

Fig. S1. Changes in ribosome composition by exposure to (PR) $\mathbf{2 0}_{20}$ peptides. a, STRING analysis (https://string-db.org) of the protein interaction networks found between the proteins identified in the purification of ribosomes from RPS9 ${ }^{\mathrm{SBP}}$-expressing HeLa cells. The node containing 60S and 40S factors (RPLs and RPSs) is indicated (arrow). The panel illustrates the ribosome composition from (PR) $)_{20}$-untreated cells. b, Protein levels of RPL factors in ribosomes purified from Hela RPS9 ${ }^{\text {SBP }}$ cells exposed to $10 \mu \mathrm{M}$ of (PR $)_{20}$ for 16 h , as identified by LC-MS/MS. c, Protein levels of RPL factors in the input extracts used for ribosome purification from Hela RPS9 ${ }^{\text {SBP }}$ cells exposed to $10 \mu \mathrm{M}$ of $(\mathrm{PR})_{20}$ for 16 h , as identified by LC-MS/MS.


Fig. S2. (PR) $\mathbf{2 0}_{20}$ peptides impair the assembly of $80 S$ ribosome particles on mRNA. a, Representative polysome profiles obtained from HeLa cells untreated or treated with $10 \mu \mathrm{M}$ of (PR)20 for 16 h . The presence of halfmers is indicated (arrows). $\mathbf{b}$, Electron microscopy images from purified 40 S and 60 S ribosomal complexes ( 1 pmol each) assembled in vitro in the presence of $\mathrm{MgCl}_{2}$, and in the presence or absence of 5 pmol of $(\mathrm{PR})_{20}$. Assembled 80S particles are indicated (red arrows). c, Quantification of 80 S particles identified in (d) ( $\mathrm{n}=1000$ ) in nonassembly ( 1 mM MgCl 2 ) or assembly ( 5 mM MgCl 2 ) conditions.


Fig. S3. Effects of (PR) $\mathbf{2 0}_{\mathbf{2 0}} \mathbf{o n} \mathbf{m R N A}$ translation. a, In vitro translation of 100 ng of luciferase mRNA (quantified by luciferase activity) in the presence of increasing doses of $(\mathrm{PR})_{20}$. $\mathbf{b}$, In vitro translation of 100 ng of luciferase mRNA in the presence or absence of $0.5 \mu \mathrm{M}(\mathrm{PR})_{20}$. Translation products were labeled with $\left[{ }^{35} \mathrm{~S}\right]$-Met/Cys and analyzed by SDS-PAGE and autorradiography. c, In vitro translation of 50 ng of luciferase mRNA was allowed for $15^{\prime}$ and stopped in ice. After inhibition of new translation initiation with lactimidomycin for $10^{\prime}$, translation was allowed in the presence or absence $0.5 \mu \mathrm{M}(\mathrm{PR})_{20}$ for the indicated times. d, In vitro translation of 100 ng of luciferase mRNA with different $5^{\prime}$ UTR lengths in the presence or absence of $0.5 \mu \mathrm{M}(\mathrm{PR})_{20}$. e, In vitro translation of 100 ng of luciferase mRNA in the presence or absence of $0.5 \mu \mathrm{M}(\mathrm{PR})_{20}$. In the right two columns, the mRNA was extracted from a translation reaction done in the presence of the DPR, and subsequently used in a new translation reaction performed in the absence of $(P R)_{20}$. *, $\mathrm{p}<0.05$.


Fig. S4. DNA- and RNA-binding by (PR) $\mathbf{2 0}_{20}$ peptides. (a, b) Binding of (PR) $)_{20}$ to DNA (a) or RNA (b) as examined by EMSA. Previously annealed 19 bp Cy3-dsDNA or Cy3-RNA oligonucleotides $(0.2 \mu \mathrm{M})$ were incubated with increased doses of $\mathrm{PR}_{20}$ for $10^{\prime}$. The images show the disappearance of free dsDNA/dsRNA and non-annealed ssDNA/ssRNA oligonucleotides, which occurs concomitant with the accumulation of DNA- or RNA-(PR $)_{20}$ complexes on the loading well. The sharp transition from unbound to unbound fractions is indicative of cooperative binding in all cases. A quantification from these assays is shown in Fig. a.


