het	-	-	-		-	-	Unknown ^t
		F Homo x WT	-	-	-		-	-	-
		M Homo x WT	-	-	-		-	-	-
<i>Exco6b</i>	Intron 1 of 21	Het x Het	2	6	11.6±0.40		31:69:0	41:59	Embryonic Lethal
		F Homo x WT	0	0	-		-	-	-
		M Homo x WT	0	0	-		-	-	-
<i>Gsgl1</i>	Intron 3 of 6	Het x Het	2	5	5.80±1.02	p=0.003*	3:69:28	52:48	Subfertile
		F Homo x WT	2	6	9.33±2.58		0:100:0	46:54	Normal
		M Homo x WT	2	5	12.8±1.16		0:100:0	48:52	Normal
<i>Grik3</i>	Exon 11 of 16	Het x Het	2	6	9.50±0.84		19:51:30	44:56	Normal
		F Homo x WT	2	5	5.80±1.07	p=0.003*	0:100:0	48:52	Subfertile
		M Homo x WT	2	6	11.3±2.07		0:100:0	54:46	Normal
<i>Fstl5</i>	Intron 3 of 15	Het x Het	2	5	12.2±0.97		27:39:34	52:48	Normal
		F Homo x WT	2	5	11.2±1.88		0:100:0	55:45	Normal
		M Homo x WT	2	3	14.0±1.00		0:100:0	50:50	Normal
<i>Pan3</i>	Intron 5 of 18	Het x Het	2	6	9.67±1.58		27:48:25	36:64	Normal
		F Homo x WT	2	6	6.00±1.55	p=0.014*	0:100:0	44:56	Subfertile
		M Homo x WT	2	0	0		-	-	Gametogenesis Defect
<i>Pclol^{fl}</i>	Intron 3 of 25	Het x Het	2	6	11.3±1.26		26:62:12	53:47	Normal <8 mo
		F Homo x WT	2	0	0		-	-	Behavioral
		M Homo x WT	2	0	0		-	-	Behavioral
<i>Rgs22</i>	Intron 2 of 24	Het x Het	2	4	14.3±1.93		24:53:23	52:48	Normal
		F Homo x WT	2	4	13.8±0.63		0:100:0	55:45	Normal

		M Homo x WT	2	4	11.8±1.77		0:100:0	53:47	Normal
<i>Slc1a3</i>	Intron 2 of 9	Het x Het	2	12	6.92±1.33	p=0.01*	22:77:1	51:49	Embryonic Lethal
		F Homo x M Het	0	0	-		-	-	-
		M Homo x WT	0	0	-		-	-	-
<i>Spaca6</i>	Intron 2 of 8	Het x Het	2	5	10.0±0.89		24:42:34	50:50	Normal
		F Homo x WT	2	5	9.40±2.20		0:100:0	51:49	Normal
		M Homo x WT	2	0	0		-	-	Fertilization
<i>Tmx4</i>	Intron 5 of 7	Het x Het	2	8	9.63±1.25		39:61:0	51:49	Embryonic Lethal
		F Homo x WT	0	0	-		-	-	-
		M Homo x WT	0	0	-		-	-	-
<i>Ube2k</i>	Intron 1 of 6	Het x Het	2	6	9.40±1.14	p=0.036*	21:53:26	55:45	Subfertile
		F Homo x WT	2	0	0		-	-	Gametogenesis Defect
		M Homo x WT	2	0	0		-	-	Gametogenesis Defect
<i>Ube2q2</i>	Intron 1 of 14	Het x Het	2	6	9.50±1.32		19:50:31	42:58	Normal
		F Homo x WT	2	5	12.2±1.93		0:100:0	47:53	Normal
		M Homo x WT	2	6	12.0±1.06		0:100:0	44:56	Normal
<i>Zmynd8</i>	Intron 3 of 21	Het x Het	2	7	7.80±1.22	p=0.003*	75:25:00	44:56	Embryonic Lethal
		F Homo x WT	0	0	-		-	-	-
		M Homo x WT	0	0	-		-	-	-

*Potentially Subfertile due to smaller mean letter sizes (p<0.1 shown versus wildtype).

[†]Heterozygous *Dlg1-wt/gt* mutant females produced from wildtype recipient males displayed problems breeding for undefined reasons.

[‡]Heterozygous *PcLo-wt/gt* females 8-14 months of age did not reproduce when paired with wildtype males of similar age (n=4 breeder pairs).

Additional analyses confirmed reproductive failure in *Spaca6^{9^{fl}/9^{fl}}* males (n=4), *PcLo^{9^{fl}/9^{fl}}* females (n=6) and *PcLo^{9^{fl}/9^{fl}}* males (n=6).

Supplementary Table 3. Body, Testis and Epididymal Weights in Mutant Rats

Mutant *	Rats (n)	Body wt (g) ±S.E.M.	Testis wt (g) ±S.E.M.		Epididymus wt (g) ±S.E.M.		Testis:Body wt [†] ±S.E.M.	Epid:Body wt [†] ±S.E.M.
<i>Atg13</i>	3	354±14	1.51±0.07	p=0.0054	0.49±0.04	p=0.025	4.27±0.08	1.38±0.01
<i>Brcc</i>	3	428±40	0.67±0.04	P<0.0001	0.30±0.03	P<0.0001	1.55±0.10	0.70±0.03
<i>Grik3</i>	4	511±63	1.86±0.13	p>0.10	0.61±0.05	p>0.10	3.64±0.24	1.19±0.11
<i>Pan3</i>	4	436±19	1.10±0.04	P<0.0001	0.45±0.02	p=0.001	2.53±0.15	1.03±0.09
<i>Pclo</i>	4	536±58	1.95±0.05	P=0.1	0.64±0.01	p>0.10	3.64±0.39	1.19±0.13
<i>Spacc6</i>	6	447±20	2.01±0.06	p>0.10	0.57±0.02	p>0.10	4.50±0.18	1.28±0.09
<i>Ube2k</i>	4	302±38 [¶]	0.35±0.01	P<0.0001	0.24±0.03	P<0.0001	1.16±0.25	0.79±0.38
<i>Ube2q2</i>	3	486±22	2.17±0.04	p>0.10	0.65±0.03	p>0.10	4.47±0.20	1.33±0.09
Wildtype	6	506±28	1.98±0.10	n.a.	0.64±0.02	n.a.	3.91±0.26	1.26±0.07

* Male rats were analyzed between 90-250 days of age; significant differences in mutant testis and epididymal weights are compared to wildtype littermates at the respective age of analysis.

