

Supplementary materials for Bayesian inference of agent-based models: a tool for studying kidney branching morphogenesis

Ben Lambert^{1,*}, Adam L. MacLean^{2,3,*}, Alexander G. Fletcher^{2,4},
Alexander N. Coombes^{5,6}, Melissa H. Little^{6,7}, Helen M. Byrne^{2,†}

¹Department of Zoology, University of Oxford, Oxford, UK

²Mathematical Institute, University of Oxford, Andrew Wiles Building,
Woodstock Road, Oxford, UK

³Current address: Department of Mathematics, University of California,
Irvine, Irvine CA, USA

⁴Current address: Bateson Centre, University of Sheffield, Sheffield, UK

⁵Department of Anatomy & Neuroscience, University of Melbourne,
Melbourne 3010 VIC, Australia

⁶Murdoch Childrens Research Institute, Flemington Rd, Parkville,
Melbourne, 3052 VIC, Australia

⁷Department of Paediatrics, The University of Melbourne, Melbourne,
3010 VIC, Australia *These authors contributed equally.

†email: helen.byrne@maths.ox.ac.uk

1 Appendix

In this section we present results of an analysis conducted to assess the sensitivity of branching to the initial distribution of epithelial cells. We use images from two other experiments presented in Watanabe & Costantini (2004) to determine the shape of the initial bulk of epithelium that we use in simulations. We then perform the same simulations as described in the main text (see Table 1). These also reveal that GDNF-stimulated proliferation coupled with anisotropic cell division (and chemotaxis) are needed to recapitulate *ex vivo* branching.

Similarly Figs. S3 and S4 (analogous to Fig. 6 in the main text) show how branching depends on a finely-tuned GDNF proliferation switch for simulations initialised with the data from the other two explant experiments in Watanabe & Costantini (2004).

| Parameter | Description | All experiments | 2,3 | 4,5A | 4,5B | 4,5C | 4,5D | 4,5E |
|--------------|---|----------------------|-------------|-------------|-----------|-----------|-----------|------------|
| L | domain width and height (cell diameters) | 400 | - | - | - | - | - | - |
| d_g | non-dimensional diffusion coefficient | 0.006 | - | - | - | - | - | - |
| g_∞ | non-dimensional GDNF at boundaries | 1 | - | - | - | - | - | - |
| p_{move} | probability of selecting move vs division | 0.3 (varied in AABC) | 0.3 | 0.3 | 0.3 | 0.3 | 0.3 | 0.3 |
| c_1 | GDNF-stimulated division switch parameter | -40 to 10 | -25 | 10 | -25 | 10 | -25 | 10 |
| c_2 | GDNF-stimulated division switch parameter | 0 to 400 | 120 | 0 | 120 | 0 | 120 | 0 |
| β_m | chemotaxis GDNF sensitivity parameter | 0 to 200 | 200 | 0 | 0 | 200 | 200 | 0 |
| β_{cd} | ACD GDNF sensitivity parameter | 0 to 200 | 200 | 0 | 0 | 0 | 0 | 200 |
| | | 4,5F | 4,5G | 4,5H | 6A | 6B | 6C | 7 |
| p_{move} | probability of selecting move vs division | 0.3 | 0.3 | 0.3 | 0.3 | 0.3 | 0.3 | 0 to 1 |
| c_1 | GDNF-stimulated division switch parameter | -25 | 10 | -25 | -25 | -25 | -25 | -40 to -20 |
| c_2 | GDNF-stimulated division switch parameter | 120 | 0 | 120 | 400 | 120 | 20 | 40 to 280 |
| β_m | chemotaxis GDNF sensitivity parameter | 0 | 200 | 200 | 200 | 200 | 200 | 200 |
| β_{cd} | ACD GDNF sensitivity parameter | 200 | 200 | 200 | 200 | 200 | 200 | 200 |

Table S1: Summary of dimensionless parameter values used in each simulation. “ACD” indicates “anisotropic cell division”.

References

Watanabe, Tomoko, & Costantini, Frank D. 2004. Real-time analysis of ureteric bud branching morphogenesis in vitro. *Developmental Biology*, **271**(1), 98–108.

