Sing cell RNA-seq reveals cellular diversity and developmental characteristics of human infant retina

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Abstract:

Retina, located in the innermost layer of the eye of human, holds the decisive role in visual perception. Dissecting the heterogeneity of retina is essential for understanding the mechanism of vision formation and even the development of central nervous system (CNS). Here, we performed single cell RNA-seq, analyzed 57,832 cells from human infant donors, resulting in 20 distinct clusters representing major cell types in retina: rod photoreceptors, cone photoreceptors, bipolar cells, horizontal cells, amacrine cells, Muller glia cells and microglia. We next constructed extensive networks of intercellular communication and identified ligand-receptor interactions playing crucial roles in regulating neural cell development and immune homeostasis in retina. Though re-clustering, we identified known subtypes in cone PRs and additional unreported subpopulations and corresponding markers in rod PRs as well as bipolar cells. Additionally, we linked inherited retinal disease to certain cell subtypes or subpopulations through enrichment analysis. Intriguingly, we found that status and functions of photoreceptors changed drastically between early and late retina. Overall, our study offers the first retinal cell atlas in human infants, dissecting the heterogeneity of retina and identifying the key molecules in the developmental process, which provides an important resource that will pave the way for retina development mechanism research and regenerative medicine concerning retinal biology.

Key words: single cell RNA sequencing; human retina heterogeneity; Muller glia;

Introduction:

Retina is the system responsible for light perception and visual signal transmission. Retina is a part of the central nervous system (CNS) and possesses diversified cell types. Thus, decoding the heterogeneity of retina is crucial for understanding the mechanism of vision formation as well as the highly complicated CNS. In general, seven major cell types (rod, cone, Müller glia cell, amacrine cell, horizontal cell, bipolar and retinal ganglion cells) constitute the three layers in retina: photoreceptors, bipolar cell layer and ganglion cell layer from inside out[1]. Photoreceptors, including rods and cones, are responsible for photo capture and convert them into electrical signal which will pass on to interneurons in bipolar cell layer for further modulation. Ganglion cells relay signals from interneurons to certain brain regions through optic nerves. Finally, the brain deciphers the electrical signals into three-dimensional vision. As the only non-neuronal cell, Muller glia cell is involved in extracellular environment regulation via nutrient intake, debris clearance, structural and functional stability maintenance[2,3].

In recent years, the boost of scRNA-seq techniques allow us to zoom in to retina and have discovered tremendous subtypes in traditionally defined classification. In human, 20009 single cells from three post-mortem donors were divided into 18 distinct clusters, including well-known retinal cells and novel rod cells subpopulations[4]. In mouse, 40 subtypes of retinal ganglion cells and their markers were identified using scRNA-seq[5]. Developmental researches also shed lights on retina and CNS morphogenesis. A recent study in mouse analyzed cells from ten developmental stages and found that RPCs displayed a transcriptomic temporal transition between early stage and late stage. Also, NFI family of TF were designated as regulator of bipolar cells and Muller cells cell fate determination due to the enriched expression in late RPCs[6]. Another research characterized 24979 cells from human fetal retina from gestational weeks 12 to 27, revealing the differentiation programs of retinal cells and specifying the potential of early born Muller cells to be RPCs[7].

Nevertheless, the retina development of postnatal infants, which impacts visual system vitally, remains elusive. In this research, we analyzed 57,832 single-nuclei transcriptomic profiles from infants in different development stages. Unsupervised clustering has defined 20 clusters, all of which were assigned with 7 major retinal cell types correspondingly. Communication network of 20 clusters connects the receptor-ligand pair with certain biological process in retina. We next performed reclustering thus providing a more delicate map of PRs and BCs. Reported subtypes of cone PR

were confirmed and new subtypes of BCs were identified.

Results:

Sample preparation and single cell transcriptome

We collected retinal biopsies from 2 donors (10-month-old and 2-year-old, hereafter termed as TenM and TwoY respectively) and dissociated them into single-cell or single nuclei suspension without surface marker pre-selection (Figure S1a). We constructed 3 snRNA-seq libraries for TenM retina and 2 scRNA-seq libraries for TwoY retina and then obtained the transcriptome of 66,362 nuclei/cells. After filtering, a total of 57,832 nuclei/cells were retained from subsequent analysis. The average number of genes detected per sample was 18,154, with a mean coverage of 57,545 reads per cell (Figure S1b). Pairwise Pearson correlation analysis demonstrated that libraries from the same donor share stronger correlation, confirming the validity of our datasets (Figure S1c). Of note, TwoYZJ and TwoY8 are standard scRNA-seq libraries thus explaining the relatively low correlations of these two libraries against others.

Cellular heterogeneity in retina

We embarked on creating a map for human infant retina. In a global view, 57,832 cells were clustered into 20 clusters (Figure1b). Batch effects from five different libraries were minimal (Figure S2a). According to canonical markers, differentially expressed genes (DEGs) and functional enrichment analysis (Figure2c, f, g), we were able to identify several major retinal cell types and 1 microglia cells: Rod photoreceptors (C0, C1, C2, C3, C4, C5, C6), cone photoreceptors (C11, C17), bipolar cells (C8, C10, C15, C19), horizontal cells (C13), amacrine cells (C16), Muller glia (C7, C12, C18), microglia (C9, C14). Rod photoreceptors highly expressed *NR2E3* and *PED6A* and cone photoreceptors were characterized by the expression of *ARR3* and *CNGB3*. Bipolar cells exhibited specific expression of *TRPM1* while *GRIA3* for horizontal cells. Apart from the common glia cells marker (*SPP1*), Muller glia cells and microglia cells can be annotated clearly by the expression of *SLC1A3* and *HLA-DPA1* respectively. Additionally, functional related GO terms were enriched in corresponding clusters, further confirming the annotation result of each cluster. For example, photoreceptors clusters were strongly associated with sensory perception of light stimulus and visual

perception while BC were enriched in GO terms related to modulation of chemical synaptic transmission. Microglia, known to be the primary resident immune cell type, displayed enrichment in neutrophil activation involved in immune response and T cell activation (Figure 2g). For ACs, though modest expression of corresponding markers were detected, hierarchical clustering and functional enrichment analysis exhibited AC characteristics in C16.

To systematically dissect the relationships between different cell types, we built correlation-based networks at the cell-type level (Figure 1d). We binned cellular cluster into sub-groups with 100 cells to reduce noise and enhance the robustness of the network. Consequently, the correlation map mimicked a cellular landscape for different retinal cell types. In general, the majority of cells showed much denser sub-network with cells from the same cell type than with others. Notably, almost all cells showed connections with rod PR, revealing high correlation with rod PR at averaged transcriptomic level. Another interesting observation was that, glia cells, including microglia cells and part of Muller cells, dispersed amidst rod PRs instead of forming distinct sub-network.

In order to assess the cellular composition in retina, we calculated the percentage of each cluster in the whole dataset (Figure 1c). The constitution of infant retina showed high similarity when compared with human adult retina dataset[4]. Rod photoreceptors took up the majority (75.32%) of the whole dataset while cone photoreceptors occupied a very small portion (3.1%). The second-largest cell type is Muller glia (9.24%). However, amacrine cells which constituted ~10% of the total cells in mouse, only account for 0.92% of our data. We also investigated the proportion of each cell type in early and late retina sample, which are shown in pie chart (absolute number) and radial barplot (proportion) (Figure 1c).

We next moved on to explore the intercellular communication network within retina. The cellular interactions were inferred using published ligand-receptor pairs (Methods). Expression patterns of ligand-receptor pairs in the networks revealed dense intercellular communication networks, especially in glia and BCs (Figure 2a). Nevertheless, rod PR subpopulations exhibited a relatively sparse network, showing less than one interaction among subpopulations within themselves and few with other cell types. This indicated that rod PRs interacted much more often with other retinal cell types than with PRs subpopulations. The most frequent interaction that appeared in our dataset is RTN4—LINGO1 which is suggested to regulate the radial migration of cortical neurons in

mouse brain[8], implying that RTN4—LINGO1 may be involved with migration of neural cells in retina. NRG1-ERBB4 and NRG3—ERBB4 pairs were also extensively expressed in retina neural cells—Muller glia interactions. *NRG3* has been reported to play a key role in regulating the growth and differentiation of glia cells and its signal transduction with ERBB4 was essential for neuron circuit formation[9] (Figure 2b). *SEMA3A*, specifically expressed in rod PR (C0, C16) can signal through PLXNA4 and engage in multiple biological activities like axon pruning and dendrites branching in adult procine[10] (Figure 2b). Particularly, Microglia_1 (C14) expressed HLA-A and its cognate receptor APLP2 were detected in the majority clusters (C0, C1, C4, C5, C6, C8, C9, C10, C11, C12, C15, C19), covering PRs, BCs and Muller cells (Figure 2b).

Developmental trajectory of Muller glia

Muller glia, as the most common type of glia cell in retina, serve to maintain the structural and functional stability of retinal cells. Previous study has reported that Muller glia showed similar properties as retinal progenitor cells[7] and were capable to differentiate into photoreceptors in human and mouse cell culture[3,11,12].

To further investigate the developmental relationship of MG clusters identified in Figure 1b, we subset MG and performed pseudo-time trajectory reconstruction using Monocle. Consequently, we obtained a putative developmental path with two branches (Figure2c). We then labeled MG subpopulations in the trajectory and found that MG_1 were primarily centralized on the start, MG_2 converged on both ends while MG_3 distributed loosely along the developmental path (Figure 2d). In order to explore the different properties of cells located in the starting point and two branches of the trajectory, we adopted branch expression analysis modeling (BEAM, Methods) to calculate DEGs. GO term enrichment analysis revealed that genes upregulated in cells in the start of the path is associated with neurogenesis and neuron development (Figure 2f). *PAX6*, a key regulator in the development of retinal neural tissues, showed relatively high expression in the start of the path and declined along with the path proceeding (Figure 2g). Cells in End1 were enriched with GO terms related to light perception whereas cells in End2 displayed features of active translation and respiration processes (Figure 2f). On top of that, cells in End1 showed *ATP8A2* gene expression, which was reported to be required for normal visual function and involved in photoreceptor survival[13]. *RPS27*, which encodes a ribosomal subunit protein and *EIF3L*, which specifically targets and initiates translation of a subset of mRNAs involved in

cell proliferation and cell cycling[14], were highly expressed in cell in End2 (Figure 2g). These results implied the distinct states of cells in two Ends.

Heterogeneity within major cell types: PRs and BCs

Due to the high heterogeneity of retinal cells, we next provided a more specific map for major cell types in retina by reclustering of corresponding clusters.

Based on previous study, there are three types of cone photoreceptors in human retina that can be differentiated by the expression of three opsins (OPN1SW, OPN1MW, OPN1LW), responsible for the perception of colors with different wavelengths[15,16]. To assess the heterogeneity of cone, we performed reclustering in cone PR clusters and divided them into 3 subclusters (Figure 3a). Due to the highly homologous sequence between *OPN1MW* and *OPN1LW*, we were only capable of distinguishing the S-cones (C2) from M/L-cones (C0, C1) using opsins encoding genes (Figure 3c). In this cone PR dataset (1569 cone PRs), M/L-cones constitute 94.77% of the total cells and the remaining 5.23% are S-cones (Figure 3b), which was a bit higher than the proportion (3.19%) from a scRNA-seq study into human adult retina[4]. Next, we conducted gene differential expression analysis to reveals new genetic characteristics for S-cones and M/L-cones. Highly and specifically expressed genes for S-cones (e.g. *KIAA1217, VWC2, SMAD5*) and M/L-cones (e.g. *RPS16, DDX5, SMTN*) were identified (Figure 3d, Table S2), providing more molecular evidences for future cone PRs studies.

We next interrogated another kind of PR in retina — rod PRs. On average, there are approximately 92 million rod cells in the human retina[17]. Rod cells are more sensitive than cone cells and are almost entirely responsible for night vision. In our dataset, 44448 rod cells were divided into 7 subpopulations (Figure 1b, Figure S7a). To investigate the sample origin of each cluster, we calculated the proportion of cell derived from the early and late retina in each cluster (Figure S7b). We found that Rod_1 and Rod_2 subpopulations were dominant by cells from early retina while the rest subpopulations were primarily comprised of cells from late retina. To assess the similarity of clusters at transcriptomic levels, we performed hierarchal clustering based on transcriptomic data. DEGs expression patterns and corresponding GO term enrichment results suggested the different features of rod subpopulations (Figure S7c,d).

Bipolar cells, served as interneurons, receive electrical impulses from photoreceptors, process them in various manners and then pass them to RGC. The diverse subtypes of BCs ensure their multi-functioning roles. Previous researches confirmed 15 subtypes

of BC experimentally using ~25,000 BCs[18]. BCs can be divided into two categories according to the type of photoreceptor which they connect to. Another taxonomy classifies BCs into ON and OFF types based on whether they will be excited or suppressed by increasing illumination. In our BC dataset (4282 BCs), unsupervised reclustering produced 9 distinct subclusters (Figure 3g). Of note, C7 were thought to be photoreceptor contamination and excluded from downstream analysis due to the modest expression of BC markers and light perception concerning GO terms enriched using DEGs of C7 (Figure S6b, c). C7 also displayed higher correlation with rod subtypes instead of BC subtypes (Figure S6d). When we took a closer look into C7 we found that cells in C7 were scattered in the whole BC clusters in Figure 2a (i.e. C8, C10, C15, C19 in Figure 2a). One possible explanation is that reclustering of BC dataset puts photoreceptors into a BC environment, extending the differences when it compared to the differences in the whole dataset situation. C0 and C5 were annotated as rod bipolar cells (RBC), showing high expression of *PRKCA*. According to the clear expression pattern of *GRM6* and *GRIK1*, C0, C2, C4, C5, C6 were considered as ON BCs while the remaining C1, C3, C8 were OFF BCs (Figure 3k). Heatmap of DEGs displayed distinct expression pattern of the defined clusters. We further explore the function of DEGs by functional enrichment analysis (Figure 3i, j). GO terms that closely related to the signal transduction like modulation of chemical synaptic transmission, neuron recognition and morphogenesis like axon guidance and synapse organization were enriched in almost all clusters to a varying extent. Next, we sought to fully describe the type-specific functional roles of BC subtypes by assessing the expression of Sodium, Calcium and Potassium ion channel subunits because ion channels control the transmit of neural signals. Sodium channel subunit-encoding genes were expressed selectively among subtypes (Figure 3l-n). Expect for *SCN1A*, *SCN2A, SCN3A* and *SCN8A* that were previously reported to be expressed in CBCs in goldfish[19], we noticed that *SCN7A* and *SCN9A* also showed specific expression among CBCs. Calcium channel alpha and delta subunits encoding genes also showed varying expression levels among subtypes. Furthermore, we observed a great deal of differentially expressed TFs, receptors and ligands, supporting our deductive functional subtypes (Figure S6e).

