## Fibroblast activation protein enzyme deficiency prevents liver steatosis, insulin resistance and glucose intolerance and increases fibroblast growth factor-21 in diet induced obese mice

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## **Supplementary Material:**

## **Supplementary Method:**

#### Generation of the FAP gene knock-in (gki) mouse strain

Fap conditional and constitutive knock-in mice containing the point mutation Ser624Ala, which ablates catalytic activity, [1] were generated by Ozgene (Bentley, Western Australia) using standard techniques. A "floxed" "mini cDNA cassette, cloned into intron 21 approximately 100 bp upstream of exon 22 (E22) (Fig. S1), consisted of a fused exon (E22-E26) derived from wildtype Fap cDNA (OriGene, Rockville, MD; #BC019190) followed by a polyadenylation signal (pA) to terminate transcription and thus force expression of wildtype Fap; and further followed by a PGK-neomycin selection cassette flanked by FRT sites. The TCC to GCC point mutation (Ser624Ala) was targeted into the first codon of endogenous E22 downstream of the mini cDNA cassette and the selection cassette.

Fap<sup>wt/flox</sup> neomycin-resistant C57BL/6 Bruce4 embryonic stem (ES) cell clones were identified by Southern blot analyses (Fig. S2) and microinjected into BALB/c blastocysts for generation of Fap<sup>wt/flox</sup> mice. Fap<sup>wt/conKl</sup> mice, which express the wildtype Fap mini cDNA cassette, were generated by flp-mediated excision of the PGK-neomycin selection cassette. Fap<sup>wt/Kl</sup> (heterozygote FAPgki) mice were generated by cre-mediated excision of the wildtype Fap mini cDNA cassette. All ES clones and mice were screened and verified by Southern Blot analyses (Fig. S2). Putative FAPgki mice were also screened by specific FAP enzyme assay of plasma [2] (Fig. S3A). We previously showed that human FAPSer624Ala is cell surface expressed and bound by the F19 MAb, which depends upon

correct protein conformation [1, 3]. As evidence for correct protein folding of FAP in mice that carry this mutation, we showed that cell surface expression of FAP was not diminished by Ser624Ala point mutation in the FAPgki mice (Fig. S3B).

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#### **Supplementary Method:**

#### Cell surface expression of FAP on fibroblasts

Lung was harvested from FAPgki mouse and cut into ~1 mm pieces that were digested with 0.5 mg/ml collagenase at 37°C for 30 min, then pushed through a 70 μm cell strainer. After centrifugation at 1200 rpm for 5 min, cells were resuspended in complete media (DMEM with 10% FCS, antibiotic and Amphotericin B) and cultured. For flow cytometry, confluent fibroblasts were harvested as cell suspensions using trypleE (Cat. no. 12605, Thermo Scientific, Carlsbad, CA, USA). Briefly, cells were fixed in 2% (v/v) paraformaldehyde (Cat. no. 18814, Polysciences, Warrington, PA, USA) and stained with primary antibody against FAP (Cat. no. BAF3715, R&D Systems, Minneapolis, MN, USA) or normal sheep IgG (Cat. no. 31243, Thermo Fisher Scientific) as a negative control. Cells were analyzed on Cano II (BD Biosciences). Flow cytometric data were analyzed with FlowJo software version 9.9 (TreeStar, Ashland, Ore).

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#### **Supplementary Method:**

#### Plasma GLP-1

For GLP-1 assay, blood samples were collected from fasting mice and mice after 15 min post glucose challenge. Glucose (2 g/kg body weight) was administered by oral gavage following 6 h fasting. Blood samples were collected on ice in 1 mL EDTA tubes (Mini Collect®, Kremsmunster, Austria). Prior to blood collection, DPP4 inhibitor, Sitagliptin (Merck, NJ, USA) was added to the tubes for a final concentration of 10 uM, to prevent ex vivo degradation of GLP-1 in the blood. Blood was centrifuged at 3000 g for 5 min and supernatant (plasma) was stored at -80°C. All steps were performed at 4°C. Active GLP-1 was measured using GLP-1 ELISA kit (Cat. No. #81508) (Crystal Chem, IL, USA) according to manufacturers instructions.

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#### **Supplementary Method:**

#### Diets:

Mice were given ad libitum access to water and either chow diet (8% kcal fat, 21% kcal protein, 71% kcal carbohydrate; Gordon's Specialty Stock Feeds, Yanderra, NSW, Australia or Specialty Feeds, WA, Australia) (control group) or to High Fat Diet (HFD).

Each HFD cage had non-edible alpha-dri bedding and, for dental health, a wooden chew block (cat no. ASAEC-A, Able Scientific, WA, Australia). HFD was purchased (Specialty Feeds; Cat. No. SF03-020) or made in-house. The Specialty Feeds HFD was hydrogenated vegetable oil (Copha) based and contained 43% kcal fat, 17% kcal protein, 40% kcal carbohydrate (with 0.19% weight/volume cholesterol) as it is designed to be atherogenic and diabetogenic (Specialty Feeds catalogue, 2009). The in-house HFD was based on rodent diet D12451 of Research Diets (New Brunswick, NJ, USA) [4, 5]. The detail of this lard based in-house HFD, which contains 45% kcal fat, 20% kcal protein, 35% kcal carbohydrate, is published [4]. Weight gain by WT mice was similar between chow and  $\gamma$ -irradiated HFD, so when  $\gamma$ -irradiation of commercially supplied HFD became mandatory in our animal facility, only in-house HFD was used.

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#### **Supplementary Method:**

#### Immunohistochemistry and immunofluorescence stain

Liver paraffin sections at 5  $\mu m$  were rehydrated and then antigen retrieved using proteinase K (20  $\mu g/ml$ ) for 20 min. F4/80 antibody supernatant (Gift from Dr S Tikoo) at 10  $\mu g/ml$  was incubated for 1.5 hour at room temperature, with rat lgG as a control. After repeated washes in PBS, sections were incubated for 30 min with horseradish peroxidase (HRP)-conjugated secondary antibody (Dako, Glostrup, Denmark). The chromogenic reaction employed 3,3-diaminobenzidine (DAB) with  $H_2O_2$ . Liver frozen sections at 5  $\mu m$  were fixed with 4% PFA and blocked with BSA. Primary antibody to CD36 (AF2519, R&D) at 33 lg/ml was incubated for 1.5 h at room temperature followed by a AF488-labelled donkey anti-sheep antibody then DAPI to counterstain nuclei. Quantification of immunostains used an LAS image software package, version 4.8. Total F4/80 staining was expressed as percentage of F4/80 immunostain in the assessed liver area at 10x magnification. Fluorescence intensity of CD36 immunostaining from four fields per section at 200x magnification was quantified in arbitrary units of fluorescent signal at 488 nm absorbance.

