

SUPPLEMENTAL INFORMATION

A taxogenomics approach uncovers a new genus in the phylum Placozoa

Michael Eitel, Warren R. Francis, Hans-Jürgen Osigus, Stefan Krebs, Sergio Vargas, Helmut Blum, Gray A. Williams, Bernd Schierwater, Gert Wörheide*

* contact: woerheide@lmu.de

NOTE RELATING TO TAXONOMIC RULES: According to the [International Code of Zoological Nomenclature](#) preprint publication of taxonomic names is discouraged. Consequently, the “*Xxxxxxxxxx yyyyyyyyyyyyyy/X. yyyyyyyyyyyyyy*” given here is a dummy only. The valid name will be available upon formal journal publication.

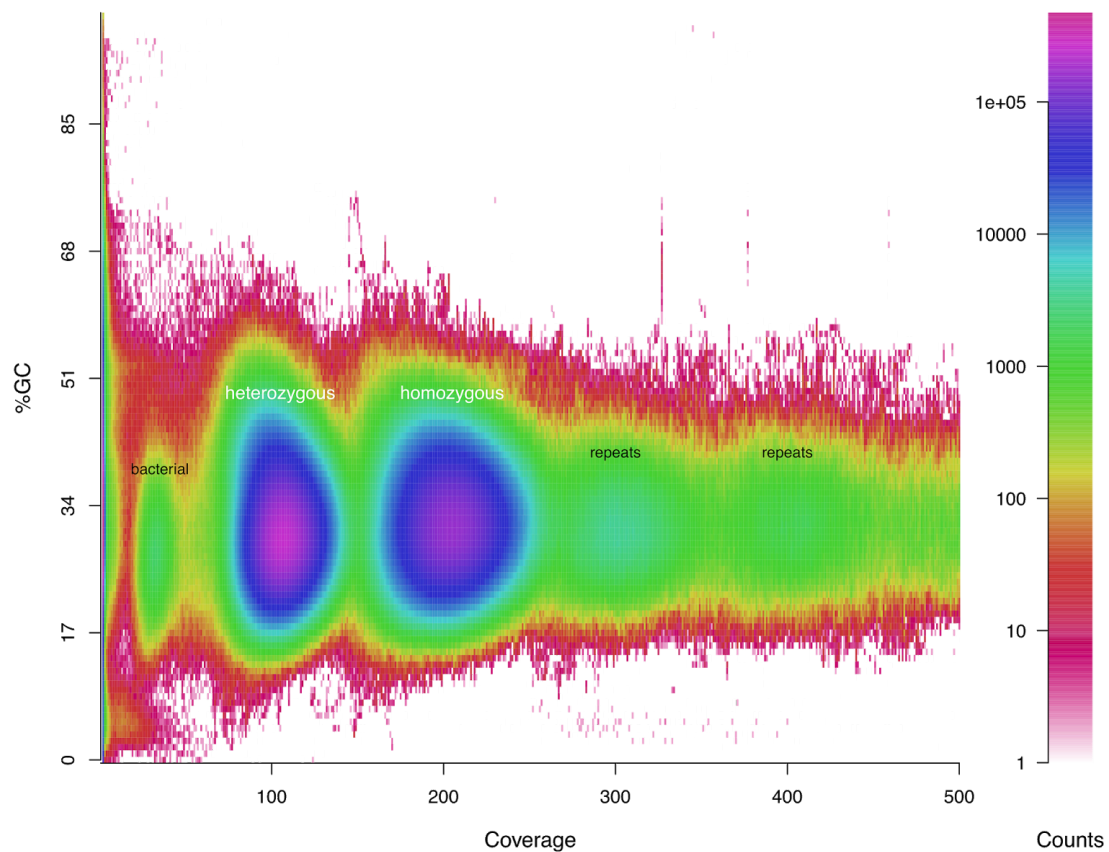


Figure S1. Lavalamp plot of kmer coverage.

The color code denotes the number of reads with a specific %GC and 31-bp kmer coverage. Heterozygous and homozygous coverage clouds show high counts at roughly 100x and 200x coverage, respectively.

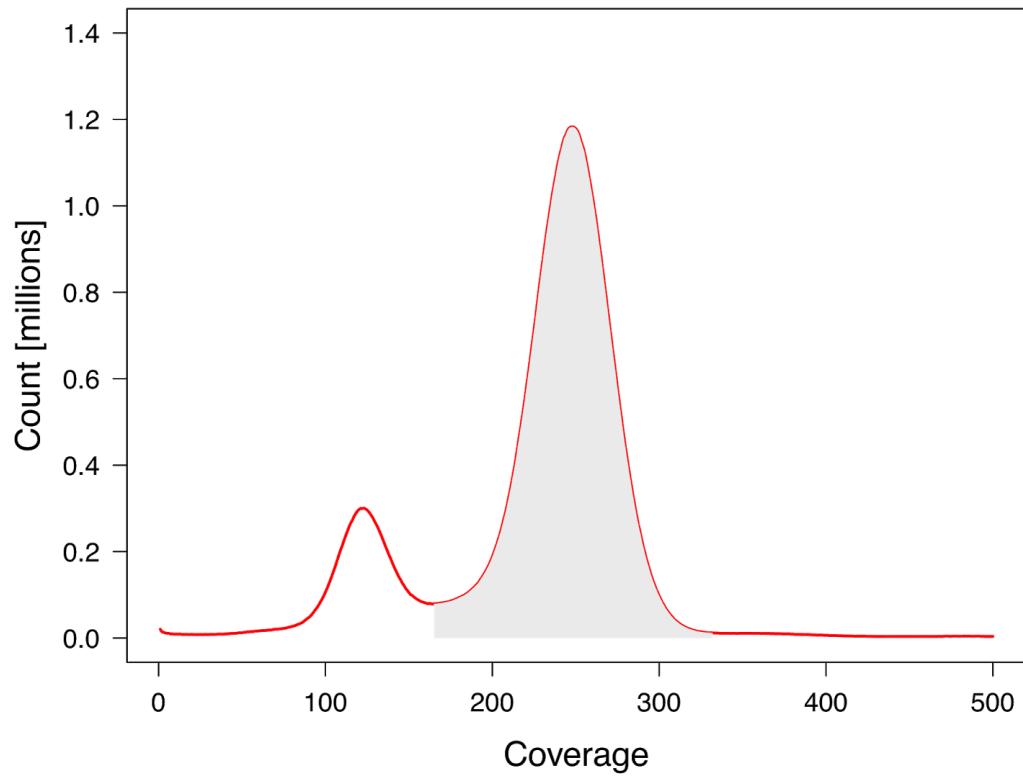


Figure S2. Per base genome coverage.

The grey area (81% of the assembly) marks bases of the reference assembly that are in the merged stage with a peak at 260x coverage.

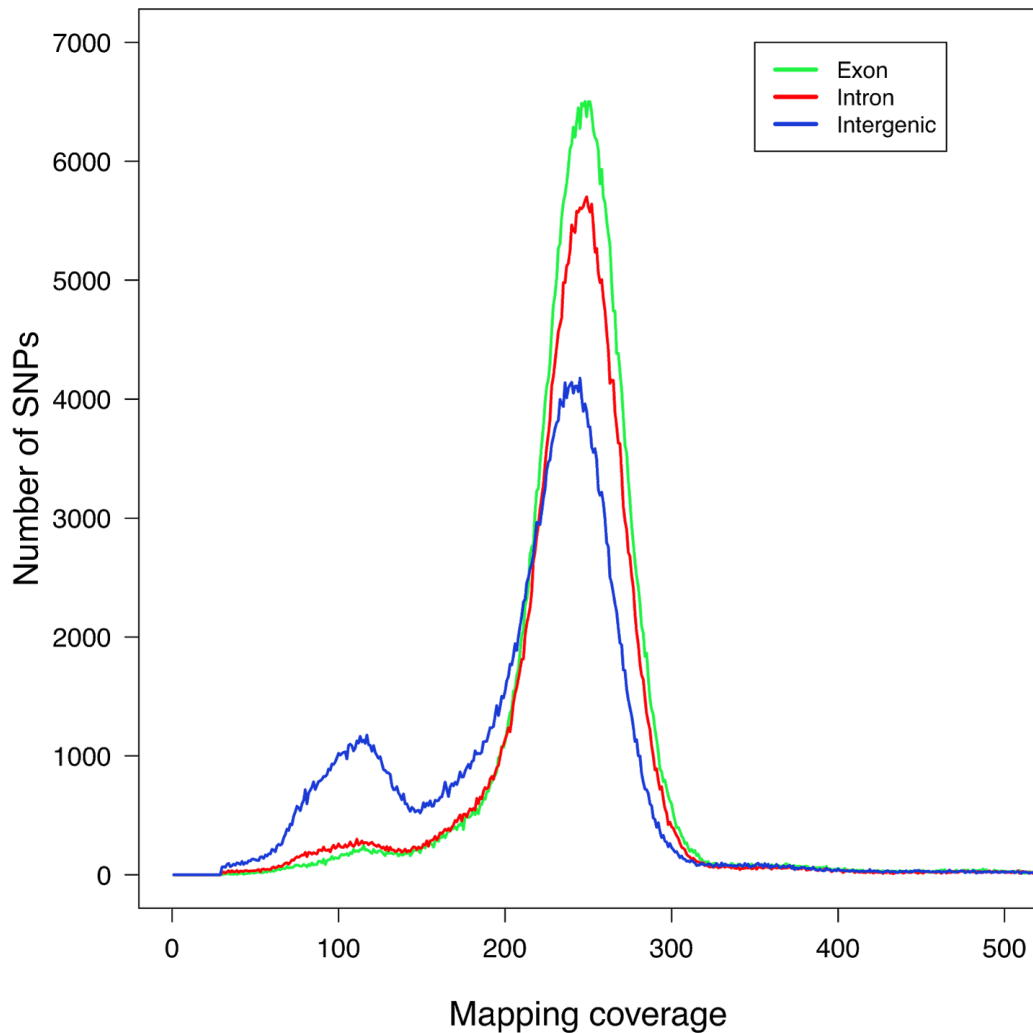


Figure S3. SNP histogram of genomic fractions.

