

### Supplementary Figure S1: Schematics of experiments from Figure 1 and Figure 4 with edit examples

Schematics on the left follow color schemes described in the main figures. Templates are shown on top and *gtbp-1* locus is shown below. Lines between templates and *gtbp-1* locus represent strand invasion/template switching events inferred from the type of edits obtained in each experiment. Images on the right show method to identify edit (PCR amplification or GFP expression).

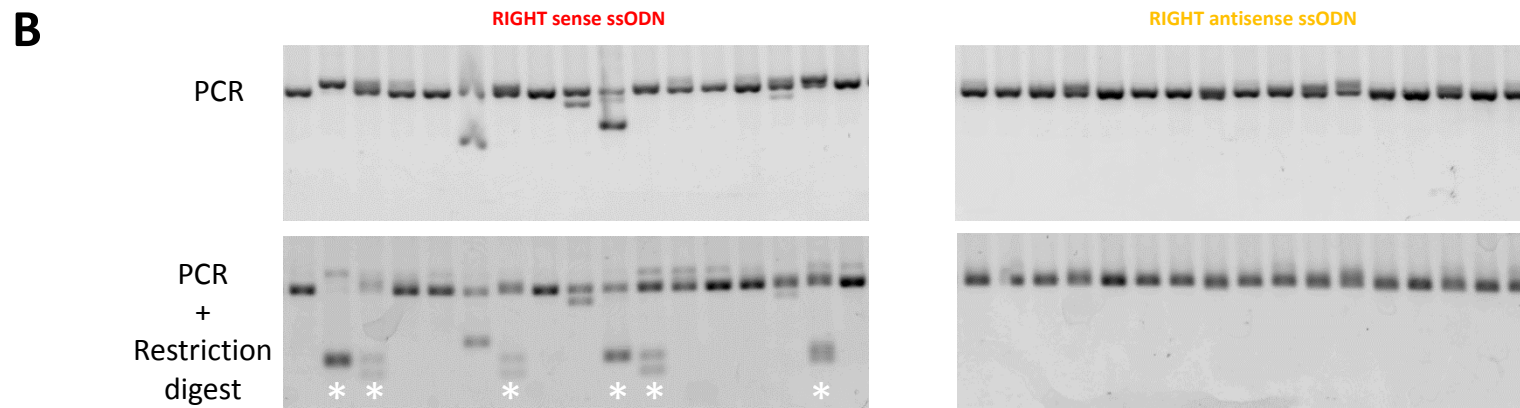
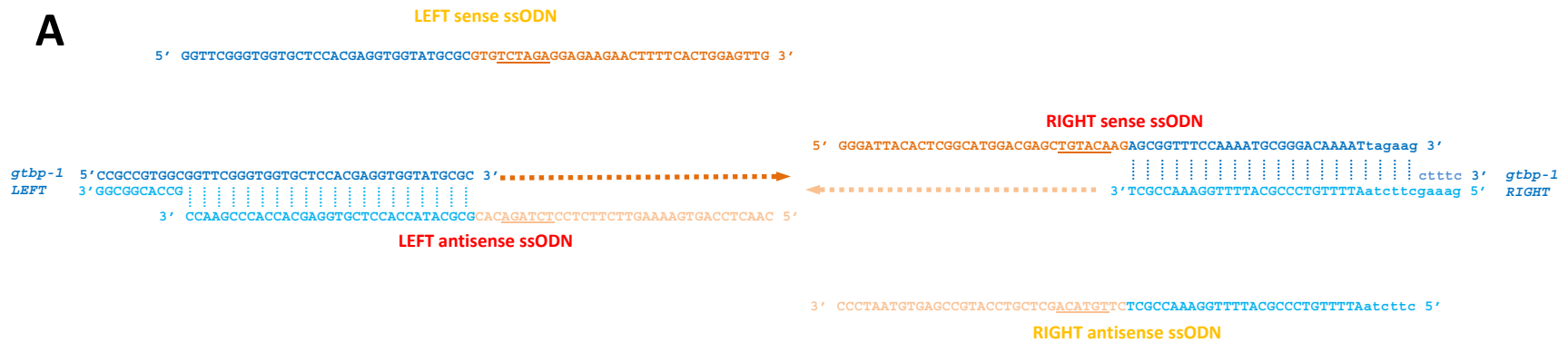
A. We identified edits based on GFP expression (not shown) and by PCR: the gel shows PCR amplification of the *gtbp-1* locus for a subset of F2 cohorts (Progenies of cloned single F1s). The lower band corresponds to the wild-type unedited allele and the upper band to the edited Myc insertion allele. Note that most edits are heterozygous, except for two, indicated by white asterisks. Heterozygous edits are most common since the injections were done in adult germlines that produce only oocytes, and the sperm were not exposed to the injection mix. Homozygous edits are obtained only in cases where the Cas9 complex (and possibly the repair template) perdures in eggs until after fertilization at which time paternal (sperm) DNA can be edited.