Cell type enrichment analysis of retinal diseases associated genes

We retrieved retinal diseases causing genes from RetNet database (https://sph.uth.edu/retnet/disease.htm) (Table S4) and performed hypergeometric test

to see whether these disease-related genes are enriched in the DEGs of certain clusters (Methods), by which means, discovering putative disease-causing cell type and therefore providing new sights for retinal diseases. Most of the disease types we investigated in this study are caused by malfunctions of PRs. Retinitis pigmentosa (RP) is characterized by retinal cells degeneration, begun with rod cells and followed by cone cells, consistent with which, enrichments of RP related genes occur in all rod subpopulations and M/L-cones (Figure 3f, Figure S7f). This indicated that all rod subpopulations but only M/L-cones are involved with RP genesis and corresponding markers we identified may help to pinpoint the scope of target cells for therapies. Cone-rod dystrophy (CRD), unlike RP, is featured by an initial loss of color vision and of visual acuity due to loss of cone function and followed by rod cells deterioration. Most rod subpopulations (Rod_1, Rod_2, Rod_5, Rod_6, Rod_7) and M/L-cones displayed close connections with CRD occurrence with enrichment significance (-log P value) over 1.5 (Figure 3e, Figure S7e). Congenital stationary night blindness (CSNB) can be classified into complete CSNB (cCSNB) and incomplete CSNB (iCSNB) which is caused by defects localize to ON BCs and PR synapse respectively. Accordant with what mentioned above, we found that ON BCs DEGs are highly enriched in CSNB causing genes, but to a substantially varying extent (Figure 3h), suggesting that different BC subtypes contribute differently to CSNB or lead to CNSB with various severities. Furthermore, DEGs of Rod_1, Rod_2, Rod_5 and Rod_7 subpopulations showed modest association with CSNB causing genes (Figure S7f), implying a moderate role in CNSB. Besides, macular degeneration (MD) were specifically associated with Rod_1, Rod_2 and Rod_7 (Figure S7h). Together with what described above, Rod_1, Rod_2 and Rod_7 showed strong enrichment of common retinal diseases. From the perspective of cone PRs, S-cones don't seem to be significantly related to any retinal disease we collected while M/L-cones are the major cone PR subtype causing or being affected by retinal diseases.

In conclusion, these results not only matched previous findings but also linked several diseases with certain retinal cell subtypes, facilitating researches and development of novel drug into retinal malfunctions.

Similarities and differences in cell proportion, cell status between retina in different developmental stages

Since cell proliferation is a key metric of cell maturation level, we next performed cell

cycle analysis. We first assigned a cell cycle phase score to each cell using gene expression signatures for the S and G2/M phases. From a global view, most of the cells were in G1 phase while relatively few were in proliferating G2/M stage (Figure S9a). Interestingly, cells in G2M stage were primarily from early retina (Figure S9c). Except for rod PR where cells from late retina diffuse more evenly, most cells from late retina in other cell types were assigned negative S scores and G2M scores, thus being considered to be in G1 phase.

We next asked what programs the postnatal retina will go through. Previous studies have shown that photoreceptors will develop from a relatively immature state into a mature state, improving the light sensitivity after birth[20]. Hence, to answer the question above, we performed differential expression analysis in the same cell type between early and late PRs.

In cone PRs, 581 upregulated and 920 down-regulated genes in total were found in early cone RPs in M/L-cones (C0, C1 in Figure 3a, Table S2) when compared with late cone RPs (Figure 4a, c). Functional enrichment demonstrated that genes upregulated in early cone PRs showed weak enrichment in terms related to light perception while highly express genes concerning photoreceptor differentiation (Figure 4b, d). In S-cones (C2 in Figure 3a), 44 down-regulated whereas no upregulated genes found to be significant (Figure S8h, Table S2). GO terms associated with cellular respiration were enrichment in genes upregulated in late PRs (Figure S8h). We next inspected those TFs, receptors and ligands that differentially express between two stages in M/L-cones (Figure 4e). We found that *ARID1B*, which were highly expressed in early cone PRs, belongs to a neuron-specific chromatin remodeling complex and involved in cell-cycle activation and neural progenitor cell fate commitment during neural development[21]. *MEF2C*, also displayed specific expression in early cone PRs, has been prove crucial for normal neuronal development, distribution, and electrical activity in the neocortex[22]. *NR2E3*, an orphan nuclear receptor of retinal photoreceptor cells, is an activator of rod development and repressor of cone development[23]. Hence, it's reasonable to infer that TFs like *ARID1B*, *MEF2C* and *NR2E3* cooperate together to drive the development process in cone maturation. Likewise, in rod PRs, according to the functional enrichment analysis, it's also showed that cells from early retina displayed strong differentiation ability while cells from late retina exhibit the competence to perceive external light and transduce signal to subsequent neurons (Figure S8).

To further identify key regulators in the course of cone PR development, we next

constructed regulatory network based on significantly high expressed TFs in early M/L-cones_1 (C1 in Figure 3a) (Figure 4f). NRL, reported as a critical intrinsic regulator of photoreceptor development and function[24], possessed most putative target genes, indicating its crucial role in the developmental course of cone PR. The targets of TCF4 were enriched in GO terms related ubiquinone and ketone biosynthetic processes (Figure 4g). Ubiquinone is the oxidized form of CoQ10, which serves as an antioxidant to help protect cells from oxidative damage. The level of CoQ10 declines with age due to decreased synthesis and increased degradation, leading to age-related neurodegenerative diseases, such as Parkinson's disease and age-related macular degeneration[25]. Besides, ZNF385B, whose targets were found to be associated with immune activities, were reported to be expressed in germinal center B cells and involved in B-cell apoptosis[26].

Discussion

This complex intrinsic composition of retina cells highlights the necessity of single-cell technologies for dissecting transcriptome signature in detail. Here, we report the first single cell atlas – to our knowledge, for human retina in different infant developmental stages. Our dataset consists of 57,832 cell and nuclei achieve unprecedentedly large scale for human retina research. In this study, we firstly established a high-resolution map for human infant retina with six out of seven major retinal cell types successfully assigned. We also identified microglia, the resident immune cells in human retina and found by ligand-receptor communication network analysis that microglia showed putative interactions primarily with rod PRs, BCs and Muller glia.

Due to the progenitor-like features of Muller glia, we performed pseudo-time trajectory analysis using MG_1, MG_2 and MG_3, which revealed a developmental path with two branches. Further characterization of the branches of the path suggested that cells in left branch displayed light perception functions and cells in the right branch showed active translation activities.

We next sought to identified subtypes and subpopulations of photoreceptors and bipolar cells through reclustering. For cone PRs, we identified S-cones but failed to distinguished M-cones from L-cones due to the high homology of opsin sequences. For BCs, reclustering resulted in 8 subpopulations, 3 of which were classified as OFF BCs and the remaining 3 were thought to be ON BCs according to the expression of *GRIK1*

and *GRM6*. We further described the different properties of each subpopulations by assessing the expression pattern of function related ion channels.

Another part of this study is to compare the similarities and differences of early and late retinas at transcriptomic level. We first evaluate the proliferation state of each cell by assigning each cell a calculated cell cycle stage. We found that except for microglia, most of the cells from late retina remain in a relatively quiescent in other cell types we identified. That is to say, cells from early retina were in G2M stage, an actively proliferating cell cycle stage. In addition, through differential gene expression analysis and functional enrichment analysis, we found M/L-cones from early retina showed higher expression of genes related to photoreceptor differentiation and neurogenesis whereas M/L-cone from late retina showed mature photoreceptor properties with enriched GO terms concerning light perception and phototransduction.

Inherited retinal diseases (IRDs) are a group of retinal disorders caused by an inherited gene mutation and can result in vision loss or blindness[27–29]. Currently, the disease diagnosis and therapies were hampered by our understanding of human retina at single cell level. In our retinal disease analysis, S-cones manifested insignificant enrichment of all common retinal diseases included in our analysis (Table S4) while M/L-cones were found to be associated with CRD and RP. For rod PRs, Rod_1, Rod_2 and Rod_7 showed strong association with many retinal genetic disease, especially with CRD and RP. CSNB were found to be associated with defects in ON BCs but the enrichment varies among the ON BC subpopulations. Taken together, our findings narrow down the disease-causing cell subtypes and subpopulations and their corresponding biomarkers, which may not only shed lights on understanding the molecular mechanisms underlying IRDs, but also provide new insights for discovering novel drug targets and developing gene therapies for IRDs.

Material and methods:

Ethics statement

Collection of human retina was approved by the Ethics Committee of the Eye & ENT Hospital of Fudan University and in accordance with the Code of Ethics of the World Medical Association (Declaration of Helsinki) for medical research involving human subjects. All protocols were in compliance with the 'Interim Measures for the Administration of Human Genetic Resources' administered by the Chinese Ministry of

Health.

Human retina collection

The retinal tissues come from a 10-month and a 2-year-old toddler donor. The donor's eyeballs were collected by Eye & ENT Hospital of Fudan University for corneal transplantation, and eyeball samples acquired within 12 hours post-mortem. The remaining eyeball tissue was used to extract retina tissue. Retina tissue could be exposed by removing iris, lens and vitreous successively. The intact retinal tissue was carefully dissociated and placed in pre-cooled HBSS without RPE/choroid layer. One of the retinal tissues was digested into single-cell suspension in Trypsin-EDTA solution (containing 0.25% Trypsin and 0.02%EDTA) for 37 ℃ 20 min, and the remaining tissues were immediately frozen in liquid nitrogen. Then, single-cell suspension was filtered through a 40μm cell strainer to eliminate any clumped cells. The freshly prepared cell suspension was immediately used for the construction of single-cell sequencing libraries.

Single cell cDNA library preparation and high-throughput sequencing

The freshly prepared cell suspensions to generate cDNA libraries with Single Cell 3' Library and Gel Bead Kit V2(120237 10x Genomics), according to the manufacturer's instructions (120237 10x Genomics). Flash-frozen tissues were homogenized in 2mL ice-cold Lysis buffer (10 mM Tris-HCl, pH 7.4, 10 mM NaCl, 3 mM MgCl2, 0.1% NP40, protease inhibitors) then dounced in an RNase-free 2ml glass dounce (D8938-1SET SIGMA) 15x with a loose pestle and 15x with tight pestle on ice. Transfer homogenization through 40 μm filter (352340 BD) and removed the block mass. The cell filtrate was proceeded to Density Gradient Centrifugation. Mixed 400 μL cell filtrate with 400 μL 50% Iodixanol Solution in 2mL lo-Bind tubes (Z666556-250EA SIGMA). Carefully layer 29% Iodixanol Solution and 35% Iodixanol Solution to the bottom of tube. Centrifuging at 3,000 xg, 4℃ for 30min.

Nuclei were resuspended in ice-cold 1 x PBS (10010-031 GIBCO) containing 0.04% BSA and spin down at 500xg for 5min. Discarded supernatant and then using regular-bore pipette tip gently pipette the cell pellet in 50 μL ice-cold 1 x PBS containing 0.04% BSA and 0.2U/μl RNase Inhibitor. Determined the nuclei concentration using a Hemocytometer (101010 QIUJING). Then, loaded on a Chromium Single Cell Controller (10x Genomics) to generate single-cell Gel Bead-In-EMulsions (GEMs) by

using Single Cell 3' Library and Gel Bead Kit V2 (120237 10x Genomics). Captured cells released RNA and barcoded in individual GEMs. Following manufacturer's instructions (120237 10x Genomics) libraries were generated from each donor sample. Indexed libraries were converted by MGIEasy Lib Trans Kit (1000004155, MGI) then sequenced on the MGISEQ 2000 (MGI) platform with pair-end 26bp+100bp+8bp (PE26+100+8).

Pre-processing and quality control of scRNA-seq data

We first used Cell Ranger 3.0.2 (10X Genomics) to process raw sequencing data and then Seurat (10.1038/nbt.3192) was applied for downstream analysis. Before we start downstream analysis, we focus on four filtering metrics to guarantee the reliability of our data. (1) We filter out genes that are detected in less than 0.1% of total cell number to guarantee the reliability of each gene; (2) We filter out cells whose percentage of expressed mitochondrial genes are greater than 10%; (3) We also filter out cells whose UMI counts are either less than 1.5 offset to the first quantile or greater than 1.5 offset to the third quantile of total UMI counts to filter out the doublet-like cells; (4) We filter out cells whose detected genes are less than 200.

Identification of cell types and subtypes by dimensional reduction

The heterogeneity of retina was determined using Seurat R package (10.1038/nbt.3192). We performed Seurat-Alignment to eliminate batch effect, allowing us to combine data from multiple samples. Then we determined significant PCs using the JackStrawPlot function. The top twelve PCs were used for cluster identification with resolution 1.0 using k-Nearest Neighbor (KNN) algorithm and visualization using t-Distributed Stochastic Neighbor Embedding (tSNE) algorithm. Cell type were assigned by the expression of known cell-type markers and functional enrichment analysis. The FindAllMarkers function in Seurat was used to identify marker genes for each cluster using default parameters.

Removal of cell cycle effects in clustering and cell cycle analysis

We collected 43 genes and 54 genes related to S phase and G2/M phase respectively[30,31]. For clustering, each cell was assigned a score to delineate its cell cycle state by CellCycleScoring function in Seurat according to the expression of these genes. Subsequently, the cell cycle effect was regressed out based on the scores, leading to a more accurate clustering result. For cell cycle analysis, cells were determined to be

quiescent (G1 stage) if their S score < 0 and G2/M score < 0; otherwise, they were deemed proliferative. In addition, proliferative cells were designated G2/M if their G2/M score > S score, whereas cells were designated S if their S score > G2/M score.

Functional enrichment analysis

Functional enrichment analysis includes two parts: GO term and KEGG pathway. Lists of genes were analyzed using clusterProfiler R package (10.1089/omi.2011.0118) and the BH method was used for multiple test correction. GO terms with a P value less than 0.01 and KEGG term with a P value less than 0.05 were considered as significantly enriched.

Construction of intercellular correlation network

To reduce noise, we averaged the expression of every 100 cells within cluster and then calculated the pairwise Pearson correlation between two dots based on their average expression profiles. Inter-dots relationship will be shown if their Pearson correlation is greater than 0.8. The visualization of the correlation network is achieved using Cytoscape (10.1101/gr.1239303).

Construction of pseu-dotime trajectory using variable genes

The Monocle2 R package (version 2.10.1) was used to construct single cell pseudo-time trajectories to discover developmental transitions[32–34]. We used highly variable genes identified by “estimateDispersions” function to sort cells in pseudo-time order. “DDRTree” was applied to reduce dimensional space, and the minimum spanning tree on cells was plotted by the visualization functions “plot_complex_cell_trajectory”. BEAM tests were performed on the first branch points of the cell lineage using all default parameters. “plot_genes_branched_pseudotime” function was performed to plot a couple of genes for each lineage.

Branch expression analysis modeling

Monocle provides the branch expression analysis modeling (BEAM) to calculate the gene expression change as cells pass from early developmental stage through the branch and DEGs between different branches. We used “BEAM” function with default parameter settings and genes with q value less than 0.01 were considered significant.

Regulatory network construction

We downloaded homo sapiens TF list from AnimalTFDB 3.0[35] as a TF reference and extracted TFs in marker genes list of each cluster to construct the regulatory network using GENIE3 R package[36]. The network was plotted using Cytoscape.

