

Supplementary Information for “Mixed model association for biobank-scale data sets”

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Supplementary Note

1 UK Biobank data

We analyzed genetic data from the UK Biobank full release consisting of 487,409 samples typed at $\sim 800,000$ markers and imputed to ~ 93 million variants [5]. We restricted the sample set to 459,327 individuals of European ancestry (based on self-reported white ethnicity), and for linear regression analyses, we further restricted the sample set to 337,539 British-ancestry individuals passing principal component analysis filters and containing no third-degree or closer relationships [5]. (The ancestry filter eliminated $\sim 50,000$ samples and the relatedness filter eliminated an additional $\sim 70,000$ samples.) We restricted the genotyped marker set to autosomal markers with missingness $< 10\%$ and minor allele frequency (MAF) $> 0.1\%$, leaving 672,292 markers. We analyzed ~ 20 million imputed variants with MAF $> 0.1\%$ (applying this filter within BOLT-LMM).

In our running time benchmarks, we also analyzed genetic data from the UK Biobank interim release of 152,249 samples imputed to ~ 72 million variants. Applying analogous exclusions produced a sample set of 145,613 European-ancestry individuals typed at 651,011 autosomal markers with missingness $< 10\%$ and MAF $> 0.1\%$. We used QCTOOL v2 to convert imputed data between the BGEN v1.1 and v1.2 formats (to benchmark previous versions of BOLT-LMM, which only support the BGEN v1.1 format).

We analyzed 23 phenotypes selected based on phenotyping rate $> 80\%$ (Supplementary Table 1), SNP-heritability $h_g^2 > 0.08$ (Supplementary Table 2), and low correlation between traits. We performed basic QC on each trait, removing outliers outside the reasonable range for each quantitative trait and quantile normalizing within sex strata after correcting for covariates as described in previous GWAS [11–16].

In all association analyses, we included assessment center, genotyping array, sex, age, and age squared as covariates. In linear regression analyses (implemented in the BOLT-LMM software), we also included 20 principal components to correct for ancestry (provided with the UK Biobank data release [5]). In our primary BOLT-LMM analyses, we included 20 principal components computed on our filtered marker set using the FastPCA [9] algorithm (as implemented in PLINK 2.0 [17] `--pca approx`). In auxiliary BOLT-LMM analyses, we varied the number of principal components included as covariates (Supplementary Table 7).

2 BOLT-LMM version 2.3

Our new release of the BOLT-LMM software (version 2.3) performs much faster processing of imputed genotypes, which we discovered was the bottleneck for analyses of extremely large imputed data sets (e.g., ~ 93 million variants in the UK Biobank $N=500K$ release). This step of the

BOLT-LMM computation, which occurs after the model-fitting steps and scales only linearly in sample size and variant count, nonetheless accounted for the large majority of running time for previous versions of BOLT-LMM on UK Biobank data. To overcome this bottleneck, we implemented fast multi-threaded support for analysis of imputed genotypes in the new BGEN v1.2 file format (used to encode genotype probabilities in the UK Biobank full release). BOLT-LMM v2.3 analyzes these imputed genotypes by reading compressed probability data for blocks of 400 variants at a time from disk and then analyzing these data in parallel compute threads. Analysis of a single variant involves decompressing the genotype probabilities, computing the variant’s allele frequency and INFO score (and exiting early if MAF or INFO thresholds are not met), projecting out covariates, and computing the BOLT-LMM test statistic (via a dot product with the residualized phenotype and rescaling term [3]).

For analyses of very large data sets with BOLT-LMM v2.3, we additionally now recommend including principal components as covariates for the purpose of increasing the rate of convergence of the iterative computations performed during BOLT-LMM’s model-fitting steps [3]. For phenotypes with high heritability, these steps of the computation account for the majority of run time (after the improvements to processing of imputed data described above) but can be sped up by including PC covariates. Projecting out top PCs improves the conditioning of the matrix computations that BOLT-LMM implicitly performs, improving convergence; details are provided in Section 5 below.

3 Power analyses

We benchmarked statistical power of three association analysis approaches: linear regression (using 20 principal component covariates), BOLT-LMM using a Gaussian SNP effect prior (BOLT-LMM-inf, equivalent to the standard “infinitesimal” mixed model [1, 2, 18–23]), and BOLT-LMM using its default mixture-of-Gaussians prior on SNP effect sizes, which accounts for larger-effect SNPs [3]. We tested all three methods on the subset of $N=337,539$ unrelated British samples, and we additionally tested BOLT-LMM-inf and BOLT-LMM (which are robust to sample structure) on the full set of $N=459,327$ European-ancestry samples.

We performed two types of benchmarks to assess statistical power afforded by each analysis. First, we counted independent genome-wide significant associations identified by each analysis. To obtain counts that were robust to linkage disequilibrium among associated variants and avoided double-counting of loci, we used PLINK’s LD clumping algorithm [17] using LD computed in $N=113,851$ unrelated British individuals [24] at 9.6 million imputed SNPs with $MAF>0.1\%$ (corresponding to a conservative genome-wide significance threshold of $p<5\times 10^{-9}$). We used a stringent 5Mb window and R^2 threshold of 0.01 for LD clumping, and we further collapsed associated SNPs within 100kb of each other. These analyses demonstrated that BOLT-LMM-inf and

BOLT-LMM achieved considerable power gains over linear regression when run on the same set of $N=337,539$ unrelated British samples (21% and 28% increases in locus discovery, respectively; Supplementary Table 2). Expanding the sample set to include all European individuals (allowing relatives) achieved even larger boosts in power (76% and 84%, respectively; Fig. 1a and Supplementary Table 2).