¶ Homozygous mutant *Ube2k* rats displayed significantly reduced body weights (p<0.001).

† Organ:body weight (wt) ratios x 1000.

Supplementary Table 4. Mutant Rat Phenotypes in Current Study* Compared Across Species.

Mutated Gene	Insertion Site	Rat*	Mouse ⁿ	Human ⁿ	Other Organisms ⁿ
<i>Abca13</i>	Intron 49 of 62	None observed	Unknown	Schizophrenia, bipolar disorder, depression ¹	Unknown
<i>Atg13</i>	Intron 16 of 16	Shortened lifespan; renal failure; inflammation; autophagy defect	Embryonic lethal, cardiovascular development defects ²	Autoimmune Disease ³	Premature senescence in plants ⁴ ; reduced lifespan in yeasts ⁵ and worms ⁶ ; embryonic lethal in flies ⁷ .
<i>Alk3</i>	Intron 1 of 10	Stunted growth in heterozygotes; embryonic lethal in homozygotes	Normal growth in heterozygotes; embryonic lethal in homozygotes; epiblast defects ⁸	Juvenile and hamartomatous polyposis syndromes ⁹⁻¹¹	Anterior-posterior wing patterning and oogenesis in flies ¹²⁻¹⁴ ;
<i>Birc</i>	Intron 1 of 14	Spermatogonial development block	Males sub-fertile ¹⁵ ; neural stem cell fate ^{16,17} ; longer circadian period ¹⁸ ; DNA stability ¹⁶	10q Medulloblastoma tumor suppressor locus ¹⁹ ; 10q split hand/foot disease ^{20,21} ; promotes HIV infection ²²	Xenopus dorsal axis duplication ²³
<i>Dlg1</i>	Intron 6 of 28	Dominant ^{†1}	Recessive effects on aging, renal function, craniofacial, vision, and urogenital defects ²⁴⁻²⁷ .	Schizophrenia ²⁸ ; 3q29 microdeletion syndrome ²⁹	Tumor suppressor ³⁰ ; alcohol tolerance in flies ³¹
<i>Exoc6b</i>	Intron 1 of 21	Embryonic lethal	Unknown	Behavioral defects, epilepsy ^{32,33}	Unknown
<i>Gsg1l</i>	Intron 3 of 6	None observed	Unknown	Candidate risk factor for colon cancer ³⁴	Unknown
<i>Grik3</i>	Exon 11 of 16	None observed	Synaptic transmission ³⁵	Behavioral disorders ³⁶ ; schizophrenia ³⁷	Unknown
<i>Fstl5</i>	Intron 3 of 15	None observed	Unknown	Adiposity-Related Anthropometric Traits ³⁸	Unknown
<i>Pan3</i>	Intron 5 of 18	Post-meiotic spermatogenesis	Unknown	Unknown	Non-lethal, polyadenylation defects in yeast ³⁹
<i>Pclo</i>	Intron 3 of 25	Female & male social behavior, mating behaviors, seizures	Adults reproduce and behave normally, postnatal growth and lifespan stunted ⁴⁰	Major depressive, bipolar disorders, seizures, pontocerebellar hypoplasia, emotional processing ⁴¹⁻⁴⁶	Unknown
<i>Rgs22t</i>	Intron 2 of 24	None observed	Unknown	Unknown	Unknown
<i>Sic1a3</i>	Intron 2 of 9	Embryonic lethal	Adult fertile, aging, behavior disorders ^{47,49}	Behavior; addiction, seizure ^{50,51}	Unknown
<i>Spaca6</i>	Intron 2 of 8	Sperm function	Sperm function ⁵²	Unknown	Unknown
<i>Tmx4</i>	Intron 5 of 7	Embryonic lethal	Unknown	Unknown	Unknown
<i>Ube2k</i>	Intron 1 of 6	Female & male meiosis	Adults fertile; resistant to amyloid- β toxicity ⁵³	Unknown	Unknown
<i>Ube2q2</i>	Intron 1 of 14	None observed	Unknown	Renal failure ^{54,55} ; arthritis ⁵⁶	Unknown
<i>Zmynd8</i>	Intron 3 of 21	Embryonic lethal	P14 viable ²¹	Unknown	Unknown

* Reproduction phenotypes observed in current study; all phenotypes recessive unless stated as dominant or occurring in heterozygotes. † Gene trap is in Reverse Orientation to the *Rgs22* gene, gene trap is predicted to be non-functional. †† Strain was not propagated from F1 *Dlg1* heterozygote mutant females for unknown reasons but displayed solitary behavior in cage (potential mating behavior or other reproduction defect). ††† PubMed citation referenced below; only spontaneous or non-conditional experimental mutations referenced.