Plotted are SNP counts in the exonic, intronic and intergenic genome fractions against the genome mapping coverage. The histogram shows that most of the genic (exonic and intronic) fraction were merged (peak at ~250x coverage) and further indicates a very low number of false gene duplications caused by genome misassembly.

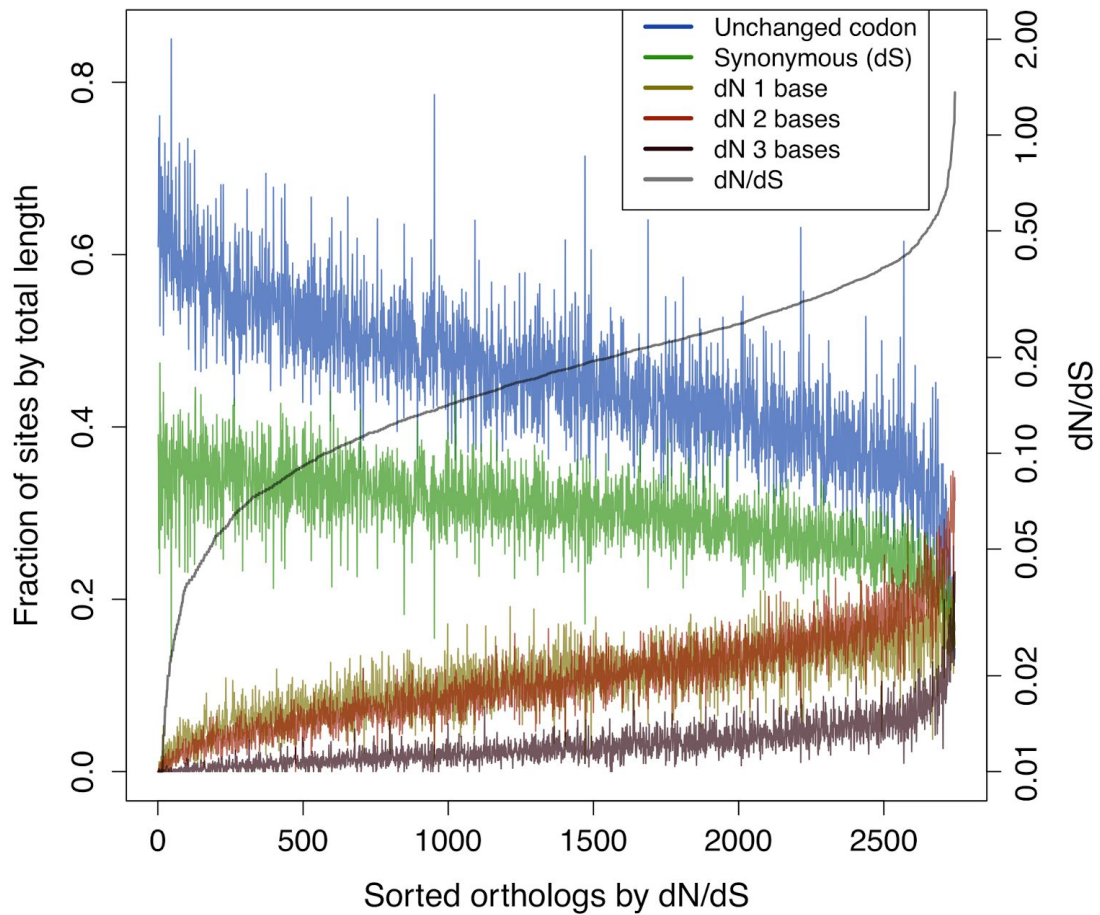


Figure S4. Evaluation of codon saturation.

Plotted are fractions of the full protein length for unchanged codons as well as synonymous and nonsynonymous sites for the 2,720 orthologs set. dN1, dN2, dN3 refer to non-synonymous sites with single, double, and triple base change, respectively. Orthologs are sorted by increasing dN/dS ratio. The percentage of unchanged sites is mostly above 40% (mean $45.8\% \pm 8.5$) and never drops below 14.2%. Third codon positions are thus never saturated and the three orthologs with $dN/dS > 1$ are truly positively selected.

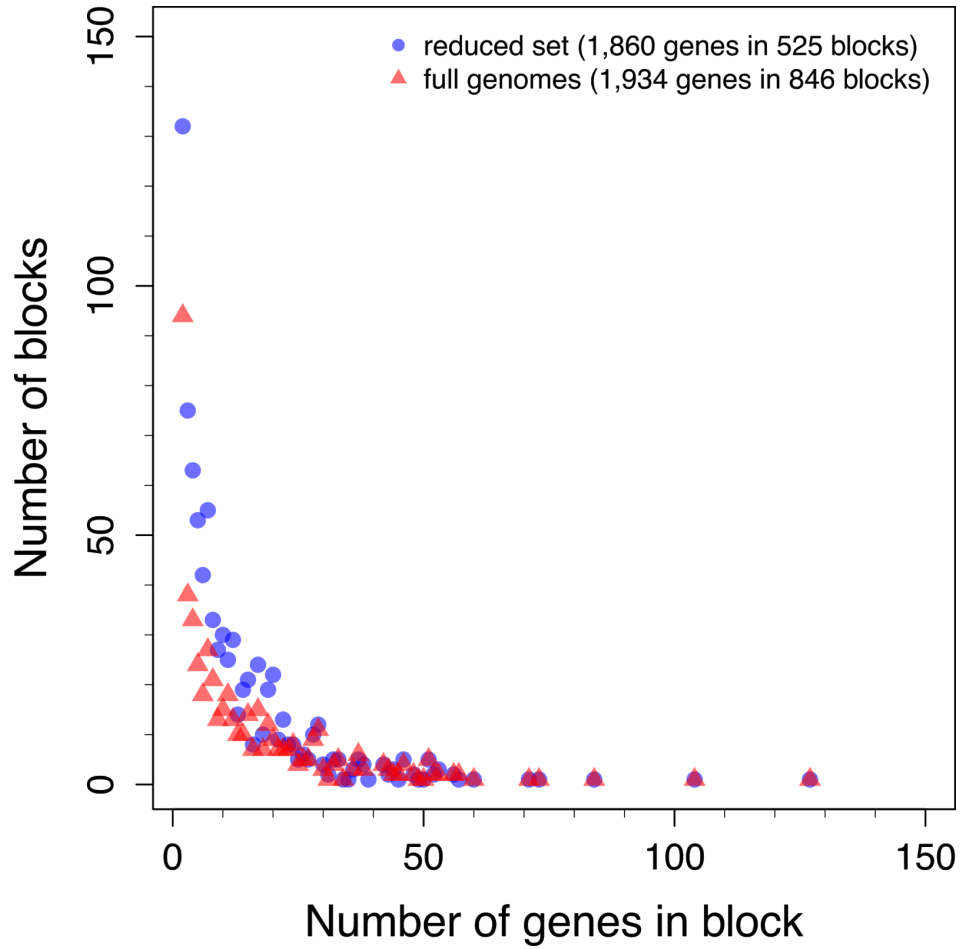


Figure S5. Length of syntenic blocks.

Shown are numbers of genes in detected syntenic blocks between a reduced set of *T. adhaerens* scaffolds and *X. yyyyyyyyyyyyyy* contigs (blue circles; same set as used for collinearity analyses, see also Figure 2 & Table S5) as well as between both whole genomes (red rectangles). Numbers of genes within blocks as well as numbers of blocks are in the same order of magnitude indicating that the reduced set is representative for full genomes.