Second, to provide additional insight into the power gain achieved by BOLT-LMM, we examined the amounts of phenotypic variance explained by BOLT-LMM’s internal linear predictors and the increases in χ^2 test statistics (for BOLT-LMM-inf and BOLT-LMM vs. linear regression) at associated SNPs. We previously showed that these two quantities are tightly coupled [3]; the intuition is that independent of its ability to analyze data sets containing sample structure (and thereby gain power by analyzing more samples), BOLT-LMM also achieves increased power by implicitly conditioning on polygenic predictions using genome-wide SNPs [3,6]. Conditioning on polygenic predictions effectively reduces noise in an association test, producing a multiplicative boost in χ^2 statistics at associated loci in a manner similar to increasing sample size.

We compared the variance explained by BOLT-LMM’s linear predictor—using either the default mixture-of-Gaussians prior on SNP effect sizes or the single-Gaussian BOLT-LMM-inf model, equivalent to best linear unbiased prediction (BLUP)—to the variance theoretically explained by an optimal linear predictor, i.e., SNP-heritability h_g^2 . Variance explained by the linear predictors within BOLT-LMM and BOLT-LMM-inf were estimated internally by the BOLT-LMM software via out-of-sample benchmarks (training on 80% of samples and testing on the remaining 20%). We observed that for several traits, BOLT-LMM successfully predicted more than half of SNP-heritability; for height and hair color, BOLT-LMM predicted $>40\%$ of phenotypic variance (Fig. 1b, Supplementary Fig. 1, and Supplementary Table 3).

To estimate the effective sample size achieved by BOLT-LMM and BOLT-LMM-inf, we then measured the boosts in χ^2 association statistics of BOLT-LMM and BOLT-LMM-inf (on either $N=337,539$ or $459,327$ samples) versus linear regression on $N=337,539$ unrelated British samples. Specifically, for BOLT-LMM on $N=459,327$ samples, we computed the median ratio of BOLT-LMM χ^2 statistics on $N=459,327$ samples to linear regression χ^2 statistics (on $N=337,539$ samples) across genotyped SNPs with $\chi^2 > 30$ in BOLT-LMM $N=337,539$ analyses. We ascertained associated SNPs in this way to avoid biasing our benchmarks in favor of either BOLT-LMM ($N=459,327$) or linear regression ($N=337,539$). We conducted the other benchmarks in an analogous manner; for each pair of association analyses that we compared, we always used a third set of association test statistics to ascertain associated SNPs. We observed boosts in χ^2 test statistics at associated SNPs (equivalently, boosts in effective sample size) that tracked closely with the proportions of variance predicted by BOLT-LMM and BOLT-LMM-inf (Fig. 1b, Supplementary Fig. 1, and Supplementary Table 3). For traits in which BOLT-LMM predicted only a very small fraction of phenotypic variance (e.g., hypothyroidism and smoking status), we observed that BOLT-LMM

$N=459,327$ analyses still achieved moderate gains in association power over linear regression on $N=337,539$ unrelated British samples; here, BOLT-LMM still benefited from the increased sample size (achieving power equivalent to $\sim 430\text{K}$ unrelated individuals; Supplementary Table 3). For traits in which BOLT-LMM predicted large fractions of phenotypic variance (e.g., height and hair color), we observed that BOLT-LMM $N=459,327$ analyses achieved power equivalent to linear regression on up to $\sim 700\text{K}$ unrelated samples (Fig. 1b, Supplementary Fig. 1, and Supplementary Table 3). As expected, BOLT-LMM achieved substantial additional gains over BOLT-LMM-inf for traits with larger-effect SNPs (e.g., hair color, tanning ability, and blood cell traits; Supplementary Table 3).

4 Calibration analyses

To assess the calibration of BOLT-LMM (i.e., control of false positives) when used to analyze all $N=459,327$ European samples (keeping related individuals) we performed benchmarks using LD score regression [7]. For each phenotype, we considered BOLT-LMM $N=459,327$ association statistics, linear regression $N=337,539$ association statistics computed using 20 principal component covariates (as a negative control robust to confounding), and linear regression $N=337,539$ association statistics computed without PC covariates (as a positive control susceptible to slight confounding from population stratification among British individuals). We used the LDSC software to run LD score regression on each set of association statistics using the baselineLD model [8] (which applies stratified LD score regression, S-LDSC [25]).

We previously proposed using the LD score regression intercept as a way of distinguishing polygenicity from confounding as possible sources of increased association test statistics [7]. In theory, SNPs with larger numbers of LD partners have more opportunities to tag causal variants, such that regressing observed χ^2 statistics (for a properly calibrated association test) against the LD score of a SNP should produce a regression line with a y-intercept of 1 (even if the mean χ^2 statistic across all SNPs is larger than 1 due to polygenicity); in contrast, the y-intercept will be larger than 1 if the association test is confounded by ancestry or relatedness. In practice, we previously observed that LD score intercepts were typically close to 1 but slightly larger than 1 due to deviations from the theoretical model (e.g., attenuation bias) [7].

Here, we observe that in highly-powered analyses of traits with substantial heritability, these deviations push the LDSC intercept well above 1 for uninflated association tests, e.g., PC-corrected linear regression on unrelated British samples (Supplementary Fig. 2a and Supplementary Table 4). The reason is that in such analyses, the mean χ^2 test statistic is much larger than 1 (e.g., ~ 4 for linear regression $N=337,539$ and ~ 7 for BOLT-LMM $N=459,327$ analysis of height, after excluding SNPs explaining $>0.1\%$ of variance), such that even a slight deviation from theory results in large intercepts (here, as high as 1.5). In general, we observe that LD score regression intercepts tend

to rise with SNP-heritability and sample size (Supplementary Fig. 2a and Supplementary Table 4). This behavior of the LDSC intercept makes test statistic inflation difficult to discern based on the value of the LDSC intercept alone: for example, for the height phenotype, linear regression on $N=337,539$ unrelated British samples *without* principal component covariates—which is susceptible to inflation—and BOLT-LMM on $N=459,327$ European samples both have LDSC intercepts of nearly 1.5.