B. All edited hermaphrodites showed the characteristic cytoplasmic pattern of GTBP-1::GFP edits.

C. All edited hermaphrodites showed GFP localized to membranes consistent with the creation of a GTBP-1::GFP::PH domain fusion.

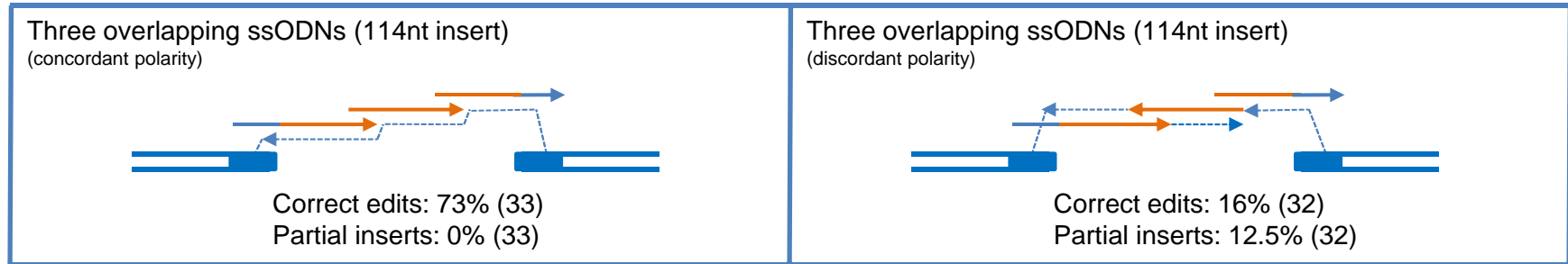
D. Edits were screened by PCR for the presence of the ZF1::3XFLAG insert. The gel shows PCR amplification of the *gtbp-1* locus for a subset of F2 cohorts. White asterisk denotes homozygous edit.

E. All edits showed the characteristic ZF1 degradation pattern, where protein is degraded in somatic, but not germline (arrow), blastomeres in embryos. Dotted line marks the embryo boundary.



### Supplementary Figure S2: Polarity requirements for donor ssODNs

- A. Sequence of *gtbp-1* locus at DSB is indicated in blue (resected for clarity). ssODNs have homology arms (blue) corresponding to sequences on left or right side of the break, and unique sequence (brown) containing a restriction site (underlined). Only the antisense ssODN can anneal (parallel dotted lines) on the left side of the resected DSB, and only the sense ssODN can anneal on the right side of the resected DSB (labeled in red). Stippled arrows show DNA synthesis templated by the annealed ssODN.
- B. Example of gels showing PCR amplification of the *gtbp-1* locus with (bottom panel) and without (top panel) restriction enzyme digestion after editing with the ssODNs with a right homology arm. Each well was loaded with DNA amplified from the progeny of one F1 *dpy-10*-edited worm. Only the sense ssODN yielded edits with the restriction enzyme site (white asterisks).