References to Supplementary Table 4

1. Knight, H.M. *et al.* A cytogenetic abnormality and rare coding variants identify ABCA13 as a candidate gene in schizophrenia, bipolar disorder, and depression. *Am J Hum Genet* **85**, 833-46 (2009).
2. Katzuka, T. & Mizushima, N. Atg13 is essential for autophagy and cardiac development in mice. *Molecular and Cellular Biology* **36**, 585-595 (2016).
3. Ferreira, R.C. *et al.* Association of IFIH1 and other autoimmunity risk alleles with selective IgA deficiency. *Nat Genet* **42**, 777-80 (2010).
4. Suttangkakul, A., Li, F., Chung, T. & Vierstra, R.D. The ATG1/ATG13 protein kinase complex is both a regulator and a target of autophagic recycling in Arabidopsis. *Plant Cell* **23**, 3761-79 (2011).
5. Funakoshi, T., Matsuyama, A., Noda, T. & Ohsumi, Y. Analyses of APG13 gene involved in autophagy in yeast, *Saccharomyces cerevisiae*. *Gene* **192**, 207-13 (1997).
6. Tian, E., Wang, F., Han, J. & Zhang, H. epg-1 functions in autophagy-regulated processes and may encode a highly divergent Atg13 homolog in *C. elegans*. *Autophagy* **5**, 608-15 (2009).
7. Chang, Y.Y. & Neufeld, T.P. An Atg1/Atg13 complex with multiple roles in TOR-mediated autophagy regulation. *Mol Biol Cell* **20**, 2004-14 (2009).
8. Mishina, Y., Suzuki, A., Ueno, N. & Behringer, R.R. Bmpr encodes a type I bone morphogenetic protein receptor that is essential for gastrulation during mouse embryogenesis. *Genes Dev* **9**, 3027-37 (1995).
9. Kim, I.J. *et al.* Identification of a novel BMPR1A germline mutation in a Korean juvenile polyposis patient without SMAD4 mutation. *Clin Genet* **63**, 126-30 (2003).
10. Zhou, X.P. *et al.* Germline mutations in BMPR1A/ALK3 cause a subset of cases of juvenile polyposis syndrome and of Cowden and Bannayan-Riley-Ruvalcaba syndromes. *Am J Hum Genet* **69**, 704-11 (2001).
11. Nieminen, T.T. *et al.* BMPR1A mutations in hereditary nonpolyposis colorectal cancer without mismatch repair deficiency. *Gastroenterology* **141**, e23-6 (2011).
12. Xia, L. *et al.* The Fused/Smurf complex controls the fate of Drosophila germline stem cells by generating a gradient BMP response. *Cell* **143**, 978-90 (2010).
13. Singer, M.A., Penton, A., Twombly, V., Hoffmann, F.M. & Gelbart, W.M. Signaling through both type I DPP receptors is required for anterior-posterior patterning of the entire Drosophila wing. *Development* **124**, 79-89 (1997).
14. Burke, R. & Basler, K. Dpp receptors are autonomously required for cell proliferation in the entire developing Drosophila wing. *Development* **122**, 2261-9 (1996).
15. Guardavaccaro, D. *et al.* Control of meiotic and mitotic progression by the F box protein beta-Trcp1 in vivo. *Dev Cell* **4**, 799-812 (2003).
16. Guardavaccaro, D. *et al.* Control of chromosome stability by the beta-TrCP-REST-Mad2 axis. *Nature* **452**, 365-9 (2008).

17. Westbrook, T.F. *et al.* SCFbeta-TRCP controls oncogenic transformation and neural differentiation through REST degradation. *Nature* **452**, 370-4 (2008).
18. Reischl, S. *et al.* Beta-TrCP1-mediated degradation of PERIOD2 is essential for circadian dynamics. *J Biol Rhythms* **22**, 375-86 (2007).
19. Scott, D.K. *et al.* Identification and analysis of tumor suppressor loci at chromosome 10q23.3-10q25.3 in medulloblastoma. *Cell Cycle* **5**, 2381-9 (2006).
20. Kano, H. *et al.* Genomic rearrangement at 10q24 in non-syndromic split-hand/split-foot malformation. *Hum Genet* **118**, 477-83 (2005).
21. Lyle, R. *et al.* Split-hand/split-foot malformation 3 (SHFM3) at 10q24, development of rapid diagnostic methods and gene expression from the region. *Am J Med Genet A* **140**, 1384-95 (2006).
22. Bour, S., Perrin, C., Akari, H. & Strebel, K. The human immunodeficiency virus type 1 Vpu protein inhibits NF-kappa B activation by interfering with beta TrCP-mediated degradation of I kappa B. *J Biol Chem* **276**, 15920-8 (2001).
23. Liu, C. *et al.* beta-Trcp couples beta-catenin phosphorylation-degradation and regulates Xenopus axis formation. *Proc Natl Acad Sci U S A* **96**, 6273-8 (1999).
24. Caruana, G. & Bernstein, A. Craniofacial dysmorphogenesis including cleft palate in mice with an insertional mutation in the discs large gene. *Mol Cell Biol* **21**, 1475-83 (2001).
25. Rivera, C. *et al.* Cell-autonomous requirements for Dlg-1 for lens epithelial cell structure and fiber cell morphogenesis. *Dev Dyn* **238**, 2292-308 (2009).
26. Mahoney, Z.X. *et al.* Discs-large homolog 1 regulates smooth muscle orientation in the mouse ureter. *Proc Natl Acad Sci U S A* **103**, 19872-7 (2006).
27. Iizuka-Kogo, A., Ishida, T., Akiyama, T. & Senda, T. Abnormal development of urogenital organs in Dlg1-deficient mice. *Development* **134**, 1799-807 (2007).
28. Mulle, J.G. *et al.* Microdeletions of 3q29 confer high risk for schizophrenia. *Am J Hum Genet* **87**, 229-36 (2010).
29. Willatt, L. *et al.* 3q29 microdeletion syndrome: clinical and molecular characterization of a new syndrome. *Am J Hum Genet* **77**, 154-60 (2005).
30. Alexander, C. *et al.* Fine scale mapping places DLG1, the gene encoding hDlg, telomeric to the OPA1 candidate region. *Mamm Genome* **8**, 795-6 (1997).
31. Maiya, R. *et al.* DLGSS97/SAP97, a neuronal isoform of discs large, regulates ethanol tolerance. *PLoS One* **7**, e48967 (2012).
32. Fruhmesser, A. *et al.* Disruption of EXOC6B in a patient with developmental delay, epilepsy, and a de novo balanced t(2;8) translocation. *Eur J Hum Genet* (2013).
33. Borsani, G. *et al.* Cytogenetic and molecular characterization of a de-novo t(2p;7p) translocation involving TNS3 and EXOC6B genes in a boy with a complex syndromic phenotype. *Eur J Med Genet* **51**, 292-302 (2008).
34. Liu, P. *et al.* Genome-wide association and fine mapping of genetic loci predisposing to colon carcinogenesis in mice. *Mol Cancer Res* **10**, 66-74 (2012).
35. Pinheiro, P.S. *et al.* GluR7 is an essential subunit of presynaptic kainate autoreceptors at hippocampal mossy fiber synapses. *Proc Natl Acad Sci U S A* **104**, 12181-6 (2007).
36. Minelli, A., Scassellati, C., Bonvicini, C., Perez, J. & Gennarelli, M. An association of GRIK3 Ser310Ala functional polymorphism with personality traits. *Neuropsychobiology* **59**, 28-33 (2009).