Fortunately, accounting for differences in mean χ^2 statistic for different phenotypes and association methods improves the interpretability of the LDSC intercept. The *attenuation ratio*, $(\text{LDSC intercept} - 1) / (\text{mean } \chi^2 - 1)$, calibrates the intercept against the overall shift in χ^2 statistics (due to polygenicity for uninflated association tests). Here we observe that for each trait, PC-corrected linear regression and BOLT-LMM have near-identical attenuation ratios, typically around 0.08 (Fig. 1c), whereas uncorrected linear regression typically has larger attenuation ratios, indicating confounding (Supplementary Fig. 2b and Supplementary Table 4). Across 23 traits, we observe mean attenuation ratios of 0.078 (s.e.m. 0.006) for PC-corrected linear regression, 0.082 (0.005) for BOLT-LMM, and 0.104 (0.012) for uncorrected linear regression, providing confidence that BOLT-LMM is successfully controlling for sample structure (as expected for mixed model methods) [1, 2]. We note that attenuation ratios are broadly smaller under the baselineLD model, which incorporates genome annotations [8], than under the original LDSC model (Supplementary Table 5), consistent with better model fit upon incorporating genome annotations.

5 Running time analyses

We benchmarked the running time of BOLT-LMM v2.3 (with 20 principal component covariates to increase convergence rate; see below), the previous version of BOLT-LMM [3], and linear regression using 20 principal component covariates (implemented efficiently within the BOLT-LMM software; cf. Bycroft et al. Table S9 [5]) in example analyses of the years-of-education phenotype. We ran each method on all European-ancestry individuals in the UK Biobank interim and full data releases, analyzing $\sim 72\text{M}$ and $\sim 93\text{M}$ imputed SNPs, respectively, and imposing a $\text{MAF} > 0.1\%$ filter on minor allele frequency. (We ran linear regression on all European-ancestry individuals for the sake of run time comparison even though this analysis would not be performed in practice due to potential confounding from sample structure. Also, for BOLT-LMM v1 analysis of the full data release, we analyzed imputed data from only chromosome 22 and extrapolated the computational cost to the full genome.) We performed all analyses using 8 threads of a 2.10 GHz Intel Xeon E5-2683 v4 processor and reported the median of 5 runs (Fig. 1d and Supplementary Table 6), observing a $\sim 4\text{x}$ speedup of BOLT-LMM v2.3 over the previous version, achieving speed comparable to linear regression.

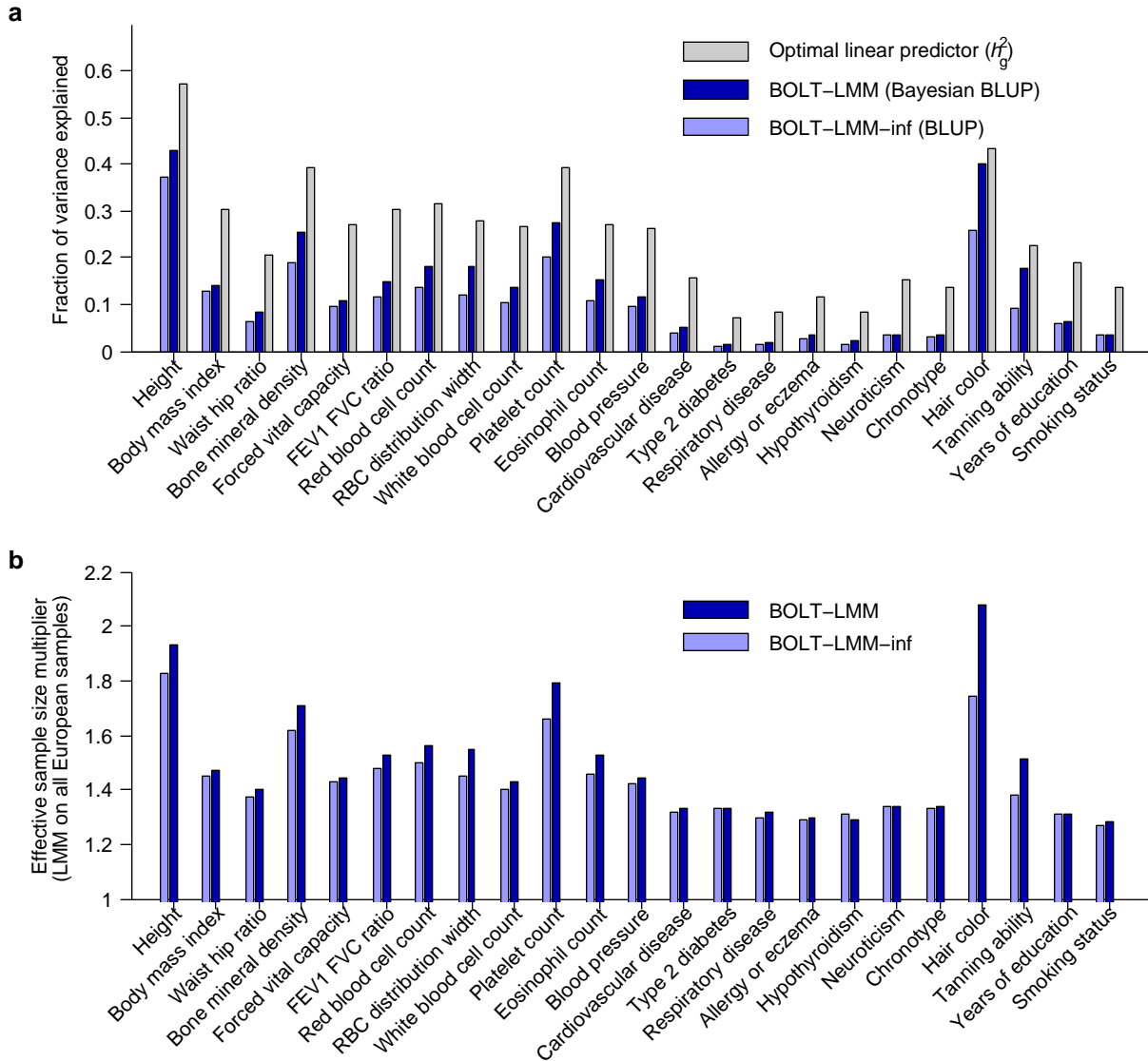
We further explored the effect of including varying numbers of principal components as co-

