

## Supplementary Figure Legends

### Supplementary 1 Recapitulation of a published CRC study

As a proof-of-concept case study, GeMSTONE (1) recapitulated every step in the original Colorectal-Cancer prioritization workflow<sup>1</sup>, (2) rescuing 26 out of 28 candidate variants from the ~30,000 variants in the raw whole exome sequencing dataset, and (3) hitting all hereditary CRC and CRC GWAS variants.

### Supplementary 2 ALS study workflow

Only simple filters were applied for the ALS case study—specifically, variants were required to be of high quality with (1) genotype quality  $\geq 40$ , (2) putative damaging effect (frameshift, in-frame indel, non-synonymous, and essential splice site variant) on protein coding transcript, (3) absent from control cohort and (4) rare in the general population defined by an allele frequency  $\leq 0.05\%$ . Two different co-segregation analyses were performed (Dominant and Recessive), in line with ALS inheritance studies<sup>2</sup>. Recurrence filters and family constraints were applied, requiring variant occurrence in at least 2 samples and at most 5 samples, in order to reduce false positives. To target for high-risk germline mutations, variants were required to co-segregate in at least one family.

### Supplementary 3 Mutation enrichment analysis on protein-protein interaction interface during different stages of GeMSTONE pipeline

**3a.** GeMSTONE's performance on the ALS study showed 122 candidate variants with a significantly increased enrichment on protein-protein interaction (PPI) interface domains and residues derived from co-crystal structures<sup>3,4</sup>. This trend is expected as a positive control based on disease mutation enrichment on PPI interfaces<sup>3,4</sup>. 54,668 HGMD disease mutations (green) were plotted alongside GeMSTONE filtering results (blue), with case-only variants and naïve filtering presented as a negative control (grey).

**3b.** The same trend holds for GeMSTONE performance on the CRC study, where F1, F2, F3 and F5 represent variants kept after different sets of filters were applied. Each F-label corresponds to the first, second, third and fifth blocks of filters from **Supplementary Figure 1** respectively.

### Supplementary 4 GeMSTONE visualization page

Through the results page of the GeMSTONE portal, users can visualize their variant statistics. The interactive menu atop the page shows the number of variants by chromosomal region, with variant quality, allele frequency (in ExAC), read depth, and insertion deletion length histograms, as well as tsv ratio and variant type comparisons. By clicking on a specific chromosome or using the filter toggle on the top left, the user can interactively explore these statistics at different resolution levels.

## Supplementary Note 1

Overall, most tools we compared GeMSTONE to accept VCF and pedigree files as inputs, and can perform routine filtering on quality control and variant consequence (**Figure 1, Raw Data Input and Prioritization**). However, GeMSTONE stands out as a more comprehensive tool by including annotations at the variant, gene, pathway, and network level (**Figure 1, Knowledge-based Annotation and Prioritization**) and flexible co-segregation analysis using different inheritance models for potential germline mutation prioritization (**Figure 1, Inheritance Models**).