Table S1. The genes studied by Taqman gene expression assay.

Gene Name	Gene ID	Assay ID
Acetyl-Coenzyme A carboxylase alpha	Acc	Mm01304276_m1
Apolipoprotein C3	Apoc3	Mm00445670_m1
Carnitine palmitoyltransferase 1	Cpt1	Mm00550438_m1
CD36 Antigen	Cd36	Mm01135198_m1
Carbohydrate Response Element Binding Protein; MLX Interacting Protein Like	Chrebp; Mlxipl	Mm00498811_m1
Dipeptidyl peptidase 4	Dpp4	Mm00494548_m1
Dipeptidyl peptidase 8	Dpp8	Mm00547049_m1
Dipeptiyl peptidase 9	Dpp9	Mm00841122_m1
Eukaryotic 18S rRNA	18S	Hs99999901_s1
Fatty acid synthase	Fasn	Mm00662319_m1
Fibroblast activation protein	Fap	Mm00484254_m1
Fibroblast growth factor 21	Fgf21	Mm00840165_g1
Glucokinase	Gck	Mm00439129_m1
Low density lipoprotein receptor	Ldlr	Mm01177349_m1
Lipin1	Lipin-1	Mm00550511_m1
Peroxisome proliferative activated receptor gamma	Ppar-γ	Mm01184322_m1
Peroxisome proliferator activated receptors α	Pparα	Mm00627559_m1
Sterol regulatory element binding protein-1c	Srebp1- c	Mm00550338_m1
Eukaryotic 18S rRNA*	18S	Hs99999901_s1
Beta-actin*	Actb	Mm00607939_s1

Housekeeping gene.

Table S2. All data of human pancreas samples obtained from nPOD.

FAP	Donor Type	AutoAb	<u>Age</u>	<u>Diabetes</u>	Gender	<u>Ethnicity</u>	C-peptide	HbA1c	ВМІ	Clinical	Cause	Histopathology	Comment
Activity		(RIA)	(years)	<u>Duration</u>			(ng/ml)			History	<u>of</u>		
(∆pmol				(years)							<u>Death</u>		
AMC/min/m													
g Protein)													
3.943	No diabetes	Negative	59		Female	Caucasian	9.89		24.8			Ins+/Gluc+ normal	
												islets. Fatty	
												infiltration-mild	
5.664	No diabetes	Negative	30		Male	Caucasian	17.91		20.6			Ins+/Gluc+ normal	
												islets. Insulin+ less	
												intense and	
												distinct as for most	
												ot	
22.957	No diabetes	Negative	27		Male	Hispanic	9.09		19.1			Normal. Very mild,	Very faint
												acute pancreatitis.	glucagon.
													Repeating
													stains.
4.044	No diabetes	Negative	27.1		Male	Caucasian	7.71	6.3	35.6			Ins+/Gluc+ normal	Multifocal, mild
												islets, many large	ductular
												especially in head	mucinous
												and body. Degree	metaplasia.
												of	
5.432	No diabetes	Negative	45.1		Female	Caucasian	0.55	6.1	35.1	Hypertension		Ins+/Gluc+ islets.	Focal islet
										Father had		Multifocal, mild	hyperplasia.
										diabetes and		periductal and peri-	
										stroke.		islet fibrosis. Mil	
4.239	No diabetes	Negative	41		Male	Caucasian	20.55		20.5			Ins+/Gluc+ islets.	Scaring one
												Mild adipose	section. 1 foci
												infiltration exocrine	mononuclear
												regions.	infiltrate peri-
													vascular.

6.524	No diabetes	Negative	24.2	Male	Caucasian	1.01		24.8	IA2A+ by screening.		Ins+/Gluc+ islets. Occ. high islet	
											Ki67.	
7.028	No diabetes	Negative	45.8	Female	Caucasian	4.45	5.6	25			Ins+/Gluc+	Multifocal
											numerous islets,	ductular
											normal sizes. No	dysplasia with
											infiltrates. Mild	epithelial
											acinar fat	proliferation.
												Extra-acinar
												islets have back
												ground staining
												of insulin into
												surrounding
												connective
												tissues. Minor
												background of
												DAB chromogen
												for CD3 too.
9.972	No diabetes	Negative	31	Female	Caucasian	6.23	5.5	26.9		Head	Ins+/Gluc+ islets,	
										Trauma	no abnormalities	
										(MVA)	observed.	
											Occasional islet	
											hyperemia.	
4.984	No diabetes	Negative	20	Female	Caucasian	6.89	5.8	25.6		Head	Ins+/Gluc+ islets.	
										trauma	No abnormalities	
										(MVA)	observed. Low	
											Ki67 all	
											compartments.	
4.153	No diabetes	Negative	31	Male	Caucasian	8.1		25.4		Head	Ins+/Gluc+ islets,	Very clean
										Trauma	numerous. Occ.	pancreas,
										(Gun	islet has high	PanTail has 1
										shot)	numbers Ki67	lobule with high
											cells. Islet	fatty infiltration.

5.286	T1D	Negative	22.4	14	Male	Caucasian	<0.05	24.1	Maternal	Ins-/Gluc+ islets.	Islets
									uncle and	Mild to moderate	reasonable
									paternal	atrophy.	number and
									grandfather		sizes.
									had T1D.		
									Retinopathy		
									and		
									neuropathy		
19.345	T1D	Negative	31.4	15	Male	Hispanic	0.24	28		Ins+ islets. Mild to	Ki67+ vascular
										moderate chronic	smooth muscle.
										pancreatitis.	Mild ductular
										Moderate	metaplasia.
										interlobular a	
7.813	T1D	GADA+	31.2	5	Male	Caucasian	<0.05	27		Ins+/Gluc+ islets	Possible
		IA-2A+								(much decreased).	insulitis. Mild to
		ZnT8A+								Insulitis. Chronic	moderate CVD.
		mIAA+								pancreatitis- mild,	Fatty infiltrate
											mild. Very mild,
											focal autolysis
											few blocks. Peri-
											ductular fibrosis
											moderate.
9.615	T1D	GADA+	43.5	21	Male	Caucasian	<0.05	28.7	GADA+ by	Ins- islets	
		mIAA+							autoab.	including extra-	
									screening.	acinar. Moderate to	
									Endstage	severe exocrine	
									renal failure	atrophy. Mo	
									(dialysis 1		
									year), HTN,		
8.954	T1D	mIAA+	49.2	41	Female	Caucasian	<0.05	33.7	Coronary	Ins-/Gluc+,	At recovery
									artery	atrophy, decreased	noted: areas of
									disease and	numbers. High	fatty pancreas
									hypertension.	acinar and duct	with evidence of
										Ki67. CD3+ inf	chronic