**A****B****C**

Left ssODN: GGTTCGGGTGGTGCTCCACGAGGTGGTATGCGCAAGCACACAGAATACAAAACGCGACTTTGTGATGCGTTCGCGCGTGAAGGATAC

Middle ssODN: CGACTTTGTGATGCGTTCGCGCGTGAAGGATACTGCCGTACAACGACAATTGCACATATGCTCACGGACAAGATGAG

Right ssODN: GACAATTGCACATATGCTCACGGACAAGATGAGCTGAGAGTTCGAGAAAGCGGTTTCCAAAATGCGGGACAAAATagaag

Full-length insert: GGTTCGGGTGGTGCTCCACGAGGTGGTATGCGCAAGCACACAGAATACAAAACGCGACTTTGTGATGCGTTCGCGCGTGAAGGATACTGCCGTACAACGACAATTGCACATATGCTCACGGACAAGATGAGCTGAGAGTTCGAGAAAGCGGTTTCCAAAATGCGGGACAAAATagaag

Partial insert 1: GGTTCGGGTGGTGCTCCACGAGGTGGT-----GCGACTTTGTGATGCGTTCGCGCGTGAAGGACA-----ATGCTCACGGACAAGATGAGCTGAGAGTTCGAGAAAGCGGTTTCCAAAATGCGGGACAAAATagaag

Partial insert 2: GGTTCGGGTGGTGCTCCACGAGGTGGT-----CACGAGGTGGAGGTGTCCGTG-----TATGCTCACGGACAAGATGAGCTGAGAGTTCGAGAAAGCGGTTTCCAAAATGCGGGACAAAATagaag

Partial insert 3: GGTTCGGGTGGTGCTCCACGAGGTGGTATG-----AGGTGGT-----ATTGCACATATGCTCACGGACAAGATGAGCTGAGAGTTCGAGAAAGCGGTTTCCAAAATGCGGGACAAAATagaag

Partial insert 4: GGTTCGGGTGGTGCTCCACGAGG-----CTTCCATTGCACATCC-----ATTGCACATATGCTCACGGACAAGATGAGCTGAGAGTTCGAGAAAGCGGTTTCCAAAATGCGGGACAAAATagaag

**Supplementary Figure S3: Template switching with ssODNs**

- A. Experimental set up as shown in Figure 2F and 2G.
- B. Gels showing PCR amplification of representative edits for each experiment shown in A. + signs indicate full length inserts, - signs indicate partial inserts
- C. Sequence of one full-length and 5 partial edits from experiment with discordant polarity ssODNs. All partial edits contain sequence from the right most ssODN. Sequence in black are non-homologous to locus or ssODN. The middle ssODN sequence is shown in the sense polarity for clarity.

A.

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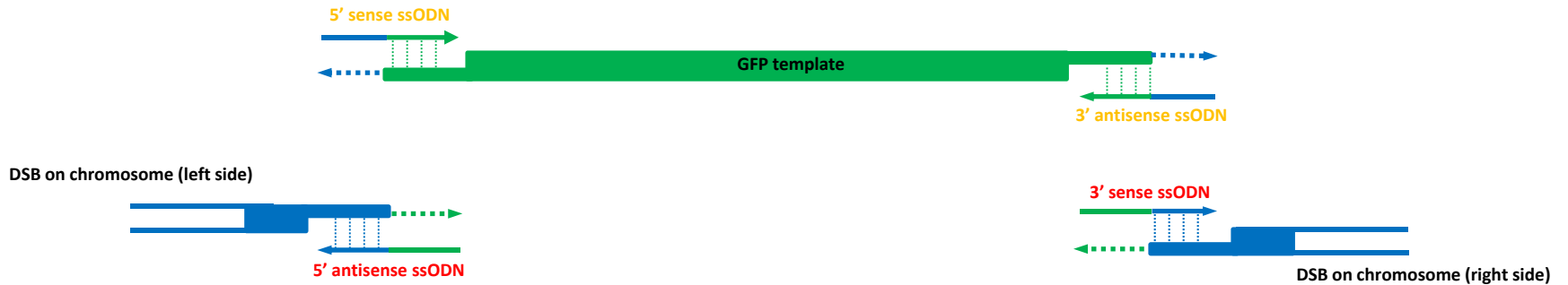
                    5' sense ssODN                                3' sense ssODN
5' GGTTCGGGTGGTGTCCACGAGGTGGTATGCGC--GTGAGTAAAGGAGAAGAAGCTTTTCACTGGAGTTG 3'
                    5' GGGATTACACTCGGCATGGACGAGCTGTACAAG--AGCGGTTTCCAAAATGCGGGACAAAATtagaag 3'