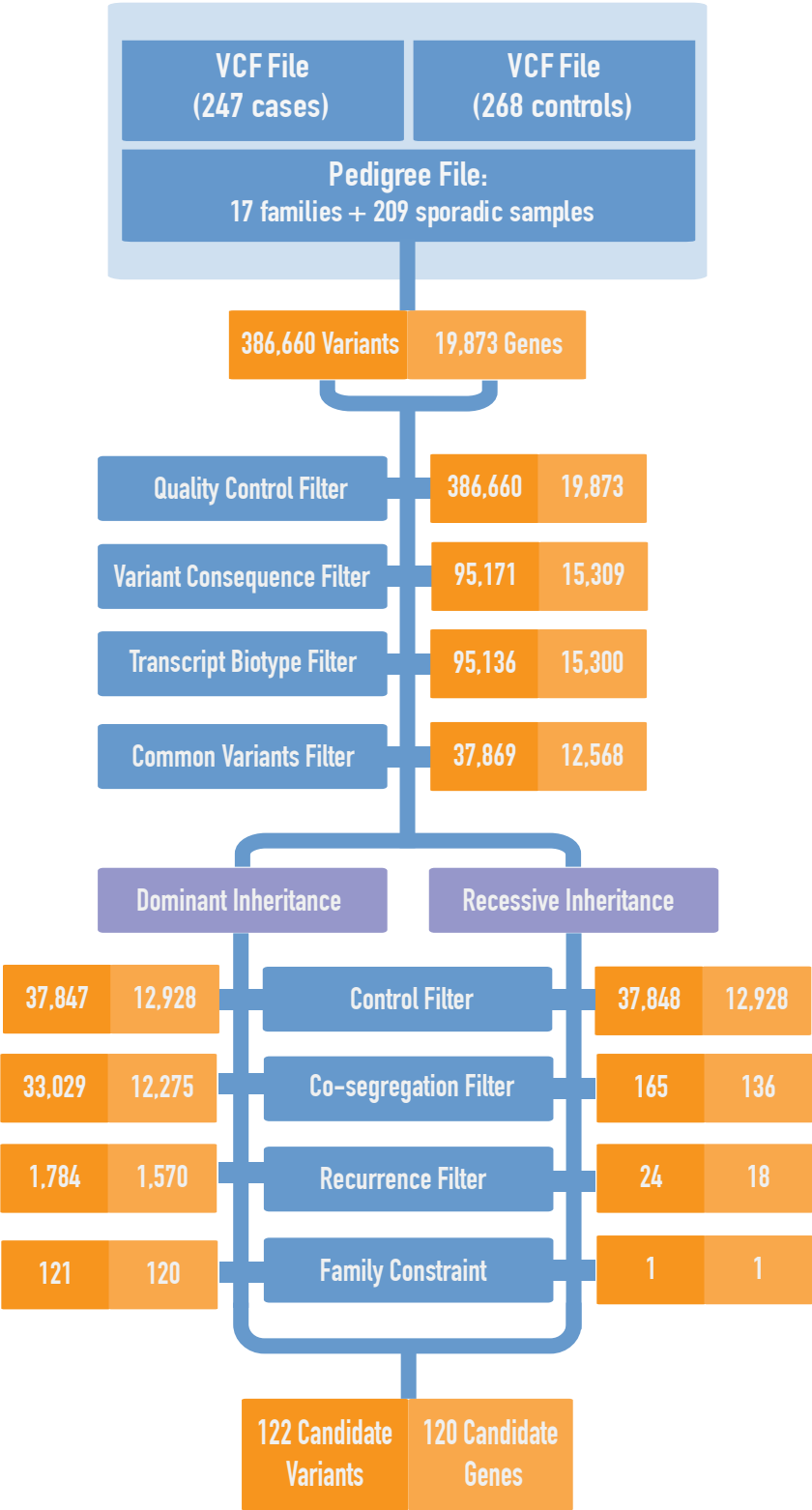
A keystone of GeMSTONE is the ‘recipe’ file (**Figure 1, Reproducibility**), which records all workflow parameters in a single file that can be shared and uploaded onto the site to reproduce a previous run. The recipe file can be used to (1) replicate results by rerunning the same workflow on the same dataset, (2) process new data with a known workflow or (3) modify parameters in a known workflow to evaluate study design. This approach has the potential to bring more transparency and openness to the bioinformatics community by enhancing the reproducibility of large-scale genomic studies.