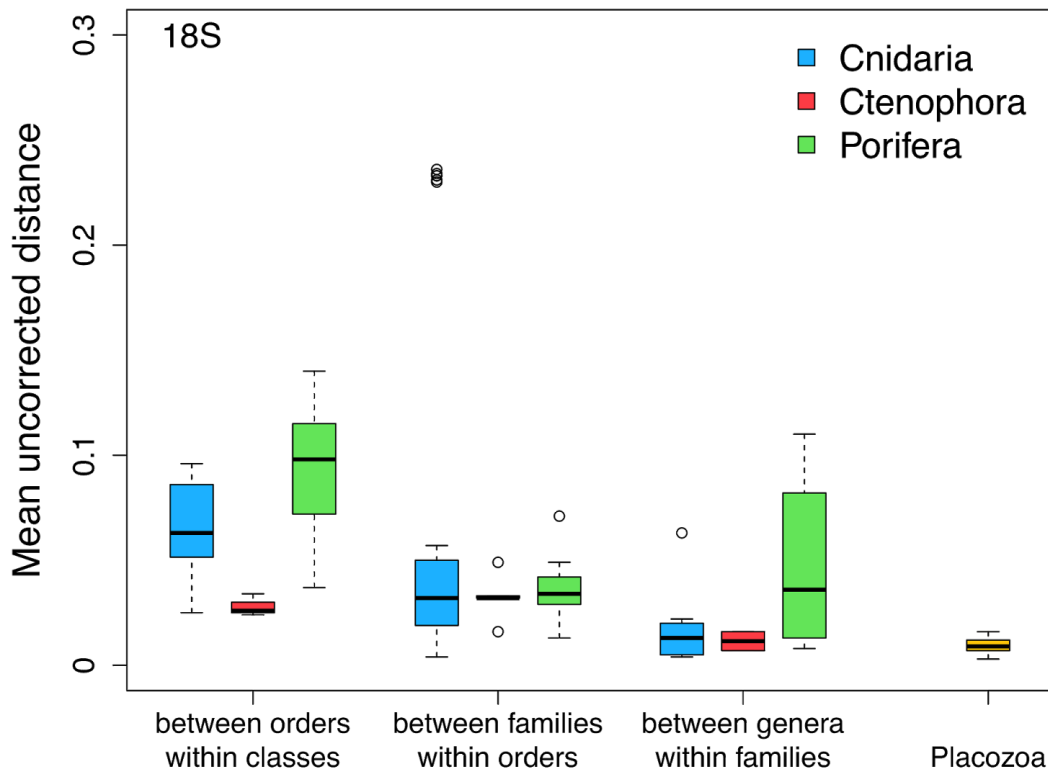


Figure S6. 18S genetic distances in non-bilateria.

Shown are mean group distances for different taxonomic ranks in the phyla Cnidaria, Ctenophora and Porifera: between orders within classes, between families within orders and between genera within families. Interspecific genetic distance between placozoans are shown on the right.

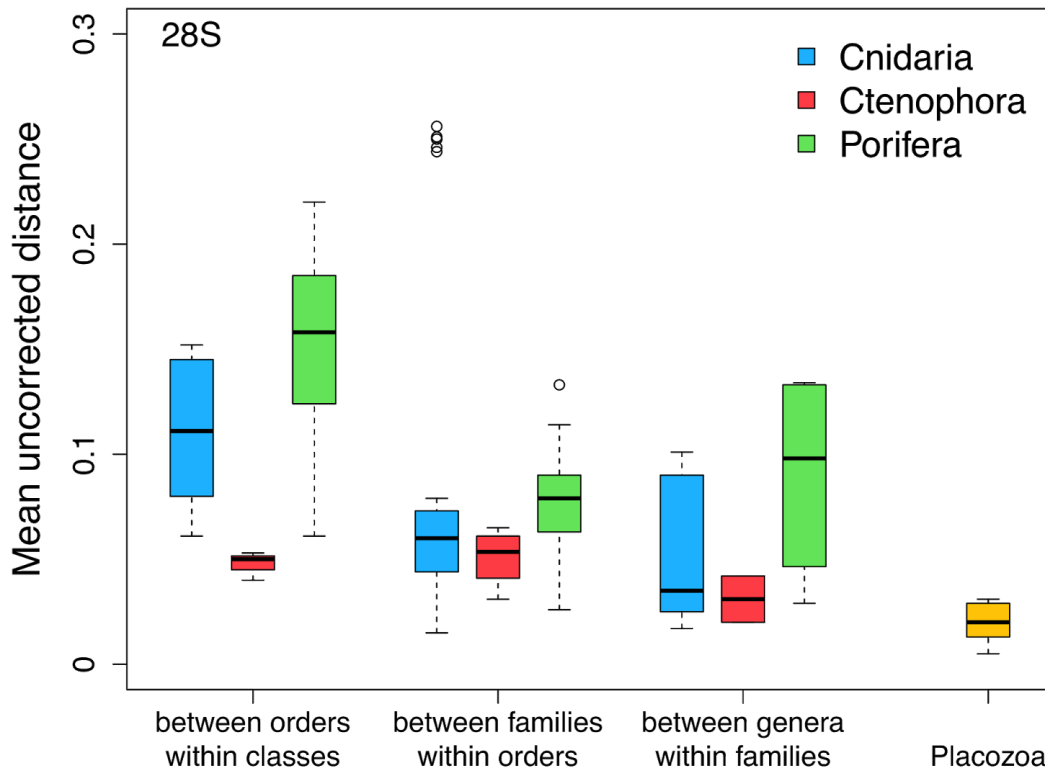


Figure S7. 28S genetic distances in non-bilateria.

Shown are mean group distances for different taxonomic ranks in the phyla Cnidaria, Ctenophora and Porifera: between orders within classes, between families within orders and between genera within families. Interspecific genetic distance between placozoans are shown on the right.

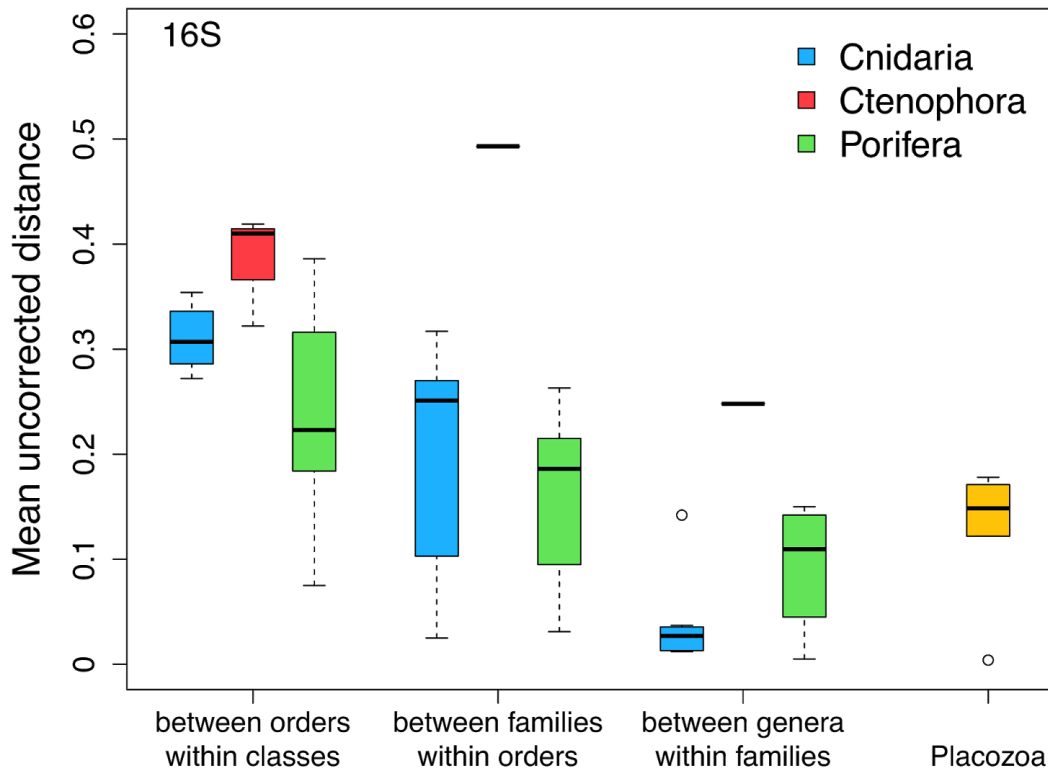


Figure S8. 16S genetic distances in non-bilateria.

Shown are mean group distances for different taxonomic ranks in the phyla Cnidaria, Ctenophora and Porifera: between orders within classes, between families within orders and between genera within families. Interspecific genetic distance between placozoans are shown on the right.