Construction of cellular communication network

The ligand-receptor interaction relationships were downloaded from the database, IUPHAR/BPS Guide to PHARMACOLOGY[37], and the Database of Ligand-Receptor Partners (DLRP)[38,39]. 0.5 was used as a threshold for the average expression level of normalized expression level. Ligands and receptors whose expression above this threshold were considered as expressed in the corresponding cluster. Cytoscape was used to visualize the interactions.

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Author contributions

XH-S, JH-W, SH-Z, F-C, Y-G and DS-C designed and initiated the project; FY-H, JJ-X, P-X, FJ-G, DD-W, DW-Z coordinated and collected the human donor retinas; XH-S, JH-W, SH-Z, and FY-H conducted the experiments of retina sampling; ZX-X, SY-W, LC-L, CC-C and J-X constructed sequencing libraries; JX-Z, SY-W and XN-D were responsible for data analysis; JC-Z, PW-D and W-L contributed to data visualization; JX-Z, DS-C, F-C, JH-W, SJ-J, JG-Z, JK-L, J-F wrote the manuscript and/or provided suggestions. All authors have revised and proofread the manuscript.

Competing interests

The authors declare no competing interests.

Data and material availability

The data that support the findings of this study have been deposited in the CNSA (https://db.cngb.org/cnsa/) of CNGBdb with accession number CNP0000436.

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Figure1. Cellular diversity in retina.

(A) Scheme of the experimental and bioinformatics design in our study.

(B) A t-SNE map of all libraries. Cells are colored by cell-type cluster.

(C) T-SNE maps of all tissues single-cell data with cell colored based on the expression of marker genes for particular cell types. Gene expression levels are indicated by shades of blue.

(D) A correlation network showing relationships among 20 cell-type clusters defined in Figure 2b. Each node represents the averaged expression of every 100 cells within each of the clusters. Each edge corresponds to a network correlation between two nodes. Each node is colored by corresponding cell type and node border is colored by cluster in Figure 2b.

(E) Barplot showing the percentage of each cell type in the whole dataset. Pie charts: the proportion of cells from a certain sample in each cell type to the total number of cells identified as this cell types. Radial barplot: the proportion of cells from a certain sample in this cell type to the total number of cells in this sample.

(F) A gene expression heatmap showing top differentially expressed gene for each cluster in all 20 clusters. Red corresponds to high expression level; white indicates intermediate level and blue correspond to low expression level.

(G) Selected GO terms enriched in each cellular cluster. Dot size denotes the gene ratio and color denotes the adjusted P value.

Figure 2.

(A) Intercellular communication within each cluster in Figure2b. The line color indicates ligands broadcast by the cell population of the same color (labeled). Lines connect to cell populations where cognate receptors are expressed. The line thickness is proportional to the number of ligands where cognate receptors are present in the recipient cell population. Lines with number of interactions lower than 1 are not shown. Map quantifies potential communication, but does not account for anatomic position or boundaries of cell populations.

(B) Sub-communication network of selected clusters.

(C-D) Cell lineage relationships of Muller cells (MG_1, MG_2 and MG_3 in Figure 1b) in the human infant retina revealed by Monocle branched single-cell trajectory analysis. Each dot represents a single cell; the color of each dot indicates the corresponding branch information assigned by Monocle(C) or the cell type information (D).

(E) Barplot showing the numbers of differentially expressed TF, receptor and ligand in each branch state.

(F) Gene ontology analysis of modules created by clustering the two main branches from the lineage tree. The analysis reflects the cell fate commitment. In this heatmap, the middle represents the start of pseudo-time. One lineage starts from the start and moves to the End1 while the other lineage moves to the End2. Rows are genes correlated into different modules.

(G) Expression of selected differentially expressed genes for each branch. The expression of cells

was ordered by Monocle analysis in pseudo-time. The full line represents the End1 while the dash line represents the End2.

Figure 3. Cellular diversity within cone PRs and BCs.

(A) A t-SNE map of cone photoreceptors single-cell data. Cells are colored by cluster (subtypes).

(B) Barplot showing the percentage of S-cones and M/L-cones in the whole cone PR dataset.

(C) Violin plot displaying the expression of opsin encoding genes in each cluster in Figure 3a.

(D) Bubble plot showing the expression of differentially expressed genes in each cluster in Figure 3a. Dot size represent the percentage of cells express corresponding gene and color represent the expression level.

(E,F,H) Cone subtype enrichment of CRD (Cone-Rod Dystrophy) (E), RP (Retinitis Pigmentosa) (F) and bipolar subtype enrichment of CSNB (Congenital Stationary Night Blindness) (H) risk genes respectively. Heatmap: Expression of corresponding disease risk genes in cell subtypes. cells are ordered by cluster. Bar graphs: numbers indicate log2 P value, the red line indicates significance.

(G) A t-SNE map of bipolar cells single-cell data. Cells are colored by cluster (subtypes).

(I) Enriched GO terms in each cluster in Figure 3f. Dot size denotes the gene ratio and color denotes the adjusted P value. Dot size represent the percentage of cells express corresponding gene and color represent the expression level.

(J) Enriched KEGG pathways in each cluster in Figure 3f. Dot size denotes the gene ratio and color denotes the adjusted P value.

(K) Expression pattern of BC subtypes classification genes.

(L-N) Bubble plot showing cluster-specific expression of Potassium (L), Sodium (M) and Calcium (N) channels encoding genes.

Figure 4. Developmental analysis of cone PR from early and late retinas.

(A) Volcano plot showing the differentially expressed genes in M/L-cone_0 (Figure 3a) between early and late retina. Genes are plotted as log2 fold change versus the −log10 of the adjusted p-value. Genes in red denotes genes upregulated in early retina while blue denotes upregulated in late retina. Insignificant genes are colored in grey. Thresholds are shown as dashed lines.

(B) GO term enrichment analysis of differentially expressed genes in M/L-cone_0 between early and late retina.

(C) Volcano plot showing the differentially expressed genes in M/L-cone_1 (Figure 3a) between early and late retina. Genes are plotted as log2 fold change versus the −log10 of the adjusted p-value. Genes in red denotes genes upregulated in early retina while blue denotes upregulated in late retina. Insignificant genes are colored in grey. Thresholds are shown as dashed lines. (D) GO term enrichment analysis of differentially expressed genes in M/L-cone_1 between early and late retina.

(E) Violin plot Differentially expressed TFs, receptors and ligands in M/L-cone_0 between early and late retina.

(F) Regulatory network constructed using genes upregulated in M/L-cone_0 in early retina. Each node represents a gene and each line represents a putative interaction. Nodes colored in red represent TF and nodes colored in blue represent target genes.

(G) GO term enriched in the target genes of TCF4, ZNF385A and ZNF385B.

Figure S1

(A) Human infant retinal sampling and sequencing information.

(B) Barplot showing the basic library information including the numbers of cells and genes that passed our QC protocol (Methods).

(C) Pearson correlation analysis of transcriptomic profiles from all sequencing libraries. Top panel shows the result of hierarchical clustering. TwoY* denote libraries constructed using samples from 2-year old infant while TenM* denote libraries constructed using samples from 10-month old infant.

(D) Violin plot showing basic information of each sequencing libraries.

(E) Venn plot showing the intersect of housekeeping genes and detected genes in our dataset.

Figure S2

(A) T-SNE maps of the whole dataset. Cells are colored according to their batches (upper left), sample source (upper right). Bottom panel shows 3D t-SNE plots of dimensions 1,2,3 from two different angles.

(B) Pairwise correlation coefficients among clusters in Figure 1b. Red indicates high correlation while blue indicates the opposite.

(C) Boxplots of differentially expressed TFs, receptors and ligands in each cluster in Figure 1b.

Figure S3

(A-L) T-SNE plots showing the expression pattern of canonical markers for each cell type identified in Figure 1b. Gene expression levels are indicated by shades of blue.

Figure S4

(A) T-SNE plots of individual clustering results of cells from two samples (left: 10 months; right: 2 years). Cells are colored by batches.

(B) T-SNE plots of individual clustering results of cells from two samples (left: 10 months; right: 2 years). Cells are colored by clusters.

(C) T-SNE plots of individual clustering results of cells from two samples (left: 10 months; right: 2 years). Cells are colored by cell types.

Figure S5

(A-T) Sub-communication network of all clusters in Figure 1b.

Figure S6

(A) T-SNE plot showing the reclustering result of BC dataset (BC_1, BC_2, BC_3 and BC_4 in Figure 1b). Cells are colored by cluster. C7 were found to be rod photoreceptors.

(B) Expression pattern of bipolar cell and rod photoreceptors markers. Dot size represent the

percentage of cells express corresponding gene and color represent the expression level.

(C) Barplot showing the GO terms enriched in C7 differentially expressed genes.

(D) Transcriptomic correlation analysis of C7 between bipolar cells and rod photoreceptors.

(E) Boxplots of differentially expressed TFs, receptors and ligands in each cluster in Figure 3g.

(F) Boxplots of differentially expressed TFs, receptors and ligands in each cluster in Figure 3a.

Figure S7

(A) T-SNE plot showing the rod dataset (C0-C6) extracted from Figure 1b. Cells are colored by cluster.

(B) Unsupervised hierarchical clustering of the 7 clusters based on the average gene expression of cells in each cluster. The proportions of different cell sources are shown.

(C) Boxplots showing the expression patterns of selected differentially expressed genes.

(D) Selected GO terms enriched in each cellular cluster. Dot size denotes the gene ratio and color denotes the adjusted P value.

(E-H) Rod subpopulation enrichment of CRD (Cone-Rod Dystrophy) (E), RP (Retinitis Pigmentosa) (F), CSNB (Congenital Stationary Night Blindness) (G) and MD (Macular Degeneration) (H) risk genes respectively. Heatmap: Expression of corresponding disease risk genes in rod subpopulations. cells are ordered by cluster. Bar graphs: numbers indicate log2 P value, the red line indicates significance.

Figure S8

(A-G) Volcano plot: The differentially expressed genes in Rod_1 (A), Rod_2 (B), Rod_3 (C), Rod_4 (E), Rod_5 (G), Rod_6 (F), Rod_7 (D) between early and late retina. Genes are plotted as log2 fold change versus the −log10 of the adjusted p-value. Genes in red denotes genes upregulated in early retina while blue denotes upregulated in late retina. Insignificant genes are colored in grey. Barplot: functional enrichment of genes that significantly upregulated in early (colored in red) or late (colored in blue) retina.

Figure S9

(A) Circular barplot of the percentage of cells in different cell cycles in each cluster in Figure 1b.

(B) Ridge plot showing the expression of selected cell cycle related genes in each cell type and sample source.

(C) Cell cycle plot for each cell type identified in Figure 1b. Each dot represents a single cell, and its color represents the sample source. The positions of dots illustrate the cell cycle phases of single cells. The inserted numbers represent the numbers of cells in G2M stage in each cell type from different sample sources.

Figure S10

(A-F) Violin plots showing different expression patterns of rhythm related genes in different cell types. Each dash box indicates an expression pattern.

Figure 1

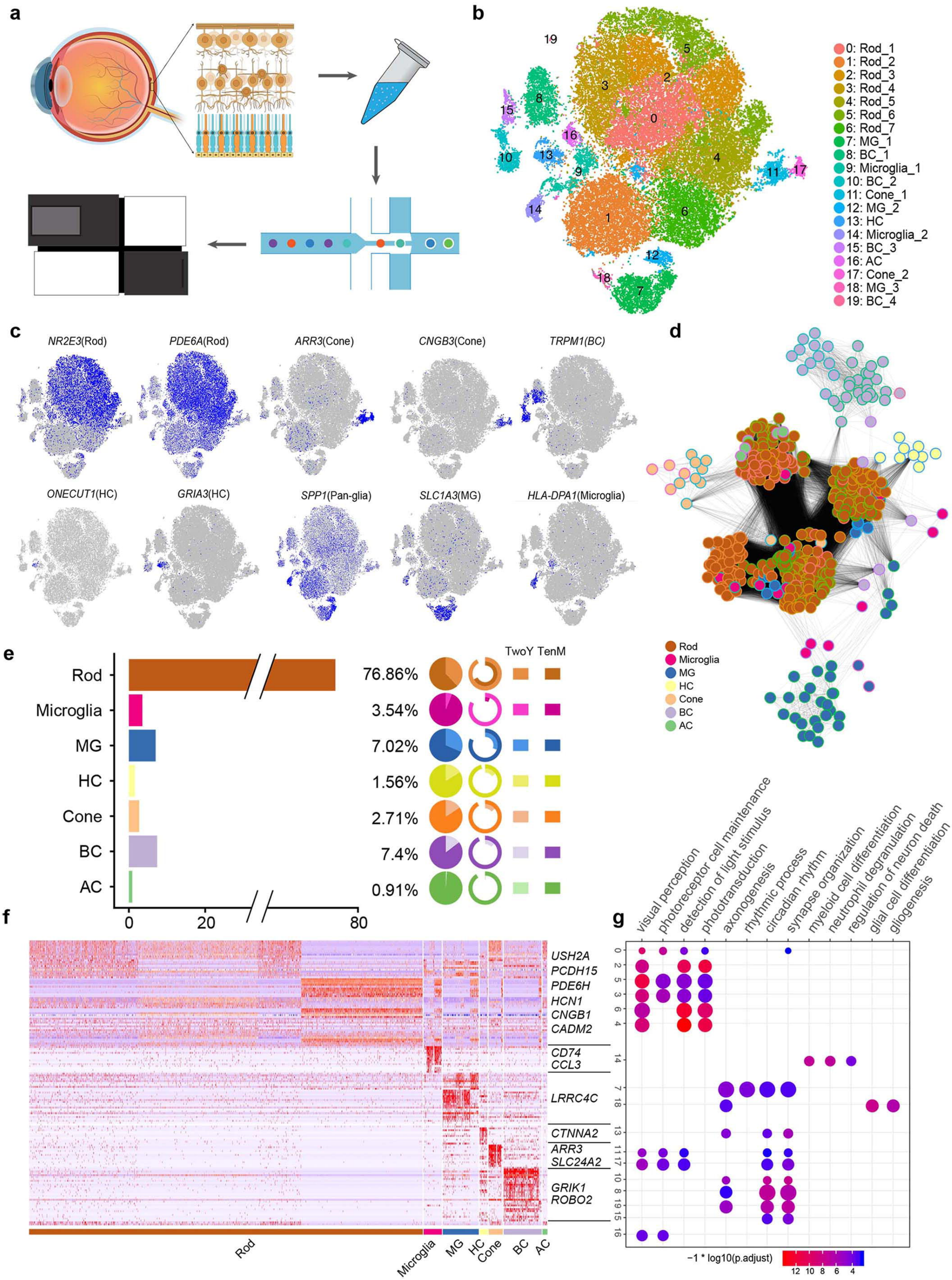

a
b
0: Rod_1
1: Rod_2
2: Rod_3
3: Rod_4
4: Rod_5
5: Rod_6
6: Rod_7
7: MG_1
8: BC_1
9: Microglia_1
10: BC_2
11: Cone_1
12: MG_2
13: HC
14: Microglia_2
15: BC_3
16: AC
17: Cone_2
18: MG_3
19: BC_4
c
NR2E3(Rod)
PDE6A(Rod)
ARR3(Cone)
CNGB3(Cone)
TRPM1(BC)
ONECUT1(HC)
GRIA3(HC)
SPP1(Pan-glia)
SLC1A3(MG)
HLA-DPA1(Microglia)
d
Rod
Microglia
MG
HC
Cone
BC
AC
e
TwoY TenM
Rod 76.86%
Microglia 3.54%
MG 7.02%
HC 1.56%
Cone 2.71%
BC 7.4%
AC 0.91%
0
20
80
f
USH2A
PCDH15
PDE6H
HCN1
CNGB1
CADM2
CD74
CCL3
LRRC4C
CTNNA2
ARR3
SLC24A2
GRIK1
ROBO2
Rod
Microglia
MG
HC
Cone
BC
AC
g
visual perception
photoreceptor cell maintenance
detection of light stimulus
phototransduction
axonogenesis
rhythmic process
circadian rhythm
synapse organization
myeloid cell differentiation
neutrophil degranulation
regulation of neuron death
glial cell differentiation
gliogenesis
−1 * log10(p.adjust)
12 10 8 6 4