												inflammation
												and pancreatitis.
												Admission
												glucose >600
												and hypo-
												kalemia. PA also
												noted gross
												enlargement
												with areas of
												saponification
												and pancreatitis.
37.748	T1D	Negative	44.1	15	Male	Caucasian	<0.05		23.9		Ins-/Gluc+ islets,	
											reduced density.	
											Acinar atrophy	
											moderate to severe	
											with	
33.881	T1D	GADA+	26	15	Female	African	0.48		26.6		Ins+ (reduced	Possible
00.00		mIAA+				American					numbers but in all	amyloid.
						7					regions)/Gluc+	<b>y</b>
											islets, normal	
											sizes, reduc	
51.234	T1D	GADA+	28	21	Male	Caucasian	<0.05	7.2		Mother had	Ins-/Gluc+ islets.	Some black
		mIAA+								gestational	High duct and	precipitate may
										diabetes	exocrine Ki67.	interfere with
										while	Small duct	image analysis.
										pregnant with	plugging with ru	Acute and
										donor.	plagging than rain	chronic
										Hypertension		pancreatitis.
										(mi		panorounio
75.039	T1D	Negative	32	16	Female	Caucasian	<0.05		23.4	<b>V</b>	Ins-/Gluc+ islets	
. 5.000				•		- 444441411	0.00				present in reduced	
											numbers. No	
											significant	
											inflammation b	
I				]							iiiiaiiiiiatioii b	

6.925	T1D	GADA+	35	11	Female	Caucasian	<0.05		27.4			Anoxia	Ins-/Gluc+	slets in		
		mIAA+											much	reduced		
													numbers. M	oderate		
													widespread			
													exocrine atı			
11.819	T1D	mIAA+	61	52	Male	Caucasian	<0.05		21.6			Cerebro	Ins-/Gluc+	islets	Reviewed	with
												vascula	(pseudoatro		Chen	Liu-
												r/Stroke	Exocrine		PanIN1	
													moderate	with	designation	on.
													focal mi		Stretch t	
															PanIN2.	
49.447	T1D	Negative	58	22	Female	Caucasian	1.28	6.1	28.6			Not	Ins+/Gluc+	islets	Chen	Liu
		13										avail		reduced	reviewed	
													numbers).		mct.	Also
													chronic		reviewed	
													pancreatitis	_	1. Treme	
													Mode	•	atrophy.	
													in out on			s. 3.
															Insulin+	
															lots. 4. F	
															PT 04, foc	
															likely.	
9.848	T1D	IA-2A+	39	19	Male	Caucasian	<0.05		19.5	Fell (	(non-	Head	Ins-/Gluc+	islets.	Chen	Liu
3.040	110	mIAA+		'	Wate	Oducasian	40.03		13.3	MVA	-11011	Trauma	Acute pand		reviewed	
		IIIIAA.								accident)		ITauma	moderate to		mct.	0, 10, 13
										accidenty	•		severe. Duc	-	mot.	
16.588	T1D	IA-2A+	41	27	Female	African	<0.05		21.1			Cerebro	Ins-/Gluc+		A*36:01,	74:01
10.300	םו ו	IA-ZA+	* '	21	remale	American	<b>\0.03</b>		21.1			vascula	reduced n		A 30.01,	74.01
						Allielicail										
												r/Stroke	Extra-acina			
													in fibrotic re	:yıo		

16.744	T2D	Negative	18.8	0.25	Female	Hispanic	10.68		39.1	DKA 3	Ins+ islets,	Islets are insulin
										months prior	clusters, single	+ with normal
										with acute	cells plentiful.	Ki67 (ie, rare).
										renal failure.	CD3+ foci intra-	Ducts have
										3 months	acinar and p	normal numbers
										duration	•	of beta cells
										diabetes.		(present yet
										Ca		exceedingly low
												proportion).
												Only occ. focal
												ductular
												proliferation or
												mucinous
												metaplasia.
												Glucagon + islet
												cells very
												heterogeneous
												distribution
												between lobules
												and faint. Fatty
												infiltrate mild-
												note high BMI.
9.384	T2D	mIAA+	48.8	0	Female	Hispanic	<0.05	8	32.5	Preclinical.	Ins+/Gluc+ islets	Mild autolysis
										Plasma (no	though in reduced	PanHead 02.
										serum)	density, especially	Islets in head
										available.	in head. Amyloid.	region arranged
											L	more in loops
												compared to
												body and tail
												regions.
12.470	T2D	Negative	20.7	0	Female	African	0.58		40	Acute onsent.	Ins+	Some lobules
						American				Gestational	(reduced)/Gluc+	have increased
										diabetes in	islets various	fibrosis between
										past. Father	sizes, some	acinar cells.

										had diabetes.	atrophied. Low Ki67. No fat	
16.204	T2D	Negative	42.8	2	Male	Caucasian	0.58	7.8	31	Diabetes medication was oral Metformin (1 g/day) for 2 years but noncomplia	Ins+ (reduced)/Gluc+ islets with severe amyloidosis. Severe exocrine atroph	Megaislets/islet hypertrophy (8/26/13)
7.236	T2D	Negative	62.3	3	Male	Caucasian	2.85		33.7	Metformin for 3 years.	Ins+/Gluc+, normal islet density. Hypertrophied islets. Amyloid. Acinar atr	Mild acinar autolysis with possible secondary changes connective tissues including islet vasculature.
1.478	T2D	mIAA+	44.8	10	Female	Caucasian	0.08		30.4		Ins+ (reduced)/Gluc+ islets. Mild adiposity exocrine regions. Low Ki67.	Amyloid noted 5/30/13 mct.
6.195	T2D	Negative	55.8	0	Female	Hispanic	0.8	9.1	44.6	Preclinical T2D ie no diagnosis or medication for diabetes; Hypertension fo	Ins+ (reduced) islets, numerous, smaller. Low Ki67. Amyloid. Exocrine atrop	

