

Sample code	Strain	BDP-acceptor
151	<i>C. coli</i> 76339	Lac
171	<i>C. coli</i> 76339	LacNAc
191	<i>C. coli</i> 76339	SiaLac
152	<i>C. coli</i> 76339 $\Delta cstV$	Lac
172	<i>C. coli</i> 76339 $\Delta cstV$	LacNAc
192	<i>C. coli</i> 76339 $\Delta cstV$	SiaLac

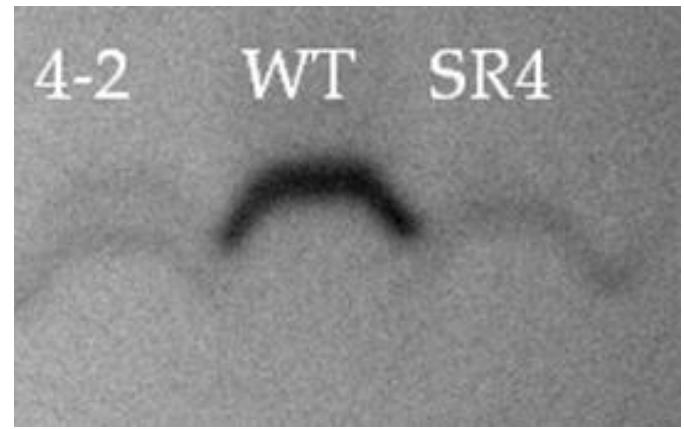
Supplemental Figure 1. Example of enzymatic test. Sialyltransferase activity of protein extracts was tested on BODIPY labelled Lac, LacNAc, and 3'Sialyllactose or FCHASE labelled α -GalNAc, β -GalNAc, GM3, α -Gal, β -GlcNAc, α -Glc, β -Glc, Hep, as donors. Reactions were performed at 37 °C in 10 μ l volumes containing 50 mM HEPES pH 7.5, 10 mM MgCl₂, 1 mM CMP-NeuAc, 0.5 mM labelled acceptor, and 6 μ l of extract. Enzymatic activity was assessed by thin-layer chromatography on silica using a solvent system of ethyl acetate/methanol/water/acetic acid 4:2:1:0.1.

Samples were incubated for 0.5, 1, 1.5, and 3 hours, and overnight (ON)

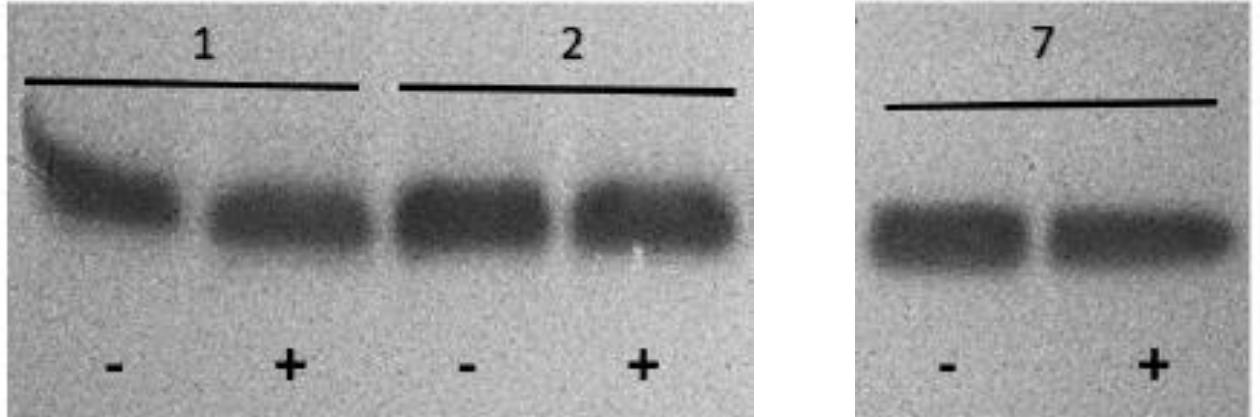


Primer name	Sequence
ProEx-cstIV-F-NdeI	GGGGGGGGGCCATATGAATAAAAATTAAATTTCACA
ProEx-cstIV-RW-SalII	ACCGTCGACCTATCTTGTATCTTGTGGGA
ProEx-cstV-R-NdeI	CCATATGATAGAAAACAATGCAGTTGTTGCA
ProEx-cstV-FW-SalII	ACCGTCGACTTATTGCCTTTTATATTAAAAA