gtbp-1 5' CCGCCGTGGCGGTTTCGGGTGGTGTCCACGAGGTGGTATGCGC GTGAGTAAAGGAGAAGAAGCTTTTCACTGGAGTTGTCCCA--GFP---CTGCTGGGATTACACTCGGCATGGACGAGCTGTACAAG AGCGGTTTCCAAAATGCGGGACAAAATtagaagcttcc 3'
        3' GCGGCACCGCCAAGCCACCAACGAGGTGCTCCACCATACGCG CACTCATTTCCTCTTCTTGAAAAGTGACCTCAACAGGGT---GFP---GACGACCCTAATGTGAGCCGTACCTGCTCGACATGTTT TCGCCAAAGGTTTTACGCCCTGTTTTAatcttcgaag 5'

3' CCAAGCCCACCACGAGGTGCTCCACCATACGCG--CACTCATTTCCTCTTCTTGAAAAGTGACCTCAAC 5'
                    5' antisense ssODN                                3' antisense ssODN
3' CCCTAATGTGAGCCGTACCTGCTCGACATGTTT--TCGCCAAAGGTTTTACGCCCTGTTTTAatcttc 5'

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B.



### Supplementary Figure S4: Sequences of bridging ssODNs used to insert GFP at *gtbp-1* locus

- Sequence from *gtbp-1* locus is blue and sequence from the GFP PCR repair template is green. crRNA sequence is underlined and PAM is double underlined in *gtbp-1* locus. Bridging ssODNs that performed best in the experiments shown in Figure 3 are labeled in yellow.
- Schematics illustrate how the ssODNs can pair with the resected GFP to add sequences homologous to the DSB (blue), or can pair with the resected DSB to add sequences homologous to GFP (bottom).