Fig. S5. Effect of heparin on the nucleolar accumulation of (PR) $\mathbf{2 0}_{\mathbf{2 0}}$. a, Immunofluorescence of HA-(PR $)_{20}$ (green) and the nucleolar factor UBF1 (red) in U2OS cells treated with $7.5 \mu \mathrm{MHA}$ $(\mathrm{PR})_{20}$ alone or together with $0.5 \mu \mathrm{M}$ heparin for 8 h . $\mathbf{b}$, HTM-mediated quantification of the nucleolar HA-(PR) $)_{20}$ intensity from U2OS cells treated as in (a).

Table S1. Proteins with reduced levels on ribosomal fractions purified from $(\mathrm{PR})_{20}$-treated HeLa-RPS $9^{\text {SBP }}$ cells.

| Gene Name | $\log _{2}\left((\mathrm{PR})_{20} /\right.$ control $)$ |
| :--- | ---: |
| PSME3 | -3.61 |
| H1FX | -1.94 |
| RPL27A | -1.76 |
| RPLP2 | -1.71 |
| BRIX1 | -1.52 |
| GTPBP4 | -1.35 |
| RPL32 | -1.32 |
| KRR1 | -1.22 |
| NAP1L4 | -1.21 |
| HP1BP3 | -1.20 |
| RPL3 | -0.99 |
| RPL14 | -0.87 |
| RPL10A | -0.70 |
| RPL6 | -0.66 |
| UTP14A | -0.65 |
| RPL9 | -0.63 |

Table S2. Proteins that show statistically significant reduced levels on chromatin after treatment of U2OS cells with (PR $)_{20}(20 \mu \mathrm{M})$ or PROTAMINE $(30 \mu \mathrm{M})$.

| Gen Symbol | Log $_{2}\left((\mathrm{PR})_{20} /\right.$ control $)$ | $\log _{2}($ Protamine/control $)$ |
| :--- | ---: | ---: |
| FURIN | -2.45 | -3.61 |
| POTEJ | -2.05 | -2.15 |
| POTEKP | -1.72 | -1.63 |
| TMEM126B | -1.40 | -1.19 |
| POTEF | -1.38 | -1.20 |
| HIST1H1C | -1.33 | -0.61 |
| FARS2 | -1.28 | -0.75 |
| HIST1H1A | -1.27 | -0.46 |
| SNX5 | -1.06 | -0.82 |
| H1F0 | -1.05 | -0.39 |
| PITPNA | -1.02 | -0.60 |
| PDK2 | -1.01 | -0.64 |
| GLTSCR2 | -1.01 | -0.42 |
| EEF1B2 | -1.01 | -0.86 |
| EEF1G | -0.98 | -0.75 |
| GAPDH | -0.97 | -0.80 |
| NRF1 | -0.96 | -0.90 |
| PSMF1 | -0.93 | -0.42 |
| AURKC | -0.92 | -0.87 |
| CORO1B | -0.88 | -0.47 |
| EBAG9 | -0.87 | -0.64 |
| APEX1 | -0.86 | -0.70 |
| EEF1D | -0.83 | -0.69 |
| ALDOC | -0.82 | -0.45 |
| REPIN1 | -0.82 | -0.54 |
| H2AFY | -0.82 | -0.43 |
| RXRB | -0.75 | -0.50 |
| HMGA1 | -0.75 | -0.68 |
| TPT1 | -0.73 | -0.65 |
| RPUSD4 | -0.72 | -0.87 |
| DNAJC15 | -0.70 | -0.40 |
| RING1 | -0.70 | -0.57 |
| PCBP3 | -0.67 | -0.41 |
|  |  |  |


