

Clustering Analysis

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1. Load packages and Data

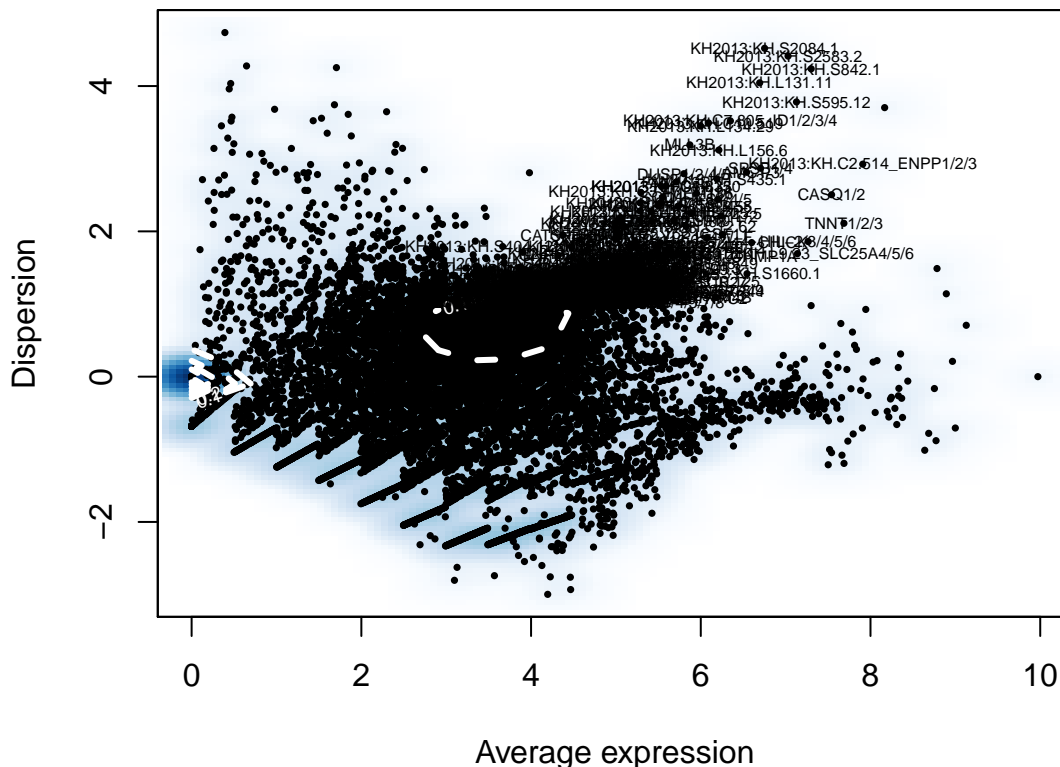
```
library(Seurat)
library(RColorBrewer)
source("functions.R")
load("hpfall.remv1.Robj")
load("hpfall.remv2.Robj")
col = colorRampPalette(rev(brewer.pal(n = 10, name = "RdBu")))
```

2. hpf12

```
# Subset data from preprocessed Seurat object
hpf12 = subsetData(hpfall.remv2, which.cells(hpfall.remv2, "hpf12"), do.scale = F)
hpf12
```

```
## An object of class seurat in project allhpf
## 14864 genes across 27 samples.
```

```
# Find variable gene with  $4 < \text{Average expression and Dispersion} > 2$ 
hpf12 = mean.var.plot(hpf12)
```



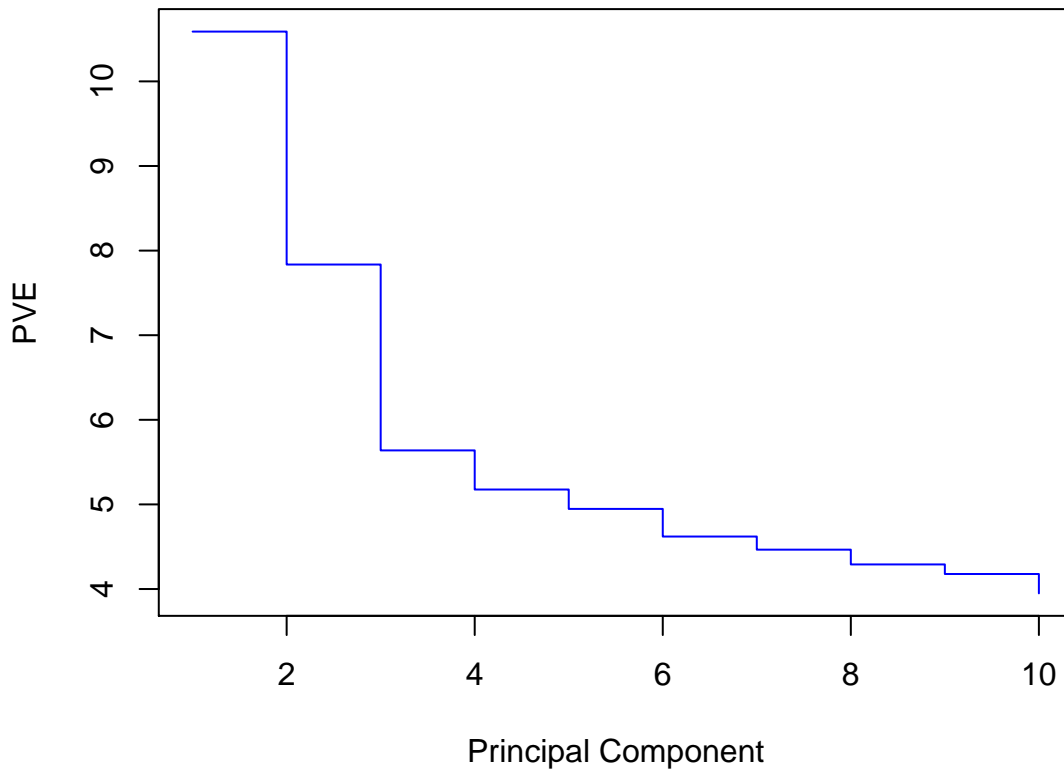
```
length(hpf12@var.genes)
```

```
## [1] 428
```

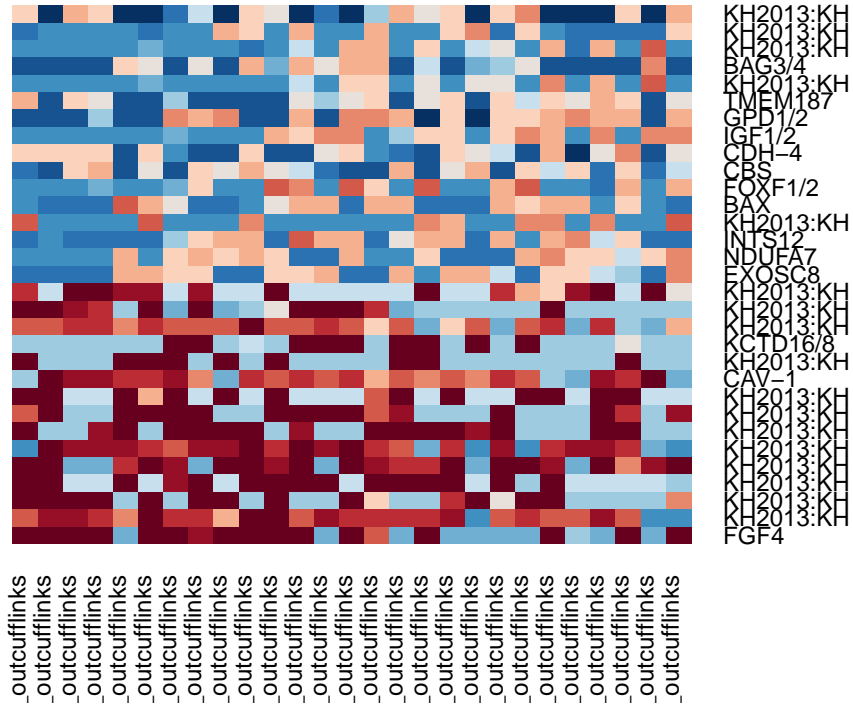
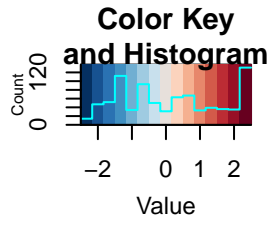
```
# Run a PCA using variable gene list
```

```
hpf12 = pca(hpf12, do.print = F)
```

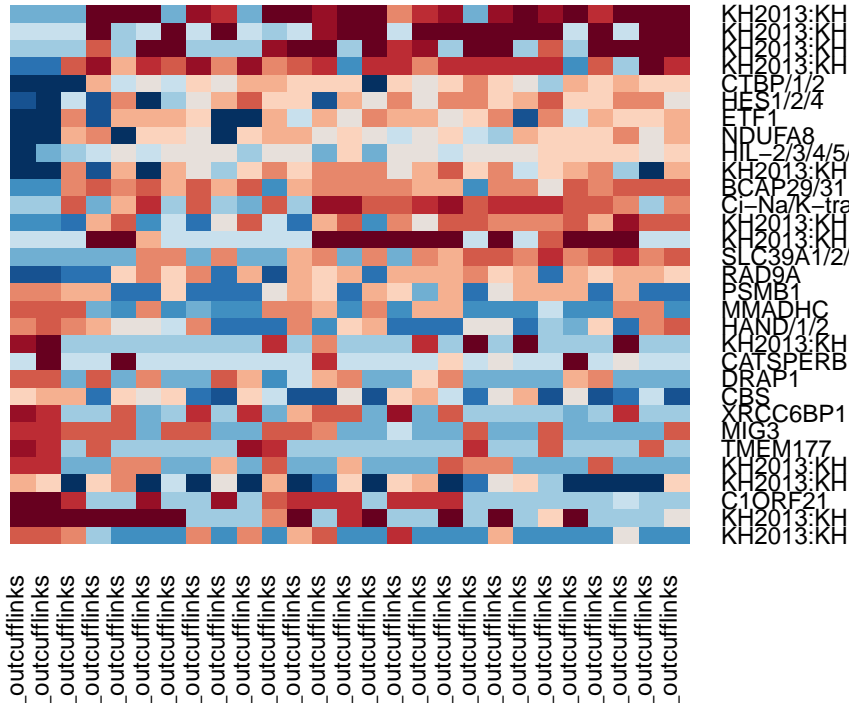
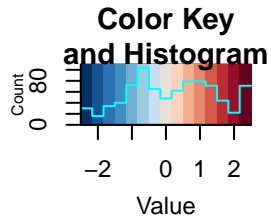
```
pcScree(hpf12, hpf12@var.genes, 10)
```



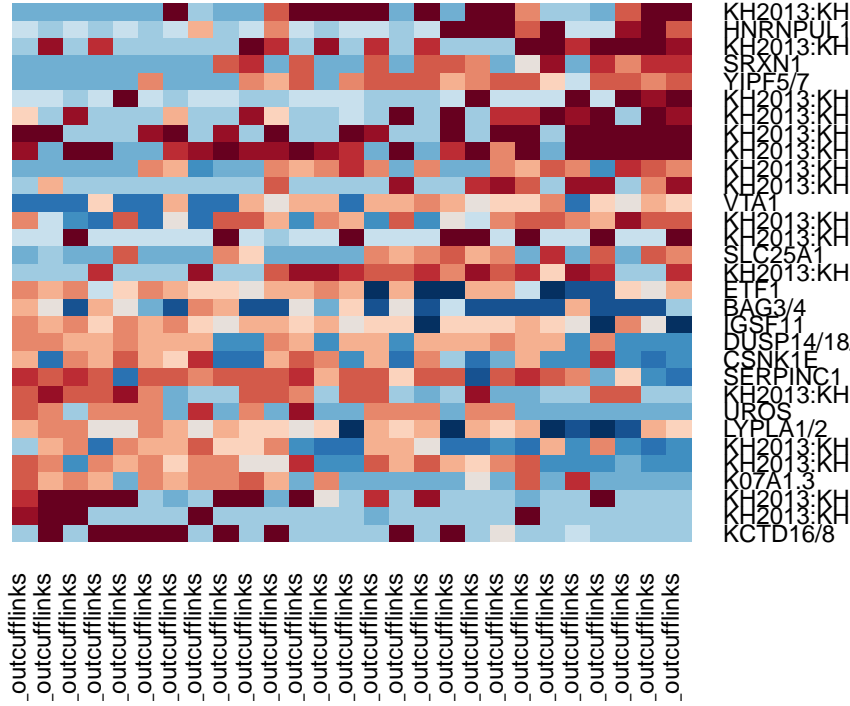
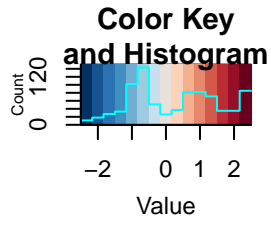
```
pcHeatmap(hpf12, 1, do.balanced = T, col.use = col)
```



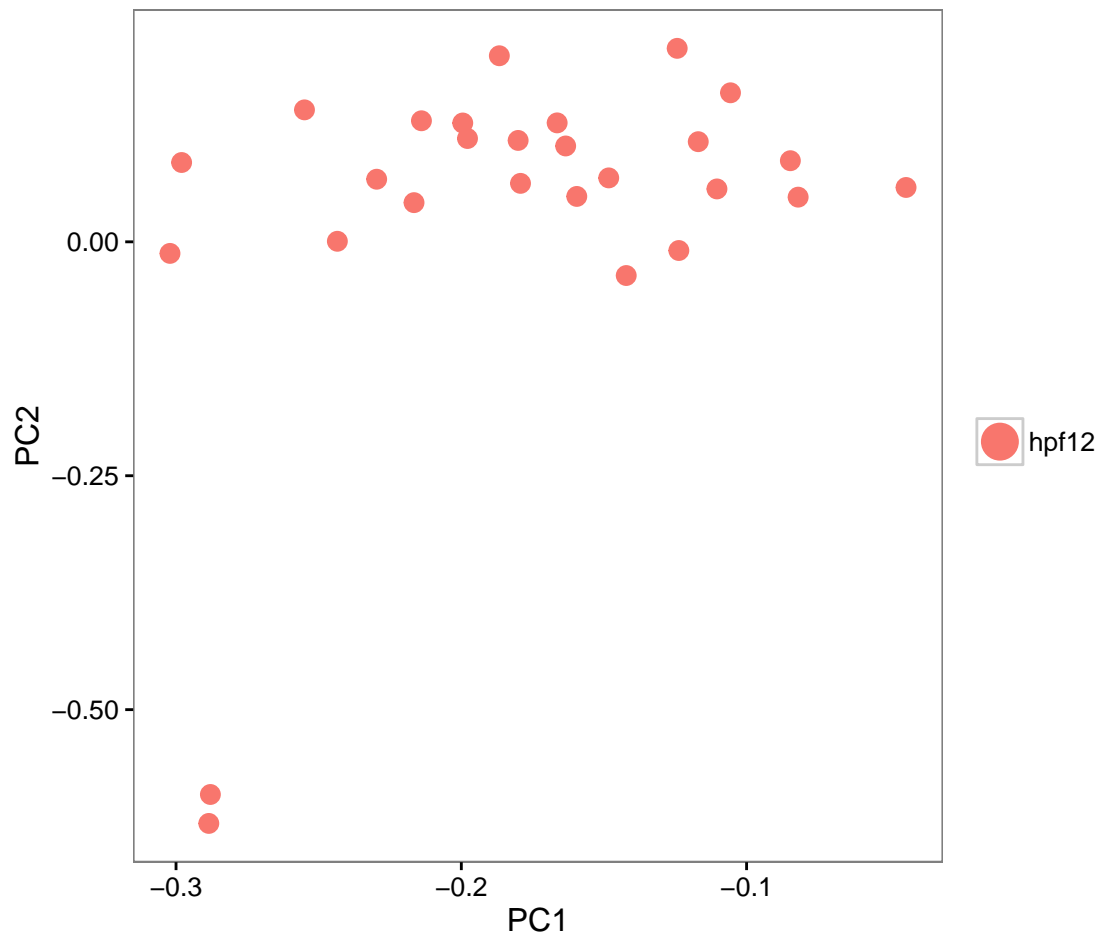
```
pcHeatmap(hpf12, 2, do.balanced = T, col.use = col)
```



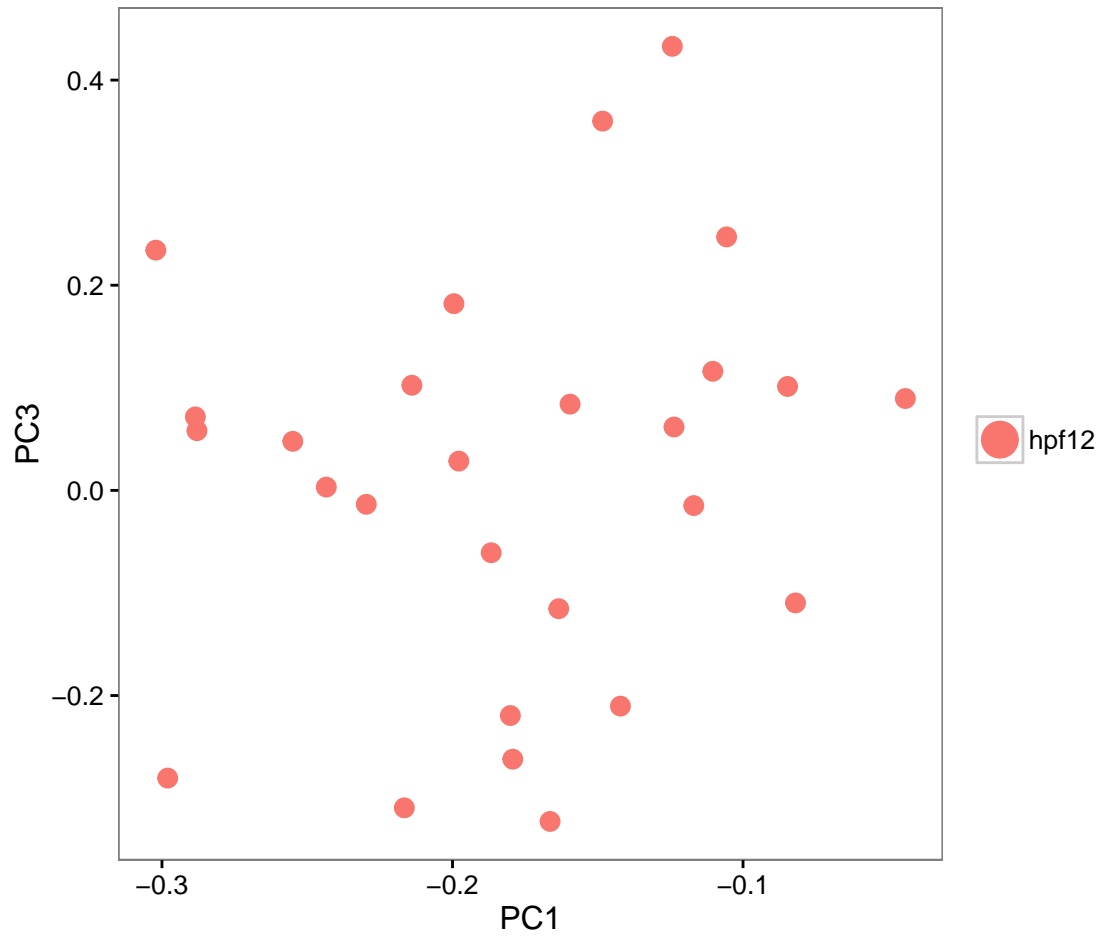
```
pcHeatmap(hpf12, 3, do.balanced = T, col.use = col)
```



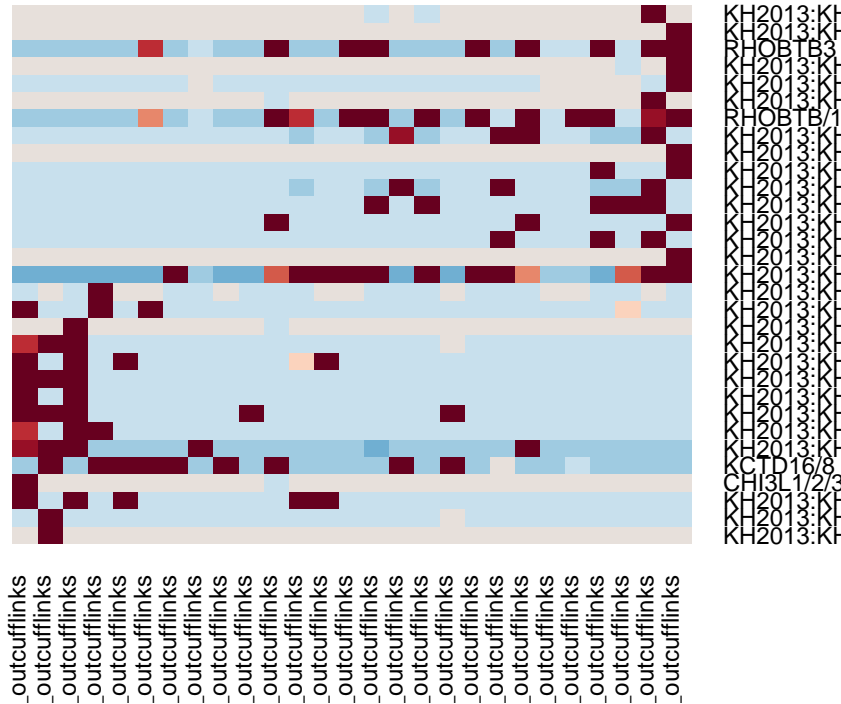
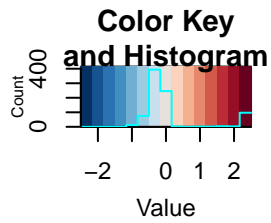
```
pca.plot(hpf12, 1, 2)
```



```
pca.plot(hpf12, 1, 3)
```



```
# Calculate PCA scores for all genes (PCA projection)  
hpf12 = project.pca(hpf12, do.print = F)  
  
# Visualize the full projected PCA, which now includes new genes which were  
# not previously (use.full=TRUE)  
pcHeatmap(hpf12, 1, use.full = T, do.balanced = T, col.use = col)
```

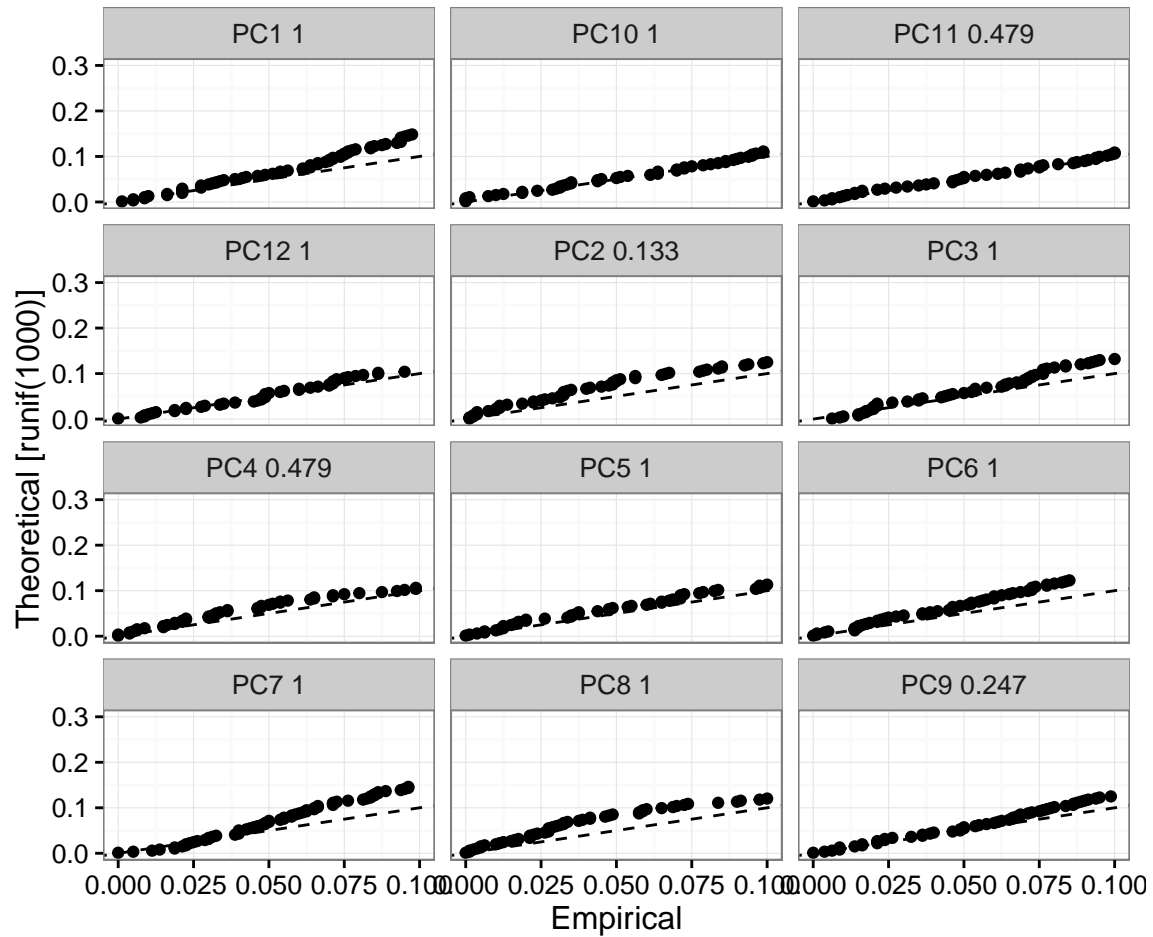



```

# Do 200 random samplings to find significant genes, each time randomly
# permute 1% of genes This returns a 'p-value' for each gene in each PC,
# based on how likely the gene/PC score woud have been observed by chance
# Note that in this case we get the same result with 200 or 1000 samplings,
# so we do 200 here for expediency (20 PCs is used for significant test)
hpf12 = jackStraw(hpf12, num.replicate = 200, do.print = F, num.pc = 20)

# The jackStraw plot compares the distribution of P-values for each PC with
# a uniform distribution (dashed line) 'Significant' PCs will have a strong
# enrichment of genes with low p-values (solid curve above dashed line)
jackStrawPlot.new(hpf12, PCs = 1:12)

```

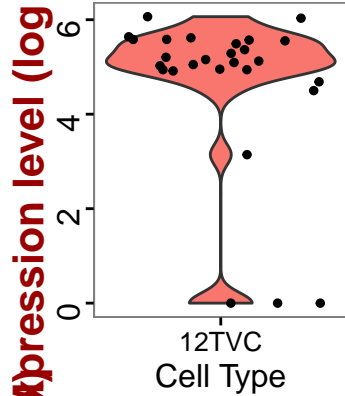


*# jackStraw plots show that none of PCs are significant enough, which
suggests homogenous population.*

With preliminary studies these cells are 12hpf TVC cells
`hpf12 = set.ident(hpf12, ident.use = "12TVC")`

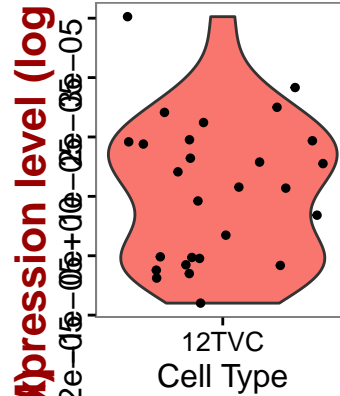
Visualize known TVC cell markers
`vlnPlot(hpf12, c("GATA4/5/6", "HAND/2", "HAND1/2", "NKX2-3"))`

GATA4/5/6



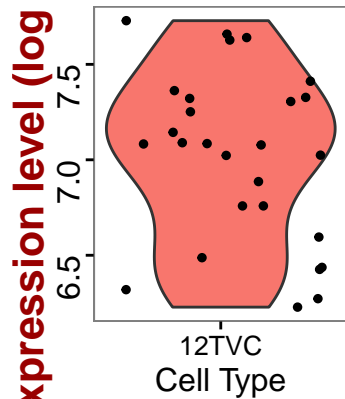
12TVC

HAND1/2



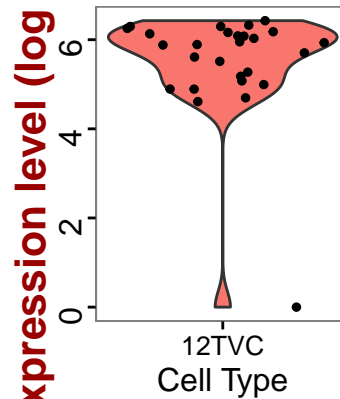
12TVC

HAND/2



12TVC

NKX2-3



12TVC

```
# Write cell names into text files
write.table(hpf12@cell.names, file = "12TVCCells.txt", sep = "\t")

# Find 12TVC marker with 12 contamination cells
mesen.cell = as.character(unlist(read.table("mesen.cellname.txt")))
contam.cell = as.character(unlist(read.table("contam.cellname.txt")))
contam.name = grep("hpf12", c(mesen.cell, contam.cell), value = T)
hpf12.new = subsetData(hpf12.remv1, which.cells(hpf12.remv1, "hpf12"), do.scale = F)
hpf12.new = set.ident(hpf12.new, hpf12@cell.names, "12TVC")
hpf12.new = set.ident(hpf12.new, contam.name, "12Contam")
tvc.marker = find.markers(hpf12.new, "12TVC", "12Contam", thresh.use = 1, test.use = "roc",
  min.pct = 0.5)
head(tvc.marker[order(tvc.marker$myAUC, decreasing = T), ], 20)
```

```
##           myAUC avg_diff power pct.1 pct.2
## TCEAL3/5/6 0.950 1.307940 0.900 1.000 0.959
## SMURF1/2   0.946 2.091587 0.892 1.000 0.306
## DCBLD2     0.946 1.868379 0.892 1.000 0.633
```

```

## KH2013:KH.C8.743          0.944 3.169373 0.888 0.926 0.143
## NAF1                      0.941 1.630562 0.882 1.000 0.510
## FARP1/2                   0.940 1.512813 0.880 1.000 0.633
## KH2013:KH.C2.118_ENPP1/2/3 0.933 1.845469 0.866 1.000 0.306
## NOLC1                     0.933 1.433875 0.866 1.000 0.755
## KH2013:KH.C2.514_ENPP1/2/3 0.925 1.807612 0.850 1.000 0.592
## KH2013:KH.C7.205_ASB2     0.923 1.886513 0.846 1.000 0.327
## KH2013:KH.L153.32_PPP1R9A 0.923 1.222093 0.846 1.000 0.694
## NOP5/58                   0.923 1.676462 0.846 0.963 0.633
## UCHL1/3/4                 0.922 1.401090 0.844 1.000 0.571
## KH2013:KH.S2435.1_ASB2    0.921 1.701182 0.842 1.000 0.245
## GPX7/8                    0.917 1.600426 0.834 1.000 0.490
## DDX56                     0.915 1.689561 0.830 1.000 0.347
## KH2013:KH.C3.665_ZAN      0.915 1.392634 0.830 0.963 0.245
## SMG5                       0.914 1.680871 0.828 0.963 0.469
## KH2013:KH.L152.12_GAS2    0.912 1.503341 0.824 1.000 0.224
## KH2013:KH.L170.61_RRP9    0.911 1.619069 0.822 1.000 0.571

```

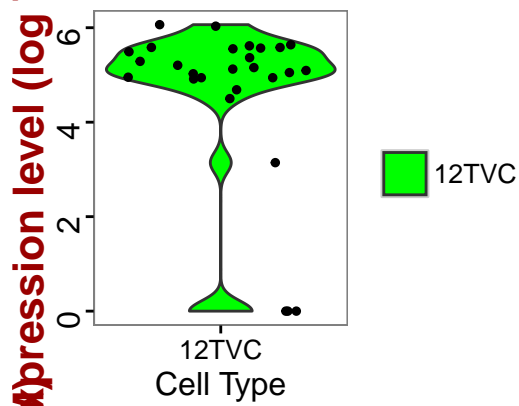
```

write.table(tvc.marker, "TVC.markers.txt", row.names = T, sep = "\t")

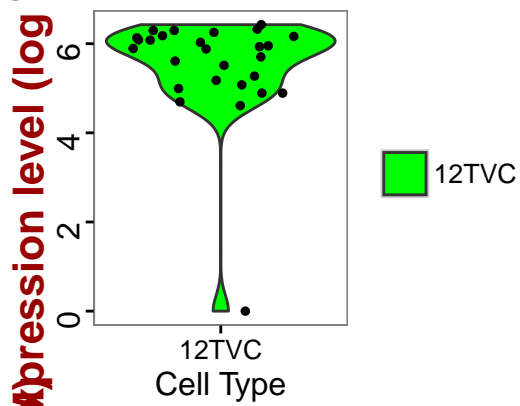
# Draw a heatmap of all cells for these marker genes
vlnPlot(hpf12, c("GATA4/5/6", "HAND/1/2", "NKX2-3", "FZD4"), cols.use = "green")

```

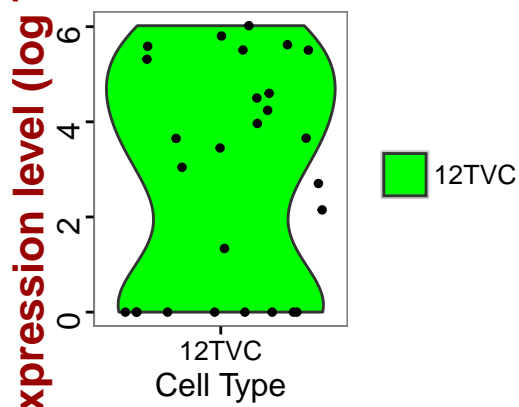
GATA4/5/6



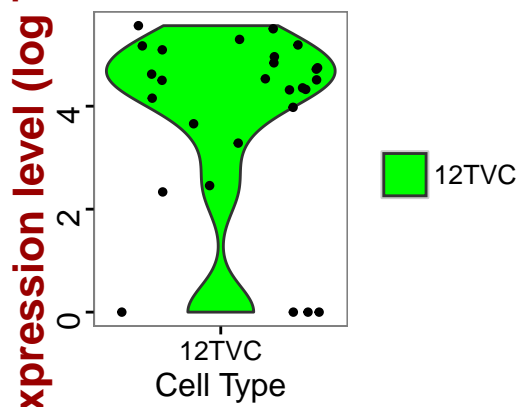
NKX2-3



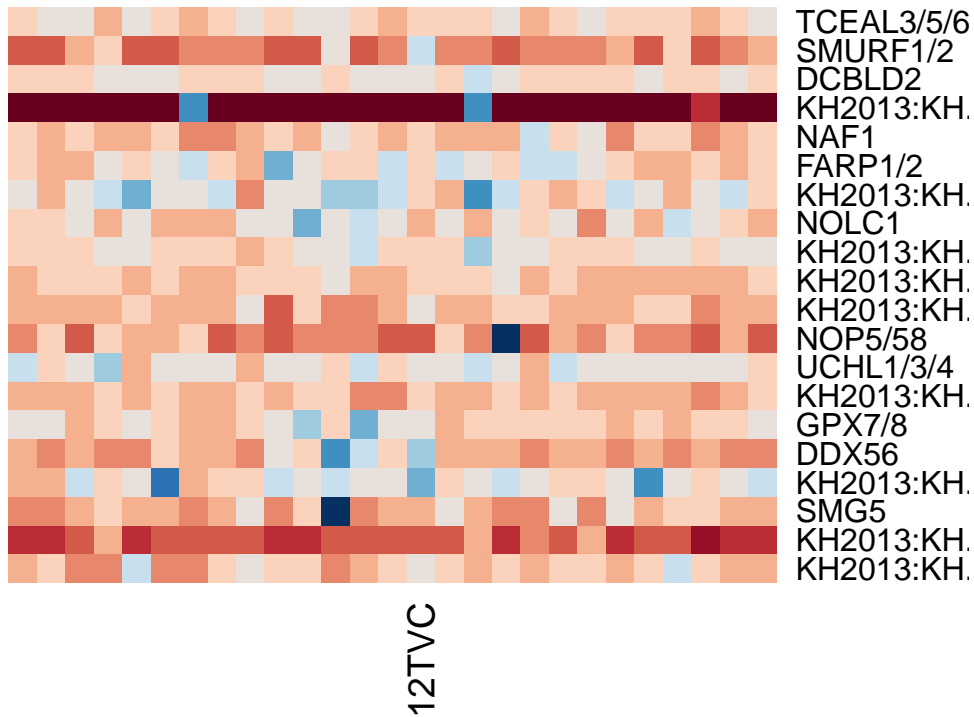
HAND/1/2



FZD4



```
doHeatMap(hpf12, genes.use = rownames(head(tvc.marker[order(tvc.marker$myAUC,
decreasing = T)], 20)), remove.key = TRUE, slim.col.label = T, cex.col = 1.2,
col.use = col, draw.line = F)
```

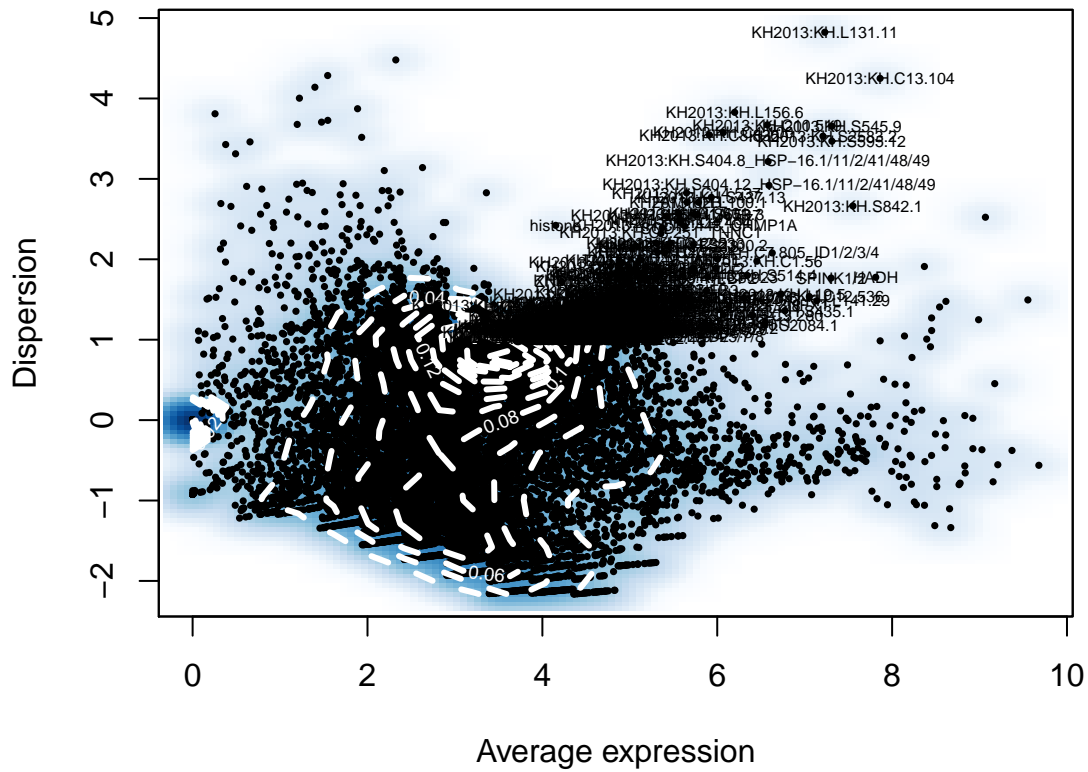


3. hpf14

```
# Subset data from preprocessed Seurat object
hpf14 = subsetData(hpfall.remv2, which.cells(hpfall.remv2, "hpf14"), do.scale = F)
hpf14

## An object of class seurat in project allhpf
## 14864 genes across 275 samples.

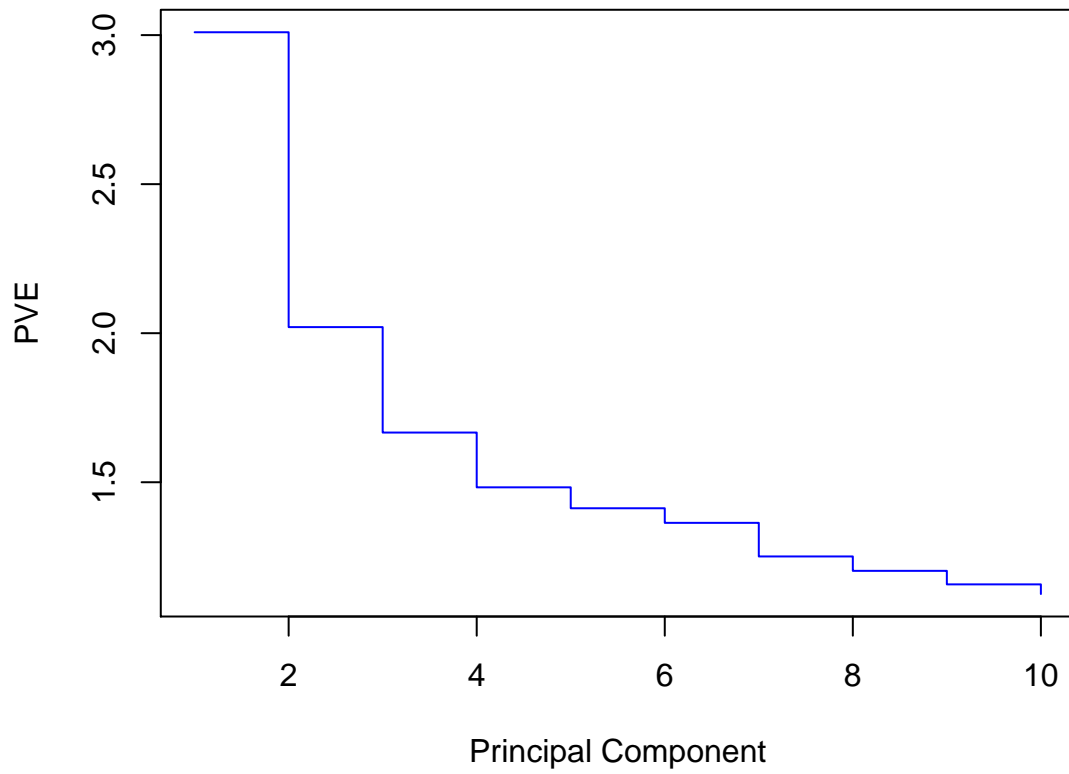
# Find variable gene with 4 < Average expression and Dispersion > 2
hpf14 = mean.var.plot(hpf14)
```



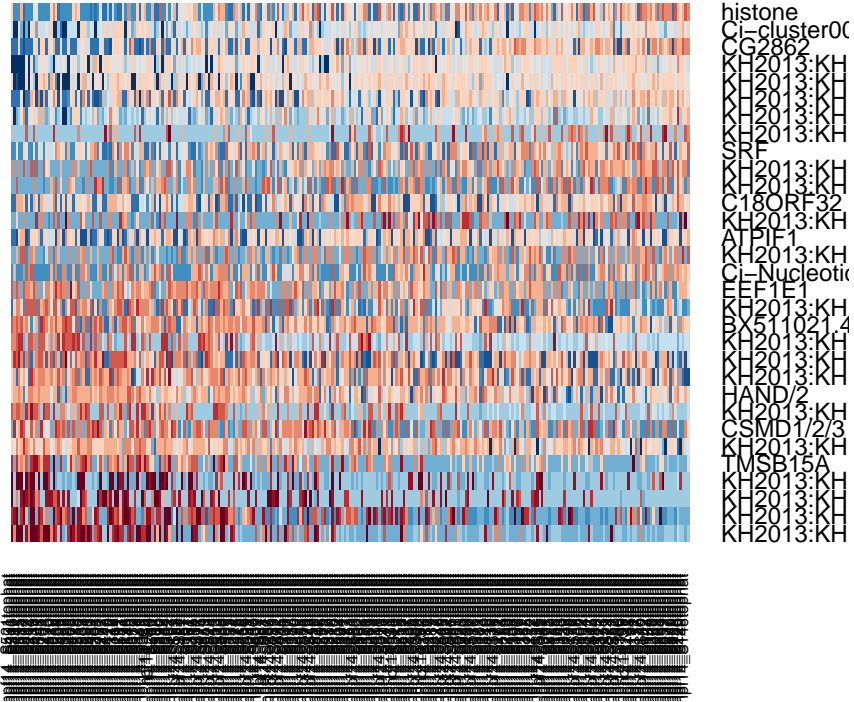
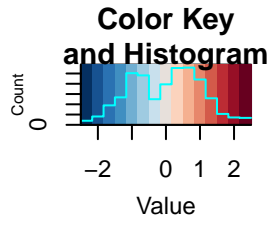
```
length(hpf14@var.genes)
```

```
## [1] 384
```

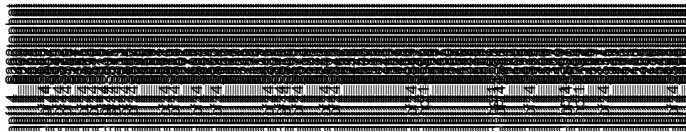
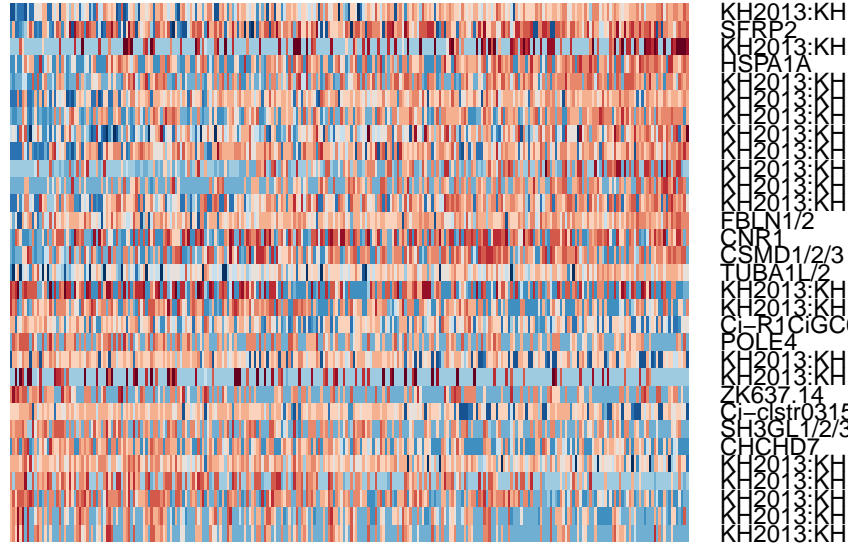
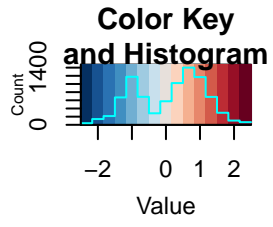
```
# Run a PCA using variable gene list
hpf14 = pca(hpf14, do.print = F)
pcScree(hpf14, hpf14@var.genes, 10)
```

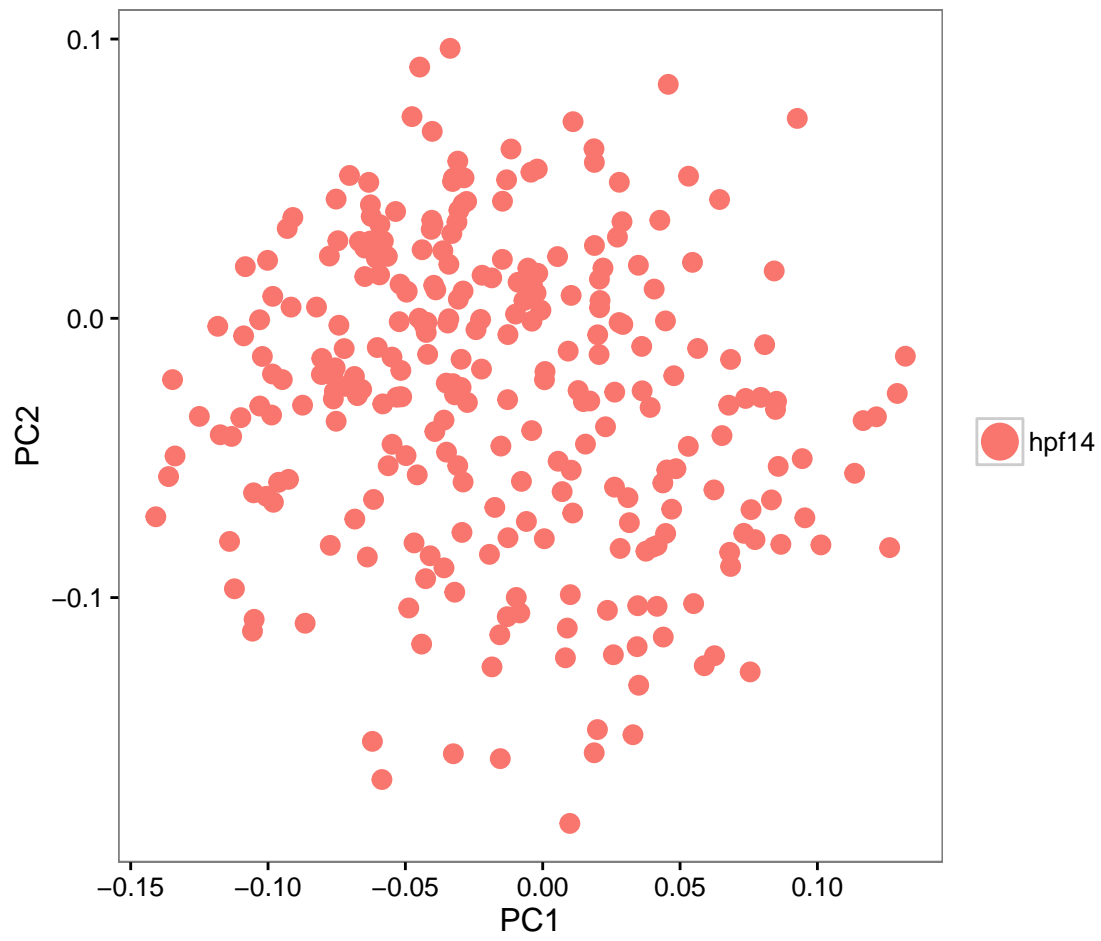
```
pcHeatmap(hpf14, 1, do.balanced = T, col.use = col)
```

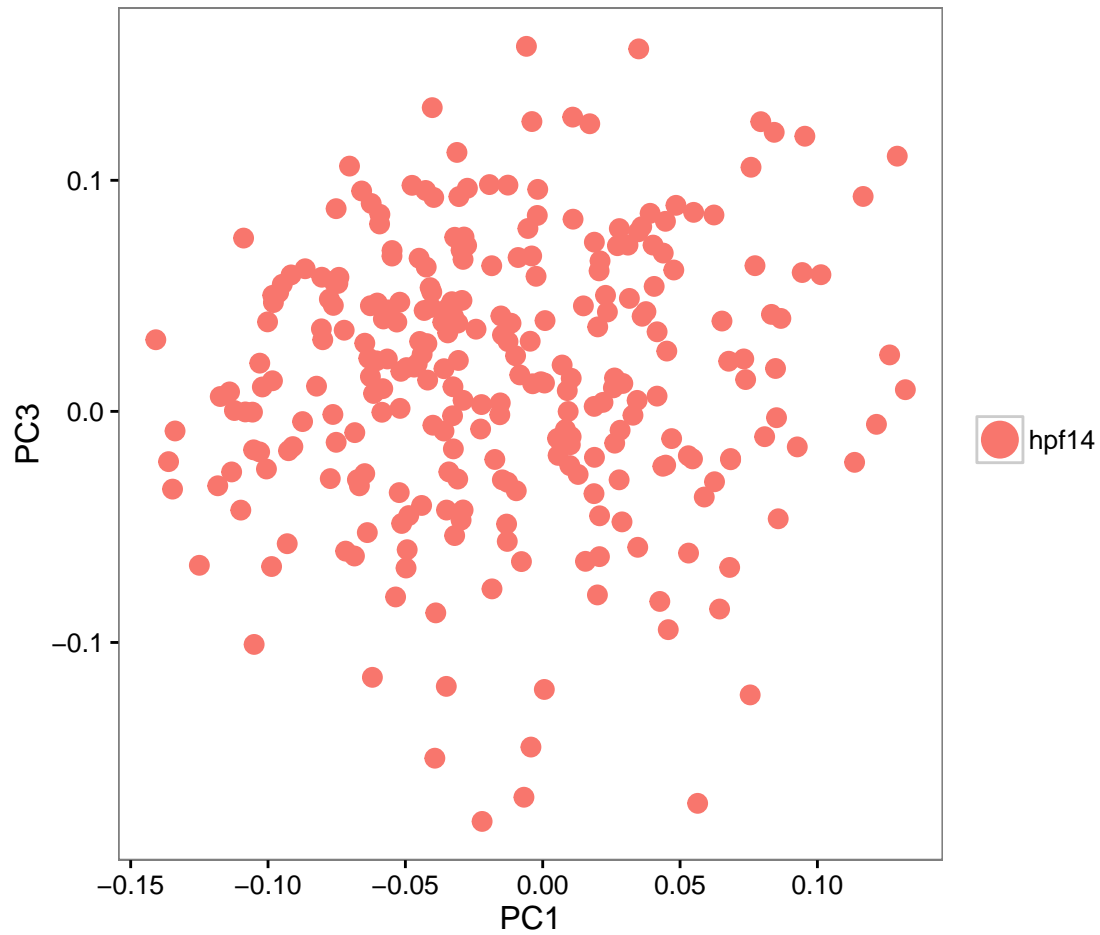
```
pcHeatmap(hpf14, 3, do.balanced = T, col.use = col)
```



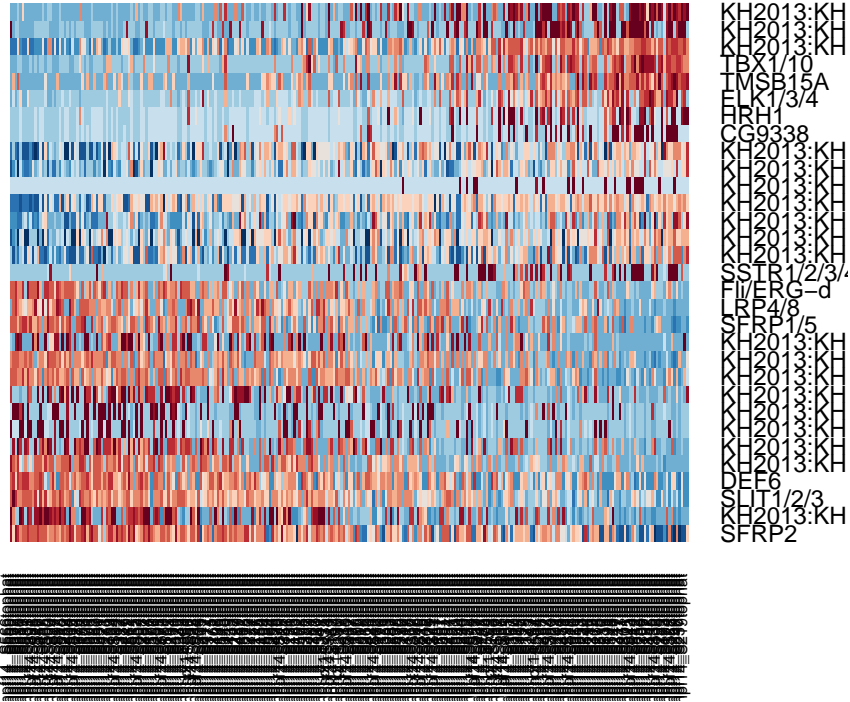
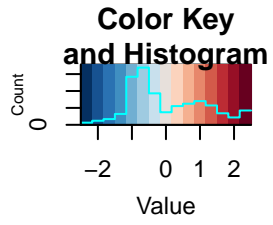
```
pca.plot(hpf14, 1, 2)
```