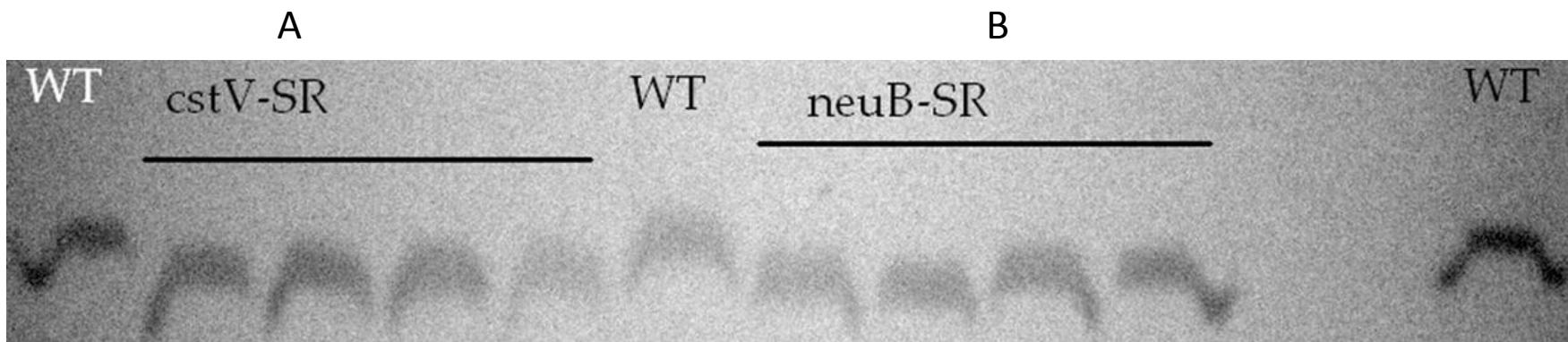
Supplemental Figure 2. Graphic representation of CstV protein expression construct. Gene *cstIV* from *C. coli* 73 and *cstV* from *C. coli* 76339 inserted into pCw and pCw-MaLET plamids. *E. coli* AD202 or *E. coli* BL21 were used as expression hosts



Supplemental Figure. 3 Electrophoresis mobility comparison of *C. coli* 76339 Δ cstV Δ ggt:cstV (4-2), *C. coli* 76339 WT, and *C. coli* 76339 Δ cstV (SR4).



Supplemental Figure 4. Silver stain analysis of neuraminidase treated crude LOS. 1. *C. coli* 76339 (*cstV*); 2. *C. coli* 65 (*cstIV*); 7. *C. coli* 73 (*cstIV*); (+) neuraminidase treated, (-) control.



Supplemental Figure 5. Electrophoresis mobility comparison of *C. coli* 76339 WT, $\Delta cstV$, and $\Delta neuB$. **A;** *C. coli* 76339 WT, $\Delta cstV$ -SR (erythromycin resistance cassette cloned in same direction as *cstV*). **B;** $\Delta neuB$ -SR (erythromycin resistance cassette cloned in same direction as *neuB*).