45.791	T2D	No	45.1	20	Male	Hispanic	No serum	8	25.3	No serum or	Ins+ (weak)/Gluc+	HbA1c added
		serum					available			cells	islets, numerous	mct 7/22/13
		available								available.	but small sized.	
										Negative for	Mild acinar	
										GADA and	atrophy.	
										IA2A by		
										autoantibody		
										scr		
7.641	T2D+Increti	Negative	68.3	5	Male	Caucasian	2.98	6.3	20.9		Ins+/Gluc+ islets,	Low Ki67.
	n										plentiful; mild,	HbA1c added
											multifocal amyloid.	mct 7/22/13
											Mild, multifocal	
											ch	
24.893	T2D	Negative	36.1	0	Male	Hispanic	3.45	7.2	30.6	Preclinical	Ins+/Gluc+ islets.	
										T2D based on	Low Ki67. No	
										HbA1c.	infiltrates or	
											amyloid observed.	
15.458	T2D+Increti	mIAA+	48.5	26	Female	Caucasian	1.85		36.1		Ins+/Gluc+	Focally severe
	n										numerous islets.	exocrine
											Rare amyloid in	atrophy.
											islets. Very mild	
											chronic pancr	
29.407	T2D	Negative	62	10	Female	Caucasian	6.14	6	19.9	Metformin 1g	Ins+/Gluc+ plentiful	
										/ twice a day.	islets, several with	
											vascular	
											congestion or	
											amyloid. S	
11.393	T2D+Increti	Negative	47.4	13	Male	Caucasian	0.16	7.3		Renal	Ins+/Gluc+ islets,	IPMN PanHead
	n									tranplant 7	numerous.	04 block (this
										years ago;	Moderate acinar	statement added
										retinopathy.	atrophy with fatty	to histopath and
											replacement	specimen levels
												1/14/14 mct)

24.064	T2D+Increti	Negative	53	20	Male	African	8.87	29.6	Father and 3	Ins+/Gluc+ islets	
	n					American			siblings with	with moderate	
									diabetes.	fibrosis and	
										variable	
										amyloidosis.	
										Moderate	

## **Supplementary Figure legends**

- **Fig. S1. Generation of conditional and constitutive** *Fap* **Ser624Ala knockin (FAPgki) mice.** Representation of the *Fap* wildtype genomic locus (A), the targeted locus  $Fap^{flox}$  (B), the FRT recombined locus  $Fap^{conKl}$  (C) and the *Cre* deleted locus  $Fap^{Kl}$  (D). 5' probe, 3' probe, enP probe and relevant restriction sites are indicated, as well as the positions of the wildtype *Fap* mini cDNA cassette and *PGK-neomycin* selection cassette.
- **Fig. S2. Southern blot verification of** *Fap*<sup>wt/conKI</sup> **and** *Fap*<sup>wt/KI</sup> **mice.** Southern blot screening tail DNA confirmed targeted recombination at each end of the Fap locus: 5' probe on *Eco*RV digest showing *Fap*<sup>wt</sup> and *Fap*<sup>flox</sup> alleles (A), 3' probe on *Kpn*I digest showing *Fap*<sup>wt</sup> and *Fap*<sup>flox</sup> alleles (B). *Fap*<sup>conKI</sup> (C) and *Fap*<sup>KI</sup> alleles (D) were identified using the enP probe on *Eco*RV digest. The use of a neomycin probe confirmed a single integration event (data not shown). As expected, intrahepatic *Fap* gene expression levels were comparable in WT and *Fap*<sup>KI/KI</sup> mice (E).
- Fig. S3. FAPgki mice lacked FAP enzyme activity but retained cell surface expression of FAP protein. (A) FAP specific enzyme assay of plasma from 3-week-old mice showed that  $Fap^{Kl/Kl}$  (FAPgki) mice lacked FAP enzyme activity whereas  $Fap^{wt/Kl}$  mice had approximately half as much FAP enzyme activity as WT mice (n=2 mice per group). (B) Flow cytometry on primary lung fibroblasts from a FAPgki mouse showed that the single point mutation at the enzyme active site serine did not prevent cell surface expression of FAP (30.9% FAP positive cells) in the FAPgki mice. (C) Clockwise flow cytometry gating strategy showed that live and single cells were gated to identify the percentage of FAP positive cells, compared with IgG control. Histogram key: blue is control IgG, red is anti-FAP.
- **Fig S4.** Improved glucose tolerance in male FAP gko mice. Oral GTT (A) and glucose AUC (B) at 14 weeks of HFD (Specialty Feeds). Ip GTT (C) and glucose AUC (D) at 20 weeks of HFD (in-house HFD). Individual replicates (B, D). Mean ± SEM. n = 6 male mice per group. \*p<0.05 by Mann-Whitney U test.

- **Fig. S5. FAPgki mice were protected from pancreatic islet hypertrophy.** FAPgki and WT male mice following 12 weeks of in-house HFD. Representative islets of those quantified for the data presented in Figure 3, immunostained for insulin and glucagon.
- **Fig. S6. FAP gko mice were resistant to HFD induced steatosis.** Intrahepatic lipid measured by quantitative calorimetric oil red O assay (fold change from average of WT Chow). Liver was obtained after overnight fasting. \*p<0.05, with or without exclusion of the highest data point, versus genotype-matched controls using Mann-Whitney U test. Individual replicates and mean ± SEM. n=5-6 female mice per group. Fold changes from livers depicted in Fig. 4 panels E and F were, respectively, 257% and 206%.
- **Fig. S7. FAP gko mouse liver.** Sirius Red staining did not detect fibrosis in either WT (A) or FAP gko (B) mice. Representative liver sections from n=5-6 female mice per group, at 20 weeks of HFD (in-house diet). Scale bars= 200 μm.
- Fig. S8. F4/80 macrophage stain in liver and adipose tissue. Liver (A), BAT (C), WAT (D) with immunoperoxidase stain of F4/80 and haematoxylin counterstain for nuclei. Quantitation of immunopositivity in liver as a percentage of area in 5 fields at x200 magnification from 5 mice per group (B). Scale bars =  $100 \mu m$ .
- **Fig. S9. FAP deficient mice were resistant to HFD induced obesity.** Change in live body weight following the start of diet (A, C) and autopsy body weight at 20 weeks of diet (HFD; Specialty Feeds) in WT and FAP gko mice (B) and 17 weeks of in-house HFD in WT and FAPgki mice (D). Individual replicates (A, C). Mean ± SEM. n = 10-12 female (A, B) or 12-14 male (C, D) mice per group. \*p<0.05 versus genotype-matched controls using Mann-Whitney U test.
- Fig. S10. Glucose tolerance AUC was not correlated with body weight or adiposity in FAP deficient mice. Regression analysis of GTT AUC versus body weight (A) or adiposity (total fat pad weight : body weight; B) of WT and FAP gko mice. Regression analysis of

GTT AUC versus body weight of WT and FAPgki mice (C). These data derive from 20 weeks of in-house HFD. Scatter plots show individual replicates and line of best fit. n=10-12 female (A, B) or 5 male (C) mice.