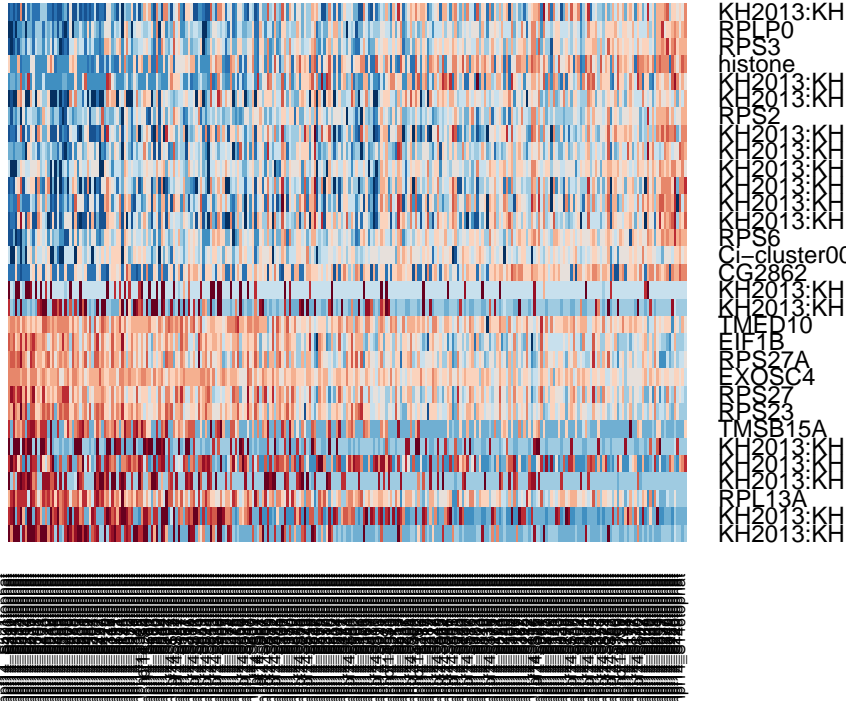
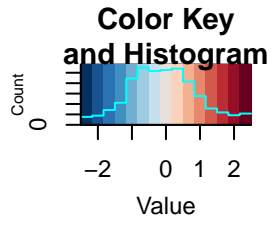
```
pca.plot(hpf14, 1, 3)
```



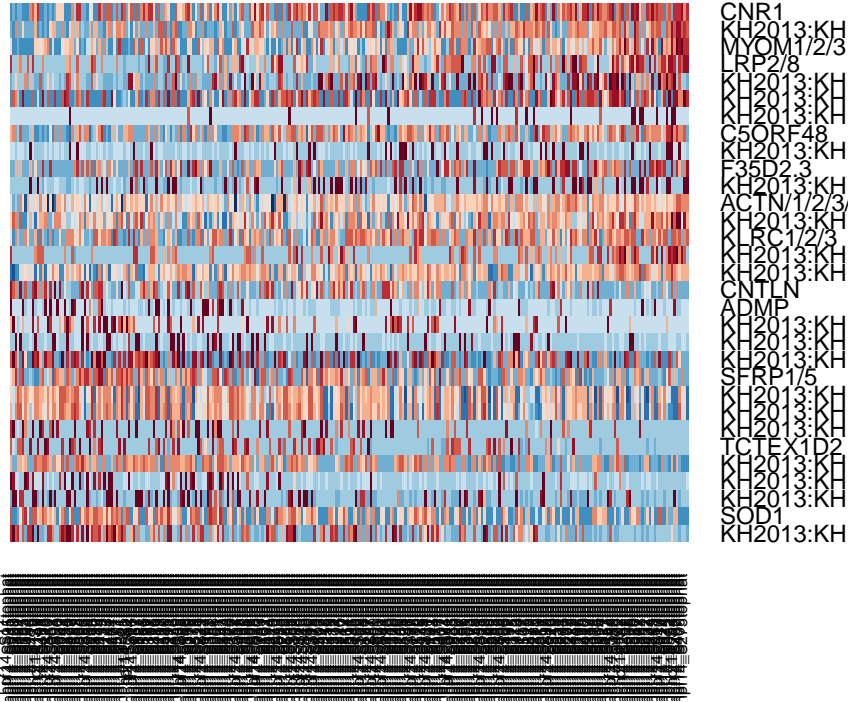
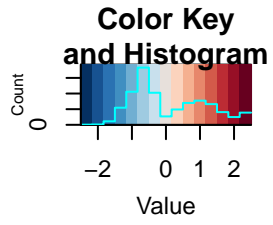
```
# Calculate PCA scores for all genes (PCA projection)  
hpf14 = project.pca(hpf14, do.print = F)  
  
# Visualize the full projected PCA, which now includes new genes which were  
# not previously (use.full=TRUE)  
pcHeatmap(hpf14, 1, use.full = T, do.balanced = T, col.use = col)
```



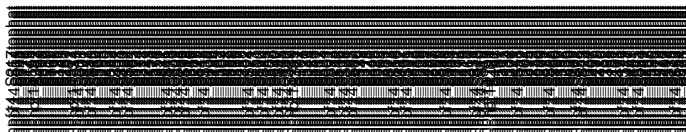
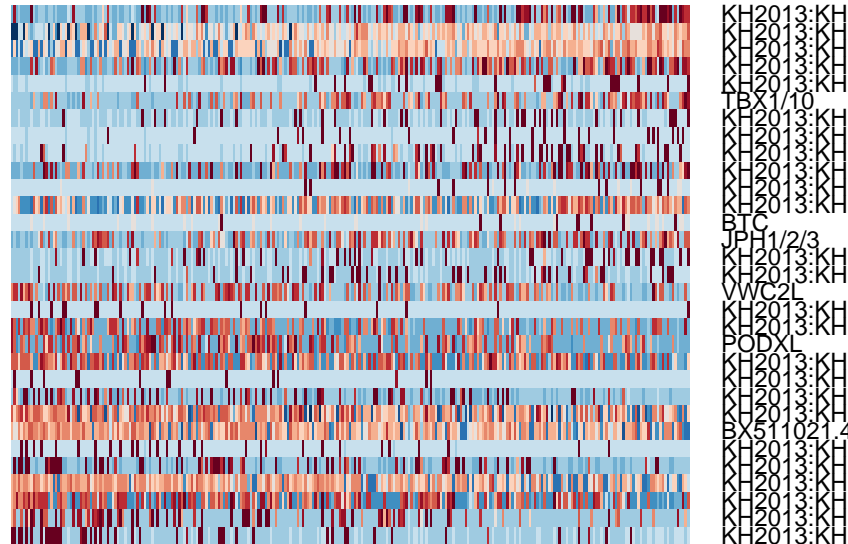
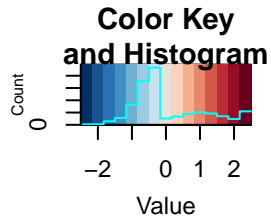
```
pcHeatmap(hpf14, 2, use.full = T, do.balanced = T, col.use = col) #technical
```



```
pcHeatmap(hpf14, 3, use.full = T, do.balanced = T, col.use = col) #technical
```

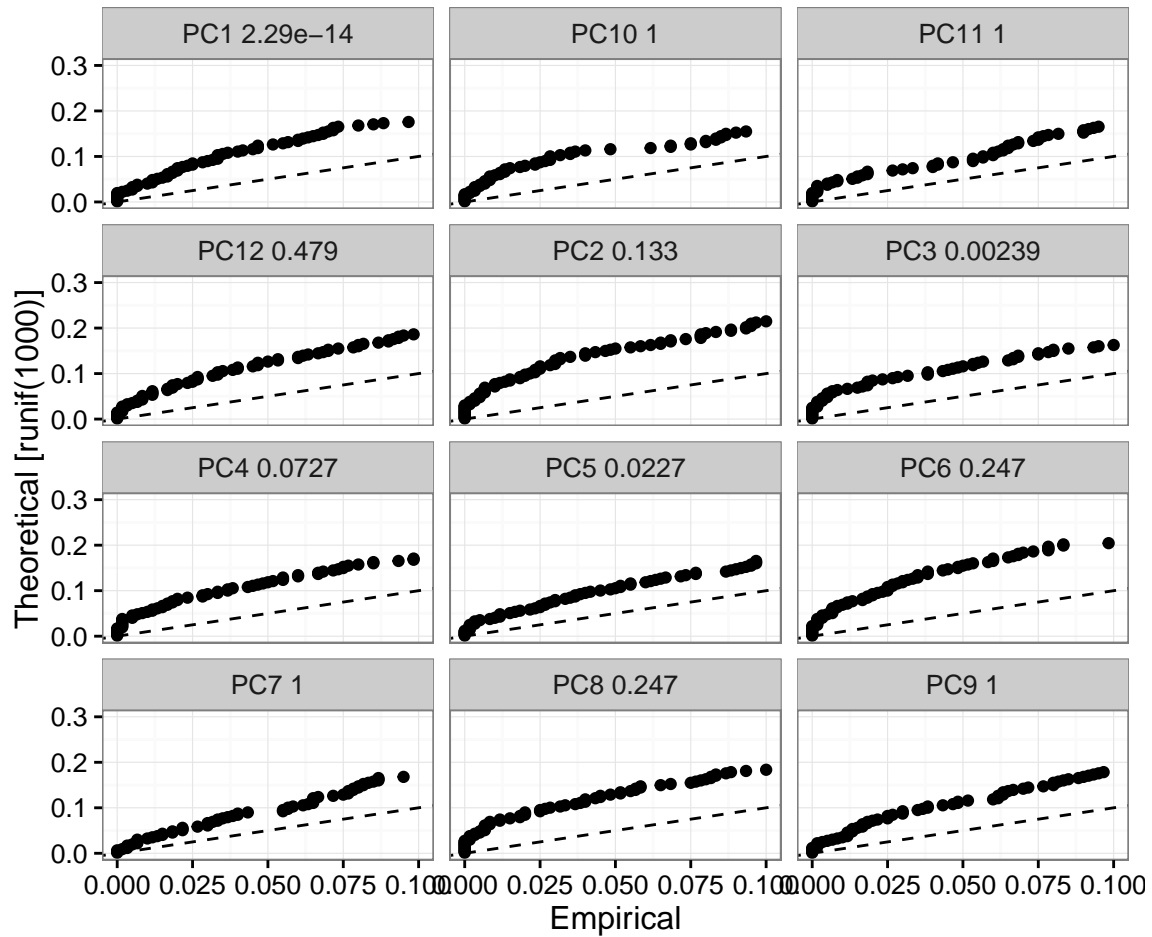



```
pcHeatmap(hpf14, 5, use.full = T, do.balanced = T, col.use = col)
```



```
# Do 200 random samplings to find significant genes, each time randomly
# permute 1% of genes This returns a 'p-value' for each gene in each PC,
# based on how likely the gene/PC score would have been observed by chance
hpf14 = jackStraw(hpf14, num.replicate = 200, do.print = F)
```

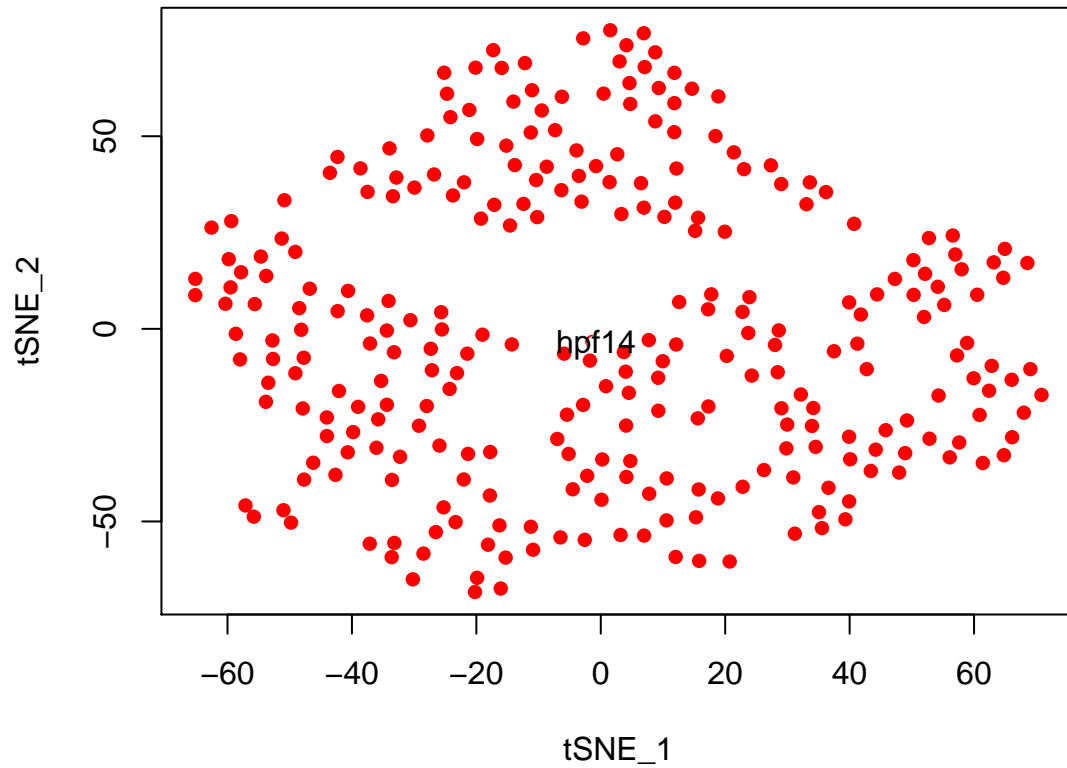
```
# The jackStraw plot compares the distribution of P-values for each PC with
# a uniform distribution (dashed line) 'Significant' PCs will have a strong
# enrichment of genes with low p-values (solid curve above dashed line)
jackStrawPlot.new(hpf14, PCs = 1:12)
```



*# In this case only PC1 is strongly significant and PC5 is significant, PC3
though significant contains technical genes*

*# Run tSNE using significant PCs as input (spectral tSNE), we get distinct
point clouds*

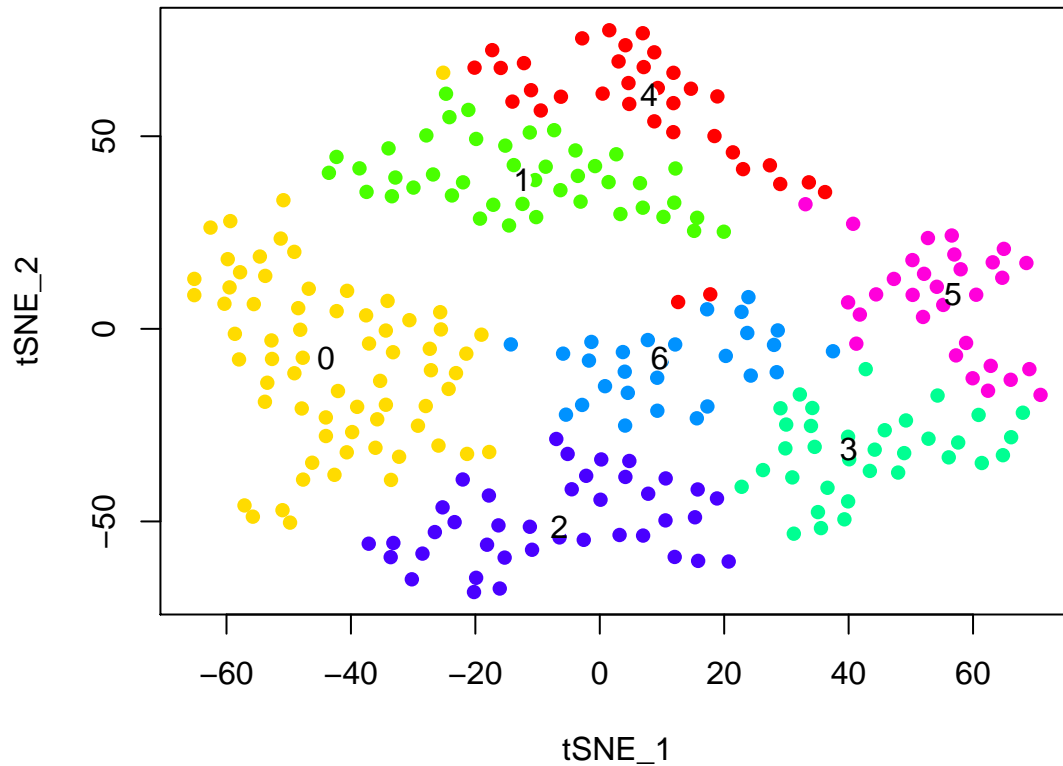
```
hpf14 = run_tsne(hpf14, max_iter = 2000, dims.use = c(1, 5))
tsne.plot(hpf14, do.label = T, label.pt.size = 1)
```



```
# Find cell clusters using Modularity optimization cluster detection.  
hpf14 = FindClusters(hpf14, pc.use = c(1, 5), do.modularity = T, resolution = 1,  
  prune.SNN = 0.1, print.output = 0, k.param = 20)
```

```
## [1] "SNN : processed 69 cells"  
## [1] "SNN : processed 138 cells"  
## [1] "SNN : processed 206 cells"  
## [1] "SNN : processed 275 cells"
```

```
tsne.plot(hpf14, do.label = T, label.pt.size = 1)
```



```
# The validity of the clusters can be validated using a classification
# scheme based on linear SVMs. (In this case cutoff of 0.86 is selected to
# optimize clustering)
```

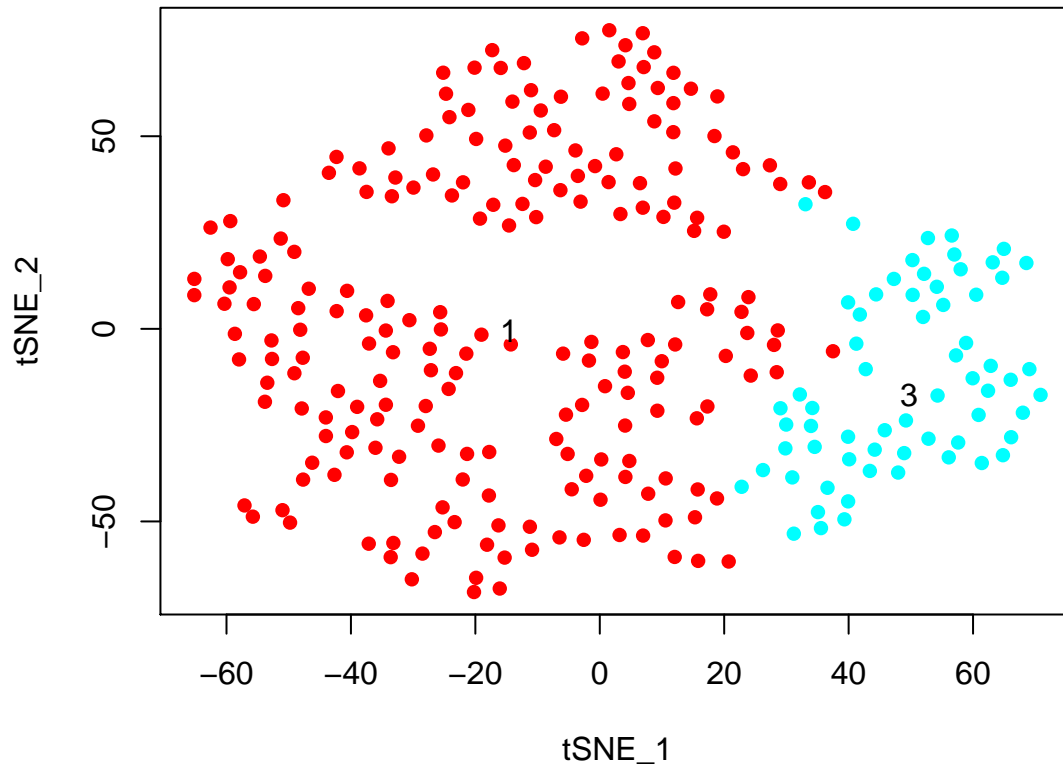
```
hpf14 = BuildSNN(hpf14, pc.use = c(1, 5), do.sparse = F, k.param = 20)
```

```
## [1] "SNN : processed 69 cells"
## [1] "SNN : processed 138 cells"
## [1] "SNN : processed 206 cells"
## [1] "SNN : processed 275 cells"
```

```
hpf14 = ValidateClusters(hpf14, pc.use = c(1, 5), min.connectivity = 0.001,
  acc.cutoff = 0.85)
```

```
## [1] " 0% complete --- merge clusters 6 and 4, classification accuracy of 0.7087"
## [1] " 23% complete --- merge clusters 5 and 3, classification accuracy of 0.7495"
## [1] " 38% complete --- merge clusters 0 and 2, classification accuracy of 0.8438"
## [1] " 54% complete --- merge clusters 4 and 1, classification accuracy of 0.7940"
## [1] " 85% complete --- merge clusters 2 and 1, classification accuracy of 0.7865"
## [1] "100% complete --- started with 7 clusters, 2 clusters remaining"
```

```
tsne.plot(hpf14, do.label = T, label.pt.size = 1)
```



*# Find cluster markers using ROC test with thresh.use = 1, min.pct = 0.5 The
 # ROC test returns the 'classification power' for any individual marker
 # (ranging from 0 - random, to 1 - perfect). Though not a statistical test,
 # it is often very useful for finding clean markers.*

```
cl1_14.markers = find.markers(hpf14, 1, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(cl1_14.markers[order(cl1_14.markers$myAUC, decreasing = T), ], 20)
```

##	myAUC	avg_diff	power	pct.1	pct.2
## SLIT1/2/3	0.797	1.550038	0.594	0.886	0.625
## LRP4/8	0.761	1.619001	0.522	0.692	0.234
## DEF6	0.749	1.325675	0.498	0.829	0.562
## SFRP2	0.739	1.183835	0.478	0.872	0.641
## KH2013:KH.S1159.1_FLI1	0.731	1.277835	0.462	0.725	0.406
## KH2013:KH.C13.152_F14B6.6	0.729	1.495670	0.458	0.654	0.266
## COL13A1	0.727	1.442988	0.454	0.645	0.250
## PDE5A	0.704	1.457533	0.408	0.621	0.312
## VWC2L	0.703	1.425926	0.406	0.559	0.156
## Fli/ERG-d	0.702	1.543703	0.404	0.611	0.281
## LYS2	0.702	1.173341	0.404	0.720	0.469
## KH2013:KH.C11.362	0.698	1.068338	0.396	0.725	0.391
## KH2013:KH.C4.547_BMP2/4	0.694	1.079053	0.388	0.706	0.422
## C5ORF48	0.691	1.172332	0.382	0.682	0.406
## EFNB1/2/3	0.690	1.372033	0.380	0.578	0.250

```

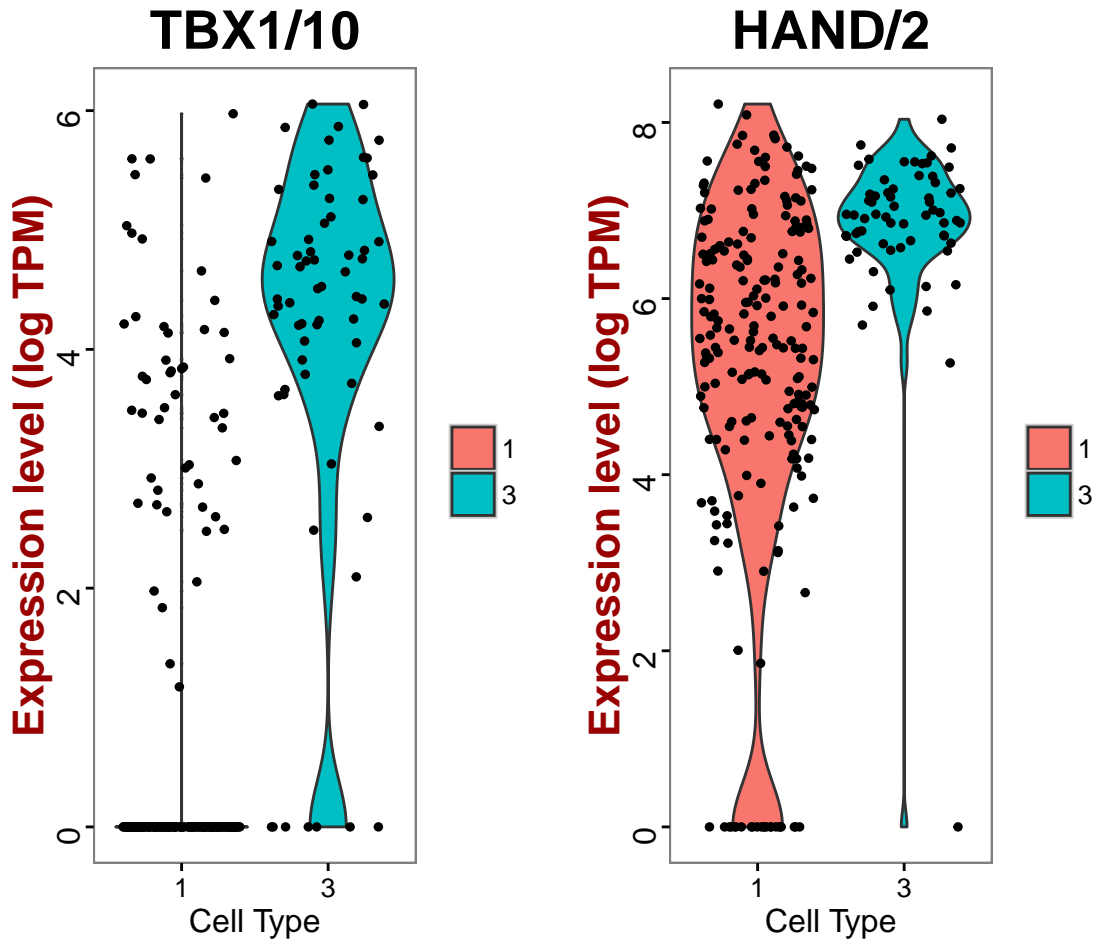
## LRRC42                0.682 1.202552 0.364 0.616 0.328
## KH2013:KH.C2.245_B3GNT2/7/9 0.682 1.169769 0.364 0.611 0.344
## RAP1GAP/2             0.671 1.000794 0.342 0.635 0.391
## KH2013:KH.C7.787_SELP 0.666 1.267515 0.332 0.517 0.219
## KH2013:KH.C3.52_EFNA1/2/3/4/5 0.649 1.747759 0.298 0.540 0.328

```

```

# Visualize known markers with a violin plot
vlnPlot(hpf14, c("TBX1/10", "HAND/2"))

```



```

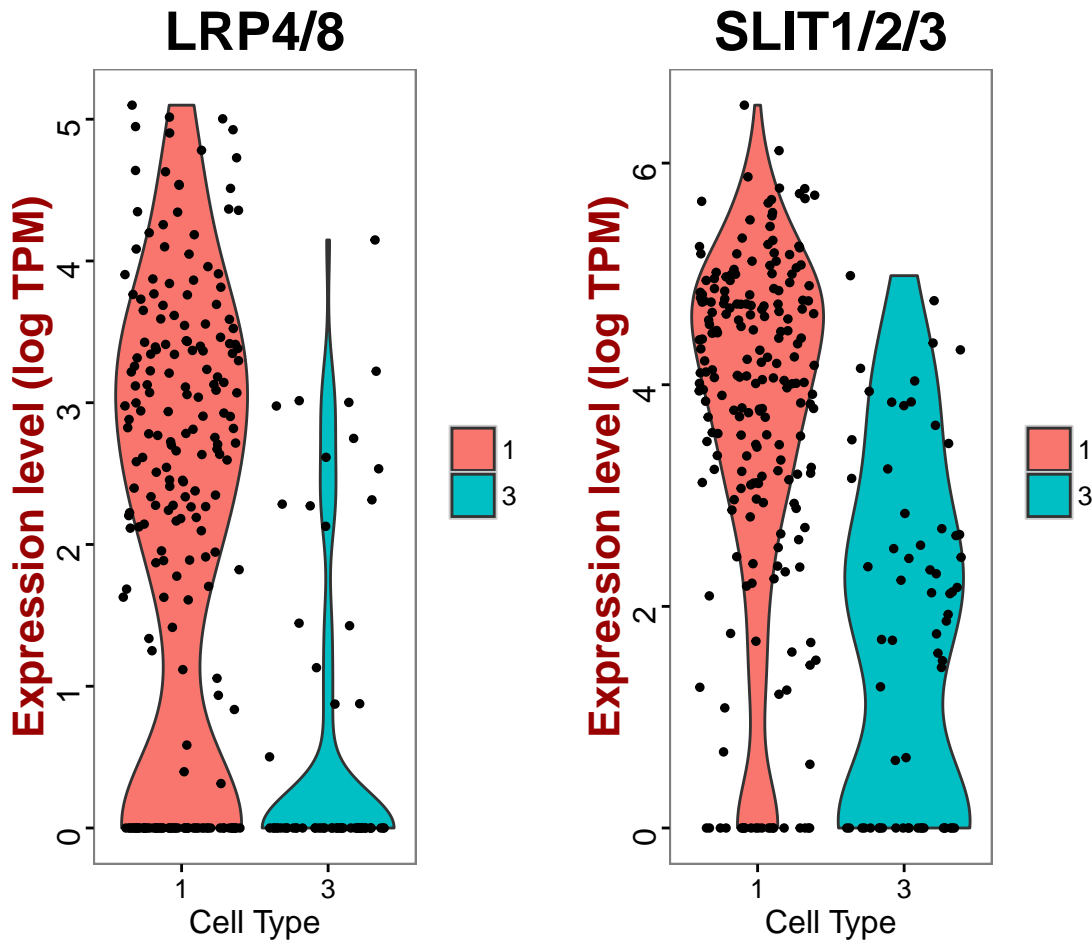
# Based on preliminary studies on TBX1/10- and HAND/2+ expression in TVC
# lineage, cluster 12 is FHP cells

```

```

# Visualize new markers with a violin plot
vlnPlot(hpf14, c("LRP4/8", "SLIT1/2/3"))

```

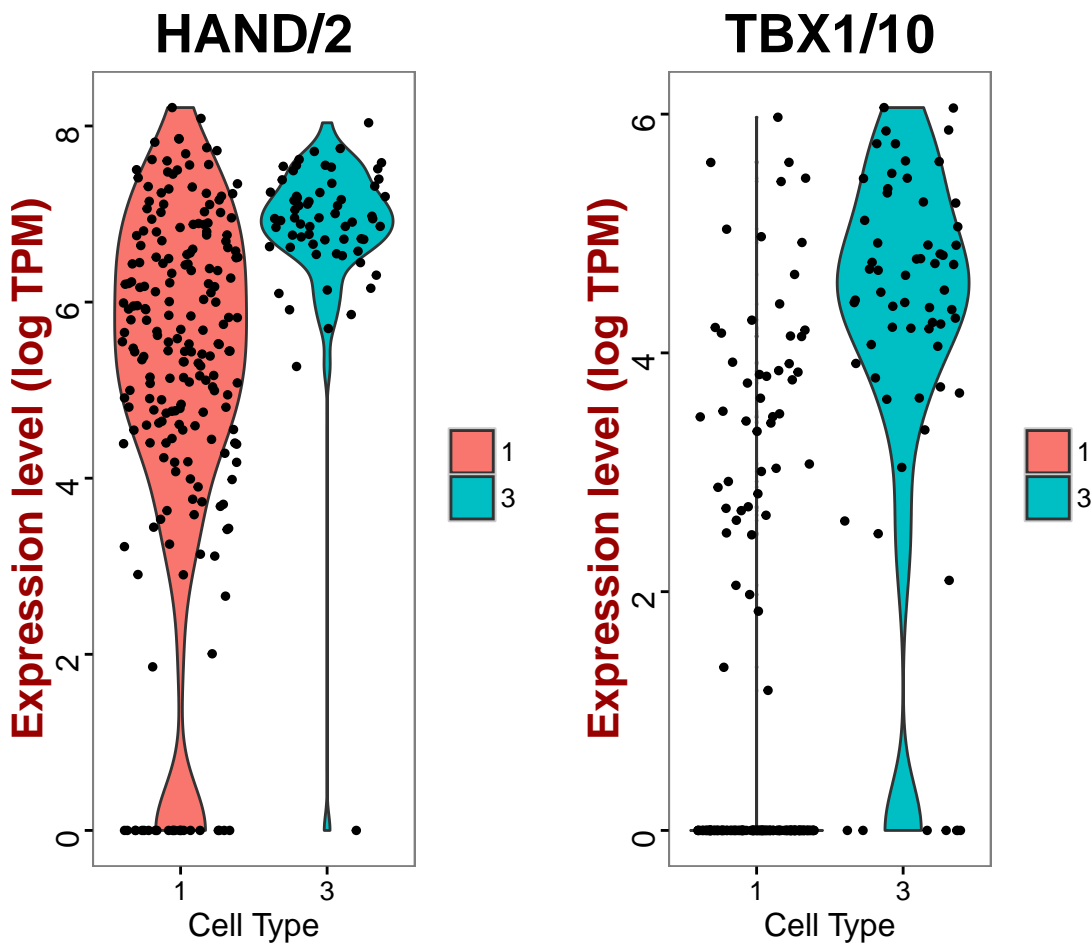
```
# Find markers for cluster 4
```

```
cl3_14.markers = find.markers(hpf14, 3, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(cl3_14.markers[order(cl3_14.markers$myAUC, decreasing = T), ], 20)
```

	myAUC	avg_diff	power	pct.1	pct.2
## KH2013:KH.S555.1_HTR7	0.895	1.787203	0.790	0.984	0.555
## TBX1/10	0.884	1.995667	0.768	0.891	0.237
## KH2013:KH.C4.404	0.866	2.276980	0.732	0.859	0.223
## TMSB15A	0.860	2.237798	0.720	0.891	0.355
## ELK1/3/4	0.819	2.005421	0.638	0.766	0.194
## HRH1	0.771	2.110109	0.542	0.594	0.057
## ZEB1/2	0.769	1.113927	0.538	0.938	0.872
## KH2013:KH.C3.696	0.758	1.277699	0.516	0.781	0.341
## KH2013:KH.C4.125_BMP2/4	0.750	1.278512	0.500	0.781	0.436
## ZK637.14	0.742	1.315203	0.484	0.781	0.351
## KH2013:KH.C10.203	0.735	1.301948	0.470	0.688	0.256
## IRX4/6	0.725	1.142938	0.450	0.734	0.384
## ITPKA	0.722	1.384907	0.444	0.750	0.464
## FOXF1/2	0.714	1.146914	0.428	0.734	0.422

```
## KH2013:KH.S1012.2      0.710 1.454288 0.420 0.578 0.190
## KH2013:KH.L108.33     0.709 1.591887 0.418 0.531 0.118
## W02G9.4                0.687 1.177161 0.374 0.594 0.246
## SSTR1/2/3/4/5         0.686 1.393247 0.372 0.516 0.161
## KH2013:KH.C1.953_CG32702 0.670 1.115838 0.340 0.609 0.322
## KH2013:KH.C9.692_GABRR1/2/3 0.669 1.146391 0.338 0.531 0.209
```

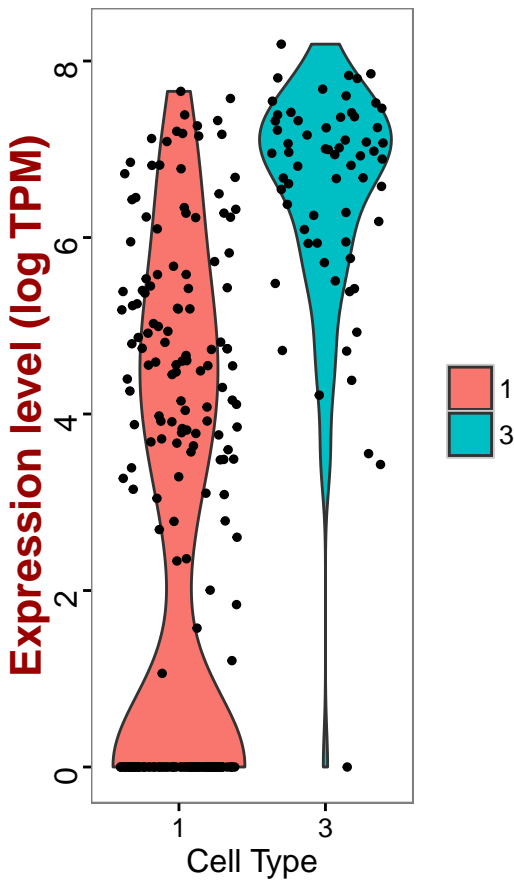
```
# Visualize known markers with a violin plot
vlnPlot(hpf14, c("HAND/2", "TBX1/10"))
```



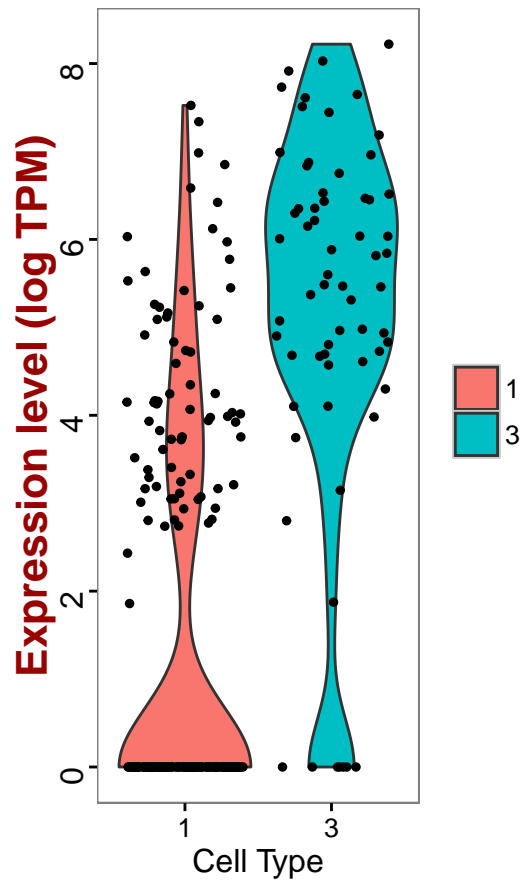
```
# Based on preliminary studies these TBX1/10- HAND/2+ cells are STVCs
```

```
# Visualize new markers with a violin plot
vlnPlot(hpf14, c("KH2013:KH.S555.1_HTR7", "TMSB15A"))
```

h2013:KH.S555.1_HTR7



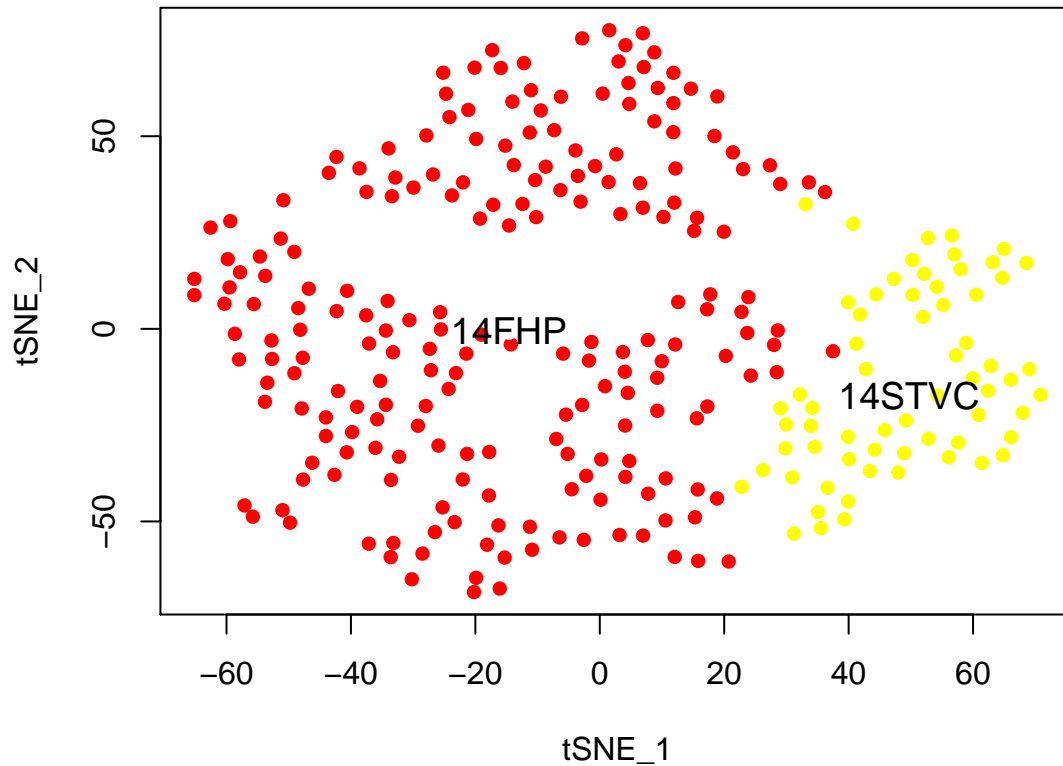
TMSB15A



```
# Write cell names into text files
write.table(which.cells(hpf14, 1), file = "14FHPCells.txt", sep = "\t")
write.table(which.cells(hpf14, 3), file = "14STVCCells.txt", sep = "\t")

# Rename cluster identities
hpf14 = rename.ident(hpf14, 1, "14FHP")
hpf14 = rename.ident(hpf14, 3, "14STVC")

# Visualize tSNE used color scheme FHP-red, STVC-yellow
tsne.plot(hpf14, do.label = T, label.pt.size = 1, label.cex.text = 1.2, label.cols.use =
  "yellow"))
```



```
# Store FHP markers in text file
FHP_14.markers = c11_14.markers
head(FHP_14.markers[order(FHP_14.markers$myAUC, decreasing = T), ], 20)
```

##	myAUC	avg_diff	power	pct.1	pct.2
## SLIT1/2/3	0.797	1.550038	0.594	0.886	0.625
## LRP4/8	0.761	1.619001	0.522	0.692	0.234
## DEF6	0.749	1.325675	0.498	0.829	0.562
## SFRP2	0.739	1.183835	0.478	0.872	0.641
## KH2013:KH.S1159.1_FLI1	0.731	1.277835	0.462	0.725	0.406
## KH2013:KH.C13.152_F14B6.6	0.729	1.495670	0.458	0.654	0.266
## COL13A1	0.727	1.442988	0.454	0.645	0.250
## PDE5A	0.704	1.457533	0.408	0.621	0.312
## VWC2L	0.703	1.425926	0.406	0.559	0.156
## Fli/ERG-d	0.702	1.543703	0.404	0.611	0.281
## LYS2	0.702	1.173341	0.404	0.720	0.469
## KH2013:KH.C11.362	0.698	1.068338	0.396	0.725	0.391
## KH2013:KH.C4.547_BMP2/4	0.694	1.079053	0.388	0.706	0.422
## C50RF48	0.691	1.172332	0.382	0.682	0.406
## EFNB1/2/3	0.690	1.372033	0.380	0.578	0.250
## LRRC42	0.682	1.202552	0.364	0.616	0.328
## KH2013:KH.C2.245_B3GNT2/7/9	0.682	1.169769	0.364	0.611	0.344
## RAP1GAP/2	0.671	1.000794	0.342	0.635	0.391

```
## KH2013:KH.C7.787_SELFP          0.666 1.267515 0.332 0.517 0.219
## KH2013:KH.C3.52_EFNA1/2/3/4/5 0.649 1.747759 0.298 0.540 0.328
```

```
write.table(FHP_14.markers, file = "FHP_14.markers.txt", sep = "\t")

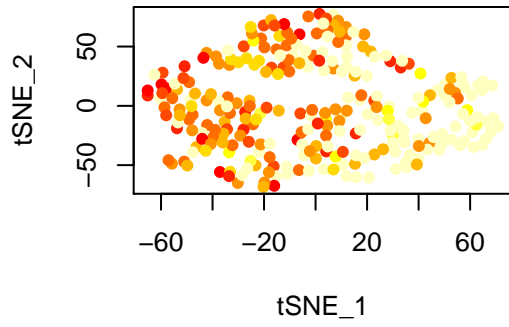
# Store STVC markers in text file
STVC_14.markers = cl3_14.markers
head(STVC_14.markers[order(STVC_14.markers$myAUC, decreasing = T), ], 20)
```

```
##
## KH2013:KH.S555.1_HTR7          0.895 1.787203 0.790 0.984 0.555
## TBX1/10                        0.884 1.995667 0.768 0.891 0.237
## KH2013:KH.C4.404              0.866 2.276980 0.732 0.859 0.223
## TMSB15A                       0.860 2.237798 0.720 0.891 0.355
## ELK1/3/4                      0.819 2.005421 0.638 0.766 0.194
## HRH1                          0.771 2.110109 0.542 0.594 0.057
## ZEB1/2                        0.769 1.113927 0.538 0.938 0.872
## KH2013:KH.C3.696              0.758 1.277699 0.516 0.781 0.341
## KH2013:KH.C4.125_BMP2/4       0.750 1.278512 0.500 0.781 0.436
## ZK637.14                      0.742 1.315203 0.484 0.781 0.351
## KH2013:KH.C10.203            0.735 1.301948 0.470 0.688 0.256
## IRX4/6                        0.725 1.142938 0.450 0.734 0.384
## ITPKA                         0.722 1.384907 0.444 0.750 0.464
## FOXF1/2                      0.714 1.146914 0.428 0.734 0.422
## KH2013:KH.S1012.2            0.710 1.454288 0.420 0.578 0.190
## KH2013:KH.L108.33            0.709 1.591887 0.418 0.531 0.118
## W02G9.4                      0.687 1.177161 0.374 0.594 0.246
## SSTR1/2/3/4/5                0.686 1.393247 0.372 0.516 0.161
## KH2013:KH.C1.953_CG32702     0.670 1.115838 0.340 0.609 0.322
## KH2013:KH.C9.692_GABRR1/2/3 0.669 1.146391 0.338 0.531 0.209
```

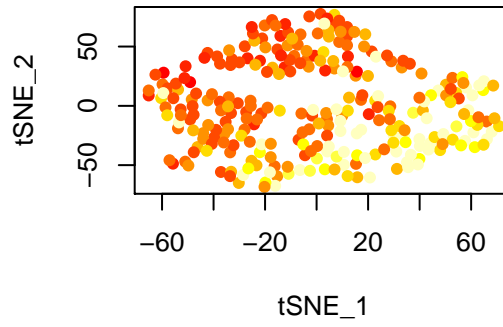
```
write.table(STVC_14.markers, file = "STVC_14.markers.txt", sep = "\t")

# Visualize markers of different clusters using violin plot and feature plot
genes.viz.14 = c("LRP4/8", "SLIT1/2/3", "TBX1/10", "KH2013:KH.S555.1_HTR7")
feature.plot(hpf14, genes.viz.14, pt.size = 1)
```

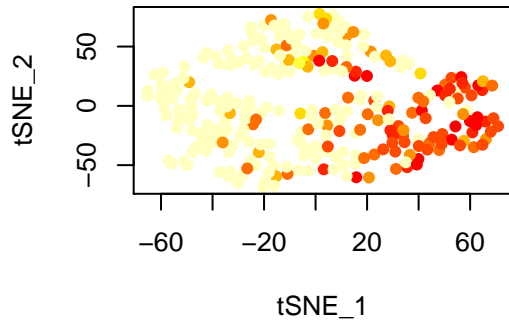
LRP4/8



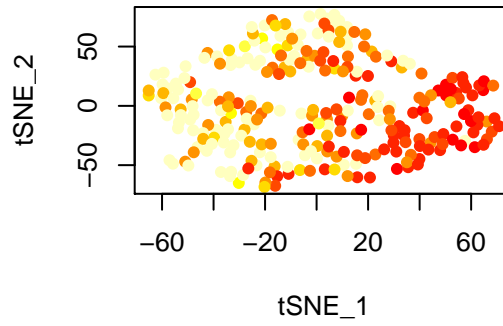
SLIT1/2/3



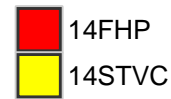
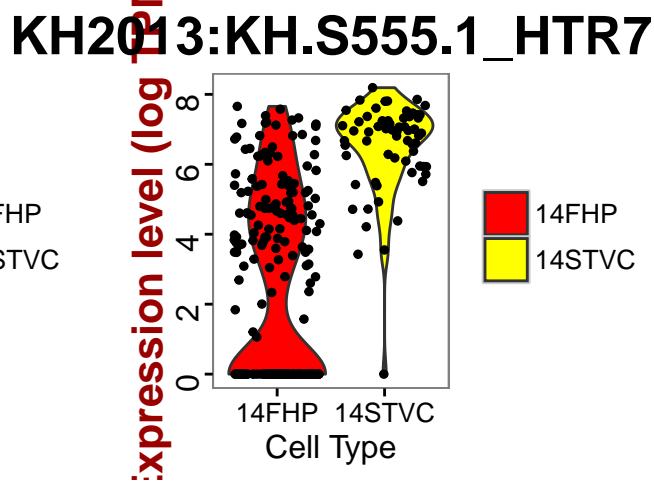
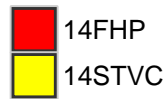
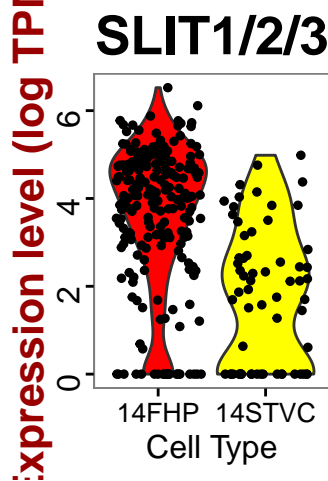
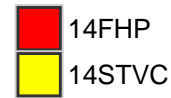
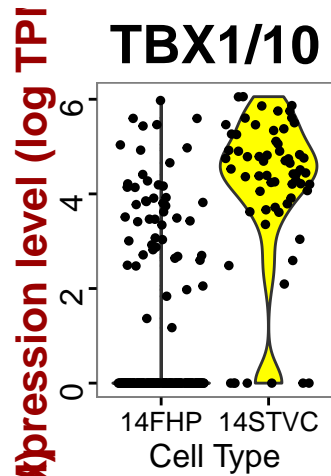
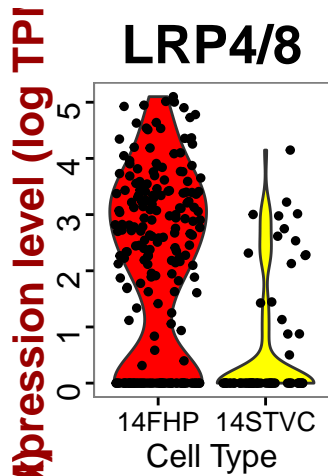
TBX1/10



KH2013:KH.S555.1_HTR7

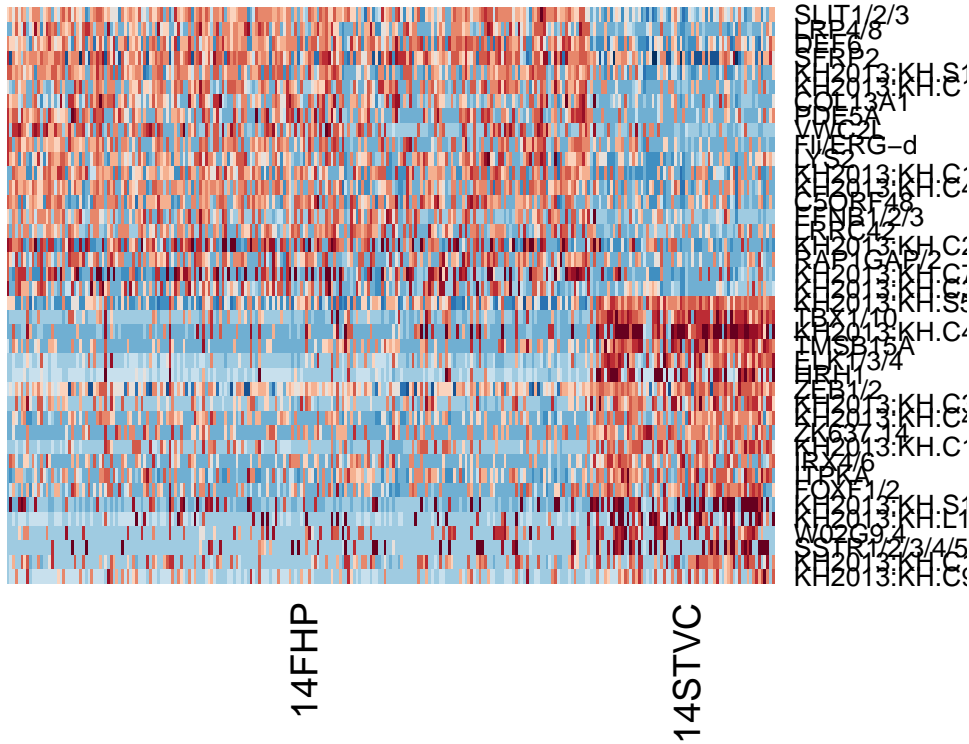


```
vlnPlot(hpf14, genes.viz.14, col = c("red", "yellow"))
```



```
# Select markers for plotting on a Heatmap (top 20 positive markers with
# high classification power)
marker14FHP = rownames(FHP_14.markers[order(FHP_14.markers$myAUC, decreasing = T)[1:20],
])
marker14STVC = rownames(STVC_14.markers[order(STVC_14.markers$myAUC, decreasing = T)[1:20],
])
marker.14 = c(marker14FHP, marker14STVC)

# Draw a heatmap of all cells for these marker genes
doHeatMap(hpf14, genes.use = marker.14, remove.key = TRUE, slim.col.label = T,
cex.col = 1.2, col.use = col, draw.line = F)
```