Supplementary Table S4: Primers and ssODN sequences

Name	Sens	Description	Sequence (5' to 3')
<b>Bridge and repair ssODNs</b>			
829	S	Bridging 5' end eGFP with <i>gfpb-2</i>	ggttccggagtgctccacaggggaggggagggaggaagggagccttccctggatg
830	S	Bridging 3' end eGFP (contain <i>NotI</i> site) with <i>gfpb-1</i>	gggattacacggcattgagagagctccagaggggtttcaaatgaggcaaaatagag
1013	S	Bridging first and second halves of eGFP together	ggggaattacacggcattgagagagctccagaggggtttcaaatgaggcaaaatagag
1015	AS	as 829 but inversed polarity	caactccagaaaagttctcttactcagcgratcacacctctggagaccaccgaac
1016	AS	as 830 but inversed polarity	cttcaatttggcctatttggaaacgcttctccaggggtttcaaatgaggcaaaatagag
1267	AS	Bridging 3' end mNeonGreen with <i>gfpb-1</i> and inserting a 3Xflag tag	cttcaatttggcctatttggaaacgcttctccaggggtttcaaatgaggcaaaatagag cgcttctccatctcagctc
1349	S	Bridging 5' end eGFP with <i>gfp-2</i>	ccagtgatcaactgctccacagaggaggagggaggaagaaagagaaactttcactgagtg
1350	AS	Bridging 3' end eGFP with <i>gfp-2</i>	gcatataaatggaacaactcaattggtcttcttctcagcagctccagctcagctcagctc
1363	AS	as 1365 but inversed polarity	ctcactctcgtgagatagtgcaattgtgtgtgagggcgatctctcaggggaaagcacaagatg
1365	S	Partial ZF1	cpactttgtgctgctcggagagatctccgctcaacaacaactgcatctgcatcagcagcagc
1376	S	Partial ZF1 with <i>gfpb-1</i> homology arm	gggtcgggtggtctccaggggtgtgagcagcacaagaatacaaaagcpxctttgtgctcgtcagagatc
1377	S	Partial ZF1 with <i>gfpb-2</i> homology arm	gkacattgcatatgctcagcagagatgagtgaggttcgagagcggttcaaatgggcaaaatagag
1378	S	ZF1 with <i>gfpb-1</i> homology arm	gggtcgggtggtctccaggggtgtgagcagcacaagaatacaaaagcpxctttgtgctcgtcagagatc
1379	S	Partial ZF1 with <i>gfpb-2</i> homology arm	gkacattgcatatgctcagcagagatgagtgaggttcgagagcggttcaaatgggcaaaatagag
1380	S	Partial ZF1 and 3Xflag with <i>gfpb-1</i> homology arm	gggtcgggtggtctccaggggtgtgagcagcacaagaatacaaaagcpxctttgtgctcgtcagagatc caatgcatatgctcagcagagatgagtgaggttcgagagcggttcaaatgggcaaaatagag
1394	S	Bridging 5' end of <i>meq-3</i> N1 domain with 5' end of <i>gfpb-1</i>	aaatgcatatgctcggctcagagatgagtgagcagcacaagaatacaaaagcpxctttgtgctcgtcagagatc
1395	AS	Bridging 3' end of <i>meq-3</i> N1 domain with 3' end of <i>gfpb-1</i> and inserting a <i>ollas</i> tag and <i>NotI</i> site	gcttcaatttggcctatttggaaacgcttctccaggggtttcaaatgaggcaaaatagag tagaacaactctgatttctcattgctg
1407	S	Left <i>gfpb-2</i> homology arm with <i>NotI</i> site	ggggaattacacggcattgagagagctccagaggggtttcaaatgaggcaaaatagag
1408	AS	as 1407 but inversed polarity	caactccagaaaagttctcttactcagcgratcacacctctggagaccaccgaac
<b>PCR primers for linear repair templates</b>			
436	F	<i>gfpb-2</i> ~60 bp homology arm amplification (plasmid AP554-1)	gagggaactctctccgctc
437	R	<i>gfpb-1</i> ~60 bp homology arm amplification (plasmid AP554-1)	caagagaaaaggaagag
527	F	<i>gfpb-1</i> ~100 bp homology arm amplification (plasmid AP553-1)	gggagcattccagagagac
528	R	<i>gfpb-2</i> ~100 bp homology arm amplification (plasmid AP553-1)	tgagaaggatagacagatc
603	F	<i>gfpb-2</i> homology arm amplification (plasmid 1297-1 and 1298-1)	ggggttgggtggtctccaggggtgtgagcagcacaagaatacaaaagcpxctttgtgctcgtcagagatc
604	R	<i>gfpb-1</i> homology arm amplification (plasmid 1297-1 and 1298-1)	cttcaagaagaaaagag
645	F	eGFP/ <i>meGFP</i> amplification with <i>gfpb-1</i> homology arm	gggttccgggtggtctccaggggtgtgagcagcacaagaatacaaaagcpxctttgtgctcgtcagagatc
646	R	eGFP amplification with <i>gfpb-1</i> homology arm	cttcaatttggcctatttggaaacgcttctccaggggtttcaaatgaggcaaaatagag
715	F	mNeonGreen amplification with <i>gfpb-2</i> homology arm	gggttccgggtggtctccaggggtgtgagcagcacaagaatacaaaagcpxctttgtgctcgtcagagatc
831	F	eGFP amplification without homology arm	gtgagaaagagagagac
832	R	eGFP amplification without homology arm	cttgcacagctcctcag
1011	F	Second half of eGFP amplification	gttgaaggatcacccttg
1012	R	First half of eGFP amplification	ttgacttgcagcctcttg
1014	R	First half of eGFP amplification with overlapping sequence with the second half	cgatcttatacaaggatc
1017	F	eGFP amplification without homology arm but with extranucleotide in order to be in frame if PCR repair template inserted at <i>Cas9</i> cut	cggtgagaaagagagagac