Fig. S11. Circulating active GLP-1 and intrahepatic *Pparα* and *Srebp1c* mRNA were unaltered by FAP deficiency in mice. GLP-1 (7-36) levels in plasma following overnight fasting (basal) and at 15 minutes after a glucose challenge at 20 weeks of diet (A). mRNA expression of *Pparα* (B, D) and *Srebp1c* (C, E) relative to 18S house keeping gene at 20 weeks of HFD. Individual replicate mice. Mean ± SEM. n=5-6 female (A), or n=10-12 female (B, C) or male (D, E) mice per group.

Fig. S12. Intrahepatic mRNA expression of *Dpp4* gene family members was not altered in FAP gko mice compared to WT mice. Transcripts relative to housekeeper gene *18S* RNA. n=10-12 female mice per group, fed either chow (A) or HFD (B) for 20 weeks. Individual replicate mice.

Fig. S13. *Dpp9* expression and its association with *Dpp4* and *Chrebp* expression. *Dpp9 mRNA* expression was downregulated in HFD mice compared to chow mice (A), and was correlated with *Dpp4* (B) and *Chrebp* mRNA (C), regardless of mouse genotype. Transcripts relative to housekeeper gene *18S*. Basal mRNA expression from overnight fasted mice. n=10-12 female mice per group. Scatter plots show individual replicates (A-C). Mean ± SEM (A). Line of best fit for each genotype (B, C). \*p<0.05 by Mann-Whitney U test.

Fig. S14. CD36 immunopositivity in hepatocytes was less in FAPgki than WT liver. Immunofluorescence of CD36 on mouse liver sections, with immunopositivity from cells having morphological characteristics of hepatocytes and macrophages (A). Quantitation of four fields per section at 200x magnification in arbitrary units of total intensity of fluorescent signal at 488 nm absorbance. Scale bars = 30  $\mu$ m.

Fig. S15. FAP depletion was not associated with changes in physical activity or energy expenditure. Physical activity (A) and energy expenditure (B) were measured by indirect calorimetry at 12 weeks of HFD (in-house diet). Mean ±SEM, n=6-8 female mice per group. \*p<0.05 versus genotype-matched controls using Mann-Whitney U test.

**Fig S16. Proposed model of effects of FAP deficiency in mice.** Effects of FAP implied from these studies of FAP deficient mice, pathways of FAP action that were identified, and molecules that are potentially involved.

## **Supplementary Material References**

- [1] Wang XM, Yu DMT, McCaughan GW, Gorrell MD. Fibroblast activation protein increases apoptosis, cell adhesion and migration by the LX-2 human stellate cell line. Hepatology 2005;42:935-945.
- [2] Keane FM, Yao T-W, Seelk S, Gall MG, Chowdhury S, Poplawski SE, et al. Quantitation of fibroblast activation protein (FAP)-specific protease activity in mouse, baboon and human fluids and organs. FEBS Open Bio 2014;4:43-54.
- [3] Osborne B, Yao T-W, Wang XM, Chen Y, Kotan LD, Nadvi NA, et al. A rare variant in human fibroblast activation protein associated with ER stress, loss of function and loss of cell surface localisation. Biochim Biophys Acta 2014;1844:1248-1259.
- [4] Lo L, McLennan SV, Williams PF, Bonner J, Chowdhury S, McCaughan GW, et al. Diabetes is a progression factor for hepatic fibrosis in a high fat fed mouse obesity model of non-alcoholic steatohepatitis. J Hepatol 2011;55:435-444.
- [5] Henderson JM, Polak N, Chen J, Roediger B, Weninger W, Kench JG, et al. Multiple liver insults synergize to accelerate experimental hepatocellular carcinoma. Scientific Reports 2018;8:10283.

# Supplementary Material

Fibroblast activation protein enzyme deficiency prevents liver steatosis, insulin resistance and glucose intolerance and increases fibroblast growth factor-21 in diet induced obese mice

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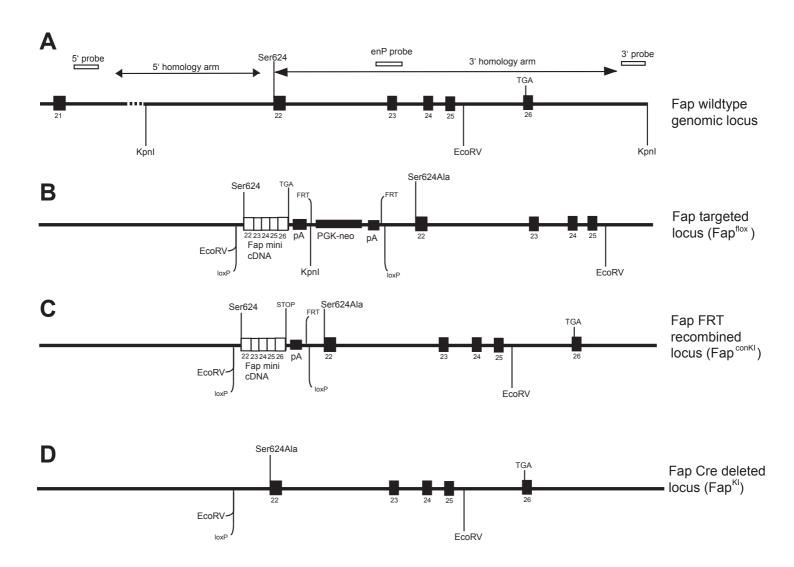


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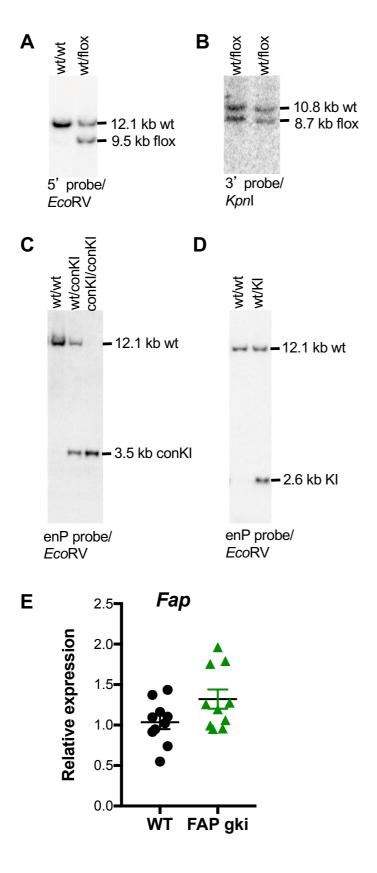