variates in BOLT-LMM analyses to improve convergence speed. During its model-fitting steps, BOLT-LMM applies iterative methods (specifically, conjugate gradient iteration and variational Bayes) to eliminate computationally expensive matrix operations that scale quadratically or cubically with sample size [3]. The cost of a single iteration scales only linearly with N ; however, we previously observed that the number of iterations required to achieve convergence increases slowly with N [3]. Our analyses here (Supplementary Table 7) demonstrate that convergence can be sped up by including principal component covariates (which effectively improve the conditioning of the underlying matrix computations), thus achieving close-to-linear scaling of run time with sample size. We note that to achieve increased convergence, principal components need to be computed on the set of SNPs used in the mixed model; PCs that do not match the implicit genetic relationship matrix (GRM) will not improve conditioning. We also note that after model-fitting, BOLT-LMM performs a linear-time association test on imputed SNPs (which we sped up separately using multi-threading; see Section 2); the speedup described here only applies to the model-fitting step.

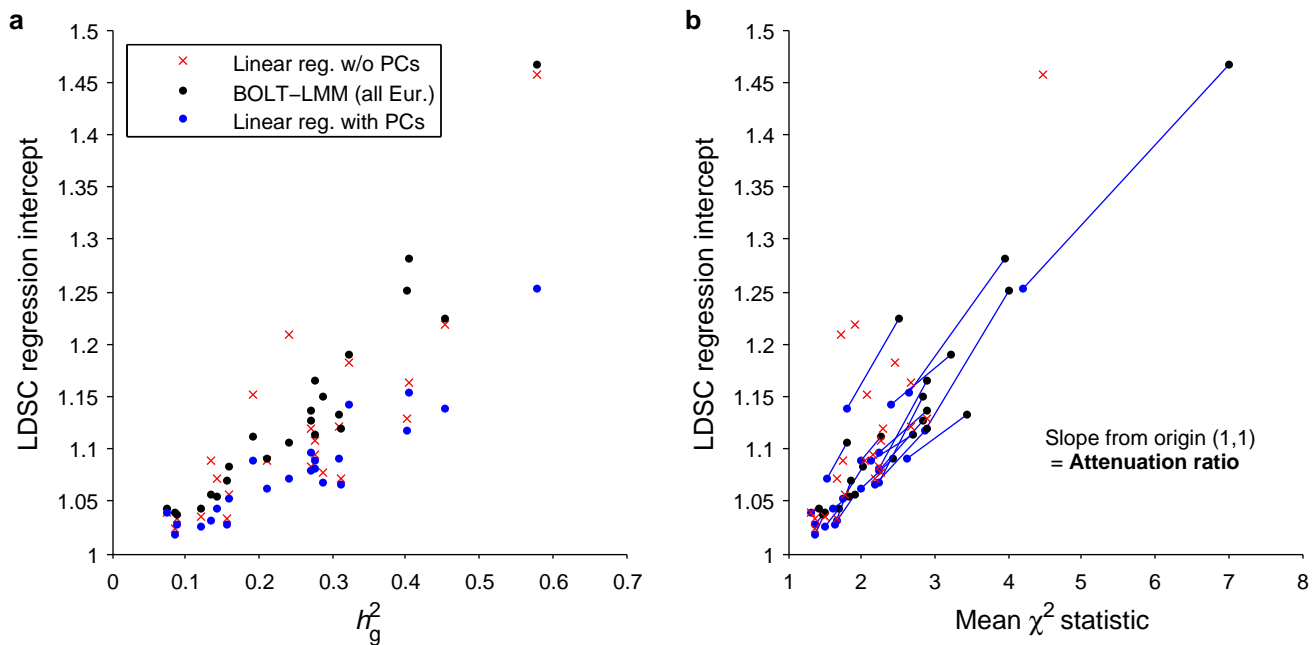
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Supplementary Figure 1. Conditioning on polygenic predictions from genome-wide SNPs boosts association power. (a) Comparison of variance explained by BOLT-LMM’s linear predictor—using either the default mixture-of-Gaussians prior on SNP effect sizes, which accounts for larger-effect SNPs [3], or the single-Gaussian “infinitesimal” model (BOLT-LMM-inf, equivalent to best linear unbiased prediction, BLUP)—and variance theoretically explained by an optimal linear predictor, i.e., SNP-heritability h_g^2 . BOLT-LMM and BOLT-LMM-inf results (on $N=459,327$ European-ancestry samples) are from out-of-sample prediction performed internally by the BOLT-LMM software (holding out 20% of samples for testing). (b) Boost in effective sample size using BOLT-LMM or BOLT-LMM-inf on $N=459,327$ European samples vs. linear regression on $N=337,539$ unrelated British samples, as assessed by multiplicative increase in χ^2 statistics at associated SNPs (Supplementary Note). Numerical data are provided in Supplementary Table 3.