Bi-Cj-cstII	MKKVIIAGNGPSLKEIDYSLRLPNDFDVFCRNQFYFEDKYYLGKKCKAVFVNPILEFFCQYYTLKHL	65
Cj-cstI	MTRTRMENELIVSKNMQNIIAGNGPSLKNINYKRLPREYDVFRCNQFYFEDKYYLGKKIKAVFFNPGEVLCQYHTAKQL	80
cstIV	-----MNKNLNFTKKSVLIAGNNGPSLKEIDYSLLPKEYDIFRCN E FYEDKYYLGKKIKAAFIWIPYEFIE E YTQMOKM	73
cstV	-----MIENNAVVVAGNGPSLKEIDYSLRPKEFDVFCRNQFYFEDKYYLGKKVKAASFSPGVEFEQYYTLNTL	68
HAC1267	-----MNKKPLIIAGNGPSIKLDYALFPKDFDVFRCRNQFYFEDKYYLGREIKGVFNAHVEDIQMKITKAI	67
HAC1268	-----MGTIKKPLIIAGNGPSIKLDYALFPKDFDVFRCRNQFYFEDKYYLGREIKGVFNPCLSSQMTVQYL	69
HBS-02	-----MPLKPLIVAGNGPSIKLDYALDYSLEPPDFDVFRCRNQFYFEDKYYLGKEVKGVFFNAQVEDMOMKTAREL	67
Pm70	-----MDKFAEHEIPKAVIVAGNGPSLQIDYRLLPKNYDVFRCRNQFYFEERYFLGNKIKAVFFTGPCVLECYYTLYHL	74
Bi-Cj-cstII	ICNQEYETEILIMCSNYNQAHLENEN----FVKTIFYDYFPDAHLGY-DFFKQLKDFNAYFKFHEIYFNQ--RITSGVYMC	137
Cj-cstI	ILKNEYEIKNIFCSTFNLPIESND----FLHQEYNFFPDALKGY-EVLENKEFYAYIKYNEIMFNK--RITSGVYMC	152
cstIV	LCNGDYEONIVCKMYNFQDRKE----KIFRENKEKYFFPAAIINGYDAFFYKIKELNSNMIDEFCMEYNTTE E YTIVI	148
cstV	MQNKEYYCCENIVCKLFLPLQHEINQKSL-RNFKKIFPLFFFYALDGNEHYFNKIKELNSFINENFLYDEG-IQITIGMYAI	146
HAC1267	VKNGEYHPDHYCCTHVEPYGYVNGNQQ--LMQEYLEKHFGVRSTYAYLKDLPEPFFILHSKRYRNFDQ--HFTTGIMML	142
HAC1268	MDGGEYSIBRFFCSVSTDRLHDFDGYQTILPVDGYLKAHYPFVCDTFSLFKGHEEILKHKVYHLKTYSK--ELSAGVMLM	147
HBS-02	SLRQEYYEDIECSTIAPFMNFGNHYT--HAQYLQYLDKHYPGARNTYALLQSLEPFYKLYTTRRNFYQO--HFTTGIMMI	142
Pm70	KRNNEYFVIVNVLSSSFNHPPTVDLEK----SQKIQALFIDVINGYEKLSKTAFDVYLRYKELYENQ--RITSGVYMC	146
Bi-Cj-cstII	AVAIAGYKEIYLGSIDFYQN-GSSYAFDTKQKNLLKLAPNKFKNDNSHYICHGSKNTDIKALEFTEKTYKIKLYCOPNSL	216
Cj-cstI	AIAIAIALGYKIYLCGIDFYEG-DVIMPFEAMSTNIKTIIFPGIK-DFKPNSQHSKEYDIEALKLIKSYKVNTYALCDDSI	230
cstIV	COAVACGTYKEIYLAGMDFGDE-KYNYFSEKIEK----IKESKKR-- T DKMHKSIDLKILDFIQQQYNVKLESICOPNSS	220
cstV	ACAVACGTYKEIYLGIDFYST-QEYAFDIKDKIGLYALNPSFKIQ--YIKSHSKETDLEIILSFLIKQTYNANEESISPKSP	223
HAC1267	LVAIQLGYKEIYLCGIDFYENGFGH YE --NQGGFFEEDSDPMHDKNIDICALELAKKY--AKIYALVPNSA	210
HAC1268	LSAVVILGYKEIYLVGIDFGASSWG HE YDE--SQSCHFSNHMADOCNIIYYDMLTICLCKQKY--AKIYALAPNSP	216
HBS-02	IVAIIVGYKEIYCAGLDFYLEGLGH YH --VKSHPHTLAPDCQHTKDIDIKGIEVAKQY--ACIYALVPNSA	210
Pm70	AVAIANGYTDIYLIGIDFYQASEENYAFDNKKPNIIRLLPDRKEKTLFSYH SKD IDLEALSFIQQHYHVNBYSISPMSP	226
Bi-Cj-cstII	LANFTIPIAPN-----	226
Cj-cstI	LANHFPPISTIN-----	240
cstIV	I N AFITPIPHPI-----	230
cstV	MTKYIPIIAPK-----	233
HAC1267	LVKMIPISSQKGVLEVKDRIGLGEFKREKFGQKELERQKELERQKELERQKELERQKELERQKELER	290
HAC1268	LSHLUTNPQAKYPPFELLDKP-IGYTSQDLIISSPLEEKLL E FKNIEEKLL E FKNIEEKLL E FKNIEEKLL	295
HBS-02	LSAIIIPISPHKNALSQEK-----	228
Pm70	LSKHPPIPTV-----	236
Bi-Cj-cstII	-----LNSN--FIIQEKNN-YTKDILIPSSEAYGKF SKN INF-----	260
Cj-cstI	-----INNN--FTLENKHNN S INDIILTDNTPGVSFYKNQLKAD--- N KIMLNFYNILH	289
cstIV	-----QANENIFKPIERPKDAIKTQ L TPPIKAVRRYKR-TYLES---NIIKFFHELQ	280
cstV	-----Q N YS--FDIEEKSSSES I KDFLIPSKKAYRNYSRALY L QN---NMFYNFIDCLK	282
HAC1267	QKELERQKELERQKELERQKELERQKELERQKELERQKELERQKELERQKELERQKELER	370
HAC1268	E F KNIEEKLL E FKNIEEKLL E FKNIEEKLL E FKNIEEKLL E FKNIEEKLL E FKNIEEKLL	375
HBS-02	-----M A ALKLGDPKPNGYIDDVCVP E FSMRATLIQAIQ S VGITP N LIYKGLNMVWR	283
Pm70	-----E D CETTFVAPLKENYINDI L LP H FVY E KLGT I VSKKS R FHS N LIVRLIR D LLK	291
Bi-Cj-cstII	-----	260
Cj-cstI	SKDN I IKFLNK-----	300
cstIV	V P RR I RHYYSKTRYTR-----	296
cstV	FPSA I KNYF K NIKK-----	296
HAC1267	LFKGCFALLD-----LKALKSIIKAFLKR-----	395
HAC1268	E F KNIEEKLLASRLNNI R KIKRKILPFFWGGGVPTLKV S FRWGA A	422
HBS-02	CASD I YRVV R G-----LYRLSLKALYFLRAWFKHRATGG-----	318
Pm70	LPSA I KHYLKEK-----	303