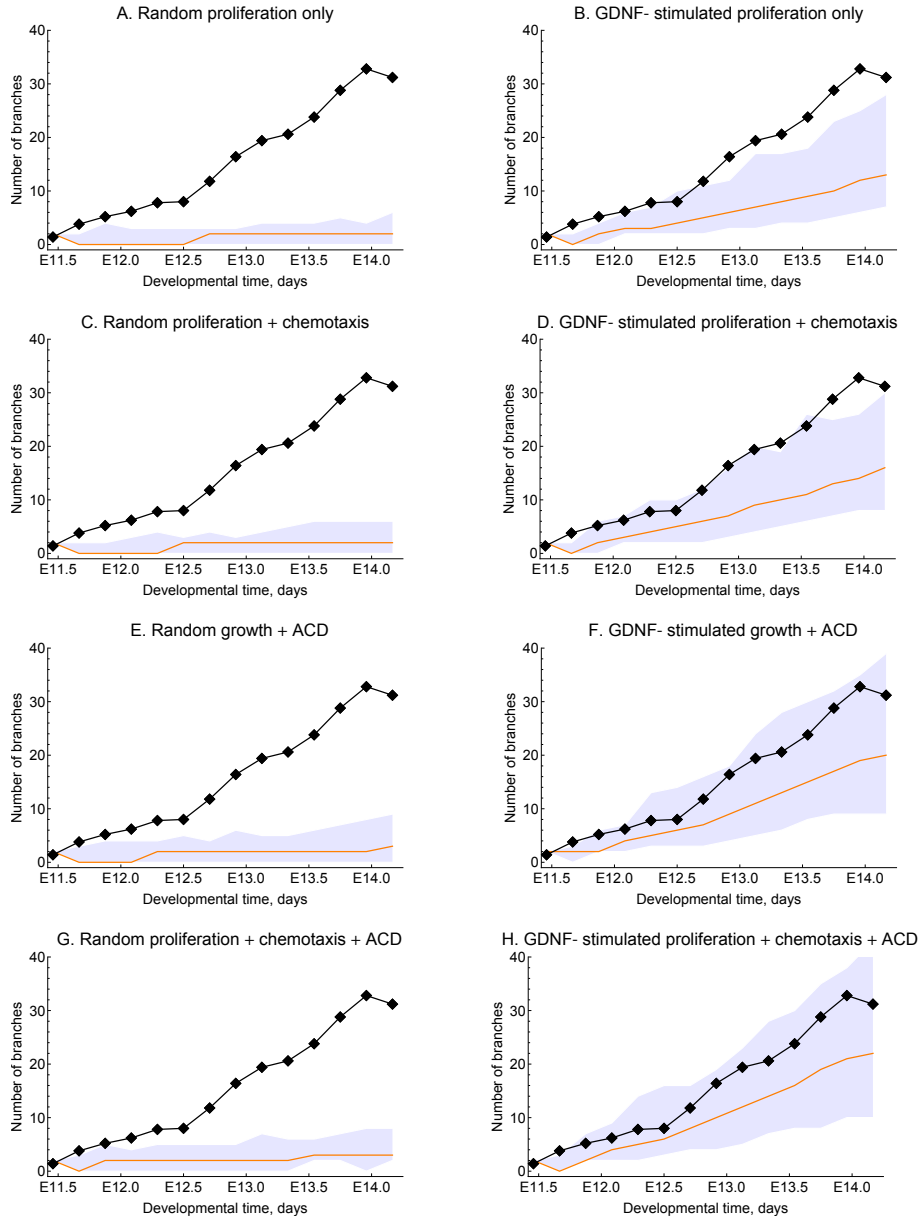


Figure S1: The effect of different GDNF signalling mechanisms on explant branching for another explant experiment. In each panel the black line and points represent the evolution of branches from an explant experiment in (Watanabe & Costantini, 2004); the orange line represents the mean branching observed by model simulation (n= 200) and the shaded region indicates the 95% confidence interval. “ACD” indicates “anisotropic cell division”. The parameter values used in each case are the same as for Fig. 4 in the main text.