Supplementary Figure 2. LD score regression intercepts. Plotted points correspond to analyses of 23 phenotypes using 3 association methods (under the baselineLD model [8] of LDSC). (a) LD score regression intercepts [7] tend to rise with SNP-heritability and sample size, even for association tests robust to confounding (e.g., linear regression on $N=337,539$ unrelated British samples using 20 principal component covariates and BOLT-LMM on $N=459,327$ European samples). This behavior of the LDSC intercept makes test statistic inflation difficult to discern based on the value of the LDSC intercept alone: for example, for the height phenotype, linear regression on $N=337,539$ unrelated British samples *without* principal component covariates—which is susceptible to inflation—and BOLT-LMM on $N=459,327$ European samples both have LDSC intercepts of nearly 1.5. (b) Accounting for differences in mean χ^2 statistic for different phenotypes and association methods improves the interpretability of the LDSC intercept. Deviations from the theoretical model assumed by LD score regression (e.g., attenuation bias [7]) push the LDSC intercept above 1—even for uninflated association tests—toward the mean χ^2 test statistic (which can be much larger than 1 for highly-powered analyses of traits with substantial heritability, e.g., ~ 7 for BOLT-LMM analysis of height, after excluding SNPs explaining $>0.1\%$ of variance). The attenuation ratio, $(\text{intercept} - 1) / (\text{mean } \chi^2 - 1)$, calibrates the intercept against the overall shift in χ^2 statistics (due to polygenicity for uninflated association tests). In these data, we observe that for each trait, PC-corrected linear regression and BOLT-LMM (connected by a line segment) have near-identical attenuation ratios, typically around 0.08, whereas uncorrected linear regression typically has larger attenuation ratios, indicating confounding. Numerical data are provided in Supplementary Table 4.

Supplementary Table 1. Number of phenotyped individuals analyzed per UK Biobank trait.

Phenotype	<i>N</i>	Fraction phenotyped
Height	458303	1.00
Body mass index	457824	1.00
Waist hip ratio	458417	1.00
Bone mineral density	445921	0.97
Forced vital capacity	371949	0.81
FEV1 FVC ratio	371949	0.81
Red blood cell count	445174	0.97
RBC distribution width	442700	0.96
White blood cell count	444502	0.97
Platelet count	444382	0.97
Eosinophil count	439938	0.96
Blood pressure (systolic)	422771	0.92
Cardiovascular disease	459324	1.00
Type 2 diabetes	459324	1.00
Respiratory disease	459324	1.00
Allergy or eczema	458699	1.00
Hypothyroidism	459324	1.00
Neuroticism	372066	0.81
Chronotype	410520	0.89
Hair color	452720	0.99
Tanning ability	449984	0.98
Years of education	454813	0.99
Smoking status	457683	1.00
Self-reported white, QC pass	459327	–

Phenotypes we analyzed were available for large majorities of the 459,327 UK Biobank participants we analyzed who self-reported white ancestry and passed genotyping QC (Supplementary Note). Throughout this manuscript, when we refer to analyses of 459K European-ancestry individuals, we take it to be understood that the actual number of individuals analyzed per phenotype is slightly smaller than 459K and varies depending on phenotyping rate.

Supplementary Table 2. Number of independent GWAS loci identified by different association analysis methods.

Phenotype	h_g^2	$N=337K$ unrelated British			$N=459K$ all European	
		Linear reg.	BOLT-LMM-inf	BOLT-LMM	BOLT-LMM-inf	BOLT-LMM
Height	0.579	1086	1479	1540	1992	2098
Body mass index	0.308	300	379	387	645	665
Waist hip ratio	0.210	217	241	255	365	384
Bone mineral density	0.401	537	681	713	947	978
Forced vital capacity	0.277	203	244	251	406	412
FEV1 FVC ratio	0.313	308	368	391	552	566
Red blood cell count	0.324	406	485	505	697	714
RBC distribution width	0.288	354	387	418	544	570
White blood cell count	0.272	347	387	404	555	584
Platelet count	0.404	558	694	751	955	1007
Eosinophil count	0.277	342	403	414	576	625
Blood pressure	0.271	282	332	346	516	522
Cardiovascular disease	0.160	126	131	135	210	213
Type 2 diabetes	0.074	38	39	41	62	62
Respiratory disease	0.086	46	46	50	75	76
Allergy or eczema	0.120	99	99	99	149	153
Hypothyroidism	0.088	69	67	70	112	111
Neuroticism	0.156	36	41	43	75	78
Chronotype	0.143	53	54	57	100	101
Hair color	0.454	210	273	326	352	436
Tanning ability	0.242	95	105	113	129	136
Years of education	0.193	95	89	91	165	172
Smoking status	0.134	32	37	36	93	96
All phenotypes		5839	7061	7436	10272	10759

Counts of independent genome-wide significant associations ($p < 5 \times 10^{-9}$) are reported for three types of association tests: linear regression using 20 principal component covariates, BOLT-LMM using a Gaussian SNP effect prior (the standard “infinitesimal” mixed model, BOLT-LMM-inf), and BOLT-LMM using its default mixture-of-Gaussians prior on SNP effect sizes, which accounts for larger-effect SNPs [3]. We tested all three methods on $N=337K$ unrelated British samples, and we additionally tested BOLT-LMM-inf and BOLT-LMM (which are robust to sample structure) on all individuals who reported white ethnicity ($N=459,327$ European-ancestry samples). For reference, we also report SNP-heritability estimated by BOLT-LMM on the $N=337K$ unrelated British samples.

For each analysis, we counted independent associations by performing stringent LD clumping (requiring $R^2 < 0.01$ in 5Mb windows) and further collapsing associated SNPs within 100kb of each other (Supplementary Note).

Supplementary Table 3. Conditioning on polygenic predictions boosts association power.