Figure 2

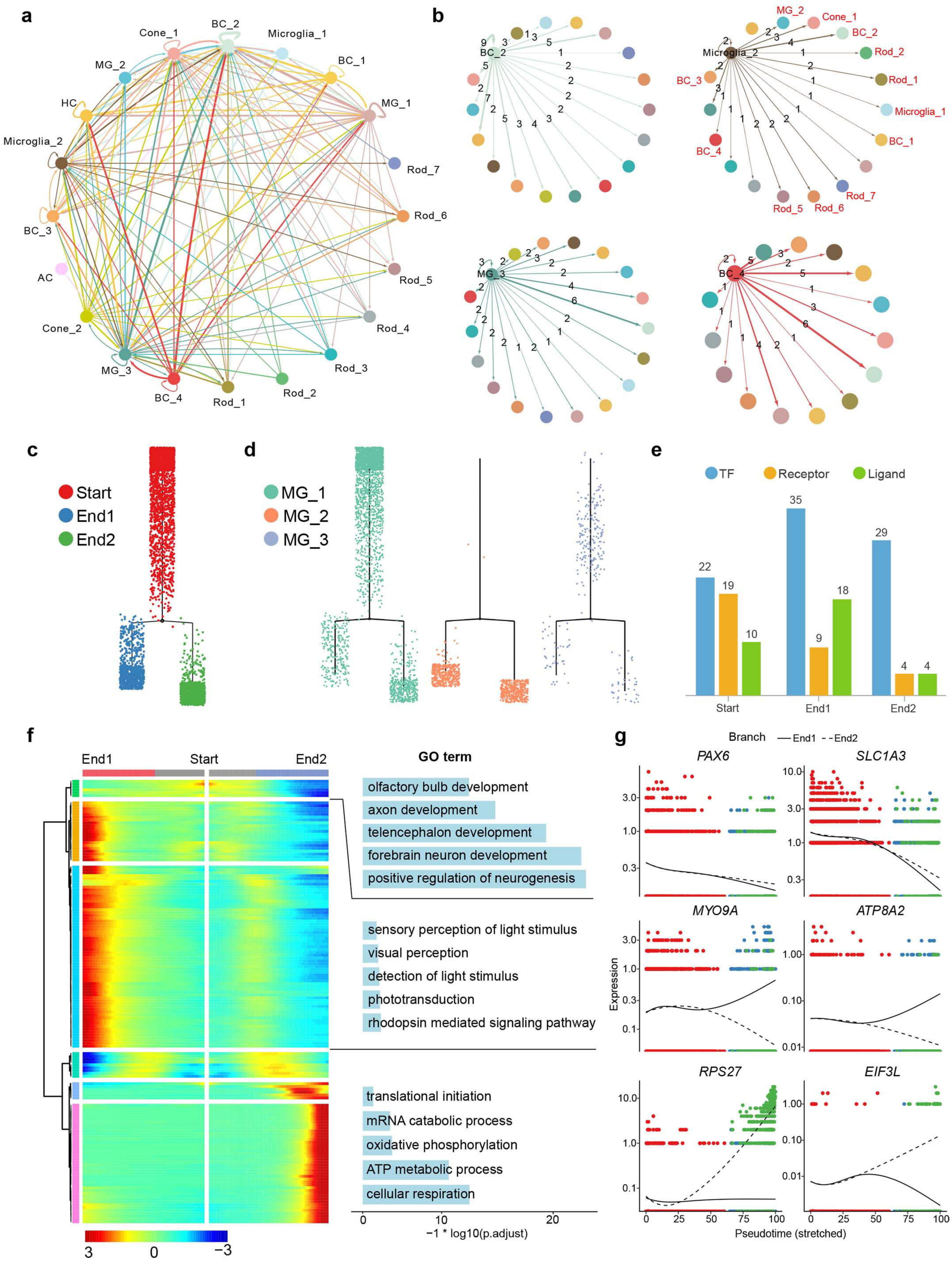
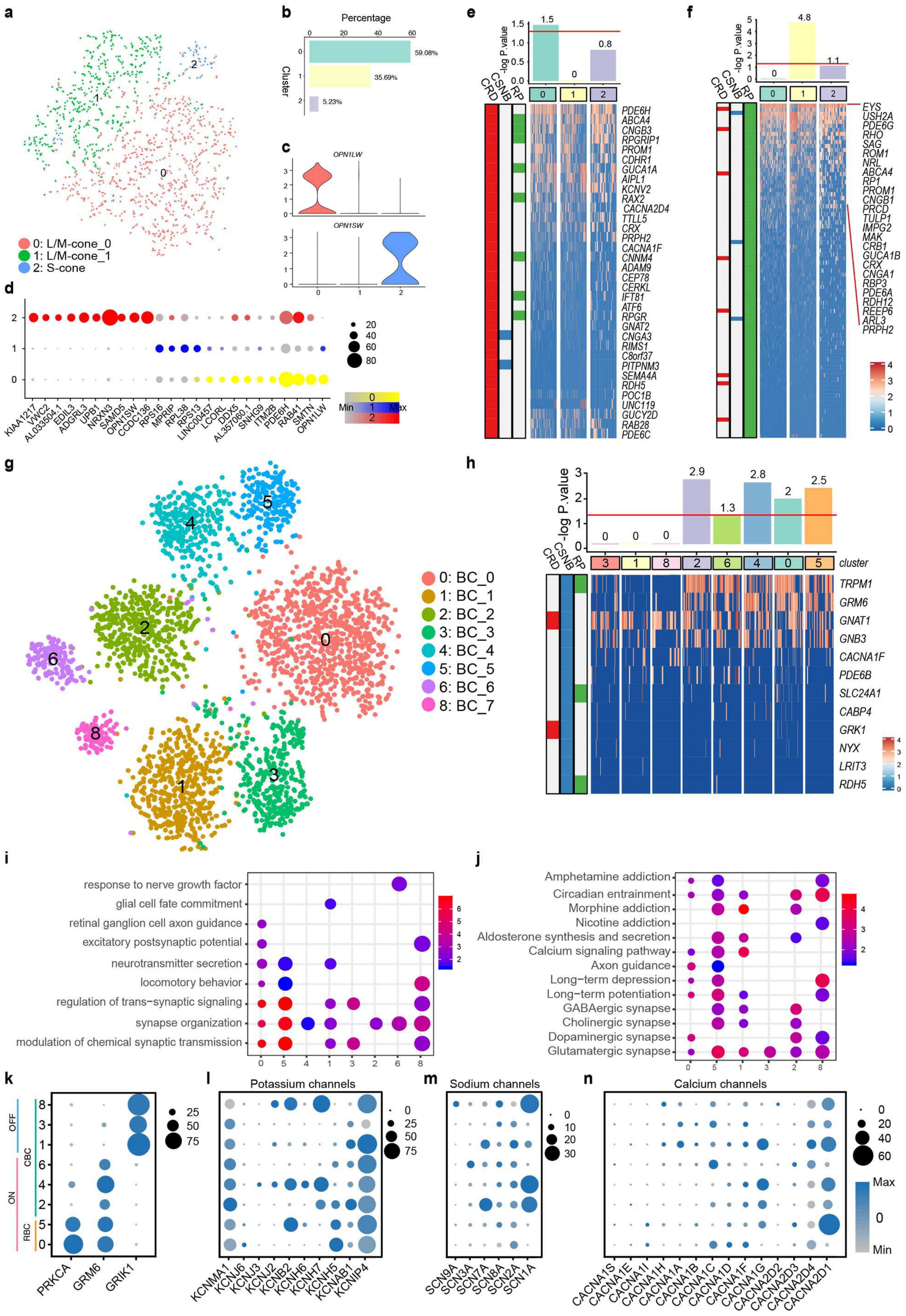

a
Cone_1
BC_2
Microglia_1
MG_2
BC_1
HC
MG_1
Microglia_2
Rod_7
Rod_6
BC_3
AC
Rod_5
Cone_2
Rod_4
MG_3
Rod_3
BC_4
Rod_1
Rod_2
b
BC_2
Microglia_2
MG_2
Cone_1
BC_2
Rod_2
BC_3
Rod_1
Microglia_1
BC_1
BC_4
Rod_7
Rod_5
Rod_6
MG_3
BC_4
c
Start
End1
End2
d
MG_1
MG_2
MG_3
e
TF
Receptor
Ligand
22
19
10
35
9
18
29
4
4
Start
End1
End2
f
End1
Start
End2
GO term
olfactory bulb development
axon development
telencephalon development
forebrain neuron development
positive regulation of neurogenesis
sensory perception of light stimulus
visual perception
detection of light stimulus
phototransduction
rhodopsin mediated signaling pathway
translational initiation
mRNA catabolic process
oxidative phosphorylation
ATP metabolic process
cellular respiration
0
10
20
−1 * log10(p.adjust)
3
0
−3
g
Branch
End1
End2
PAX6
SLC1A3
MYO9A
ATP8A2
RPS27
EIF3L
Expression
Pseudotime (stretched)

Figure 3


a
0: L/M-cone_0
1: L/M-cone_1
2: S-cone
b
Percentage
Cluster
59.08%
35.69%
5.23%
c
OPN1LW
OPN1SW
d
KIAA1217
VWC2
AL033504.1
EDIL3
ADGRL3
UPB1
NRXN3
SAMD5
OPN1SW
CCDC136
RPS16
MPRIP
RPL38
RPS13
LINC00457
LCORL
DDX5
AL357060.1
SNHG9
ITM2B
PDE6H
RAB41
SMTN
OPN1LW
Min
Max
e
-log P.value
CRD
CSNB
RP
PDE6H
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PROM1
CDHR1
GUCA1A
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TTLL5
CRX
PRPH2
CACNA1F
CNNM4
ADAM9
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CERKL
IFT81
ATF6
RPGR
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RDH5
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PDE6C
f
EYS
USH2A
PDE6G
RHO
SAG
ROM1
NRL
ABCA4
RP1
PROM1
CNGB1
PRCD
TULP1
IMPG2
MAK
CRB1
GUCA1B
CRX
CNGA1
RBP3
PDE6A
RDH12
REEP6
ARL3
PRPH2
g
0: BC_0
1: BC_1
2: BC_2
3: BC_3
4: BC_4
5: BC_5
6: BC_6
8: BC_7
h
-log P.value
cluster
TRPM1
GRM6
GNAT1
GNB3
CACNA1F
PDE6B
SLC24A1
CABP4
GRK1
NYX
LRIT3
RDH5
i
response to nerve growth factor
glial cell fate commitment
retinal ganglion cell axon guidance
excitatory postsynaptic potential
neurotransmitter secretion
locomotory behavior
regulation of trans−synaptic signaling
synapse organization
modulation of chemical synaptic transmission
j
Amphetamine addiction
Circadian entrainment
Morphine addiction
Nicotine addiction
Aldosterone synthesis and secretion
Calcium signaling pathway
Axon guidance
Long−term depression
Long−term potentiation
GABAergic synapse
Cholinergic synapse
Dopaminergic synapse
Glutamatergic synapse
k
OFF
ON
CBC
RBC
PRKCA
GRM6
GRIK1
l
Potassium channels
KCNMA1
KCNJ6
KCNJ3
KCNJ2
KCNB2
KCNH6
KCNH7
KCNH5
KCNAB1
KCNIP4
m
Sodium channels
SCN9A
SCN3A
SCN7A
SCN8A
SCN2A
SCN1A
n
Calcium channels
CACNA1S
CACNA1E
CACNA1I
CACNA1H
CACNA1A
CACNA1B
CACNA1C
CACNA1D
CACNA1F
CACNA1G
CACNA2D2
CACNA2D3
CACNA2D4
CACNA2D1
Max
Min