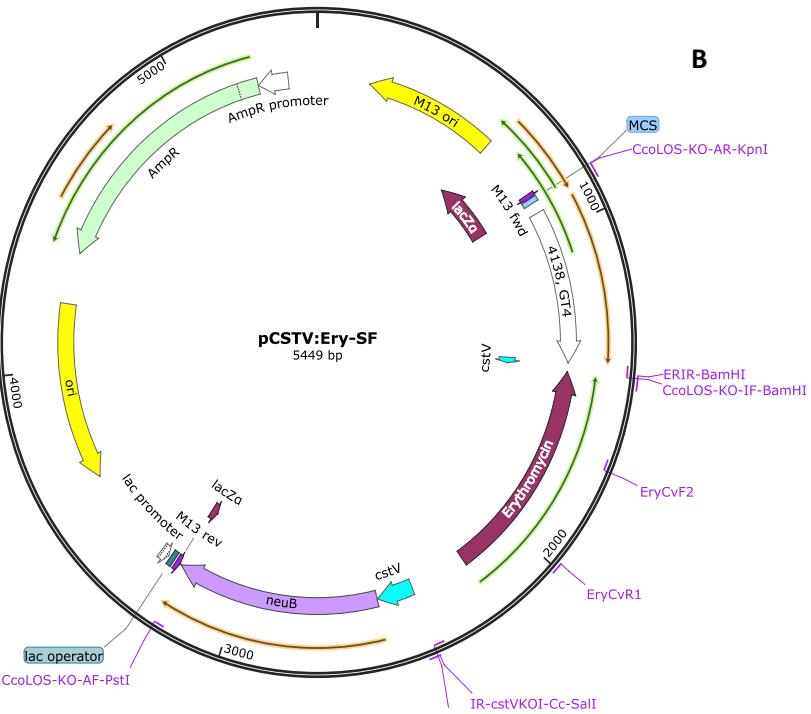
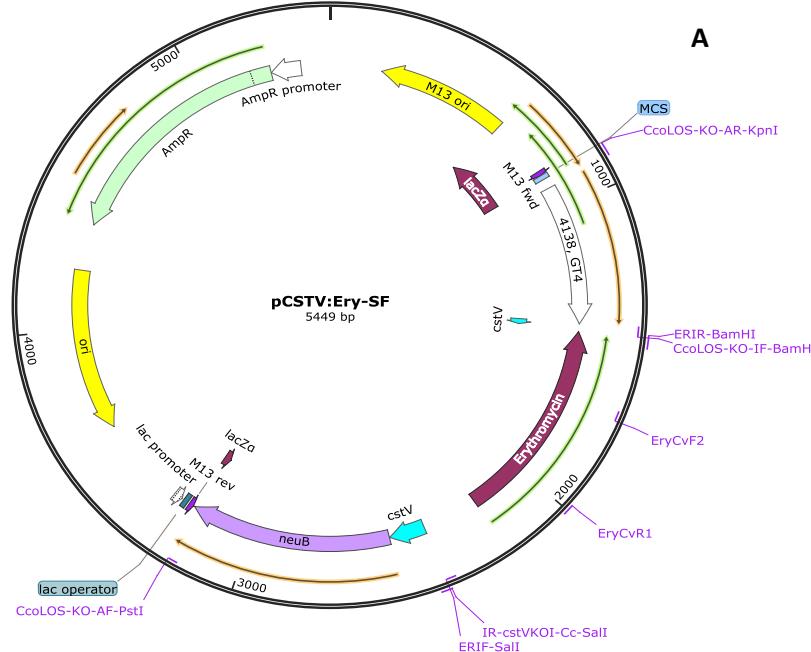
Important residues for CMP	
CstII	CstIV
Thr131	Leu142
Ser132	Thr143
Important residues for Neu5Ac	
CstII	CstIV
Gln32	His40
Asn51	Val59
Gln58	Glu66
Arg129	Glu140
Ser132	Thr143
Tyr185	Thr189

Supplemental Figure 6. Multiple sequence alignment of characterized GT-42 sialyltransferases and *C. coli* 76339 *cstV* and *C. coli* 73 *cstIV*.

Supplemental Material

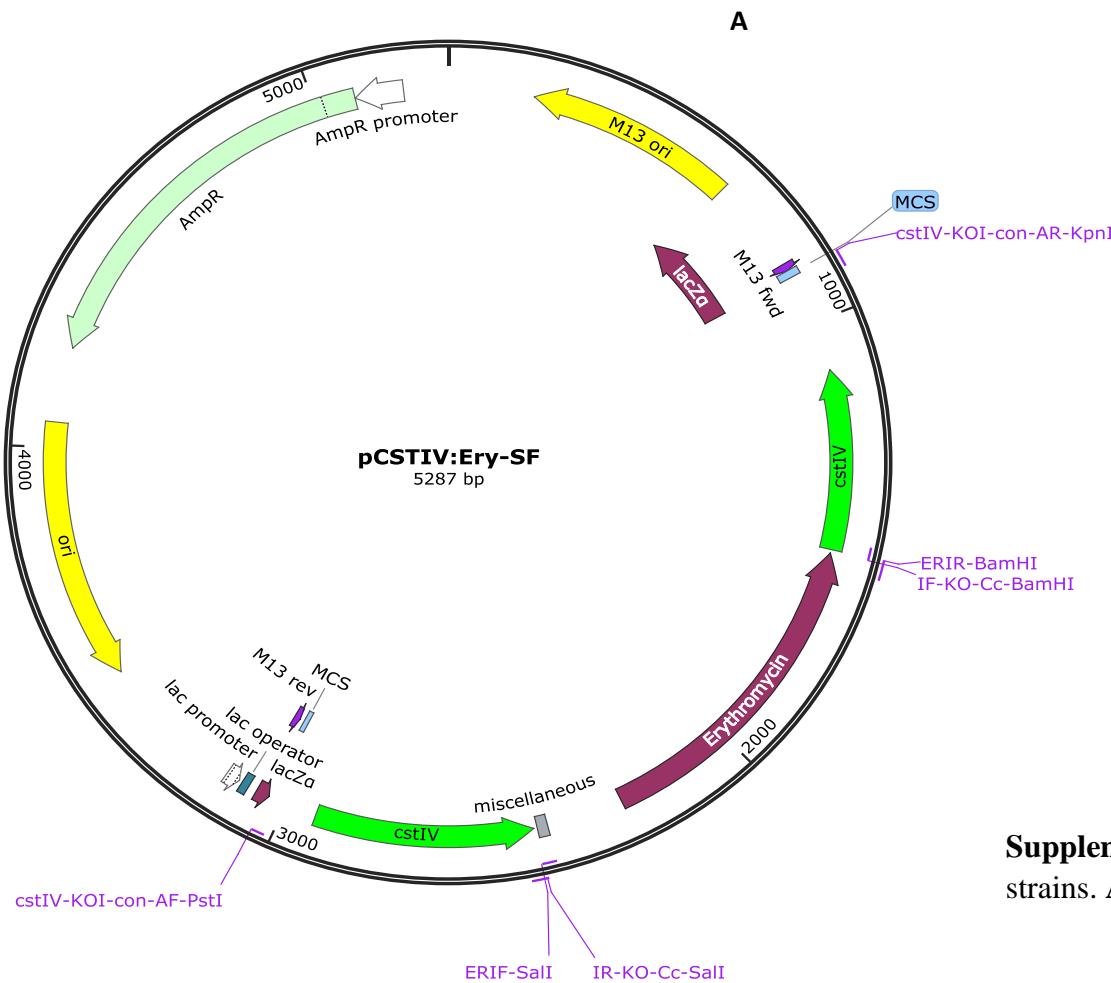
Supplemental Table 1. Bacterial strains and protein expression constructs