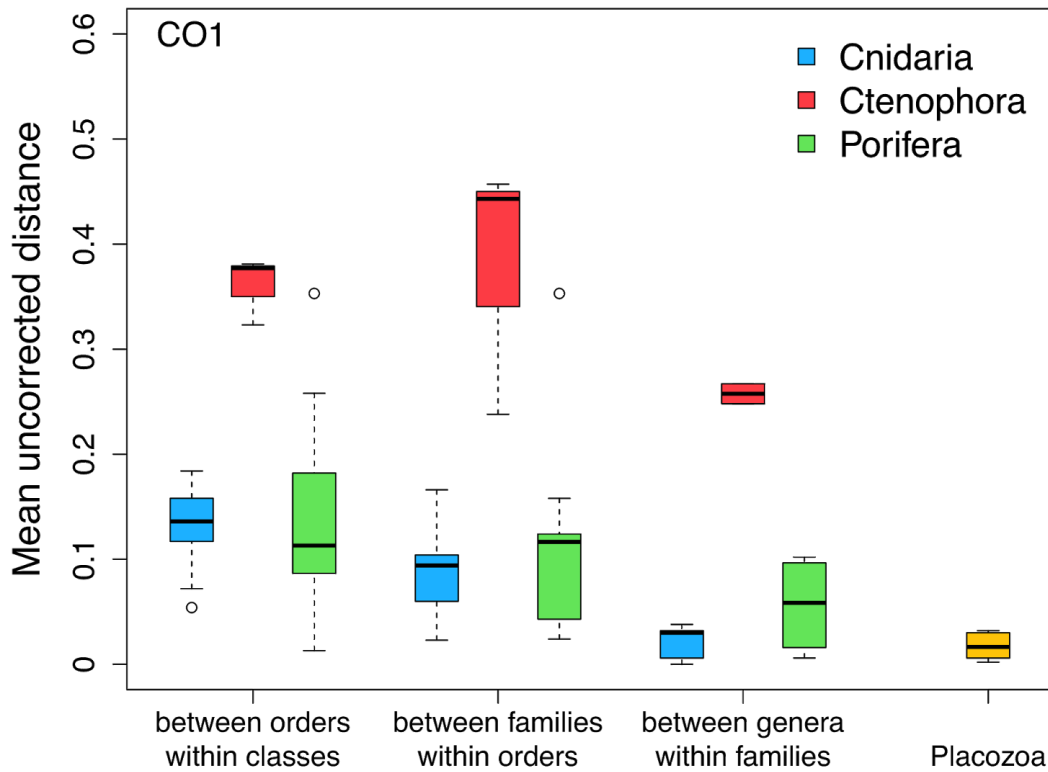


Figure S9. CO1 genetic distances in non-bilateria.

Shown are mean group distances for different taxonomic ranks in the phyla Cnidaria, Ctenophora and Porifera: between orders within classes, between families within orders and between genera within families. Interspecific genetic distance between placozoans are shown on the right.

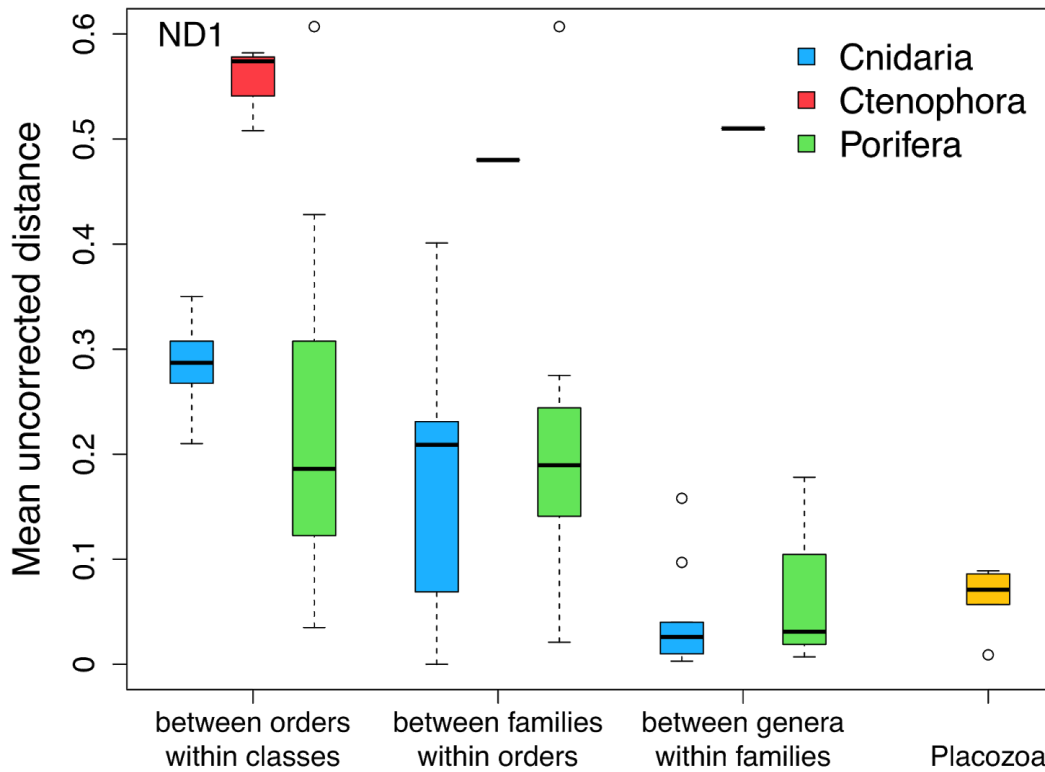


Figure S10. ND1 genetic distances in non-bilateria.

Shown are mean group distances for different taxonomic ranks in the phyla Cnidaria, Ctenophora and Porifera: between orders within classes, between families within orders and between genera within families. Interspecific genetic distance between placozoans are shown on the right.

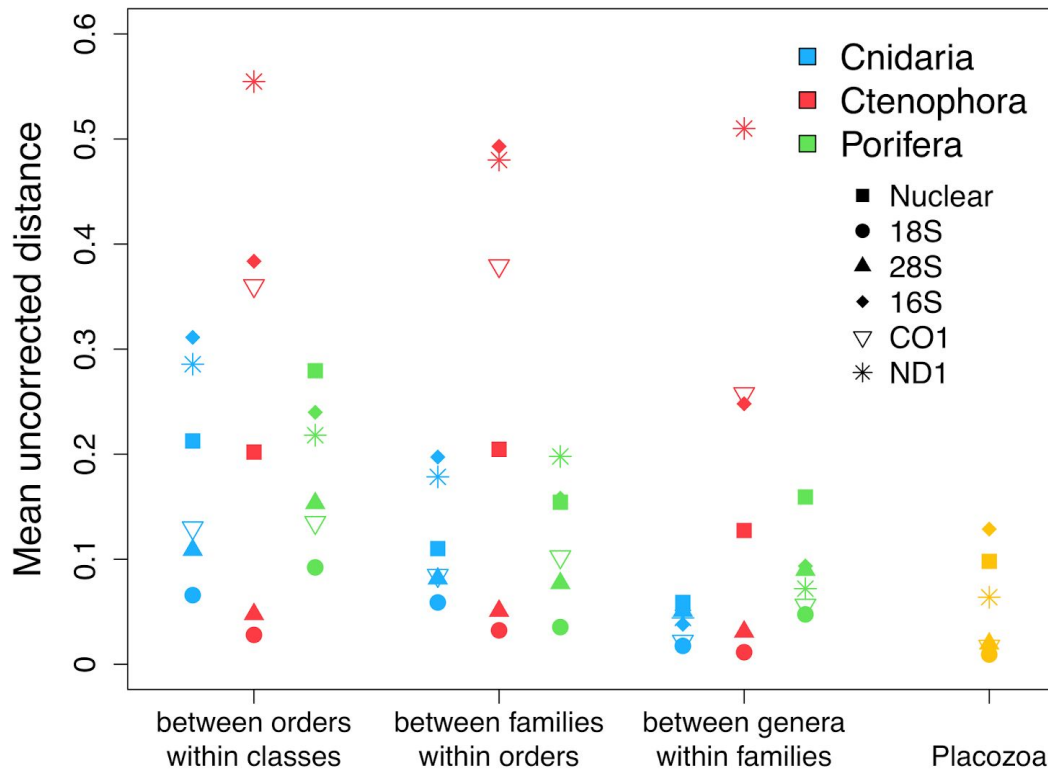


Figure S11. Summary of genetic distances of all markers.

Shown are the means of all mean group distances for different taxonomic ranks in the non-bilaterian phyla Cnidaria, Ctenophora and Porifera. Interspecific genetic distance between placozoans are shown on the right (yellow).