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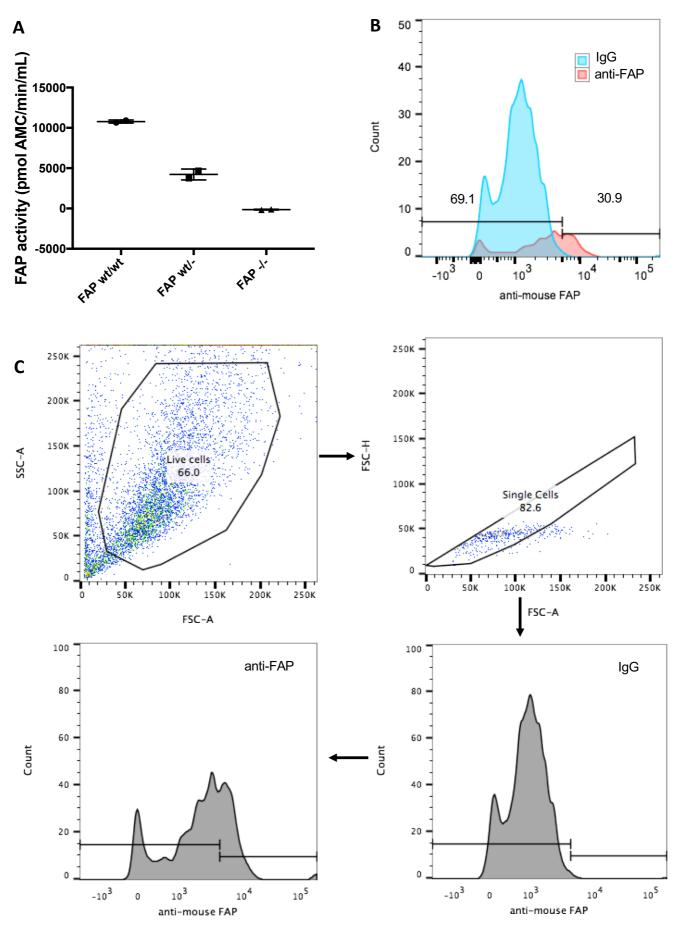


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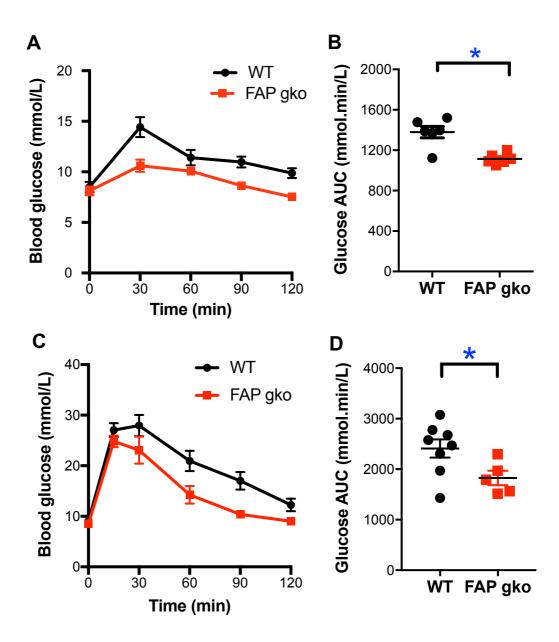


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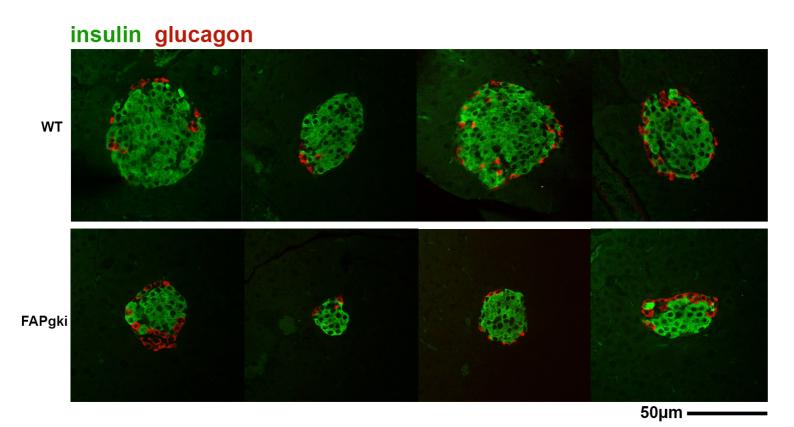


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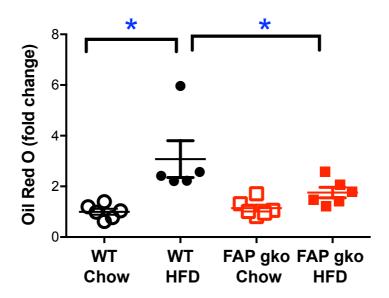


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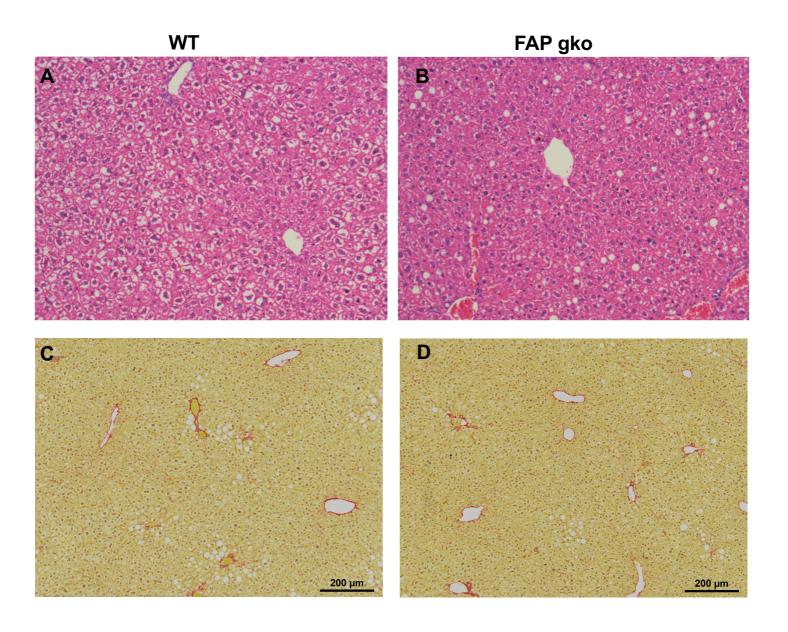