Figure 4

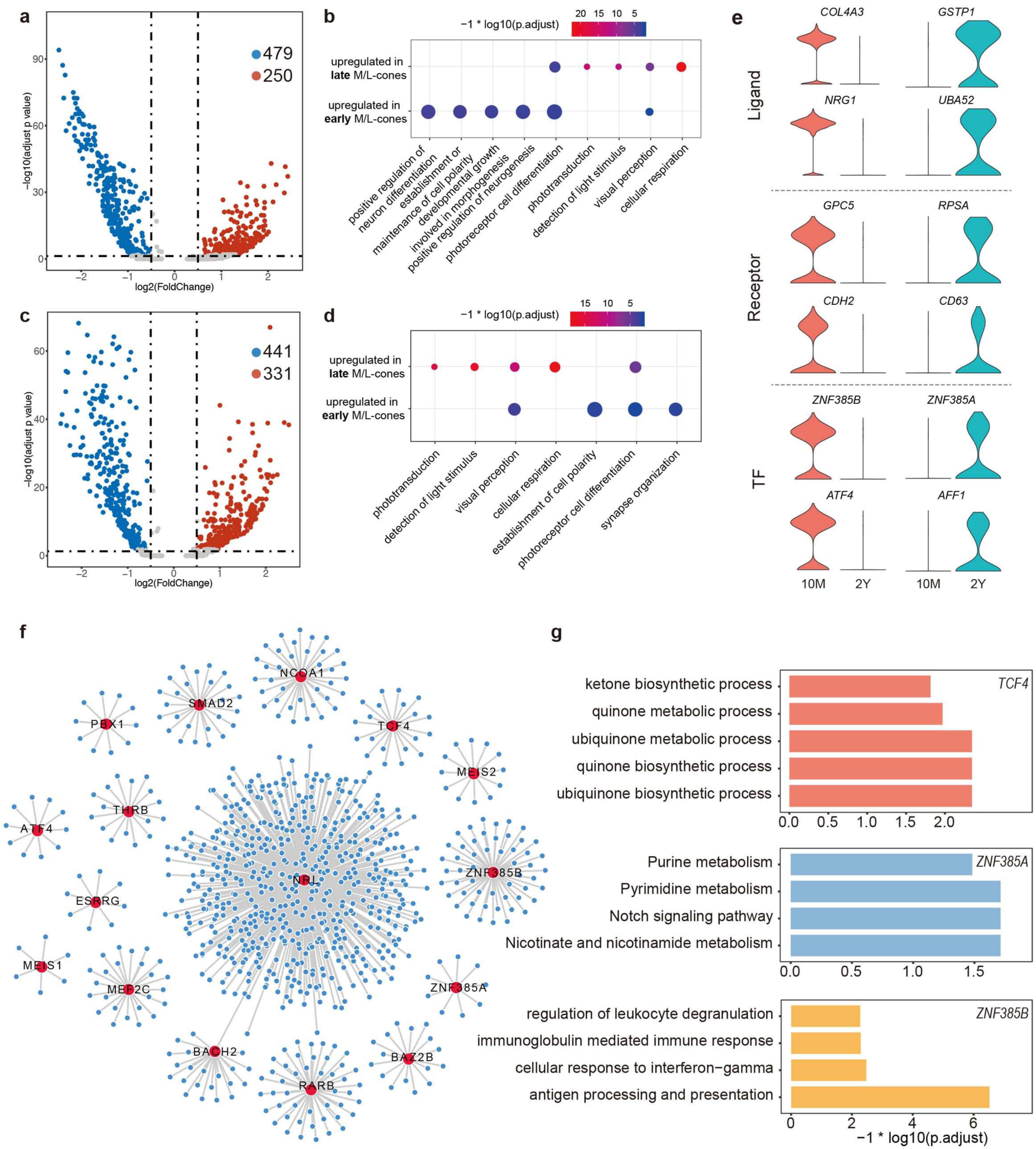
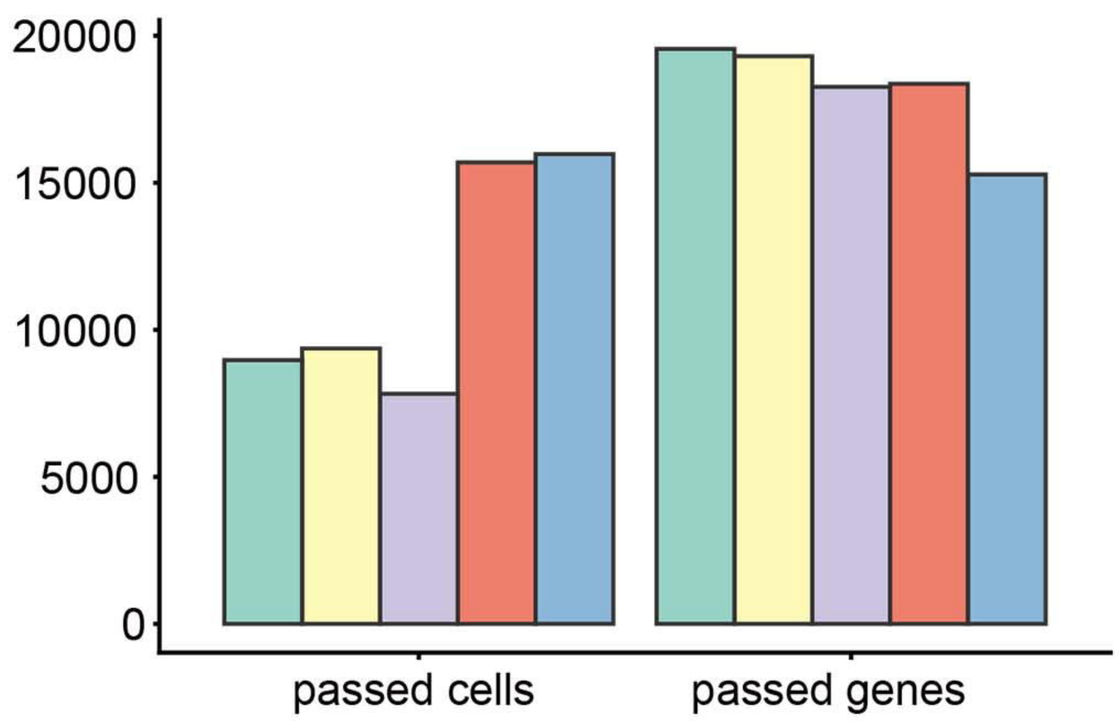

a
479
250
−log10(adjust p value)
log2(FoldChange)
b
−1 * log10(p.adjust)
upregulated in late M/L-cones
upregulated in early M/L-cones
positive regulation of neuron differentiation
establishment or maintenance of cell polarity
developmental growth involved in morphogenesis
positive regulation of photoreceptor cell differentiation
phototransduction
detection of light stimulus
visual perception
cellular respiration
c
441
331
−log10(adjust p value)
log2(FoldChange)
d
−1 * log10(p.adjust)
upregulated in late M/L-cones
upregulated in early M/L-cones
phototransduction
detection of light stimulus
visual perception
cellular respiration
establishment of cell polarity
photoreceptor cell differentiation
synapse organization
e
Ligand
COL4A3
GSTP1
NRG1
UBA52
Receptor
GPC5
RPSA
CDH2
CD63
TF
ZNF385B
ZNF385A
ATF4
AFF1
10M
2Y
10M
2Y
f
NCOA1
SMAD2
PBX1
TCF4
MEIS2
THRB
ATF4
NRL
ZNF385B
ESRRG
MEIS1
MEF2C
ZNF385A
BACH2
BAZ2B
RARB
g
ketone biosynthetic process
quinone metabolic process
ubiquinone metabolic process
quinone biosynthetic process
ubiquinone biosynthetic process
TCF4
Purine metabolism
Pyrimidine metabolism
Notch signaling pathway
Nicotinate and nicotinamide metabolism
ZNF385A
regulation of leukocyte degranulation
immunoglobulin mediated immune response
cellular response to interferon−gamma
antigen processing and presentation
ZNF385B
−1 * log10(p.adjust)

Figure S1

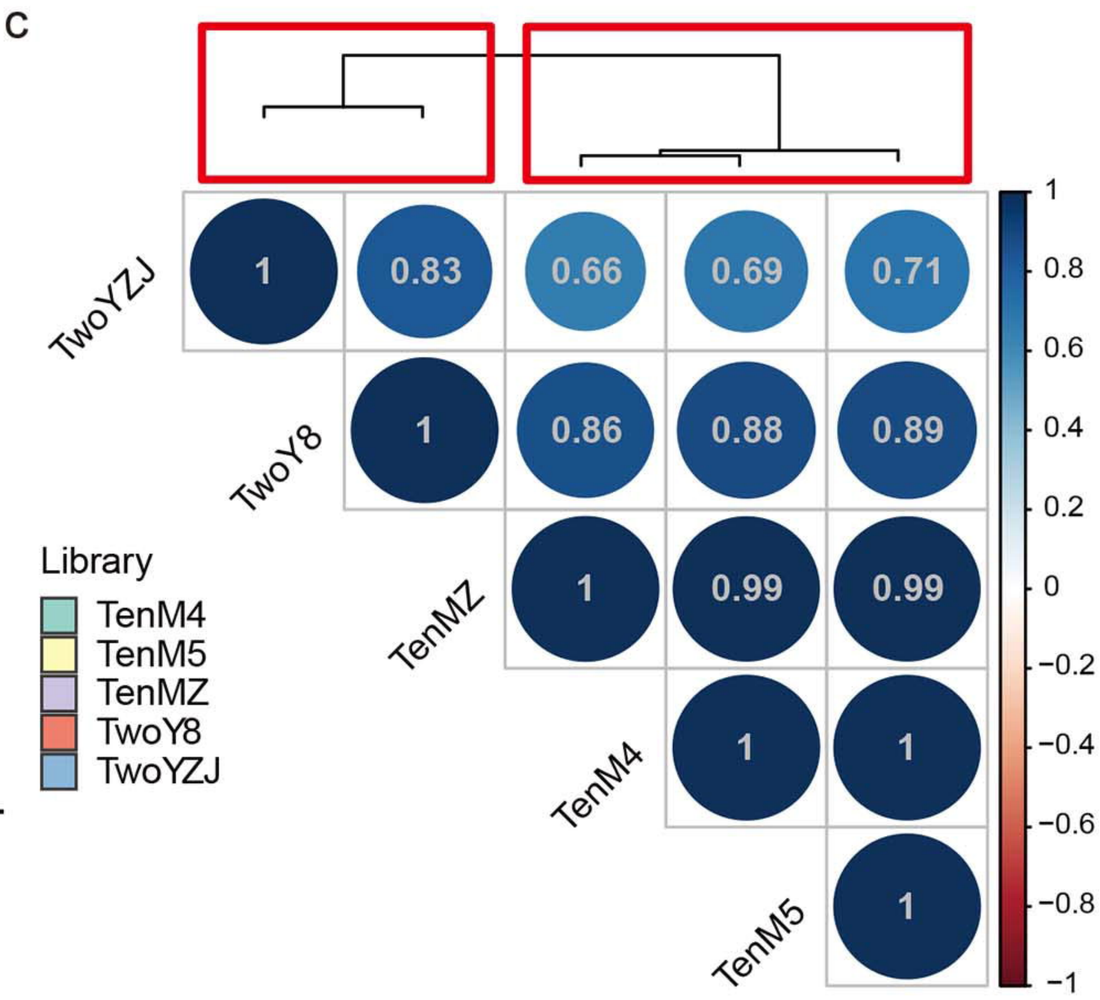
a

Library	TenM4	TenM5	TenMZ	TwoY8	TwoYZJ
Sample	SampleA	SampleA	SampleA	SampleB	SampleB
Age	10 Months	10 Months	10 Months	2 Years	2 Years
Mean reads per cell	128,753	58,402	38,802	39,066	22,701
Median genes per cell	461	381	426	680	513
Sequencing saturation	90.80%	89.10%	81.10%	85.10%	82.70%

