

1 **Supplementary Information S2**

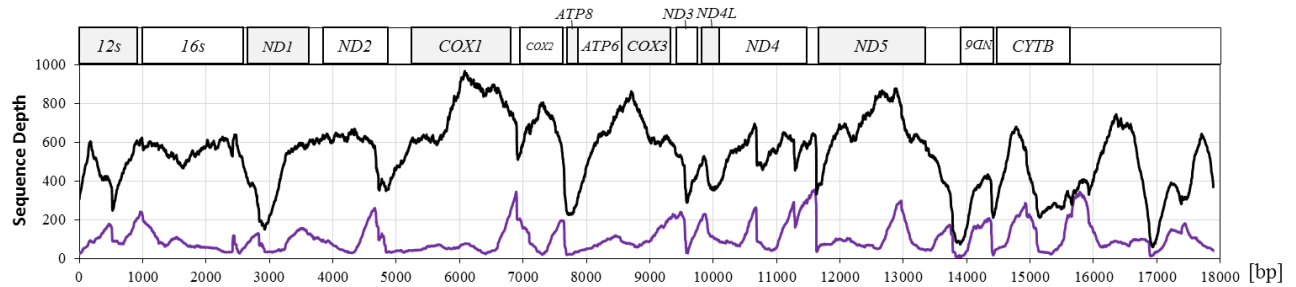
2 DNA and LR-PCR products were amplified for *CO1* with the primers 5'-
3 CGCCTGTTTAyCAAAAACAT -3' and 5'- GAGTCCGTcGcNAndGGTTG -3' (1) and for the partial *CYTB*
4 with the primers 5'- TACCATGAGGACArATrTCnTTyTG -3' and 5'- GGrATdGAdCGdAGrATdGCrTAnGC
5 -3'. Partial *ND6-CYTB* of *Quasipaa yei* was amplified with the primers 5'- AACGCAGCACTCTTGTGACC
6 -3' and 5'-GGrATdGAdCGdAGrATdGCrTAnGC-3'. PCR was carried out in a 13.5 µl PCR reaction of 20
7 ng of DNA, 0.2 µl *taq* DNA Polymerase and 0.1 µl *pfu* (TaKaRa), 0.8 µl 2.5 mM dNTP (TaKaRa), and 0.8
8 µl 5 µM forward and reverse primers (Invitrogen). PCR was conducted at 94°C for 5 min, followed by 35
9 cycles of 94°C for 30 s, 50°C for 30 s, and 72°C for 2 min, and with a final extension at 72°C for 10 min
10 and holding at 10°C. Sequencing PCR is adding 20 µg of PCR product, 2 µl ABI BigDye3 diluted 6×, and
11 1.5 µM primer (2 µl) and conducted 25 cycles of 96 °C for 10 s, 50°C for 5 s, and 60°C for 4 min and
12 holding at 4°C. Sequencing used an ABI-3730 capillary DNA sequencer. The results were assessed by
13 using SeqMan v.7 (DNASar, WI, USA).

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15 **Supplementary Information S3**

16 The Ion Torrent platform yields uneven and biased coverage across MtGs as well as nuclear genome (2).
17 We discovered that there are exists amount of small fragments in all the libraries. For example, there are
18 27.65% (8,538/30,883) reads sequenced incompletely in *Babina adenopleura*. We use the following
19 standard to check sequence incompleteness. During synthesis-by-sequencing, extension is from sequence
20 primer across the mtDNA to the end of adapter P1 (Figure S4). When sequence incomplete, generated
21 fragment will be short and do not have the P1 region. Library *Babina adenopleura* had 27.65% reads that
22 do not have a region similar to the P1 adaptor. Such reads involve low-coverage regions that tend to have
23 gaps (Figure S3: purple). This indicated the sequence incomplete is one of the reason result of coverage
24 unevenness. In addition, sequence chip is vital in improving sequence-quality. Difference batches of chips