Strain or plasmid	Genotype and/or phenotype	Reference or source
<i>C. coli</i> 76339	<i>cstI</i> , <i>cstV</i> , <i>neuB</i>	¹
<i>C. coli</i> 76339 Δ <i>cstV</i> -SF1	<i>cstI</i> , Δ <i>cstV</i> : Ery, <i>neuB</i>	This study
<i>C. coli</i> 76339 Δ <i>cstV</i> -SR4	<i>cstI</i> , Δ <i>cstV</i> : Ery, <i>neuB</i>	This study
<i>C. coli</i> 76339 Δ <i>neuB</i> -SR1	<i>cstI</i> , Δ <i>neuB</i> : Ery, <i>cstV</i>	This study
<i>C. coli</i> 76339 Δ <i>cstI</i> -XR3	Δ <i>cstI</i> : CAT, <i>cstV</i> , <i>neuB</i>	This study
<i>C. coli</i> 76339 Δ <i>cstV</i> -SR4 Δ <i>cstI</i> -XR1	Δ <i>cstI</i> : CAT, Δ <i>cstV</i> : Ery, <i>neuB</i>	This study
<i>C. coli</i> 76339 Δ <i>neuB</i> -SR2	<i>cstI</i> , Δ <i>neuB</i> : Ery, <i>cstV</i>	This study
<i>C. coli</i> 76339 Δ <i>cstV</i> -SR4 Δ <i>ggt:cstV</i> -2	<i>cstI</i> , Δ <i>cstV</i> : Ery, <i>neuB</i> , Δ <i>ggt:cstV</i> :CAT	This study
<i>C. coli</i> 65	<i>cstIV</i>	²
<i>C. coli</i> 65 Δ <i>cstIV</i> -SF5	Δ <i>cstIV</i> :Ery	This study
<i>C. coli</i> 73	<i>cstIV</i>	²
<i>C. coli</i> 73 Δ <i>cstIV</i> -SF3	Δ <i>cstIV</i> :Ery	This study
<i>C. coli</i> 73 Δ <i>neuB</i> 2	Δ <i>neuB</i> 2:Ery	This study
<i>C. jejuni</i> 81-176		
<i>E. coli</i> AD202		³
<i>E. coli</i> BL21		
Protein expression constructs		
pCwMal-ET		³
pCw		
pCwMal-ET_51cstV		This study
pCwMal-ET_73cstIV		This study
pCw_51cstV		This study
pCw_73cstIV		This study