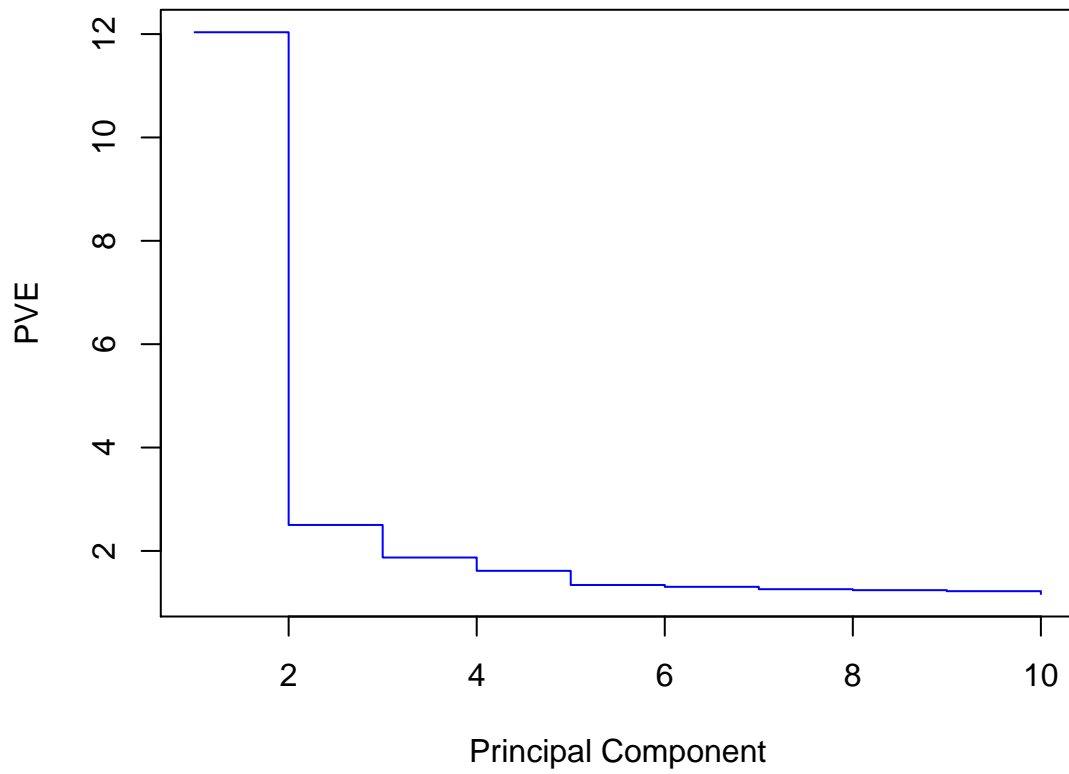


4. hpf20

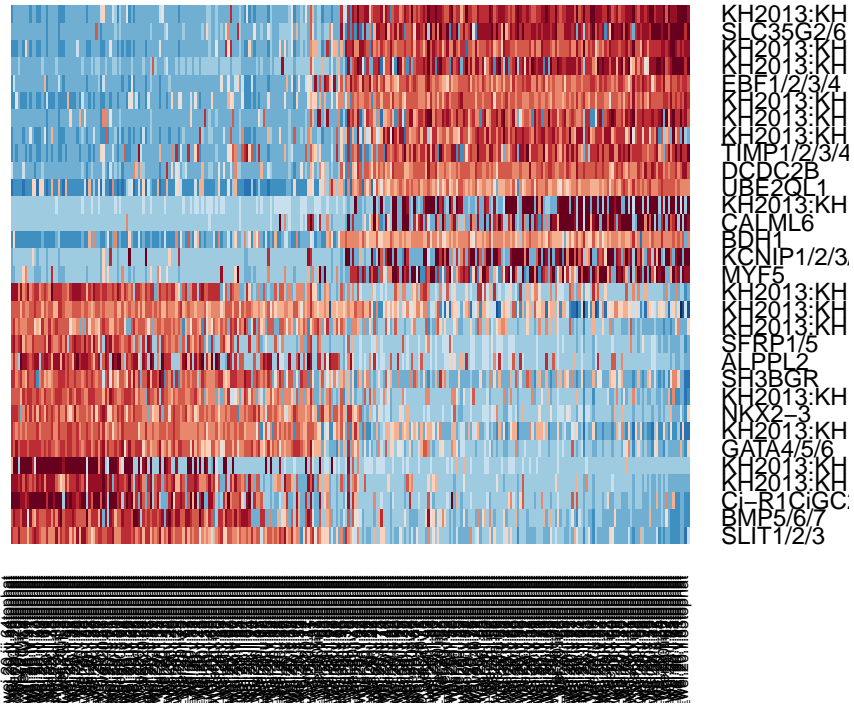
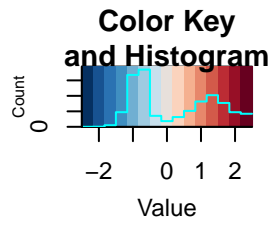
```
# Subset data from preprocessed Seurat object
hpf20 = subsetData(hpfall.remv2, which.cells(hpfall.remv2, "hpf20"), do.center = F,
  do.scale = F)
hpf20
```

```
## An object of class seurat in project allhpf
## 14864 genes across 288 samples.
```

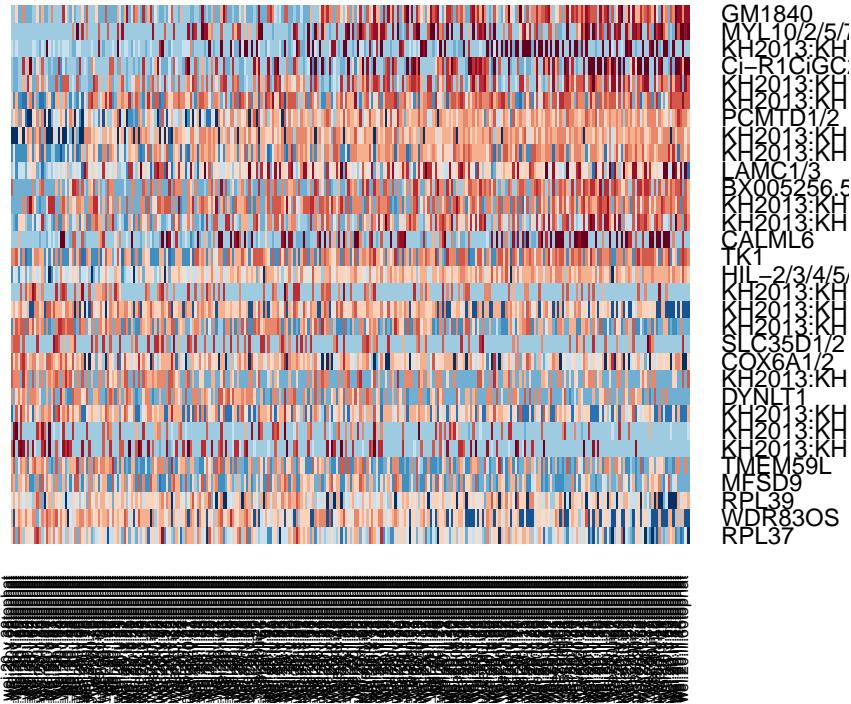
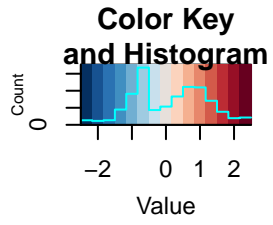
```
# Find variable gene with 4 < Average expression and Dispersion > 2
hpf20 = mean.var.plot(hpf20)
```

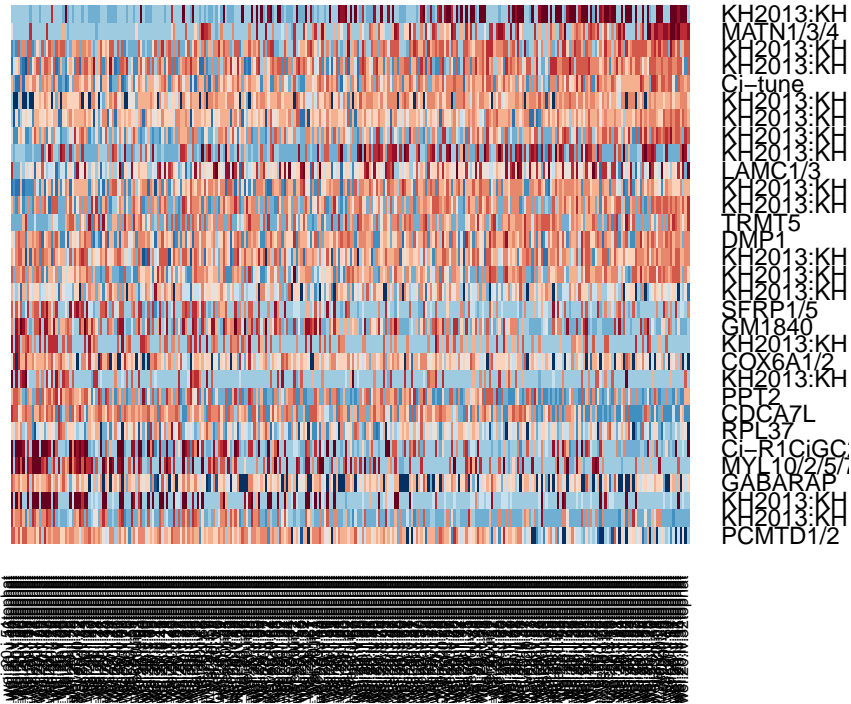
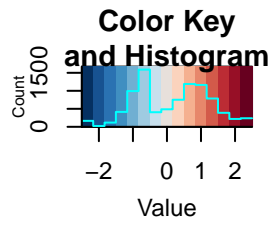
```
pcHeatmap(hpf20, 1, do.balanced = T, col.use = col)
```



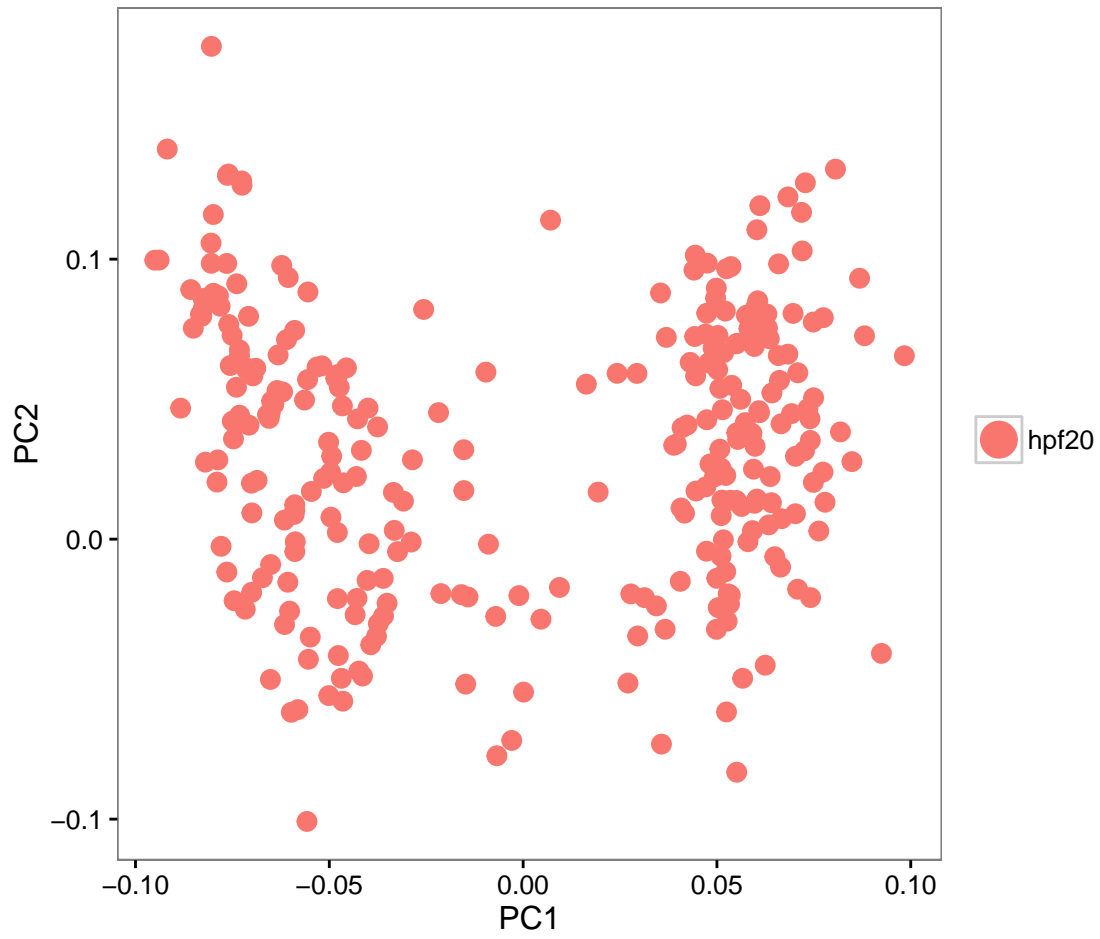
```
pcHeatmap(hpf20, 2, do.balanced = T, col.use = col)
```



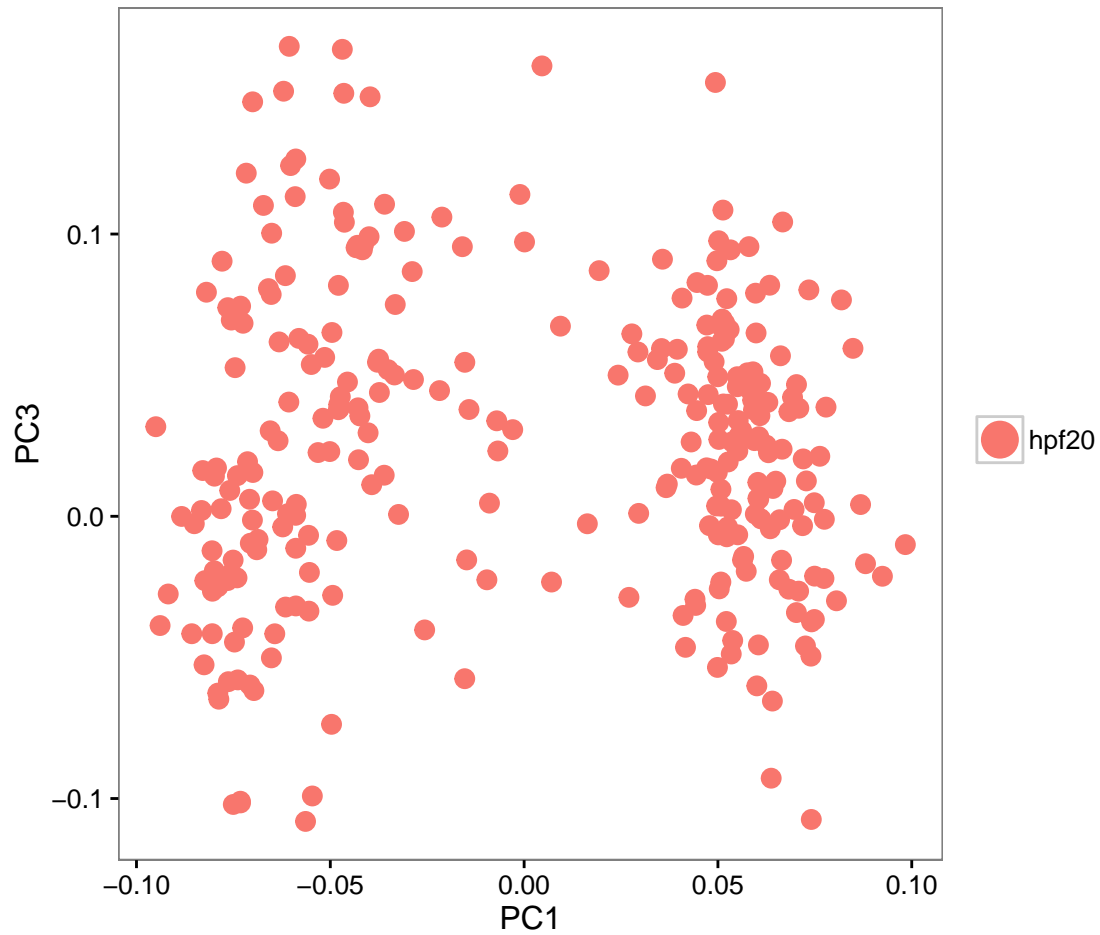
```
pcHeatmap(hpf20, 3, do.balanced = T, col.use = col)
```



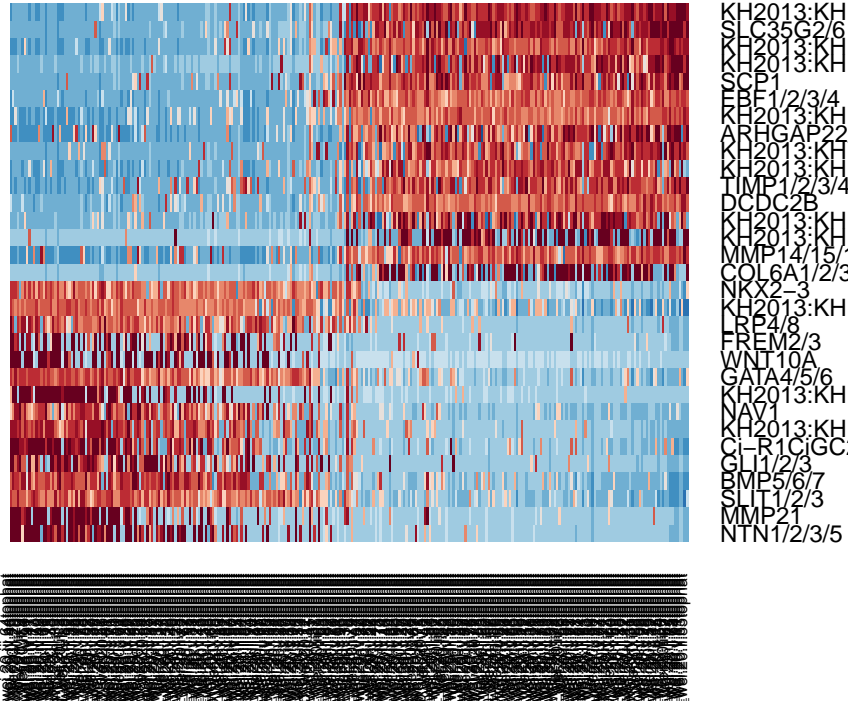
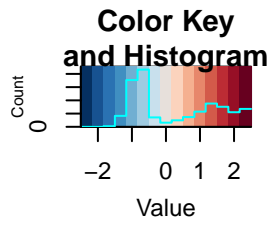
```
pca.plot(hpf20, 1, 2)
```



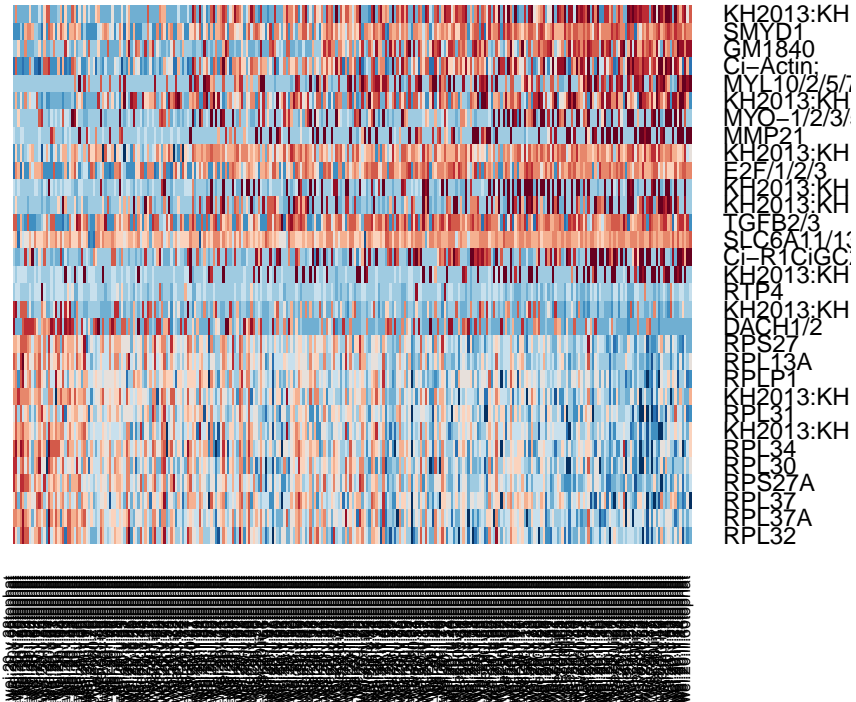
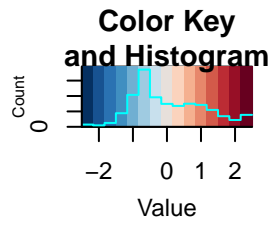
```
pca.plot(hpf20, 1, 3)
```



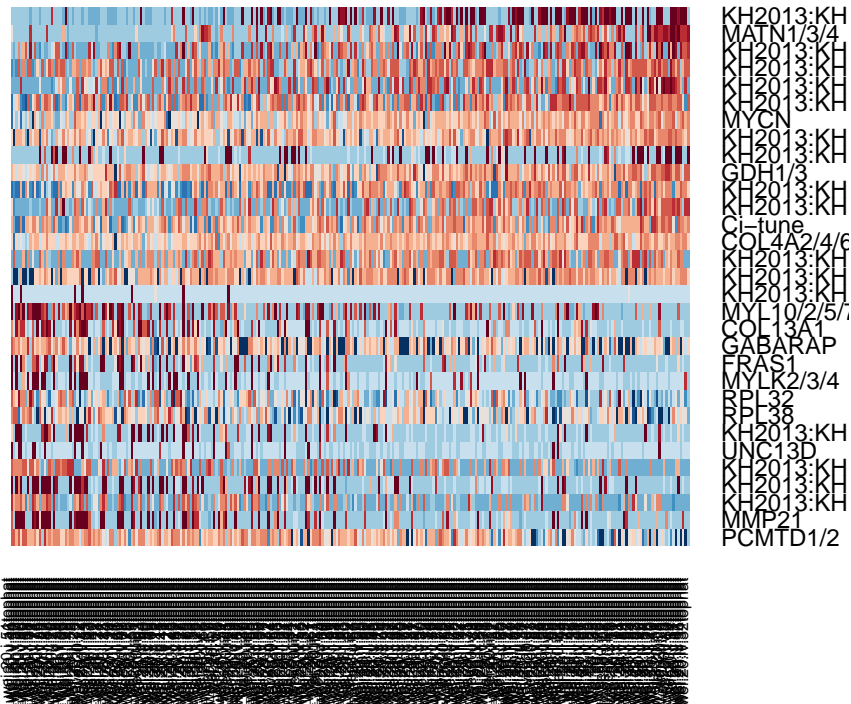
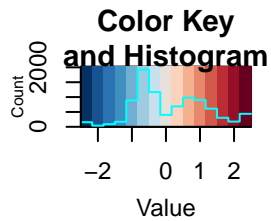
```
# Calculate PCA scores for all genes (PCA projection)  
hpf20 = project.pca(hpf20, do.print = F)  
  
# Visualize the full projected PCA, which now includes new genes which were  
# not previously (use.full=TRUE)  
pcHeatmap(hpf20, 1, use.full = T, do.balanced = T, col.use = col)
```



```
pcHeatmap(hpf20, 2, use.full = T, do.balanced = T, col.use = col) #technical
```

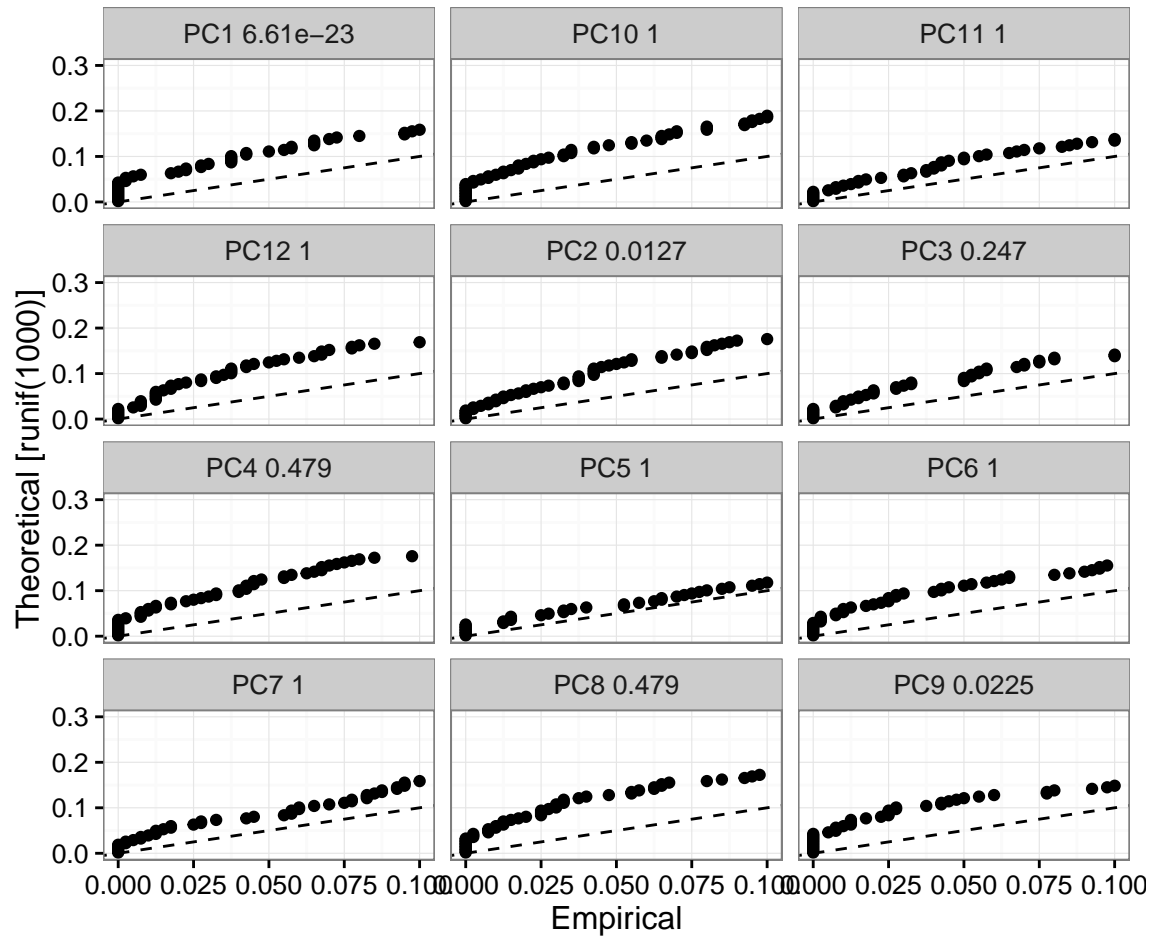



```
pcHeatmap(hpf20, 3, use.full = T, do.balanced = T, col.use = col)
```



```
# Do 200 random samplings to find significant genes, each time randomly
# permute 1% of genes This returns a 'p-value' for each gene in each PC,
# based on how likely the gene/PC score would have been observed by chance
hpf20 = jackStraw(hpf20, num.replicate = 200, do.print = F)
```

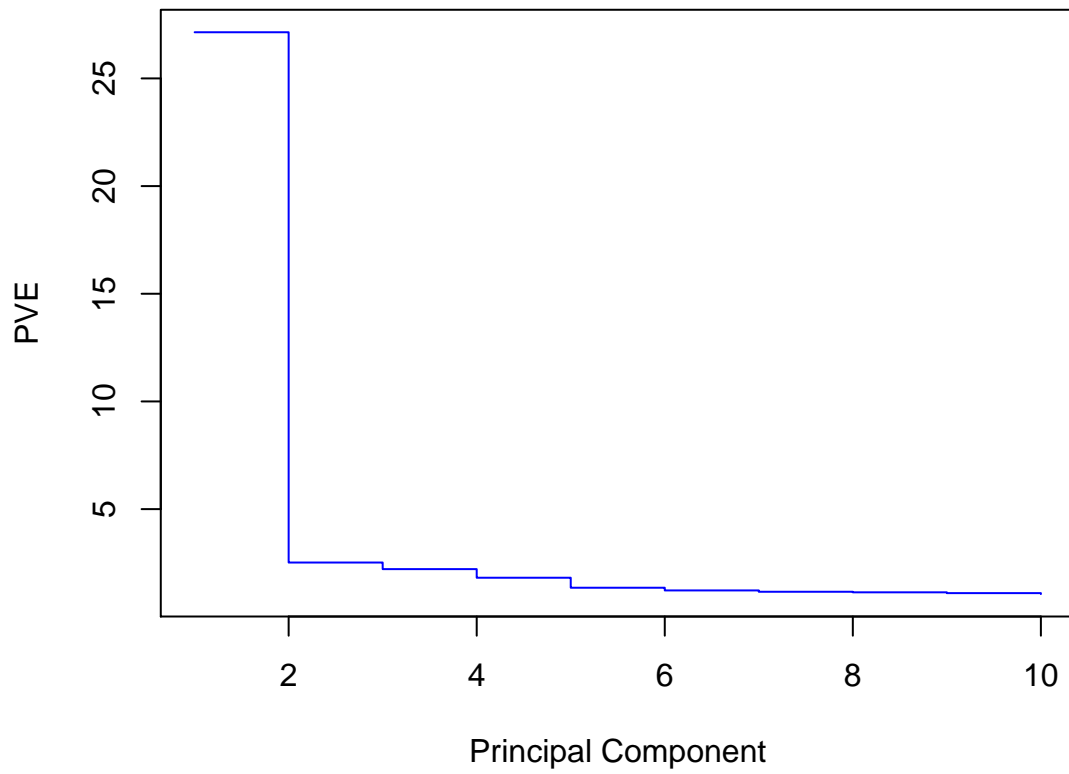
```
# The jackStraw plot compares the distribution of P-values for each PC with
# a uniform distribution (dashed line) 'Significant' PCs will have a strong
# enrichment of genes with low p-values (solid curve above dashed line)
jackStrawPlot.new(hpf20, PCs = 1:12)
```



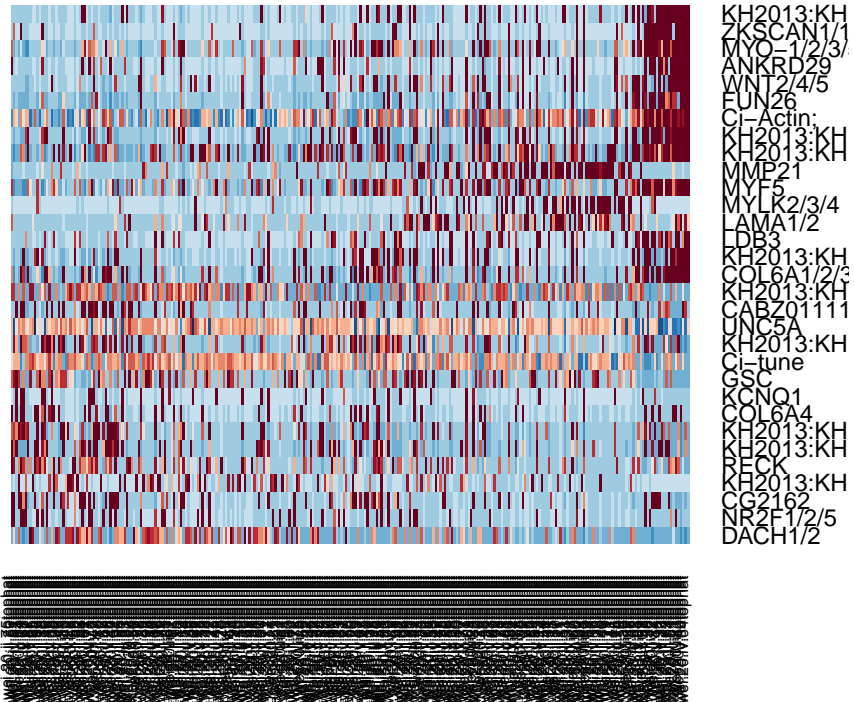
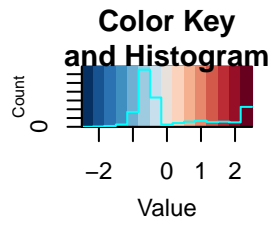
*# In this case only PC1 is strongly significant, PC2 is significant but
contain technical genes*

Select 300 genes from PC1 and rerun PCA
`good.genes20 = pcTopGenes(hpf20, 1, 300, T, T)`

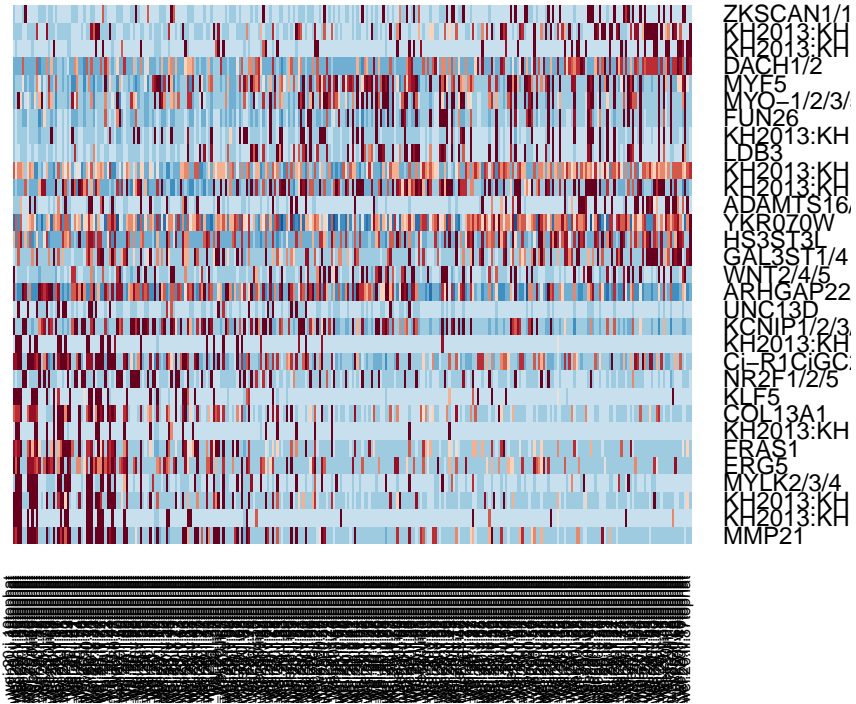
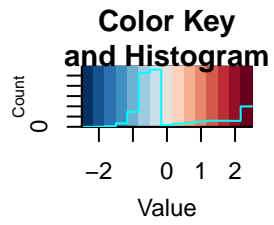
Run a PCA using selected gene list
`hpf20 = pca(hpf20, pc.genes = good.genes20, do.print = F)`
`pcScree(hpf20, good.genes20, 10)`



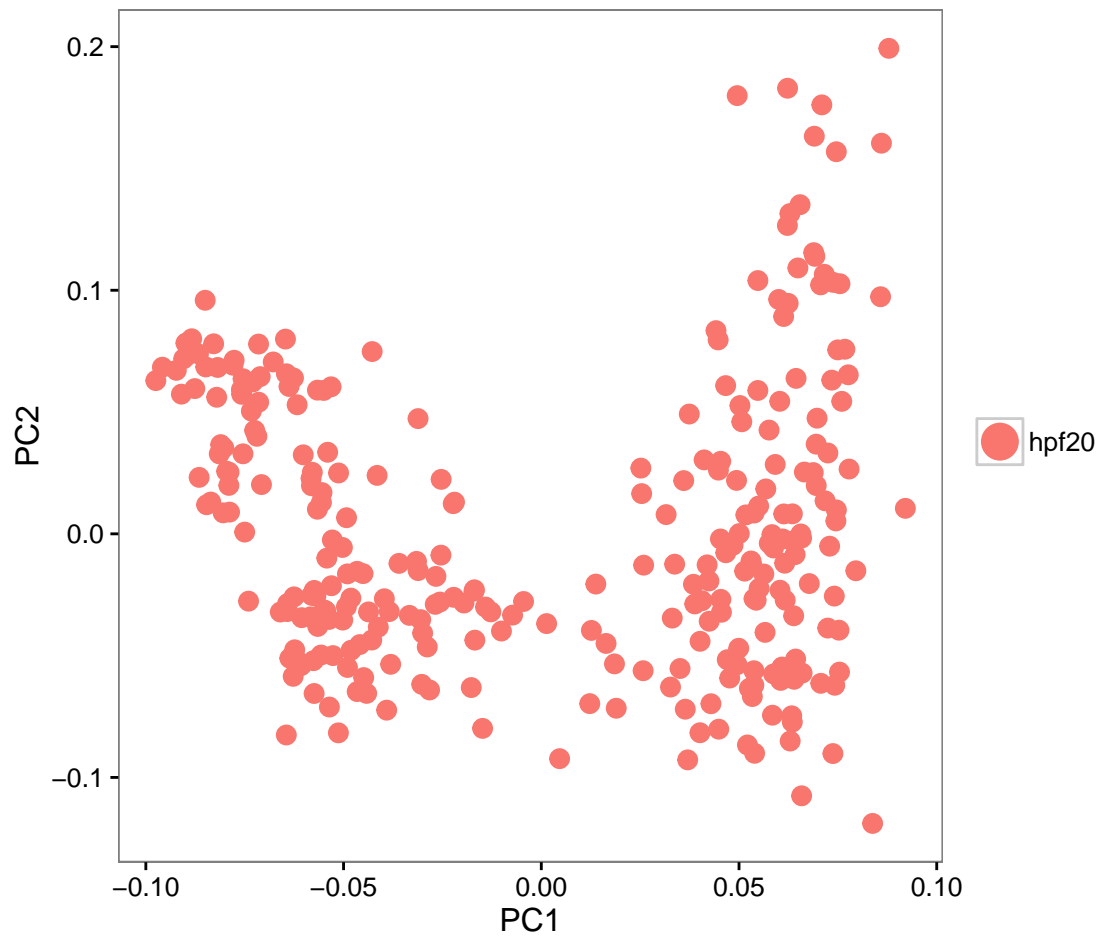
```
pcHeatmap(hpf20, 1, do.balanced = T, col.use = col)
```

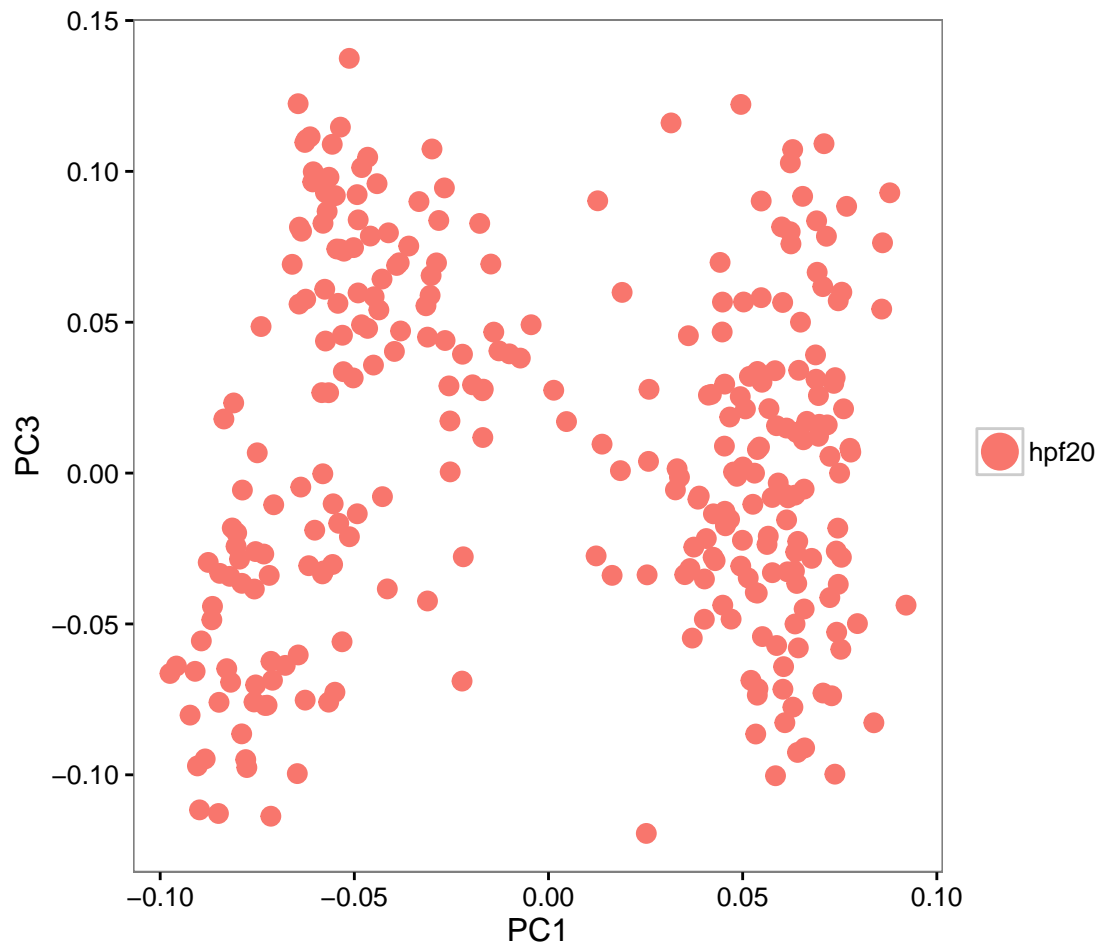
```
pcHeatmap(hpf20, 3, do.balanced = T, col.use = col)
```



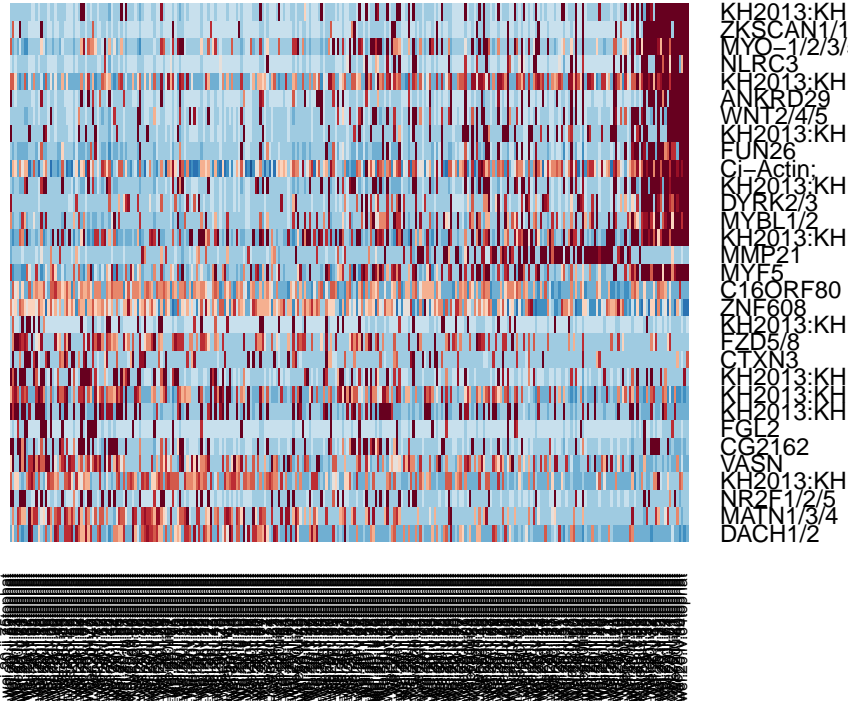
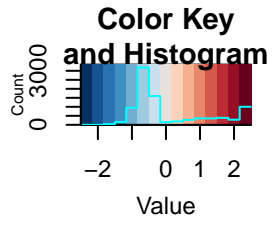
```
pca.plot(hpf20, 1, 2)
```



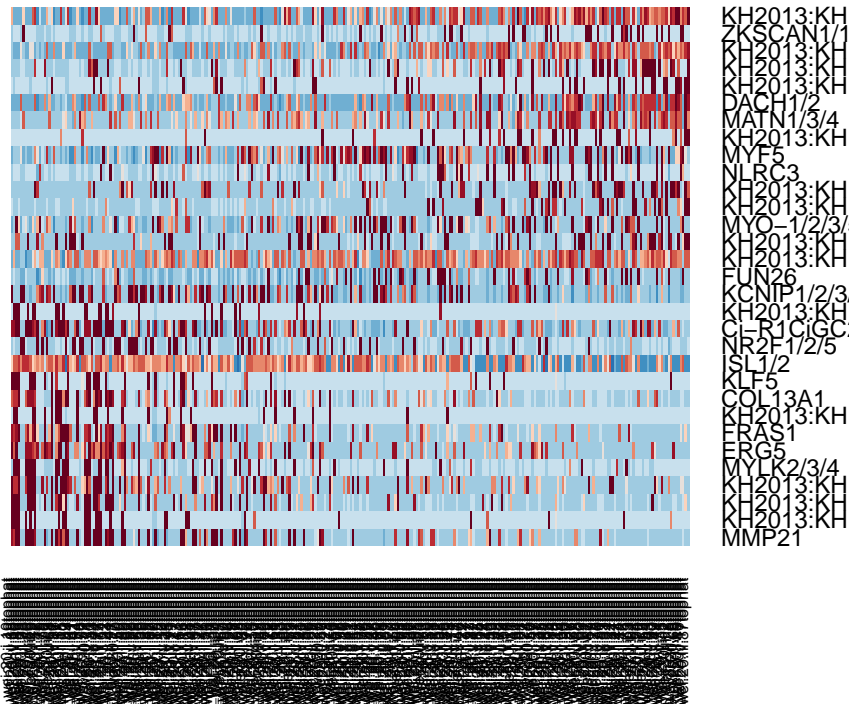
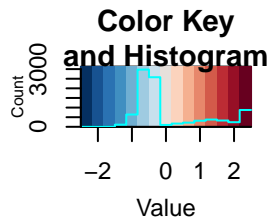
```
pca.plot(hpf20, 1, 3)
```

```
# Calculate PCA scores for all genes (PCA projection)  
hpf20 = project.pca(hpf20, do.print = F)  
  
# Visualize the full projected PCA, which now includes new genes which were  
# not previously (use.full=TRUE)  
pcHeatmap(hpf20, 1, use.full = T, do.balanced = T, col.use = col)
```

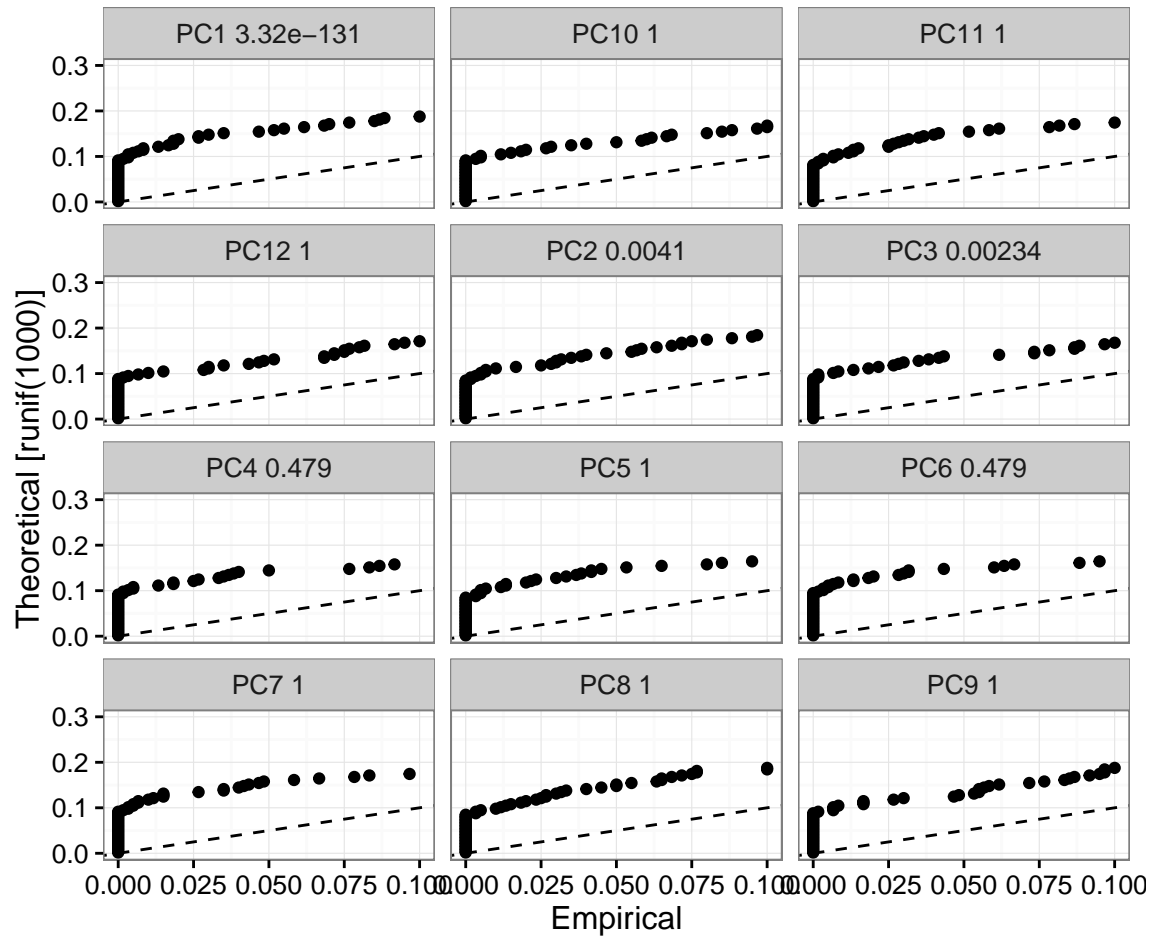



```
pcHeatmap(hpf20, 3, use.full = T, do.balanced = T, col.use = col)
```



```
# Do 200 random samplings to find significant genes, each time randomly
# permute 1% of genes This returns a 'p-value' for each gene in each PC,
# based on how likely the gene/PC score would have been observed by chance
hpf20 = jackStraw(hpf20, num.replicate = 200, do.print = F)
```

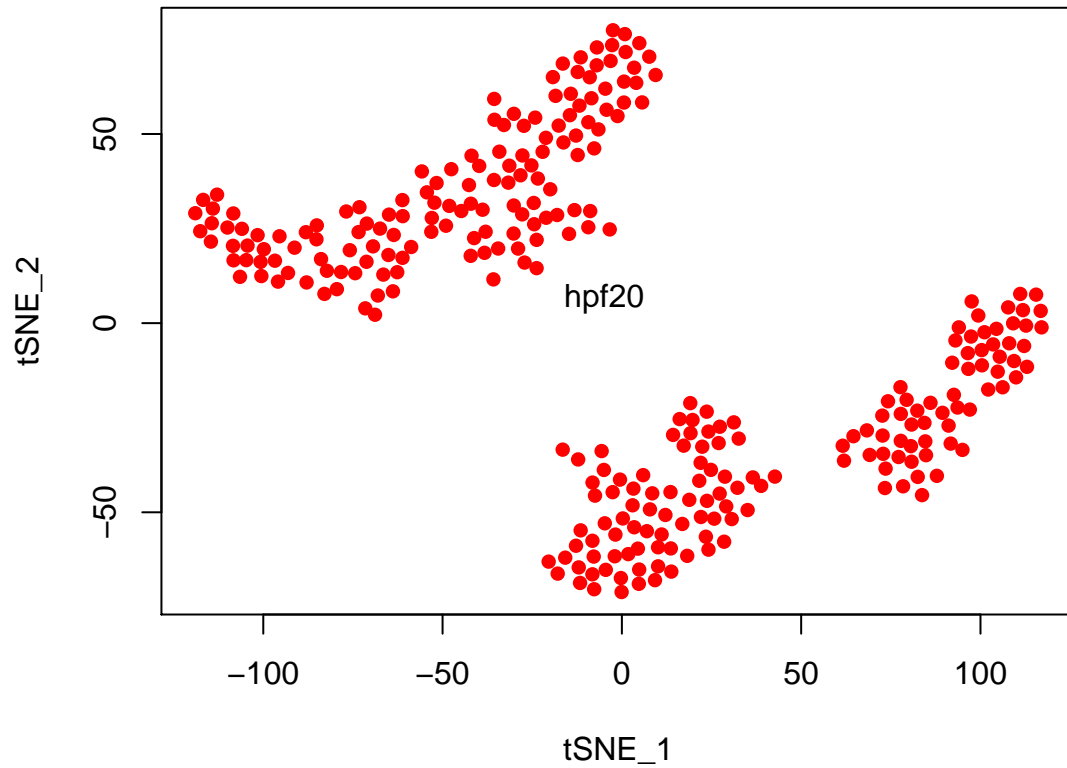
```
# The jackStraw plot compares the distribution of P-values for each PC with
# a uniform distribution (dashed line) 'Significant' PCs will have a strong
# enrichment of genes with low p-values (solid curve above dashed line)
jackStrawPlot.new(hpf20, PCs = 1:12)
```



In this case PC1-3 are significant

*# Run tSNE using significant PCs as input (spectral tSNE), we get distinct
point clouds*

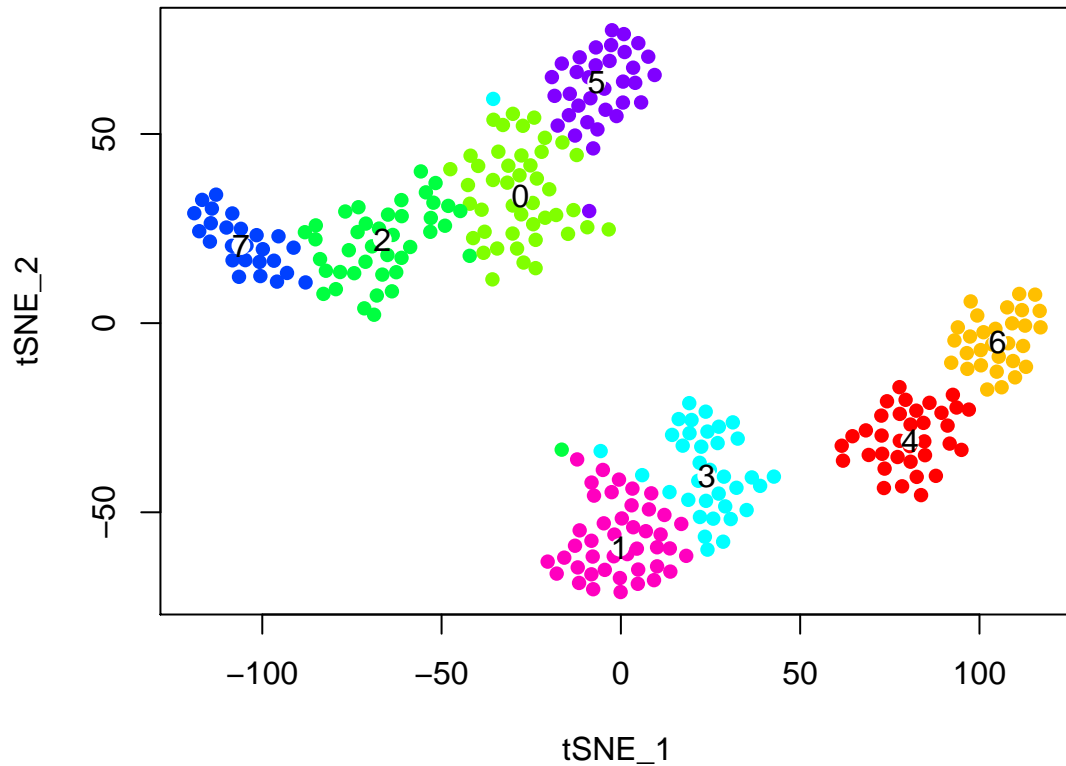
```
hpf20 = run_tsne(hpf20, max_iter = 2000, dims.use = 1:3)
tsne.plot(hpf20, do.label = T, label.pt.size = 1)
```



```
# Find cell clusters using Modularity optimization cluster detection.  
hpf20 = FindClusters(hpf20, pc.use = 1:3, do.modularity = T, resolution = 1,  
  prune.SNN = 0.1, print.output = 0, k.param = 20)
```

```
## [1] "SNN : processed 72 cells"  
## [1] "SNN : processed 144 cells"  
## [1] "SNN : processed 216 cells"  
## [1] "SNN : processed 288 cells"
```

```
tsne.plot(hpf20, do.label = T, label.pt.size = 1)
```



```
# The validity of the clusters can be validated using a classification  
# scheme based on linear SVMs.
```