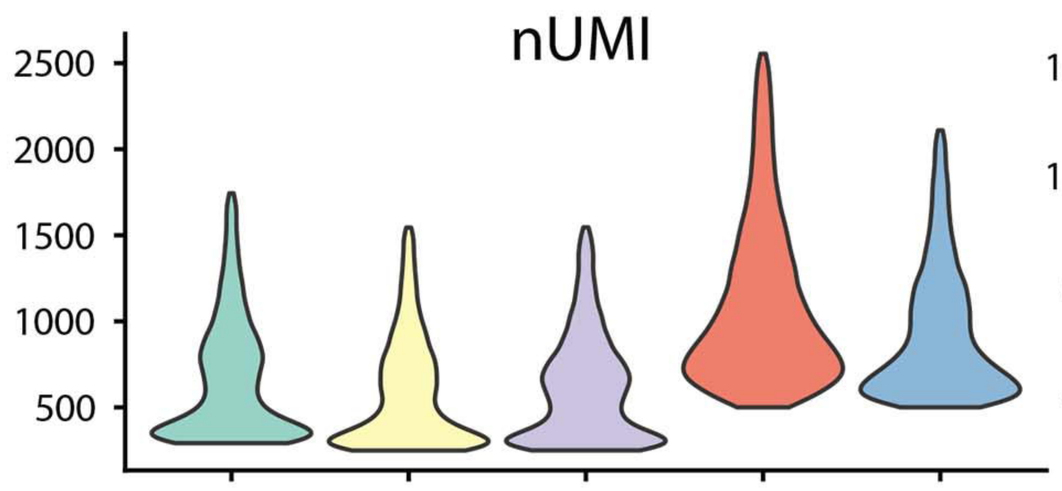
b

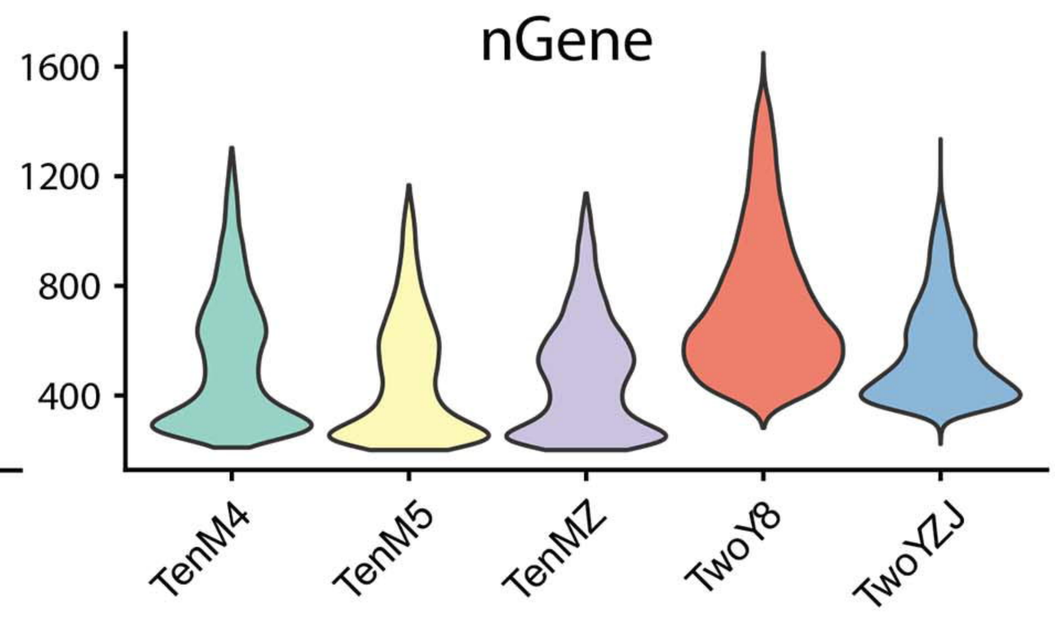


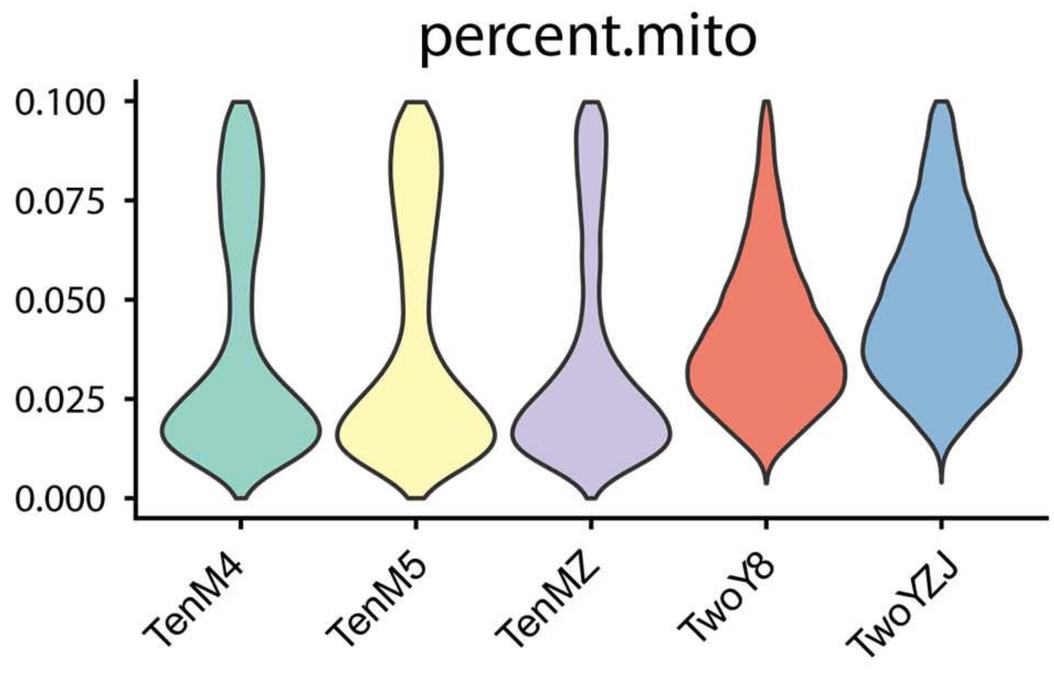
c



d







e

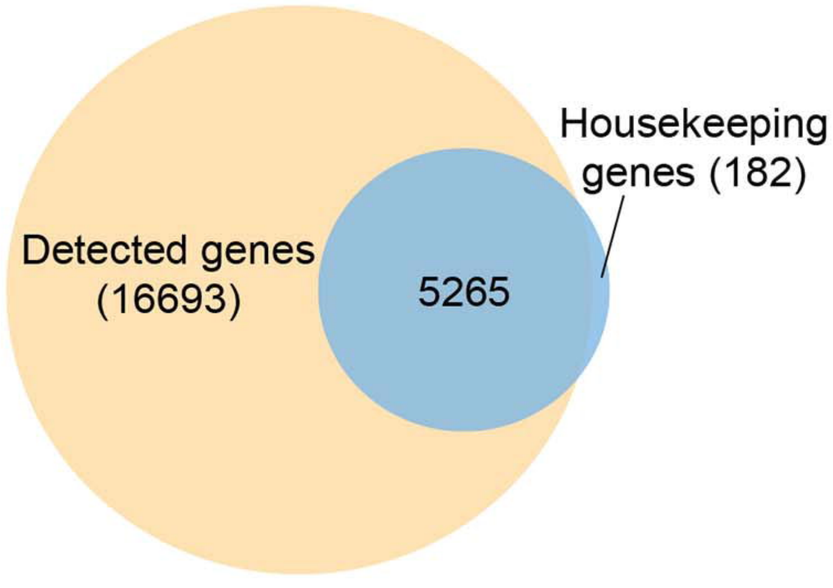
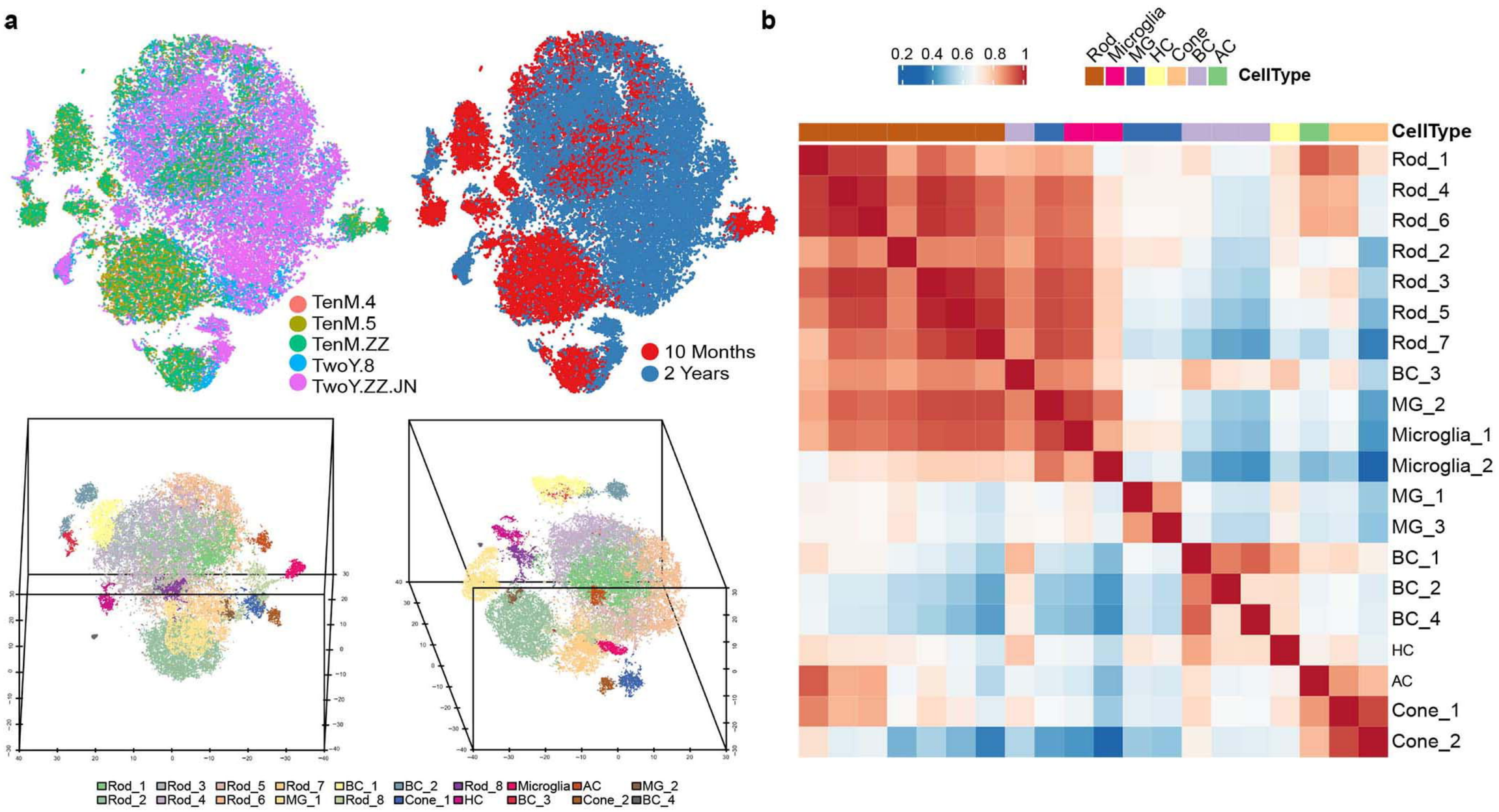


Figure S2


a
TenM.4
TenM.5
TenM.ZZ
TwoY.8
TwoY.ZZ.JN
10 Months
2 Years
Rod_1 Rod_2 Rod_3 Rod_4 Rod_5 Rod_6 Rod_7 MG_1 Rod_8 BC_1 BC_2 Cone_1 Rod_8 HC Microglia BC_3 AC Cone_2 MG_2 BC_4
b
0.2 0.4 0.6 0.8 1
Rod Microglia MG HC Cone BC AC
CellType
Rod_1
Rod_4
Rod_6
Rod_2
Rod_3
Rod_5
Rod_7
BC_3
MG_2
Microglia_1
Microglia_2
MG_1
MG_3
BC_1
BC_2
BC_4
HC
AC
Cone_1
Cone_2

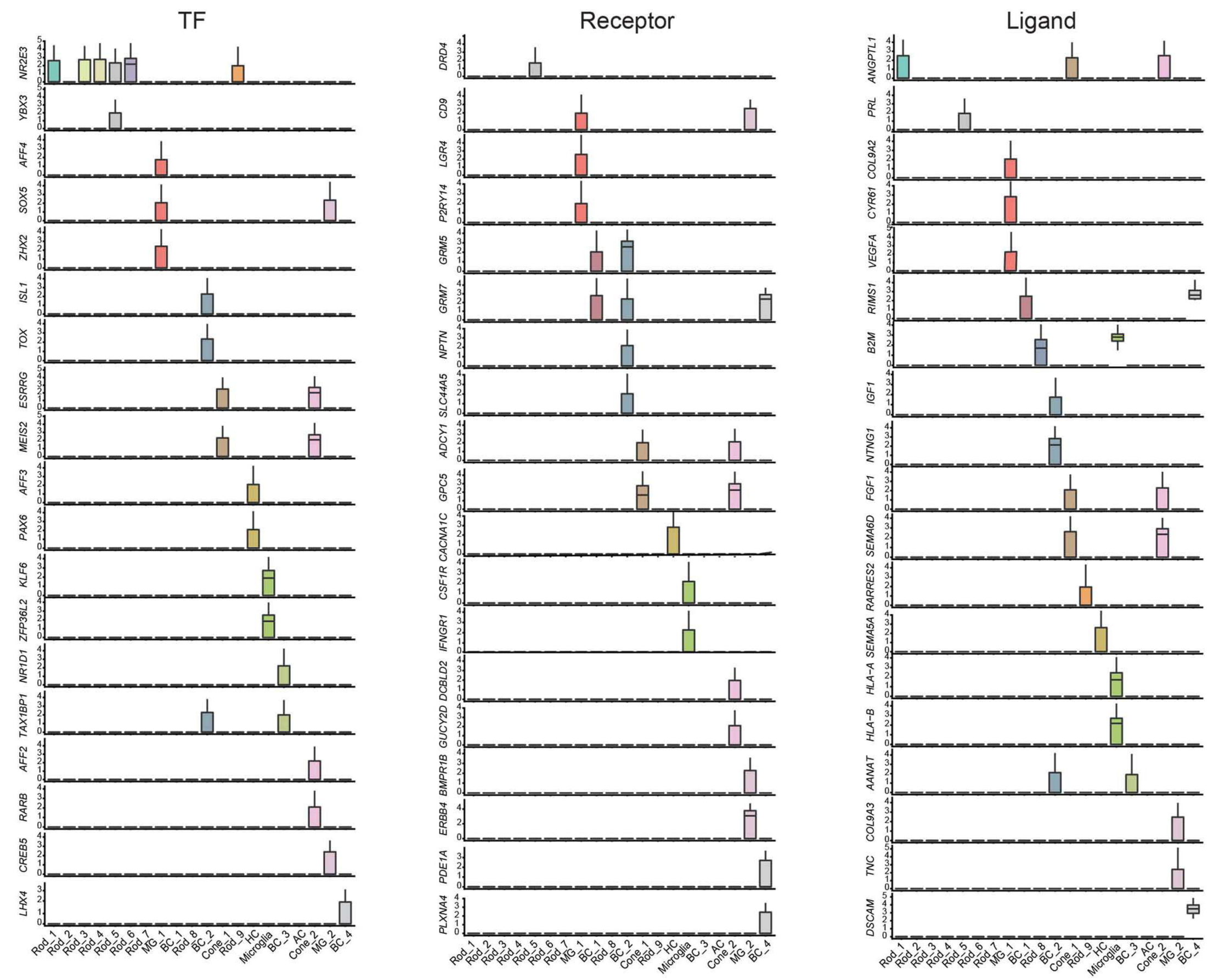

c
TF
Receptor
Ligand
NR2E3
YBX3
AFF4
SOX5
ZHX2
ISL1
TOX
ESRRG
MEIS2
AFF3
PAX6
KLF6
ZFP36L2
NR1D1
TAX1BP1
AFF2
RARB
CREB5
LHX4
DRD4
CD9
LGR4
P2RY14
GRM5
GRM7
NPTN
SLC44A5
ADCY1
GPC5
CACNA1C
CSF1R
IFNGR1
DCBLD2
GUCY2D
BMPR1B
ERBB4
PDE1A
PLXNA4
ANGPTL1
PRL
COL9A2
CYR61
VEGFA
RIMS1
B2M
IGF1
NTNG1
FGF1
SEMA6D
RARRES2
SEMA5A
HLA-A
HLA-B
AANAT
COL9A3
TNC
DSCAM
Rod_1 Rod_2 Rod_3 Rod_4 Rod_5 Rod_6 Rod_7 MG_1 BC_1 Rod_8 BC_2 Cone_1 Rod_9 HC Microglia BC_3 AC Cone_2 MG_2 BC_4

Figure S3

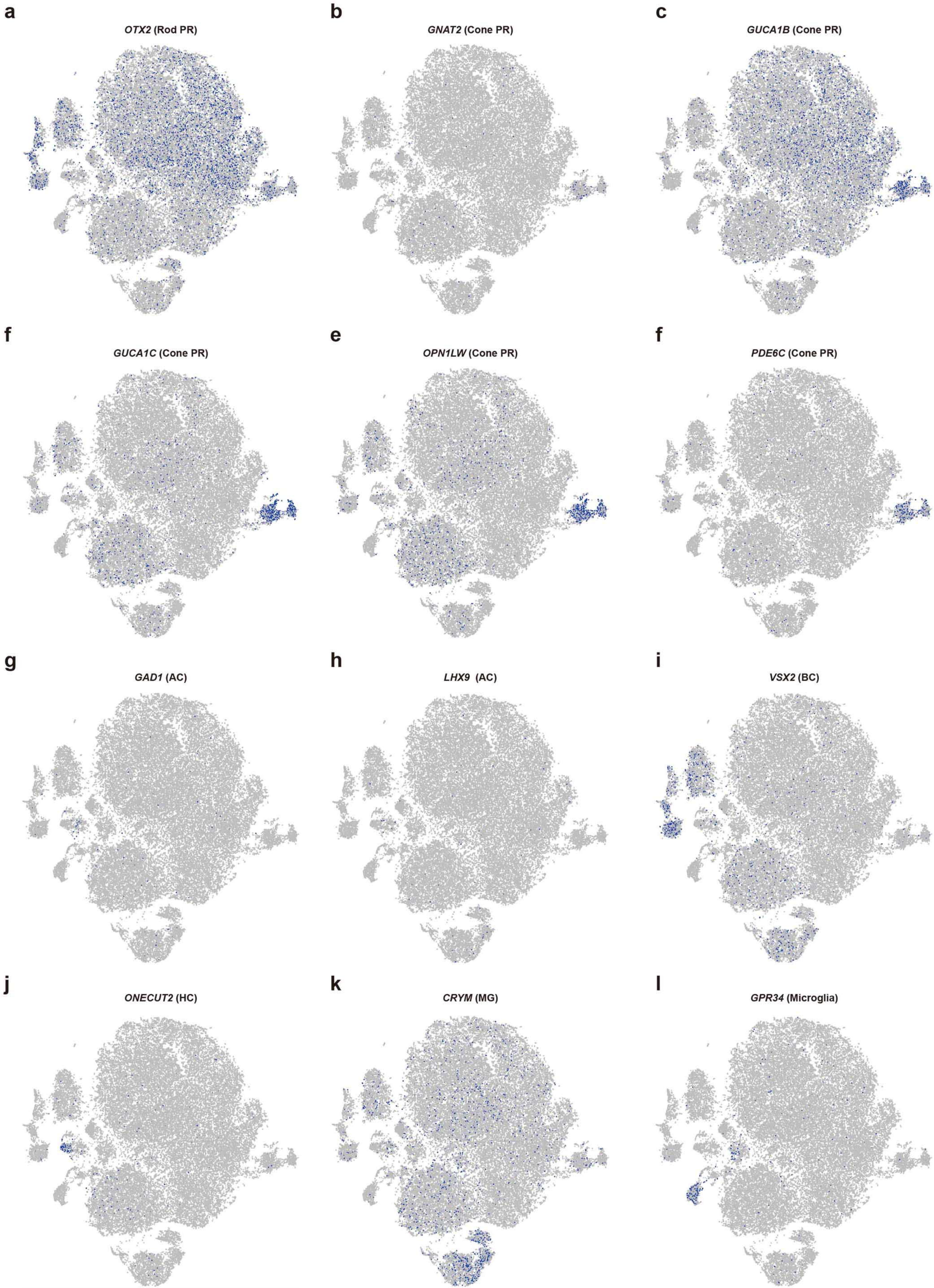

a
OTX2 (Rod PR)
b
GNAT2 (Cone PR)
c
GUCA1B (Cone PR)
f
GUCA1C (Cone PR)
e
OPN1LW (Cone PR)
f
PDE6C (Cone PR)
g
GAD1 (AC)
h
LHX9 (AC)
i
VSX2 (BC)
j
ONECUT2 (HC)
k
CRYM (MG)
l
GPR34 (Microglia)