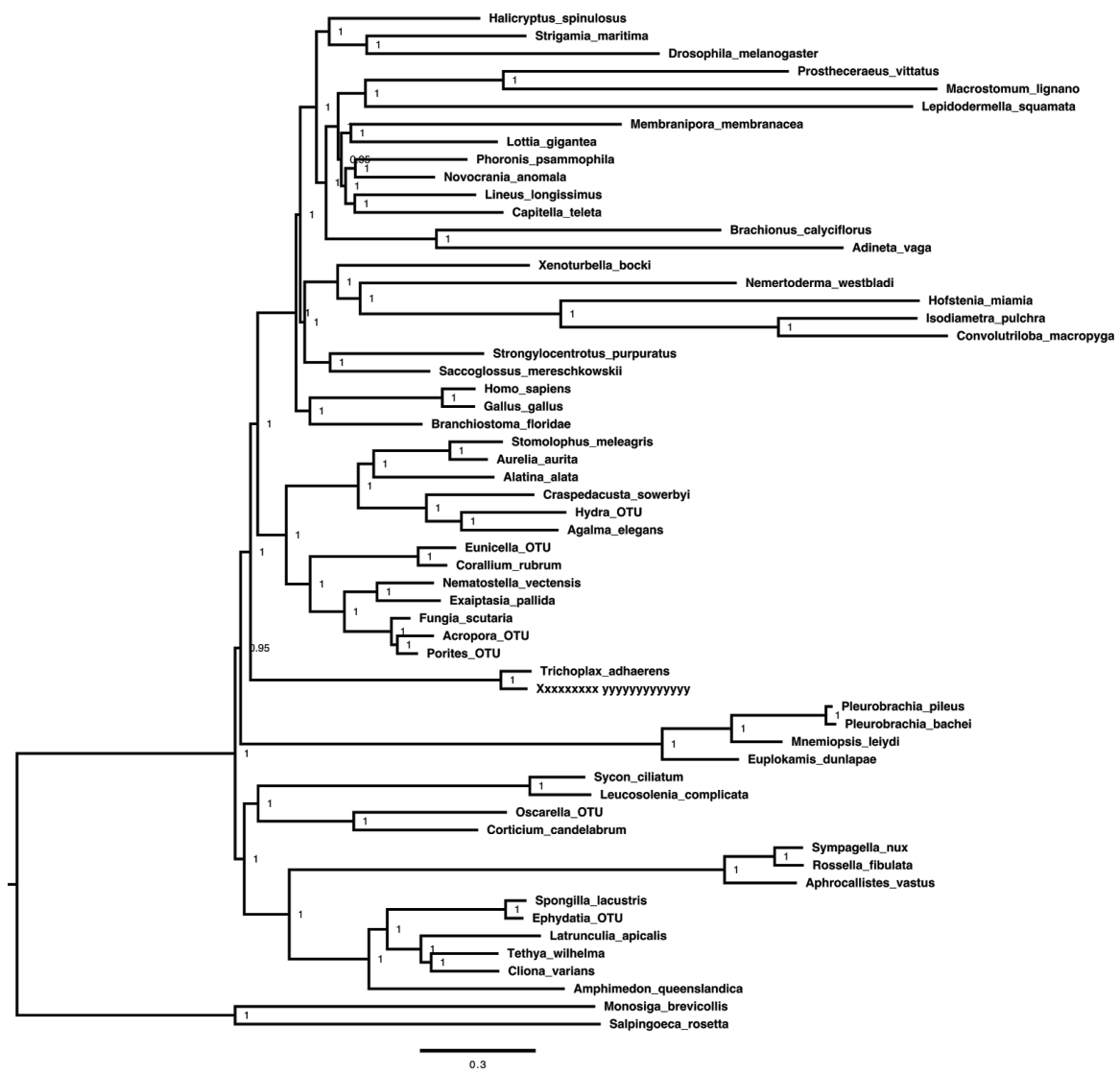


Figure S12. PhyloBayes tree of the dataset 2 protein matrix.

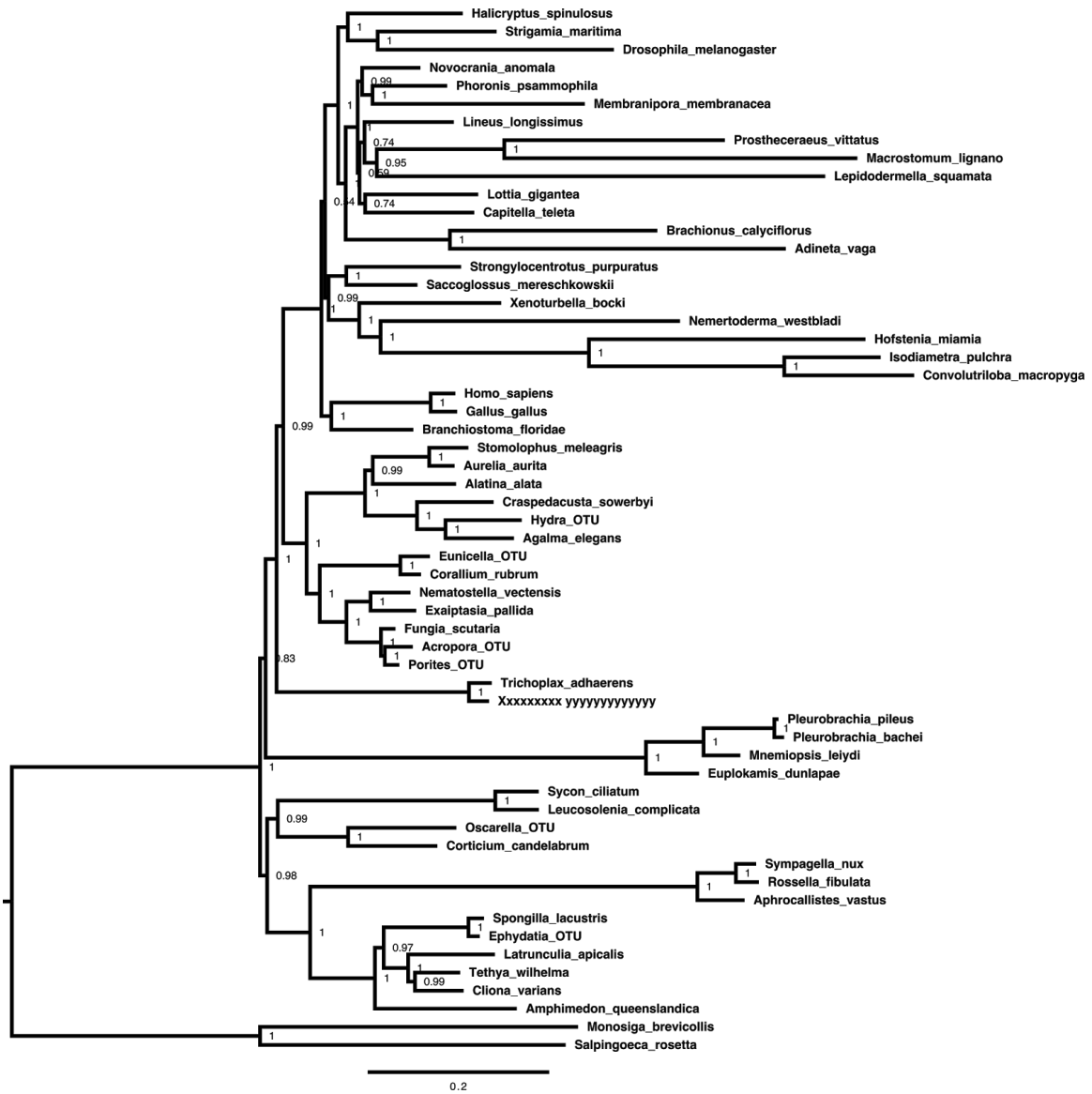


Figure S13. PhyloBayes tree of the dataset 2 Dayhoff-6 recoded matrix.

Table S1. Genome assembly statistics.

Xxxxxxxxxx *yyyyyyyyyyyyyy* genome assembly and annotation statistics in comparison to *Trichoplax adhaerens*.

		<i>Xxxxxxxxxx</i> <i>yyyyyyyyyyyyyy</i>	<i>Trichoplax</i> <i>adhaerens</i>
Contigs	assembly size	87.19Mb	94.75Mb
	numbers	669	3,217
	N50	407.8Kb	204.2Kb
	mean length	130.3kb	29.5kb
	longest	2.06Mb	803.8Kb
Scaffolds	assembly size	-	105.6Mb
	numbers	-	1,414
	N50	-	5.98Mb
	mean length	-	74.65kb
	longest	-	13.26Mb
	gaps	-	10.86Mb
mt genome size		36.5Kb	43.1Kb
GC content		31.30%	32.70%
JGI best gene models		-	11,520
BRAKER1 gene models	<i>ab initio</i> genes	12,010	12,521
	<i>ab initio</i> transcripts	12,575	26,796
	unexpressed <i>ab initio</i> genes	422 (3.5%)	-
Trinity transcripts	total transcripts	124,155	-
	N50	2,550	
	protein coding	58,838	-
StringTie transcripts	total transcripts	25,266	-
	protein coding	19,795	-

Table S2. Summary of SNP counts.

Shown are summaries from the genomic and transcriptomic datasets.

source	total SNP count	exonic SNP count	total exonic SNPs per 1kb CDS	unique exonic SNPs per 1kb CDS
RNA	302,430	138,302	6.4	1.0
DNA	1,397,488	202,901	9.5	4.0

Table S3. Read and transcript mapping statistics.

data type	reads or transcripts	mapped	mapping %
DNA reads	206,777,776	172,868,338	83.6
Moleculo (long) reads	79,974	74,626	93.3
RNAseq reads	114,475,046	92,154,755	80.5
Trinity mRNA	124,155	110,509	89.0
Trinity transdecoder mRNA	64,753	59,137	91.3
Trinity transdecoder CDS	64,753	58,838	90.9

Table S4. Results of the BUSCO v2 gene set searches.

protein set	complete (total)	complete (single)	complete (duplicate)	fragmented	missing
BRAKER1	92.4%	86.7%	5.7%	1.0%	6.6%
StringTie	89.5%	70.1%	19.4%	1.0%	9.5%
Trinity	93.0%	41.5%	51.5%	2.5%	4.5%

Table S5. Collinearity analyses.

Collinearity between the ten largest *Trichoplax adhaerens* (*T.a.*) scaffolds and the associated 144 *Xxxxxxxx yyyyyyyyyyyy* (*X.y.*) supercontigs larger than 100Kb.

<i>Trichoplax</i> scaffold ID	scaffold size [Mbp]	gene number in <i>T.a.</i> scaffold	collinear <i>X.y.</i> genes	percentage of collinear genes
ta1	13.2	1,581	1,199	75.8
ta2	9.7	1,168	887	75.9
ta3	9.6	1,109	812	73.2
ta4	8.6	1,027	789	76.8
ta5	8.2	955	664	69.5
ta6	5.9	757	592	78.2
ta7	5.8	669	527	78.8
ta8	3.4	342	205	59.9
ta9	3.2	373	278	74.5
ta10	2.4	279	206	73.8
total	70.0	8,260	6,159	73.6 (± 5.5)

Table S6. Transcriptomic and genomic data resources.