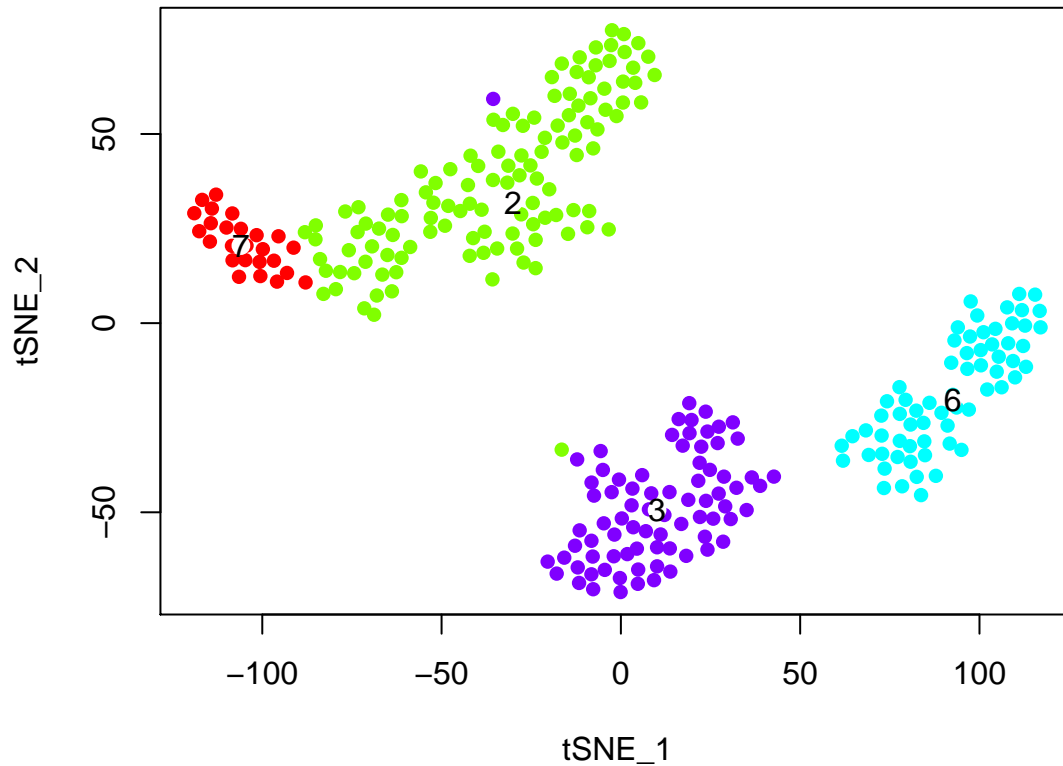
```
hpf20 = BuildSNN(hpf20, pc.use = 1:3, do.sparse = T, k.param = 20)
```

```
## [1] "SNN : processed 72 cells"  
## [1] "SNN : processed 144 cells"  
## [1] "SNN : processed 216 cells"  
## [1] "SNN : processed 288 cells"
```

```
hpf20 = ValidateClusters(hpf20, pc.use = 1:3, min.connectivity = 0.001, acc.cutoff = 0.8)
```

```
## [1] " 0% complete --- merge clusters 0 and 5, classification accuracy of 0.6994"  
## [1] " 14% complete --- merge clusters 1 and 3, classification accuracy of 0.7552"  
## [1] " 57% complete --- merge clusters 5 and 2, classification accuracy of 0.8306"  
## [1] " 57% complete --- merge clusters 4 and 6, classification accuracy of 0.8324"  
## [1] "100% complete --- started with 8 clusters, 4 clusters remaining"
```

```
tsne.plot(hpf20, do.label = T, label.pt.size = 1)
```



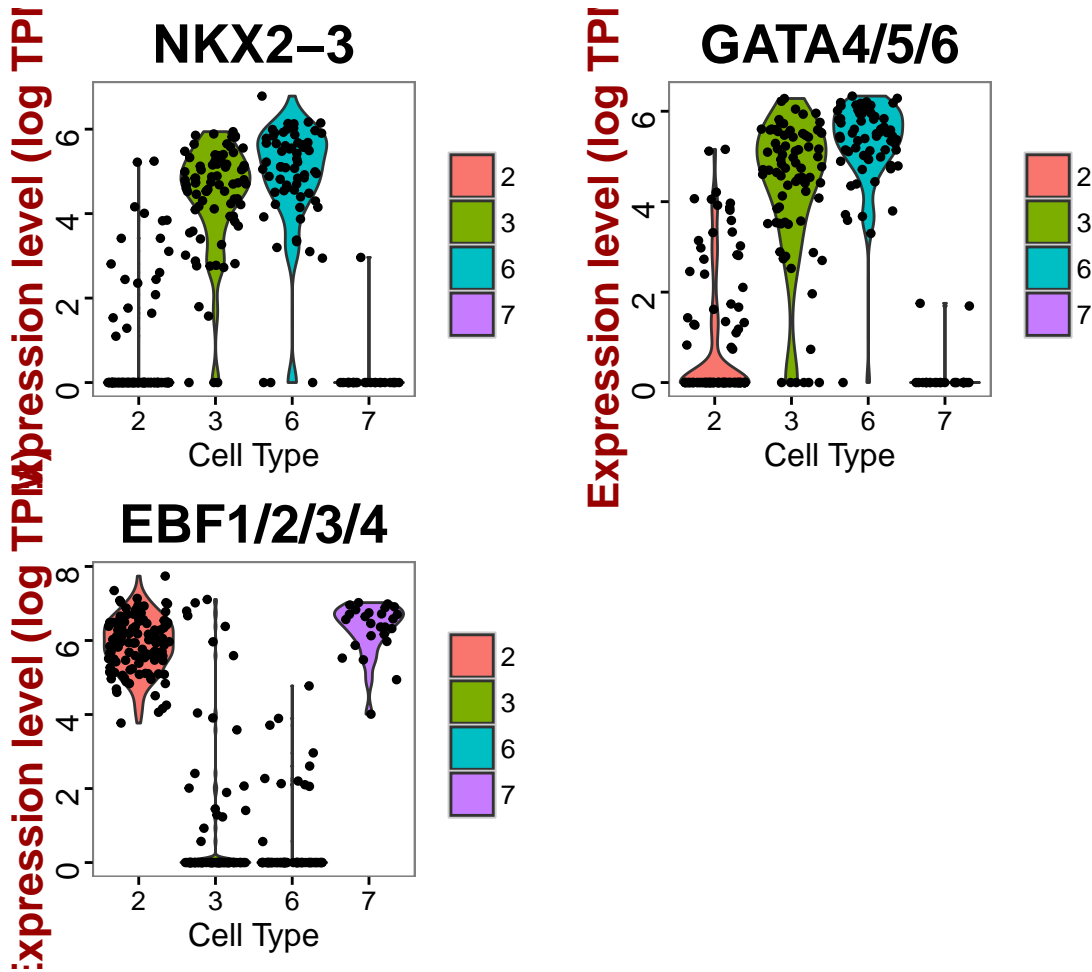
```
# Find cluster markers using ROC test with thresh.use = 1, min.pct = 0.5 The
# ROC test returns the 'classification power' for any individual marker
# (ranging from 0 - random, to 1 - perfect). Though not a statistical test,
# it is often very useful for finding clean markers. Find markers for
# cluster 2
```

```
c12_20.markers = find.markers(hpf20, 2, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(c12_20.markers[order(c12_20.markers$myAUC, decreasing = T), ], 20)
```

##	myAUC	avg_diff	power	pct.1	pct.2
## KH2013:KH.C11.139	0.930	2.042404	0.860	0.992	0.337
## KH2013:KH.L25.4	0.918	1.888086	0.836	0.992	0.231
## KH2013:KH.C5.62_ODF3L2	0.899	1.562319	0.798	0.983	0.278
## KH2013:KH.L18.10_THSD7A	0.894	1.571177	0.788	0.983	0.331
## SCP1	0.883	1.728818	0.766	0.958	0.278
## KH2013:KH.C4.506_HMCN1	0.882	1.291218	0.764	1.000	0.858
## KH2013:KH.S1269.1	0.880	1.388814	0.760	0.992	0.533
## SLC35G2/6	0.875	1.853114	0.750	0.899	0.225
## UBE2QL1	0.874	1.422110	0.748	0.966	0.432
## KH2013:KH.C6.201	0.870	1.898804	0.740	0.874	0.183
## MMP14/15/16/24	0.868	1.387104	0.736	0.975	0.355
## ARHGAP22/24/25	0.868	1.743081	0.736	0.891	0.260
## FRS2/3	0.866	1.408759	0.732	0.975	0.609
## EBF1/2/3/4	0.865	1.253061	0.730	1.000	0.314


```
## DCDC2B          0.863 1.184517 0.726 0.983 0.325
## BDH1            0.862 1.240573 0.724 0.983 0.396
## NOVA1/2        0.856 1.161588 0.712 1.000 0.462
## Ci-R1CiGC09b24; 0.853 1.216660 0.706 0.983 0.888
## TIMP1/2/3/4    0.852 1.920971 0.704 0.882 0.349
## HLH-4          0.845 1.462400 0.690 0.950 0.544
```

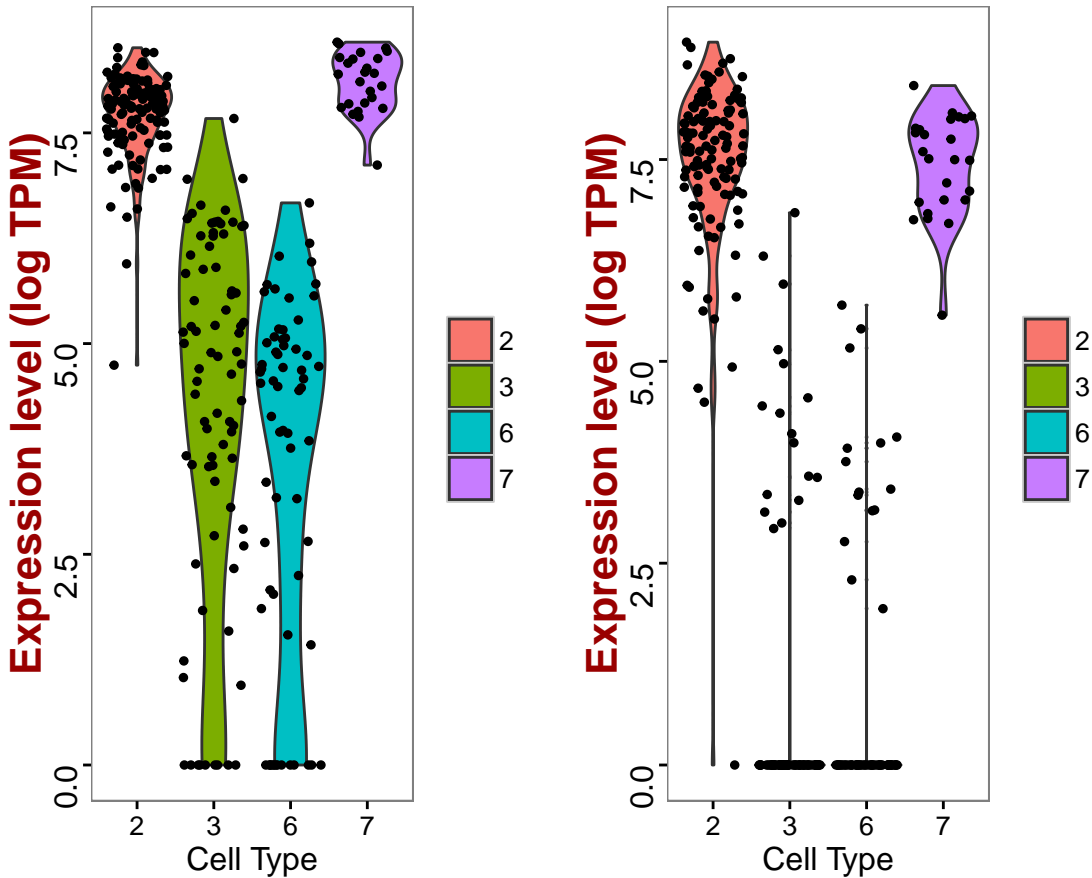
```
# Visualize known markers with a violin plot
vlnPlot(hpf20, c("NKX2-3", "EBF1/2/3/4", "GATA4/5/6"))
```



```
# Based on preliminary studies, these EBF1/2/3/4+ NKX- GATA- cells are ASM1
# cells
```

```
# Visualize new markers with a violin plot
vlnPlot(hpf20, c("KH2013:KH.C4.506_HMCN1", "KH2013:KH.C11.139"))
```

2013:KH.C4.506_HMCN KH2013:KH.C11.139



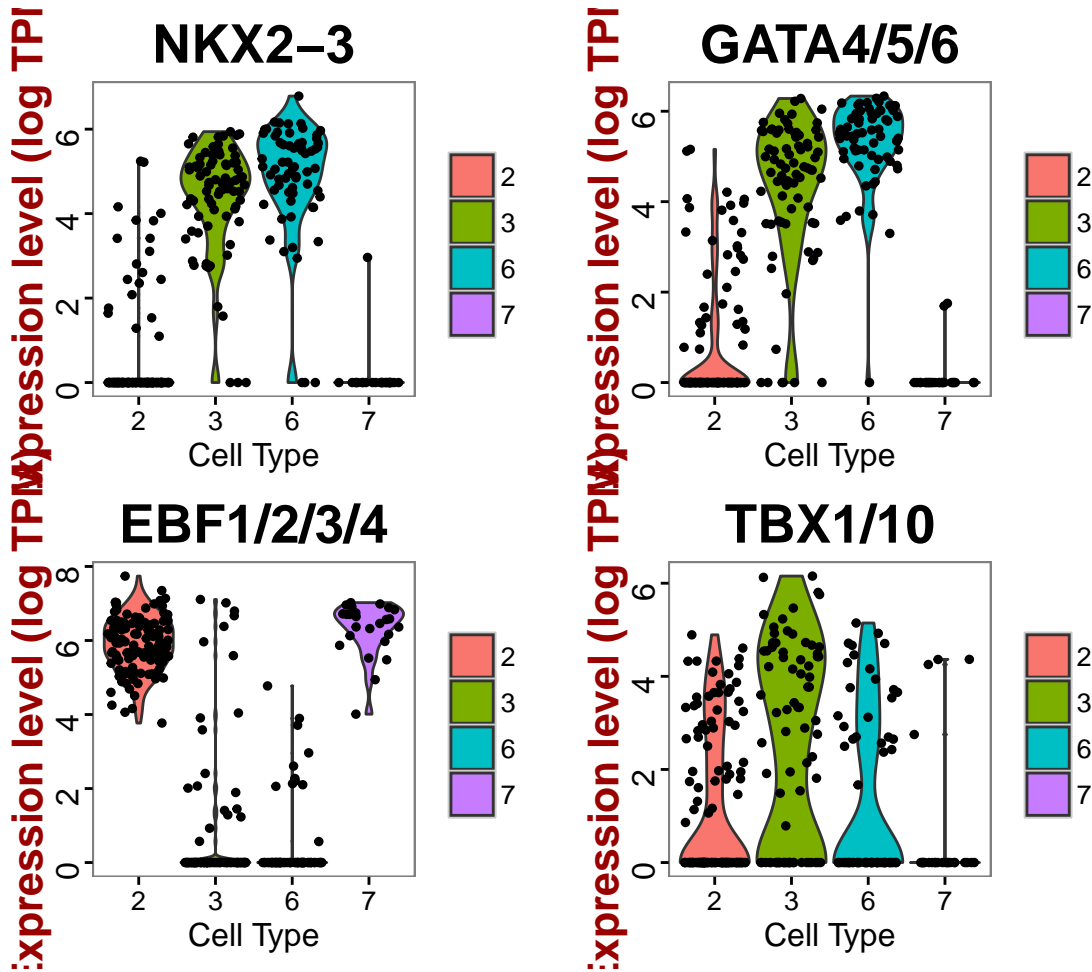
```
# Find markers for cluster 3
```

```
cl3_20.markers = find.markers(hpf20, 3, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(cl3_20.markers[order(cl3_20.markers$myAUC, decreasing = T), ], 20)
```

##	myAUC	avg_diff	power	pct.1	pct.2
## KH2013:KH.C1.638_C17ORF105	0.842	1.727786	0.684	0.861	0.325
## KH2013:KH.C4.125_BMP2/4	0.841	1.652993	0.682	0.937	0.517
## DACH1/2	0.788	1.930722	0.576	0.734	0.278
## KH2013:KH.C4.226	0.787	1.372513	0.574	0.911	0.574
## KH2013:KH.C2.209_CALM1/3	0.775	1.004900	0.550	0.924	0.646
## FOXF1/2	0.767	1.249064	0.534	0.873	0.512
## YKR070W	0.750	1.143960	0.500	0.899	0.699
## KH2013:KH.L153.109	0.745	2.030674	0.490	0.595	0.124
## SNX11	0.739	1.187810	0.478	0.747	0.297
## KH2013:KH.S1887.1	0.716	1.458843	0.432	0.620	0.254
## KH2013:KH.C11.667	0.714	1.038638	0.428	0.886	0.727
## PCTP	0.704	1.024480	0.408	0.709	0.368
## MATN1/3/4	0.695	1.452748	0.390	0.633	0.368
## HS3ST3L	0.685	1.075695	0.370	0.671	0.378

```
## TBX1/10          0.681  1.420956 0.362 0.633 0.373
## KANK1/2/3/4     0.640  1.073334 0.280 0.595 0.411
## KH2013:KH.C9.40_GST01/2 0.615  1.051532 0.230 0.506 0.335
## PIGT            0.502 -5.064610 0.004 0.570 0.569
## RBM15B          0.433 -3.685868 0.134 0.646 0.789
## PSMD14          0.430 -3.980281 0.140 0.835 0.928
```

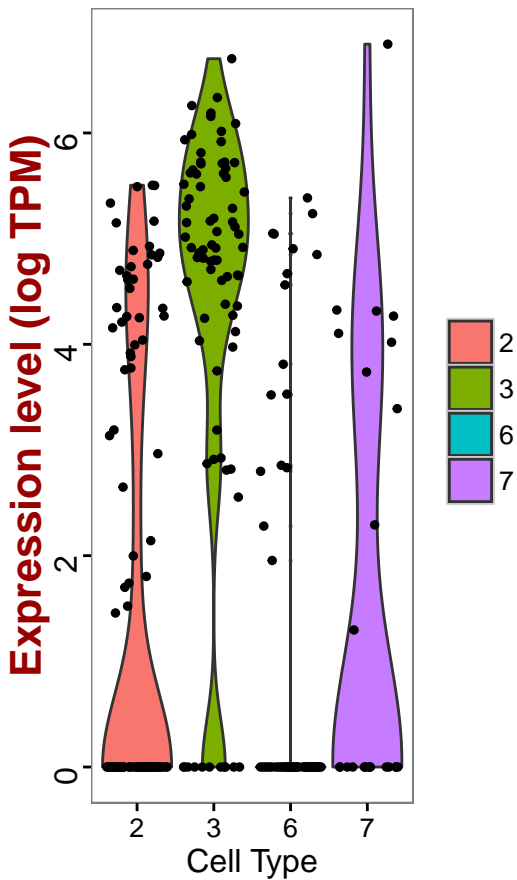
```
# Visualize known markers with a violin plot
vlnPlot(hpf20, c("NKX2-3", "EBF1/2/3/4", "GATA4/5/6", "TBX1/10"))
```



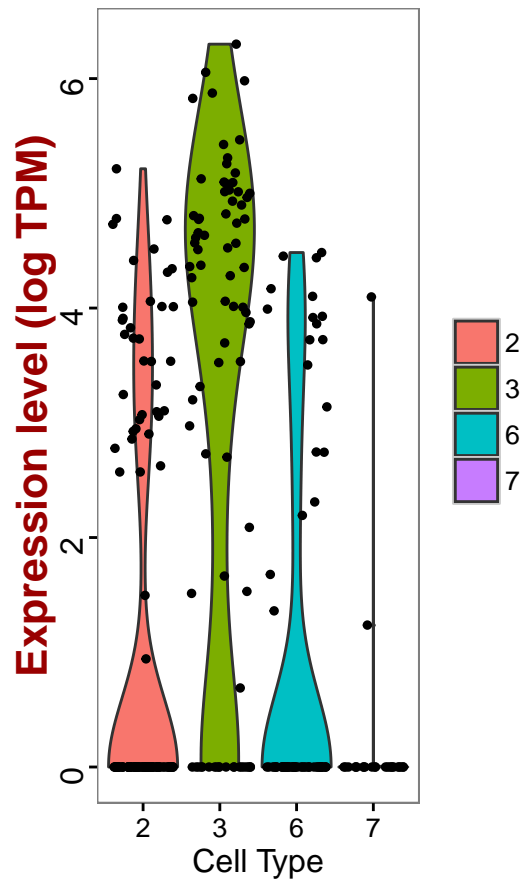
```
# Based on preliminary studies, these EBF1/2/3/4- NKX+ GATA- TBX1/10+ cells
# are SHP cells
```

```
# Visualize new markers with a violin plot
vlnPlot(hpf20, c("KH2013:KH.C1.638_C17ORF105", "DACH1/2"))
```

13:KH.C1.638_C17ORF



DACH1/2



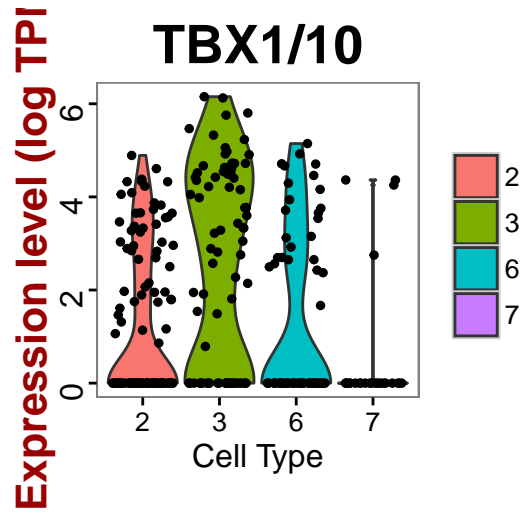
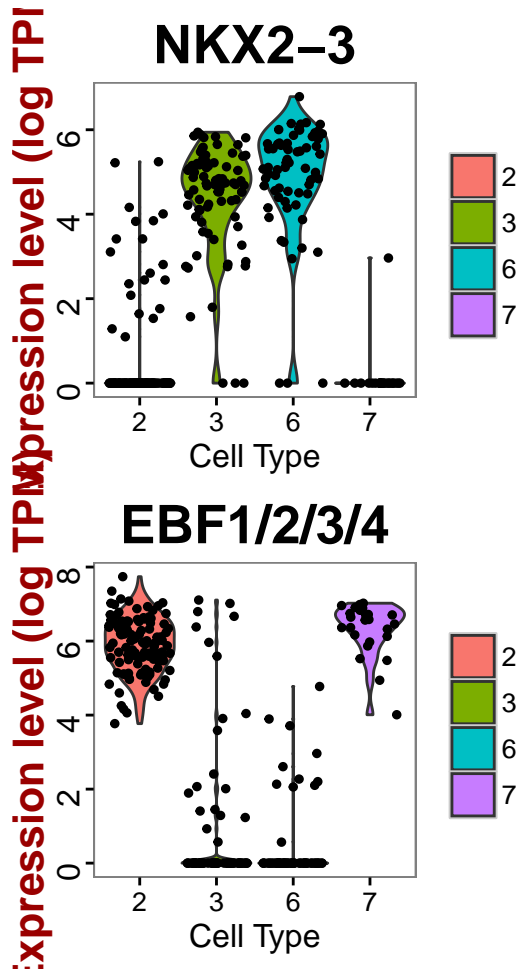
Find markers for cluster 6

```
cl6_20.markers = find.markers(hpf20, 6, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(cl6_20.markers[order(cl6_20.markers$myAUC, decreasing = T), ], 20)
```

Gene	myAUC	avg_diff	power	pct.1
Ci-R1CiGC27a04	0.962	3.284688	0.924	0.954
KH2013:KH.C4.547_BMP2/4	0.951	2.195473	0.902	0.985
KH2013:KH.C5.227_F56C4.4	0.944	1.991633	0.888	1.000
KH2013:KH.C11.378_CLDN1/10/14/19/2/3/4/5/6/7/9	0.937	1.539702	0.874	1.000
KH2013:KH.C1.479_NEB	0.927	1.680541	0.854	1.000
MMP21	0.927	3.901756	0.854	0.877
FRAS1	0.911	2.683945	0.822	0.831
SFRP1/5	0.910	2.269762	0.820	0.923
KH2013:KH.C10.203	0.909	2.466237	0.818	0.923
COL13A1	0.907	3.395970	0.814	0.846
KH2013:KH.C1.953_CG32702	0.906	2.177912	0.812	0.938
KH2013:KH.C7.205_ASB2	0.904	1.818015	0.808	0.969
NTN1/2/3/5	0.902	2.350696	0.804	0.892
KH2013:KH.C2.994_RNF149	0.900	1.278227	0.800	1.000

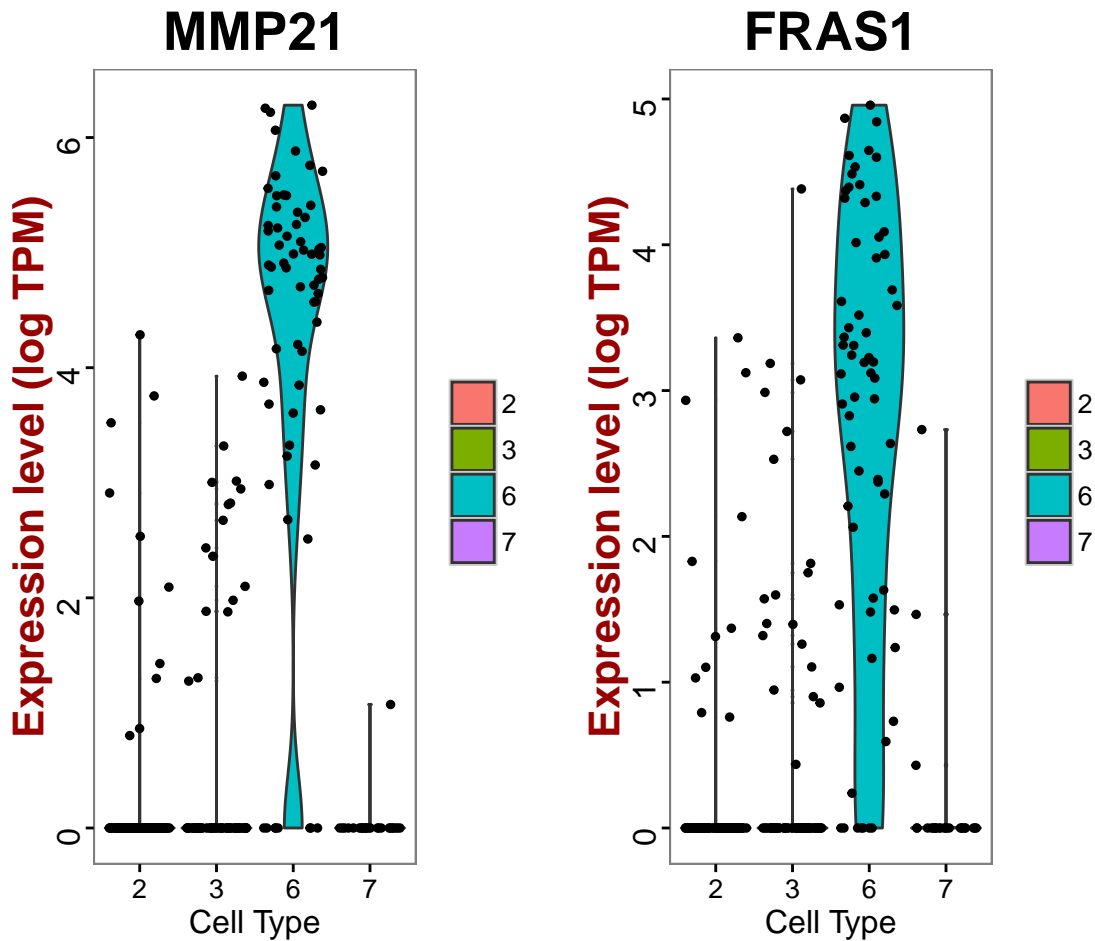
## KH2013:KH.C3.665_ZAN	0.896 1.455140 0.792 0.985
## MTDH	0.895 1.022437 0.790 1.000
## C5ORF48	0.894 2.399851 0.788 0.892
## GATA4/5/6	0.891 1.513036 0.782 0.985
## KH2013:KH.C3.191_WBSCR27	0.886 1.950294 0.772 0.938
## KH2013:KH.C1.738_CG9550	0.881 1.235709 0.762 0.985
##	pct.2
## Ci-R1CiGC27a04	0.238
## KH2013:KH.C4.547_BMP2/4	0.381
## KH2013:KH.C5.227_F56C4.4	0.973
## KH2013:KH.C11.378_CLDN1/10/14/19/2/3/4/5/6/7/9	0.946
## KH2013:KH.C1.479_NEB	0.605
## MMP21	0.117
## FRAS1	0.117
## SFRP1/5	0.193
## KH2013:KH.C10.203	0.247
## COL13A1	0.108
## KH2013:KH.C1.953_CG32702	0.309
## KH2013:KH.C7.205_ASB2	0.430
## NTN1/2/3/5	0.166
## KH2013:KH.C2.994_RNF149	0.857
## KH2013:KH.C3.665_ZAN	0.628
## MTDH	0.991
## C5ORF48	0.229
## GATA4/5/6	0.471
## KH2013:KH.C3.191_WBSCR27	0.435
## KH2013:KH.C1.738_CG9550	0.897

```
# Visualize known markers with a violin plot
vlnPlot(hpf20, c("NKX2-3", "EBF1/2/3/4", "TBX1/10"))
```



```
# Confirmed with these markers cluster 5 NKX+ EBF1/2/3/4- TBX1/10- cells
# are FHP cells
```

```
# Visualize new markers with a violin plot
vlnPlot(hpf20, c("MMP21", "FRAS1"))
```



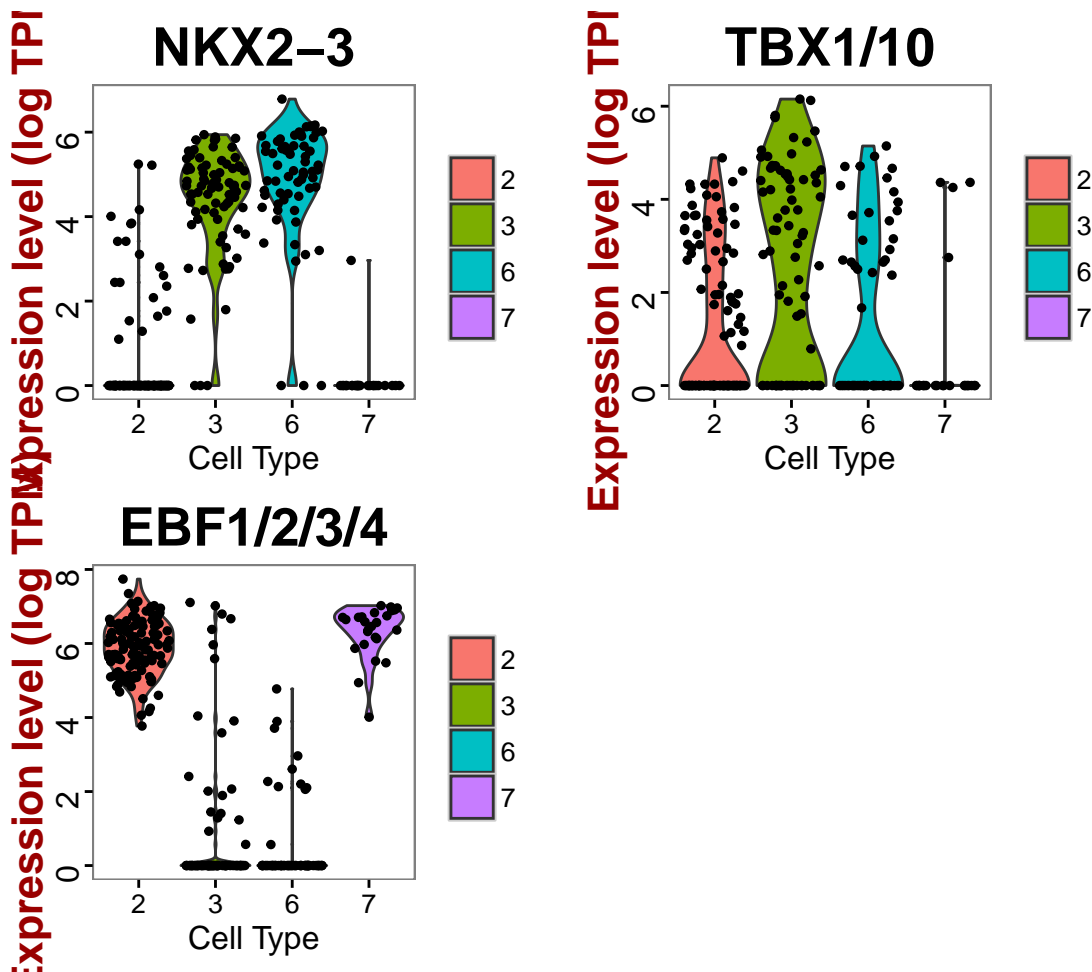
```
# Find markers for cluster 7
```

```
cl7_20.markers = find.markers(hpf20, 7, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(cl7_20.markers[order(cl7_20.markers$myAUC, decreasing = T), ], 20)
```

##	myAUC	avg_diff	power	pct.1	pct.2
## MYF5	0.936	2.237196	0.872	1.00	0.403
## KH2013:KH.C8.649_ACTA1/2	0.927	2.296572	0.854	1.00	0.589
## DCDC2B	0.914	1.380557	0.828	1.00	0.559
## MYO-1/2/3/5	0.914	2.429771	0.828	0.92	0.278
## CALML6	0.905	2.418975	0.810	0.96	0.304
## FUN26	0.905	2.611697	0.810	0.88	0.106
## RBM24/38	0.902	1.965112	0.804	0.96	0.384
## SMYD1	0.902	1.604893	0.804	1.00	0.802
## MYO10	0.899	1.617349	0.798	1.00	0.441
## Ci-Actin;	0.894	2.092404	0.788	0.96	0.738
## KH2013:KH.C4.506_HMCN1	0.887	1.052516	0.774	1.00	0.909
## KH2013:KH.C5.522_SLC6A13	0.887	1.453291	0.774	1.00	0.471
## ZKSCAN1/14/17/2/3/6	0.885	3.396235	0.770	0.80	0.080
## KH2013:KH.C12.669_PHEX	0.883	1.820194	0.766	0.96	0.407

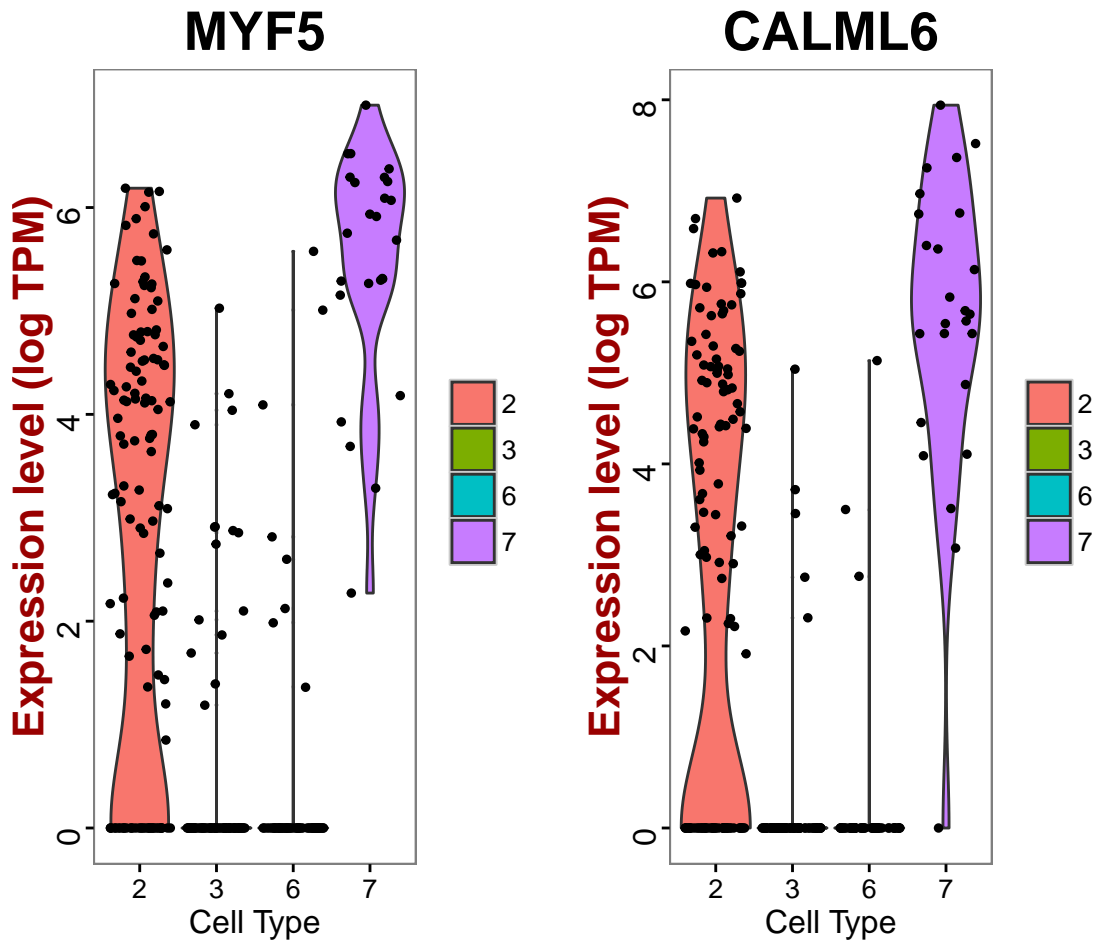
```
## KH2013:KH.C7.476_CHRNB1      0.880 1.952671 0.760 0.92 0.304
## KH2013:KH.C12.419_TSPAN4/9  0.878 1.252526 0.756 1.00 0.821
## KH2013:KH.C1.509             0.876 2.059997 0.752 0.88 0.205
## KH2013:KH.S643.6_CDKN1B     0.876 1.562963 0.752 1.00 0.612
## RBF1/2/3                     0.875 1.347928 0.750 1.00 0.863
## KH2013:KH.C5.404_MBL1/2     0.875 2.442091 0.750 0.84 0.114
```

```
# Visualize known markers with a violin plot
vlnPlot(hpf20, c("NKX2-3", "EBF1/2/3/4", "TBX1/10"))
```



```
# Confirmed with these markers cluster 5 NKX+ EBF1/2/3/4- TBX1/10- cells
# are ASM2 cells
```

```
# Visualize new markers with a violin plot
vlnPlot(hpf20, c("MYF5", "CALML6"))
```

```
# MYF5 is a more differentiated marker, this group of ASM is more
# deferentiated ASM cells.
```

```
# Write cell names into text files
```

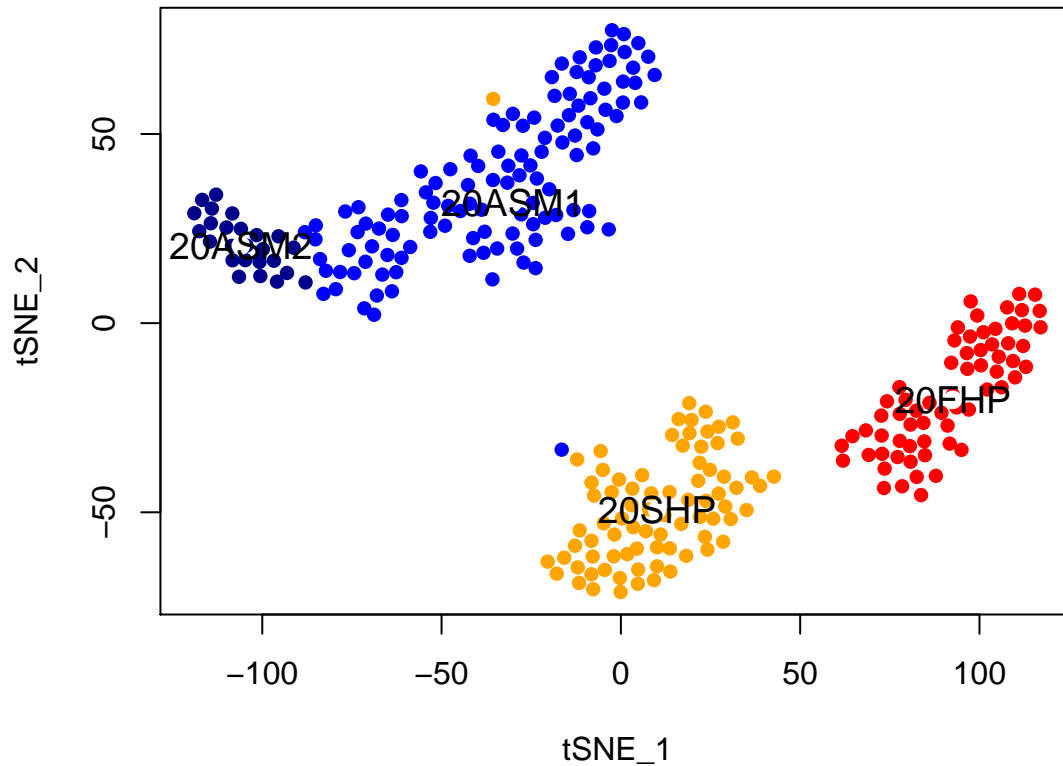
```
write.table(which.cells(hpf20, 2), file = "20ASM1Cells.txt", sep = "\t")
write.table(which.cells(hpf20, 3), file = "20SHPCells.txt", sep = "\t")
write.table(which.cells(hpf20, 6), file = "20FHPCells.txt", sep = "\t")
write.table(which.cells(hpf20, 7), file = "20ASM2Cells.txt", sep = "\t")
```

```
# Rename cluster identities
```

```
hpf20 = rename.ident(hpf20, 2, "20ASM1")
hpf20 = rename.ident(hpf20, 3, "20SHP")
hpf20 = rename.ident(hpf20, 6, "20FHP")
hpf20 = rename.ident(hpf20, 7, "20ASM2")
```

```
# Visualize tSNE used color scheme FHP-red, SHP-orange, ASM-blue
```

```
tsne.plot(hpf20, do.label = T, label.pt.size = 1, label.cex.text = 1.2, label.cols.use =
  "blue", "red", "orange"))
```



```
# Store FHP markers in text file
FHP_20.markers = cl6_20.markers
head(FHP_20.markers[order(FHP_20.markers$myAUC, decreasing = T), ], 20)
```

##	myAUC	avg_diff	power	pct.1
## Ci-R1CiGC27a04	0.962	3.284688	0.924	0.954
## KH2013:KH.C4.547_BMP2/4	0.951	2.195473	0.902	0.985
## KH2013:KH.C5.227_F56C4.4	0.944	1.991633	0.888	1.000
## KH2013:KH.C11.378_CLDN1/10/14/19/2/3/4/5/6/7/9	0.937	1.539702	0.874	1.000
## KH2013:KH.C1.479_NEB	0.927	1.680541	0.854	1.000
## MMP21	0.927	3.901756	0.854	0.877
## FRAS1	0.911	2.683945	0.822	0.831
## SFRP1/5	0.910	2.269762	0.820	0.923
## KH2013:KH.C10.203	0.909	2.466237	0.818	0.923
## COL13A1	0.907	3.395970	0.814	0.846
## KH2013:KH.C1.953_CG32702	0.906	2.177912	0.812	0.938
## KH2013:KH.C7.205_ASB2	0.904	1.818015	0.808	0.969
## NTN1/2/3/5	0.902	2.350696	0.804	0.892
## KH2013:KH.C2.994_RNF149	0.900	1.278227	0.800	1.000
## KH2013:KH.C3.665_ZAN	0.896	1.455140	0.792	0.985
## MTDH	0.895	1.022437	0.790	1.000
## C5ORF48	0.894	2.399851	0.788	0.892
## GATA4/5/6	0.891	1.513036	0.782	0.985

```

## KH2013:KH.C3.191_WBSCR27          0.886 1.950294 0.772 0.938
## KH2013:KH.C1.738_CG9550          0.881 1.235709 0.762 0.985
##                                     pct.2
## Ci-R1CiGC27a04                    0.238
## KH2013:KH.C4.547_BMP2/4          0.381
## KH2013:KH.C5.227_F56C4.4        0.973
## KH2013:KH.C11.378_CLDN1/10/14/19/2/3/4/5/6/7/9 0.946
## KH2013:KH.C1.479_NEB             0.605
## MMP21                              0.117
## FRAS1                              0.117
## SFRP1/5                            0.193
## KH2013:KH.C10.203                0.247
## COL13A1                            0.108
## KH2013:KH.C1.953_CG32702        0.309
## KH2013:KH.C7.205_ASB2           0.430
## NTN1/2/3/5                       0.166
## KH2013:KH.C2.994_RNF149         0.857
## KH2013:KH.C3.665_ZAN            0.628
## MTDH                              0.991
## C5ORF48                           0.229
## GATA4/5/6                         0.471
## KH2013:KH.C3.191_WBSCR27        0.435
## KH2013:KH.C1.738_CG9550        0.897

```

```

write.table(FHP_20.markers, file = "FHP_20.markers.txt", sep = "\t")

# Store SHP markers in text file
SHP_20.markers = cl3_20.markers
head(SHP_20.markers[order(SHP_20.markers$myAUC, decreasing = T), ], 20)

```

```

##                                     myAUC  avg_diff power pct.1 pct.2
## KH2013:KH.C1.638_C170RF105 0.842  1.727786 0.684 0.861 0.325
## KH2013:KH.C4.125_BMP2/4    0.841  1.652993 0.682 0.937 0.517
## DACH1/2                    0.788  1.930722 0.576 0.734 0.278
## KH2013:KH.C4.226           0.787  1.372513 0.574 0.911 0.574
## KH2013:KH.C2.209_CALM1/3  0.775  1.004900 0.550 0.924 0.646
## FOXF1/2                    0.767  1.249064 0.534 0.873 0.512
## YKR070W                    0.750  1.143960 0.500 0.899 0.699
## KH2013:KH.L153.109        0.745  2.030674 0.490 0.595 0.124
## SNX11                      0.739  1.187810 0.478 0.747 0.297
## KH2013:KH.S1887.1         0.716  1.458843 0.432 0.620 0.254
## KH2013:KH.C11.667        0.714  1.038638 0.428 0.886 0.727
## PCTP                      0.704  1.024480 0.408 0.709 0.368
## MATN1/3/4                 0.695  1.452748 0.390 0.633 0.368

```

```
## HS3ST3L          0.685  1.075695 0.370 0.671 0.378
## TBX1/10         0.681  1.420956 0.362 0.633 0.373
## KANK1/2/3/4    0.640  1.073334 0.280 0.595 0.411
## KH2013:KH.C9.40_GST01/2 0.615  1.051532 0.230 0.506 0.335
## PIGT           0.502 -5.064610 0.004 0.570 0.569
## RBM15B         0.433 -3.685868 0.134 0.646 0.789
## PSMD14         0.430 -3.980281 0.140 0.835 0.928
```

```
write.table(SHP_20.markers, file = "SHP_20.markers.txt", sep = "\t")
```

```
# Store ASM1 markers in text file
```

```
ASM1_20.markers = c12_20.markers
```

```
head(ASM1_20.markers[order(ASM1_20.markers$myAUC, decreasing = T), ], 20)
```

```
##          myAUC avg_diff power pct.1 pct.2
## KH2013:KH.C11.139 0.930 2.042404 0.860 0.992 0.337
## KH2013:KH.L25.4 0.918 1.888086 0.836 0.992 0.231
## KH2013:KH.C5.62_ODF3L2 0.899 1.562319 0.798 0.983 0.278
## KH2013:KH.L18.10_THSD7A 0.894 1.571177 0.788 0.983 0.331
## SCP1 0.883 1.728818 0.766 0.958 0.278
## KH2013:KH.C4.506_HMCN1 0.882 1.291218 0.764 1.000 0.858
## KH2013:KH.S1269.1 0.880 1.388814 0.760 0.992 0.533
## SLC35G2/6 0.875 1.853114 0.750 0.899 0.225
## UBE2QL1 0.874 1.422110 0.748 0.966 0.432
## KH2013:KH.C6.201 0.870 1.898804 0.740 0.874 0.183
## MMP14/15/16/24 0.868 1.387104 0.736 0.975 0.355
## ARHGAP22/24/25 0.868 1.743081 0.736 0.891 0.260
## FRS2/3 0.866 1.408759 0.732 0.975 0.609
## EBF1/2/3/4 0.865 1.253061 0.730 1.000 0.314
## DCDC2B 0.863 1.184517 0.726 0.983 0.325
## BDH1 0.862 1.240573 0.724 0.983 0.396
## NOVA1/2 0.856 1.161588 0.712 1.000 0.462
## Ci-R1CiGC09b24; 0.853 1.216660 0.706 0.983 0.888
## TIMP1/2/3/4 0.852 1.920971 0.704 0.882 0.349
## HLH-4 0.845 1.462400 0.690 0.950 0.544
```

```
write.table(ASM1_20.markers, file = "ASM1_20.markers.txt", sep = "\t")
```

```
# Store ASM2 markers in text file
```

```
ASM2_20.markers = c17_20.markers
```

```
head(ASM2_20.markers[order(ASM2_20.markers$myAUC, decreasing = T), ], 20)
```

```
##          myAUC avg_diff power pct.1 pct.2
```

```

## MYF5 0.936 2.237196 0.872 1.00 0.403
## KH2013:KH.C8.649_ACTA1/2 0.927 2.296572 0.854 1.00 0.589
## DCDC2B 0.914 1.380557 0.828 1.00 0.559
## MYO-1/2/3/5 0.914 2.429771 0.828 0.92 0.278
## CALML6 0.905 2.418975 0.810 0.96 0.304
## FUN26 0.905 2.611697 0.810 0.88 0.106
## RBM24/38 0.902 1.965112 0.804 0.96 0.384
## SMYD1 0.902 1.604893 0.804 1.00 0.802
## MYO10 0.899 1.617349 0.798 1.00 0.441
## Ci-Actin; 0.894 2.092404 0.788 0.96 0.738
## KH2013:KH.C4.506_HMCN1 0.887 1.052516 0.774 1.00 0.909
## KH2013:KH.C5.522_SLC6A13 0.887 1.453291 0.774 1.00 0.471
## ZKSCAN1/14/17/2/3/6 0.885 3.396235 0.770 0.80 0.080
## KH2013:KH.C12.669_PHEX 0.883 1.820194 0.766 0.96 0.407
## KH2013:KH.C7.476_CHRNB1 0.880 1.952671 0.760 0.92 0.304
## KH2013:KH.C12.419_TSPAN4/9 0.878 1.252526 0.756 1.00 0.821
## KH2013:KH.C1.509 0.876 2.059997 0.752 0.88 0.205
## KH2013:KH.S643.6_CDKN1B 0.876 1.562963 0.752 1.00 0.612
## RBFox1/2/3 0.875 1.347928 0.750 1.00 0.863
## KH2013:KH.C5.404_MBL1/2 0.875 2.442091 0.750 0.84 0.114

```

```

write.table(ASM2_20.markers, file = "ASM2_20.markers.txt", sep = "\t")

# Find pan Heart Progenitor markers
panHP_20.markers = find.markers(hpf20, c("20SHP", "20FHP"), thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
head(panHP_20.markers[order(panHP_20.markers$myAUC, decreasing = T), ], 20)

```

```

## myAUC avg_diff power pct.1
## KH2013:KH.C2.994_RNF149 0.977 2.285073 0.954 0.993
## SLIT1/2/3 0.961 3.358580 0.922 0.965
## NKX2-3 0.959 3.261362 0.918 0.958
## KH2013:KH.C1.738_CG9550 0.953 1.858233 0.906 0.993
## KH2013:KH.C3.665_ZAN 0.951 3.369149 0.902 0.951
## GATA4/5/6 0.951 3.224164 0.902 0.944
## QKI 0.933 1.992776 0.866 0.986
## KH2013:KH.C10.174 0.927 2.971638 0.854 0.917
## KH2013:KH.C2.514_ENPP1/2/3 0.927 1.030617 0.854 1.000
## LRP4/8 0.915 2.558266 0.830 0.889
## UNC5A 0.913 1.610891 0.826 1.000
## KH2013:KH.C9.650_SULT1ST1/2/3/4 0.910 1.510445 0.820 0.993
## KH2013:KH.C2.209_CALM1/3 0.903 2.226478 0.806 0.944
## KH2013:KH.C4.547_BMP2/4 0.884 3.020256 0.768 0.840
## CG15828 0.882 1.072115 0.764 1.000

```

```

## KH2013:KH.C11.378_CLDN1/10/14/19/2/3/4/5/6/7/9 0.880 1.799816 0.760 0.972
## NAV1 0.879 2.707348 0.758 0.799
## KH2013:KH.C2.935_S1PR1/2/3/4/5 0.875 1.985909 0.750 0.910
## KH2013:KH.C3.716_EFNA1/2/3/4/5 0.872 2.300622 0.744 0.868
## FOXG1 0.872 2.046786 0.744 0.889
## pct.2
## KH2013:KH.C2.994_RNF149 0.785
## SLIT1/2/3 0.229
## NKX2-3 0.146
## KH2013:KH.C1.738_CG9550 0.840
## KH2013:KH.C3.665_ZAN 0.465
## GATA4/5/6 0.229
## QKI 0.771
## KH2013:KH.C10.174 0.243
## KH2013:KH.C2.514_ENPP1/2/3 1.000
## LRP4/8 0.111
## UNC5A 0.757
## KH2013:KH.C9.650_SULT1ST1/2/3/4 0.861
## KH2013:KH.C2.209_CALM1/3 0.500
## KH2013:KH.C4.547_BMP2/4 0.194
## CG15828 0.972
## KH2013:KH.C11.378_CLDN1/10/14/19/2/3/4/5/6/7/9 0.944
## NAV1 0.118
## KH2013:KH.C2.935_S1PR1/2/3/4/5 0.410
## KH2013:KH.C3.716_EFNA1/2/3/4/5 0.285
## FOXG1 0.312