Figure S4

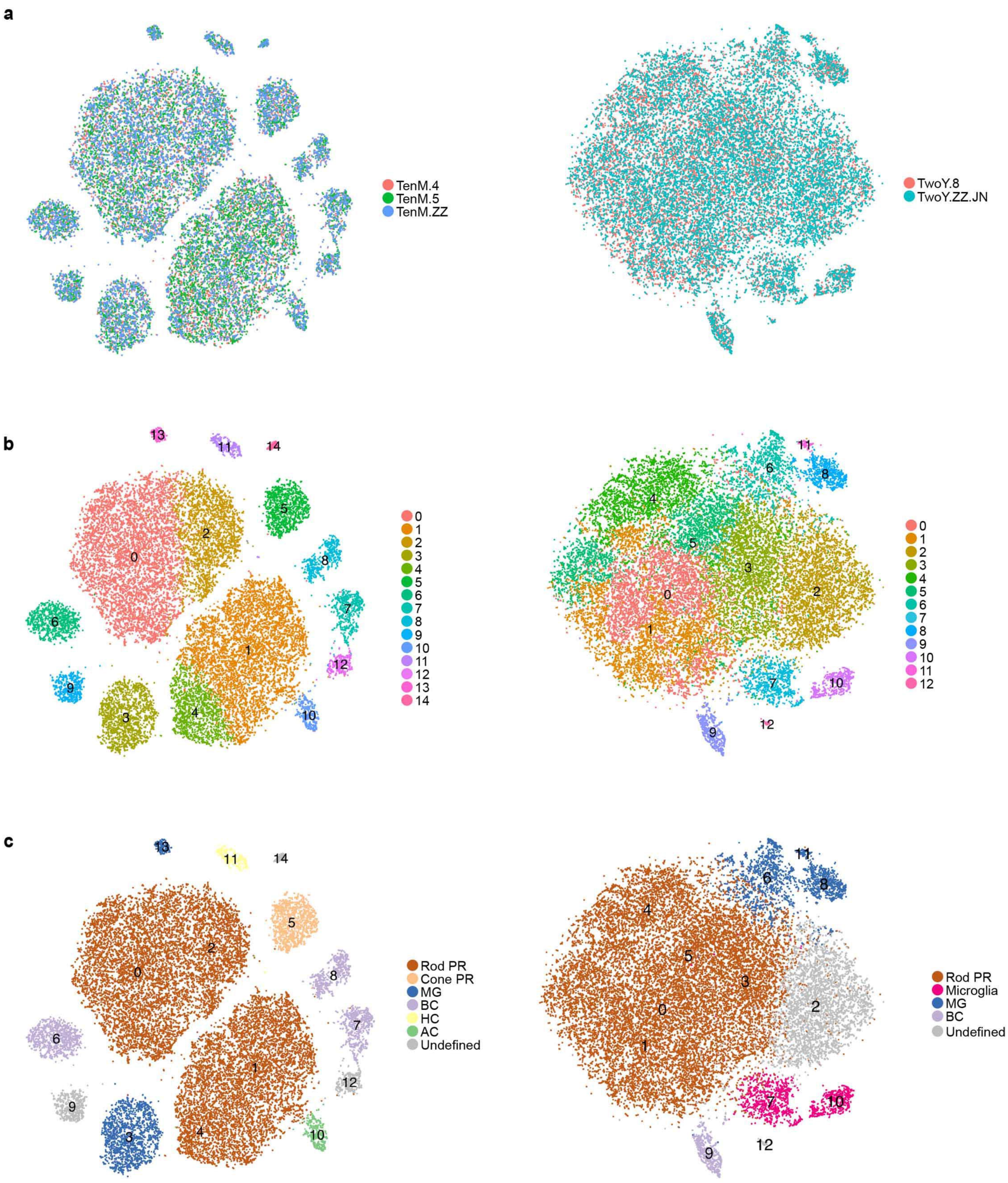

a
TenM.4
TenM.5
TenM.ZZ
TwoY.8
TwoY.ZZ.JN
b
0
1
2
3
4
5
6
7
8
9
10
11
12
13
14
c
Rod PR
Cone PR
MG
BC
HC
AC
Undefined
Rod PR
Microglia
MG
BC
Undefined

Figure S5

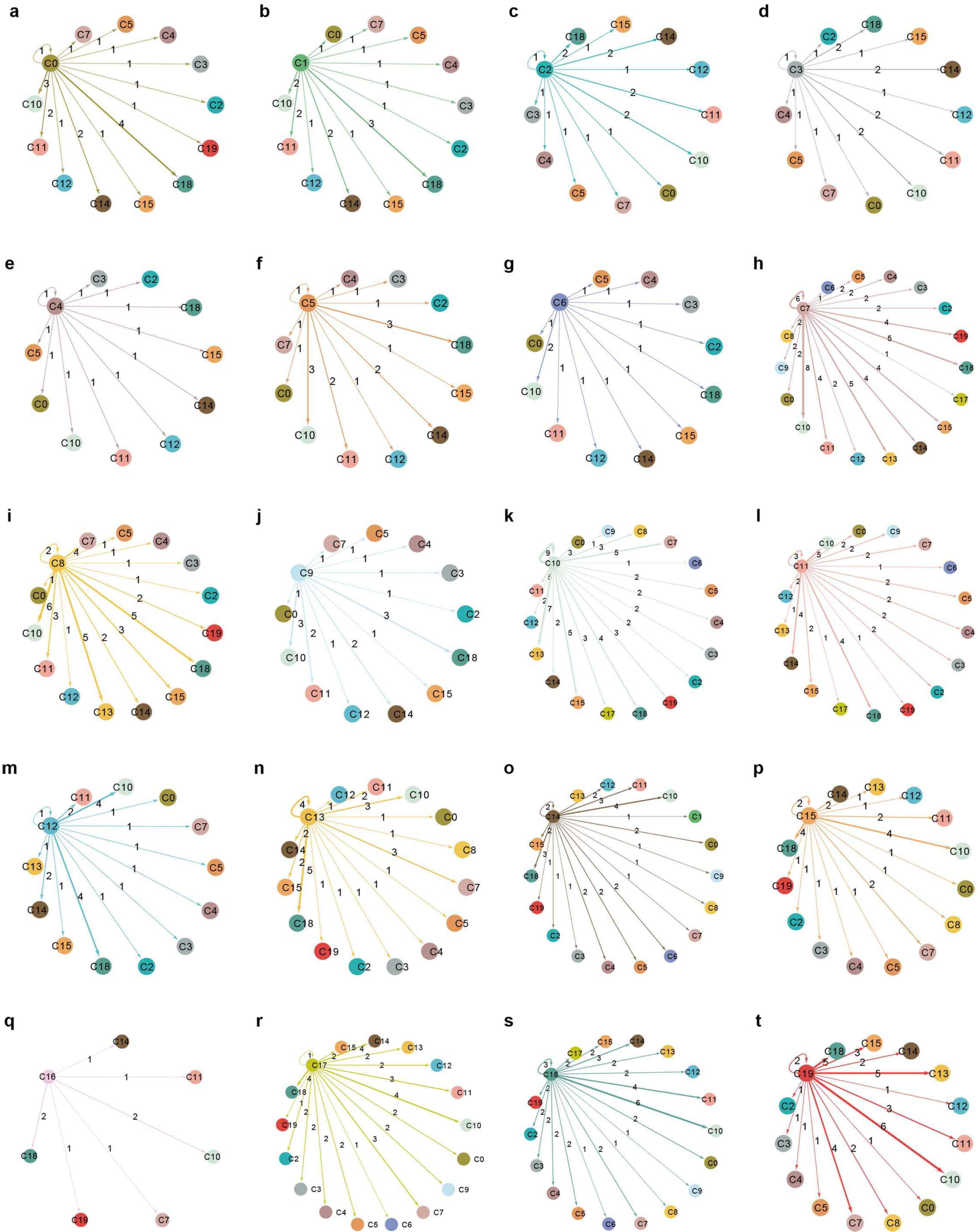


Figure S6

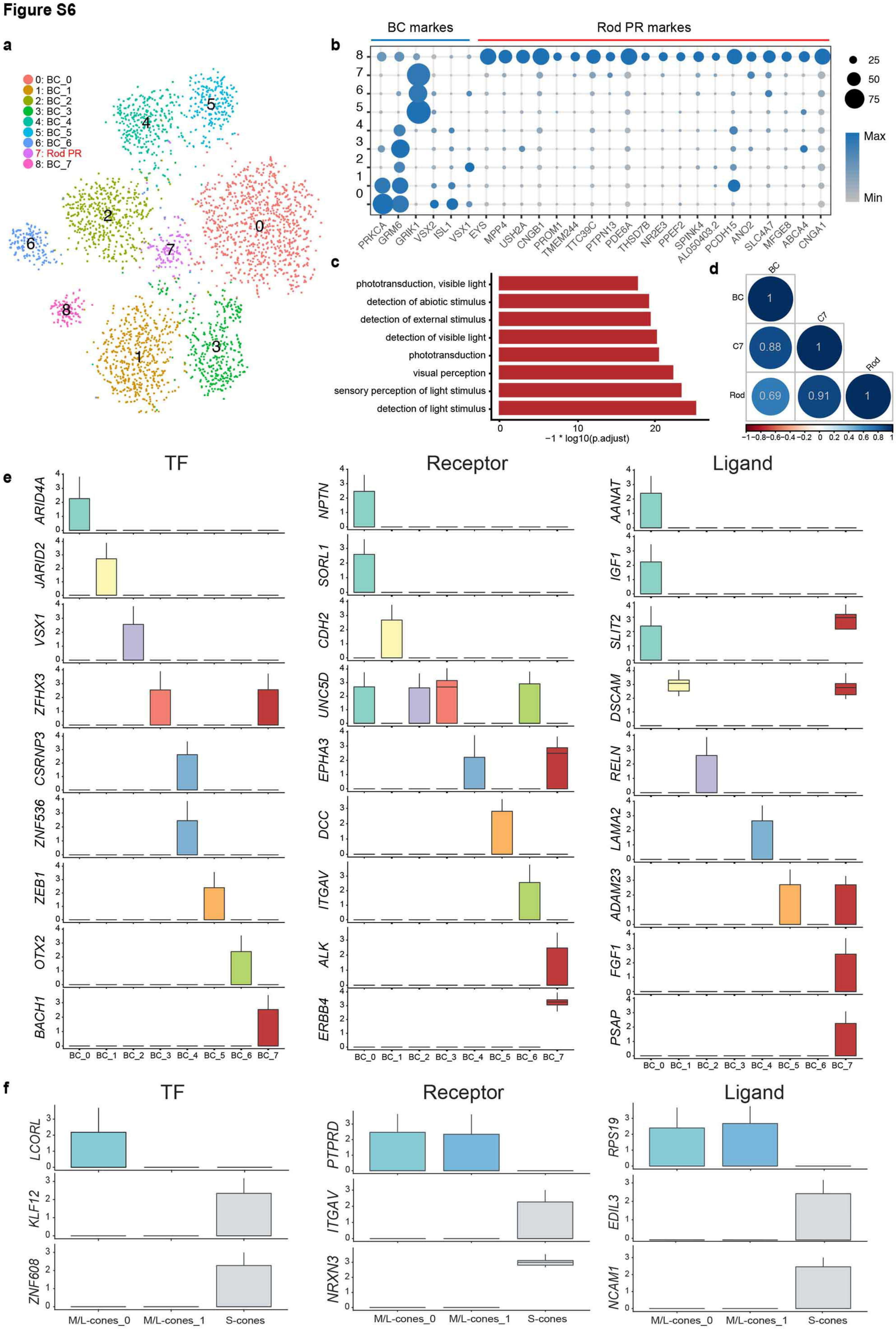

a
0: BC_0
1: BC_1
2: BC_2
3: BC_3
4: BC_4
5: BC_5
6: BC_6
7: Rod PR
8: BC_7
b
BC markes
Rod PR markes
PRKCA GRM6 GRIK1 VSX2 ISL1 VSX1 EYS MPP4 USH2A CNGB1 PROM1 TMEM244 TTC39C PTPN13 PDE6A THSD7B NR2E3 PPEF2 SPINK4 AL050403.2 PCDH15 ANO2 SLC4A7 MFGE8 ABCA4 CNGA1
25
50
75
Max
Min
c
phototransduction, visible light
detection of abiotic stimulus
detection of external stimulus
detection of visible light
phototransduction
visual perception
sensory perception of light stimulus
detection of light stimulus
−1 * log10(p.adjust)
d
BC
C7
Rod
0.88
0.69
0.91
e
TF
Receptor
Ligand
ARID4A
JARID2
VSX1
ZFHX3
CSRNP3
ZNF536
ZEB1
OTX2
BACH1
NPTN
SORL1
CDH2
UNC5D
EPHA3
DCC
ITGAV
ALK
ERBB4
AANAT
IGF1
SLIT2
DSCAM
RELN
LAMA2
ADAM23
FGF1
PSAP
BC_0 BC_1 BC_2 BC_3 BC_4 BC_5 BC_6 BC_7
f
TF
Receptor
Ligand
LCORL
KLF12
ZNF608
PTPRD
ITGAV
NRXN3
RPS19
EDIL3
NCAM1
M/L-cones_0
M/L-cones_1
S-cones