## References

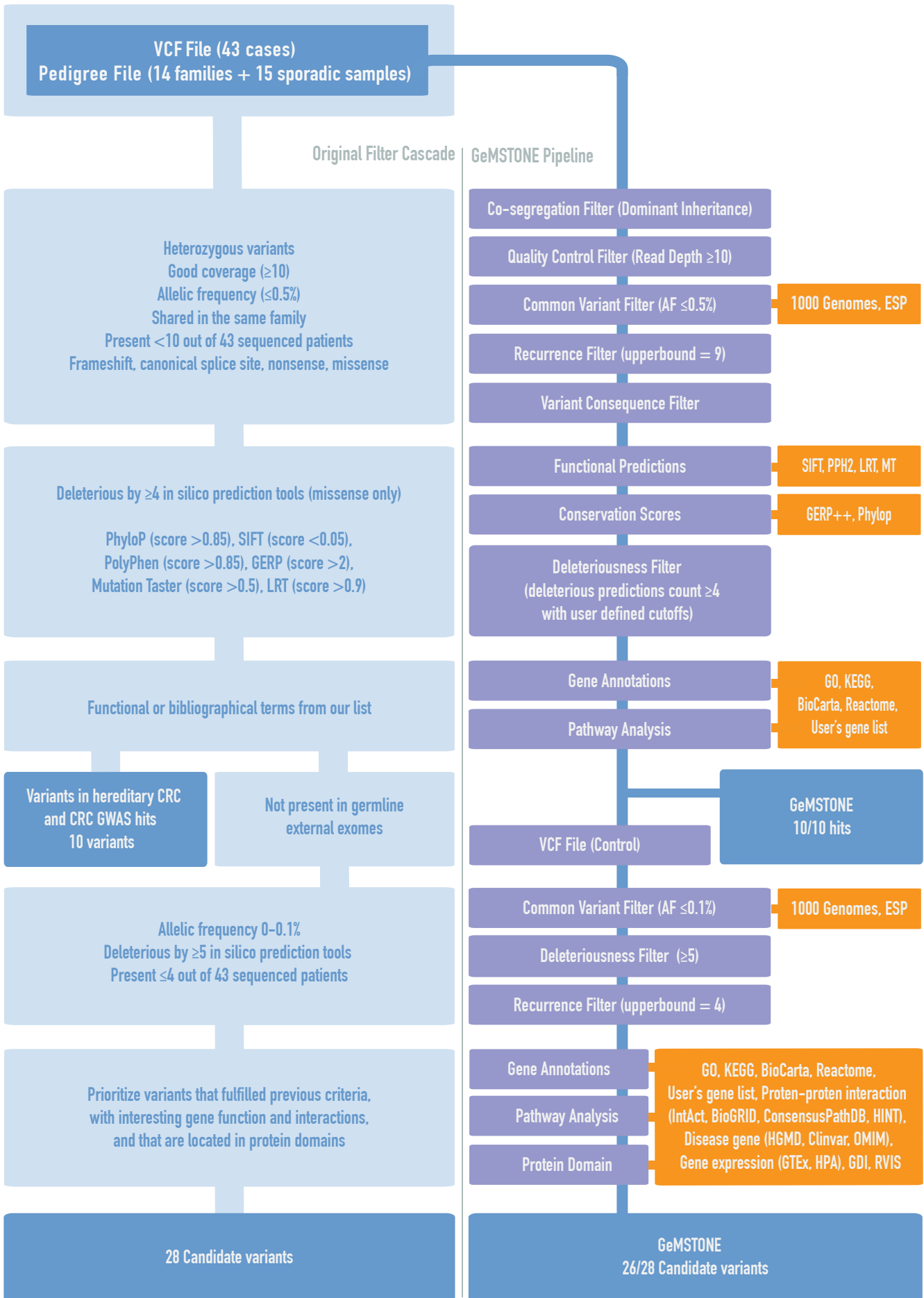
1. Esteban-Jurado, C. et al. *Genet Med* **17**, 131-142 (2015).
2. Cirulli, E.T. et al. *Science* **347**, 1436-1441 (2015).
3. Wang, X. et al. *Nat Biotechnol* **30**, 159-164 (2012).
4. Das, J. et al. *Hum Mutat* **35**, 585-593 (2014).
5. Aleman, A., Garcia-Garcia, F., Salavert, F., Medina, I. & Dopazo, J. *Nucleic Acids Res* **42**, W88-93 (2014).
6. Robinson, P.N. et al. *Genome Res* **24**, 340-348 (2014).
7. Smedley, D. et al. *Bioinformatics* **30**, 3215-3222 (2014).
8. Li, M.J. et al. *Hum Mutat* **36**, 496-503 (2015).
9. Ge, D. et al. *Bioinformatics* **27**, 1998-2000 (2011).
10. Sifrim, A. et al. *Genome Med* **4**, 73 (2012).
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12. Shetty, A.C. et al. *BMC Bioinformatics* **11**, 471 (2010).