| HMGA2 | -0.66 | -0.51 |
| :--- | ---: | ---: |
| SLC25A40 | -0.65 | -0.41 |
| HN1L | -0.65 | -0.40 |
| MEN1 | -0.65 | -0.40 |
| MSN | -0.64 | -0.58 |
| HDGF | -0.62 | -0.44 |
| HCFC1 | -0.62 | -0.52 |
| TGIF2LX | -0.62 | -0.52 |
| HMGB1;HMGB1P1 | -0.62 | -0.50 |
| LRRC57 | -0.60 | -0.49 |
| FLNA | -0.60 | -0.50 |
| RANBP1 | -0.60 | -0.59 |
| NCKIPSD | -0.59 | -0.61 |
| EZR | -0.58 | -0.51 |
| CMSS1 | -0.58 | -0.35 |
| LANCL2 | -0.58 | -0.58 |
| CSRP2 | -0.58 | -0.48 |
| TMF1 | -0.58 | -0.44 |
| EEF1A2 | -0.56 | -0.54 |
| TARS | -0.56 | -0.45 |
| CDYL | -0.56 | -0.75 |
| DIDO1 | -0.55 | -0.44 |
| AHNAK | -0.55 | -0.48 |
| ACAP2 | -0.55 | -0.39 |
| SYNGR3 | -0.55 | -0.74 |
| PALM2 | -0.55 | -0.60 |
| ARHGAP17 | -0.55 | -0.48 |
| ID1 | -0.55 | -0.57 |
| CCAR1 | -0.54 | -0.39 |
| RPL18 | -0.54 | -0.40 |
| COPS7A | -0.54 | -0.38 |
| CCDC50 | -0.53 | -0.51 |
| ZBTB10 | -0.53 | -0.53 |
| CCNC | -0.53 | -0.88 |
| SFSWAP | -0.52 | -0.38 |
| SLC43A3 | -0.52 | -0.60 |
| WASF2 | -0.52 | -0.45 |
| SPR | -0.61 |  |
| GAP43 | -1 | -1 |


| FSCN1 | -0.51 | -0.40 |
| :---: | :---: | :---: |
| VPS39 | -0.51 | -0.57 |
| FUS | -0.50 | -0.46 |
| SARS | -0.50 | -0.46 |
| FIP1L1 | -0.49 | -0.38 |
| DIAPH2 | -0.49 | -0.67 |
| CSTF2 | -0.49 | -0.40 |
| SCAF4 | -0.49 | -0.36 |
| CBX8 | -0.48 | -0.47 |
| PRPF38B | -0.48 | -0.35 |
| DCUN1D1 | -0.47 | -0.61 |
| DDX31 | -0.47 | -0.45 |
| PDLIM7 | -0.47 | -0.41 |
| RECQL | -0.47 | -0.36 |
| FNBP4 | -0.47 | -0.40 |
| EHD3 | -0.46 | -0.38 |
| SCAF1 | -0.46 | -0.37 |
| MRGBP | -0.46 | -0.54 |
| MAVS | -0.45 | -0.38 |
| CORO1C | -0.45 | -0.55 |
| MINA | -0.45 | -0.48 |
| COPS8 | -0.45 | -0.74 |
| TCERG1 | -0.45 | -0.42 |
| NEDD1 | -0.44 | -0.59 |
| SEPT9 | -0.44 | -0.43 |
| HDGFRP3 | -0.44 | -0.36 |
| FAHD1 | -0.44 | -0.73 |
| FKBP3 | -0.43 | -0.42 |
| PA2G4 | -0.43 | -0.42 |
| DDX42 | -0.43 | -0.40 |
| CASK | -0.42 | -0.69 |
| GABPA | -0.41 | -0.36 |
| MAD2L1BP | -0.40 | -0.40 |
| ITPRIP | -0.40 | -0.67 |
| EHD4 | -0.40 | -0.36 |
| NARS | -0.39 | -0.41 |
| EEF1A1;EEF1A1P5 | -0.39 | -0.47 |
| WDR33 | -0.39 | -0.42 |
| DEK | -0.39 | -0.44 |


| TBP | -0.39 | -0.36 |
| :--- | ---: | ---: |
| DOK1 | -0.39 | -0.35 |
| IFIT2 | -0.39 | -0.72 |
| UBAP2L | -0.38 | -0.49 |
| SEPT11 | -0.38 | -0.36 |
| RSU1 | -0.37 | -0.38 |
| NACA | -0.37 | -0.51 |
| ORC1 | -0.37 | -0.42 |
| CCNL1 | -0.37 | -0.37 |
| CEP97 | -0.37 | -0.45 |
| UBE2I | -0.36 | -0.42 |
| VPS13C | -0.36 | -0.41 |
| PRDX1 | -0.36 | -0.40 |
| DR1 | -0.36 | -0.64 |
| BCL7B | -0.35 | -0.36 |
| KDM2A | -0.35 | -0.41 |