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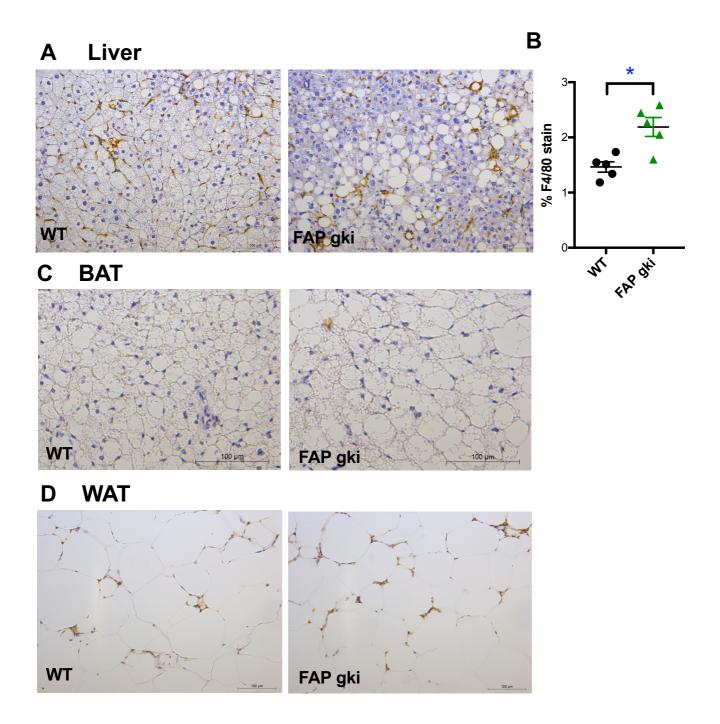


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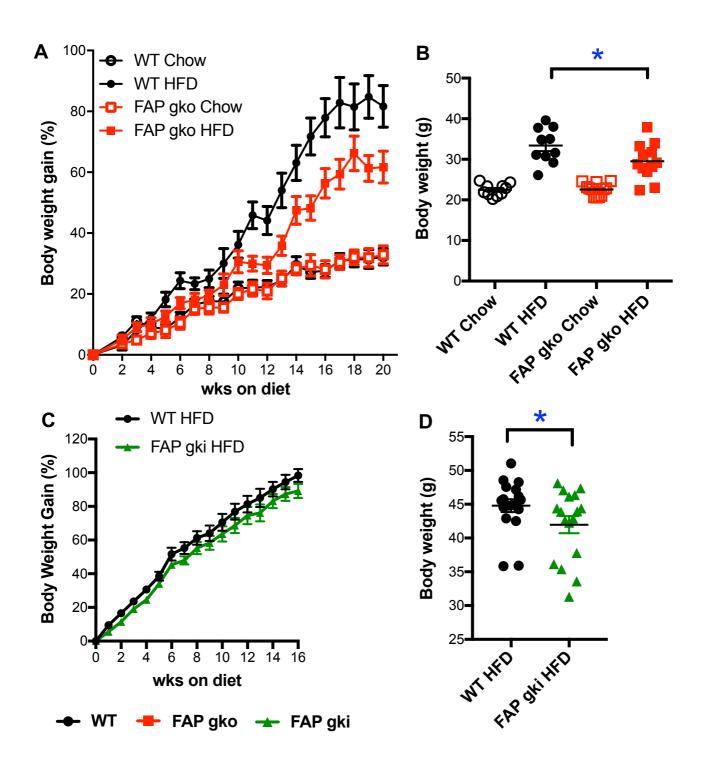


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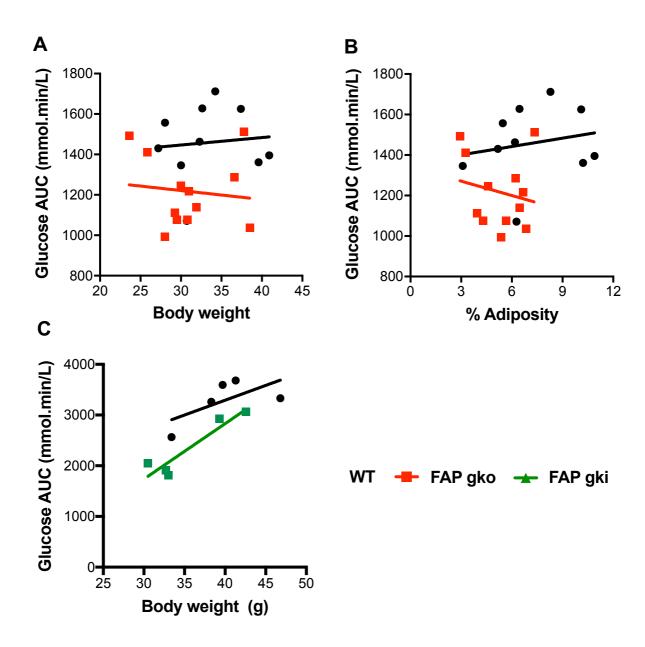
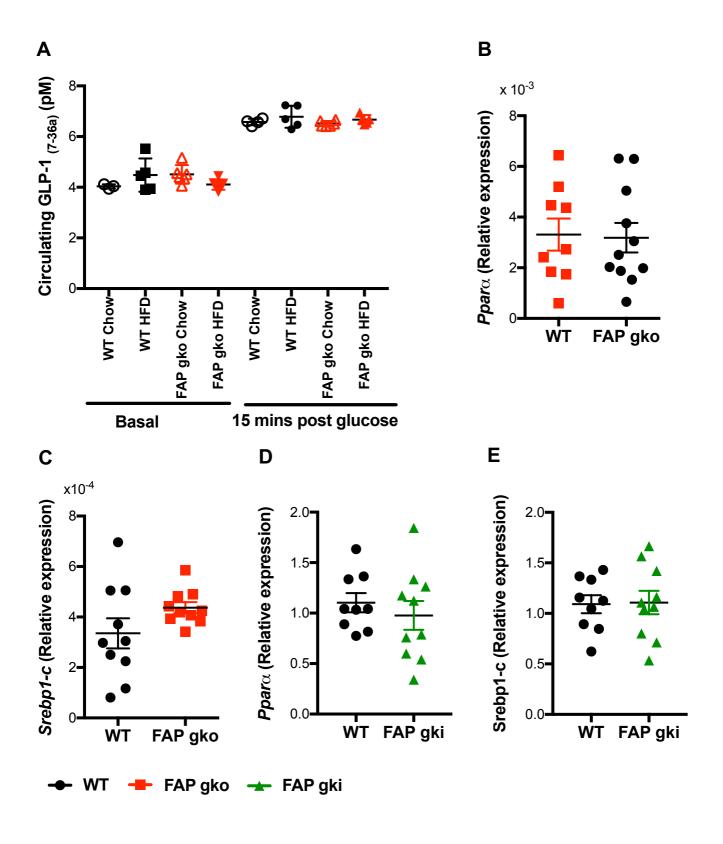
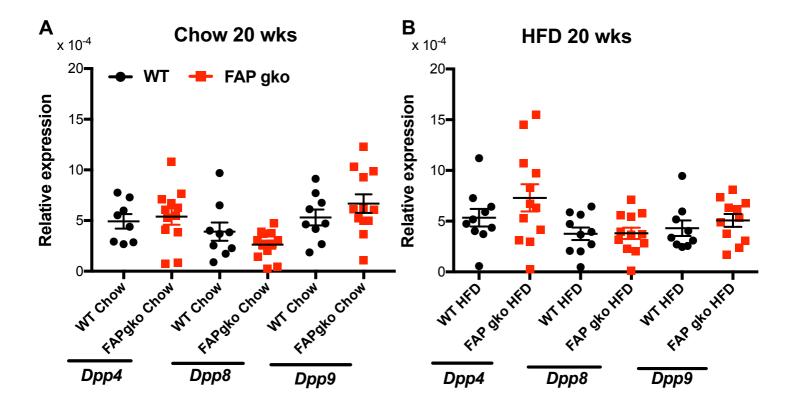