Figure S7

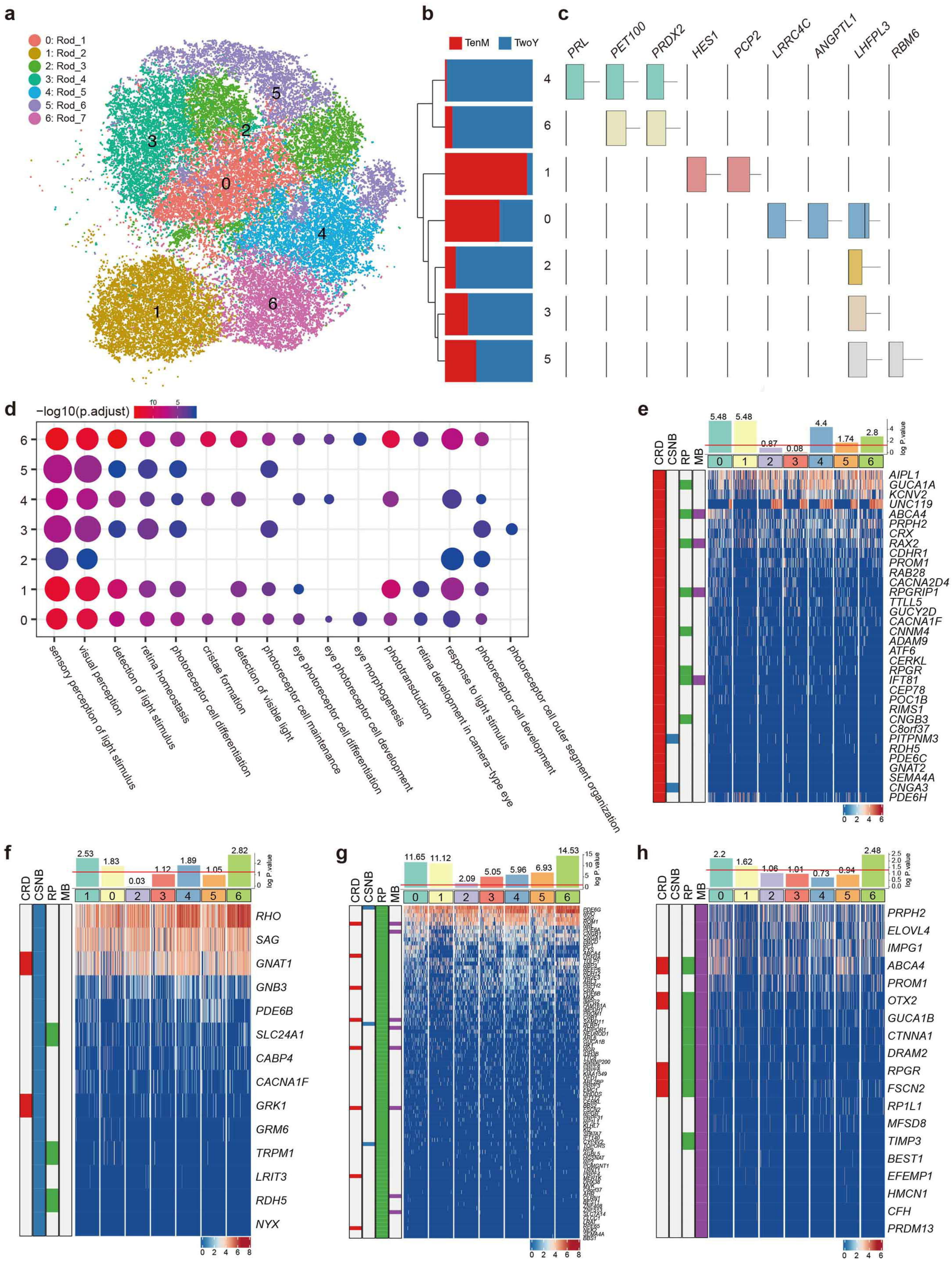

a
0: Rod_1
1: Rod_2
2: Rod_3
3: Rod_4
4: Rod_5
5: Rod_6
6: Rod_7
b
TenM
TwoY
c
PRL
PET100
PRDX2
HES1
PCP2
LRRC4C
ANGPTL1
LHFPL3
RBM6
d
−log10(p.adjust)
sensory perception of light stimulus
visual perception
detection of light stimulus
retina homeostasis
photoreceptor cell differentiation
cristae formation
detection of visible light
photoreceptor cell maintenance
eye photoreceptor cell differentiation
eye photoreceptor cell development
eye morphogenesis
phototransduction
retina development in camera-type eye
response to light stimulus
photoreceptor cell development
photoreceptor cell outer segment organization
e
CRD
CSNB
RP
MB
log P.value
AIPL1
GUCA1A
KCNV2
UNC119
ABCA4
PRPH2
CRX
RAX2
CDHR1
PROM1
RAB28
CACNA2D4
RPGRIP1
TTLL5
GUCY2D
CACNA1F
CNNM4
ADAM9
ATF6
CERKL
RPGR
IFT81
CEP78
POC1B
RIMS1
CNGB3
C8orf37
PITPNM3
RDH5
PDE6C
GNAT2
SEMA4A
CNGA3
PDE6H
f
RHO
SAG
GNAT1
GNB3
PDE6B
SLC24A1
CABP4
CACNA1F
GRK1
GRM6
TRPM1
LRIT3
RDH5
NYX
g
h
PRPH2
ELOVL4
IMPG1
ABCA4
PROM1
OTX2
GUCA1B
CTNNA1
DRAM2
RPGR
FSCN2
RP1L1
MFSD8
TIMP3
BEST1
EFEMP1
HMCN1
CFH
PRDM13

Figure S8

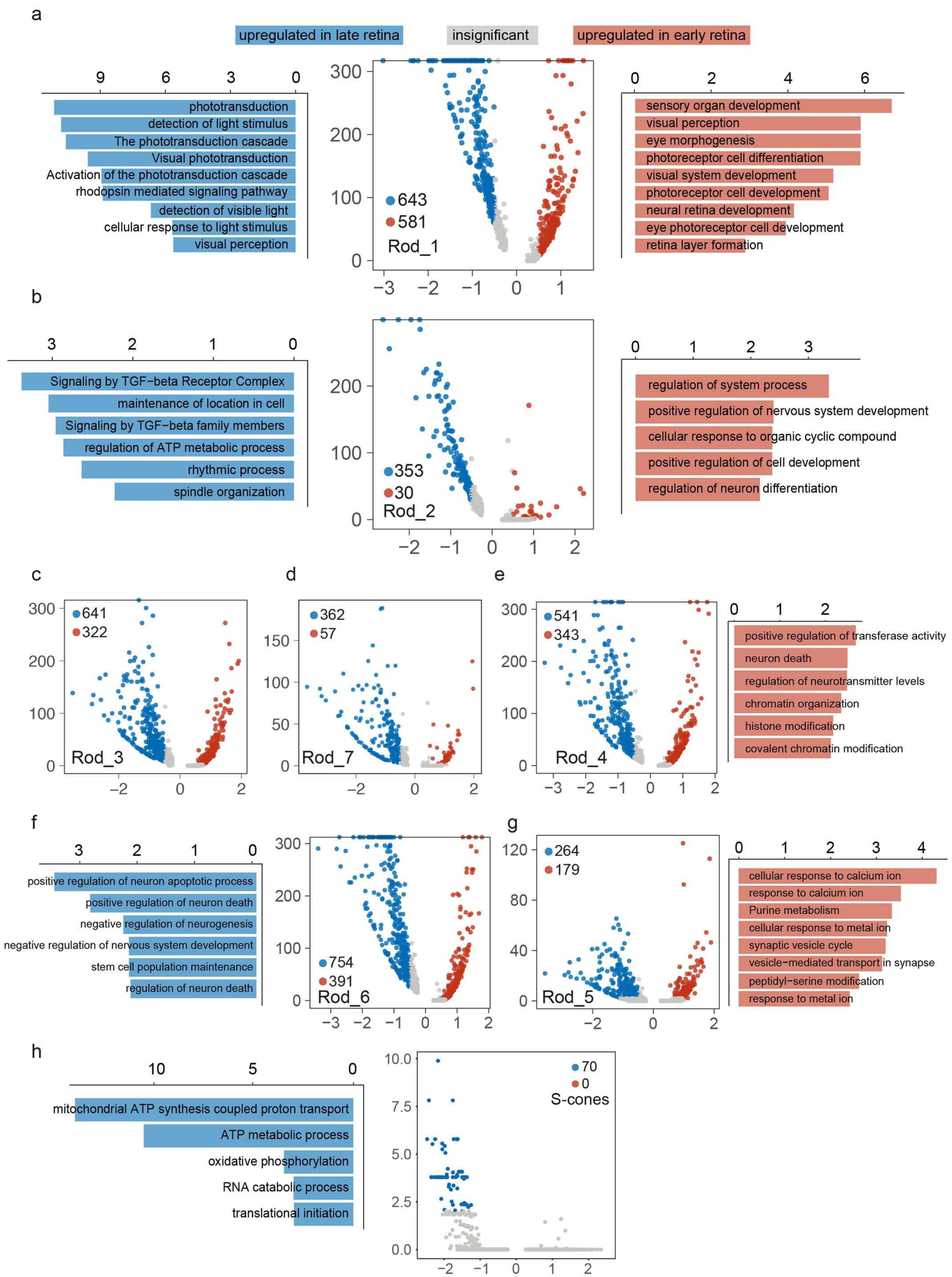

a
upregulated in late retina
insignificant
upregulated in early retina
phototransduction
detection of light stimulus
The phototransduction cascade
Visual phototransduction
Activation of the phototransduction cascade
rhodopsin mediated signaling pathway
detection of visible light
cellular response to light stimulus
visual perception
643
581
Rod_1
sensory organ development
visual perception
eye morphogenesis
photoreceptor cell differentiation
visual system development
photoreceptor cell development
neural retina development
eye photoreceptor cell development
retina layer formation
b
Signaling by TGF−beta Receptor Complex
maintenance of location in cell
Signaling by TGF−beta family members
regulation of ATP metabolic process
rhythmic process
spindle organization
353
30
Rod_2
regulation of system process
positive regulation of nervous system development
cellular response to organic cyclic compound
positive regulation of cell development
regulation of neuron differentiation
c
641
322
Rod_3
d
362
57
Rod_7
e
541
343
Rod_4
positive regulation of transferase activity
neuron death
regulation of neurotransmitter levels
chromatin organization
histone modification
covalent chromatin modification
f
positive regulation of neuron apoptotic process
positive regulation of neuron death
negative regulation of neurogenesis
negative regulation of nervous system development
stem cell population maintenance
regulation of neuron death
754
391
Rod_6
g
264
179
Rod_5
cellular response to calcium ion
response to calcium ion
Purine metabolism
cellular response to metal ion
synaptic vesicle cycle
vesicle−mediated transport in synapse
peptidyl−serine modification
response to metal ion
h
mitochondrial ATP synthesis coupled proton transport
ATP metabolic process
oxidative phosphorylation
RNA catabolic process
translational initiation
70
0
S-cones

Figure S9

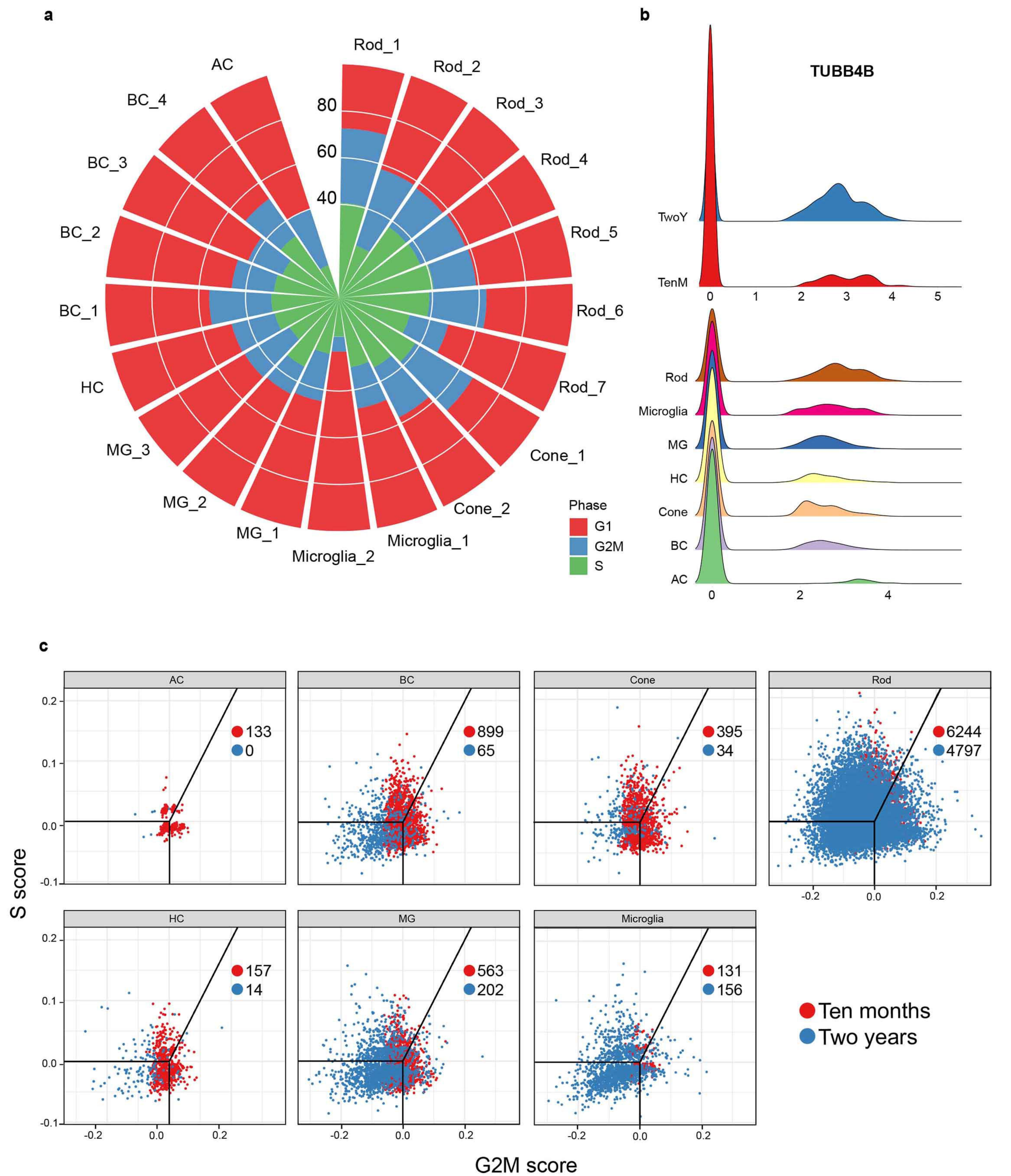

a
Rod_1
Rod_2
Rod_3
Rod_4
Rod_5
Rod_6
Rod_7
Cone_1
Cone_2
Microglia_1
Microglia_2
MG_1
MG_2
MG_3
HC
BC_1
BC_2
BC_3
BC_4
AC
80
60
40
Phase
G1
G2M
S
b
TUBB4B
TwoY
TenM
0
1
2
3
4
5
Rod
Microglia
MG
HC
Cone
BC
AC
0
2
4
c
AC
BC
Cone
Rod
HC
MG
Microglia
133
0
899
65
395
34
6244
4797
157
14
563
202
131
156
0.2
0.1
0.0
-0.1
-0.2
0.0
0.2
S score
G2M score
Ten months
Two years

Figure S10

a
ADCY1
ADK
ARR3
BTBD9
FRMPD1
GLMN
KIF2A
PCMTD2
SV2B
STXBP1
b
CSTB
FUS
H1F0
H3F3A
HMGB2
TSC22D3
c
ATP1B3
EPAS1
MXI1
MKNK2
IDI1
d
NR2E3
PDE6B
e
TRPM1
PER3
KHDRBS3
f
ACTB
ACTG1
PPP2R2B
RIMS2
AC
BC
Cone
HC
MG
Microglia
Rod