Used data for genetic distance calculations and phylogenetic inferences. A Trinity transcriptome assembly has been generated for each species with a given accession number. Otherwise transcriptomes were used from the reference.

phylum	species	Genbank accession	reference
Porifera	<i>Amphimedon queenslandica</i>		[1]
	<i>Aphrocallistes vastus</i>		[2,3]
	<i>Chondrilla nucula</i>		[2,3]
	<i>Cliona varians</i>	SRP028615	[4]
	<i>Corticium candelabrum</i>	SRP012620	[2,3]
	<i>Crambe crambe</i>	ERP008520	[5]
	<i>Crella elegans</i>	SRP017286	[6]
	<i>Ephydatia fluviatilis</i>		[7]
	<i>Ephydatia muelleri</i>		Nichols lab; through www.compagen.org
	<i>Halisarca dujardini</i>		[8]
	<i>Hyalonema populiferum</i>	SRP056162	[9]
	<i>Ircinia fasciculata</i>		[2,3]
	<i>Kirkpatrickia variolosa</i>	SRP056162	[9]
	<i>Latrunculia apicalis</i>	SRP056162	[9]
	<i>Lendenfeldia sp.</i>		Vargas <i>et al.</i> , unpubl.
	<i>Leucosolenia complicata</i>		Adamska lab; through www.compagen.org
	<i>Mycale phyllophila</i>	SRP051131	[10]
<i>Oscarella carmela</i>		Nichols lab; through www.compagen.org	

	<i>Oscarella sp.</i>		Nichols lab; through www.compagen.org
	<i>Petrosia ficiformis</i>	SRP012619	[2,3]
	<i>Pseudospongosorites suberitoides</i>	SRP037540	[2,3]
	<i>Rossella fibulata</i>	SRP056162	[9]
	<i>Scopalina sp.</i>	SRP061923	[11]
	<i>Spongilla lacustris</i>		[2,3]
	<i>Stylissa carteri</i>	SRP051559	[12]
	<i>Sycon ciliatum</i>		Adamska lab; through www.compagen.org
	<i>Sycon coactum</i>		[2,3]
	<i>Sympagella nux</i>	SRP056162	[9]
	<i>Tedania anhelans</i>	SRP061923	[11]
	<i>Tethya wilhelma</i>		[13]
	<i>Xestospongia testudinaria</i>	SRP051561	[12]
Cnidaria	<i>Abylopsis tetragona</i>		[14,15]
	<i>Acropora digitifera</i>		[16]
	<i>Acropora millepora</i>		[17,18]
	<i>Agalma elegans</i>		[14,15]
	<i>Alatina alata</i>	SRP056878	[15]
	<i>Anthopleura elegantissima</i>		[19]
	<i>Astreopora sp</i>		[19]
	<i>Aurelia aurita</i>		[15,20]
	<i>Chironex fleckeri</i>	SRP055535	[21]
	<i>Chrysaora fuscescens</i>	SRP070629	[22]
	<i>Clytia hemisphaerica</i>		www.compagen.org
	<i>Corallium rubrum</i>		[23]

<i>Craseoa lathetica</i>		[14,15]
<i>Craspedacusta sowerbyi</i>		[15,24]
<i>Cyanea capillata</i>	SRP056566	[25]
<i>Dendrogramma enigmatica</i>		[26]
<i>Ectopleura larynx</i>		[27]
<i>Eunicella cavolinii</i>	SRP042651	[28]
<i>Eunicella verrucosa</i>	SRP042651	[28]
<i>Exaiptasia pallida</i>		[15,29–31]
<i>Fungia scutaria</i>		[19]
<i>Gorgonia ventalina</i>		[32]
<i>Hydra magnipapillata</i>		[19,33]
<i>Hydra vulgaris</i>		[34]
<i>Hydractinia symbiolongicarpus</i>		[15,35,36]
<i>Madracis auretenra</i>		[19]
<i>Montastraea cavernosa</i>		[19]
<i>Nanomia bijuga</i>		[15,37]
<i>Nematostella vectensis</i>		[38,39]
<i>Physalia physalis</i>		[14,15]
<i>Pocillopora damicornis</i>		[40]
<i>Porites astreoides</i>		[19]
<i>Porites australiensis</i>		[19]
<i>Pseudodiploria strigosa</i>		[19]
<i>Rhopilema esculentum</i>	SRP073155	[41]
<i>Seriatopora hystrix</i>		[19]
<i>Seriatopora sp</i>		[19]
<i>Siderastrea siderea</i>		[42]
<i>Stomolophus meleagris</i>	SRP037742	[43]

	<i>Stylophora pistillata</i>		[44]
Ctenophora	<i>Beroe abyssicola</i>		[45]
	<i>Beroe sp</i>		[46]
	<i>Bolinopsis infundibulum</i>		[45]
	<i>Coeloplana atericola</i>		[45]
	<i>Dryodora glandiformis</i>		[45]
	<i>Euplokamis dunlapae</i>		[45]
	<i>Hormiphora californensis</i>	SRP057600	[47]
	<i>Mertensiidae sp</i>		[45]
	<i>Mnemiopsis leidyi</i>		[45,47]
	<i>Pleurobrachia bachei</i>		[45]
	<i>Pleurobrachia pileus</i>	SRP001155	[45]
	<i>Vallicula multiformis</i>		[45]
Placozoa	<i>Trichoplax adhaerens</i>		[48]

Table S7. Created OTUs.

Operational Taxonomic Units (OUTs) used for distance calculations (dataset 1, all) and phylogenetic analyses (dataset 2, as specified). The first species each participated most.

OTU	used species	in dataset 2
Acropora	<i>Acropora digitifera</i> , <i>Acropora millepora</i>	+
Ephydatia	<i>Ephydatia fluviatilis</i> , <i>Ephydatia muelleri</i>	+
Hydra	<i>Hydra vulgaris</i> , <i>Hydra magnipapillata</i>	+
Porites	<i>Porites astreoides</i> , <i>Porites australiensis</i>	+
Seriatopora	<i>Seriatopora hystrix</i> , <i>Seriatopora sp</i>	-
Beroe	<i>Beroe abyssicola</i> , <i>Beroe sp</i>	-
Eunicella	<i>Eunicella cavolinii</i> , <i>Eunicella verrucosa</i>	+
Oscarella	<i>Oscarella carmela</i> , <i>Oscarella sp</i>	+

Table S8. Datasets used for distance calculation and phylogenetic inferences.

Summary of protein datasets used for distance calculations (dataset 1) and phylogenetic inferences (dataset 2). Matrix length is 38,869 amino acid characters (dataset 1) and 35,799 (dataset 2), respectively.

data- set #	description	total taxa	outgroup taxa	non- bilaterian taxa	% missing characters
1	full dataset	139	4	77	29.0
2	100% gene content in outgroup taxa; overall gene coverage >50%	58	2	32	10.8

Table S9. Genetic distances (%) between the two placozoan species in comparison to Cnidaria.

Shown are averages of all mean genetic distances (\pm SD) between genera within families and families within orders for the Cnidaria as well as genetic distances between *X. yyyyyyyyyyyyyy* (*X.y.*) and *T. adhaerens* (*T.a.*). Number of all mean pairwise distances used to calculate the average are given in parentheses.

taxonomic level	212 nuclear proteins	18S	28S	16S	CO1	ND1
Cnidarian genera	5.9 \pm 2.7 (10)	3.8 \pm 4.3 (8)	1.8 \pm 1.9 (9)	5.1 \pm 3.4 (9)	2.2 \pm 1.5 (9)	4.4 \pm 5.1 (9)
Cnidarian families	11.0 \pm 4.2 (41)	5.9 \pm 7.4 (41)	8.2 \pm 7.2 (41)	19.7 \pm 9.4 (26)	8.5 \pm 3.8 (37)	17.8 \pm 10.5 (37)
<i>X.y. / T.a.</i>	9.8	1.2	2.9	17.8	3.0	8.6

Table S10. Accession numbers of used mitochondrial genomes.

phylum	class	species	accession
Porifera	Calcarea	<i>Leucosolenia complicata</i>	KT997975.1, KT997966.1
		<i>Sycon ciliatum</i>	KU244276.1, KU244278.1
		<i>Sycon coactum</i>	-
Demospongiae		<i>Amphimedon queenslandica</i>	NC_008944.1
		<i>Chondrilla nucula</i>	NC_010208
		<i>Cliona varians</i>	-
		<i>Crambe crambe</i>	-
		<i>Crella elegans</i>	NC_027520.1
		<i>Ephydatia muelleri</i>	NC_010202.1
		<i>Halisarca dujardini</i>	NC_010212.1
		<i>Ircinia fasciculata</i>	-
		<i>Kirkpatrickia variolosa</i>	-
		<i>Latrunculia apicalis</i>	-
		<i>Lendenfeldia sp.</i>	-
		<i>Mycale phyllophila</i>	-
		<i>Petrosia ficiformis</i>	NC_027519.1
		<i>Pseudospongosorites suberitoides</i>	-
		<i>Scopalina sp.</i>	-
		<i>Spongilla lacustris</i>	LT158503.1, KU759841.1
		<i>Stylissa carteri</i>	-
<i>Tedania anhelans</i>	-		
<i>Tethya wilhelma</i>	-		
<i>Xestospongia testudinaria</i>	-		