(a) Proportion of variance explained (BOLT-LMM prediction R^2 in cross-validation and h_g^2)

Phenotype	$N=337K$ unrelated British			$N=459K$ all European		
	BOLT-LMM-inf	BOLT-LMM	h_g^2	BOLT-LMM-inf	BOLT-LMM	h_g^2
Height	0.332	0.397	0.579	0.373	0.429	0.570
Body mass index	0.111	0.122	0.308	0.129	0.139	0.303
Waist hip ratio	0.055	0.075	0.210	0.063	0.084	0.205
Bone mineral density	0.168	0.236	0.401	0.191	0.255	0.391
Forced vital capacity	0.079	0.089	0.277	0.095	0.108	0.272
FEV1 FVC ratio	0.102	0.136	0.313	0.115	0.147	0.303
Red blood cell count	0.119	0.166	0.324	0.137	0.181	0.314
RBC distribution width	0.101	0.166	0.288	0.119	0.180	0.279
White blood cell count	0.091	0.124	0.272	0.105	0.137	0.266
Platelet count	0.170	0.255	0.404	0.201	0.275	0.394
Eosinophil count	0.091	0.139	0.277	0.108	0.154	0.272
Blood pressure	0.079	0.101	0.271	0.096	0.116	0.264
Cardiovascular disease	0.034	0.042	0.160	0.039	0.049	0.155
Type 2 diabetes	0.009	0.014	0.074	0.010	0.015	0.073
Respiratory disease	0.013	0.019	0.086	0.014	0.020	0.083
Allergy or eczema	0.022	0.033	0.120	0.025	0.035	0.115
Hypothyroidism	0.012	0.023	0.088	0.014	0.024	0.085
Neuroticism	0.027	0.028	0.156	0.033	0.036	0.151
Chronotype	0.024	0.027	0.143	0.030	0.034	0.138
Hair color	0.234	0.397	0.454	0.257	0.401	0.434
Tanning ability	0.080	0.173	0.242	0.092	0.177	0.226
Years of education	0.050	0.052	0.193	0.061	0.064	0.188
Smoking status	0.025	0.026	0.134	0.033	0.035	0.136

(b) Boost in effective sample size (vs. linear regression on $N=337K$ unrelated British samples)

Phenotype	$N=337K$ unrelated British		$N=459K$ all European		
	BOLT-LMM-inf	BOLT-LMM	BOLT-LMM-inf	BOLT-LMM	BOLT-LMM N_{eff}
Height	1.37x	1.45x	1.83x	1.93x	650K
Body mass index	1.14x	1.15x	1.45x	1.47x	500K
Waist hip ratio	1.06x	1.08x	1.37x	1.40x	470K
Bone mineral density	1.21x	1.29x	1.62x	1.71x	580K
Forced vital capacity	1.10x	1.11x	1.43x	1.44x	490K
FEV1 FVC ratio	1.13x	1.17x	1.48x	1.53x	520K
Red blood cell count	1.14x	1.19x	1.50x	1.56x	530K
RBC distribution width	1.12x	1.19x	1.45x	1.55x	520K
White blood cell count	1.09x	1.11x	1.40x	1.43x	480K
Platelet count	1.23x	1.33x	1.66x	1.79x	600K
Eosinophil count	1.11x	1.17x	1.46x	1.53x	520K
Blood pressure	1.13x	1.15x	1.42x	1.44x	480K
Cardiovascular disease	1.04x	1.04x	1.32x	1.33x	450K
Type 2 diabetes	1.01x	1.01x	1.33x	1.33x	450K
Respiratory disease	1.02x	1.02x	1.30x	1.32x	440K
Allergy or eczema	1.05x	1.05x	1.29x	1.30x	440K
Hypothyroidism	1.03x	1.05x	1.31x	1.29x	430K
Neuroticism	1.02x	1.02x	1.34x	1.34x	450K
Chronotype	1.04x	1.05x	1.33x	1.34x	450K
Hair color	1.30x	1.60x	1.74x	2.08x	700K
Tanning ability	1.06x	1.17x	1.38x	1.51x	510K
Years of education	1.02x	1.03x	1.31x	1.31x	440K
Smoking status	1.04x	1.05x	1.27x	1.28x	430K

Supplementary Table 4. LDSC intercepts increase with mean χ^2 statistics while attenuation ratios are consistently low for BOLT-LMM and linear regression with PC covariates.

(a) LDSC intercept (jackknife s.e.) and mean χ^2 statistic for different association methods

Phenotype	h_g^2	LDSC intercept (baselineLD model)			Mean χ^2 statistic		
		Lin. reg. w/o PCs	Lin. regression	BOLT-LMM	LR w/o PCs	LR	BOLT-LMM
Height	0.579	1.456 (0.035)	1.252 (0.033)	1.468 (0.057)	4.48	4.19	7.00
Body mass index	0.308	1.121 (0.016)	1.090 (0.015)	1.132 (0.019)	2.68	2.63	3.45
Waist hip ratio	0.210	1.088 (0.016)	1.062 (0.015)	1.091 (0.019)	2.05	2.00	2.44
Bone mineral density	0.401	1.129 (0.034)	1.117 (0.034)	1.250 (0.048)	2.88	2.86	4.01
Forced vital capacity	0.277	1.095 (0.013)	1.089 (0.013)	1.113 (0.015)	2.14	2.13	2.70
FEV1 FVC ratio	0.313	1.071 (0.015)	1.065 (0.016)	1.119 (0.021)	2.20	2.18	2.89
Red blood cell count	0.324	1.183 (0.025)	1.142 (0.026)	1.189 (0.036)	2.46	2.39	3.23
RBC distribution width	0.288	1.078 (0.025)	1.067 (0.025)	1.151 (0.032)	2.26	2.24	2.83
White blood cell count	0.272	1.120 (0.018)	1.097 (0.018)	1.137 (0.021)	2.28	2.24	2.89
Platelet count	0.404	1.163 (0.029)	1.154 (0.029)	1.281 (0.044)	2.68	2.66	3.96
Eosinophil count	0.277	1.108 (0.021)	1.082 (0.021)	1.165 (0.030)	2.26	2.23	2.89
Blood pressure	0.271	1.084 (0.015)	1.079 (0.015)	1.128 (0.018)	2.24	2.23	2.84
Cardiovascular disease	0.160	1.056 (0.013)	1.053 (0.013)	1.083 (0.016)	1.77	1.76	2.03
Type 2 diabetes	0.074	1.040 (0.012)	1.040 (0.012)	1.043 (0.015)	1.31	1.31	1.41
Respiratory disease	0.086	1.024 (0.011)	1.019 (0.011)	1.040 (0.013)	1.37	1.36	1.49
Allergy or eczema	0.120	1.034 (0.014)	1.026 (0.014)	1.043 (0.016)	1.51	1.50	1.70
Hypothyroidism	0.088	1.033 (0.013)	1.029 (0.013)	1.036 (0.013)	1.37	1.36	1.48
Neuroticism	0.156	1.034 (0.015)	1.028 (0.015)	1.069 (0.011)	1.66	1.65	1.85
Chronotype	0.143	1.072 (0.012)	1.043 (0.012)	1.055 (0.013)	1.66	1.62	1.84
Hair color	0.454	1.218 (0.057)	1.139 (0.054)	1.224 (0.078)	1.92	1.82	2.52
Tanning ability	0.242	1.209 (0.032)	1.071 (0.029)	1.105 (0.042)	1.73	1.53	1.81
Years of education	0.193	1.152 (0.013)	1.089 (0.012)	1.112 (0.013)	2.09	1.98	2.26
Smoking status	0.134	1.088 (0.011)	1.032 (0.010)	1.057 (0.011)	1.76	1.67	1.92