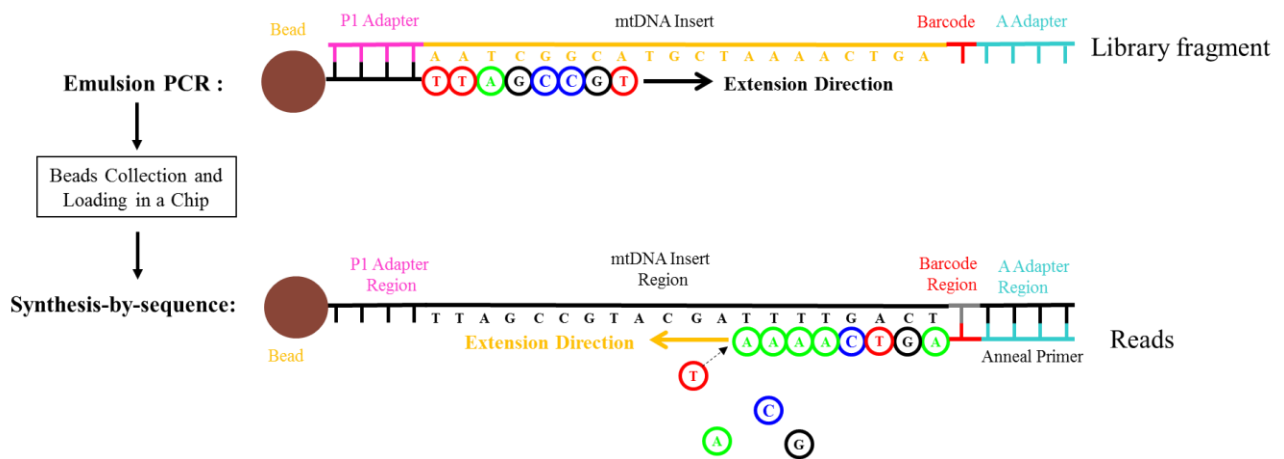
25 vary in sequence-quality (3) and we also recommend Ion 316 Chip v2. To reduced coverage unevenness,
 26 another sequence platform Illumina is good choice since human MtG was coverage evenness in it (4).
 27



28
 29 Figure S3. Coverage distributions of *Babina adenopleura* for different read types. Purple line represents
 30 coverage using the reads without adaptor sequences; black line showing the overall coverage by using all
 31 the reads.

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33 **Supplementary Information S4**



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 35 Figure S4. PCR extension during emulsion PCR and Synthesis-by-sequence. In emulsion PCR, the
 36 fragment is extend from P1 adapter to end of A adapter according to the library sequence. In the
 37 Synthesis-by-sequence, the extend is from A adapter to P1 adapter to get signal base by base at a
 38 constant temperature.

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40 **REFERENCES**

- 41 1. Che, J., Chen, H.M., Yang, J.X., Jin, J.Q., Jiang, K., Yuan, Z.Y., Murphy, R.W. and Zhang, Y.P. (2012)
42 Universal COI primers for DNA barcoding amphibians. *Mol. Ecol. Resour.*, **12**, 247–258.
- 43 2. Quail, M.A., Smith, M., Coupland, P., Otto, T.D., Harris, S.R., Connor, T.R., Bertoni, A., Swerdlow, H.P.
44 and Gu, Y. (2012) A tale of three next generation sequencing platforms: comparison of Ion Torrent,
45 Pacific Biosciences and Illumina MiSeq sequencers. *BMC Genomics*, **13**, 341.
- 46 3. Wang, Y., Wen, Z., Shen, J., Cheng, W., Li, J., Qin, X., Ma, D. and Shi, Y. (2014) Comparison of the
47 performance of Ion Torrent chips in noninvasive prenatal trisomy detection. *J. Hum. Genet.*, **59**, 393–396.
- 48 4. Cui, H., Li, F.Y., Chen, D., Wang, G.L., Truong, C.K., Enns, G.M., Graham, B., Milone, M., Landsverk,
49 M.L., Wang, J. *et al.* (2013) Comprehensive next-generation sequence analyses of the entire
50 mitochondrial genome reveal new insights into the molecular diagnosis of mitochondrial DNA disorders.
51 *Genet. Med.*, **15**, 388-394.

52 **Supplementary Table S2**53 **Table S2.** MtDNA read-distribution and gaps in the library of 33 mixed samples (Samples ranked by read-number).