	Hexactinellida	<i>Aphrocallistes vastus</i>	NC_010769.1
		<i>Hyalonema populiferum</i>	-
		<i>Rossella fibulata</i>	-
		<i>Sympagella nux</i>	EF537577.1
	Homoscleromorpha	<i>Corticium candelabrum</i>	NC_014872.1
		<i>Oscarella carmela</i>	NC_009090.1
Cnidaria	Anthozoa	<i>Acropora digitifera</i>	NC_022830.1
		<i>Anthopleura elegantissima</i>	-
		<i>Corallium rubrum</i>	-
		<i>Eunicella verrucosa</i>	-
		<i>Exaiptasia pallida</i>	-
		<i>Fungia scutaria</i>	-
		<i>Gorgonia ventalina</i>	-
		<i>Madracis auretenra</i>	-
		<i>Montastraea cavernosa</i>	-
		<i>Nematostella vectensis</i>	NC_008164.1
		<i>Pocillopora damicornis</i>	NC_009797.1
		<i>Porites astreoides</i>	-
		<i>Pseudodiploria strigosa</i>	
		<i>Seriatopora hystrix</i>	NC_010244.1
		<i>Siderastrea siderea</i>	-
		<i>Stylophora pistillata</i>	NC_011162.1
	Cubozoa	<i>Alatina alata</i>	-
		<i>Chironex fleckeri</i>	-
	Hydrozoa	<i>Abylopsis tetragona</i>	-
		<i>Agalma elegans</i>	-
		<i>Clytia hemisphaerica</i>	-

		<i>Craseoa lathetica</i>	-
		<i>Craspedacusta sowerbyi</i>	NC_018537.1
		<i>Dendrogramma enigmatica</i>	-
		<i>Ectopleura larynx</i>	-
		<i>Hydra vulgaris</i>	NC_011221.1
		<i>Hydractinia symbiolongicarpus</i>	LN901197.1
		<i>Nanomia bijuga</i>	-
		<i>Physalia physalis</i>	KT809328.1
	Scyphozoa	<i>Aurelia aurita</i>	NC_008446.1
		<i>Chrysaora fuscescens</i>	-
		<i>Cyanea capillata</i>	JN700937.1
		<i>Rhopilema esculentum</i>	
		<i>Stomolophus meleagris</i>	
Ctenophora	Tentaculata	<i>Bolinopsis infundibulum</i>	-
		<i>Coeloplana atericola</i>	-
		<i>Dryodora glandiformis</i>	-
		<i>Euplokamis dunlapae</i>	-
		<i>Hormiphora californensis</i>	-
		<i>Mertensiidae sp</i>	-
		<i>Mnemiopsis leidyi</i>	NC_016117.1
		<i>Pleurobrachia bachei</i>	NC_016697.1
		<i>Pleuronrachia pileus</i>	-
		<i>Vallicula multiformis</i>	-
Placozoa		<i>Trichoplax adhaerens</i>	NC_008151.2
		Placozoa sp. H4	NC_008833.2
		Placozoa sp. H8	NC_008832.2

Supplemental References

1. Fernandez-Valverde, S.L., Calcino, A.D., and Degnan, B.M. (2015). Deep developmental transcriptome sequencing uncovers numerous new genes and enhances gene annotation in the sponge *Amphimedon queenslandica*. *BMC Genomics* *16*, 720.
2. Riesgo, A., Farrar, N., Windsor, P.J., Giribet, G., and Leys, S.P. (2014). The analysis of eight transcriptomes from all poriferan classes reveals surprising genetic complexity in sponges. *Mol. Biol. Evol.* *31*, 1102–1120.
3. Ludeman, D.A., Farrar, N., Riesgo, A., Paps, J., and Leys, S.P. (2014). Evolutionary origins of sensation in metazoans: functional evidence for a new sensory organ in sponges. *BMC Evol. Biol.*, 1–11.
4. Riesgo, A., Peterson, K., Richardson, C., Heist, T., Strehlow, B., McCauley, M., Cotman, C., Hill, M., and Hill, A. (2014). Transcriptomic analysis of differential host gene expression upon uptake of symbionts: a case study with *Symbiodinium* and the major bioeroding sponge *Cliona varians*. *BMC Genomics* *15*, 376.
5. Versluis, D., D’Andrea, M.M., Ramiro Garcia, J., Leimena, M.M., Hugenholtz, F., Zhang, J., Öztürk, B., Nylund, L., Sipkema, D., van Schaik, W., *et al.* (2015). Mining microbial metatranscriptomes for expression of antibiotic resistance genes under natural conditions. *Sci. Rep.* *5*, 11981.
6. Pérez-Porro, A.R.P., Navarro-Gomez, D., Uriz, M.J., and Giribet, G. (2013). A NGS approach to the encrusting Mediterranean sponge *Crella elegans* (Porifera, Demospongiae, Poecilosclerida): transcriptome sequencing, characterization and overview of the gene expression along three life cycle stages. *Molecular Ecology Resources* *13*, 494–509.
7. Alié, A., Hayashi, T., Sugimura, I., Manuel, M., Sugano, W., Mano, A., Satoh, N., Agata, K., and Funayama, N. (2015). The ancestral gene repertoire of animal stem cells. *Proc. Natl. Acad. Sci. U. S. A.* *112*, E7093–100.
8. Borisenko, I., Adamski, M., Ereskovsky, A., and Adamska, M. (2016). Surprisingly rich repertoire of Wnt genes in the demosponge *Halisarca dujardini*. *BMC Evol. Biol.* *16*, 123.
9. Whelan, N.V., Kocot, K.M., Moroz, L.L., and Halanych, K.M. (2015). Error, signal, and the placement of Ctenophora sister to all other animals. *Proc. Natl. Acad. Sci. U. S. A.* *112*, 5773–5778.
10. Qiu, F., Ding, S., Ou, H., Wang, D., Chen, J., and Miyamoto, M.M. (2015).

Transcriptome Changes during the Life Cycle of the Red Sponge, *Mycale phyllophila* (Porifera, Demospongiae, Poecilosclerida). *Genes* 6, 1023–1052.

11. Díez-Vives, C., Moitinho-Silva, L., Nielsen, S., Reynolds, D., and Thomas, T. (2017). Expression of eukaryotic-like protein in the microbiome of sponges. *Mol. Ecol.* 26, 1432–1451.
12. Ryu, T., Seridi, L., Moitinho-Silva, L., Oates, M., Liew, Y.J., Mavromatis, C., Wang, X., Haywood, A., Lafi, F.F., Kupresanin, M., *et al.* (2016). Hologenome analysis of two marine sponges with different microbiomes. *BMC Genomics* 17, 158.
13. Francis, W.R., Eitel, M., Vargas, S., and Adamski, M. (2017). The Genome Of The Contractile Demosponge *Tethya wilhelma* And The Evolution Of Metazoan Neural Signalling Pathways. *bioRxiv*. <http://biorxiv.org/content/early/2017/03/27/120998>.
14. Dunn, C.W., Howison, M., and Zapata, F. (2013). Agalma: an automated phylogenomics workflow. *BMC Bioinformatics*, 1–9.
15. Zapata, F., Goetz, F.E., Smith, S.A., Howison, M., Siebert, S., Church, S.H., Sanders, S.M., Ames, C.L., McFadden, C.S., France, S.C., *et al.* (2015). Phylogenomic Analyses Support Traditional Relationships within Cnidaria. *PLoS One* 10, e0139068.
16. Shinzato, C., Shoguchi, E., Kawashima, T., Hamada, M., Hisata, K., Tanaka, M., Fujie, M., Fujiwara, M., Koyanagi, R., Ikuta, T., *et al.* (2011). Using the *Acropora digitifera* genome to understand coral responses to environmental change. *Nature* 476, 320–323.
17. Moya, A., Huisman, L., Ball, E.E., Hayward, D.C., Grasso, L.C., Chua, C.M., Woo, H.N., Gattuso, J.-P., Forêt, S., and Miller, D.J. (2012). Whole Transcriptome Analysis of the Coral *Acropora millepora* Reveals Complex Responses to CO₂-driven Acidification during the Initiation of Calcification. *Mol. Ecol.* 21, 2440–2454.
18. Weiss, Y., Forêt, S., Hayward, D.C., Ainsworth, T., King, R., Ball, E.E., and Miller, D.J. (2013). The acute transcriptional response of the coral *Acropora millepora* to immune challenge: expression of GiMAP/IAN genes links the innate immune responses of corals with those of mammals and plants. *BMC Genomics* 14, 400.
19. Bhattacharya, D., Agrawal, S., Aranda, M., Baumgarten, S., Belcaid, M., Drake, J.L., Erwin, D., Foret, S., Gates, R.D., Gruber, D.F., *et al.* (2016). Comparative genomics explains the evolutionary success of reef-forming corals. *Elife* 5.

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4878875>.