37. Ahmad, Y. *et al.* Association between the ionotropic glutamate receptor kainates3 (GRIK3) Ser310A1a polymorphism and schizophrenia in the Indian population. *World J Biol Psychiatry* **10**, 330-3 (2009).
38. Croteau-Chonka, D.C. *et al.* Genome-wide association study of anthropometric traits and evidence of interactions with age and study year in Filipino women. *Obesity (Silver Spring)* **19**, 1019-27 (2011).
39. Brown, C.E. & Sachs, A.B. Poly(A) tail length control in *Saccharomyces cerevisiae* occurs by message-specific deadenylation. *Mol Cell Biol* **18**, 6548-59 (1998).
40. Mukherjee, K. *et al.* Piccolo and bassoon maintain synaptic vesicle clustering without directly participating in vesicle exocytosis. *Proc Natl Acad Sci U S A* **107**, 6504-9 (2010).
41. Sullivan, P.F. *et al.* Genome-wide association for major depressive disorder: a possible role for the presynaptic protein piccolo. *Mol Psychiatry* **14**, 359-75 (2009).
42. Choi, K.H. *et al.* Gene expression and genetic variation data implicate PCLO in bipolar disorder. *Biol Psychiatry* **69**, 353-9 (2011).
43. Aragam, N., Wang, K.S. & Pan, Y. Genome-wide association analysis of gender differences in major depressive disorder in the Netherlands NESDA and NTR population-based samples. *J Affect Disord* **133**, 516-21 (2011).
44. Woudstra, S. *et al.* Piccolo genotype modulates neural correlates of emotion processing but not executive functioning. *Transl Psychiatry* **2**, e99 (2012).
45. Ahmed, M.Y. *et al.* Loss of PCLO function underlies pontocerebellar hypoplasia type III. *Neurology* **84**, 1745-50 (2015).
46. Woudstra, S. *et al.* Modulatory effects of the piccolo genotype on emotional memory in health and depression. *PLoS One* **8**, e61494 (2013).
47. Watase, K. *et al.* Motor discoordination and increased susceptibility to cerebellar injury in GLAST mutant mice. *Eur J Neurosci* **10**, 976-88 (1998).
48. Stoffel, W., Korner, R., Wachtmann, D. & Keller, B.U. Functional analysis of glutamate transporters in excitatory synaptic transmission of GLAST1 and GLAST1/EAAAC1 deficient mice. *Brain Res Mol Brain Res* **128**, 170-81 (2004).
49. Karlsson, R.M. *et al.* Assessment of glutamate transporter GLAST (EAAAT1)-deficient mice for phenotypes relevant to the negative and executive/cognitive symptoms of schizophrenia. *Neuropsychopharmacology* **34**, 1578-89 (2009).
50. de Vries, B. *et al.* Episodic ataxia associated with EAAAT1 mutation C186S affecting glutamate reuptake. *Arch Neurol* **66**, 97-101 (2009).
51. Jen, J.C., Wan, J., Palos, T.P., Howard, B.D. & Baloh, R.W. Mutation in the glutamate transporter EAAAT1 causes episodic ataxia, hemiplegia, and seizures. *Neurology* **65**, 529-34 (2005).
52. Lorenzetti, D. *et al.* A transgenic insertion on mouse chromosome 17 inactivates a novel immunoglobulin superfamily gene potentially involved in sperm-egg fusion. *Mamm Genome* **25**, 141-8 (2014).
53. Song, S. *et al.* E2-25K/Hip-2 regulates caspase-12 in ER stress-mediated Abeta neurotoxicity. *J Cell Biol* **182**, 675-84 (2008).
54. Boger, C.A. *et al.* Association of eGFR-Related Loci Identified by GWAS with Incident CKD and ESRD. *PLoS Genet* **7**, e1002292 (2011).
55. Kottgen, A. *et al.* New loci associated with kidney function and chronic kidney disease. *Nat Genet* **42**, 376-84 (2010).
56. Kottgen, A. *et al.* Genome-wide association analyses identify 18 new loci associated with serum urate concentrations. *Nat Genet* **45**, 145-54 (2013).

Supplementary Material

Supplementary Results

A majority of protein coding genes are essential for rat reproduction

Rats harboring *Sleeping Beauty* genetraps within 18 distinct protein coding genes were crossed with wildtype rats to generate heterozygous breeding pairs (Supplementary Table 1). Heterozygous breeding pairs were successfully generated for 17 of 18 mutant alleles (Supplementary Table 2). Only female rats heterozygous for a genetraps (*gt*) mutation within intron 5 of *Dlg1* (i.e. *Dlg1*^{wt/*gt*}) failed to transmit their mutation more than one generation (Supplementary Table 2). Male and female heterozygotes for the remaining 17 strains were paired to generate homozygous mutant strains. Mean litter sizes produced by crossing heterozygotes for 12 of 18 mutant strains were not significantly different from wildtype breeder pairs (Supplementary Table 2). Heterozygous breeding pairs from each *Alk3*^{wt/*gt*}, *Abca13*^{wt/*gt*}, *Gsg1l*^{wt/*gt*}, *Slc1a3*^{wt/*gt*}, *Ube2k*^{wt/*gt*} and *Zmynd8*^{wt/*gt*} strains consistently reproduced, but their litter sizes were significantly smaller compared to wildtype (Supplementary Table 2).