1018	R	eGFP amplification without homology arm but with extranucleotide in order to be in frame if PCR repair template inserted at <i>Cas9</i> cut	cttctgcacagctcctcag
1084	R	mNeonGreen amplification with <i>gfpb-1</i> homology arm	cttcaatttggcctatttggaaacgcttctccaggggtttcaaatgaggcaaaatagag
1085	R	tagRFP amplification with <i>gfpb-1</i> homology arm	cttcaatttggcctatttggaaacgcttctccaggggtttcaaatgaggcaaaatagag
1096	R	meGFP:Linker amplification	actagttctagagggcgc
1099	F	3Xflag:tagRFP amplification	gactacaagcattgagc
1152	F	second half of mNeonGreen (with 1 intron):3Xflag:tagRFP amplification	cggtcaactatgctaac
1153	R	meGFP:Linker::first half of mNeonGreen (with 1 intron) amplification	gacttccctctagctg
1255	F	Linker::mNeonGreen amplification	gagttccacgggg
1256	R	mNeonGreen:3Xflag amplification	gacttctatacctcagctgctc
1263	R	eGFP amplification with <i>ollas</i> homology arm	ccatgagcgtggtcaggggttctggagactcagcagctcagcctccatg
1264	F	PH amplification with <i>ollas</i> homology arm	gattgccaaagcctgagcagctctcagggaaagcagggctccagatgacc
1265	R	PH amplification with <i>gfpb-1</i> homology arm	cttcaatttggcctatttggaaacgcttctccaggggtttcaaatgaggcaaaatagag
1266	R	mNeonGreen amplification without homology arm	cttgaagttgctcattc
1272	R	eGFP amplification with ZF1 homology arm	gatttctggtgcttcttctcagcctcctcag
1273	F	ZF1 amplification with eGFP homology arm	catggagcagctacaagagcagcagatcaaaaagc
1274	R	ZF1 amplification with <i>gfpb-1</i> homology arm	cttcaatttggcctatttggaaacgcttctccaggggtttcaaatgaggcaaaatagag
1292	F	eGFP amplification (1st PCR round) with <i>gfpb-1</i> (27bp) and partial <i>Myc</i>	aaaactgatacagagaggtctgaggggtttcaaatgaggcaaaatagag
1293	F	Partial <i>Myc</i> amplification (2nd PCR round) with <i>gfpb-1</i> homology arm	gggttccgggtggtctccaggggtgtgagcagcacaagaatacaaaagcpxctttgtgctcgtcagagatc
1294	R	eGFP amplification (1st PCR round) with <i>gfpb-2</i> homology arm	cttcaagaagaaaagagagaggaagcgttcttactgactcgtcctgacc
1294	R	eGFP amplification with <i>gfpb-1</i> homology arm away from the cut	cttcaagaagaaaagagagaggaagcgttcttactgactcgtcctgacc
1295	R	<i>gfpb-1</i> homology arm amplification (2nd PCR round)	cttcaagaagaaaagagaggaag
1357	F	Additional non-homologous bases (partial 3Xflag tag) and <i>gfpb-2</i> homology arm amplification (plasmid 554-1)	catgatatgattacagatgagcagaggggttccgggtgctccac
1358	R	Additional non-homologous bases (partial <i>bbp-2</i> 3' UTR) and <i>gfpb-1</i> homology arm amplification (plasmid 554-1)	gggagttgaaagatttgcattataccttcttaatttccgctc
1375	F	eGFP amplification with <i>gfpb-2</i> homology arm away from the cut	gtttcaaccaagggggaacttctcgcgctgagaaagagagagac
1392	F	<i>meq-3</i> N1 domain amplification without homology arm	atgattctcaaaccttacc
1393	R	<i>meq-3</i> N1 domain amplification without homology arm	gtctagcaacatctcagttta
1506	F	Mutated <i>meq-3</i> coding sequence amplification with <i>meq-3</i> deletion: <i>hotspot</i> homology arm	tcaaaccttaccagagaggtctcagcagacgggttcccaaacgtcagagaaaagggaggg
1507	R	First half of mutated <i>meq-3</i> coding sequence amplification	tgtctgactctgaccac
1508	F	Second half of mutated <i>meq-3</i> coding sequence amplification	acgctcagatgctgccc
1509	R	Mutated <i>meq-3</i> coding sequence amplification with <i>meq-3</i> deletion: <i>hotspot</i> homology arm	gtaatagataaattgttacaatagactcttccacttccatgagcagctctgttgggaactcctctctgctccacaacttcc ggcaggtctcttcaacatcttccagctg
<b>PCR primers for genotyping</b>			
321	F	<i>gfpb-1</i> genotyping (3' end cut)	agtaaaagccatttgaaggg
322	R	<i>gfpb-1</i> genotyping (3' end cut)	gataagtggaatggatgc
390	F	<i>gfpb-2</i> genotyping (gene replacement)	gtgctgcaatcaactgg
391	R	<i>gfpb-2</i> genotyping (gene replacement)	gagcgggtgattgattg
402	F	<i>gfpb-1</i> edits sequencing (3' end cut)	caactcaactggatcattg
408	F	<i>gfpb-1</i> edits sequencing (gene replacement)	ctttgaatttgggtctgg
412	R	<i>gfpb-1</i> edits sequencing (3' end cut)	gacttcaaaaagctacagc
1510	F	<i>meq-3</i> genotyping	catcttctgctgcttccc
1511	R	<i>meq-3</i> genotyping	tcaaaagagagagcagctg