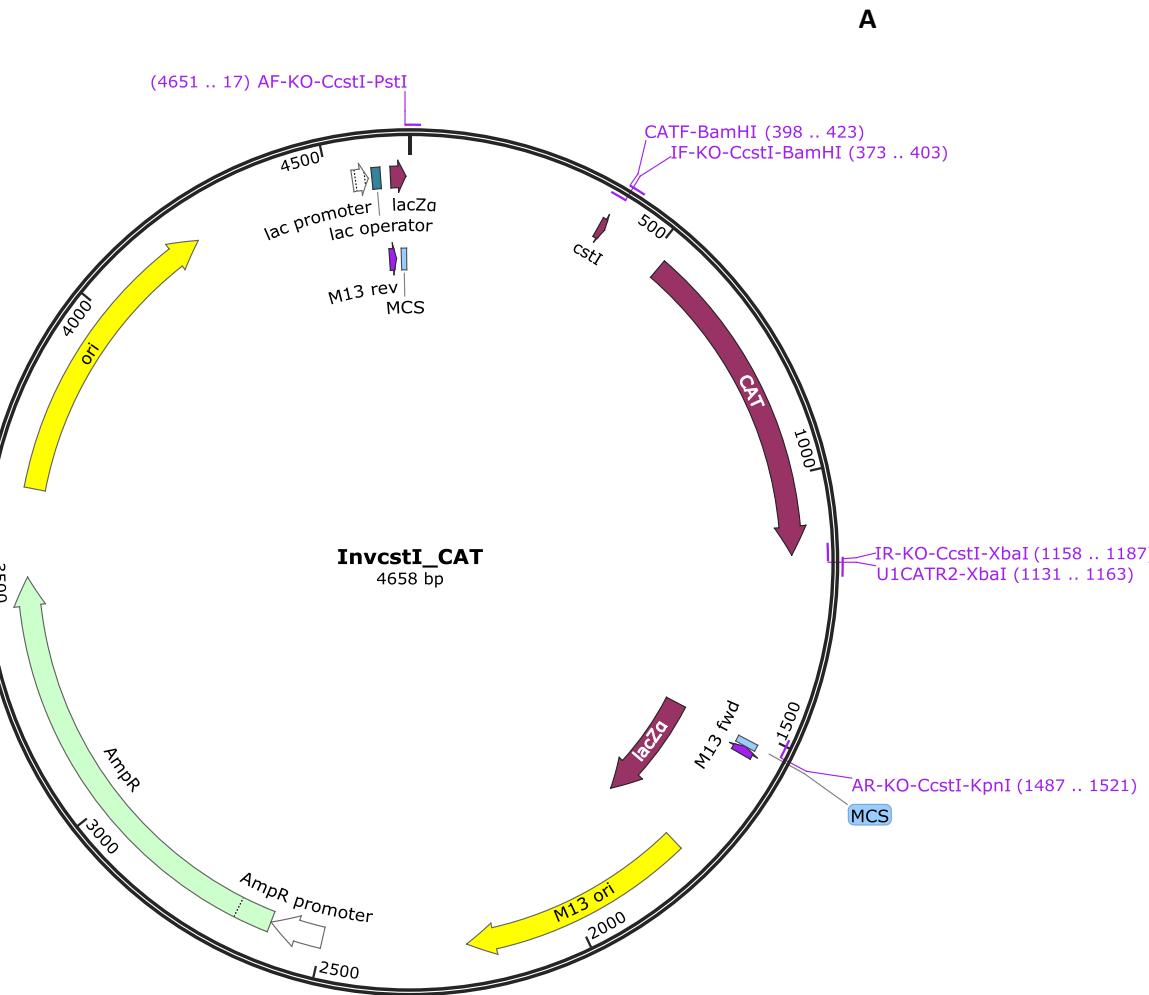
Primer name	Sequence
cstV amplification	
CcoLOS-KO-AF-PstI	ATCTGCAGCCTAATGCAACCGCACCCAAGC
CcoLOS-KO-AR-KpnI	AGGTACCATTATCCCAAAAAGTTGTCTTAAGCGTGG
Inverse PCR	
IR-cstVKOI-Cc-SalII	ACCGTCGACCATATGAAATTATTCTAGAGCTC
CcoLOS-KO-IF-BamHI	AGGATCCGTTTCTATCATATCTGCTCATAG
Erythromycin cassette	
ERIF-SalII	ACCGTCGACAGTATAAACCTTAAGAACTTTC
ERIR-BamHI	ACCGGATCCACTTACTTATTAAATAATTATAGCTATTG
ERIR-SalII	ACCGTCGACACTTACTTATTAAATAATTATAGCTATTG
ERIF-BamHI	ACCGGATCCAGTATAAACCTTAAGAACTTTC
Mutant verification	
EryCvF1	CCTCTTATTATGCCATTGTTGC
EryCvF2	ATATTTCATCCTAACCTAAAGTGAATAGC
EryCvR1	TTATTTCTGTAGTTTGCTAATTATGG
EryCvR2	GGAGAAAGAGTTGTGCTAATCTC
Gene expression verification	
09870cFv	TAA GTT TTT GGT AGT TTT TGC CTC G
09870cRv	ATTAGCACTAGATGATACAACCAGTG
09880cFv	CAACTAAGATCCCATTATGCCAAG
09880cRv	TAGAAGGTGGAGAGCTTCAGG
09890cFv	ACATAGCCTGTAATCTTACTAAATACG
09890cRv	ATAGGTTCAGGAGAATGTAATAACTATCC
09910cFv	GTC TAG TAA GTG CAT AAA TAC TTG C
09910cRv	CGATCAAAGAAAAGTATTATCCCAA
09930cFv	TGCGTGGTAGAGCTAGGATG
09930cRv	ACAATGAAAGCACTGATGACACTC

Supplemental Figure 7. Plasmids and primers for the generation of *C. coli* 76339 Δ cstV mutant strains. **A;** pCSTV:Ery-SF plasmid, **B;** pCSTV:Ery-RF plasmid, **C;** primer list

B

Primer name	Sequence
cstIV amplification	
cstIV-KOI-con-AF-PstI	ATCTGCAGGCCAAAACCACTCACTAAAAG
cstIV-KOI-con-AR-KpnI	AGGTACCGCAATCGATACTGATAATTAAACGCT
Inverse PCR	
IF-KO-Cc-BamHI	AGGATCCGCTGGCATGGATTGGAGATG
IR-KO-Cc-SalI	ATAGTCGACTCTTATAGCCACATGCAACAGC
Erythromycin cassette	
ERIF-SalI	ACCGTCGACAGTATAAACCTTAAGAACCTTC
ERIR-BamHI	ACCGGATCCACTTACTTATTAAATAATTATAGCTAT TG
Mutant verification	
CcCstIVvF	AACAGAAATGCTTACTGGCAC
EryCvF2	ATATTTCATCCTAACCTAAAGTGAATAGC
EryCvR1	TTATTTCTGTAGTTTGATAATTATGG