20. Brekhman, V., Malik, A., Haas, B., Sher, N., and Lotan, T. (2015). Transcriptome profiling of the dynamic life cycle of the scyphozoan jellyfish *Aurelia aurita*. *BMC Genomics* *16*, 74.
21. Brinkman, D.L., Jia, X., Potriquet, J., Kumar, D., Dash, D., Kvaskoff, D., and Mulvenna, J. (2015). Transcriptome and venom proteome of the box jellyfish *Chironex fleckeri*. *BMC Genomics* *16*, 407.
22. Ponce, D., Brinkman, D.L., Potriquet, J., and Mulvenna, J. (2016). Tentacle Transcriptome and Venom Proteome of the Pacific Sea Nettle, *Chrysaora fuscescens* (Cnidaria: Scyphozoa). *Toxins* *8*, 102.
23. Pratlong, M., Haguenaer, A., Chabrol, O., Klopp, C., Pontarotti, P., and Aurelle, D. (2015). The red coral (*Corallium rubrum*) transcriptome: a new resource for population genetics and local adaptation studies. *Mol. Ecol. Resour.* *15*, 1205–1215.
24. Hroudova, M., Vojta, P., Strnad, H., Krejcik, Z., Ridl, J., Paces, J., Vleck, C., and Paces, V. (2012). Diversity, phylogeny and expression patterns of Pou and Six homeodomain transcription factors in hydrozoan jellyfish *Craspedacusta sowerbyi*. *PLoS One* *7*, e36420.
25. Liu, G., Zhou, Y., Liu, D., Wang, Q., Ruan, Z., He, Q., and Zhang, L. (2015). Global Transcriptome Analysis of the Tentacle of the Jellyfish *Cyanea capillata* Using Deep Sequencing and Expressed Sequence Tags: Insight into the Toxin- and Degenerative Disease-Related Transcripts. *PLoS One* *10*, e0142680.
26. O'Hara, T.D., Hugall, A.F., MacIntosh, H., Naughton, K.M., Williams, A., and Moussalli, A. (2016). *Dendrogramma* is a siphonophore. *Curr. Biol.* *26*, R457–R458.
27. Fidler, A.L., Vanacore, R.M., Chetyrkin, S.V., Pedchenko, V.K., Bhave, G., Yin, V.P., Stothers, C.L., Rose, K.L., McDonald, W.H., Clark, T.A., *et al.* (2014). A unique covalent bond in basement membrane is a primordial innovation for tissue evolution. *Proc. Natl. Acad. Sci. U. S. A.* *111*, 331–336.
28. Romiguier, J., Gayral, P., Ballenghien, M., Bernard, A., Cahais, V., Chenuil, A., Chiari, Y., Dernet, R., Duret, L., Faivre, N., *et al.* (2014). Comparative population genomics in animals uncovers the determinants of genetic diversity. *Nature* *515*, 261–263.
29. Lehnert, E.M., Burriesci, M.S., and Pringle, J.R. (2012). Developing the anemone *Aiptasia* as a tractable model for cnidarian-dinoflagellate symbiosis: the

- transcriptome of aposymbiotic *A. pallida*. *BMC Genomics* *13*, 271.
30. Lehnert, E.M., Mouchka, M.E., Burriesci, M.S., Gallo, N.D., Schwarz, J.A., and Pringle, J.R. (2014). Extensive differences in gene expression between symbiotic and aposymbiotic cnidarians. *G3* *4*, 277–295.
 31. Baumgarten, S., Simakov, O., Esherick, L.Y., Liew, Y.J., Lehnert, E.M., Michell, C.T., Li, Y., Hambleton, E.A., Guse, A., Oates, M.E., *et al.* (2015). The genome of *Aiptasia*, a sea anemone model for coral symbiosis. *Proc. Natl. Acad. Sci. U. S. A.* *112*, 11893–11898.
 32. Burge, C.A., Mouchka, M.E., Harvell, C.D., and Roberts, S. (2013). Immune response of the Caribbean sea fan, *Gorgonia ventalina*, exposed to an *Aplanochytrium* parasite as revealed by transcriptome sequencing. *Front. Physiol.* *4*, 180.
 33. Chapman, J.A., Kirkness, E.F., Simakov, O., Hampson, S.E., Mitros, T., Weinmaier, T., Rattei, T., Balasubramanian, P.G., Borman, J., Busam, D., *et al.* (2010). The dynamic genome of *Hydra*. *Nature* *464*, 592–596.
 34. Wenger, Y., and Galliot, B. (2013). RNAseq versus genome-predicted transcriptomes: a large population of novel transcripts identified in an Illumina-454 *Hydra* transcriptome. *BMC Genomics* *14*, 204.
 35. Sanders, S.M., Shcheglovitova, M., and Cartwright, P. (2014). Differential gene expression between functionally specialized polyps of the colonial hydrozoan *Hydractinia symbiolongicarpus* (Phylum Cnidaria). *BMC Genomics* *15*, 406.
 36. Plachetzki, D.C., Sabrina Pankey, M., Johnson, B.R., Ronne, E.J., Kopp, A., and Grosberg, R.K. (2014). Gene co-expression modules underlying polymorphic and monomorphic zooids in the colonial hydrozoan, *Hydractinia symbiolongicarpus*. *Integr. Comp. Biol.* *54*, 276–283.
 37. Siebert, S., Robinson, M.D., Tintori, S.C., Goetz, F., Helm, R.R., Smith, S.A., Shaner, N., Haddock, S.H.D., and Dunn, C.W. (2011). Differential Gene Expression in the Siphonophore *Nanomia bijuga* (Cnidaria) Assessed with Multiple Next-Generation Sequencing Workflows. *PLoS One* *6*, e22953.
 38. Putnam, N.H., Srivastava, M., Hellsten, U., Dirks, B., Chapman, J., Salamov, A., Terry, A., Shapiro, H., Lindquist, E., Kapitonov, V.V., *et al.* (2007). Sea anemone genome reveals ancestral eumetazoan gene repertoire and genomic organization. *Science* *317*, 86–94.
 39. Moran, Y., Fredman, D., Praher, D., Li, X.Z., Wee, L.M., Rentzsch, F., Zamore,

- P.D., Technau, U., and Seitz, H. (2014). Cnidarian microRNAs frequently regulate targets by cleavage. *Genome Res.* *24*, 651–663.
40. Vidal-Dupiol, J., Dheilly, N.M., Rondon, R., Grunau, C., Cosseau, C., Smith, K.M., Freitag, M., Adjeroud, M., and Mitta, G. (2014). Thermal Stress Triggers Broad *Pocillopora damicornis* Transcriptomic Remodeling, while *Vibrio coralliilyticus* Infection Induces a More Targeted Immuno-Suppression Response. *PLoS One*, 1–15.
 41. Li, Y., Zhou, Z., Tian, M., Tian, Y., Dong, Y., Li, S., Liu, W., and He, C. (2017). Exploring single nucleotide polymorphism (SNP), microsatellite (SSR) and differentially expressed genes in the jellyfish (*Rhopilema esculentum*) by transcriptome sequencing. *Mar. Genomics*.
<http://dx.doi.org/10.1016/j.margen.2017.01.007>.
 42. Davies, S.W., Marchetti, A., Ries, J.B., and Castillo, K.D. (2016). Thermal and pCO₂ Stress Elicit Divergent Transcriptomic Responses in a Resilient Coral. *Front. Mar. Sci.* *3*.
 43. Li, R., Yu, H., Xue, W., Yue, Y., Liu, S., Xing, R., and Li, P. (2014). Jellyfish venomomics and venom gland transcriptomics analysis of *Stomolophus meleagris* to reveal the toxins associated with sting. *J. Proteomics* *106*, 17–29.
 44. Liew, Y.J., Aranda, M., Carr, A., Baumgarten, S., Zoccola, D., Tambutté, S., Allemand, D., Micklem, G., and Voolstra, C.R. (2014). Identification of microRNAs in the coral *Stylophora pistillata*. *PLoS One* *9*, e91101.
 45. Moroz, L.L., Kocot, K.M., Citarella, M.R., Dosung, S., Norekian, T.P., Povolotskaya, I.S., Grigorenko, A.P., Dailey, C., Berezikov, E., Buckley, K.M., *et al.* (2014). The ctenophore genome and the evolutionary origins of neural systems. *Nature* *510*, 109–114.
 46. Nosenko, T., Schreiber, F., Adamska, M., Adamski, M., Eitel, M., Hammel, J., Maldonado, M., Müller, W.E.G., Nickel, M., Schierwater, B., *et al.* (2013). Deep metazoan phylogeny: when different genes tell different stories. *Mol. Phylogenet. Evol.* *67*, 223–233.
 47. Ryan, J.F., Pang, K., Schnitzler, C.E., Nguyen, A.D., Moreland, R.T., Simmons, D.K., Koch, B.J., Francis, W.R., Havlak, P., Smith, S.A., *et al.* (2013). The genome of the ctenophore *Mnemiopsis leidyi* and its implications for cell type evolution. *Science* *342*, 1242592.
 48. Srivastava, M., Begovic, E., Chapman, J., Putnam, N.H., Hellsten, U., Kawashima, T., Kuo, A., Mitros, T., Salamov, A., Carpenter, M.L., *et al.* (2008). The *Trichoplax*

genome and the nature of placozoans. *Nature* 454, 955–960.