(b) LDSC attenuation ratio: $(\text{intercept} - 1) / (\text{mean } \chi^2 - 1)$; jackknife s.e.

Phenotype	Lin. reg. w/o PCs	Lin. regression	BOLT-LMM
Height	0.131 (0.010)	0.079 (0.010)	0.078 (0.009)
Body mass index	0.072 (0.009)	0.055 (0.009)	0.054 (0.008)
Waist hip ratio	0.084 (0.015)	0.061 (0.015)	0.063 (0.013)
Bone mineral density	0.069 (0.018)	0.063 (0.018)	0.083 (0.016)
Forced vital capacity	0.083 (0.012)	0.079 (0.012)	0.066 (0.009)
FEV1 FVC ratio	0.060 (0.013)	0.055 (0.013)	0.063 (0.011)
Red blood cell count	0.125 (0.018)	0.102 (0.018)	0.085 (0.016)
RBC distribution width	0.062 (0.020)	0.054 (0.020)	0.082 (0.017)
White blood cell count	0.093 (0.014)	0.078 (0.014)	0.072 (0.011)
Platelet count	0.097 (0.017)	0.093 (0.017)	0.095 (0.015)
Eosinophil count	0.085 (0.017)	0.067 (0.017)	0.087 (0.016)
Blood pressure	0.067 (0.012)	0.065 (0.012)	0.069 (0.010)
Cardiovascular disease	0.073 (0.017)	0.070 (0.017)	0.081 (0.016)
Type 2 diabetes	0.130 (0.040)	0.129 (0.041)	0.105 (0.036)
Respiratory disease	0.064 (0.030)	0.052 (0.031)	0.080 (0.026)
Allergy or eczema	0.067 (0.028)	0.051 (0.029)	0.061 (0.023)
Hypothyroidism	0.089 (0.035)	0.079 (0.035)	0.076 (0.026)
Neuroticism	0.051 (0.024)	0.043 (0.024)	0.082 (0.013)
Chronotype	0.109 (0.018)	0.070 (0.019)	0.066 (0.015)
Hair color	0.236 (0.062)	0.170 (0.066)	0.147 (0.051)
Tanning ability	0.288 (0.045)	0.134 (0.054)	0.129 (0.051)
Years of education	0.140 (0.012)	0.091 (0.012)	0.089 (0.010)
Smoking status	0.117 (0.015)	0.047 (0.015)	0.062 (0.012)

Supplementary Table 5. LDSC intercepts and attenuation ratios are higher under the original LDSC model (vs. baselineLD model).

(a) LDSC intercept (jackknife s.e.) and mean χ^2 statistic for different association methods

Phenotype	h_g^2	LDSC intercept (original LDSC model)			Mean χ^2 statistic		
		Lin. reg. w/o PCs	Lin. regression	BOLT-LMM	LR w/o PCs	LR	BOLT-LMM
Height	0.579	1.706 (0.031)	1.493 (0.030)	1.870 (0.043)	4.70	4.40	7.62
Body mass index	0.308	1.235 (0.017)	1.202 (0.016)	1.318 (0.019)	2.69	2.64	3.48
Waist hip ratio	0.210	1.217 (0.015)	1.185 (0.015)	1.267 (0.018)	2.05	2.01	2.44
Bone mineral density	0.401	1.315 (0.022)	1.305 (0.022)	1.497 (0.028)	3.12	3.10	4.51
Forced vital capacity	0.277	1.194 (0.014)	1.190 (0.014)	1.267 (0.017)	2.14	2.13	2.70
FEV1 FVC ratio	0.313	1.218 (0.015)	1.212 (0.014)	1.326 (0.018)	2.24	2.22	2.96
Red blood cell count	0.324	1.323 (0.020)	1.272 (0.019)	1.393 (0.026)	2.58	2.51	3.44
RBC distribution width	0.288	1.181 (0.017)	1.171 (0.017)	1.274 (0.022)	2.42	2.40	3.15
White blood cell count	0.272	1.264 (0.018)	1.238 (0.018)	1.349 (0.023)	2.34	2.30	2.97
Platelet count	0.404	1.283 (0.020)	1.272 (0.020)	1.461 (0.026)	2.85	2.83	4.35
Eosinophil count	0.277	1.232 (0.019)	1.207 (0.019)	1.324 (0.024)	2.38	2.36	3.15
Blood pressure	0.271	1.200 (0.014)	1.195 (0.013)	1.303 (0.017)	2.24	2.23	2.85
Cardiovascular disease	0.160	1.127 (0.012)	1.123 (0.012)	1.181 (0.015)	1.77	1.76	2.03
Type 2 diabetes	0.074	1.066 (0.009)	1.065 (0.009)	1.084 (0.010)	1.31	1.31	1.42
Respiratory disease	0.086	1.076 (0.010)	1.071 (0.010)	1.103 (0.011)	1.37	1.36	1.49
Allergy or eczema	0.120	1.114 (0.011)	1.107 (0.011)	1.148 (0.012)	1.51	1.50	1.70
Hypothyroidism	0.088	1.081 (0.011)	1.078 (0.011)	1.103 (0.012)	1.37	1.36	1.49
Neuroticism	0.156	1.079 (0.010)	1.074 (0.010)	1.113 (0.010)	1.66	1.65	1.85
Chronotype	0.143	1.114 (0.011)	1.082 (0.010)	1.103 (0.011)	1.66	1.62	1.84
Hair color	0.454	1.212 (0.017)	1.133 (0.015)	1.238 (0.021)	3.02	2.89	4.80
Tanning ability	0.242	1.219 (0.013)	1.075 (0.011)	1.109 (0.013)	2.35	2.12	2.70
Years of education	0.193	1.216 (0.012)	1.147 (0.011)	1.187 (0.012)	2.09	1.98	2.26
Smoking status	0.134	1.149 (0.010)	1.085 (0.010)	1.125 (0.011)	1.76	1.67	1.92