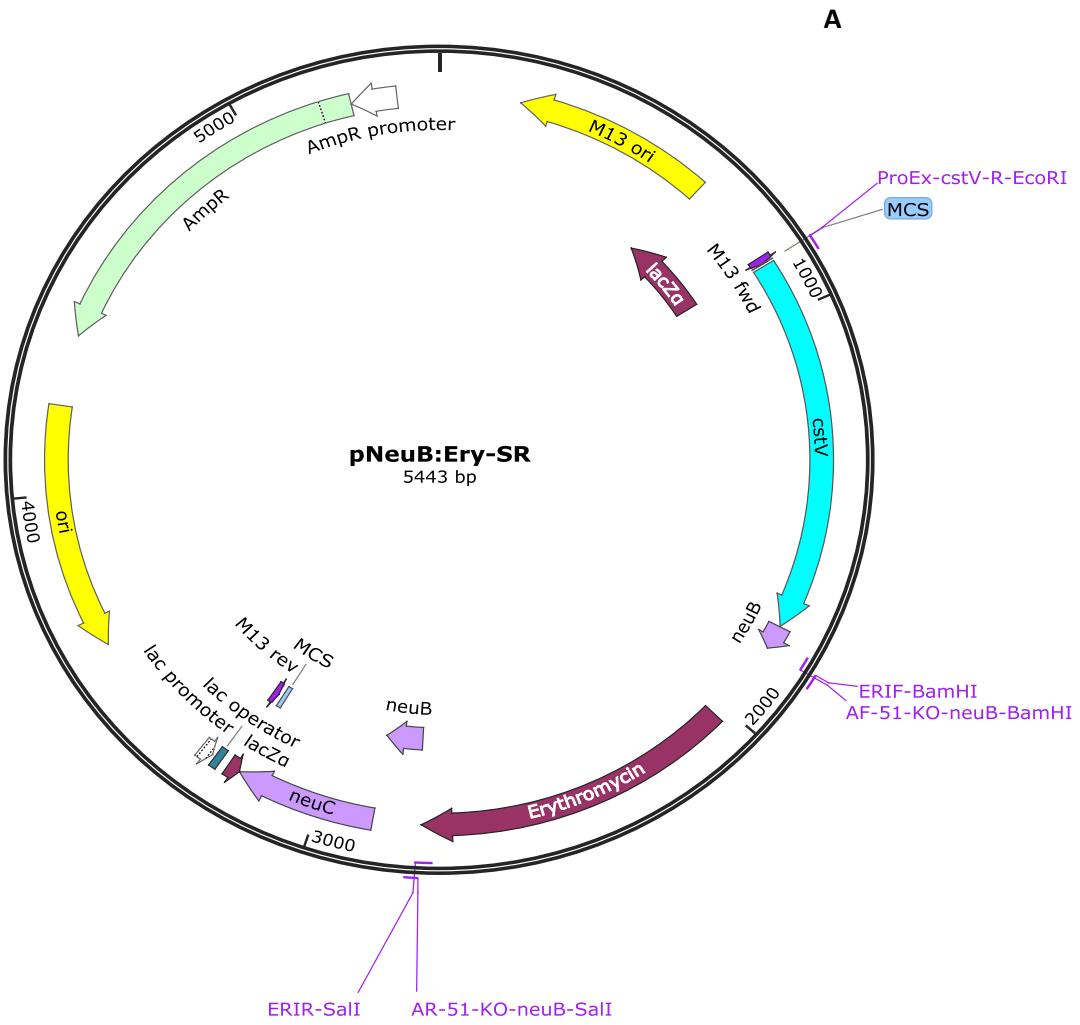
Supplemental Figure 8. Plasmids and primers for the generation of *C. coli* 65 and 73 Δ cstIV mutant strains. **A;** pCSTIV:Ery-SF plasmid, **B;** primer list



B

Primer name	Sequence
cstI amplification	
AF-KO-CcstI-PstI	ATCTGCAGAGGAAATGGGCCTAGTCTT
AR-KO-CcstI-KpnI	AGGTACCCACTATGGATTTCATAAGAATTCTACCT
Inverse PCR	
IR-KO-CcstI-XbaI	ATTCTAGATGAATTGGCTAACAGACTGTAGGTG
IF-KO-CcstI-BamHI	ACCGGATCCGCTATGGCGCACATATAAATACCAAG
CAT cassette	
U1CATR2-XbaI	ATTCTAGAGGGATTATTATTATTAGCAAGTCTTG
CATF-BamHI	AGGATCCC GGCGGTGTTCCCTTCCAAG
Mutant verification	
cstI-CvR	GCATTCTTCTATACAATTAACTCCC
cstI-CvF	ACTAAAAAACGGGAGGGAAAGC
CAT-CvR	CTCAGTCAAATACTCGAAAAGG
CAT-CvF	TCTATGATACCGTGGACAAGC

