# Regulation of sleep plasticity by a thermo-sensitive circuit in *Drosophila*

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### SUPPLEMENTAL INFORMATION



**Fig. S1: Alterations in locomotion and sleep architecture due to increased ambient temperature in** *Drosophila* **males and females.** (**a**) Average number of beam crossings per 30 min over 24 h for wild-type males shifted from 22°C to either 27°C (**a**) or 30°C (**b**). Black arrowheads indicate a delay in evening anticipation observed at either 27°C or 30°C relative to 22°C. Grey arrowhead denotes advanced morning anticipation observed at 30°C, but not at 27°C. (**c**) Average sleep patterns of adult mated female flies shifted from 22°C to 30°C. n = 32. (**d**) Latency to initiate the first day sleep bout in adult males and females shifted from 22°C to 30°C. \*\*\*p < 0.0005, ns – p > 0.05, Kruskal-Wallis test with Dunn's post-hoc test.



### **FIGURE S2**

Fig. S2: Locomotor and sleep patterns of *timeless* null flies during warm conditions. (a-b) Number of beam breaks per 30 min in wild type iso31 controls (n = 31) or *timeless* null (*tim*<sup>ko</sup>) homozygotes (n = 33) during consecutive days at either 22°C or 30°C. Locomotor activity was measured under 12 h light: 12 h dark conditions (white/grey bars) with Zeitgeber Times (ZT) shown below. Data are presented as mean  $\pm$  SEM for each time point. (c) Sleep patterns of *tim*<sup>ko</sup> homozygotes at either 22°C or 30°C. (d) Change in latency to the first day sleep episode in iso31 controls (n = 69) or *timeless* null (*tim*<sup>ko</sup>) homozygotes (n = 64) following a shift from 22°C to 30°C. \*p < 0.05, Mann-Whitney U-test.

## Figure S3



Fig. S3: The Gr28b and Pyrexia thermo-receptors do not mediate PMW. (a) Change in latency to the first day sleep episode in iso31 controls (n = 69),  $Gr28b^{MB03888}$  (n = 27) or *pyrexia*<sup>3</sup> (*pyx*<sup>3</sup>; n = 26) homozygotes following a shift from 22°C to 30°C. All genotypes show a significant increase in latency following an increase in ambient temperature to 30°C. \*\*\*p < 0.0005, compared to a median of zero, Wilcoxon signed rank test. (b) Pan-neuronal knockdown of TrpA1 expressing using a *UAS-TrpA1* RNAi transgene under control of *elav*-GAL4 suppresses PMW. n = 33-120. \*\*\*p < 0.0005, Kruskal-Wallis test with Dunn's post-hoc test.





*ppk*[200871]-GAL4 along the anterior/posterior axis of the adult brain. Right, z-stack projections. Neurons are labeled using a UAS-CD4::TdTom transgene (Red). Synaptic neuropil is labeled using an anti-Bruchpilot (BRP) antibody. Scale bar: 50  $\mu$ m. (**b**) Visualization of mdIV neurons on the adult body wall. White stars: v'ada neurons; blue stars: ddaC neurons. Scale bar: 100  $\mu$ m. (**c**) Acute inhibition of synaptic output (using UAS-*shi*<sup>ts</sup>) from *TrpA1*[SH]- and *ppk*-neurons does not suppress the delay in initiation of night sleep in response to increased ambient temperature. n = 24-77. \*p < 0.05, \*\*\*p < 0.0005, ns – p > 0.05, Kruskal-Wallis test with Dunn's post-hoc test.



**Fig. S5: The** *ppk*-**GAL4 driver stochastically labels AC neurons in the adult** *Drosophila* **<b>brain.** Confocal z-stacks showing *TrpA1*[SH]- and *ppk*-neurons labeled using UAS-CD4::TdTom. Synaptic neuropil is labeled using an anti-Bruchpilot (BRP) antibody. Scale bar: 50 μm. Star: AC neuron cell bodies observed when labeling *ppk*-neurons.





ambient temperature. +>UAS-*shi*<sup>ts</sup> n=25, *mai179>+* n=7, *mai179>shi*<sup>ts</sup> n=21. (**b**) Average sleep patterns of adult *pdf*<sup>01</sup> homozygote males shifted from 22°C to 30°C at ZT0. (**c**) Comparison of PMW between iso31 controls and *pdf*<sup>01</sup> homozygotes, Mann-Whitney test. iso31: n = 79, *pdf*<sup>01</sup>: n = 65. Iso31 p<0.0001, *pdf*<sup>01</sup> p=0.0007, Wilcoxon signed rank test. (**d**) GRASP between *pdf*- and *ppk*-neurons. Scale bar: 15  $\mu$ m. (**e**) Change in latency to the first night sleep episode in *4.1m* > shi<sup>ts</sup> males and associated controls. n = 37-53. \*\*p < 0.0005, ns – p > 0.05, Kruskal-Wallis test with Dunn's post-hoc test.