```

```

write.table(panHP_20.markers, file = "panHP_20.markers.txt", sep = "\t")

# Find SHP specific markers that distinguish tow heart progenitors FHP and
# SHP
SHPspecific_20.markers = find.markers(hpf20, "20SHP", "20FHP", thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
head(SHPspecific_20.markers[order(SHPspecific_20.markers$myAUC, decreasing = T),
  ], 20)

```

```

## myAUC avg_diff power pct.1 pct.2
## KH2013:KH.C1.638_C17ORF105 0.861 2.080418 0.722 0.861 0.246
## DACH1/2 0.788 1.960855 0.576 0.734 0.292
## MATN1/3/4 0.786 3.205297 0.572 0.633 0.138
## PPAP2B 0.784 1.536951 0.568 0.886 0.585
## KH2013:KH.C10.172 0.724 1.018103 0.448 0.759 0.369
## TIMP4 0.695 1.525484 0.390 0.608 0.277
## SPINK1/2 0.689 1.225729 0.378 0.570 0.215

```

```

## KH2013:KH.L154.26_CSMD2          0.686 1.969010 0.372 0.557 0.277
## MIG3                             0.684 1.056078 0.368 0.570 0.231
## KANK1/2/3/4                      0.684 1.139440 0.368 0.595 0.262
## NVL                               0.680 1.009940 0.360 0.747 0.554
## KH2013:KH.L153.109              0.680 1.170249 0.360 0.595 0.246
## C12ORF52                         0.679 1.332137 0.358 0.557 0.292
## KH2013:KH.S1269.1               0.672 1.295684 0.344 0.582 0.292
## KH2013:KH.S1140.1_GPM6B         0.670 1.050835 0.340 0.671 0.415
## TBX1/10                          0.661 1.134796 0.322 0.633 0.385
## GPR89B                           0.656 1.013249 0.312 0.519 0.231
## KH2013:KH.S404.8_HSP-16.1/11/2/41/48/49 0.654 1.208633 0.308 0.747 0.523
## MXRA7                            0.653 1.006081 0.306 0.759 0.600
## COPS2                            0.646 1.083980 0.292 0.519 0.292

```

```
write.table(SHPspecific_20.markers, file = "SHPspecific_20.markers.txt", sep = "\t")
```

```
# Find FHP specific markers that distinguish two heart progenitors FHP and
# SHP
```

```
FHPspecific_20.markers = find.markers(hpf20, "20FHP", "20SHP", thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
```

```
head(FHPspecific_20.markers[order(FHPspecific_20.markers$myAUC, decreasing = T),
  ], 20)
```

```

##                               myAUC avg_diff power pct.1 pct.2
## Ci-R1CiGC27a04              0.945 2.550143 0.890 0.954 0.443
## KH2013:KH.C5.227_F56C4.4    0.922 1.583096 0.844 1.000 0.962
## MMP21                       0.920 3.614448 0.840 0.877 0.203
## COL13A1                     0.891 2.810263 0.782 0.846 0.190
## FRAS1                       0.887 2.287808 0.774 0.831 0.190
## KH2013:KH.C4.547_BMP2/4    0.887 1.342081 0.774 0.985 0.722
## KH2013:KH.C10.203          0.857 1.758780 0.714 0.923 0.418
## SMYD1                       0.848 1.757263 0.696 0.923 0.506
## SMTN                        0.838 1.238689 0.676 0.985 0.848
## BX005256.5                 0.837 1.667243 0.674 0.938 0.519
## C5ORF48                     0.836 1.655992 0.672 0.892 0.418
## SFRP1/5                     0.835 1.369541 0.670 0.923 0.405
## NTN1/2/3/5                 0.829 1.383314 0.658 0.892 0.380
## KH2013:KH.C1.1083_EPHA10/2/3/4/5/6/7/8 0.827 2.507890 0.654 0.708 0.089
## LAMA1/2                     0.825 1.628374 0.650 0.738 0.165
## ISL1/2                     0.819 1.549760 0.638 0.846 0.342
## FOXP1/2/4                  0.814 2.022667 0.628 0.769 0.203
## KH2013:KH.C1.953_CG32702    0.807 1.211560 0.614 0.938 0.646
## KH2013:KH.C12.244          0.801 1.641349 0.602 0.785 0.266
## KH2013:KH.C3.191_WBSCR27    0.796 1.211239 0.592 0.938 0.620

```



```

write.table(FHPspecific_20.markers, file = "FHPspecific_20.markers.txt", sep = "\t")

# Find ASM1 specific markers that distinguish two heart progenitors ASM1 and
# ASM2
ASM1specific_20.markers = find.markers(hpf20, "20ASM1", "20ASM2", thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
head(ASM1specific_20.markers[order(ASM1specific_20.markers$myAUC, decreasing = T),
  ], 20)

```

##	myAUC	avg_diff	power	pct.1	pct.2
## GPX7/8	0.765	1.262169	0.530	0.882	0.64
## SFXN2	0.739	1.422001	0.478	0.739	0.44
## LATS1/2	0.737	1.317006	0.474	0.697	0.32
## UNC5A	0.728	1.247983	0.456	0.815	0.48
## KH2013:KH.C10.340_DHRS11	0.715	1.095224	0.430	0.672	0.28
## FZD5/8	0.705	1.552283	0.410	0.555	0.20
## KH2013:KH.S2635.1_FNBP1	0.703	1.304549	0.406	0.681	0.48
## KH2013:KH.C2.646	0.699	1.046240	0.398	0.765	0.60
## RASSF10A	0.697	1.149363	0.394	0.521	0.16
## VASN	0.684	1.168025	0.368	0.504	0.12
## Ci-tune	0.678	1.178515	0.356	0.689	0.40
## MRRF	0.677	1.570648	0.354	0.580	0.32
## CD2BP2	0.674	1.258361	0.348	0.706	0.60
## KH2013:KH.C1.29 EIF4EBP1	0.673	1.261134	0.346	0.639	0.40
## RGS17/18/19/20	0.655	1.096347	0.310	0.571	0.32
## ARRDC2/3	0.654	1.148108	0.308	0.538	0.28
## HAND/1/2	0.645	1.345731	0.290	0.521	0.36
## KH2013:KH.L124.2	0.645	2.411661	0.290	0.966	1.00
## CHCHD7	0.645	1.266356	0.290	0.563	0.32
## KH2013:KH.C2.209_CALM1/3	0.636	1.193868	0.272	0.538	0.32

```

write.table(ASM1specific_20.markers, file = "ASM1specific_20.markers.txt", sep = "\t")

# Find ASM2 specific markers that distinguish two heart progenitors ASM2 and
# ASM1
ASM2specific_20.markers = find.markers(hpf20, "20ASM2", "20ASM1", thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
head(ASM2specific_20.markers[order(ASM2specific_20.markers$myAUC, decreasing = T),
  ], 20)

```

##	myAUC	avg_diff	power	pct.1	pct.2
## KH2013:KH.C8.649_ACTA1/2	0.886	1.832513	0.772	1.00	0.706
## MYF5	0.873	1.562098	0.746	1.00	0.697


```

## SMYD1                0.873 1.325876 0.746 1.00 0.933
## ZKSCAN1/14/17/2/3/6 0.869 2.751014 0.738 0.80 0.151
## FUN26                0.864 1.904252 0.728 0.88 0.235
## Ci-Actin;           0.861 1.712992 0.722 0.96 0.782
## MYO-1/2/3/5         0.845 1.814388 0.690 0.92 0.563
## DYRK2/3             0.842 2.433072 0.684 0.84 0.252
## KH2013:KH.C12.669_PHEX 0.839 1.439534 0.678 0.96 0.597
## KH2013:KH.C7.476_CHRNB1 0.830 1.470585 0.660 0.92 0.454
## KH2013:KH.C5.404_MBL1/2 0.826 1.706676 0.652 0.84 0.235
## CALML6              0.820 1.671514 0.640 0.96 0.605
## KH2013:KH.S489.1_ARF1/3 0.819 1.012200 0.638 1.00 0.765
## CCDC66              0.818 1.216917 0.636 0.96 0.647
## RBM24/38            0.817 1.232162 0.634 0.96 0.748
## KH2013:KH.S643.6_CDKN1B 0.809 1.113191 0.618 1.00 0.815
## MYBL1/2             0.803 1.214419 0.606 0.92 0.437
## KH2013:KH.C1.509   0.799 1.285231 0.598 0.88 0.445
## LGALS4/8/9          0.797 1.557405 0.594 0.80 0.445
## NLRC3               0.790 1.435796 0.580 0.60 0.160

```

```
write.table(ASM2specific_20.markers, file = "ASM2specific_20.markers.txt", sep = "\t")
```

```
# Find ASM markers
```

```

ASM_20.markers = find.markers(hpf20, c("20ASM1", "20ASM2"), "20FHP", thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
head(ASM_20.markers[order(ASM_20.markers$myAUC, decreasing = T), ], 20)

```

```

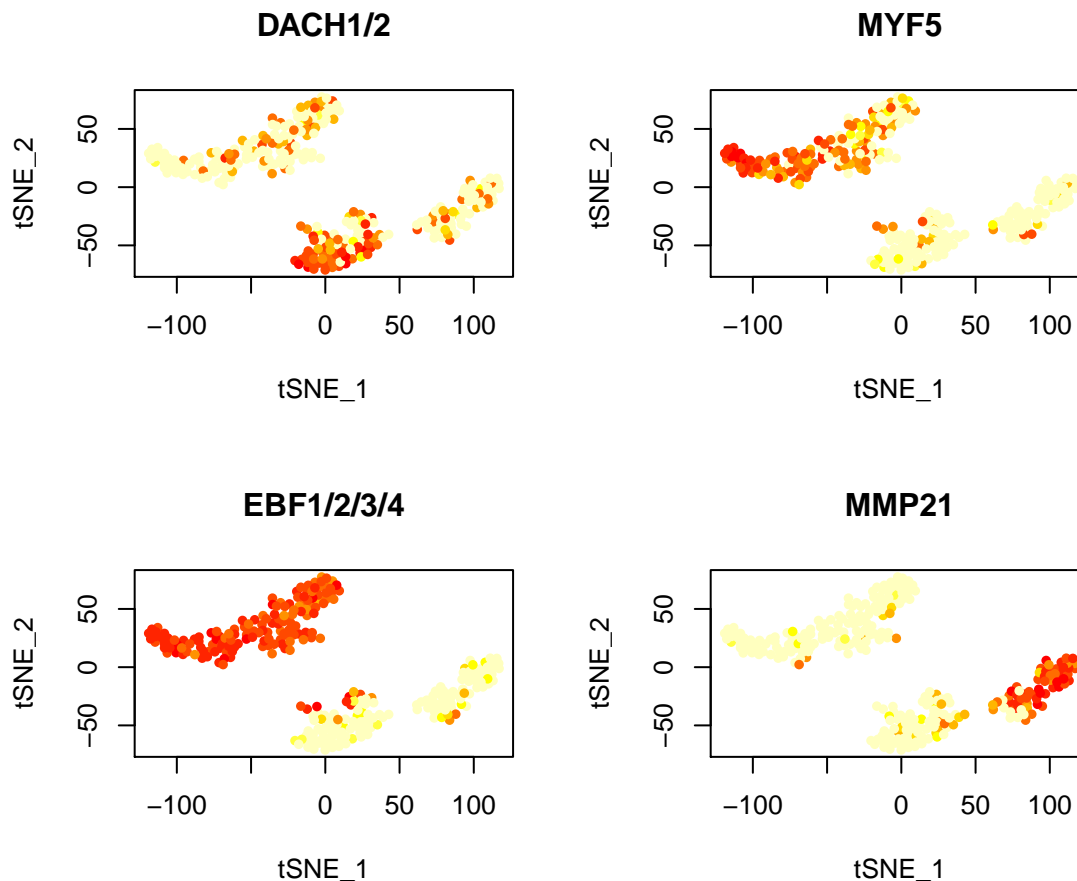
##                myAUC avg_diff power pct.1 pct.2
## EBF1/2/3/4     0.999 4.550177 0.998 1.000 0.154
## KH2013:KH.C4.506_HMCN1 0.997 3.159464 0.994 1.000 0.785
## KH2013:KH.C11.139 0.994 4.919472 0.988 0.993 0.231
## KH2013:KH.L25.4 0.994 4.655595 0.988 0.993 0.062
## DCDC2B         0.989 4.127543 0.978 0.986 0.154
## KH2013:KH.C5.62_ODF3L2 0.988 4.192618 0.976 0.986 0.108
## KH2013:KH.S1269.1 0.983 3.046282 0.966 0.993 0.292
## BDH1           0.981 3.236993 0.962 0.986 0.215
## NOVA1/2        0.980 2.940057 0.960 1.000 0.215
## SCP1           0.978 4.539122 0.956 0.958 0.046
## KH2013:KH.C8.892_TUB1/3 0.967 2.799581 0.934 0.993 0.508
## KH2013:KH.C5.522_SLC6A13 0.962 3.947985 0.924 0.931 0.092
## UBE2QL1        0.957 2.591103 0.914 0.972 0.369
## KH2013:KH.L18.10_THSD7A 0.956 2.568935 0.912 0.972 0.231
## ELK1/3/4       0.949 3.539293 0.898 0.917 0.108
## PCBP3          0.948 2.392518 0.896 0.965 0.169
## SLC35G2/6      0.942 4.200869 0.884 0.903 0.108

```

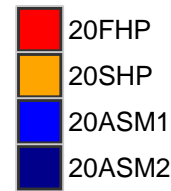
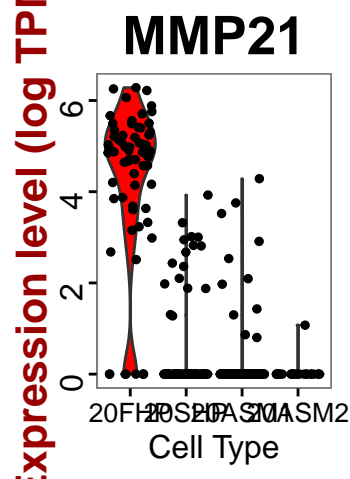
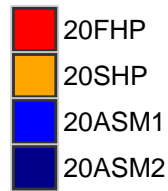
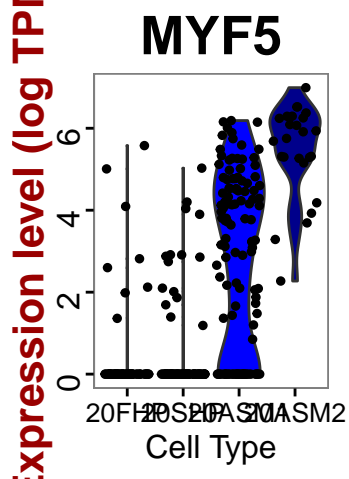
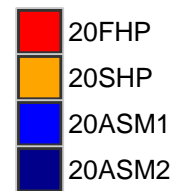
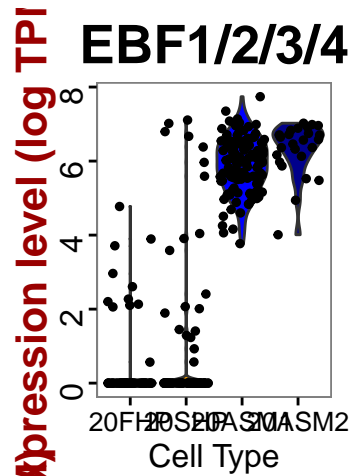
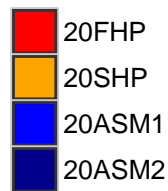
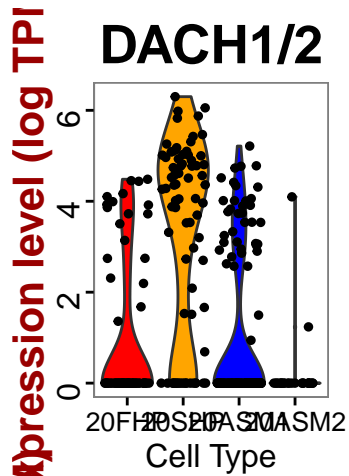
```
## KH2013:KH.L154.26_CSMD2 0.942 3.077734 0.884 0.938 0.277
## TIMP1/2/3/4 0.941 4.461382 0.882 0.903 0.123
## MMP14/15/16/24 0.940 2.208247 0.880 0.979 0.308
```

```
# ASM_20.markers=find.markers(hpf20,c('20ASM1','20ASM2'),thresh.use =
# 1,test.use = 'roc',min.pct = 0.5)
# head(ASM_20.markers[order(ASM_20.markers$myAUC,decreasing = T),],20)
write.table(ASM_20.markers, file = "ASM_20.markers.txt", sep = "\t")

# Visualize markers of different clusters using violin plot and feature plot
hpf20@ident = factor(hpf20@ident, ordered = T, levels = c("20FHP", "20SHP",
"20ASM1", "20ASM2"))
genes.viz.20 = c("DACH1/2", "MYF5", "EBF1/2/3/4", "MMP21")
feature.plot(hpf20, genes.viz.20, pt.size = 0.8)
```



```
vlnPlot(hpf20, genes.viz.20, cols.use = c("red", "orange", "blue", "blue4"))
```



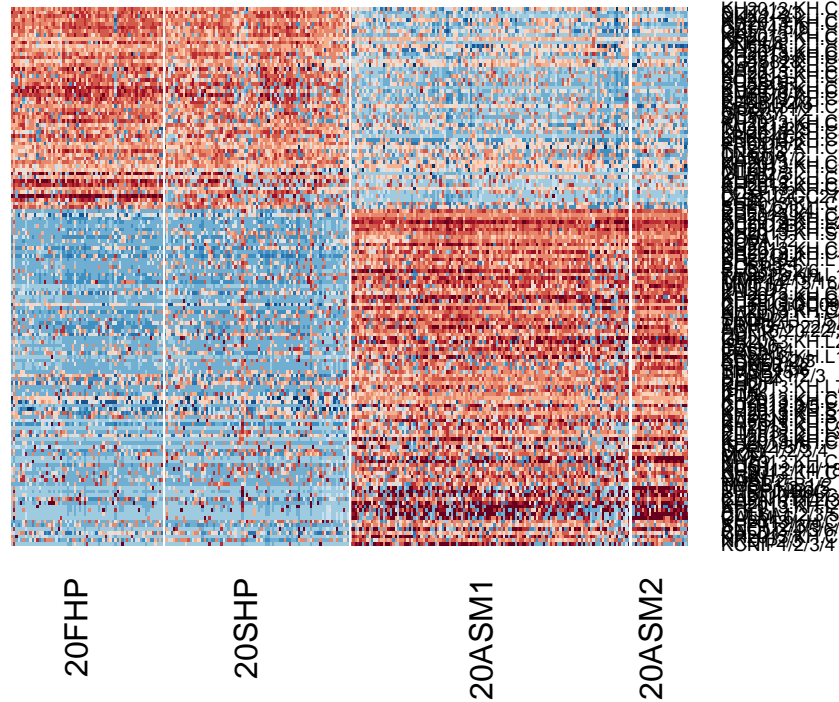
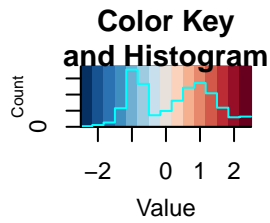
```

# Select markers for plotting on a Heatmap (top 10 positive markers with
# high classification power(myAUC))
marker20FHP = rownames(FHP_20.markers[order(FHP_20.markers$myAUC, decreasing = T)[1:15],
])
marker20SHP = rownames(SHP_20.markers[order(SHP_20.markers$myAUC, decreasing = T)[1:15],
])
marker20ASM1 = rownames(ASM1_20.markers[order(ASM1_20.markers$myAUC, decreasing = T)[1:15],
])
marker20ASM2 = rownames(ASM2_20.markers[order(ASM2_20.markers$myAUC, decreasing = T)[1:15],
])
marker.20 = c(marker20FHP, marker20SHP, marker20ASM1, marker20ASM2)

panHeart.select = rownames(subset(panHP_20.markers, power > 0.6 & avg_diff >
0))
asm.select = rownames(subset(ASM_20.markers, power > 0.6 & avg_diff > 0))

# Draw a heatmap of all cells for these marker genes
doHeatMap(hpf20, genes.use = marker.20, remove.key = F, slim.col.label = T,
draw.line = T, cex.col = 1.2, col.use = col)

```

5. hpf18

```
# Subset data from preprocessed Seurat object
hpf18 = subsetData(hpfall.remv2, which.cells(hpfall.remv2, "hpf18"), do.center = F,
  do.scale = F)
hpf18
```

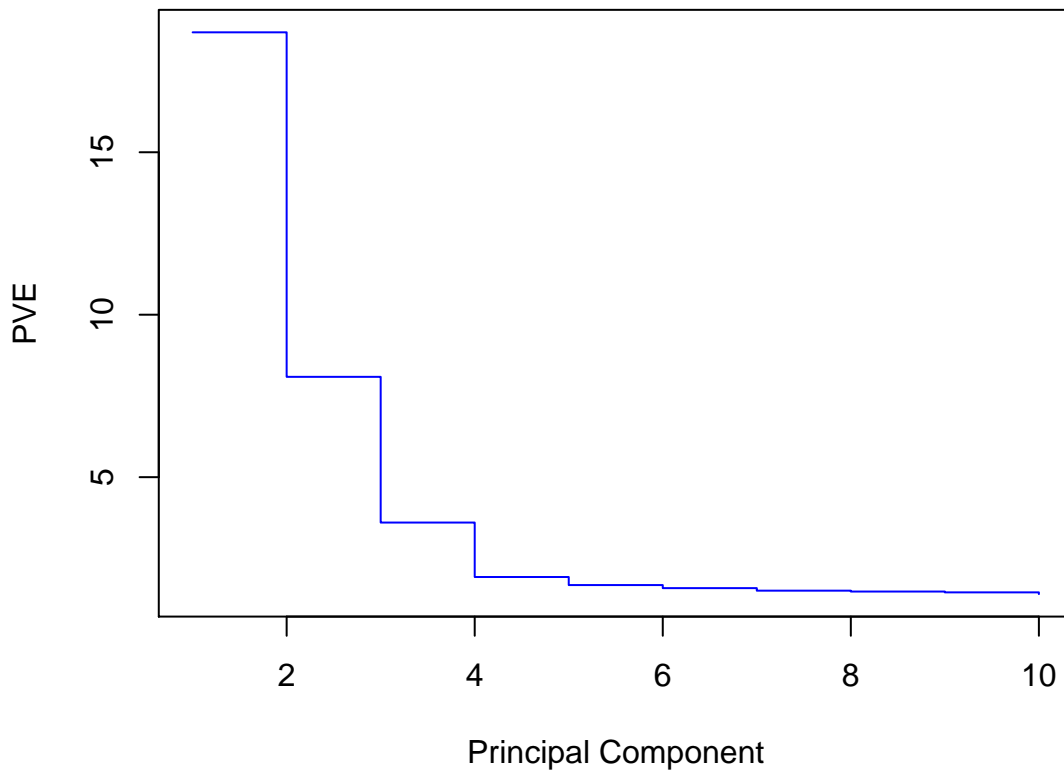
```
## An object of class seurat in project allhpf
## 14864 genes across 177 samples.
```

```
# Based on hpf12 hpf14 and hpf20 data, we have successfully uncovered all
# the TVC lineage types: TVC, STVC, FHP, SHP, ASM Based on preliminary
# studies, hpf18 contains three TVC lineage cell types: FHP, SHP and ASM
# Therefore we can run PCA using 20hpf cell markers (20FHP, 20SHP, 20ASM)
# with power > 0.5 and positive expressions
marker20FHP.use = rownames(subset(FHP_20.markers, power > 0.5 & avg_diff > 0))
marker20SHP.use = rownames(subset(SHP_20.markers, power > 0.5 & avg_diff > 0))
marker20ASM1.use = rownames(subset(ASM1_20.markers, power > 0.5 & avg_diff >
```

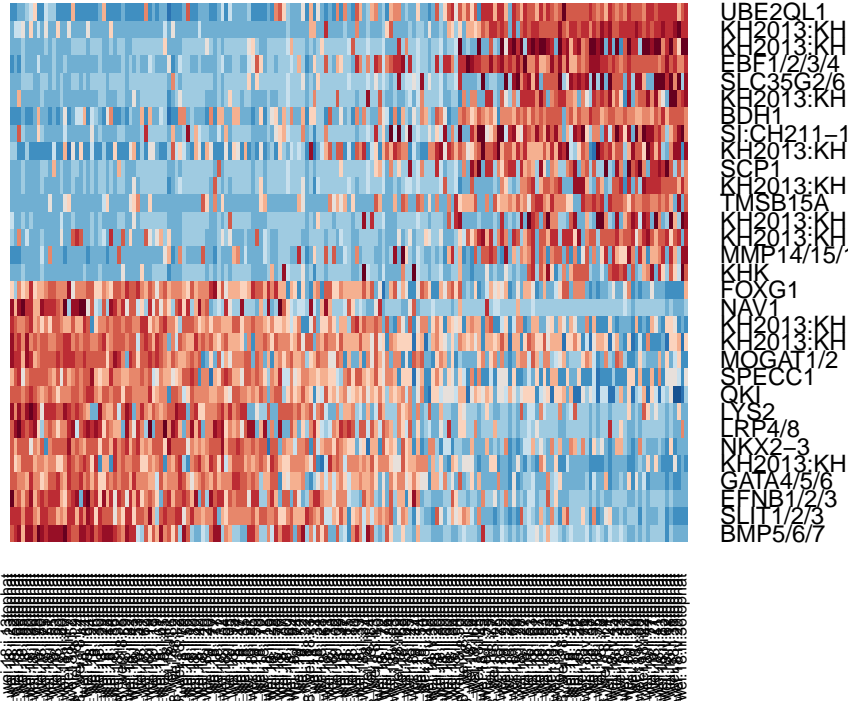
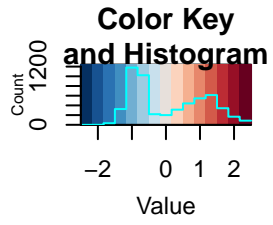
```
0))
marker20ASM2.use = rownames(subset(ASM2_20.markers, power > 0.5 & avg_diff >
0))
marker.20.use = unique(c(marker20FHP.use, marker20SHP.use, marker20ASM1.use,
marker20ASM2.use))
length(marker.20.use)
```

```
## [1] 200
```

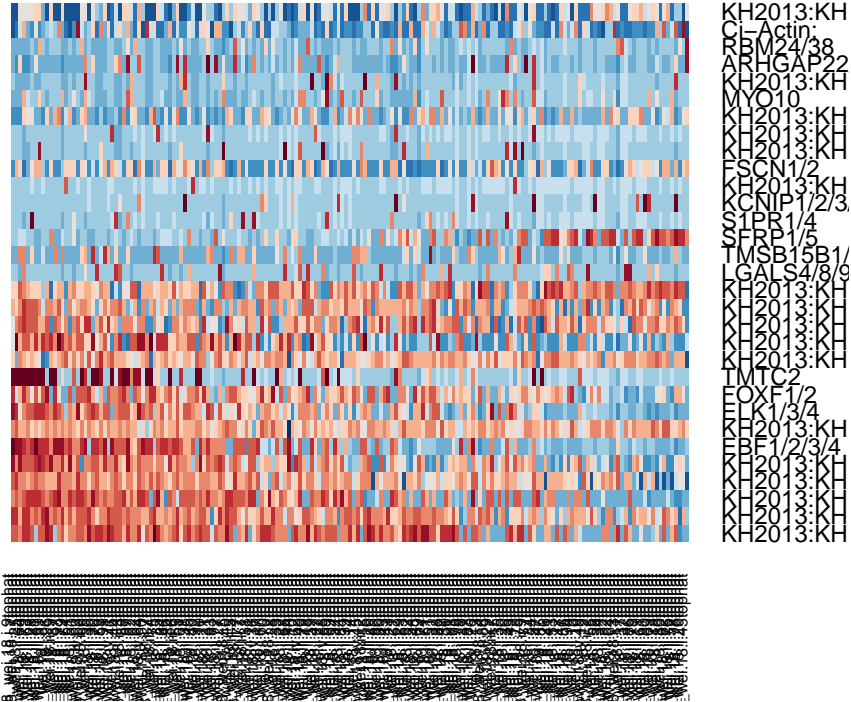
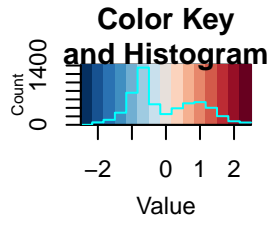
```
hpf18 = pca(hpf18, pc.genes = marker.20.use, do.print = F)
pcScree(hpf18, marker.20.use, 10)
```



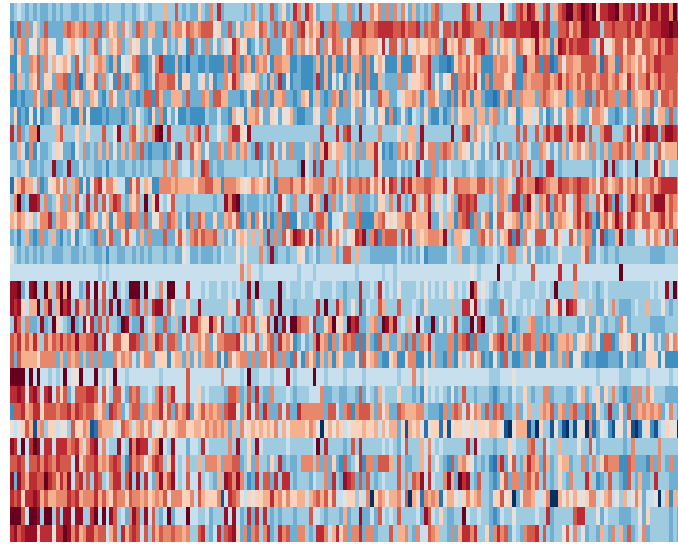
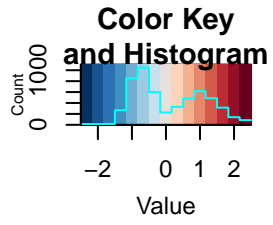
```
pcHeatmap(hpf18, 1, do.balanced = T, col.use = col)
```



```
pcHeatmap(hpf18, 2, do.balanced = T, col.use = col)
```



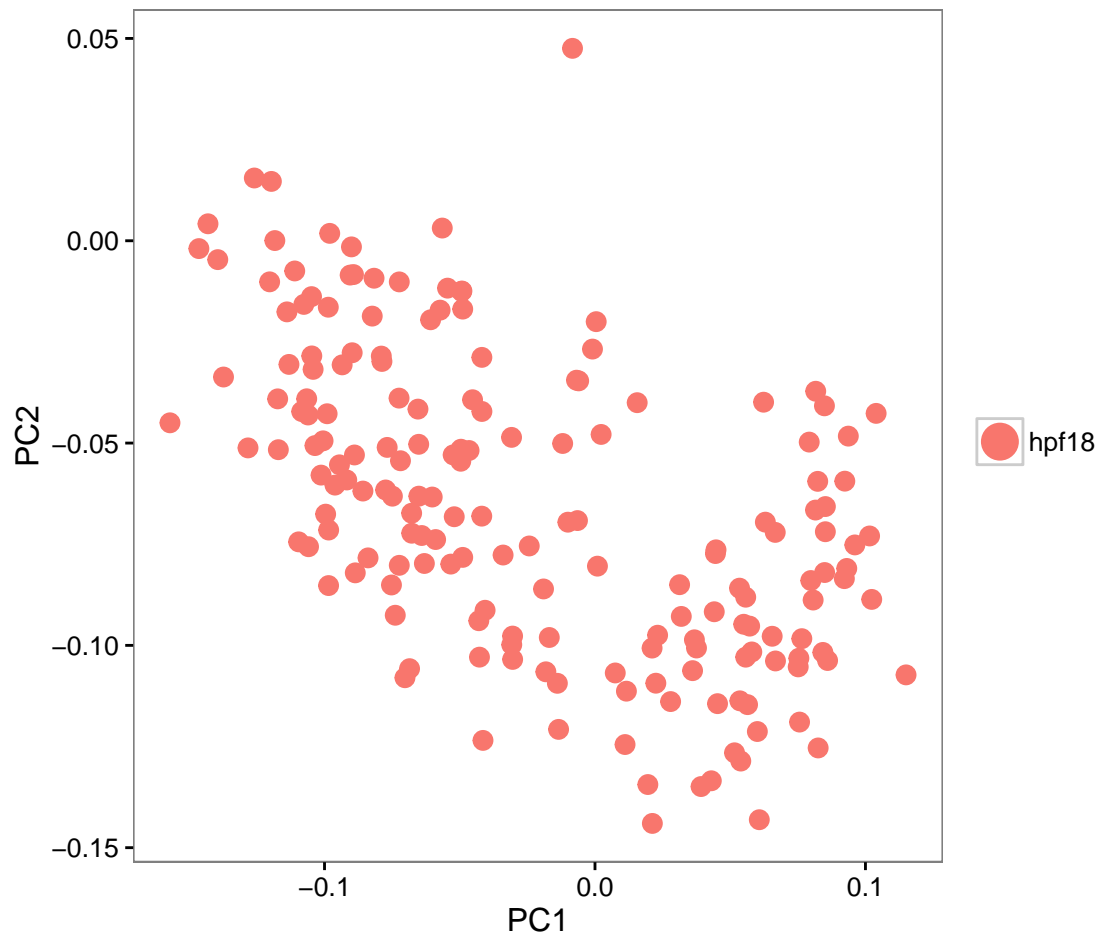
```
pcHeatmap(hpf18, 3, do.balanced = T, col.use = col)
```

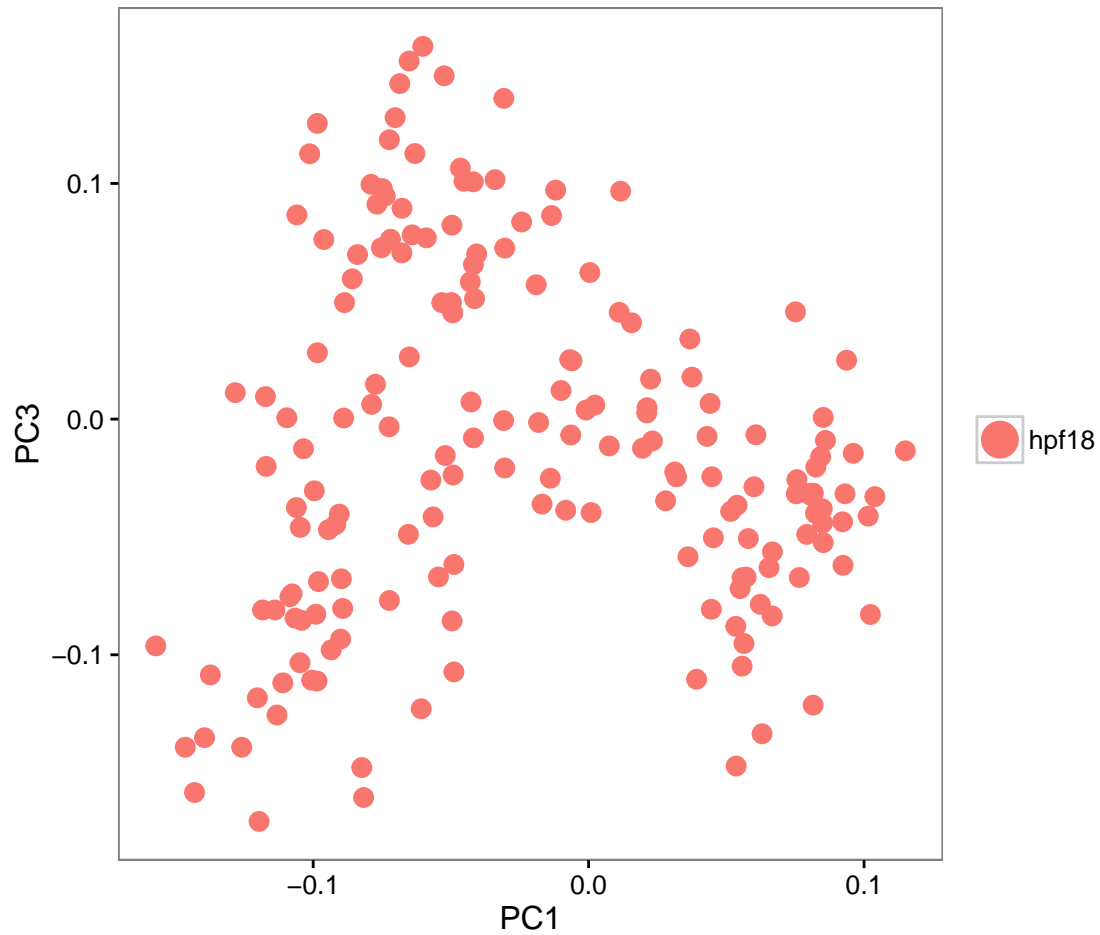
DACH1/2
 KL2013:KH
 KO2F1/2
 SOX1
 SCYL
 XH2013:KH
 XH2013:KH
 XH2013:KH
 KH2013:KH
 CDC25A
 GFI
 KH2013:KH
 RP4/8
 ITI
 OXG1
 ITI
 K1/3/4
 XBM24/38
 KH2013:KH
 NTN1/2/3/5
 NAV1
 S:CH211-1
 MOGAT1/2
 SEPT12/3/9
 MYLK2/3/4
 SFRP1/5
 KL2013:KH
 KL2013:KH
 COL13A1
 XH2013:KH
 C5ORF48
 KH2013:KH
 Ci-R1CiGC
 KH2013:KH



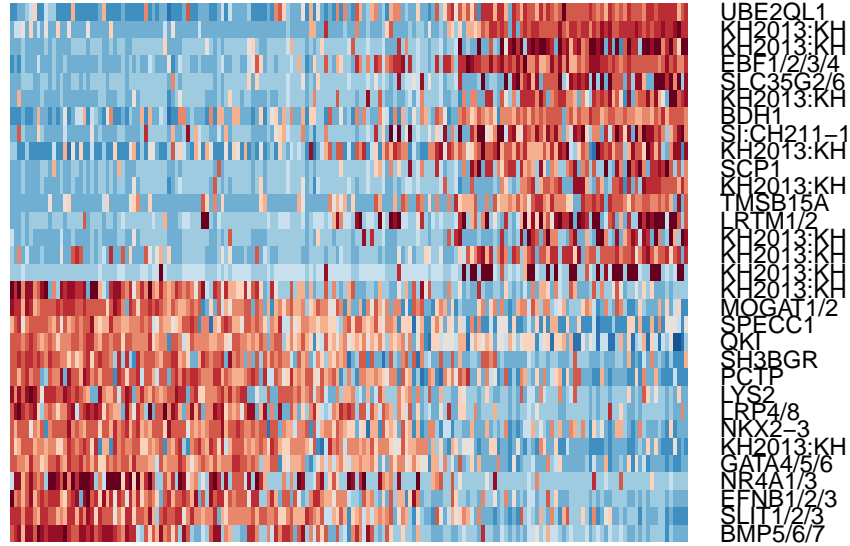
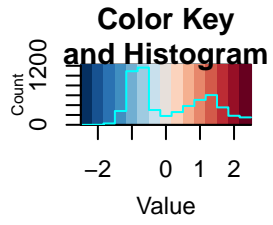
```
pca.plot(hpf18, 1, 2)
```



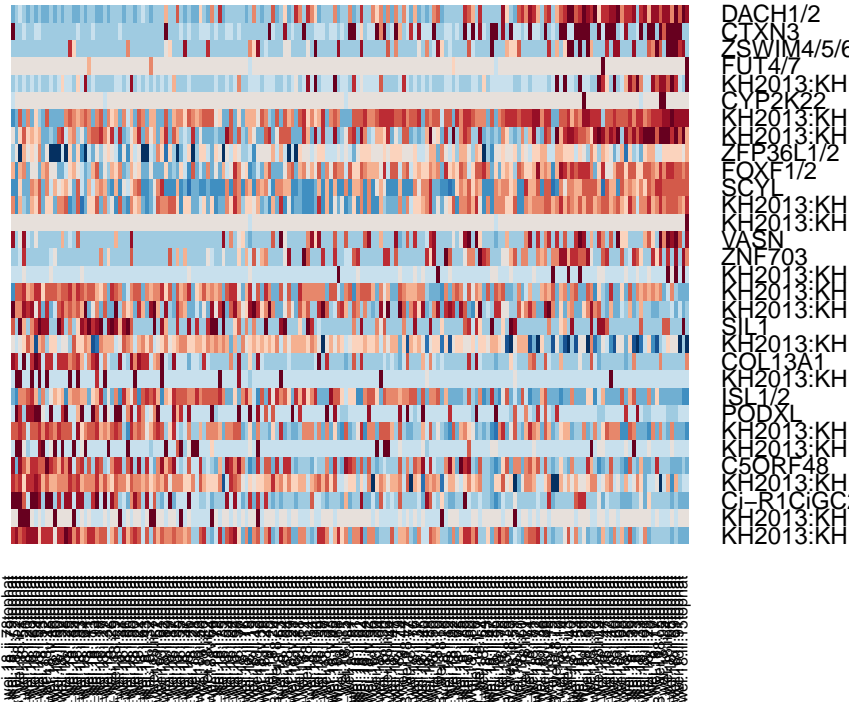
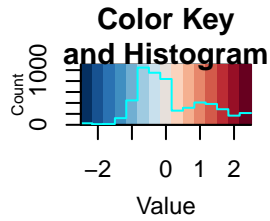
```
pca.plot(hpf18, 1, 3)
```



```
# Calculate PCA scores for all genes (PCA projection)  
hpf18 = project.pca(hpf18, do.print = F)  
  
# Visualize the full projected PCA, which now includes new genes which were  
# not previously (use.full=TRUE)  
pcHeatmap(hpf18, 1, use.full = T, do.balanced = T, col.use = col)
```

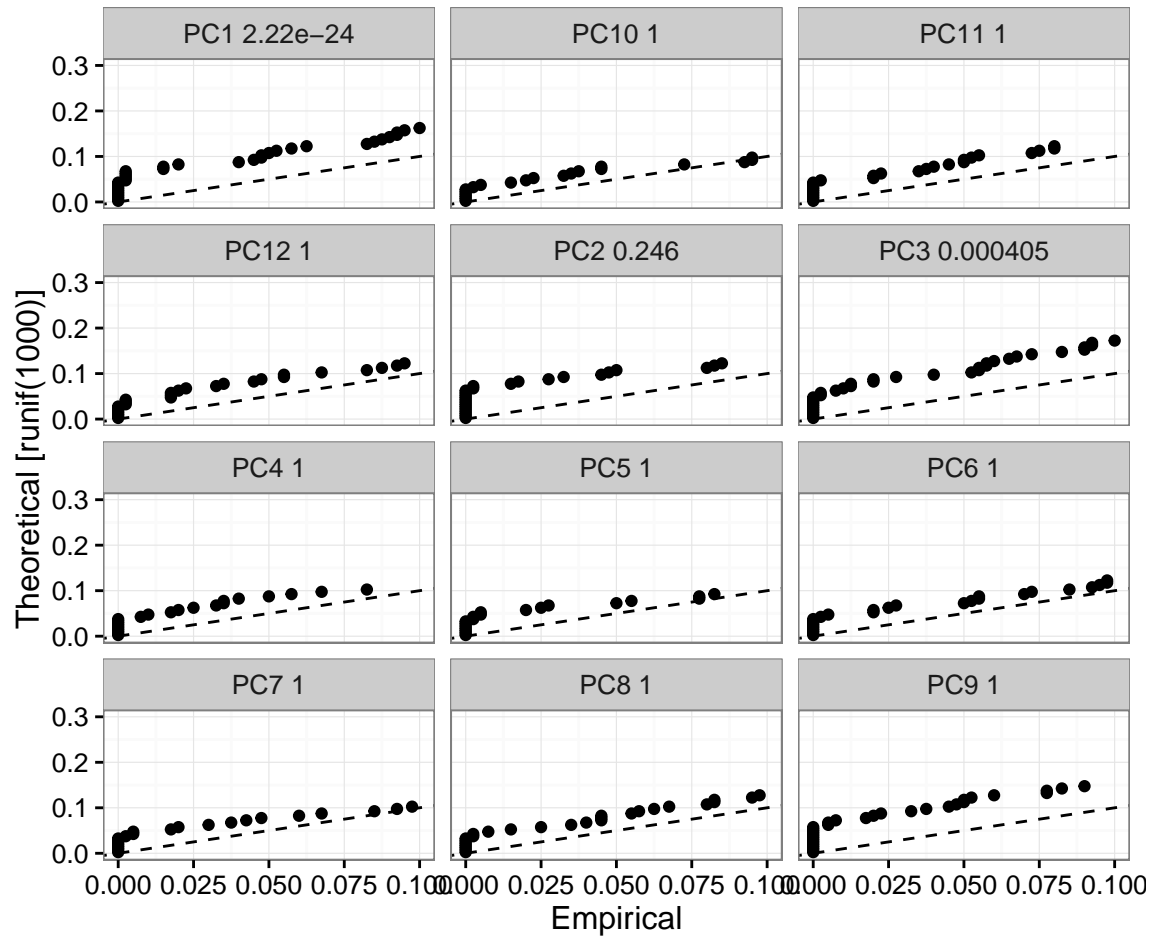


```
pcHeatmap(hpf18, 2, use.full = T, do.balanced = T, col.use = col) #technical
```

```
# Do 200 random samplings to find significant genes, each time randomly
# permute 1% of genes This returns a 'p-value' for each gene in each PC,
# based on how likely the gene/PC score would have been observed by chance
hpf18 = jackStraw(hpf18, num.replicate = 200, do.print = F)
```

```
# The jackStraw plot compares the distribution of P-values for each PC with
# a uniform distribution (dashed line) 'Significant' PCs will have a strong
# enrichment of genes with low p-values (solid curve above dashed line)
jackStrawPlot.new(hpf18, PCs = 1:12)
```

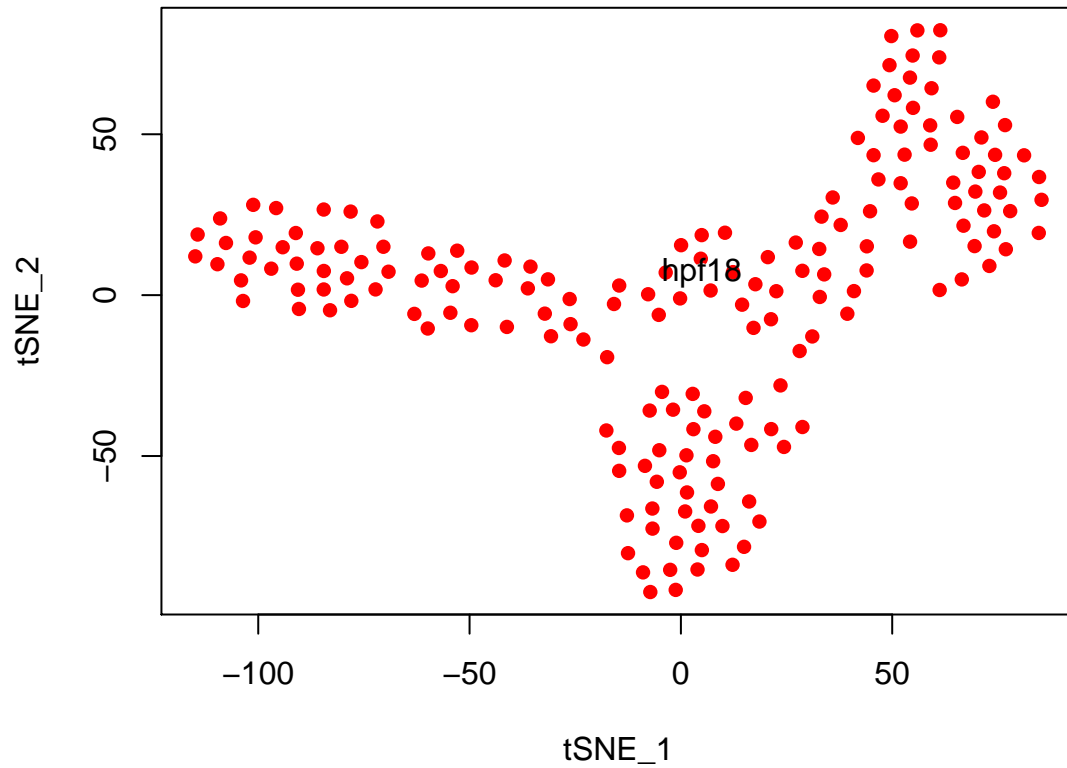


```

# In this case only PC1 and PC3 are strongly significant.

# Run tSNE using significant PCs as input (spectral tSNE), we get distinct
# point clouds
hpf18 = run_tsne(hpf18, max_iter = 2000, dims.use = c(1, 3))
tsne.plot(hpf18, do.label = T, label.pt.size = 1)

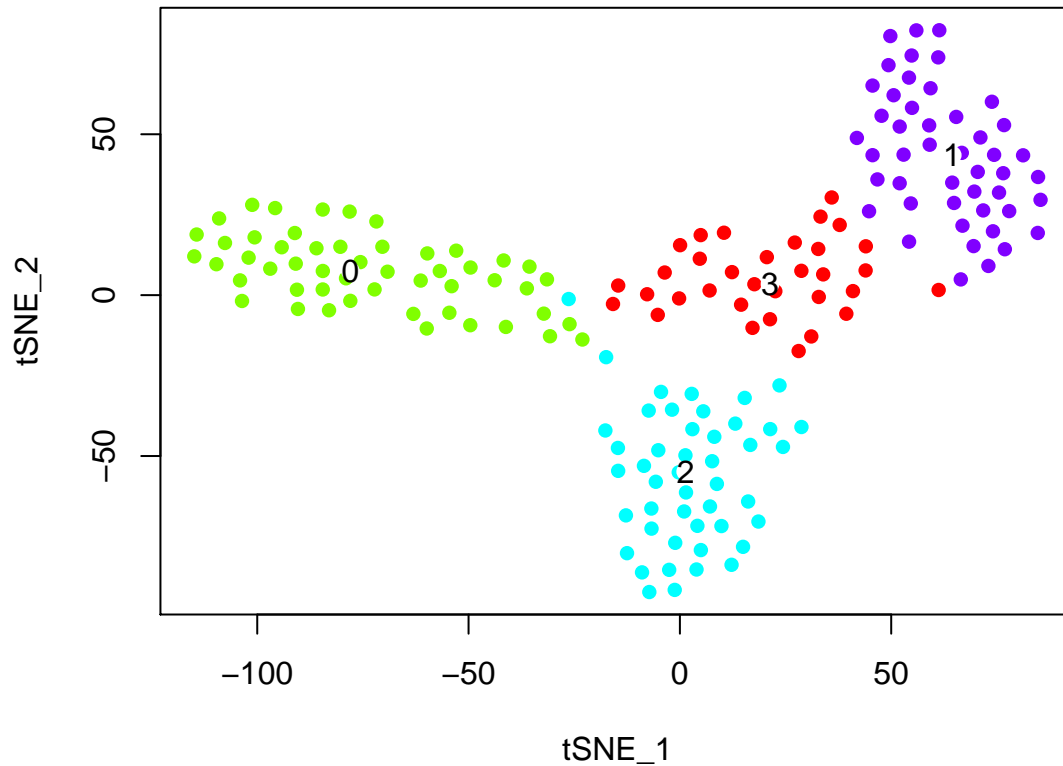
```



```
# Find cell clusters using Modularity optimization cluster detection.  
hpf18 = FindClusters(hpf18, pc.use = c(1, 3), do.modularity = T, resolution = 1,  
  prune.SNN = 0.1, print.output = 0, k.param = 20, k.scale = floor(177/20))
```

```
## [1] "SNN : processed 44 cells"  
## [1] "SNN : processed 88 cells"  
## [1] "SNN : processed 133 cells"  
## [1] "SNN : processed 177 cells"
```

```
tsne.plot(hpf18, do.label = T, label.pt.size = 1)
```

```
# The validity of the clusters can be validated using a classification
# scheme based on linear SVMs.
```

```
hpf18 = BuildSNN(hpf18, pc.use = c(1, 3), do.sparse = T, k.param = 20, k.scale = floor(
```

```
## [1] "SNN : processed 44 cells"
## [1] "SNN : processed 88 cells"
## [1] "SNN : processed 133 cells"
## [1] "SNN : processed 177 cells"
```

```
hpf18 = ValidateClusters(hpf18, pc.use = c(1, 3), min.connectivity = 0.001,
  acc.cutoff = 0.85)
```

```
## [1] "100% complete --- started with 4 clusters, 4 clusters remaining"
```