Supplementary Figures

Supplementary 1 Recapitulation of a published CRC study

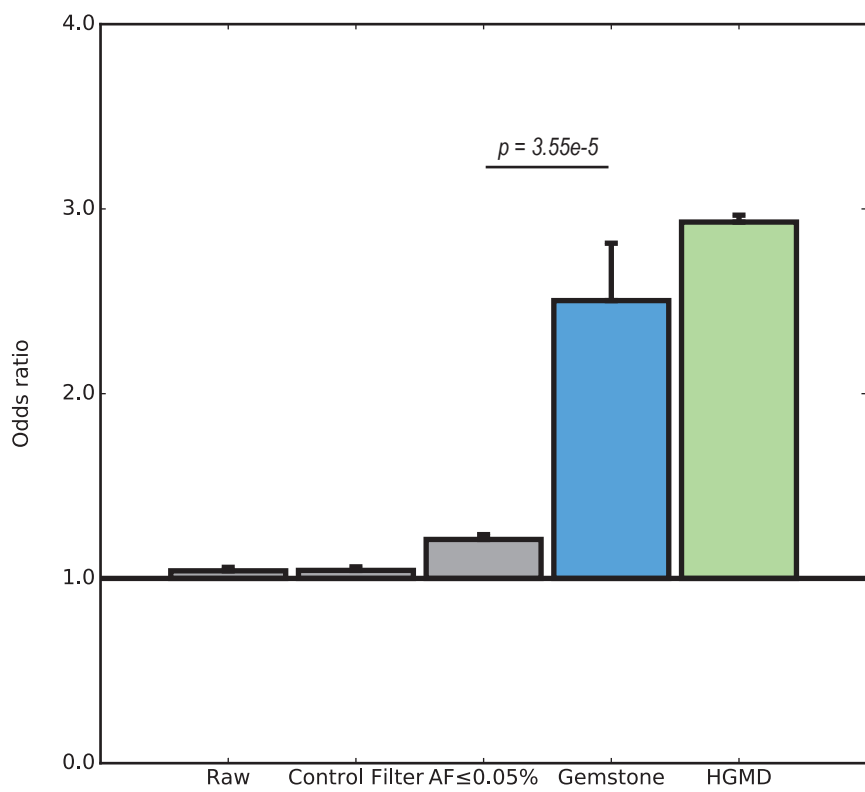


## Supplementary 2 ALS study workflow

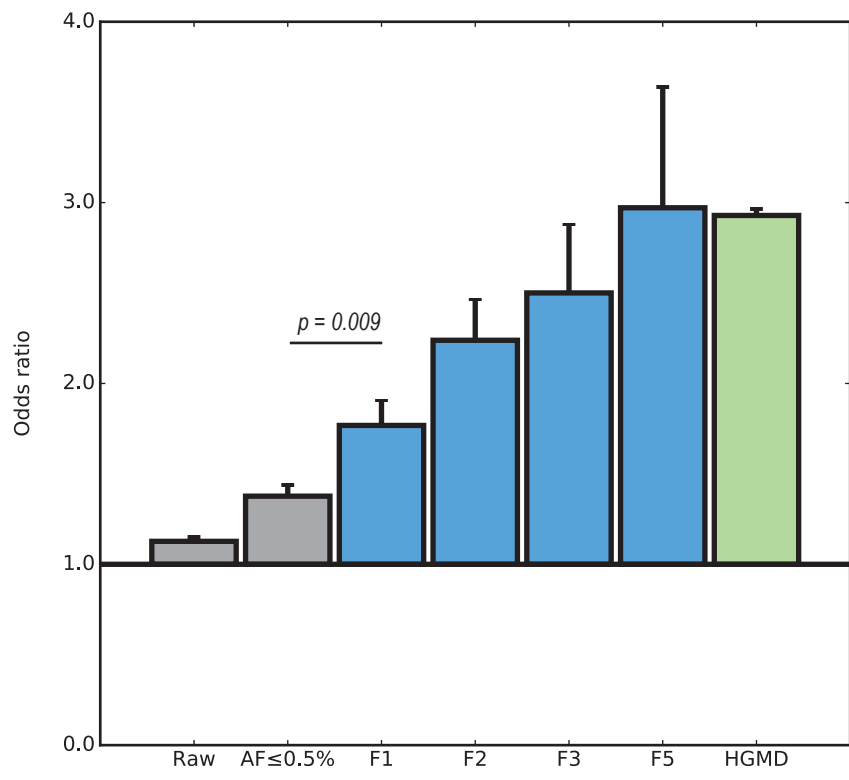


**Supplementary 3** Mutation enrichment on protein interface during different stages of GemSTONE pipeline

**a** Mutations At Protein-Protein Interaction Interface from the ALS Dataset



**b** Mutations At Protein-Protein Interaction Interface from the Colo-Rectal Cancer Dataset

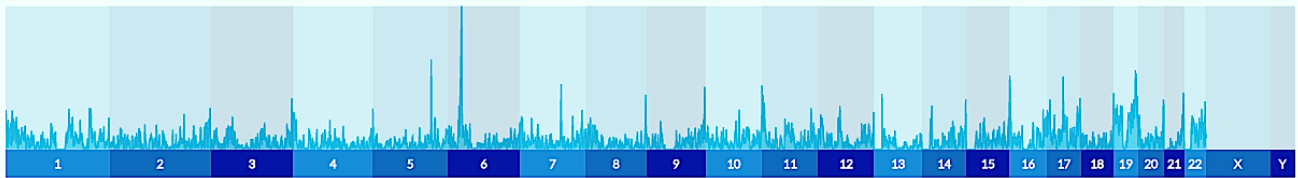