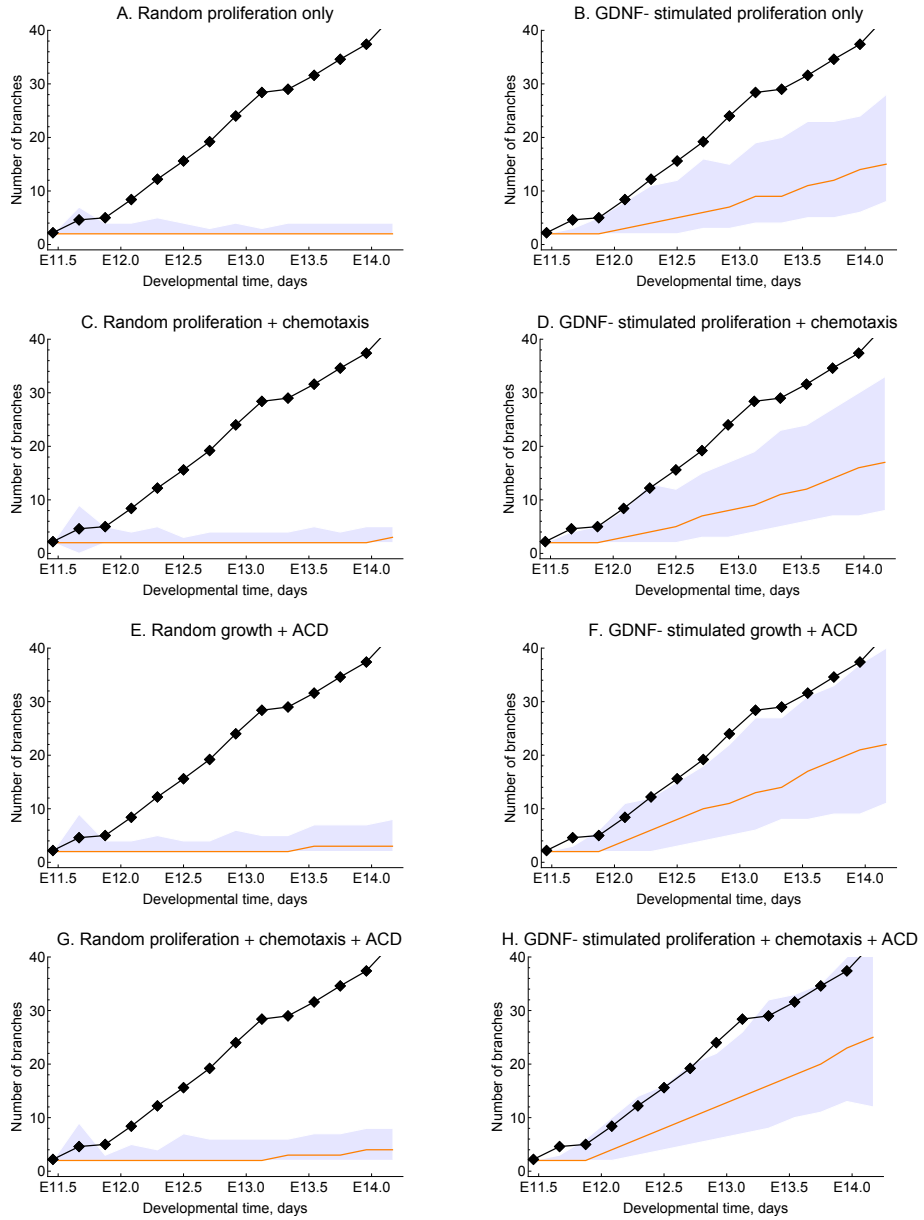


Figure S2: The effect of different GDNF signalling mechanisms on explant branching for a further explant experiment. In each panel the black line and points represent the evolution of branches from an explant experiment in (Watanabe & Costantini, 2004); the orange line represents the mean branching observed by model simulation (n= 200) and the shaded region indicates the 95% confidence interval. “ACD” indicates “anisotropic cell division”. The parameter values used in each case are the same as for Fig. 4 in the main text.

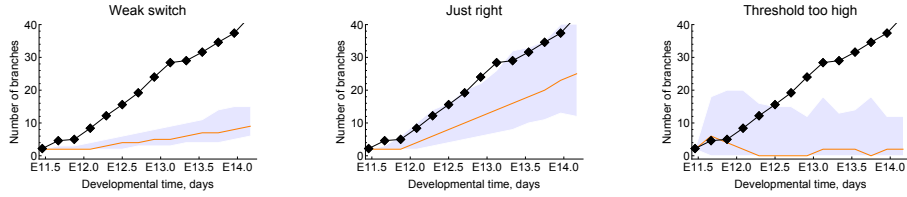


Figure S3: The effect of a GDNF-mediated proliferation growth switch on simulated explant branching for the same explant as in figure S1. In each panel the black line and points represent the evolution of branches from an explant experiment in (Watanabe & Costantini, 2004); the orange line represents the mean branching observed by model simulation ($n=200$) and the shaded region indicates the 95% confidence interval. The parameter values used in each case are the same as for Fig. 6 in the main text.

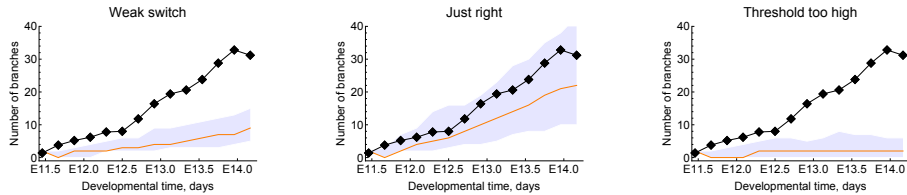


Figure S4: The effect of a GDNF-mediated proliferation growth switch on simulated explant branching for the same explant as in figure S2. In each panel the black line and points represent the evolution of branches from an explant experiment in (Watanabe & Costantini, 2004); the orange line represents the mean branching observed by model simulation ($n=200$) and the shaded region indicates the 95% confidence interval. The parameter values used in each case are the same as for Fig. 6 in the main text.