Mean litter sizes generated by crossing homozygous mutant *Abca13*^{gt/*gt*}, *Fstl5*^{gt/*gt*}, *Grik3*^{gt/*gt*} (males only), *Gsg1l*^{gt/*gt*}, *Rg22*^{gt/*gt*} and *Ube2q2*^{gt/*gt*} rats to wildtype rats appeared normal (Supplementary Table 2). *Grik3*^{gt/*gt*} and *Pan3*^{gt/*gt*} females were fertile, but produced smaller litters after crossing with wildtype males (p=0.003). In contrast, homozygous genetraps mutations in *Btrc*, *Ube2k*, *Pan3*, *Atg13*, *Spaca6* and *Pclo* blocked reproduction by males during peak reproductive age (Fig. 2a and Supplementary Table 2); in this group, *Ube2k*^{gt/*gt*}, *ATG13*^{gt/*gt*} and *Pclo*^{gt/*gt*} mutant females also did not reproduce (Supplementary Table 2). *Btrc*^{gt/*gt*} and *Pan3*^{gt/*gt*} female rats were fertile (Supplementary Table 2). Despite their inability to reproduce, *Ube2k*^{gt/*gt*} and *Pclo*^{gt/*gt*} female rats developed oocytes (Supplementary Fig. 2c).

Spermatogenic cells in *Ube2k^{gt/gt}* rats arrested early during meiosis I postnatally and in young adults (Fig. 2a, b), and progressively regressed to arrest at pre-meiotic steps during adulthood (Supplementary Fig. 2a). *Ube2k^{gt/gt}* mutant rats were also significantly smaller in body size compared to wildtype littermates ($p < 0.001$) (Supplementary Table 3). Spermatogenic cells in *Pan3^{gt/gt}* mutant rats arrested during spermatid elongation (Fig. 2a, b and Supplementary Fig. 2). Rats with heterozygous genetrapp mutations predicted to disrupt expression of *Alk3*, *Exoc6b*, *Slc1a3*, *Tmx4* and *Zmynd8* reproduced, but demonstrated reduced Mendelian rates towards generation of homozygous mutant progeny (Supplementary Table 2). Homozygous genetrapp mutations in these later genes were classified as embryonic lethal. Heterozygous *Alk3^{wt/gt}* mutants also displayed a reduced growth phenotype compared to wildtype littermates (D24 mean body weight, *Alk3^{wt/wt}* 69.4 ± 1.7 g, *Alk3^{wt/gt}* 58.6 ± 2.7 g, $n = 36$ total pups, 3 litters; \pm SD $p < 0.001$).

Comparative Analysis of Rat Genetrapp Mutations

Phenotypes transmitted by rat genetrapp mutations were compared across different species, as follows:

Alk3^{gt/gt}: Distinct from reduced growth in *Alk3^{wt/gt}* rats, heterozygous mutant *Alk3* mice display normal growth¹. Consistent with current Mendelian rates of germline transmission in *Alk3^{gt/gt}* rats, homozygous mutations are embryonic lethal in mice, which caused defects stemming from epiblast development¹. In flies, homozygous *Alk3* mutations disrupt wing patterning^{2,3}, as well as germline stem cell development⁴. In humans, recessive *Alk3* mutations are linked to Juvenile and Hamartomatous Polyposis syndromes⁵⁻⁷.

Atg13^{gt/gt}: Analogous to reduced postnatal lifespan in *Atg13^{gt/gt}* rats, disrupting *Atg13* expression induces nutrient dependent senescence in plants⁸ and reduces lifespan in *C. Elegans*⁹. However, *Atg13* loss-of-function mutations are embryonic lethal in flies¹⁰. Spermatozoa from the cauda

epididymus of *Atg13^{gt/gt}* mutants appeared immotile when analyzed between 60 and 120 days of age. In addition to the reduced sperm count (Supplementary Fig. 3), flagella from *Atg13^{gt/gt}* cauda epididymal spermatozoa were largely immotile, and often displayed detached heads and tails (not shown). Notably, the health decline of *Atg13^{gt/gt}* rats was observed ~90-100 days of age, which is only 2-3 weeks after rats reach reproductive age (*also see* Discussion section in manuscript). Livers and kidneys in *Atg13^{gt/gt}* rats appeared abnormal (Fig. 3c). In the liver, occasional vacuolated cells were scattered throughout histological sections. These cells contained small spherical vacuoles consistent with accumulation of triglycerides (Supplementary Fig. 4a).

All kidneys examined displayed marked glomerulonephritis and moderate tubulointerstitial disease (Fig. 3d). Glomeruli were diffusely altered by membranous change; glomerular tufts were thickened by expansion of mesangium and thickening of basement membranes (Figs. 3d and Supplementary 4a). Bowman's capsule was also thickened, and in some glomeruli, Bowman's space was filled with crescentic proliferations (Fig. 3d). Numerous tubules were dilated and contained protein-rich casts (Fig. 3d and Supplementary 4a). PAS and Trichrome stains indicated that hyaline deposits consisted of both PAS positive material and collagen. There was scattered interstitial inflammation consisting of lymphocytes and plasma cells; and, small numbers of tubules contained neutrophils (Supplementary Fig. 4a).