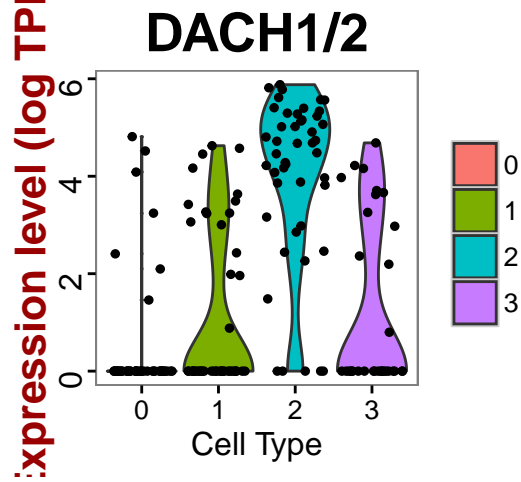
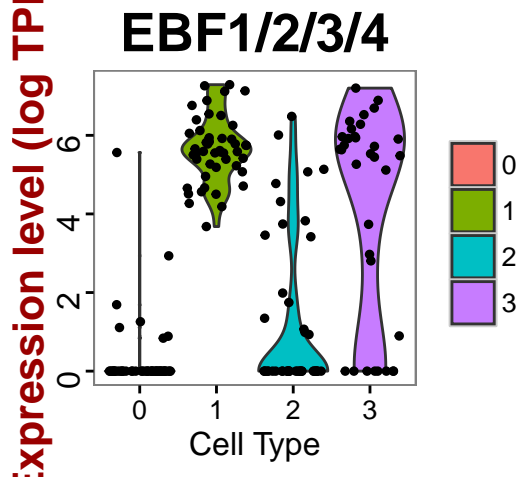
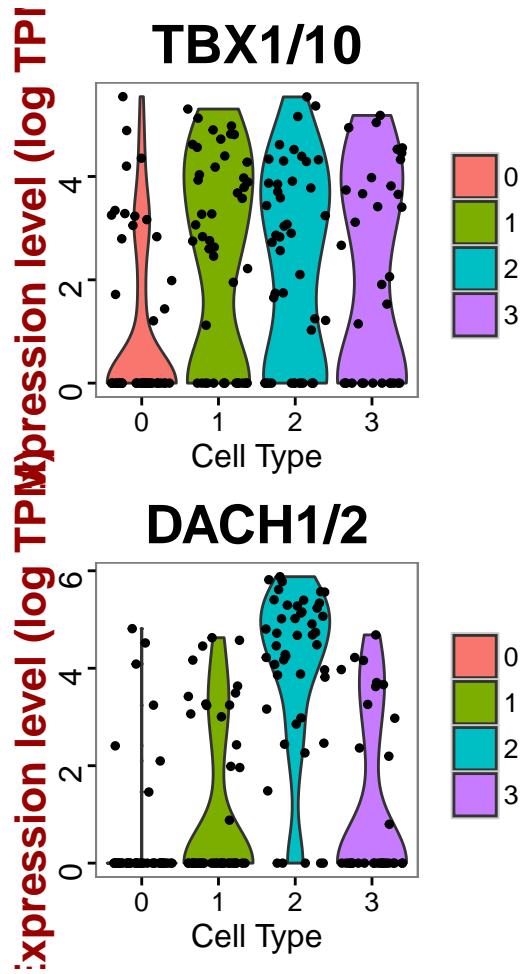
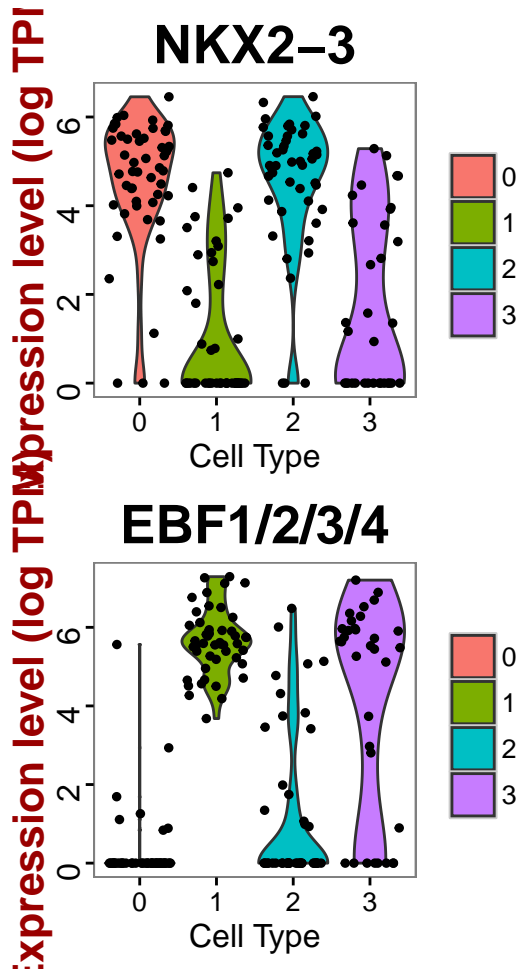
```
tsne.plot(hpf18, do.label = T, label.pt.size = 1)
```

```
# Find cluster markers using ROC test with thresh.use = 1, min.pct = 0.5 The
# ROC test returns the 'classification power' for any individual marker
# (ranging from 0 - random, to 1 - perfect). Though not a statistical test,
# it is often very useful for finding clean markers. Find markers for
# cluster 0
```

```
c10_18.markers = find.markers(hpf18, 0, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(c10_18.markers[order(c10_18.markers$myAUC, decreasing = T), ], 20)
```

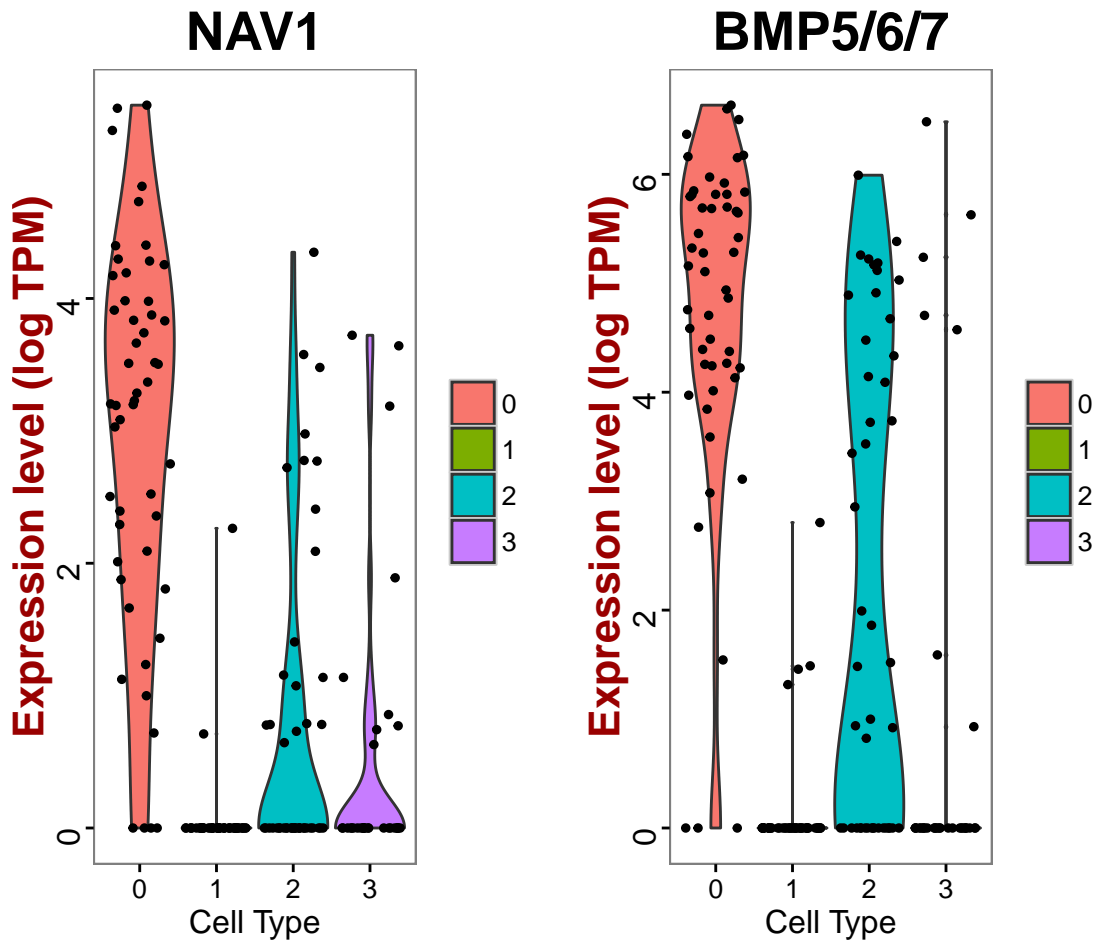
##	myAUC	avg_diff	power	pct.1	pct.2
## NAV1	0.906	2.416886	0.812	0.882	0.151
## KH2013:KH.C3.665_ZAN	0.905	1.528765	0.810	0.980	0.825
## MOGAT1/2	0.899	1.491851	0.798	1.000	0.579
## BMP5/6/7	0.890	1.943625	0.780	0.941	0.270
## KH2013:KH.C4.547_BMP2/4	0.881	1.778261	0.762	0.961	0.437
## KH2013:KH.C5.227_F56C4.4	0.862	1.196972	0.724	1.000	0.960
## KH2013:KH.C1.953_CG32702	0.847	1.778144	0.694	0.902	0.508
## LAMA1/2	0.835	1.664197	0.670	0.588	0.063
## SFRP1/5	0.833	3.061270	0.666	0.745	0.151
## COL13A1	0.828	2.536370	0.656	0.725	0.119
## Ci-R1CiGC27a04	0.821	3.809702	0.642	0.686	0.095
## FZD4	0.821	1.729355	0.642	0.784	0.159
## LYS2	0.814	1.294364	0.628	0.863	0.405
## EFNB1/2/3	0.805	1.119200	0.610	0.922	0.397
## KH2013:KH.C7.695_CYP24A1	0.798	1.849781	0.596	0.863	0.429
## KH2013:KH.C10.203	0.786	1.130693	0.572	0.902	0.651
## NTN1/2/3/5	0.782	2.734596	0.564	0.588	0.048
## RAC1/2	0.781	1.024299	0.562	0.941	0.611
## C5ORF48	0.778	1.347507	0.556	0.784	0.341
## SPECC1	0.769	1.041143	0.538	0.980	0.698

```
# Visualize known markers with a violin plot
vlnPlot(hpf18, c("NKX2-3", "EBF1/2/3/4", "TBX1/10", "DACH1/2"))
```



Based on EBF1/2/3/4- and NKX+, DACH1/2-, these are FHP cells

Visualize new markers with a violin plot
`vlnPlot(hpf18, c("NAV1", "BMP5/6/7"))`



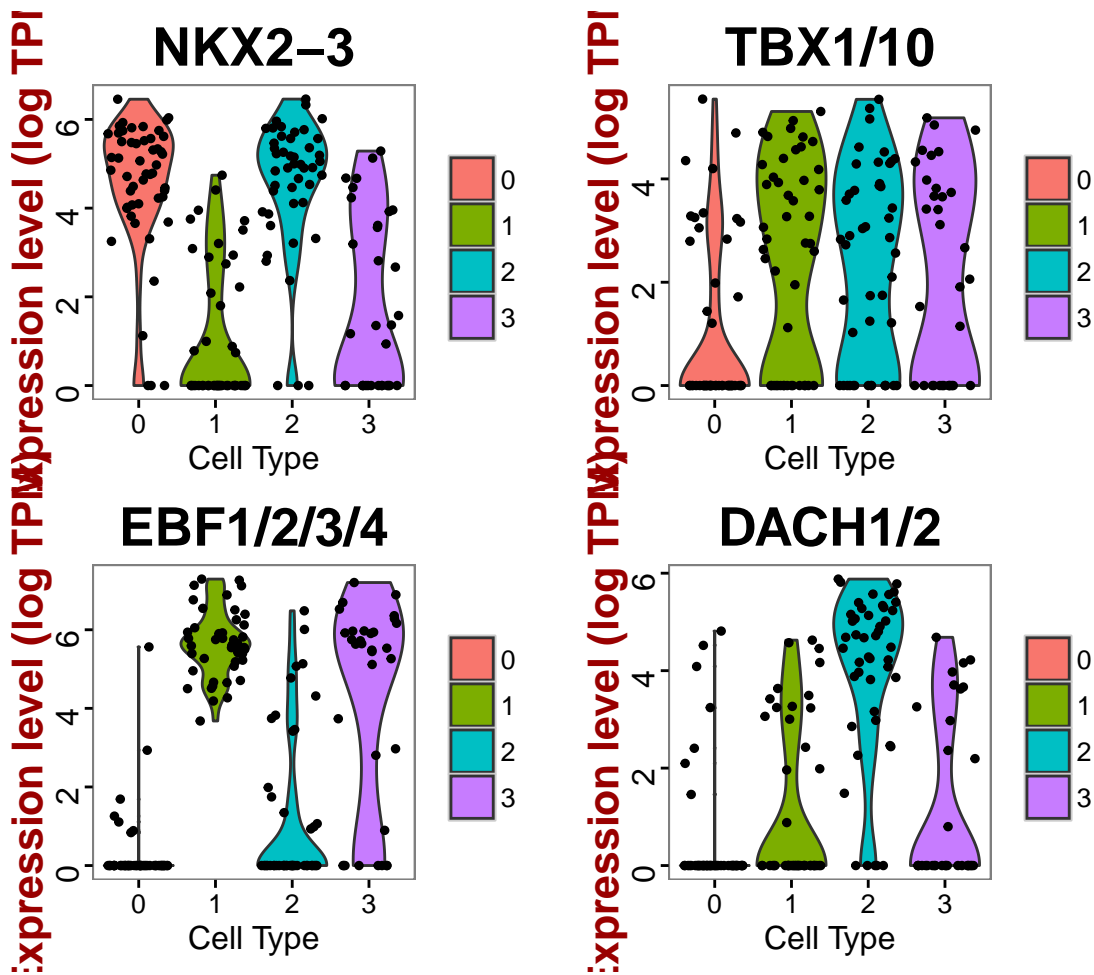
```
# Find markers for cluster 1
```

```
c11_18.markers = find.markers(hpf18, 1, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(c11_18.markers[order(c11_18.markers$myAUC, decreasing = T), ], 20)
```

Gene	myAUC	avg_diff	power	pct.1	pct.2
## KH2013:KH.C11.139	0.980	3.608234	0.960	0.979	0.254
## UBE2QL1	0.951	2.206912	0.902	1.000	0.354
## BDH1	0.931	1.935226	0.862	0.979	0.446
## KH2013:KH.C4.506_HMCN1	0.928	1.584676	0.856	1.000	0.808
## KH2013:KH.C5.127_FMO1	0.909	2.870722	0.818	0.872	0.108
## KH2013:KH.C5.62_ODF3L2	0.904	2.904233	0.808	0.872	0.108
## KH2013:KH.L18.10_THSD7A	0.893	1.859869	0.786	0.872	0.231
## KH2013:KH.C10.209	0.882	1.205272	0.764	1.000	0.777
## EBF1/2/3/4	0.875	1.459145	0.750	1.000	0.331
## Ci-R1CiGC09b24;	0.869	1.385696	0.738	1.000	0.785
## KH2013:KH.S1269.1	0.866	1.686455	0.732	0.915	0.331
## KH2013:KH.C8.782_LPHN1/2/3	0.859	1.144203	0.718	1.000	0.692
## SI:CH211-15101.2	0.856	1.477531	0.712	0.894	0.254
## KH2013:KH.C7.805_ID1/2/3/4	0.855	1.197553	0.710	0.979	0.815

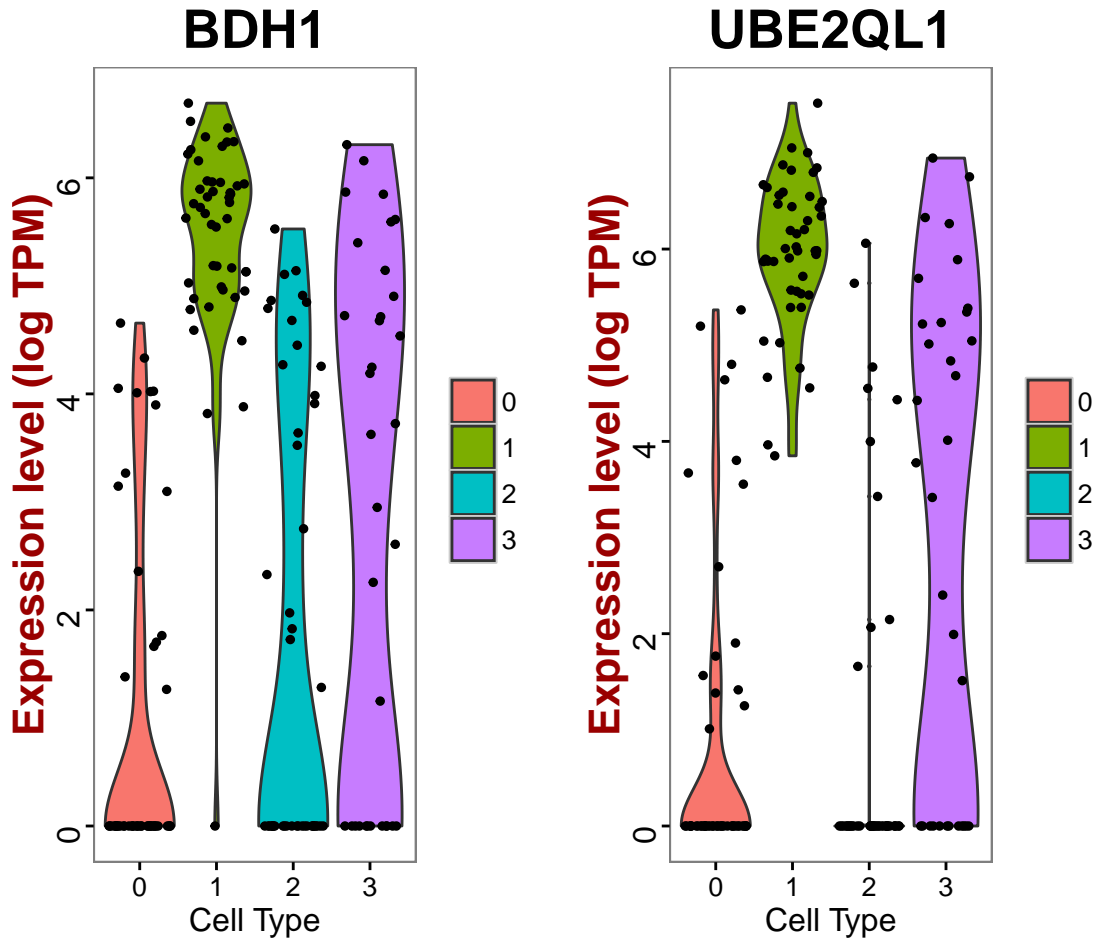
```
## SLC35G2/6          0.851 2.414261 0.702 0.766 0.100
## DCDC2B             0.847 1.681428 0.694 0.872 0.300
## KH2013:KH.L25.4   0.845 3.078121 0.690 0.723 0.062
## KH2013:KH.S511.3 0.839 1.126590 0.678 0.979 0.523
## TMSB15A           0.838 1.960725 0.676 0.872 0.346
## KH2013:KH.L154.26_CSMD2 0.834 1.168114 0.668 0.957 0.577
```

```
# Visualize known markers with a violin plot
vlnPlot(hpf18, c("NKX2-3", "EBF1/2/3/4", "TBX1/10", "DACH1/2"))
```



```
# Based on previous discoveries NKX- EBF+ TBX+ DACH- cells are ASM
```

```
# Visualize new markers with a violin plot
vlnPlot(hpf18, c("BDH1", "UBE2QL1"))
```



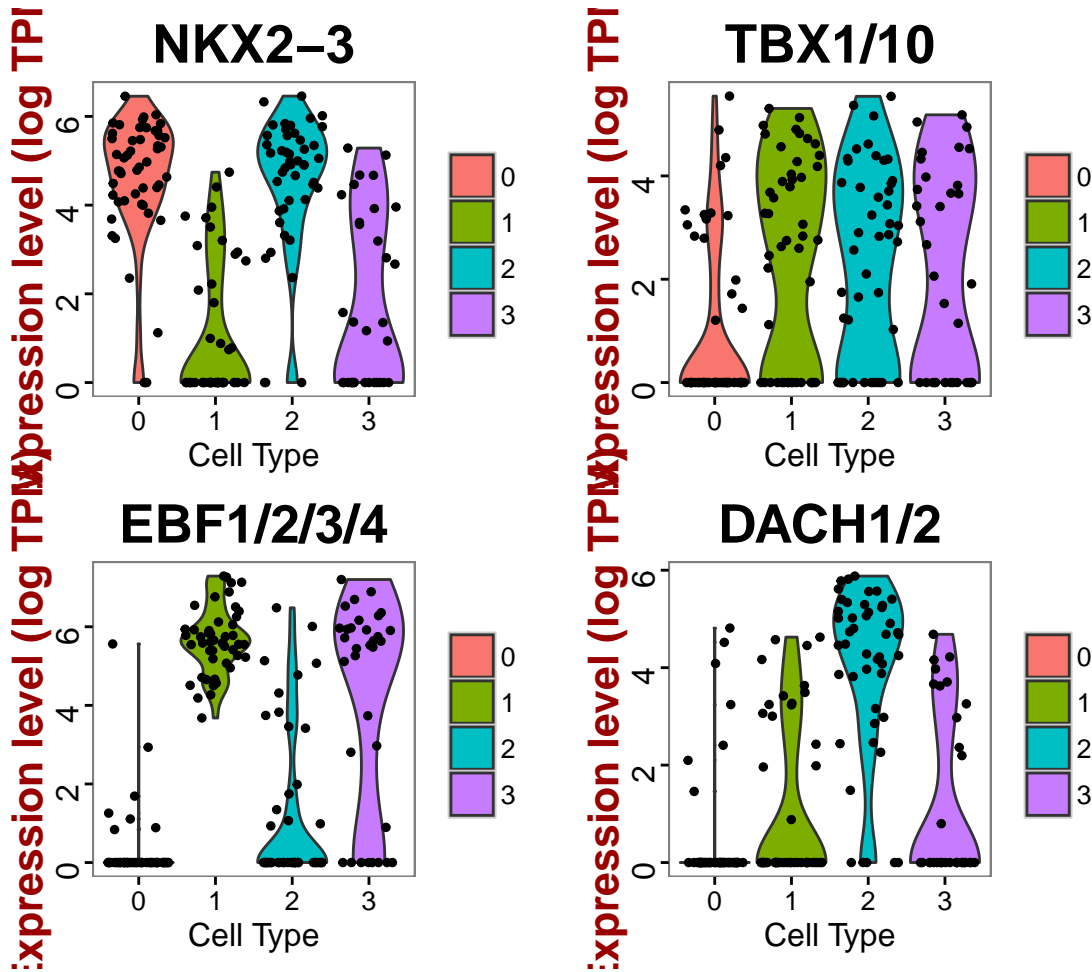
```
# Find markers for cluster 2
```

```
c12_18.markers = find.markers(hpf18, 2, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(c12_18.markers[order(c12_18.markers$myAUC, decreasing = T), ], 20)
```

	myAUC	avg_diff	power	pct.1	pct.2
## DACH1/2	0.885	2.322664	0.770	0.891	0.252
## KH2013:KH.C1.638_C17ORF105	0.841	1.244018	0.682	0.935	0.740
## KH2013:KH.C10.174	0.832	1.082488	0.664	1.000	0.580
## SCYL	0.797	1.327073	0.594	0.891	0.527
## KH2013:KH.C10.172	0.771	1.475241	0.542	0.848	0.557
## FOXF1/2	0.738	1.061960	0.476	0.913	0.763
## SUDS3	0.732	1.072577	0.464	0.913	0.718
## LENG8	0.700	1.019048	0.400	0.630	0.389
## KH2013:KH.C3.716_EFNA1/2/3/4/5	0.699	1.004073	0.398	0.870	0.618
## KH2013:KH.C6.44_KCNK10/2/4	0.692	1.027052	0.384	0.674	0.298
## KH2013:KH.C3.299	0.691	1.150896	0.382	0.587	0.252
## DLG5	0.675	1.005763	0.350	0.630	0.321
## KH2013:KH.S725.5	0.670	1.190847	0.340	0.652	0.382
## KH2013:KH.C2.147	0.659	1.120100	0.318	0.522	0.244

```
## KH2013:KH.L108.49      0.638  1.017853  0.276  0.543  0.290
## MATN1/3/4              0.628  1.046960  0.256  0.652  0.481
## FIR1                   0.620  1.138595  0.240  0.543  0.336
## ODF2                   0.589  1.003972  0.178  0.565  0.458
## MICALL1/2             0.532 -1.529114  0.064  0.565  0.481
## SMEK1/2               0.529  1.923853  0.058  0.565  0.534
```

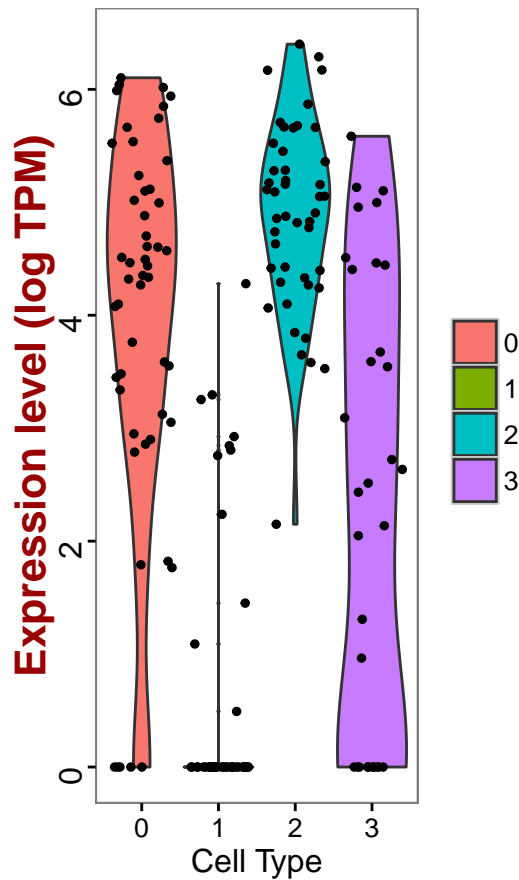
```
# Visualize known markers with a violin plot
vlnPlot(hpf18, c("NKX2-3", "EBF1/2/3/4", "TBX1/10", "DACH1/2"))
```



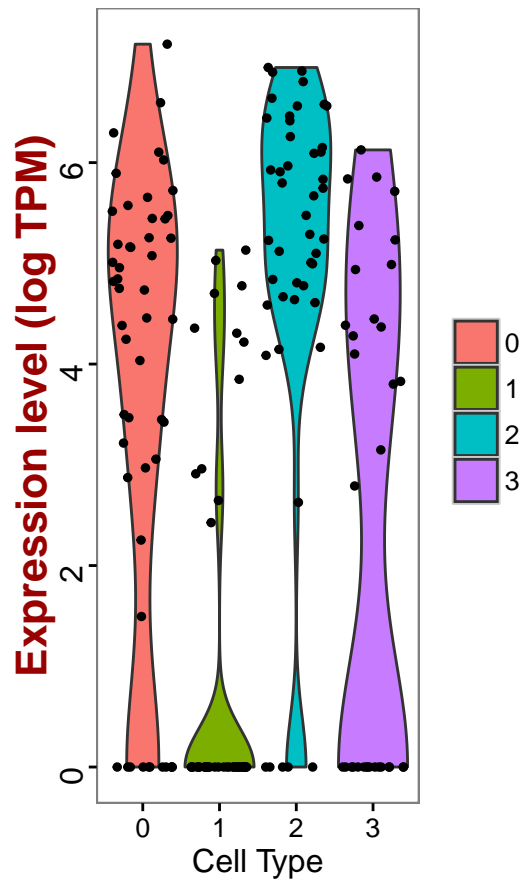
```
# Based on preliminary discoveries these NKX+ EBF1/2/3/4- TBX+ DACH1/2+
# cells are SHP's
```

```
# Visualize new markers with a violin plot
vlnPlot(hpf18, c("KH2013:KH.C10.174", "SCYL"))
```

KH2013:KH.C10.174



SCYL



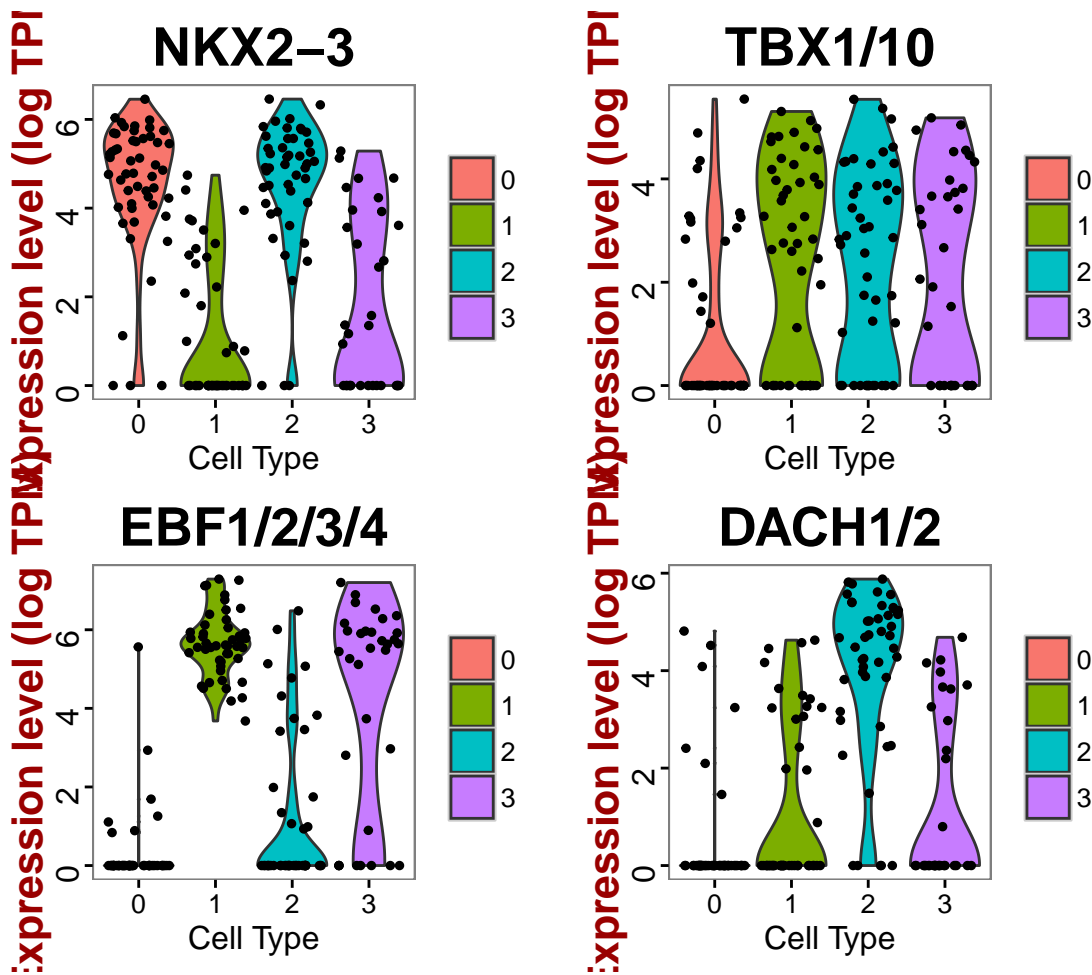
Find markers for cluster 3

```
cl3_18.markers = find.markers(hpf18, 3, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(cl3_18.markers[order(cl3_18.markers$myAUC, decreasing = T), ], 20)
```

	myAUC	avg_diff	power	pct.1	pct.2
## TMEM144	0.652	1.230936	0.304	0.515	0.243
## LGR4/5/6	0.643	1.121841	0.286	0.727	0.542
## BECN1	0.640	1.131217	0.280	0.576	0.319
## SLC6A1/5	0.621	1.602470	0.242	0.636	0.361
## PARP2	0.613	1.028850	0.226	0.576	0.396
## KH2013:KH.C9.182_AADAT	0.585	1.062296	0.170	0.515	0.438
## ITPKA	0.544	1.161089	0.088	0.515	0.535
## MICALL1/2	0.535	-1.558863	0.070	0.576	0.486
## PIGT	0.489	-4.667773	0.022	0.545	0.556
## RBM15B	0.473	-2.221725	0.054	0.697	0.792
## SMEK1/2	0.471	-2.776749	0.058	0.515	0.549
## KH2013:KH.C1.953_CG32702	0.405	-1.000614	0.190	0.576	0.632
## RARG	0.392	-1.025694	0.216	0.455	0.549
## TTC19	0.369	-1.209717	0.262	0.273	0.514

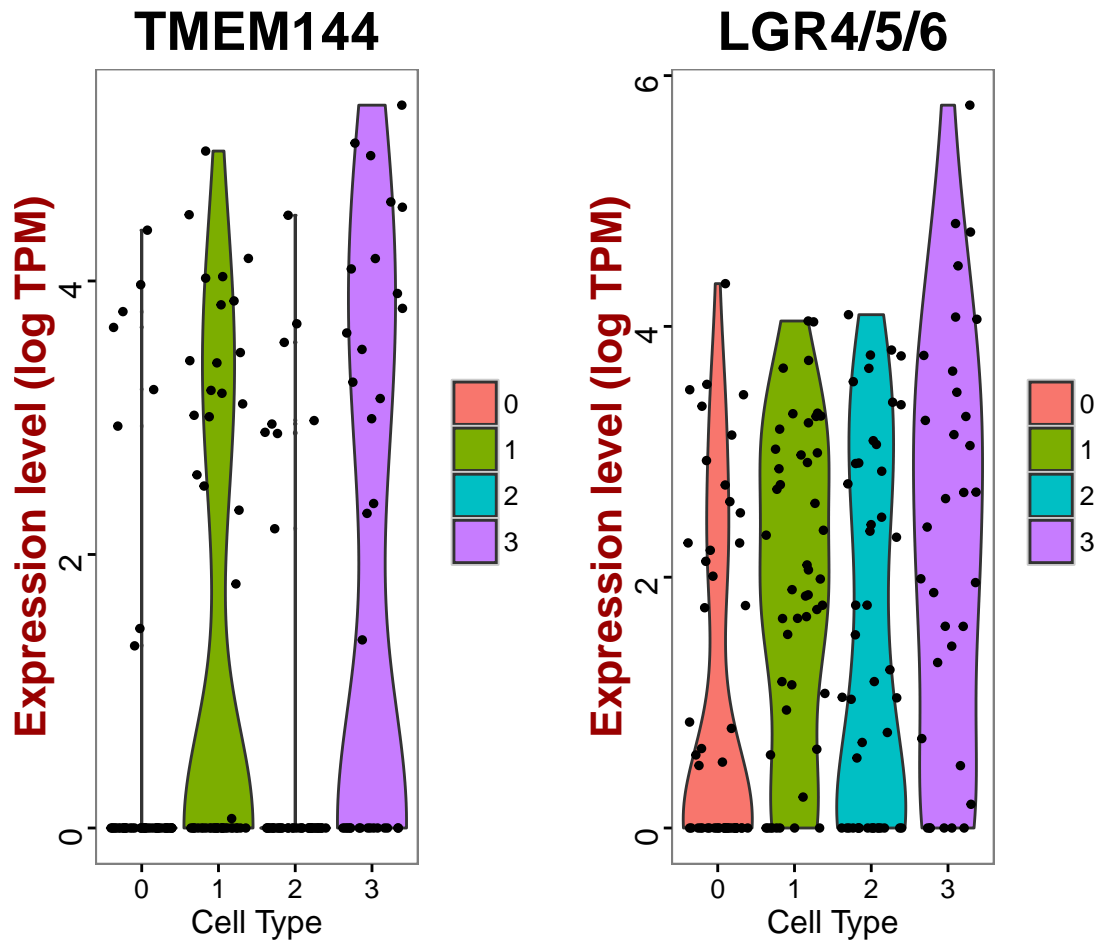

```
## KH2013:KH.C8.190      0.358 -2.057851 0.284 0.333 0.549
## GPR37                  0.354 -1.300561 0.292 0.424 0.583
## KH2013:KH.C7.695_CYP24A1 0.340 -1.290304 0.320 0.333 0.604
## SLIT1/2/3             0.318 -1.109890 0.364 0.545 0.743
## RBMS1/2/3            0.302 -1.117102 0.396 0.394 0.701
## NKX2-3                0.291 -1.361200 0.418 0.515 0.729
```

```
# Visualize known markers with a violin plot
vlnPlot(hpf18, c("NKX2-3", "EBF1/2/3/4", "TBX1/10", "DACH1/2"))
```

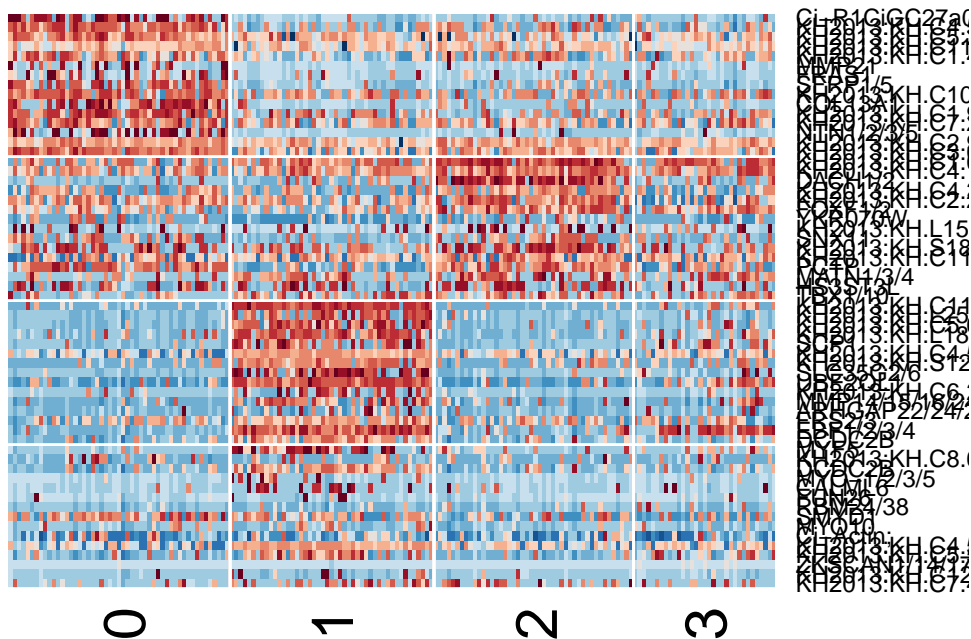


```
# Based on preliminary discoveries these NKX- EBF1/2/3/4+ TBX+ DACH1/2-
# cells look like ASM but has few highly expressed markers
```

```
# Visualize new markers with a violin plot
vlnPlot(hpf18, c("TMEM144", "LGR4/5/6"))
```



```
# Plot hpf20 markers
doHeatMap(hpf18, genes.use = marker.20, slim.col.label = T, remove.key = T,
  rowsep = seq(0, 60, 15), col.use = col)
```



```
# Based on the heatmap this group of cells do not express 20ASM2
# markers(later differentiated markers), also it tend to express some heart
# progenitor markers. Named Unknown for late analysis
```

```
# Write cell names and markers into text files
```

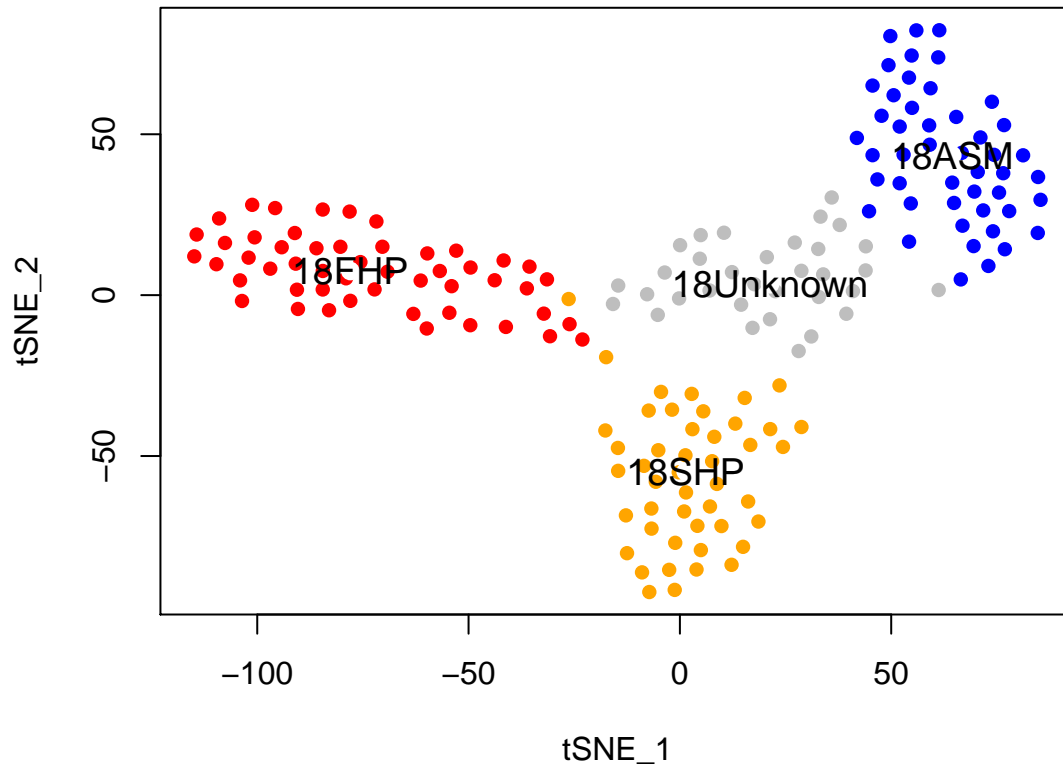
```
write.table(which.cells(hpf18, 0), file = "18FHPCells.txt", sep = "\t")
write.table(which.cells(hpf18, 1), file = "18ASMCells.txt", sep = "\t")
write.table(which.cells(hpf18, 2), file = "18SHPCells.txt", sep = "\t")
```

```
# Rename cluster identities
```

```
hpf18 = rename.ident(hpf18, 0, "18FHP")
hpf18 = rename.ident(hpf18, 1, "18ASM")
hpf18 = rename.ident(hpf18, 2, "18SHP")
hpf18 = rename.ident(hpf18, 3, "18Unknown")
```

```
# Visualize tSNE used color scheme FHP-red, SHP-orange, ASM-blue
```

```
tsne.plot(hpf18, do.label = T, label.pt.size = 1, label.cex.text = 1.2, label.cols.use =
  "red", "orange", "blue"))
```



```
# Store FHP markers in text file
FHP_18.markers = find.markers(hpf18, "18FHP", c("18SHP", "18ASM"), thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
head(FHP_18.markers[order(FHP_18.markers$myAUC, decreasing = T), ], 20)
```

##	myAUC	avg_diff	power	pct.1	pct.2
## MOGAT1/2	0.921	1.695660	0.842	1.000	0.591
## NAV1	0.909	2.469266	0.818	0.882	0.151
## KH2013:KH.C3.665_ZAN	0.909	1.565571	0.818	0.980	0.817
## KH2013:KH.C5.227_F56C4.4	0.892	1.683649	0.784	1.000	0.946
## BMP5/6/7	0.891	2.057759	0.782	0.941	0.301
## KH2013:KH.C4.547_BMP2/4	0.886	1.917017	0.772	0.961	0.452
## KH2013:KH.C1.953_CG32702	0.849	1.780812	0.698	0.902	0.484
## LAMA1/2	0.838	1.650821	0.676	0.588	0.054
## SFRP1/5	0.828	2.941007	0.656	0.745	0.172
## COL13A1	0.826	2.513739	0.652	0.725	0.118
## Ci-R1CiGC27a04	0.824	4.002483	0.648	0.686	0.086
## C5ORF48	0.821	2.357489	0.642	0.784	0.280
## FZD4	0.821	1.810412	0.642	0.784	0.183
## KH2013:KH.C10.203	0.804	1.277857	0.608	0.902	0.645
## LYS2	0.803	1.212652	0.606	0.863	0.430
## EFNB1/2/3	0.798	1.092338	0.596	0.922	0.387
## KH2013:KH.C11.362	0.792	2.079359	0.584	0.784	0.355

```
## NTN1/2/3/5          0.786 2.737731 0.572 0.588 0.043
## KH2013:KH.C7.695_CYP24A1 0.786 1.784354 0.572 0.863 0.462
## WNT9B              0.763 1.601544 0.526 0.725 0.301
```

```
write.table(FHP_18.markers, file = "FHP_18.markers.txt", sep = "\t")
```

```
# Store SHP markers in text file
```

```
SHP_18.markers = find.markers(hpf18, "18SHP", c("18FHP", "18ASM"), thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
head(SHP_18.markers[order(SHP_18.markers$myAUC, decreasing = T), ], 20)
```

```
##          myAUC avg_diff power pct.1 pct.2
## DACH1/2    0.892 2.438529 0.784 0.891 0.224
## KH2013:KH.C1.638_C17ORF105 0.860 1.393115 0.720 0.935 0.714
## SCYL       0.794 1.296783 0.588 0.891 0.531
## FOXF1/2    0.790 1.416604 0.580 0.913 0.704
## KH2013:KH.C10.172 0.769 1.482743 0.538 0.848 0.582
## SPSB1/4    0.753 1.110795 0.506 0.978 0.796
## SUDS3      0.749 1.180373 0.498 0.913 0.704
## ZNF703     0.719 1.080167 0.438 0.652 0.245
## KH2013:KH.C6.44_KCNK10/2/4 0.704 1.173353 0.408 0.674 0.296
## KH2013:KH.C3.232 0.703 1.013660 0.406 0.674 0.347
## CCNB3      0.692 1.069960 0.384 0.717 0.408
## KH2013:KH.C3.299 0.688 1.224314 0.376 0.587 0.276
## DLG5       0.681 1.004789 0.362 0.630 0.316
## KH2013:KH.C2.147 0.671 1.243503 0.342 0.522 0.224
## KH2013:KH.S725.5 0.668 1.163315 0.336 0.652 0.398
## MATN1/3/4  0.649 1.204521 0.298 0.652 0.439
## KH2013:KH.C14.406 0.638 1.101730 0.276 0.717 0.531
## KH2013:KH.C12.536 0.630 1.082419 0.260 0.848 0.786
## histone    0.629 1.006188 0.258 0.739 0.673
## KH2013:KH.S1660.1 0.591 1.010002 0.182 0.739 0.684
```

```
write.table(SHP_18.markers, file = "SHP_18.markers.txt", sep = "\t")
```

```
# Store ASM markers in text file
```

```
ASM_18.markers = find.markers(hpf18, "18ASM", c("18SHP", "18FHP"), thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
head(ASM_18.markers[order(ASM_18.markers$myAUC, decreasing = T), ], 20)
```

```
##          myAUC avg_diff power pct.1 pct.2
## KH2013:KH.C11.139 0.984 4.588292 0.968 0.979 0.196
## UBE2QL1     0.984 3.208404 0.968 1.000 0.258
```

```

## EBF1/2/3/4          0.971 2.877694 0.942 1.000 0.196
## BDH1                0.964 2.570204 0.928 0.979 0.381
## KH2013:KH.C4.506_HMCN1 0.942 1.738256 0.884 1.000 0.794
## Ci-R1CiGC09b24;    0.939 2.065758 0.878 1.000 0.753
## KH2013:KH.C10.209  0.931 1.544434 0.862 1.000 0.732
## KH2013:KH.C5.127_FM01 0.928 4.482840 0.856 0.872 0.082
## KH2013:KH.C5.62_ODF3L2 0.919 3.552450 0.838 0.872 0.072
## SI:CH211-15101.2   0.917 2.834658 0.834 0.894 0.155
## KH2013:KH.C8.782_LPHN1/2/3 0.910 1.483750 0.820 1.000 0.670
## KH2013:KH.L18.10_THSD7A 0.910 2.158503 0.820 0.872 0.175
## KH2013:KH.S1269.1  0.890 1.958529 0.780 0.915 0.289
## KH2013:KH.L154.26_CSMD2 0.889 1.614030 0.778 0.957 0.536
## TMSB15A            0.886 2.831017 0.772 0.872 0.247
## KH2013:KH.S511.3   0.882 1.513598 0.764 0.979 0.454
## SLC35G2/6          0.877 4.418579 0.754 0.766 0.041
## KH2013:KH.C7.805_ID1/2/3/4 0.874 1.352515 0.748 0.979 0.804
## ELK1/3/4           0.864 1.368382 0.728 0.979 0.423
## KH2013:KH.S555.1_HTR7 0.862 1.327817 0.724 0.979 0.443

```

```
write.table(ASM_18.markers, file = "ASM_18.markers.txt", sep = "\t")
```

```
# Find pan Heart Progenitor markers
```

```
panHP_18.markers = find.markers(hpf18, c("18SHP", "18FHP"), "18ASM", thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
```

```
head(panHP_18.markers[order(panHP_18.markers$myAUC, decreasing = T), ], 20)
```

```

##                myAUC avg_diff power pct.1 pct.2
## KH2013:KH.C10.174 0.952 3.337930 0.904 0.948 0.213
## SLIT1/2/3        0.949 2.523519 0.898 0.979 0.255
## GATA4/5/6        0.946 3.202716 0.892 0.938 0.191
## QKI              0.931 2.260126 0.862 0.979 0.723
## NKX2-3           0.928 2.720123 0.856 0.938 0.298
## SPECC1           0.916 2.230076 0.832 0.979 0.489
## PCTP             0.910 2.477431 0.820 0.938 0.362
## TNS1/3           0.901 2.394328 0.802 0.928 0.255
## EFN1/2/3         0.900 3.684817 0.800 0.825 0.064
## KH2013:KH.C8.80_SI:DKEY-29D8.3 0.895 1.182032 0.790 1.000 0.872
## LRP4/8           0.892 2.625513 0.784 0.825 0.085
## KH2013:KH.C8.489_SI:DKEY-29D8.3 0.887 2.125379 0.774 0.928 0.617
## KH2013:KH.C2.994_RNF149 0.885 1.044521 0.770 1.000 0.979
## BMP5/6/7         0.877 4.532942 0.754 0.742 0.085
## KH2013:KH.C2.935_S1PR1/2/3/4/5 0.875 1.766753 0.750 0.969 0.468
## SH3BGR           0.873 2.986558 0.746 0.804 0.128
## KH2013:KH.C3.665_ZAN 0.868 1.999101 0.736 0.959 0.702

```

```
## SCYL                0.855 2.519750 0.710 0.835 0.255
## IGF1/2              0.854 2.586108 0.708 0.794 0.149
## LYS2                0.850 2.201142 0.700 0.773 0.191
```

```
write.table(panHP_18.markers, file = "panHP_18.markers.txt", sep = "\t")

# Find SHP specific markers that distinguish tow heart progenitors FHP and
# SHP
SHPspecific_18.markers = find.markers(hpf18, "18SHP", "18FHP", thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
head(SHPspecific_18.markers[order(SHPspecific_18.markers$myAUC, decreasing = T),
  ], 20)
```

```
##                myAUC avg_diff power pct.1 pct.2
## DACH1/2        0.911 2.783039 0.822 0.891 0.137
## KH2013:KH.C1.638_C17ORF105 0.878 1.574147 0.756 0.935 0.588
## GDH1/3         0.807 1.282798 0.614 0.978 0.824
## ZFP318         0.797 1.205806 0.594 0.870 0.588
## KH2013:KH.L154.26_CSMD2 0.794 1.907616 0.588 0.783 0.314
## KH2013:KH.C9.692_GABRR1/2/3 0.768 2.048822 0.536 0.717 0.333
## TUBA1L/2       0.767 1.329805 0.534 0.935 0.941
## FOXF1/2        0.761 1.189351 0.522 0.913 0.765
## ZNF703         0.757 1.547560 0.514 0.652 0.157
## ELK1/3/4       0.746 1.261431 0.492 0.652 0.216
## KH2013:KH.C10.172 0.731 1.248617 0.462 0.848 0.725
## PCBP3          0.720 1.574723 0.440 0.630 0.314
## MATN1/3/4      0.708 1.603016 0.416 0.652 0.294
## NOTCH1/4       0.708 2.316566 0.416 0.522 0.118
## KH2013:KH.C9.208_TTN 0.705 1.052186 0.410 0.630 0.314
## ZNF367         0.702 1.026594 0.404 0.652 0.275
## PRKD1/2/3      0.697 1.127657 0.394 0.587 0.353
## CCNB3          0.696 1.039161 0.392 0.717 0.373
## NOVA1/2        0.693 1.657795 0.386 0.609 0.314
## KH2013:KH.C14.406 0.689 1.426562 0.378 0.717 0.471
```

```
write.table(SHPspecific_18.markers, file = "SHPspecific_18.markers.txt", sep = "\t")

# Find FHP specific markers that distinguish tow heart progenitors FHP and
# SHP
FHPspecific_18.markers = find.markers(hpf18, "18FHP", "18SHP", thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
head(FHPspecific_18.markers[order(FHPspecific_18.markers$myAUC, decreasing = T),
  ], 20)
```

```
##          myAUC avg_diff power pct.1 pct.2
## MOGAT1/2      0.866 1.227366 0.732 1.000 0.696
## KH2013:KH.C1.953_CG32702 0.865 1.958750 0.730 0.902 0.391
## NAV1          0.862 1.946789 0.724 0.882 0.283
## KH2013:KH.C5.227_F56C4.4 0.859 1.443269 0.718 1.000 0.935
## KH2013:KH.C3.665_ZAN    0.844 1.114215 0.688 0.980 0.935
## KH2013:KH.C4.547_BMP2/4 0.841 1.478739 0.682 0.961 0.565
## BMP5/6/7      0.814 1.381609 0.628 0.941 0.522
## COL13A1       0.813 2.370790 0.626 0.725 0.174
## Ci-R1CiGC27a04 0.809 3.468222 0.618 0.686 0.152
## C5ORF48       0.808 2.160508 0.616 0.784 0.326
## LAMA1/2       0.806 1.428195 0.612 0.588 0.109
## KH2013:KH.C10.203      0.802 1.238416 0.604 0.902 0.587
## SFRP1/5       0.801 2.553373 0.602 0.745 0.283
## NTN1/2/3/5    0.769 2.193801 0.538 0.588 0.087
## KH2013:KH.C3.21_HSPG2  0.751 2.223697 0.502 0.549 0.130
## FZD4          0.751 1.169858 0.502 0.784 0.370
## KH2013:KH.C11.362      0.749 1.660434 0.498 0.784 0.478
## KH2013:KH.C7.695_CYP24A1 0.738 1.434126 0.476 0.863 0.587
## DSEL          0.734 1.339817 0.468 0.667 0.239
## KH2013:KH.C7.649_ENPPP7 0.723 1.007389 0.446 0.765 0.435
```

```
write.table(FHPspecific_18.markers, file = "FHPspecific_18.markers.txt", sep = "\t")
```

```
# Find gene expression percentage among single cells
```

```
hpf18.pct = cluster.alpha(hpf18, thresh.min = 0)
```

```
# Find top 50 pan heart marker expression percentage in 18Unknown cells
```

```
sum(hpf18.pct[rownames(head(panHP_18.markers[order(panHP_18.markers$myAUC, decreasing =
  ], 50))), 4] > 0.5)/50
```

```
## [1] 0.7
```

```
# Unknown cells have 70% top 50 pan heart marker expressed
```

```
sum(hpf18.pct[rownames(head(panHP_18.markers[order(panHP_18.markers$myAUC, decreasing =
  ], 50))), c(1, 3)] > 0.5)/100
```

```
## [1] 0.98
```

```
# FHP and SHP cells have only 98% top 50 pan heart marker expressed
```

```
sum(hpf18.pct[rownames(head(panHP_18.markers[order(panHP_18.markers$myAUC, decreasing =
  ], 50))), 2] > 0.5)/100
```

```
## [1] 0.21
```



```
# ASM cells have 21% top 50 pan heart marker expressed based on pan heart  
# markers the Unknwon cells express more heart progenitors markers
```

```
# Find top 50 pan heart marker expression percentage in 18Unknown cells
```

```
sum(hpf18.pct[rownames(head(ASM_18.markers[order(ASM_18.markers$myAUC, decreasing = T),  
], 50)), 4] > 0.5)/50
```

```
## [1] 0.64
```

```
# Unknown cells have 74% top 50 ASM marker expressed
```

```
sum(hpf18.pct[rownames(head(ASM_18.markers[order(ASM_18.markers$myAUC, decreasing = T),  
], 50)), c(1, 3)] > 0.5)/100
```

```
## [1] 0.3
```

```
# FHP and SHP cells have only 30% top 50 ASM marker expressed
```