Fig. S10



**Fig. S11** 



**Fig. S12** 

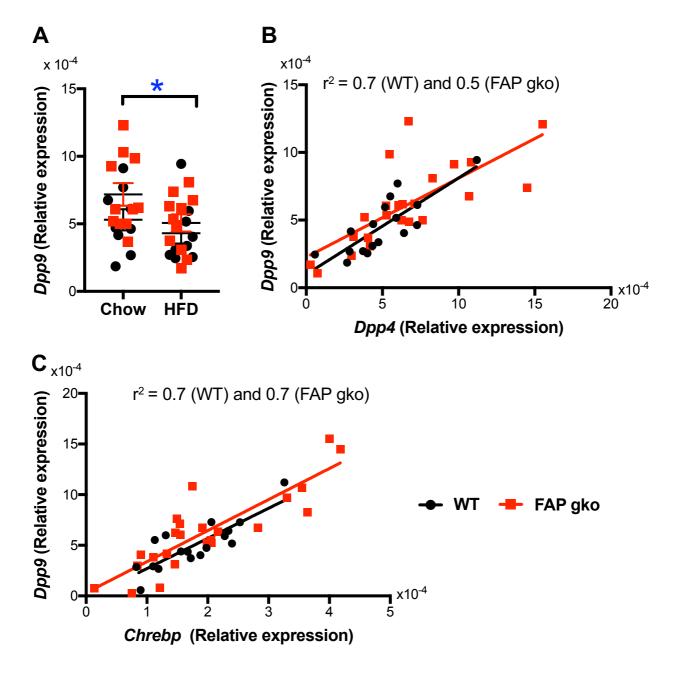


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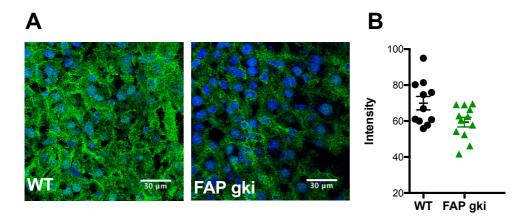


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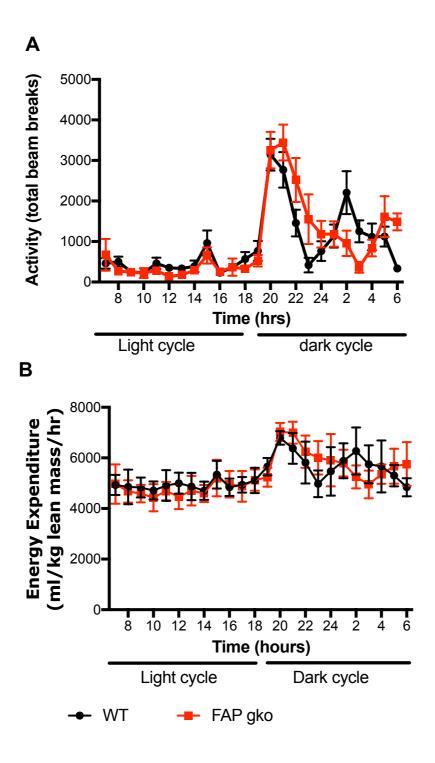


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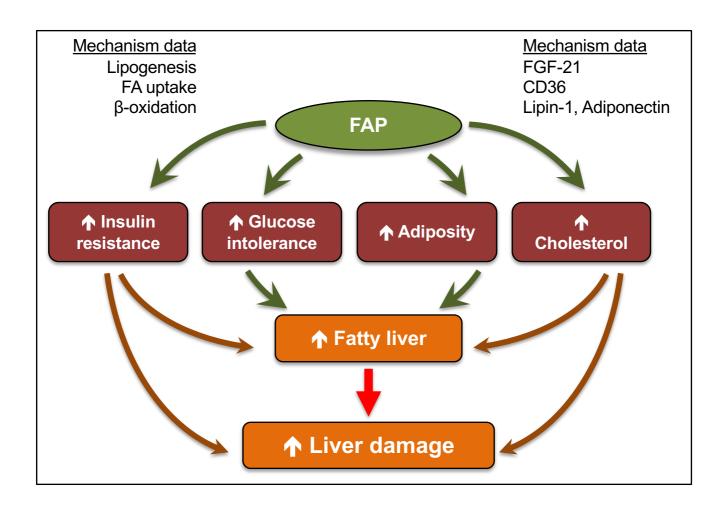


Fig. S16