Btrc^{gt/gt}: Distinct from *Btrc^{gt/gt}* rats, *Btrc*-deficient mice develop sufficient numbers of functional sperm to reproduce by natural mating, but their litter sizes are significantly smaller when compared to wildtype¹¹. Reduced litter size in *Btrc*-deficient mice is thought to be caused by abnormal meiotic divisions in some segments of seminiferous tubules, which would reduce total sperm counts¹¹. In contrast, normal elongating spermatids were not observed in *Btrc^{gt/gt}* rats (Supplementary Figs. 2a, b). And, most seminiferous tubule cross sections from *Btrc^{gt/gt}* rats

contained only Sertoli cells and type A spermatogonia (~85%, n=2 rats; Supplementary Figs. 2a, b). A lower percent tubules (~15%) uniquely contained spermatocytes, round spermatids and abnormal elongating spermatids (Supplementary Fig. 2b). *Btrc* encodes an E3 ubiquitin ligase, and analogous to effects observed here on spermatogonial development, *Btrc* is essential for proper differentiation of neural progenitor cells^{12,13}. In *Xenopus*, dominant negative *Btrc* constructs missing the N-terminal F-box domain, but retaining its C-terminal WD40 repeat domains induce duplication of the dorsal tadpole axis¹⁴. Interestingly, absolute testis weights in *Btrc*^{gt/gt} rats are greater than in *Ube2k*^{gt/gt} rats (Supplementary Table 3). This is despite the fact that spermatogenesis arrested at later developmental steps in *Ube2k*^{gt/gt} compared to a vast majority of tubules in *Btrc*^{gt/gt} rats (Supplementary Fig. 2a, b). However, it should be noted that *Ube2k*^{gt/gt} male rats are between 30-40% smaller in body size compared to *Btrc*^{gt/gt} or wildtype rats (Supplementary Table 3).

Dlg1^{gt/gt}: Female *Dlg1*^{wt/gt} rats were consistently observed in back cage corners, remained socially isolated following pairing with respective wildtype males, did not produce offspring, and appeared otherwise healthy. In contrast to the reproduction deficiency of heterozygous female *Dlg1*^{wt/gt} rats, heterozygous *Dlg1* mutant female mice breed normally¹⁵⁻¹⁸. However, homozygous *Dlg1* mouse mutants display overt urogenital, craniofacial and lymphocyte developmental defects linked to shorter postnatal lifespan¹⁵⁻¹⁸. Here, a similar *Dlg1*^{wt/gt} mutation with respect to deleted domains was transmitted through the rat male germline by spermatogonial stem cells transplanted into the somatic background of wildtype founder rats. This, and the abnormal breeding by female *Dlg1*^{wt/gt} rats, is consistent with species-dependent dominant and recessive somatic defects in rats and mice, respectively.

Pan3^{gt/gt}: Like in *Pan3*^{gt/gt} rats, yeast harboring *Pan3* deletion mutations are viable; however, effects on yeast gametogenesis were not reported. Here, spermatogenic cells in *Pan3*^{gt/gt} mutant rats arrested during spermatid elongation (Fig. 2a, b and Supplementary Fig. 2a). One hypothesis is that the *Pan3* mutation in rats disrupts control over mRNA stability and/or translation during steps critical for spermatid elongation. This is because PAN3 functions as a co-activator of the PAN2 polyA specific ribonuclease subunit homolog¹⁹.

Pclo^{gt/gt}: Unlike effects of *Pclo*^{gt/gt} in rats, current strains of mutant *Pclo* mice reproduce normally and do not display neurological and/or behavioral phenotypes (*also see* Discussion section in manuscript)²⁰.

Slc1a3^{gt/gt}: Unlike embryonic lethality in *Slc1a3*^{gt/gt} rats, adult homozygous mutant *Slc1a3* mice are viable and reproduce normally, but display measurable behavioral phenotypes²¹⁻²³.

Spaca6^{gt/gt}: Here, based on normal testis histology, epididymal sperm counts and mating behavior, the rat *Spaca6*^{gt/gt} mutation is predicted to disrupt spermatozoan function at some point within the female reproductive tract. In mutant mice lacking a 11kb region of chromosome 17, *Spaca6* was initially implicated in gamete membrane fusion²⁴.

Ube2k^{gt/gt}: Unlike in rats, male and female mice harboring a very similar *Ube2k*^{gt/gt} mutation reproduced normally, had normal body weights and did not show age related myogenic/motor neuron-like phenotypes²⁵. *Ube2k* encodes the ubiquitin conjugating enzyme E2-25k²⁶.

Zmynd8^{gt/gt}: Distinct from embryonic lethal effects of *Zmynd8^{gt/gt}* rats, *Zmynd8*-deficient mice are reported as postnatal lethal, surviving to postnatal day 14 (IMPC database. <http://mousephenotype.org/>)²⁷.

Supplementary Figure Legends

Supplementary Fig. 1. Rat gene-trap mutations screened for reproduction effects.

Schematic of mutant proteins predicted to be produced in rat strains harboring intronic insertions of *Sleeping Beauty* β -geo genetraps²⁸. Exon sequences predicted to be excluded (Δ) from mRNAs encoding truncated polypeptides (aa) generated by imposed splicing to the genetraps are shown below respective wildtype proteins for 17 of the 18 mutant rat strains screened for effects on reproduction. A transposon insertion within intron 2 of *Rgs22* is not shown, and is not predicted to truncate the RGS22 open reading frame due to its genetraps cassette inserting in the 3' to 5' orientation to *Rgs22*. See Supplementary Table S1 for full amino acid sequences of the predicted mutant proteins, which contain additional epitopes of either 3, 24 or 1319 (β -GEO) amino acids derived from the genetraps construct.