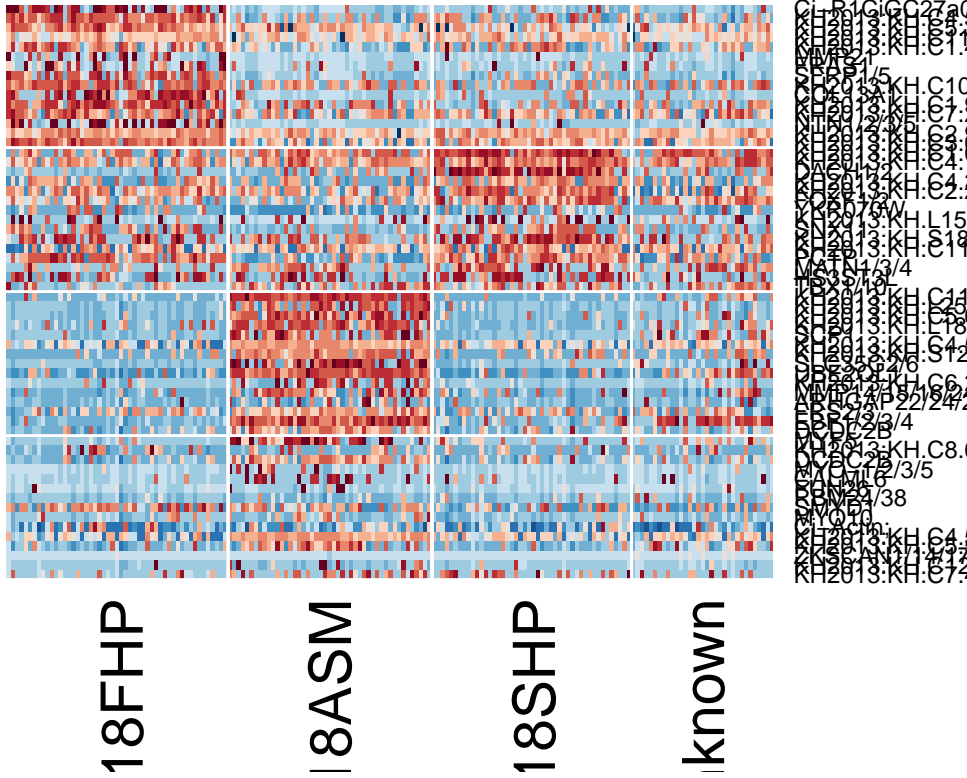
```
sum(hpf18.pct[rownames(head(ASM_18.markers[order(ASM_18.markers$myAUC, decreasing = T),  
], 50)), 2] > 0.5)/50
```

```
## [1] 1
```

```
# ASM cells have 100% top 50 ASM marker expressed based on ASM markers the  
# Unknwon cells express more ASM progenitors markers
```

```
# Heat map of 20hpf markers also demonstrates that the 18Unknown cells tend  
# to have both heart and muscle progenitor characteristics
```

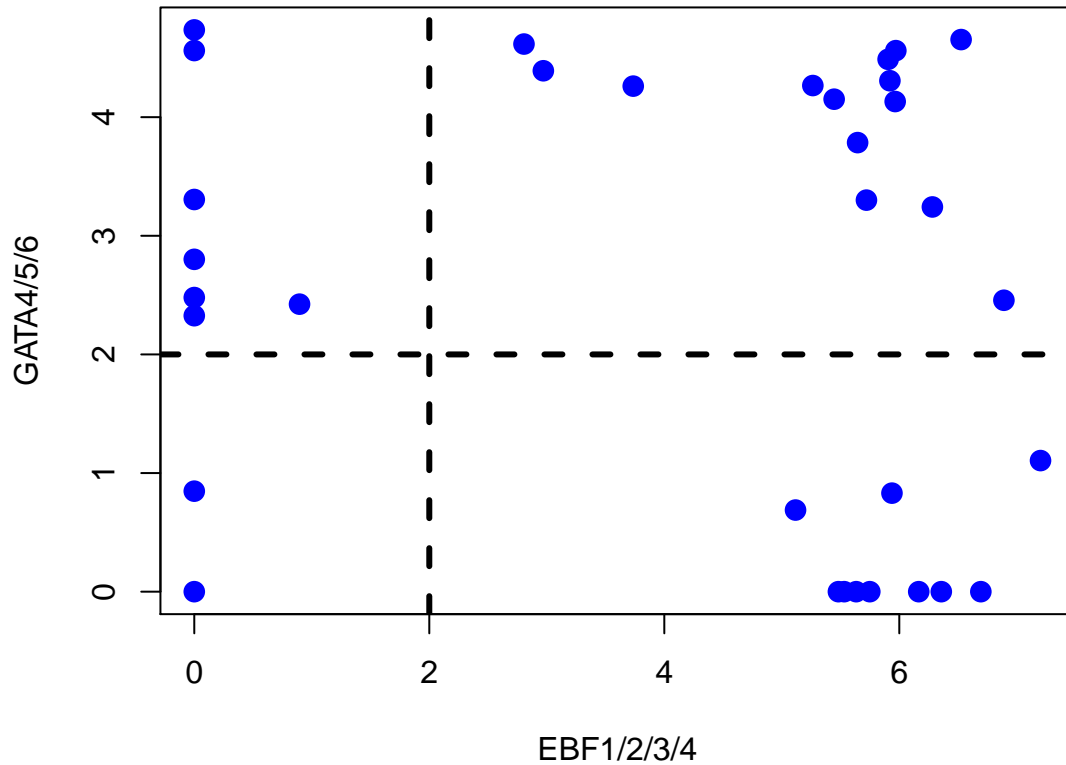
```
doHeatMap(hpf18, genes.use = marker.20, slim.col.label = T, remove.key = T,  
rowsep = seq(0, 60, 15), col.use = col)
```



Based on available published data EBF1/2/3/4 and GATA4/5/6 inhibiting each other during the fate determination of ASM and SHP. No cells were previously reported to have both EBF and GATA expression in TVC lineage (FISH data).

```
genePlot(hpf18, cell.ids = which.cells(hpf18, "18Unknown"), "EBF1/2/3/4", "GATA4/5/6")
abline(v = 2, h = 2, lwd = 3, lty = 2)
```

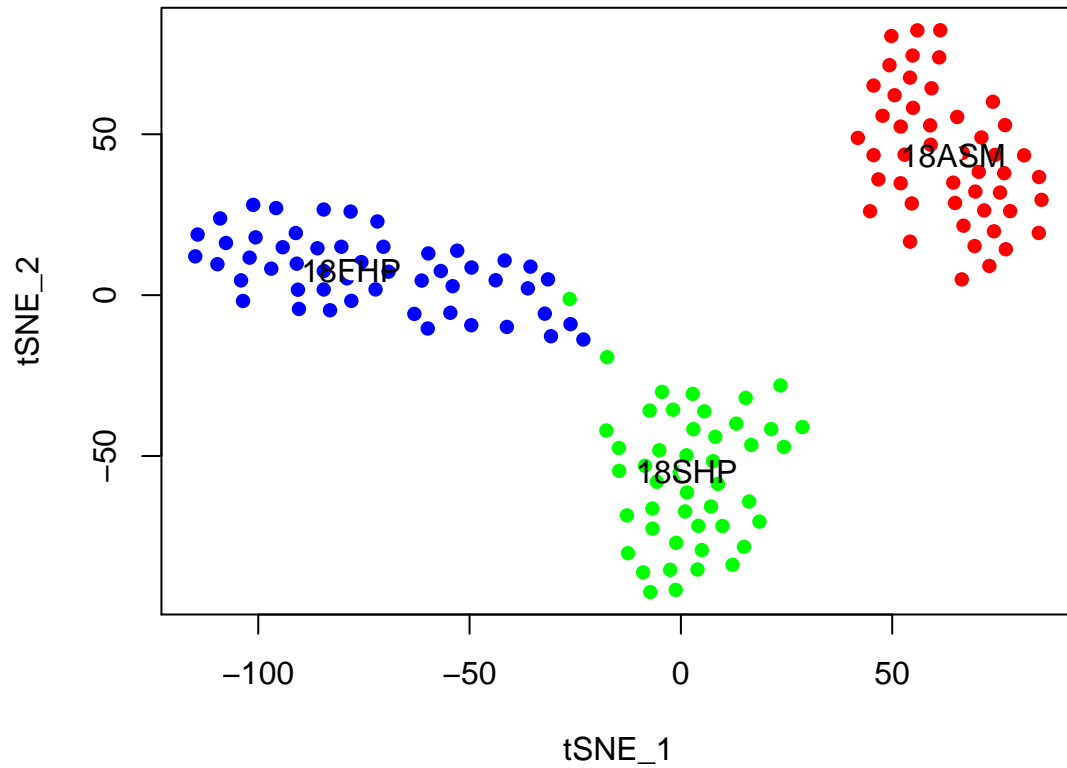
-0.15



*# There are 14 out of 33 cells (42%) have good expression level(2 logFPKM)
of both genes. Thus, upon reasoning above, we conclude that this cluster
contains cells that contradict to experimental discoveries which has a
high probability to be doublet cells due to technical error.*

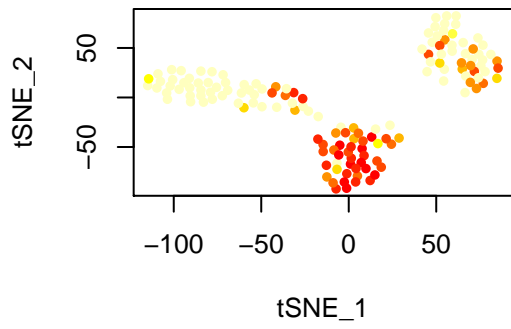
Filter the cells based on reasoning above

```
hpf18.new = subsetData(hpf18, which.cells(hpf18, c("18ASM", "18SHP", "18FHP")),  
  do.scale = F)  
tsne.plot(hpf18.new, do.label = T, cols.use = c("red", "orange", "blue"))
```

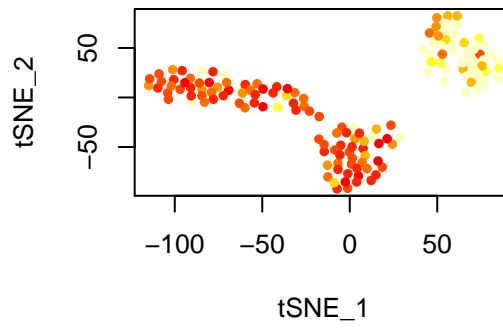


```
# Visualize markers of different clusters using violin plot and feature plot
hpf18.new@ident = factor(hpf18.new@ident, ordered = T, levels = c("18FHP", "18SHP",
  "18ASM"))
genes.viz.18 = c("DACH1/2", "NKX2-3", "EBF1/2/3/4", "SFRP1/5")
feature.plot(hpf18.new, genes.viz.18, pt.size = 0.8)
```

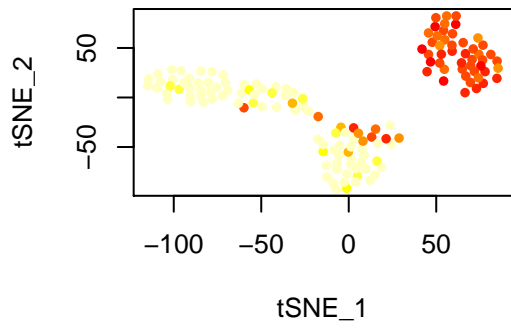
DACH1/2



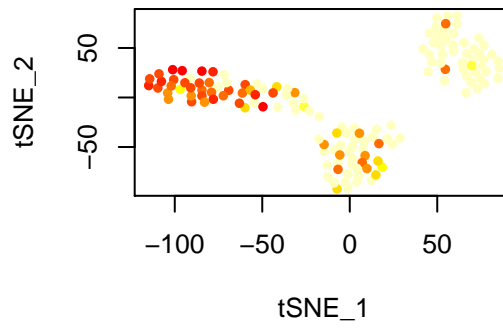
NKX2-3



EBF1/2/3/4

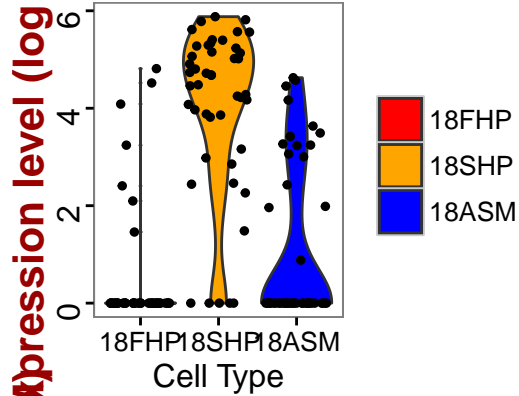


SFRP1/5

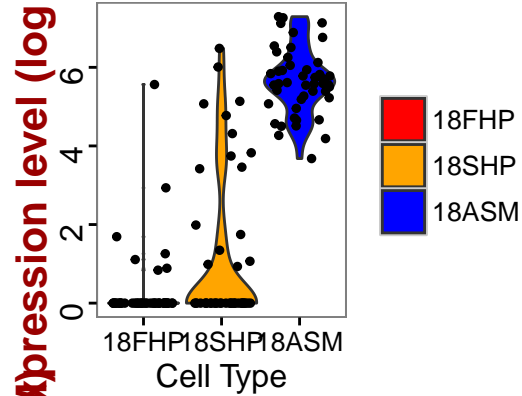


```
vlnPlot(hpf18.new, genes.viz.18, cols.use = c("red", "orange", "blue"))
```

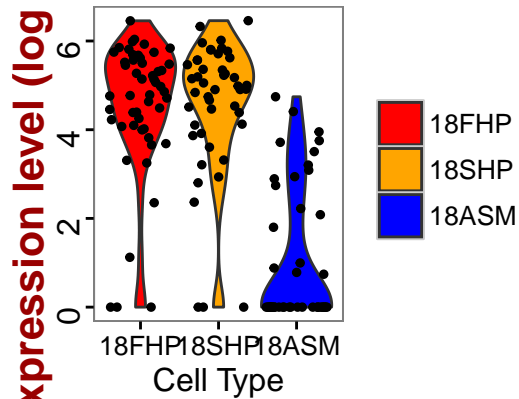
DACH1/2



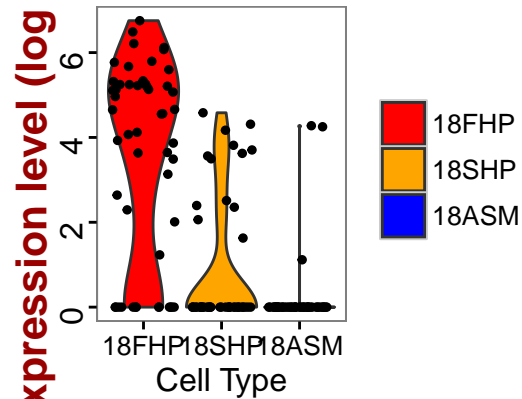
EBF1/2/3/4



NKX2-3



SFRP1/5



```
# Select markers for plotting on a Heatmap (top 10 positive markers with  
# high classification power(myAUC))
```

```
marker18FHP = rownames(FHP_18.markers[order(FHP_18.markers$myAUC, decreasing = T)[1:15],  
])
```

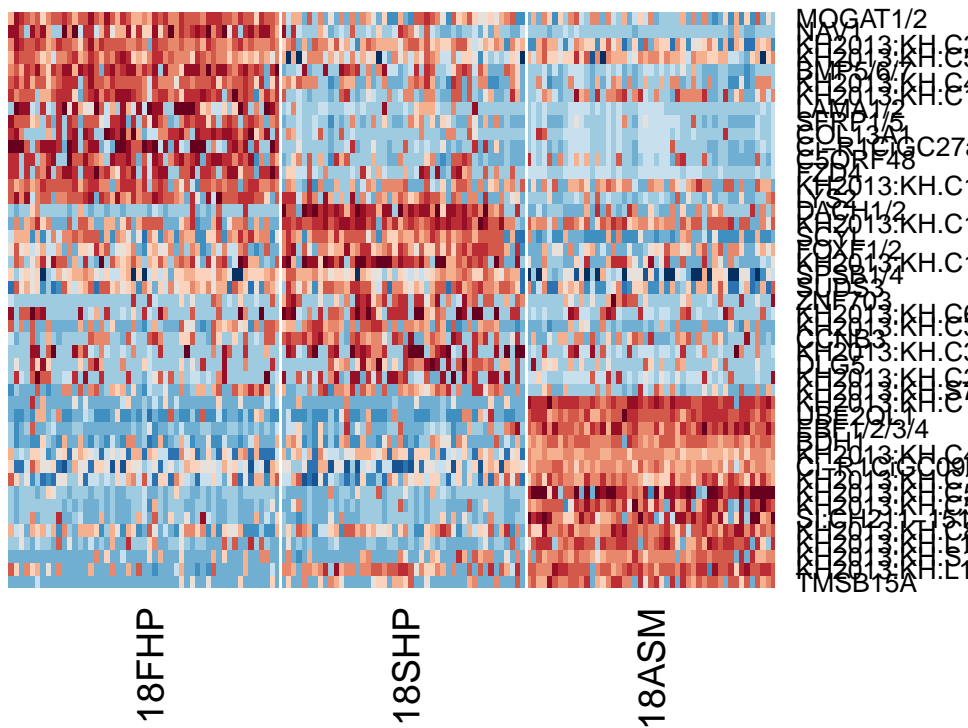
```
marker18SHP = rownames(SHP_18.markers[order(SHP_18.markers$myAUC, decreasing = T)[1:15],  
])
```

```
marker18ASM = rownames(ASM_18.markers[order(ASM_18.markers$myAUC, decreasing = T)[1:15],  
])
```

```
marker.18 = c(marker18FHP, marker18SHP, marker18ASM)
```

```
# Draw a heatmap of all cells for these marker genes
```

```
doHeatMap(hpf18.new, genes.use = marker.18, remove.key = TRUE, slim.col.label = T,  
cex.col = 1.2, col.use = col)
```



6. hpf16

```
# Subset data from preprocessed Seurat object
hpf16 = subsetData(hpfall.remv2, which.cells(hpfall.remv2, "hpf16"), do.scale = F)
hpf16
```

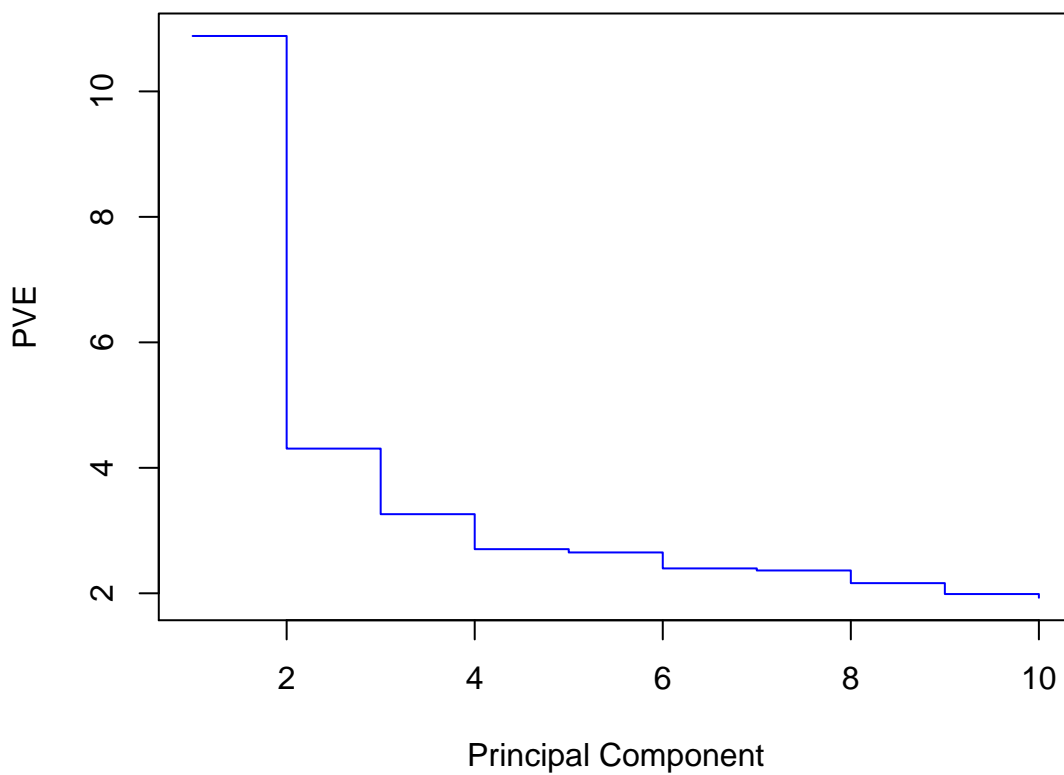
```
## An object of class seurat in project allhpf
## 14864 genes across 114 samples.
```

```
# Based on hpf12 hpf14 hpf18 and hpf20 data, we have successfully uncovered
# all the TVC lineage types: TVC, STVC, FHP, SHP, ASM Based on preliminary
# studies, hpf16 contains three TVC lineage cell types: FHP, late STVC,
# early SHP and early ASM which are divided from early STVC. Therefore hpf16
# is an intermediate stage between hpf14 and hpf18, we can run PCA with both
# hpf14 (14STVC,14FHP) and hpf18 cell markers (18FHP, 18SHP, 18ASM).
# Power>0.4 is used to obtain more markers from hpf14 cells.
marker20FHP.use = rownames(subset(FHP_20.markers, power > 0.5 & avg_diff > 0))
marker20SHP.use = rownames(subset(SHP_20.markers, power > 0.5 & avg_diff > 0))
marker20ASM1.use = rownames(subset(ASM1_20.markers, power > 0.5 & avg_diff >
0))
marker20ASM2.use = rownames(subset(ASM2_20.markers, power > 0.5 & avg_diff >
0))
marker14FHP.use = rownames(subset(FHP_14.markers, power > 0.4 & avg_diff > 0))
marker14STVC.use = rownames(subset(STVC_14.markers, power > 0.4 & avg_diff >
```

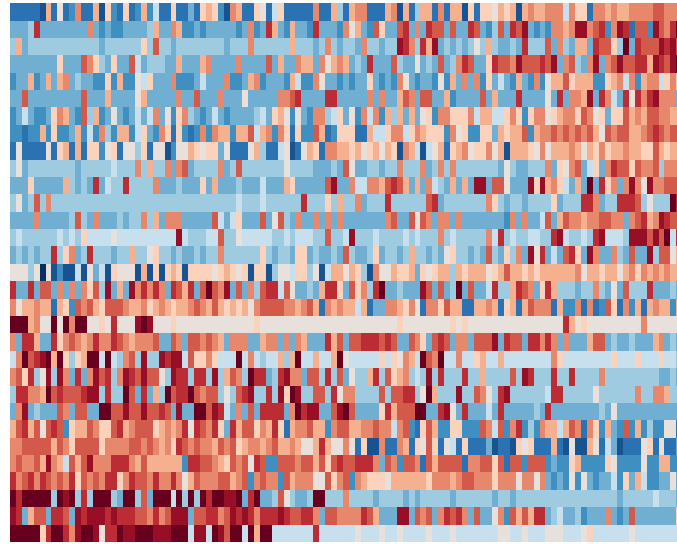
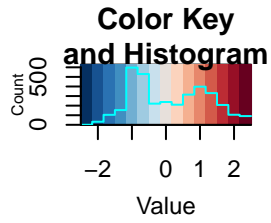
```
0))
marker.16.use = unique(c(marker20FHP.use, marker20SHP.use, marker20ASM1.use,
  marker20ASM2.use, marker14FHP.use, marker14STVC.use))
length(marker.16.use)
```

```
## [1] 217
```

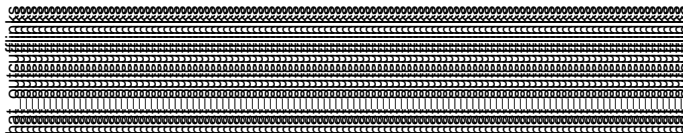
```
# Run a PCA using marker list
hpf16 = pca(hpf16, pc.genes = marker.16.use, do.print = F)
pcScree(hpf16, marker.16.use, 10)
```



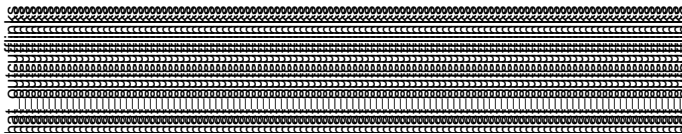
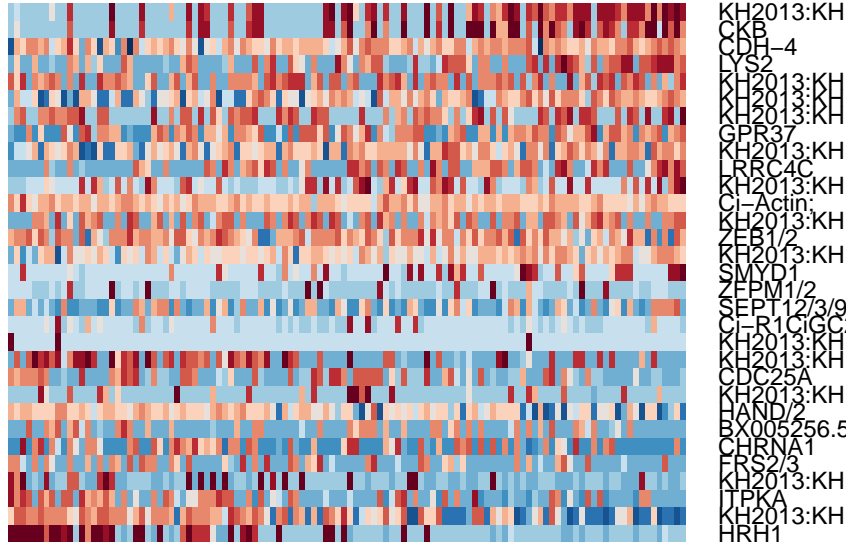
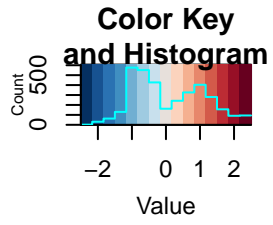
```
pcHeatmap(hpf16, 1, do.balanced = T, col.use = col)
```

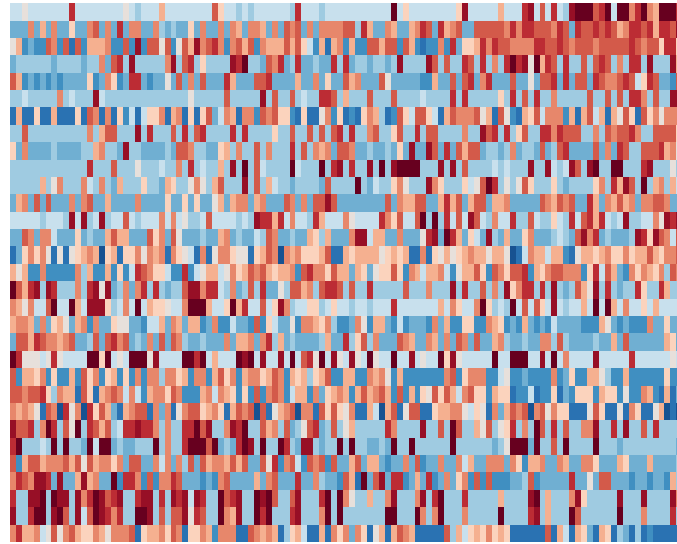
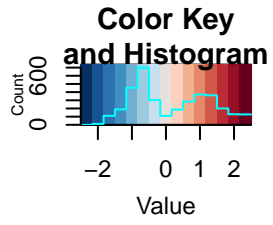
NKX2-3
SFRP2
COL13A1
LGS2
SEPT12/3/9
WNT9B
SLIT1/2/3
MOGAT1/2
SCY1
SFRP1/5
LRP4/8
ITFNB1/2/3
NLI2013:KH
DMP5/6/7
DDR1/2
CK1
KH2013:KH
KH2013:KH
S:CH211-1
KH2013:KH
IRX1/2/3/5
ELK1/3/4
IRX4/6
KH2013:KH
NOVA1/2
KH2013:KH
FOXF1/2
TMSB15A
HRH1
TBX1/10
EBF1/2/3/4



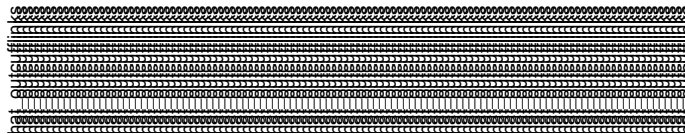
```
pcHeatmap(hpf16, 2, do.balanced = T, col.use = col)
```



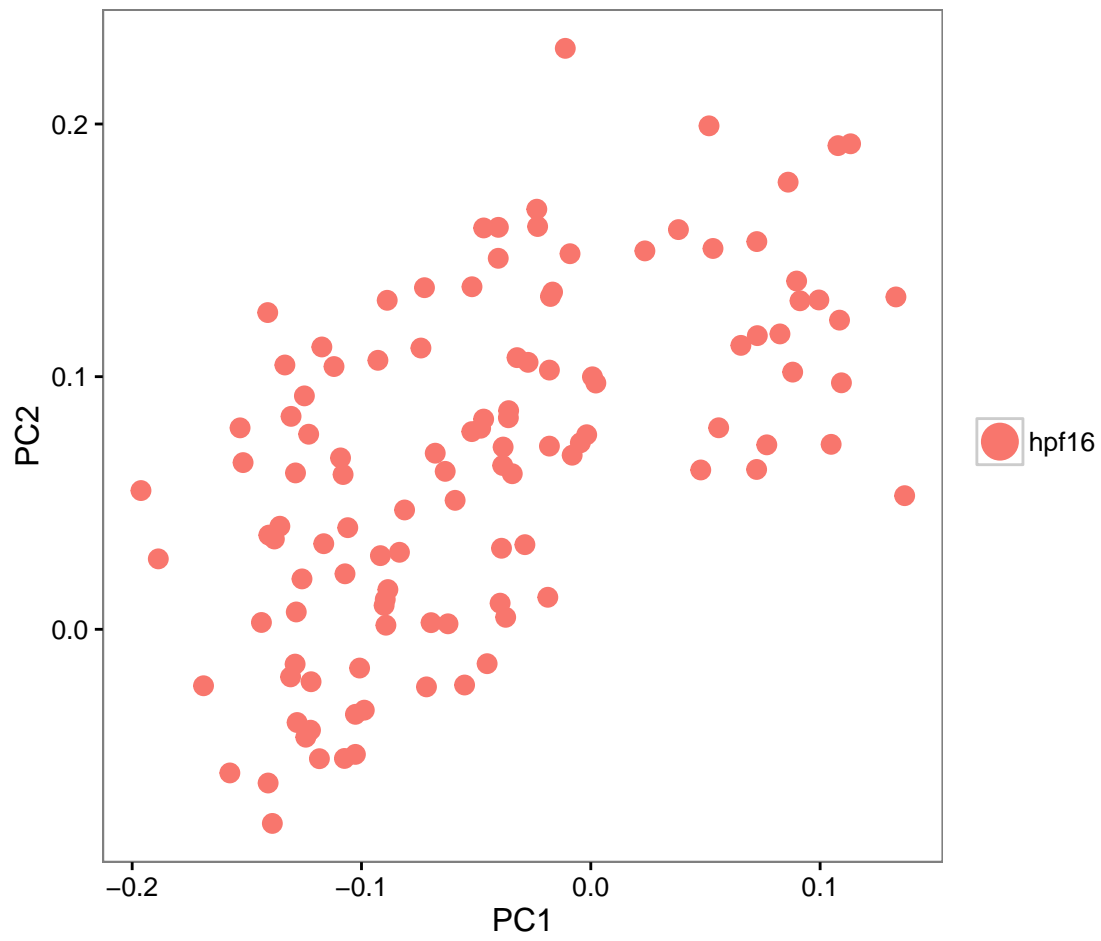
```
pcHeatmap(hpf16, 3, do.balanced = T, col.use = col)
```



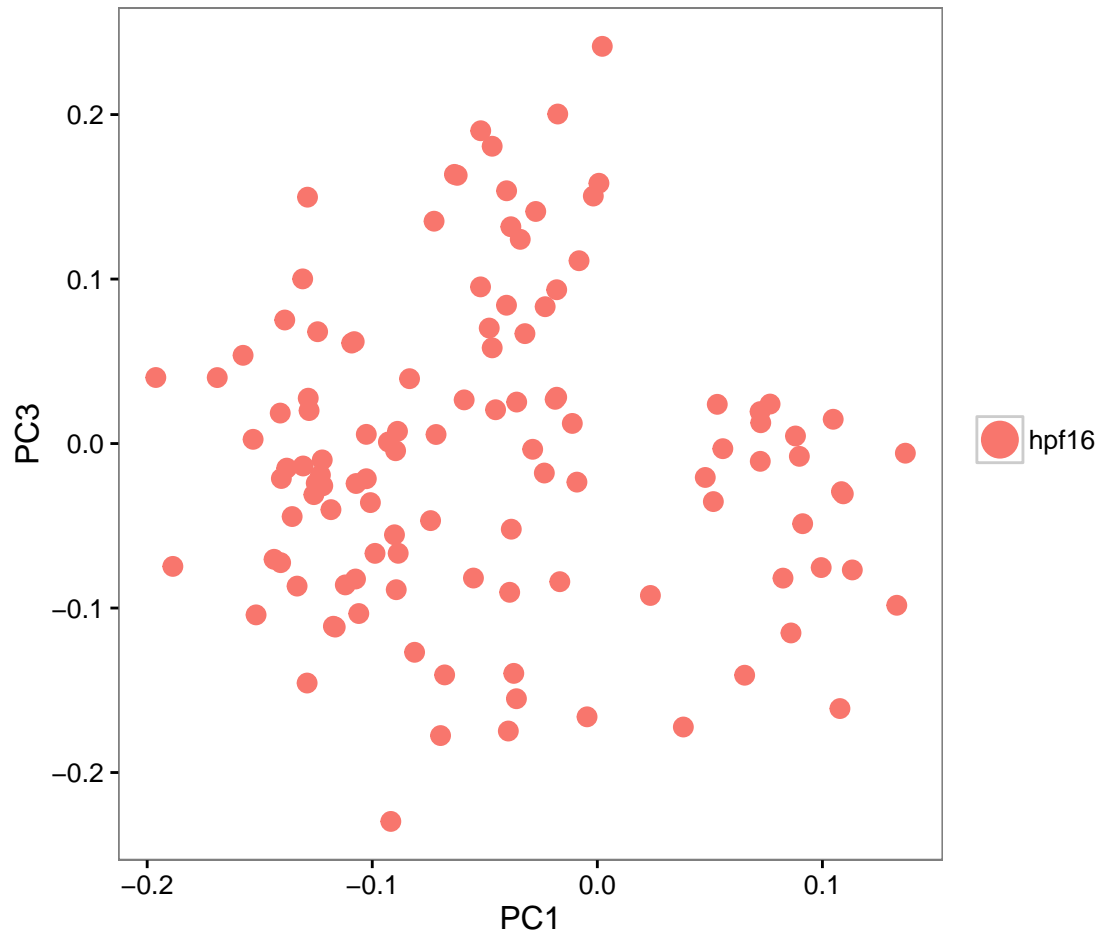
DACH1/2
 KL2013:KH
 TOX1/2
 STXF1/2
 SLC38A7
 NHS
 OTUD3
 TOXG1
 KH2013:KH
 CISH
 KH2013:KH
 CCDC66
 NK637-14
 NL2013:KH
 PPA8
 XH2013:KH
 C1-R1CiGC
 FLK1/3/4
 TRX1/2/3/5
 SEPT12/3/9
 C5ORF48
 TIBF1/2/3/4
 XH2013:KH
 VOGAT1/2
 KL2013:KH
 TRX4/6
 LIP4
 LIP1
 XH2013:KH
 SFRP2
 KH2013:KH
 CKB
 TNS1/3



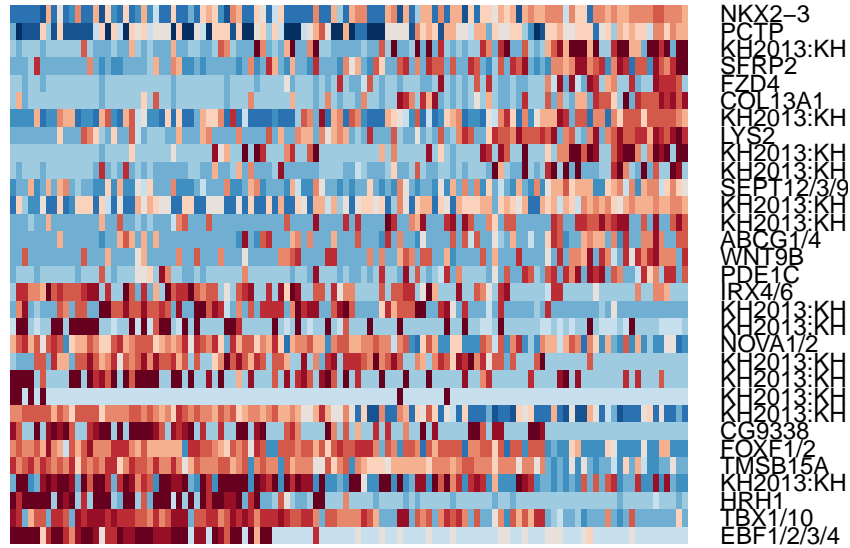
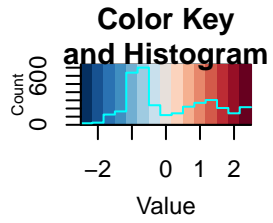
`pca.plot(hpf16, 1, 2)`



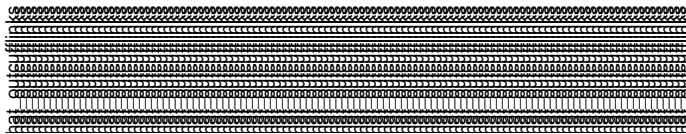
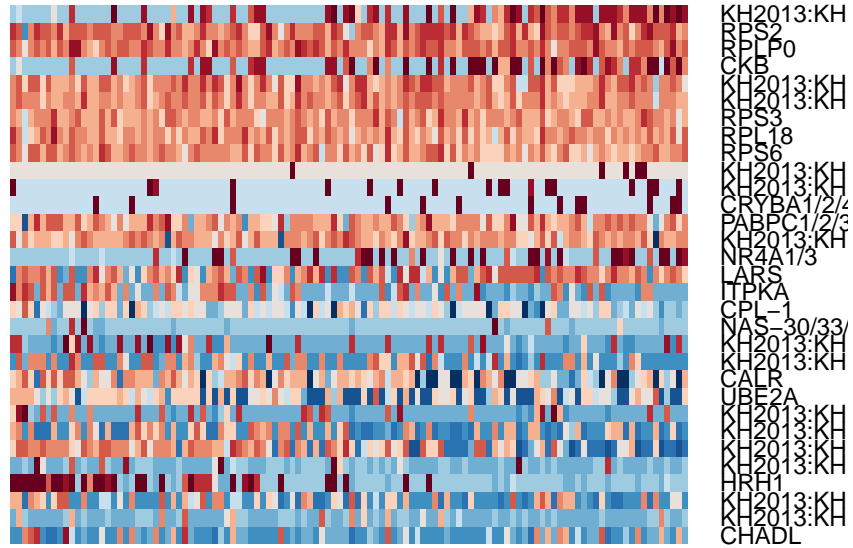
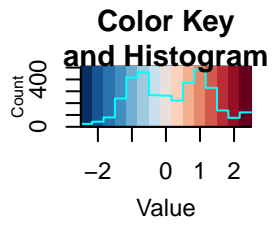
```
pca.plot(hpf16, 1, 3)
```



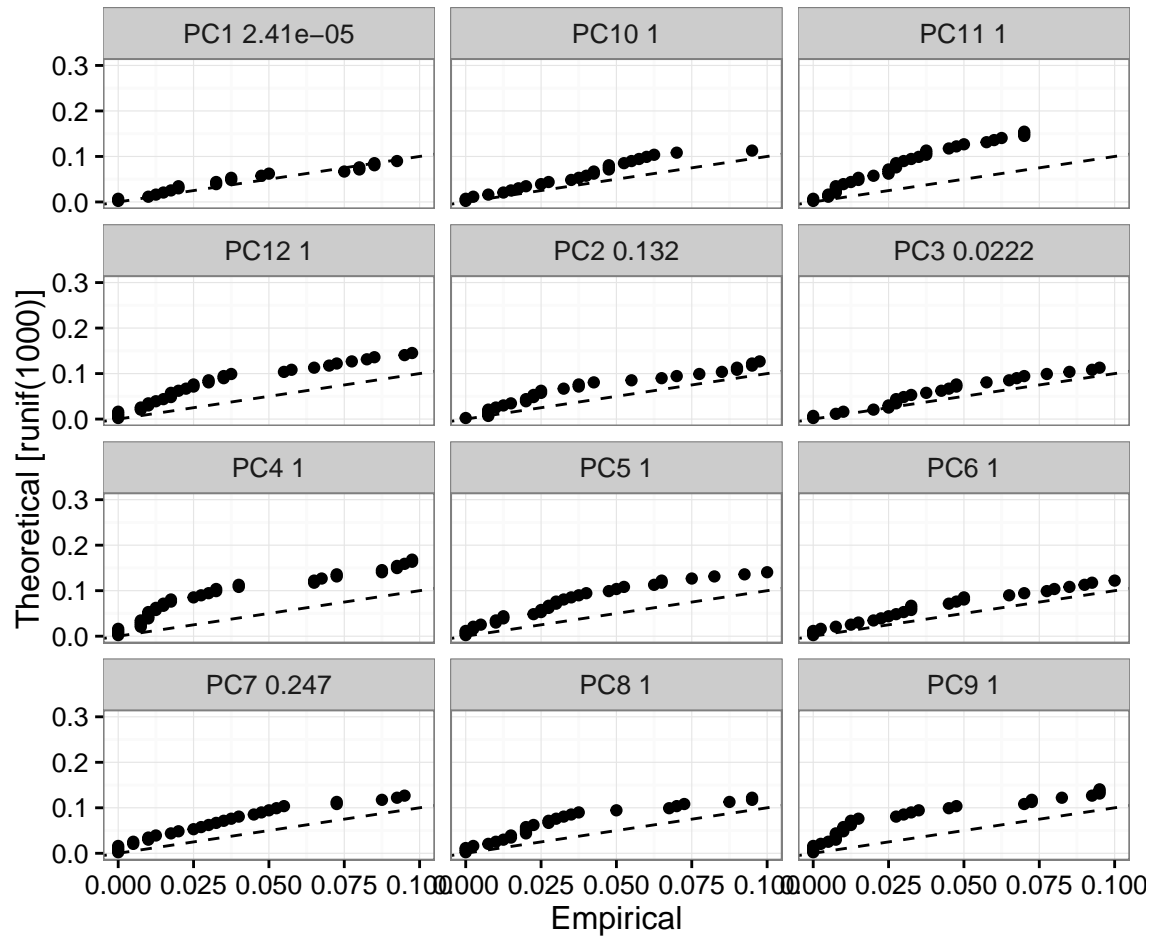
```
# Calculate PCA scores for all genes (PCA projection)  
hpf16 = project.pca(hpf16, do.print = F)  
  
# Visualize the full projected PCA, which now includes new genes which were  
# not previously (use.full=TRUE)  
pcHeatmap(hpf16, 1, use.full = T, do.balanced = T, col.use = col)
```



```
pcHeatmap(hpf16, 2, use.full = T, do.balanced = T, col.use = col) #technical
```



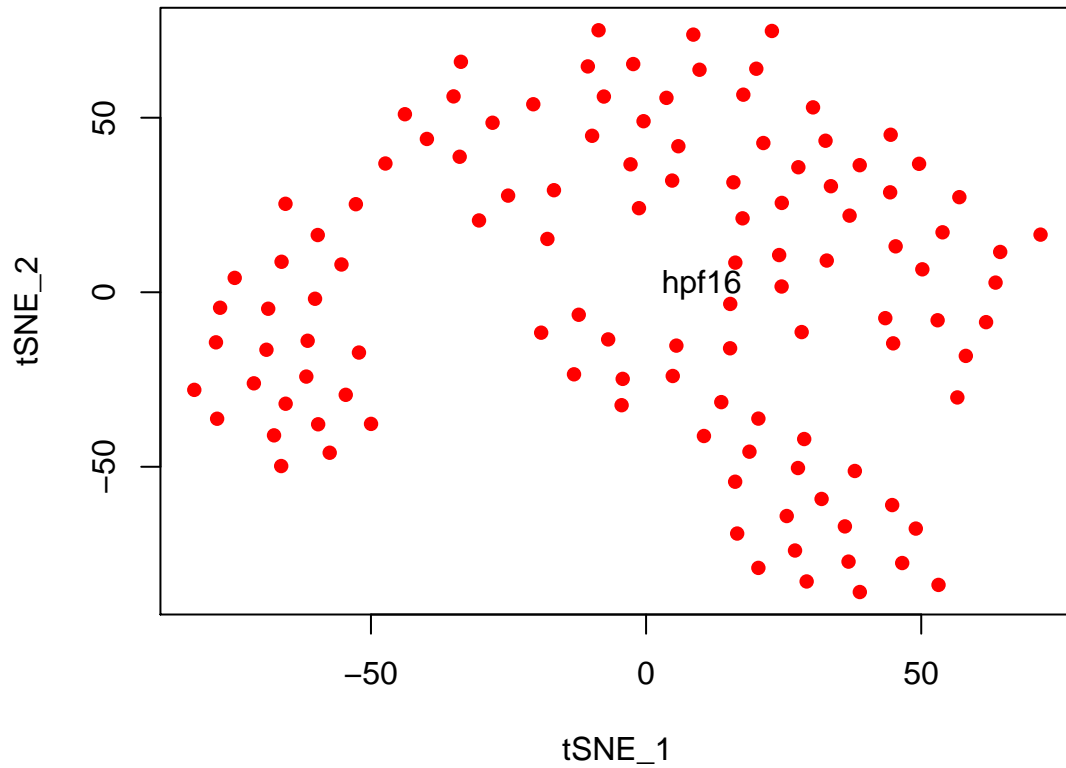
```
pcHeatmap(hpf16, 3, use.full = T, do.balanced = T, col.use = col)
```

In this case PC1 and PC3 are significant

*# Run tSNE using significant PCs as input (spectral tSNE), we get distinct
point clouds*

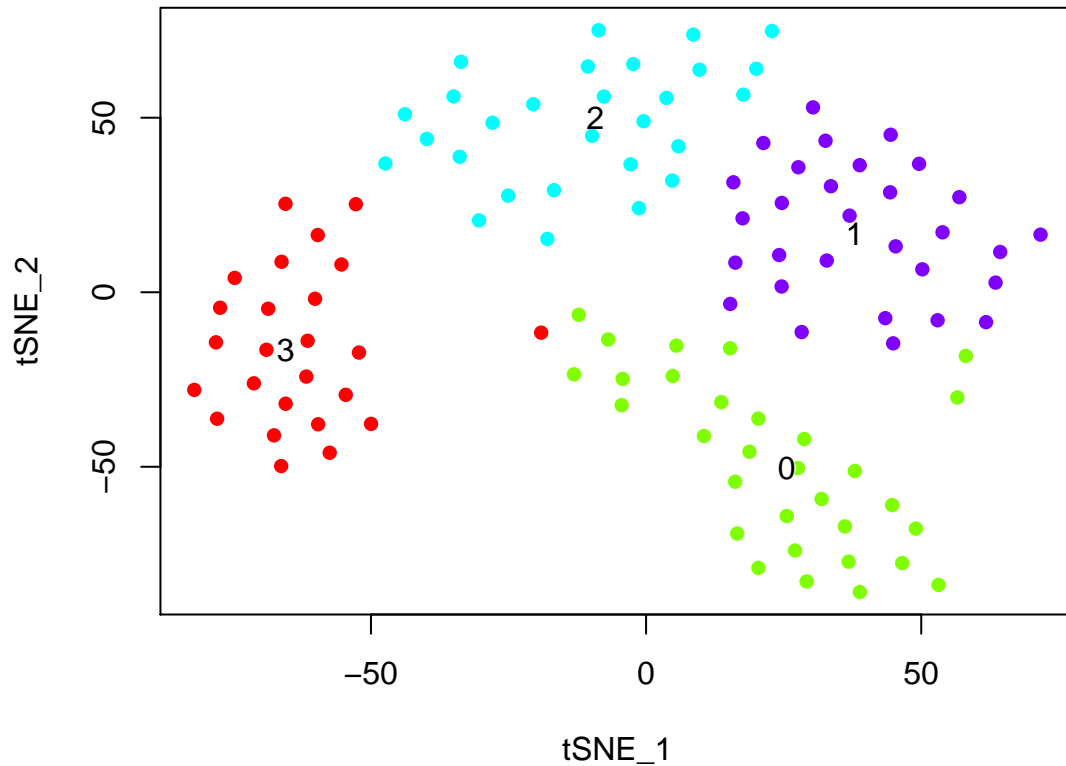
```
hpf16 = run_tsne(hpf16, max_iter = 2000, dims.use = c(1, 3))
tsne.plot(hpf16, do.label = T, label.pt.size = 1)
```



```
# Find cell clusters using Modularity optimization cluster detection. hpf16
# dataset contain fewer cells than the others a k.param=10.
hpf16 = FindClusters(hpf16, pc.use = c(1, 3), do.modularity = T, resolution = 1,
  prune.SNN = 0.1, print.output = 0, k.param = 20, k.scale = floor(114/20))
```

```
## [1] "SNN : processed 28 cells"
## [1] "SNN : processed 57 cells"
## [1] "SNN : processed 86 cells"
## [1] "SNN : processed 114 cells"
```

```
tsne.plot(hpf16, do.label = T, label.pt.size = 1)
```



```
# The validity of the clusters can be validated using a classification
# scheme based on linear SVMs.
```

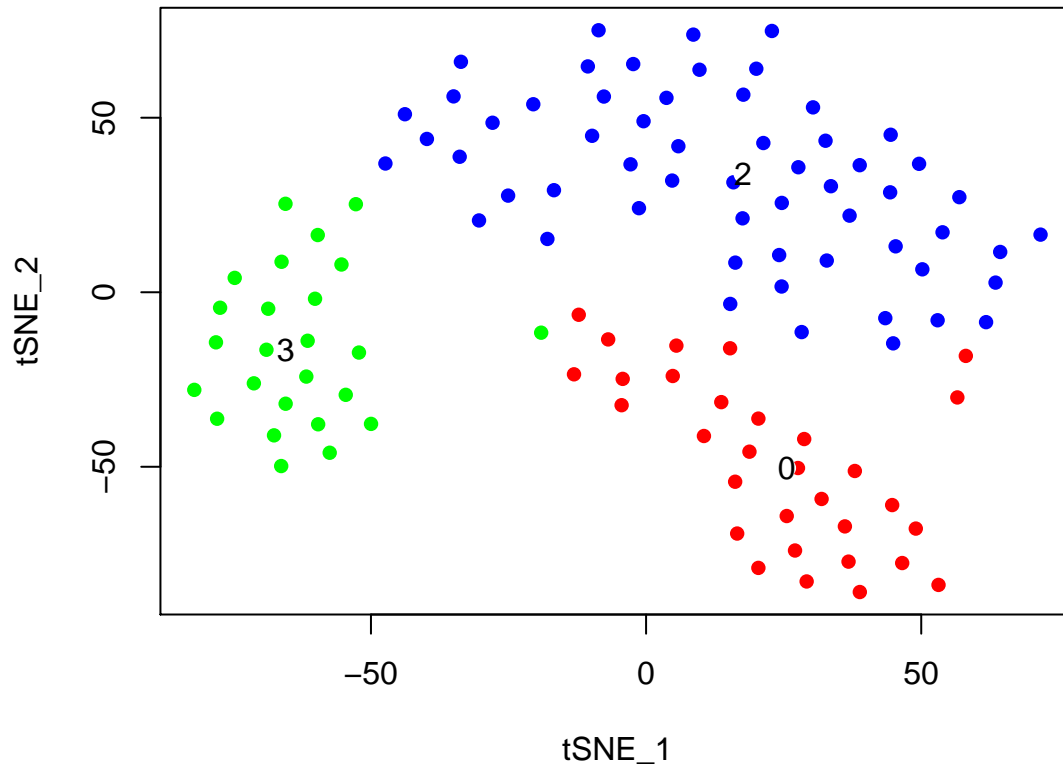
```
hpf16 = BuildSNN(hpf16, pc.use = c(1, 3), do.sparse = T, k.param = 20, k.scale = floor(1
```

```
## [1] "SNN : processed 28 cells"
## [1] "SNN : processed 57 cells"
## [1] "SNN : processed 86 cells"
## [1] "SNN : processed 114 cells"
```

```
hpf16 = ValidateClusters(hpf16, pc.use = c(1, 3), min.connectivity = 0.001,
  acc.cutoff = 0.85)
```

```
## [1] " 0% complete --- merge clusters 1 and 2, classification accuracy of 0.7733"
## [1] "100% complete --- started with 4 clusters, 3 clusters remaining"
```

```
tsne.plot(hpf16, do.label = T, label.pt.size = 1)
```



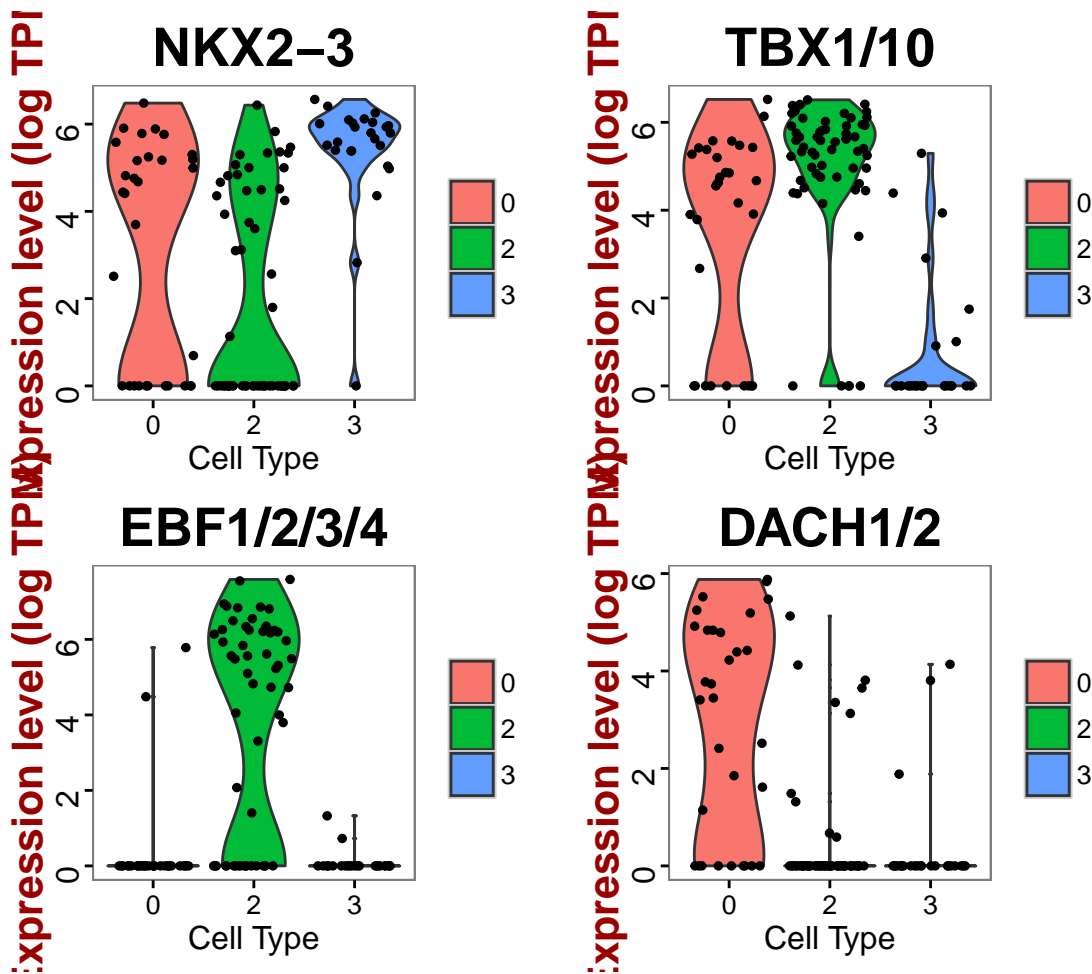
```
# Find cluster markers using ROC test with thresh.use = 1, min.pct = 0.5 The
# ROC test returns the 'classification power' for any individual marker
# (ranging from 0 - random, to 1 - perfect). Though not a statistical test,
# it is often very useful for finding clean markers. Find markers for
# cluster 0
```

```
cl0_16.markers = find.markers(hpf16, 0, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(cl0_16.markers[order(cl0_16.markers$myAUC, decreasing = T), ], 20)
```