F and R : Forward and Reverse primers for PCR amplification. S and AS: Sens and Antisense polarity for ssODNs.



**Supplementary Table S5: Plasmids and new strains**

Plasmid name	Backbone	Insert
AP553-1	pUC19	gtbp-1 (~500 bp flanking sequence)::GFP (with 3 introns)::gtbp-1 (flanking sequence)
AP554-1	pUC19	gtbp-1 (~60bp flanking sequence)::GFP (with 3 introns)::gtbp-1 (flanking sequence)
AP625-1	Paix <i>et al.</i> , 2015	eGFP
AP763-1	pUC19	Pleckstrin Homology Domain From Phospholipase C Delta (Green <i>et al.</i> , 2011)
AP931-8N	pUC19	TEV::meGFP::Linker::mNeonGreen (with 2 introns)::3Xflag::tagRFP
AP1297-1	pUC19	gtbp-1 (flanking sequence)::eGFP::gtbp-1 (27bp)::PH::gtbp-1 (flanking sequence)
AP1298-1	pUC19	gtbp-1 (flanking sequence)::eGFP::recoded gtbp-1(8/27bp, every 3bp)::PH::gtbp-1 (flanking sequence)

Plasmid containing the *pie-1* ZF1 domain was a gift from Chih-Yung Lee, Seydoux lab

Experimental variant	CRISPR alleles
Figure 1-Panel F	<i>gtbp-1::Myc</i>
Figure 1-Panel H	<i>gtbp-1::eGFP::recoded gtbp-1(8/27bp, every 3bp)::PH (at Stop)</i>
Figure 2-Panel F	<i>gtbp-1::ZF1</i>
Figure 4-Panel A	<i>gtbp-1::ZF1::3Xflag</i>
Figure 4-Panel B	<i>gtbp-1::eGFP::ZF1</i>
Figure 4-Panel C	<i>gtbp-1::eGFP::ollas::PH</i>
Figure 4-Panel D	<i>gtbp-1:: meGFP::Linker::mNeon</i>
Figure 4-Panel E	<i>gtbp-1:: meGFP::Linker::mNeon::3Xflag::tagRFP</i>
Figure 4-Panel G	<i>glh-1::eGFP</i>
Figure 4-Panel H	<i>gtbp-1::mNeon::3Xflag</i>
Figure 4-Panel I	<i>gtbp-1 replacement with meg-3 Nt domain::hotspot::ollas</i>
Figure 4-Panel J	<i>mutated meg-3 coding sequence (122 mutations spread on 1.7 kb)</i>