TM, Transmembrane domain; AAA, ATPase Associated with a variety of cellular activities; GS, GS Motif; L27, domain in receptor targeting proteins Lin-2 and Lin-7; MG-PEST, Polyubiquitination (PEST) N-terminal domain of MAGUK; PDZ, Domain present in PSD-95; β -TrCP, D domain of beta-TrCP; FBOX, A Receptor for Ubiquitination Targets; Dlg, and ZO-1/2; SH3, Src homology 3 domain; GuKc, Guanylate kinase homologue; CC, coil coil region; KAZAL, Kazal type serine protease inhibitors; IGc2, Immunoglobulin C-2 Type; Lg-Ch-Bd, Ligated ion channel L-glutamate- and glycine-binding site; ZnF_C3H1, Zinc Finger Domain; STYKc, Protein kinase; unclassified specificity; C2, Protein kinase C conserved region 2 (CalB); UBCC, Ubiquitin-conjugating enzyme E2, catalytic domain homologue; UBA, Ubiquitin associated domain; RWD, domain in RING finger and WD repeat containing proteins and DEXDc-like helicases subfamily related to the UBCC domain; PHD, PHD zinc finger; BROMO, bromo domain; PWWP, domain with conserved PWWP motif.

Supplementary Fig. 2. Gametogenesis defects in mutant rats

- a) H&E stained testis sections from wildtype and respective homozygous mutant rats. Scale bar, 100 μm .
- b) Immunofluorescence labeling of cells in wildtype and mutant *Btrc*^{gt/gt} rat testis sections using an antibody to γH2AX and Hoechst 43332 dye. Scale bar, 100 μm .
- c) H&E stained ovarian sections from wildtype and respective homozygous mutant rats. Scale bar, 100 μm .

Supplementary Fig. 3. Sperm counts correlate with infertility in mutant rats

Mean testis weight in Grams (tan bars; left y-axis) and epididymal sperm counts plotted as Millions/Rat (blue bars; right y-axis) from respective homozygous mutant rat strains ($\pm\text{SEM}$, n=4-6 rats/strain). Measurements taken between postnatal days 120-180. Caudal epididymal spermatozoa from *Spaca6*^{gt/gt} (n=6) and *Pclo*^{gt/gt} (n=4) rats displayed similar basal activity compared to wildtype. *Atg13*^{gt/gt} (n=3) and *Btrc*^{gt/gt} (n=2) rat caudal epididymal sperm were immotile and morphologically abnormal compared to wildtype.

Supplementary Fig. 4. Pathology in *Atg13* mutant rats

- a) Hematoxylin and Eosin stained sections illustrating fatty liver (Left) and pyelonephritis (Right) in *Atg13*^{gt/gt} rats. Note: Renal tubules (Right) are dilated and filled with protein-rich filtrate (asterisks) and numerous neutrophils (arrows). Scale bar, 50 μm .
- b) Western blot analysis of autophagy marker proteins (ATG13, p62) and TUBA1a in wildtype and *Atg13*^{gt/gt} primary rat embryonic fibroblast cultures shown in Fig. 3f. Cultures were pretreatment with rapamycin (100 nM), ammonium chloride (3 mM) and/or Bafilomycin A1 (3 nM). The loading control blot shown for TUBA1a is the same blot shown in Fig. 3f.

Supplementary Fig 5. Synaptic transmission is downregulated in the brain of mutant *Piccolo* rats.

Scatter plots of Log2-fold change in transcript abundance versus mean relative expression levels (Log2 FPKM values) in *Pclo*^{wt/gt} and *Pclo*^{gt/gt} brain (left panels) and testis (right panel) compared to wildtype. Differentially expressed genes (DEGS) are shown in red (increased abundance) and blue (decreased abundance), respectively (log2-fold change >1 or <-1; FDR < 0.05). Note that the *Pclo* mutation caused more DEGS in the brain vs testis transcriptome. *Gabra6* and *Gabrg3*, the most significantly affected genes in brain and testis, respectively, have similar expression pattern in the homozygous and heterozygous mutant rats.

Supplementary Fig. 6. Gonadotropin Releasing Hormone (GnRH) pathway genes are downregulated in the brain of mutant *Piccolo* rats.

- a) Panther Pathway finder analysis on down regulated Hormonal Secretion GO: gene sets identify GnRH signaling pathway as a prominent cluster of down-regulated genes in *Pclo*^{gt/gt} rat brain versus *Pclo*^{wt/wt} rat brains.
- b) Coronal brain sections from *Pclo*^{wt/wt} or *Pclo*^{gt/gt} postnatal day 100 rat brains immuno-stained with GnRH antibodies. Lower panel, 5x magnification of the preoptic area (POA) located in the boxed area reveal presence of somata and GnRH positive neuron processes flanking the third ventricle. Scale bars = upper panel 0.3 cm, lower panel 1000 μ m.
- c) Relative abundance of GnRH signaling pathway GO: gene set components in *Pclo*^{gt/gt} rat brains versus *Pclo*^{wt/wt} rat brains.
- d) KEGG pathway analysis predicts downregulation of GnRH signaling pathway in *Pclo*^{gt/gt} rat brains versus *Pclo*^{wt/wt} rat brains.

Supplementary Fig. 7. Misregulated gene networks downstream of GnRH Signaling.

a) Relative abundance of G-Protein Coupled Receptors (GPCRs) involved in conducting GnRH signaling on gonadotropes GO: gene set components in *Pclo^{gt/gt}* (KO) and *Pclo^{wt/gt}* (Het) rat brains versus *Pclo^{wt/wt}* (WT) rat brains.

b) Relative abundance of Calcium Signaling GO: gene set components in *Pclo^{gt/gt}* (KO) rat brains versus *Pclo^{wt/wt}* (WT) rat brains.

c) Relative abundance of Gonadotropin Regulated Genes in *Pclo^{gt/gt}* (KO) rat testes versus *Pclo^{wt/wt}* (WT) rat testes. FSH, Follicle Stimulating Hormone; LH, Luteinizing Hormone, T, Testosterone.