##	myAUC	avg_diff	power	pct.1	pct.2
## KH2013:KH.C1.638_C17ORF105	0.812	1.121174	0.624	0.935	0.578
## DACH1/2	0.803	2.475778	0.606	0.710	0.133
## KH2013:KH.C9.692_GABRR1/2/3	0.740	1.120309	0.480	0.839	0.518
## KH2013:KH.C3.616_GNRHR/1/2/4	0.722	1.529193	0.444	0.613	0.193
## SFR1	0.704	1.048728	0.408	0.710	0.434
## ORC1	0.701	1.215804	0.402	0.645	0.289
## GM1840	0.698	1.469439	0.396	0.613	0.265
## KH2013:KH.C10.172	0.691	1.726626	0.382	0.548	0.193
## ZGC:174877	0.685	1.114684	0.370	0.645	0.349
## IKBKG	0.684	1.433664	0.368	0.581	0.301
## KH2013:KH.L150.4	0.661	1.021943	0.322	0.581	0.349
## PPT2	0.654	1.024035	0.308	0.613	0.349
## USP48	0.649	1.120253	0.298	0.581	0.458
## KH2013:KH.C6.155_FLRT1/2/3	0.621	1.114470	0.242	0.581	0.410

```
## RBM15B 0.581 -1.461119 0.162 0.774 0.590
## KH2013:KH.L124.2 0.526 1.806418 0.052 0.806 0.916
## ZNRD1 0.507 -3.447701 0.014 0.516 0.494
## KH2013:KH.L39.6 0.473 -4.277293 0.054 0.452 0.542
## SMEK1/2 0.461 -1.636758 0.078 0.613 0.602
## KH2013:KH.C8.649_ACTA1/2 0.390 -2.190225 0.220 0.355 0.590
```

```
# Visualize known markers with a violin plot
vlnPlot(hpf16, c("NKX2-3", "EBF1/2/3/4", "TBX1/10", "DACH1/2"))
```



```
# Based on preliminary studies these NKX2-3+ EBF1/2/3/4- DACH1/2+ TBX1/10+
# cells are SHPs
```

```
# Find markers for cluster 2
c12_16.markers = find.markers(hpf16, 2, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(c12_16.markers[order(c12_16.markers$myAUC, decreasing = T), ], 20)
```

```
## myAUC avg_diff power pct.1 pct.2
```

```

## KH2013:KH.S555.1_HTR7 0.939 2.307248 0.878 0.983 0.464
## KH2013:KH.C1.21_CG2781 0.852 1.103323 0.704 1.000 0.946
## TBX1/10 0.820 1.194245 0.640 0.914 0.500
## EBF1/2/3/4 0.817 3.644445 0.634 0.672 0.054
## CDC6 0.816 1.009440 0.632 1.000 0.929
## HAND/2 0.810 1.079622 0.620 0.966 0.786
## ZCCHC24 0.797 1.156534 0.594 0.931 0.536
## HRH1 0.796 3.209113 0.592 0.603 0.018
## SMURF1/2 0.787 1.502168 0.574 0.828 0.536
## TMSB15A 0.784 1.133905 0.568 0.966 0.679
## ELK1/3/4 0.749 1.134942 0.498 0.741 0.286
## IRX4/6 0.735 1.253873 0.470 0.759 0.357
## KH2013:KH.C4.404 0.733 1.737132 0.466 0.690 0.304
## KH2013:KH.L134.29 0.724 1.222385 0.448 0.741 0.446
## IRX1/2/3/5 0.705 1.109414 0.410 0.569 0.214
## NOX5 0.696 1.016322 0.392 0.638 0.286
## KIAA0513 0.693 1.031799 0.386 0.759 0.500
## KH2013:KH.C9.223_CDC42 0.689 1.056894 0.378 0.914 0.821
## ITPKA 0.666 1.211642 0.332 0.586 0.339
## KH2013:KH.C12.239 0.647 1.110297 0.294 0.534 0.304

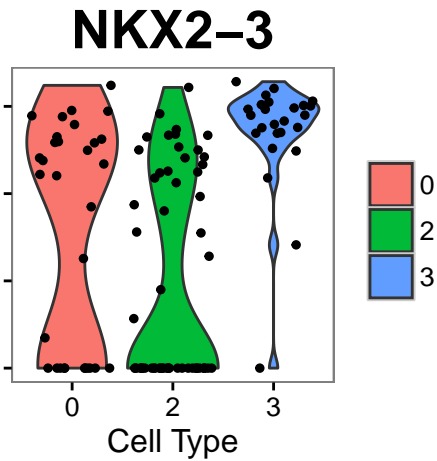
```

```

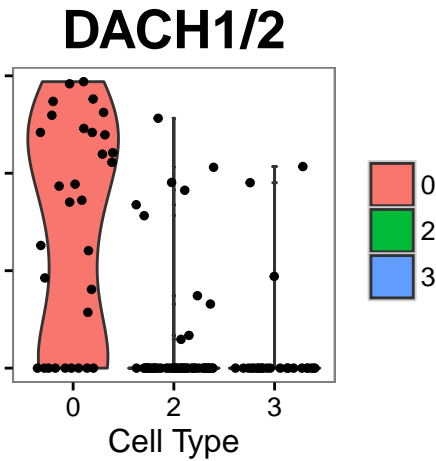
# Visualize known markers with a violin plot
vlnPlot(hpf16, c("NKX2-3", "EBF1/2/3/4", "DACH1/2", "TBX1/10"))

```

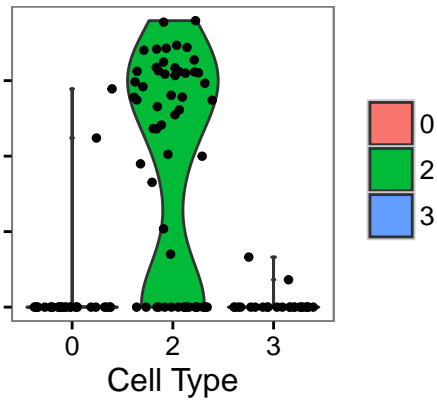
expression level (log TPM)



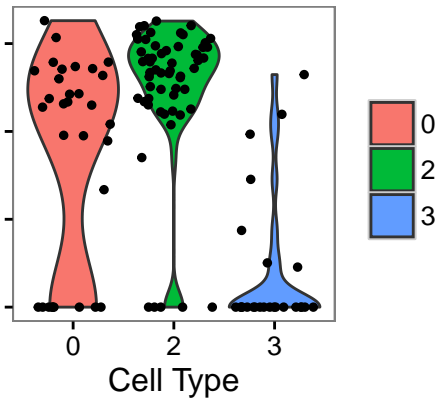
expression level (log TPM)



EBF1/2/3/4



TBX1/10



```
# Based on preliminary studies these NKX2-3- EBF1/2/3/4+ DACH1/2+ TBX1/10+
# cells are ASM
```

```
# Find markers for cluster 3
```

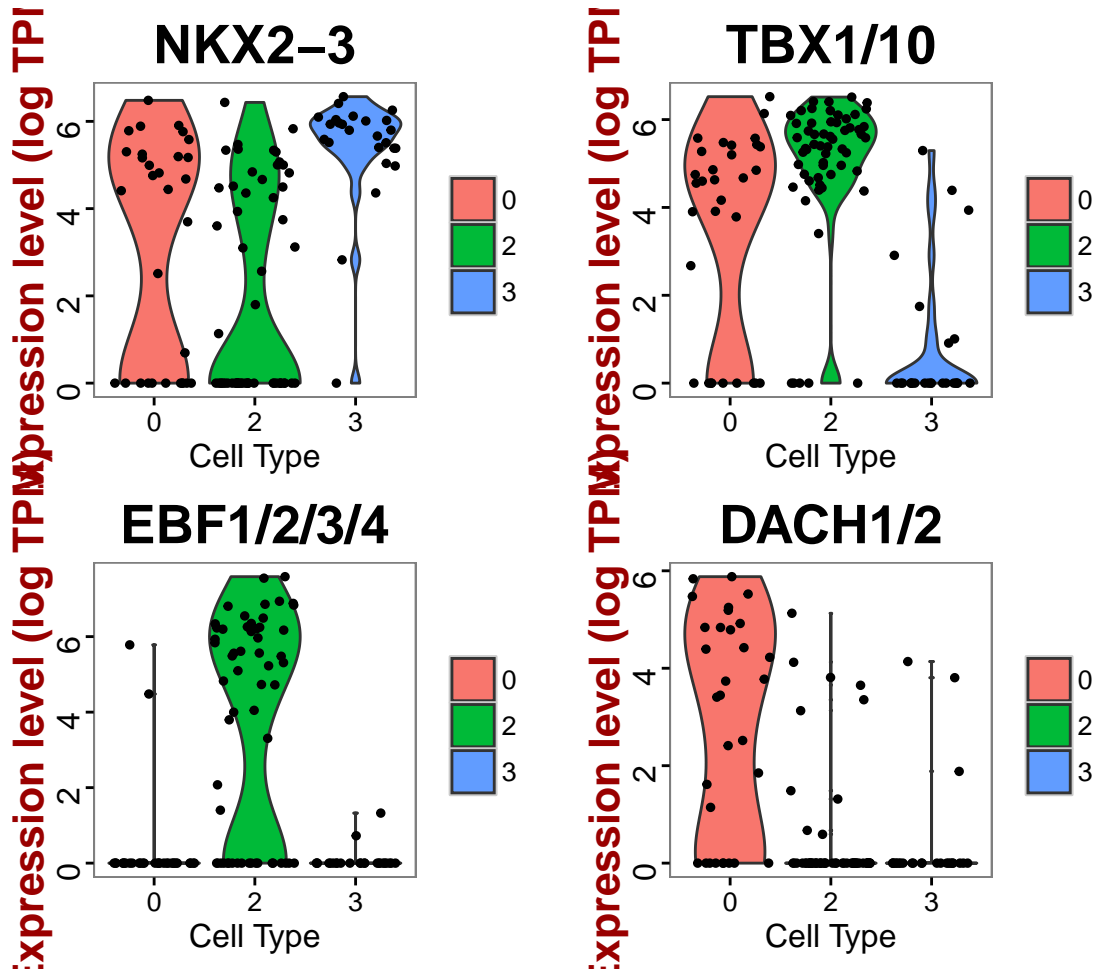
```
c13_16.markers = find.markers(hpf16, 3, thresh.use = 1, test.use = "roc", min.pct = 0.5)
head(c13_16.markers[order(c13_16.markers$myAUC, decreasing = T), ], 20)
```

```
## myAUC
## KH2013:KH.C5.227_F56C4.4 0.950
## SMTN 0.931
## MOGAT1/2 0.919
## NKX2-3 0.879
## KH2013:KH.C9.40_GST01/2 0.873
## SI:DKEY-79F11.5 0.852
## KH2013:KH.C9.770_RGR 0.851
## SCYL 0.831
## KH2013:KH.C8.489_SI:DKEY-29D8.3 0.830
## KH2013:KH.C4.547_BMP2/4 0.829
```

## KH2013:KH.C9.247_BX119910.1/10/11/12/13/14/16/2/3/4/5/6/7/8/9	0.828		
## CDH-4	0.827		
## SFRP1/5	0.826		
## TIMP4	0.820		
## KH2013:KH.C2.22	0.818		
## SFRP2	0.818		
## SLIT1/2/3	0.816		
## LYS2	0.816		
## KH2013:KH.C4.376_UNC93A	0.806		
## COL13A1	0.792		
##	avg_diff		
## KH2013:KH.C5.227_F56C4.4	1.269902		
## SMTN	1.785546		
## MOGAT1/2	1.568086		
## NKX2-3	1.329046		
## KH2013:KH.C9.40_GST01/2	1.365326		
## SI:DKEY-79F11.5	1.537301		
## KH2013:KH.C9.770_RGR	1.117639		
## SCYL	1.030414		
## KH2013:KH.C8.489_SI:DKEY-29D8.3	1.243672		
## KH2013:KH.C4.547_BMP2/4	1.630799		
## KH2013:KH.C9.247_BX119910.1/10/11/12/13/14/16/2/3/4/5/6/7/8/9	1.348407		
## CDH-4	1.034349		
## SFRP1/5	1.934746		
## TIMP4	1.211017		
## KH2013:KH.C2.22	1.290851		
## SFRP2	1.820400		
## SLIT1/2/3	1.306153		
## LYS2	1.452199		
## KH2013:KH.C4.376_UNC93A	1.210027		
## COL13A1	2.009872		
##	power	pct.1	
## KH2013:KH.C5.227_F56C4.4	0.900	1.00	
## SMTN	0.862	1.00	
## MOGAT1/2	0.838	1.00	
## NKX2-3	0.758	0.96	
## KH2013:KH.C9.40_GST01/2	0.746	1.00	
## SI:DKEY-79F11.5	0.704	0.92	
## KH2013:KH.C9.770_RGR	0.702	1.00	
## SCYL	0.662	1.00	
## KH2013:KH.C8.489_SI:DKEY-29D8.3	0.660	0.96	
## KH2013:KH.C4.547_BMP2/4	0.658	0.88	
## KH2013:KH.C9.247_BX119910.1/10/11/12/13/14/16/2/3/4/5/6/7/8/9	0.656	0.92	
## CDH-4	0.654	1.00	
## SFRP1/5	0.652	0.80	

## TIMP4	0.640	0.92
## KH2013:KH.C2.22	0.636	0.92
## SFRP2	0.636	0.84
## SLIT1/2/3	0.632	0.96
## LYS2	0.632	0.84
## KH2013:KH.C4.376_UNC93A	0.612	0.72
## COL13A1	0.584	0.72
##	pct.2	
## KH2013:KH.C5.227_F56C4.4	1.000	
## SMTN	0.438	
## MOGAT1/2	0.618	
## NKX2-3	0.506	
## KH2013:KH.C9.40_GST01/2	0.652	
## SI:DKEY-79F11.5	0.685	
## KH2013:KH.C9.770_RGR	0.708	
## SCYL	0.742	
## KH2013:KH.C8.489_SI:DKEY-29D8.3	0.742	
## KH2013:KH.C4.547_BMP2/4	0.337	
## KH2013:KH.C9.247_BX119910.1/10/11/12/13/14/16/2/3/4/5/6/7/8/9	0.494	
## CDH-4	0.944	
## SFRP1/5	0.191	
## TIMP4	0.438	
## KH2013:KH.C2.22	0.438	
## SFRP2	0.315	
## SLIT1/2/3	0.584	
## LYS2	0.404	
## KH2013:KH.C4.376_UNC93A	0.236	
## COL13A1	0.180	

```
# Visualize known markers with a violin plot
vlnPlot(hpf16, c("NKX2-3", "EBF1/2/3/4", "TBX1/10", "DACH1/2"))
```



```
# Based known markers, this group is FHP
```

```
# Write cell names into text files
```

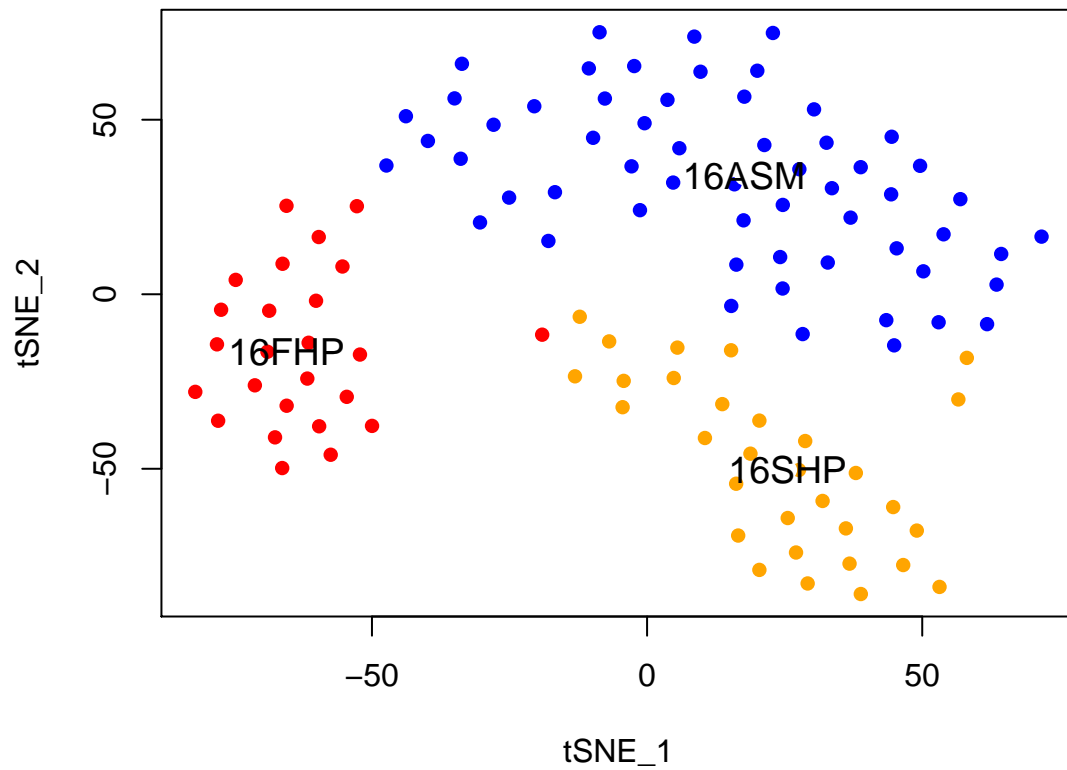
```
write.table(which.cells(hpf16, 0), file = "16SHPCells.txt", sep = "\t")
write.table(which.cells(hpf16, 2), file = "16ASMCells.txt", sep = "\t")
write.table(which.cells(hpf16, 3), file = "16FHPCells.txt", sep = "\t")
```

```
# Rename cluster identities
```

```
hpf16 = rename.ident(hpf16, 0, "16SHP")
hpf16 = rename.ident(hpf16, 2, "16ASM")
hpf16 = rename.ident(hpf16, 3, "16FHP")
```

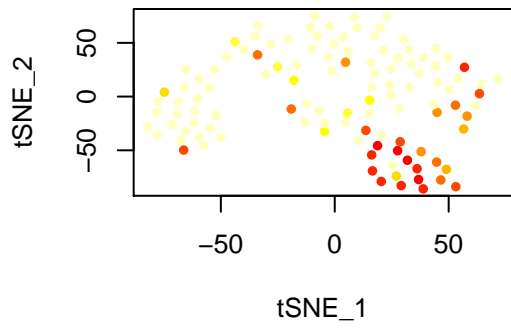
```
# Visualize tSNE used color scheme
```

```
tsne.plot(hpf16, do.label = T, label.pt.size = 1, label.cex.text = 1.2, label.cols.use =
  "red", "blue"))
```

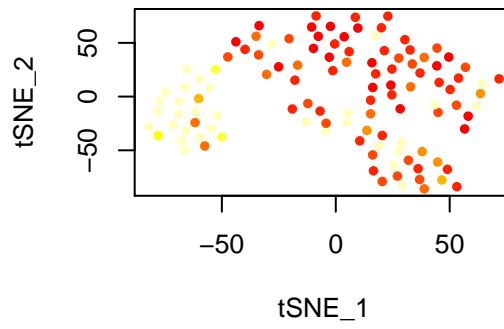


```
# Visualize markers of different clusters using violin plot and feature plot
hpf16@ident = factor(hpf16@ident, ordered = T, levels = c("16FHP", "16SHP",
  "16ASM"))
genes.viz.16 = c("DACH1/2", "TBX1/10", "NKX2-3", "EBF1/2/3/4")
feature.plot(hpf16, genes.viz.16, pt.size = 0.8)
```

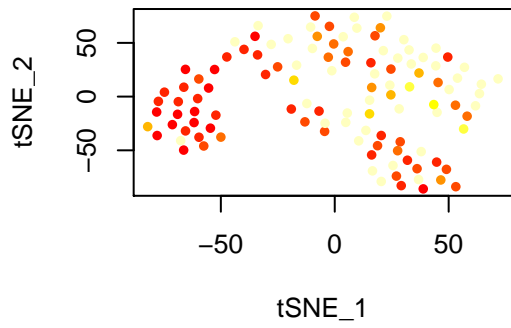
DACH1/2



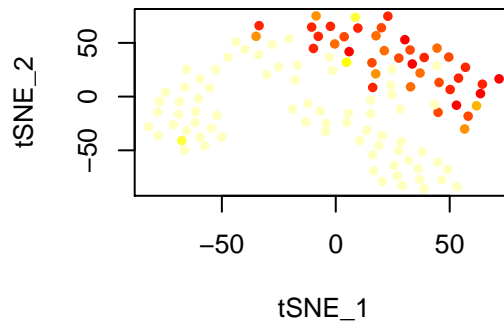
TBX1/10



NKX2-3

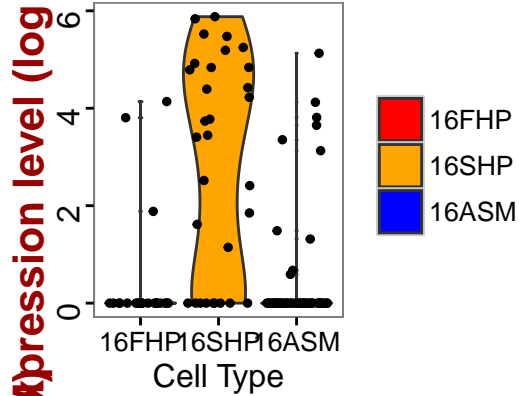


EBF1/2/3/4

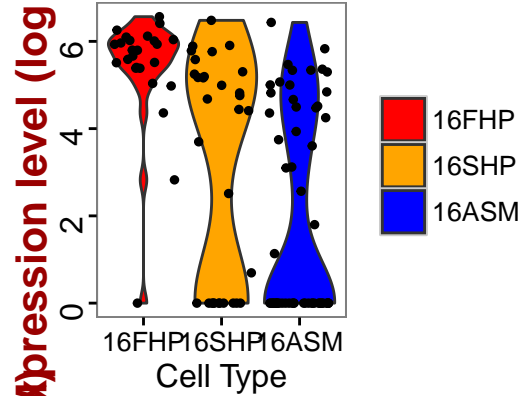


```
vlnPlot(hpf16, genes.viz.16, cols.use = c("red", "orange", "blue"))
```

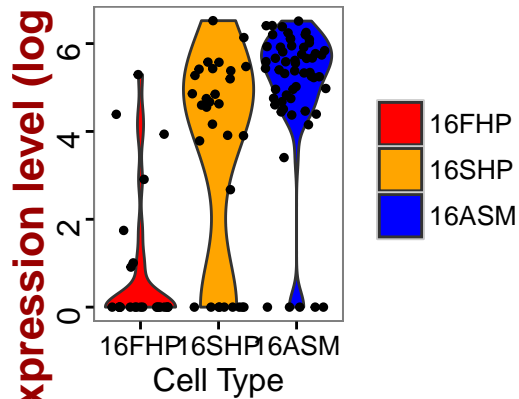
DACH1/2



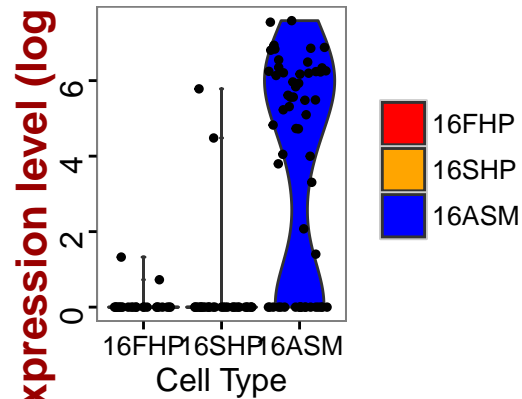
NKX2-3



TBX1/10



EBF1/2/3/4



```
# Store FHP markers in text file
```

```
FHP_16.markers = c13_16.markers
```

```
head(FHP_16.markers[order(FHP_16.markers$myAUC, decreasing = T), ], 20)
```

```
##                                                                 myAUC
## KH2013:KH.C5.227_F56C4.4                                     0.950
## SMTN                                                         0.931
## MOGAT1/2                                                    0.919
## NKX2-3                                                       0.879
## KH2013:KH.C9.40_GST01/2                                     0.873
## SI:DKEY-79F11.5                                             0.852
## KH2013:KH.C9.770_RGR                                       0.851
## SCYL                                                         0.831
## KH2013:KH.C8.489_SI:DKEY-29D8.3                            0.830
## KH2013:KH.C4.547_BMP2/4                                    0.829
## KH2013:KH.C9.247_BX119910.1/10/11/12/13/14/16/2/3/4/5/6/7/8/9 0.828
## CDH-4                                                        0.827
## SFRP1/5                                                     0.826
```

## TIMP4	0.820
## KH2013:KH.C2.22	0.818
## SFRP2	0.818
## SLIT1/2/3	0.816
## LYS2	0.816
## KH2013:KH.C4.376_UNC93A	0.806
## COL13A1	0.792
##	avg_diff
## KH2013:KH.C5.227_F56C4.4	1.269902
## SMTN	1.785546
## MOGAT1/2	1.568086
## NKX2-3	1.329046
## KH2013:KH.C9.40_GST01/2	1.365326
## SI:DKEY-79F11.5	1.537301
## KH2013:KH.C9.770_RGR	1.117639
## SCYL	1.030414
## KH2013:KH.C8.489_SI:DKEY-29D8.3	1.243672
## KH2013:KH.C4.547_BMP2/4	1.630799
## KH2013:KH.C9.247_BX119910.1/10/11/12/13/14/16/2/3/4/5/6/7/8/9	1.348407
## CDH-4	1.034349
## SFRP1/5	1.934746
## TIMP4	1.211017
## KH2013:KH.C2.22	1.290851
## SFRP2	1.820400
## SLIT1/2/3	1.306153
## LYS2	1.452199
## KH2013:KH.C4.376_UNC93A	1.210027
## COL13A1	2.009872
##	power pct.1
## KH2013:KH.C5.227_F56C4.4	0.900 1.00
## SMTN	0.862 1.00
## MOGAT1/2	0.838 1.00
## NKX2-3	0.758 0.96
## KH2013:KH.C9.40_GST01/2	0.746 1.00
## SI:DKEY-79F11.5	0.704 0.92
## KH2013:KH.C9.770_RGR	0.702 1.00
## SCYL	0.662 1.00
## KH2013:KH.C8.489_SI:DKEY-29D8.3	0.660 0.96
## KH2013:KH.C4.547_BMP2/4	0.658 0.88
## KH2013:KH.C9.247_BX119910.1/10/11/12/13/14/16/2/3/4/5/6/7/8/9	0.656 0.92
## CDH-4	0.654 1.00
## SFRP1/5	0.652 0.80
## TIMP4	0.640 0.92
## KH2013:KH.C2.22	0.636 0.92
## SFRP2	0.636 0.84

## SLIT1/2/3	0.632	0.96
## LYS2	0.632	0.84
## KH2013:KH.C4.376_UNC93A	0.612	0.72
## COL13A1	0.584	0.72
##	pct.2	
## KH2013:KH.C5.227_F56C4.4	1.000	
## SMTN	0.438	
## MOGAT1/2	0.618	
## NKX2-3	0.506	
## KH2013:KH.C9.40_GST01/2	0.652	
## SI:DKEY-79F11.5	0.685	
## KH2013:KH.C9.770_RGR	0.708	
## SCYL	0.742	
## KH2013:KH.C8.489_SI:DKEY-29D8.3	0.742	
## KH2013:KH.C4.547_BMP2/4	0.337	
## KH2013:KH.C9.247_BX119910.1/10/11/12/13/14/16/2/3/4/5/6/7/8/9	0.494	
## CDH-4	0.944	
## SFRP1/5	0.191	
## TIMP4	0.438	
## KH2013:KH.C2.22	0.438	
## SFRP2	0.315	
## SLIT1/2/3	0.584	
## LYS2	0.404	
## KH2013:KH.C4.376_UNC93A	0.236	
## COL13A1	0.180	

```
write.table(FHP_16.markers, file = "FHP_16.markers.txt", sep = "\t")

# Store SHP markers in text file
SHP_16.markers = c10_16.markers
head(SHP_16.markers[order(SHP_16.markers$myAUC, decreasing = T), ], 20)
```

##	myAUC	avg_diff	power	pct.1	pct.2
## KH2013:KH.C1.638_C17ORF105	0.812	1.121174	0.624	0.935	0.578
## DACH1/2	0.803	2.475778	0.606	0.710	0.133
## KH2013:KH.C9.692_GABRR1/2/3	0.740	1.120309	0.480	0.839	0.518
## KH2013:KH.C3.616_GNRHR/1/2/4	0.722	1.529193	0.444	0.613	0.193
## SFR1	0.704	1.048728	0.408	0.710	0.434
## ORC1	0.701	1.215804	0.402	0.645	0.289
## GM1840	0.698	1.469439	0.396	0.613	0.265
## KH2013:KH.C10.172	0.691	1.726626	0.382	0.548	0.193
## ZGC:174877	0.685	1.114684	0.370	0.645	0.349
## IKBKG	0.684	1.433664	0.368	0.581	0.301
## KH2013:KH.L150.4	0.661	1.021943	0.322	0.581	0.349

```
## PPT2                0.654  1.024035 0.308 0.613 0.349
## USP48               0.649  1.120253 0.298 0.581 0.458
## KH2013:KH.C6.155_FLRT1/2/3 0.621  1.114470 0.242 0.581 0.410
## RBM15B             0.581 -1.461119 0.162 0.774 0.590
## KH2013:KH.L124.2   0.526  1.806418 0.052 0.806 0.916
## ZNRD1              0.507 -3.447701 0.014 0.516 0.494
## KH2013:KH.L39.6   0.473 -4.277293 0.054 0.452 0.542
## SMEK1/2            0.461 -1.636758 0.078 0.613 0.602
## KH2013:KH.C8.649_ACTA1/2 0.390 -2.190225 0.220 0.355 0.590
```

```
write.table(SHP_16.markers, file = "SHP_16.markers.txt", sep = "\t")
```

```
# Store ASM markers in text file
```

```
ASM_16.markers = cl2_16.markers
```

```
head(ASM_16.markers[order(ASM_16.markers$myAUC, decreasing = T), ], 20)
```

```
##                myAUC avg_diff power pct.1 pct.2
## KH2013:KH.S555.1_HTR7 0.939 2.307248 0.878 0.983 0.464
## KH2013:KH.C1.21_CG2781 0.852 1.103323 0.704 1.000 0.946
## TBX1/10            0.820 1.194245 0.640 0.914 0.500
## EBF1/2/3/4        0.817 3.644445 0.634 0.672 0.054
## CDC6               0.816 1.009440 0.632 1.000 0.929
## HAND/2            0.810 1.079622 0.620 0.966 0.786
## ZCCHC24           0.797 1.156534 0.594 0.931 0.536
## HRH1              0.796 3.209113 0.592 0.603 0.018
## SMURF1/2          0.787 1.502168 0.574 0.828 0.536
## TMSB15A           0.784 1.133905 0.568 0.966 0.679
## ELK1/3/4          0.749 1.134942 0.498 0.741 0.286
## IRX4/6            0.735 1.253873 0.470 0.759 0.357
## KH2013:KH.C4.404   0.733 1.737132 0.466 0.690 0.304
## KH2013:KH.L134.29 0.724 1.222385 0.448 0.741 0.446
## IRX1/2/3/5        0.705 1.109414 0.410 0.569 0.214
## NOX5              0.696 1.016322 0.392 0.638 0.286
## KIAA0513          0.693 1.031799 0.386 0.759 0.500
## KH2013:KH.C9.223_CDC42 0.689 1.056894 0.378 0.914 0.821
## ITPKA             0.666 1.211642 0.332 0.586 0.339
## KH2013:KH.C12.239 0.647 1.110297 0.294 0.534 0.304
```

```
# Find HP markers (pan cardiac marker)
```

```
HP_16.markers = find.markers(hpf16, c("16SHP", "16FHP"), thresh.use = 1, test.use = "roc",
  min.pct = 0.5)
```

```
head(HP_16.markers[order(HP_16.markers$myAUC, decreasing = T), ], 20)
```

```
##                myAUC avg_diff power pct.1 pct.2
```



```

## QKI 0.845 1.109346 0.690 0.982 0.776
## SCYL 0.833 1.217014 0.666 0.964 0.638
## SLIT1/2/3 0.805 1.300099 0.610 0.893 0.448
## KH2013:KH.C8.489_SI:DKEY-29D8.3 0.789 1.261880 0.578 0.893 0.690
## HAND/1/2 0.786 1.013146 0.572 0.893 0.586
## LRP4/8 0.784 1.805954 0.568 0.696 0.224
## LYS2 0.774 1.779484 0.548 0.714 0.293
## NKX2-3 0.759 1.255571 0.518 0.768 0.448
## IGF1/2 0.733 1.277706 0.466 0.786 0.500
## SI:DKEY-79F11.5 0.715 1.158875 0.430 0.839 0.638
## KH2013:KH.C9.608_PDE9A 0.710 1.064685 0.420 0.643 0.276
## KH2013:KH.C3.716_EFNA1/2/3/4/5 0.706 1.252546 0.412 0.714 0.379
## TANC1/2 0.704 1.290037 0.408 0.518 0.207
## KH2013:KH.L119.21_POLA1 0.686 1.000919 0.372 0.696 0.431
## SMTN 0.685 1.142035 0.370 0.679 0.448
## ZGC:174877 0.684 1.254427 0.368 0.589 0.276
## LMOD1/3 0.683 1.012676 0.366 0.607 0.241
## USP48 0.665 1.094911 0.330 0.607 0.379
## KH2013:KH.C13.152_F14B6.6 0.661 1.103362 0.322 0.571 0.328
## KH2013:KH.C3.52_EFNA1/2/3/4/5 0.661 1.102856 0.322 0.571 0.259

```

```

write.table(HP_16.markers, file = "panHP_16.markers.txt", sep = "\t")

# Find SHP specific markers that distinguish two heart progenitors FHP and
# SHP
SHPspecific_16.markers = find.markers(hpf16, "16SHP", "16FHP", thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
head(SHPspecific_16.markers[order(SHPspecific_16.markers$myAUC, decreasing = T),
  ], 20)

```

```

## myAUC avg_diff power pct.1 pct.2
## TMSB15A 0.937 2.553345 0.874 0.968 0.32
## PCMTD1/2 0.929 1.542177 0.858 1.000 0.88
## FOXF1/2 0.926 2.612649 0.852 0.903 0.32
## KH2013:KH.C9.692_GABRR1/2/3 0.897 3.262190 0.794 0.839 0.08
## KH2013:KH.C1.638_C17ORF105 0.835 1.377923 0.670 0.935 0.40
## CHID1 0.828 1.637985 0.656 0.806 0.28
## ZNF703 0.813 2.656630 0.626 0.710 0.12
## KH2013:KH.S1660.1 0.812 1.157955 0.624 1.000 0.76
## DACH1/2 0.810 2.692742 0.620 0.710 0.12
## E2F/1/2/3 0.801 1.314878 0.602 0.903 0.60
## SFR1 0.794 2.042680 0.588 0.710 0.28
## GDH1/3 0.788 1.844341 0.576 0.742 0.36
## TBX1/10 0.783 2.133826 0.566 0.710 0.24

```

```
## KH2013:KH.C4.230_TAGLN/2/3    0.775 1.300167 0.550 0.806 0.36
## histone                       0.772 1.009498 0.544 0.871 0.52
## PRMT7                         0.772 1.300034 0.544 0.839 0.48
## KH2013:KH.C3.616_GNRHR/1/2/4 0.743 1.895749 0.486 0.613 0.16
## ZK637.14                     0.743 1.290958 0.486 0.710 0.24
## ITPKA                         0.735 2.323179 0.470 0.548 0.08
## KH2013:KH.C5.329             0.734 1.284407 0.468 0.710 0.36
```

```
write.table(SHPspecific_16.markers, file = "SHPspecific_16.markers.txt", sep = "\t")

# Find FHP specific markers that distinguish two heart progenitors FHP and
# SHP
FHPspecific_16.markers = find.markers(hpf16, "16FHP", "16SHP", thresh.use = 1,
  test.use = "roc", min.pct = 0.5)
head(FHPspecific_16.markers[order(FHPspecific_16.markers$myAUC, decreasing = T),
  ], 20)
```

```
##                                                                 myAUC
## MOGAT1/2                                                         0.954
## SMTN                                                             0.934
## KH2013:KH.C5.227_F56C4.4                                         0.932
## KH2013:KH.C9.770_RGR                                             0.883
## KH2013:KH.C9.40_GST01/2                                          0.858
## KH2013:KH.C7.205_ASB2                                            0.853
## KH2013:KH.C2.668_SLC38A9                                         0.850
## SFRP1/5                                                           0.843
## TIMP4                                                             0.830
## KH2013:KH.C4.457_UNC93A                                          0.827
## SI:DKEY-79F11.5                                                  0.826
## CDH-4                                                             0.826
## SFRP2                                                             0.815
## KH2013:KH.C9.247_BX119910.1/10/11/12/13/14/16/2/3/4/5/6/7/8/9 0.815
## KH2013:KH.C4.547_BMP2/4                                          0.808
## KH2013:KH.C2.22                                                 0.806
## COL13A1                                                           0.801
## SEPT12/3/9                                                       0.797
## KH2013:KH.C11.362                                               0.780
## TNS1/3                                                            0.772
##                                                                 avg_diff
## MOGAT1/2                                                         1.931942
## SMTN                                                             1.823235
## KH2013:KH.C5.227_F56C4.4                                         1.122933
## KH2013:KH.C9.770_RGR                                             1.359233
## KH2013:KH.C9.40_GST01/2                                          1.277879
```

## KH2013:KH.C7.205_ASB2	1.045074
## KH2013:KH.C2.668_SLC38A9	2.120019
## SFRP1/5	2.235711
## TIMP4	1.119125
## KH2013:KH.C4.457_UNC93A	1.331659
## SI:DKEY-79F11.5	1.317059
## CDH-4	1.039662
## SFRP2	2.048199
## KH2013:KH.C9.247_BX119910.1/10/11/12/13/14/16/2/3/4/5/6/7/8/9	1.313579
## KH2013:KH.C4.547_BMP2/4	1.472345
## KH2013:KH.C2.22	1.218241
## COL13A1	2.130965
## SEPT12/3/9	1.331829
## KH2013:KH.C11.362	1.416327
## TNS1/3	1.102989
##	power pct.1
## MOGAT1/2	0.908 1.00
## SMTN	0.868 1.00
## KH2013:KH.C5.227_F56C4.4	0.864 1.00
## KH2013:KH.C9.770_RGR	0.766 1.00
## KH2013:KH.C9.40_GST01/2	0.716 1.00
## KH2013:KH.C7.205_ASB2	0.706 1.00
## KH2013:KH.C2.668_SLC38A9	0.700 0.88
## SFRP1/5	0.686 0.80
## TIMP4	0.660 0.92
## KH2013:KH.C4.457_UNC93A	0.654 0.72
## SI:DKEY-79F11.5	0.652 0.92
## CDH-4	0.652 1.00
## SFRP2	0.630 0.84
## KH2013:KH.C9.247_BX119910.1/10/11/12/13/14/16/2/3/4/5/6/7/8/9	0.630 0.92
## KH2013:KH.C4.547_BMP2/4	0.616 0.88
## KH2013:KH.C2.22	0.612 0.92
## COL13A1	0.602 0.72
## SEPT12/3/9	0.594 0.84
## KH2013:KH.C11.362	0.560 0.88
## TNS1/3	0.544 0.88
##	pct.2
## MOGAT1/2	0.613
## SMTN	0.419
## KH2013:KH.C5.227_F56C4.4	1.000
## KH2013:KH.C9.770_RGR	0.645
## KH2013:KH.C9.40_GST01/2	0.710
## KH2013:KH.C7.205_ASB2	0.871
## KH2013:KH.C2.668_SLC38A9	0.323
## SFRP1/5	0.161

```

## TIMP4 0.355
## KH2013:KH.C4.457_UNC93A 0.161
## SI:DKEY-79F11.5 0.774
## CDH-4 0.935
## SFRP2 0.355
## KH2013:KH.C9.247_BX119910.1/10/11/12/13/14/16/2/3/4/5/6/7/8/9 0.484
## KH2013:KH.C4.547_BMP2/4 0.323
## KH2013:KH.C2.22 0.548
## COL13A1 0.194
## SEPT12/3/9 0.258
## KH2013:KH.C11.362 0.452
## TNS1/3 0.548

```

```
write.table(FHPspecific_16.markers, file = "FHPspecific_16.markers.txt", sep = "\t")
```

```
# Select markers for plotting on a Heatmap (top 10 positive markers with  
# high discriminatory power) (5 markers are shown for STVC and SHP due to  
# less markers)
```

```
marker16FHP = rownames(FHP_16.markers[order(FHP_16.markers$myAUC, decreasing = T)[1:15],  
])
```

```
marker16SHP = rownames(SHP_16.markers[order(SHP_16.markers$myAUC, decreasing = T)[1:15],  
])
```

```
marker16ASM = rownames(ASM_16.markers[order(ASM_16.markers$myAUC, decreasing = T)[1:15],  
])
```

```
marker.16 = c(marker16FHP, marker16SHP, marker16ASM)
```

```
# Redefine ASM markers
```

```
ASM_16.markers = find.markers(hpf16, "16ASM", "16FHP", thresh.use = 1, test.use = "roc",  
min.pct = 0.5)
```

```
head(ASM_16.markers[order(ASM_16.markers$myAUC, decreasing = T), ], 20)
```

```

## myAUC avg_diff power pct.1 pct.2
## KH2013:KH.S555.1_HTR7 0.968 3.169129 0.936 0.983 0.40
## TMSB15A 0.953 3.156753 0.906 0.966 0.32
## TBX1/10 0.929 2.827890 0.858 0.914 0.24
## CDC6 0.921 1.599529 0.842 1.000 0.84
## TP53I11 0.890 1.224223 0.780 1.000 0.72
## FOXF1/2 0.888 2.343817 0.776 0.897 0.32
## ZCCHC24 0.881 1.848557 0.762 0.931 0.40
## SELT/2 0.876 1.038634 0.752 0.983 0.96
## KH2013:KH.C1.21_CG2781 0.872 1.198799 0.744 1.000 0.92
## HAND/2 0.859 1.436427 0.718 0.966 0.64
## E2F/1/2/3 0.845 1.243666 0.690 0.914 0.60
## SLC16A12/7 0.839 1.432206 0.678 0.931 0.64

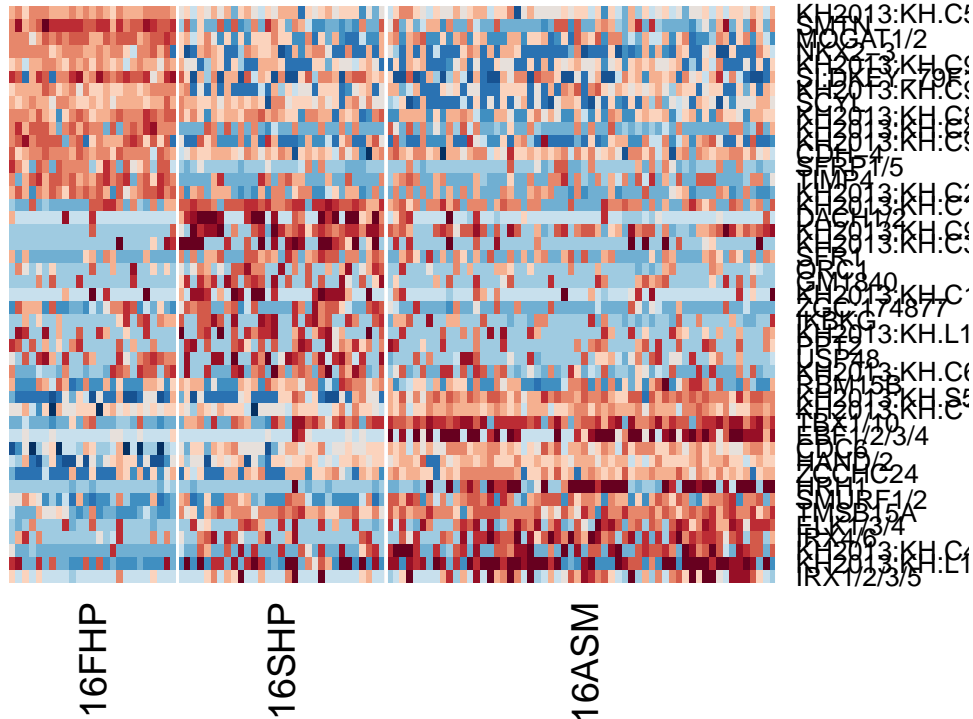
```

```
## EBF1/2/3/4          0.823 5.630515 0.646 0.672 0.04
## KH2013:KH.C9.692_GABRR1/2/3 0.818 2.464266 0.636 0.707 0.08
## KH2013:KH.C4.404    0.817 3.428063 0.634 0.690 0.08
## SMURF1/2           0.817 1.927637 0.634 0.828 0.52
## KH2013:KH.L134.29  0.809 2.316999 0.618 0.741 0.28
## ELK1/3/4           0.807 1.756193 0.614 0.741 0.16
## HRH1                0.802 4.432899 0.604 0.603 0.00
## CG9338              0.802 4.212724 0.604 0.603 0.00
```

```
write.table(ASM_16.markers, file = "ASM_16.markers.txt", sep = "\t")
```

```
# Draw a heatmap of all cells for these marker genes
```

```
doHeatMap(hpf16, genes.use = marker.16, remove.key = TRUE, slim.col.label = T,
          cex.col = 1.2, col.use = col)
```



```
# Store clustering information into a master Seurat object
```

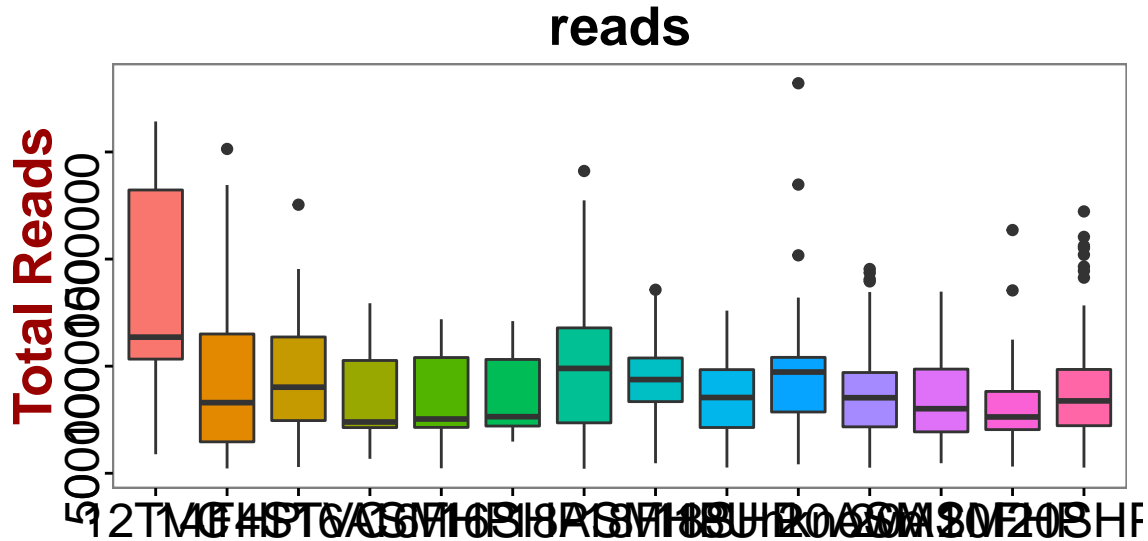
```
hpfall.cluster = hpfall.remv2
hpfall.cluster = set.ident(hpfall.cluster, which.cells(hpfall.cluster, "hpf12"),
                           hpf12@ident)
hpfall.cluster = set.ident(hpfall.cluster, which.cells(hpfall.cluster, "hpf14"),
                           hpf14@ident)
hpfall.cluster = set.ident(hpfall.cluster, which.cells(hpfall.cluster, "hpf16"),
                           hpf16@ident)
hpfall.cluster = set.ident(hpfall.cluster, which.cells(hpfall.cluster, "hpf18"),
```

```

hpf18@ident)
hpfall.cluster = set.ident(hpfall.cluster, which.cells(hpfall.cluster, "hpf20"),
hpf20@ident)

boxPlot.FPKM(hpfall.cluster, "reads", name.y = "Total Reads", name.x = "", ratio.plot =

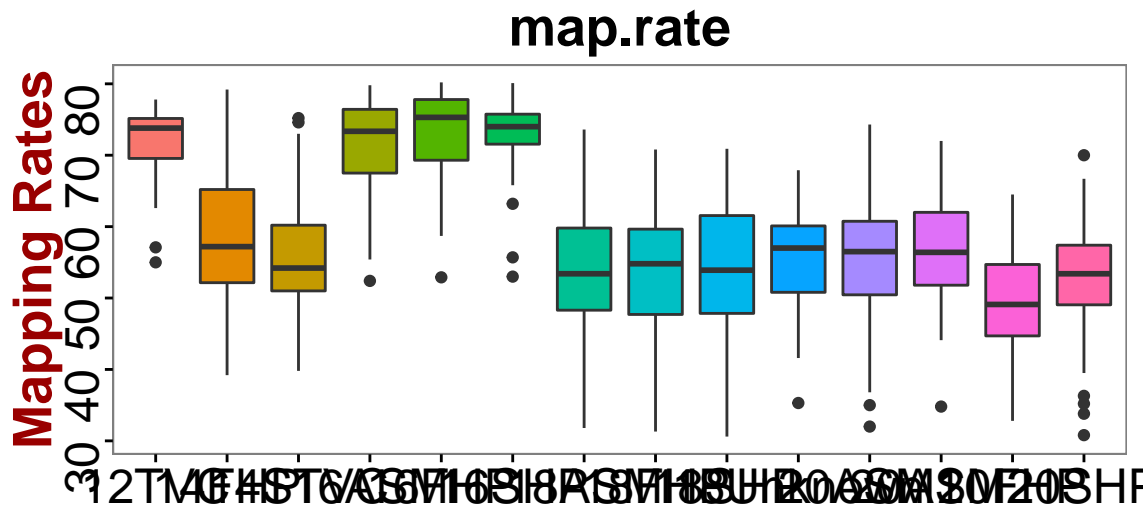
```



```

boxPlot.FPKM(hpfall.cluster, "map.rate", name.y = "Mapping Rates", name.x = "",
ratio.plot = 0.1)

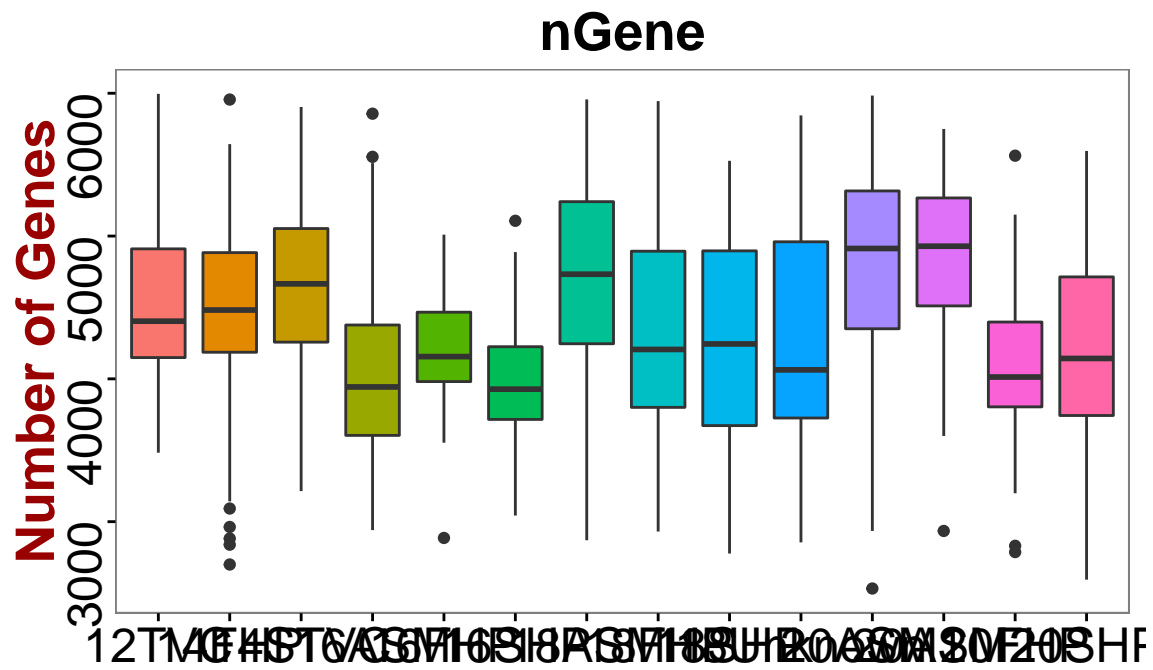
```



```

boxPlot.FPKM(hpfall.cluster, "nGene", name.y = "Number of Genes", name.x = "",
ratio.plot = 0.002)

```



```

save(hpfall.cluster, file = "hpfallCluster.Robj")
save(hpf12, hpf14, hpf16, hpf18, hpf20, file = "hpfall.Robj")

```