(b) LDSC attenuation ratio: $(\text{intercept} - 1) / (\text{mean } \chi^2 - 1)$; jackknife s.e.

Phenotype	Lin. reg. w/o PCs	Lin. regression	BOLT-LMM
Height	0.191 (0.008)	0.145 (0.009)	0.132 (0.006)
Body mass index	0.139 (0.010)	0.123 (0.010)	0.128 (0.008)
Waist hip ratio	0.206 (0.015)	0.184 (0.015)	0.185 (0.013)
Bone mineral density	0.148 (0.010)	0.145 (0.010)	0.142 (0.008)
Forced vital capacity	0.170 (0.013)	0.168 (0.013)	0.157 (0.010)
FEV1 FVC ratio	0.176 (0.012)	0.173 (0.012)	0.166 (0.009)
Red blood cell count	0.204 (0.013)	0.180 (0.013)	0.161 (0.011)
RBC distribution width	0.127 (0.012)	0.122 (0.012)	0.128 (0.010)
White blood cell count	0.197 (0.014)	0.184 (0.014)	0.177 (0.012)
Platelet count	0.153 (0.011)	0.149 (0.011)	0.138 (0.008)
Eosinophil count	0.168 (0.014)	0.153 (0.014)	0.150 (0.011)
Blood pressure	0.162 (0.011)	0.159 (0.011)	0.164 (0.009)
Cardiovascular disease	0.165 (0.016)	0.162 (0.016)	0.177 (0.014)
Type 2 diabetes	0.212 (0.029)	0.211 (0.030)	0.202 (0.025)
Respiratory disease	0.207 (0.028)	0.198 (0.028)	0.209 (0.022)
Allergy or eczema	0.223 (0.022)	0.211 (0.022)	0.211 (0.017)
Hypothyroidism	0.220 (0.029)	0.214 (0.029)	0.212 (0.025)
Neuroticism	0.120 (0.015)	0.113 (0.015)	0.134 (0.012)
Chronotype	0.172 (0.016)	0.134 (0.016)	0.123 (0.013)
Hair color	0.104 (0.008)	0.070 (0.008)	0.063 (0.005)
Tanning ability	0.162 (0.009)	0.067 (0.010)	0.064 (0.008)
Years of education	0.199 (0.011)	0.149 (0.011)	0.148 (0.010)
Smoking status	0.197 (0.014)	0.127 (0.015)	0.137 (0.012)

Supplementary Table 6. Running time of association methods on UK Biobank data.

Data set	Linear regression	BOLT-LMM v1	BOLT-LMM v2.3
$N=150K$	0.49 days	2.43 days	0.62 days
$N=500K$	1.62 days	9.34 days	2.54 days

Run time benchmarks for association analyses using BOLT-LMM v2.3 (with 20 principal component covariates to increase convergence rate; Supplementary Table 7), the previous version of BOLT-LMM [3], and linear regression using 20 principal component covariates (implemented efficiently within the BOLT-LMM software; cf. Bycroft et al. Table S9 [5]). We analyzed the years-of-education phenotype as a representative trait, and we ran all methods on the same set of all European-ancestry individuals in the UK Biobank $N=150K$ and $N=500K$ data releases (Supplementary Note), analyzing $\sim 72M$ and $\sim 93M$ imputed SNPs, respectively, and imposing a $MAF > 0.1\%$ filter on minor allele frequency. Analyses used 8 threads on a 2.10 GHz Intel Xeon E5-2683 v4 processor.

Supplementary Table 7. Faster convergence of BOLT-LMM iterative computations using principal component covariates.

Principal component covariates	Conjugate gradient iterations	Variational Bayes iterations
0	85	178
10	75	82
20	63	79
30	55	80
40	49	73

During its model-fitting steps, BOLT-LMM applies iterative methods (specifically, conjugate gradient iteration and variational Bayes) to eliminate computationally expensive matrix operations that scale quadratically or cubically with sample size [3]. The cost of a single iteration scales only linearly with N ; however, we previously observed that the number of iterations required to achieve convergence increases slowly with N [3]. Our analyses here demonstrate that convergence can be sped up by including principal component covariates (which effectively improve the conditioning of the underlying matrix computations), thus achieving close-to-linear scaling of run time with sample size. The iteration counts reported in this table are for total numbers of iterations performed in BOLT-LMM’s conjugate gradient steps (for estimating parameters and fitting the infinitesimal mixed model) and variational Bayes steps (for estimating parameters and fitting the mixture-of-Gaussians model) [3]. We note that to achieve increased convergence, principal components need to be computed on the set of SNPs used in the mixed model; PCs that do not match the implicit genetic relationship matrix (GRM) will not improve conditioning. We also note that after model-fitting, BOLT-LMM performs a linear-time association test on imputed SNPs; the speedup described here only applies to the model-fitting step.