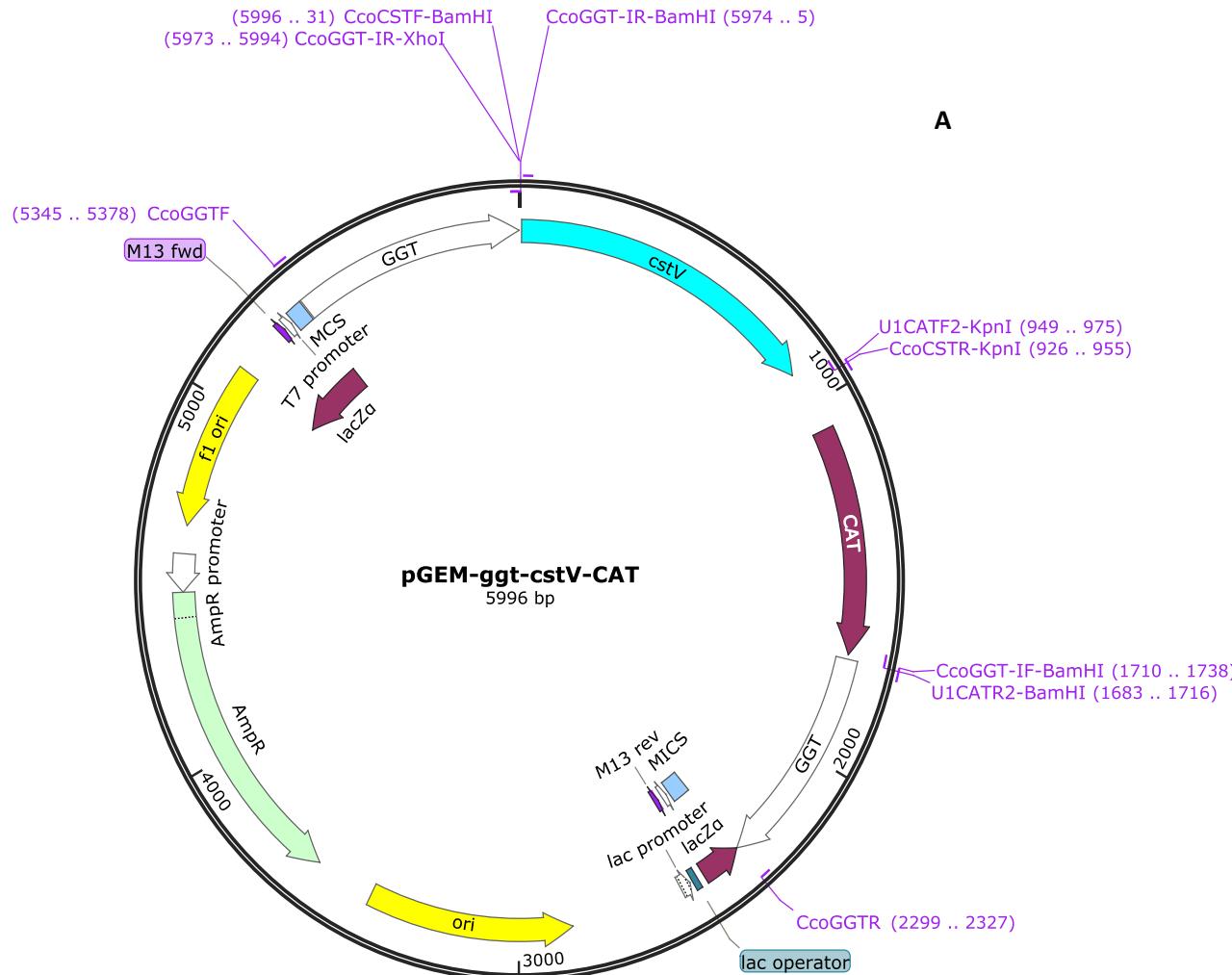
Supplemental Figure 9. Plasmids and primers for the generation of *C. coli* 76339 Δ cstI mutant strains. A; InvcstI_CAT plasmid, B; primer list



B

Primer name	Sequence
neuB amplification	
AF-51-KO-neuB-PstI	ATCTGCAGCTCTCCACCTTCTATATGTGCTAC
ProEx-cstV-R-EcoRI	ACTGAATTCA TAGAAAACAATGCAGTTGGTGT
Inverse PCR	
AR-51-KO-neuB-SalI	ACCGTCGACGGGACTCGGTGGAATCAGC
AF-51-KO-neuB-BamHI	AGGATCCCTAAAGGTGGTTTCTTGGGATATG
Erythromycin cassette	
ERIR-SalI	ACCGTCGACACTTACTTATTAAATAATTATAGCTAT TG
ERIF-BamHI	ACCGGATCCAGTATAAACCTTAAGAACTTTC
Mutant verification	
EryCvF2	ATATTTCATCCTAAACCTAAAGTGAATAGC
EryCvR1	TTATTTCTGTAGTTGCATAATTATGG

Supplemental Figure 10. Plasmids and primers for the generation of *C. coli* 76339 Δ neuB1 mutant strains. A; pNeuB:Ery-SR plasmid, B; primer list

**A****B**

Primer name	Sequence
ggt amplification	
CcoGGTF	ATTTAGTTATATTGTGATTCAATCACGCTAGG
CcoGGTR	AAAACCTGTTGTGATGATTCTAGAGCCACC
Inverse PCR	
CcoGGT-IF-BamHI	AGGATCCTCTATGTCGCCACCTAGTAGCG
CcoGGT-IR-BamHI	AGGATCCCAGGGCCTTGGCGATGAG
cstV amplification	
CcoCSTR-KpnI	ACCGGTACCTCTGGGATATGGTTAATTATC
CcoCSTF-BamHI	AGGATCCAATGATAAGAAACAATGCAGTTGTG
CAT cassette	
U1CATR2-KpnI	AGGTACCGGGATTATTATTAGCAAGTCTG
U1CATR2-BamHI	AGGATCCGGGATTATTATTAGCAAGTCTG

Supplemental Figure 11. Plasmids and primers for the generation of *cstV* complemented strain. A; pGEM-ggt-cstV-CAT plasmid, B; primer list