Species	Read-number*	Gap position	Reference	Family
<i>Limnonectes bannaensis</i>	19267	Circle	This study	Dicroglossidae
<i>Megophrys palpebralespinosa</i>	11315	Circle	This study	Megophryidae
<i>Leptobrachium ailaonicum</i>	6994	ND6: 13990-14016	This study	Megophryidae
<i>Rhacophorus translineatus</i>	6399	Non-coding region between ND5 and 12s rRNA	This study	Rhacophoridae
<i>Kurixalus odontotarsus</i>	6354	Circle	This study	Rhacophoridae
<i>Leptobrachium liui</i>	5975	near ND6	This study	Megophryidae
<i>Glandirana tientaiensis</i>	5659	ND6: 16006-16090	NC_025226	Ranidae
<i>Bufo pageoti</i>	5646	Circle	This study	Bufoidea
<i>Hylarana taipehensis</i>	5629	1. Non-coding region next to ND1: 4503-4542; 2. Non-coding region next to ND2: 6426-6463; 3. ND6: 14713-14738	This study	Ranidae
<i>Babina adenopleura</i>	5181	Circle	This study	Ranidae
<i>Kaloula borealis</i>	5011	ND6: 13721-13741	This study	Microhylidae
<i>Leptolalax oshanensis</i>	4940	Circle	This study	Megophryidae
<i>Feihyla vittatus</i>	4596	Almost circle	This study	Rhacophoridae
<i>Occidozyga martensii</i>	4254	Circle	This study	Occidozygidae
<i>Rhacophorus bipunctatus</i>	4122	1. ND6: 13573-13639; 2. Non-coding region between ND5 and 12s rRNA	This study	Rhacophoridae
<i>Ichthyophis bannanicus</i>	3661	Circle	This study	Ichthyophiidae
<i>Liurana alpinus</i>	3583	1. The end of ND4 and its next non-coding region: 12030-12147, 12148-12377; 2. The front of ND5: 12654-	This study	Occidozygidae

		12696; 3. <i>ND6</i> : 14541-14666		
<i>Fejervarya kawamurai</i>	2802	Circle	This study	Dicroglossidae
<i>Raorchestes longchuanensis</i>	2518	Absent <i>ND5</i>	This study	Rhacophoridae
<i>Bufo stejnegeri</i>	2371	1. <i>ND2</i> : 4135-4147; 2. <i>apt6</i> : 8135-8161; 3. <i>ND4</i> : 11245-11257; 4. <i>ND6</i> : 13557-13660	This study	Bufoidea
<i>Leptobranchium chapaense</i>	2103	Almost circle	This study	Megophryidae
<i>Oreolalax xiangchengensis</i>	2032	Circle	This study	Megophryidae
<i>Hyla chinensis</i>	1840	1. Non-coding region next to <i>apt8</i> : 8088-8165	This study	Hylidae
<i>Scutigera wuguanfui</i>	1767	1. <i>ND2</i> : 4924-4950, 5230-5530; 2. Non-coding region next to <i>ND5</i> : 14192-14238; 3. <i>ND6</i> : 14445-14474, 14577-14582, 14629-14650	This study	Megophryidae
<i>Parapelophryne scalpta</i>	1690	1. Non-coding region next to <i>ND5</i> : 12103-12244; 2. Non-coding region next to <i>ND6</i> : 14174-14194	This study	Bufoidea
<i>Bufo tibetanus</i>	1434	1. <i>apt6</i> : 8416-8527; 2. The end of <i>ND6</i> : 13787-13922	NC_020048	Bufoidea
<i>Pelophylax plancyi</i>	1344	circle	JF730436	Ranidae
<i>Nanorana maculosa</i>	1324	1. non-coding region: 2789-2812, 2869-2897; 2. The end of <i>ND2</i> : 5202-5292; 3. <i>COX1</i> : 6117-6147, 6239-6339	This study	Dicroglossidae
<i>Bufo gargarizans</i>	1118	1. <i>ND6</i> : 13782-13947; 2. Some gaps in control region	NC_008410	Bufoidea
<i>Hynobius amjiensis</i>	824	Large gaps	NC_008076	Hynobiidae
<i>Pachytriton granulosus</i>	640	Large gaps	This study	Salamandridae
<i>Paramesotriton hongkongensis</i>	153	Large gaps	NC_006407	Salamandridae
<i>Bombina orientalis</i>	2*	Large gaps	NC_006689	Bombinatoridae