# GeMSTONE visualizer

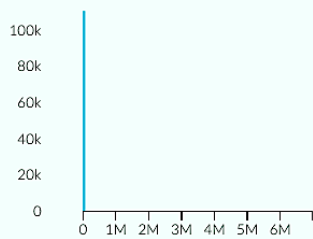
Yu Lab

Unfiltered **Filtered**

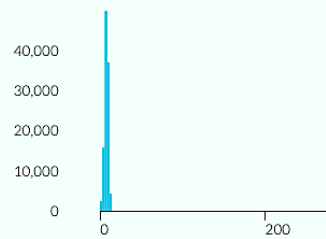
Variant Density



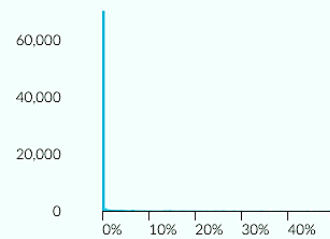
Variant Quality



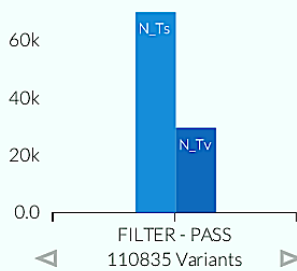
Mean Depth



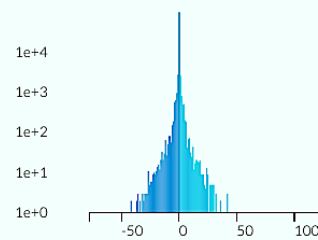
Allele Frequency Spectrum



TSTV Ratio = 2.36



Insertion/Deletion Lengths



Variant Type