Supplementary References

1. Mishina, Y., Suzuki, A., Ueno, N. & Behringer, R.R. Bmpr encodes a type I bone morphogenetic protein receptor that is essential for gastrulation during mouse embryogenesis. *Genes Dev* **9**, 3027-37 (1995).
2. Burke, R. & Basler, K. Dpp receptors are autonomously required for cell proliferation in the entire developing Drosophila wing. *Development* **122**, 2261-9 (1996).
3. Singer, M.A., Penton, A., Twombly, V., Hoffmann, F.M. & Gelbart, W.M. Signaling through both type I DPP receptors is required for anterior-posterior patterning of the entire Drosophila wing. *Development* **124**, 79-89 (1997).
4. Xia, L. *et al.* The Fused/Smurf complex controls the fate of Drosophila germline stem cells by generating a gradient BMP response. *Cell* **143**, 978-90 (2010).
5. Kim, I.J. *et al.* Identification of a novel BMPR1A germline mutation in a Korean juvenile polyposis patient without SMAD4 mutation. *Clin Genet* **63**, 126-30 (2003).
6. Nieminen, T.T. *et al.* BMPR1A mutations in hereditary nonpolyposis colorectal cancer without mismatch repair deficiency. *Gastroenterology* **141**, e23-6 (2011).
7. Zhou, X.P. *et al.* Germline mutations in BMPR1A/ALK3 cause a subset of cases of juvenile polyposis syndrome and of Cowden and Bannayan-Riley-Ruvalcaba syndromes. *Am J Hum Genet* **69**, 704-11 (2001).
8. Suttangkakul, A., Li, F., Chung, T. & Vierstra, R.D. The ATG1/ATG13 protein kinase complex is both a regulator and a target of autophagic recycling in Arabidopsis. *Plant Cell* **23**, 3761-79 (2011).
9. Tian, E., Wang, F., Han, J. & Zhang, H. epg-1 functions in autophagy-regulated processes and may encode a highly divergent Atg13 homolog in *C. elegans*. *Autophagy* **5**, 608-15 (2009).
10. Chang, Y.Y. & Neufeld, T.P. An Atg1/Atg13 complex with multiple roles in TOR-mediated autophagy regulation. *Mol Biol Cell* **20**, 2004-14 (2009).
11. Guardavaccaro, D. *et al.* Control of meiotic and mitotic progression by the F box protein beta-Trcp1 in vivo. *Dev Cell* **4**, 799-812 (2003).
12. Guardavaccaro, D. *et al.* Control of chromosome stability by the beta-TrCP-REST-Mad2 axis. *Nature* **452**, 365-9 (2008).
13. Westbrook, T.F. *et al.* SCFbeta-TRCP controls oncogenic transformation and neural differentiation through REST degradation. *Nature* **452**, 370-4 (2008).
14. Liu, C. *et al.* beta-Trcp couples beta-catenin phosphorylation-degradation and regulates Xenopus axis formation. *Proc Natl Acad Sci U S A* **96**, 6273-8 (1999).
15. Caruana, G. & Bernstein, A. Craniofacial dysmorphogenesis including cleft palate in mice with an insertional mutation in the discs large gene. *Mol Cell Biol* **21**, 1475-83 (2001).
16. Rivera, C. *et al.* Cell-autonomous requirements for Dlg-1 for lens epithelial cell structure and fiber cell morphogenesis. *Dev Dyn* **238**, 2292-308 (2009).
17. Mahoney, Z.X. *et al.* Discs-large homolog 1 regulates smooth muscle orientation in the mouse ureter. *Proc Natl Acad Sci U S A* **103**, 19872-7 (2006).
18. Iizuka-Kogo, A., Ishida, T., Akiyama, T. & Senda, T. Abnormal development of urogenital organs in Dlg1-deficient mice. *Development* **134**, 1799-807 (2007).
19. Brown, C.E. & Sachs, A.B. Poly(A) tail length control in *Saccharomyces cerevisiae* occurs by message-specific deadenylation. *Mol Cell Biol* **18**, 6548-59 (1998).
20. Mukherjee, K. *et al.* Piccolo and bassoon maintain synaptic vesicle clustering without directly participating in vesicle exocytosis. *Proc Natl Acad Sci U S A* **107**, 6504-9 (2010).
21. Karlsson, R.M. *et al.* Assessment of glutamate transporter GLAST (EAAT1)-deficient mice for phenotypes relevant to the negative and executive/cognitive symptoms of schizophrenia. *Neuropsychopharmacology* **34**, 1578-89 (2009).

22. Stoffel, W., Korner, R., Wachtmann, D. & Keller, B.U. Functional analysis of glutamate transporters in excitatory synaptic transmission of GLAST1 and GLAST1/EAAC1 deficient mice. *Brain Res Mol Brain Res* **128**, 170-81 (2004).
23. Watase, K. *et al.* Motor discoordination and increased susceptibility to cerebellar injury in GLAST mutant mice. *Eur J Neurosci* **10**, 976-88 (1998).
24. Lorenzetti, D. *et al.* A transgenic insertion on mouse chromosome 17 inactivates a novel immunoglobulin superfamily gene potentially involved in sperm-egg fusion. *Mamm Genome* **25**, 141-8 (2014).
25. Song, S. *et al.* E2-25K/Hip-2 regulates caspase-12 in ER stress-mediated Abeta neurotoxicity. *J Cell Biol* **182**, 675-84 (2008).
26. Chen, Z.J., Niles, E.G. & Pickart, C.M. Isolation of a cDNA encoding a mammalian multiubiquitinating enzyme (E225K) and overexpression of the functional enzyme in *Escherichia coli*. *J Biol Chem* **266**, 15698-704 (1991).
27. Nijman, I.J. *et al.* Mutation discovery by targeted genomic enrichment of multiplexed barcoded samples. *Nat Methods* **7**, 913-5 (2010).
28. Izsvak, Z. *et al.* Generating knockout rats by transposon mutagenesis in spermatogonial stem cells. *Nat Methods* **7**, 443-5 (2010).
29. Petrulis, A. Chemosignals and hormones in the neural control of mammalian sexual behavior. *Front Neuroendocrinol* **34**, 255-67 (2013).
30. Sokolowski, K. & Corbin, J.G. Wired for behaviors: from development to function of innate limbic system circuitry. *Front Mol Neurosci* **5**, 55 (2012).